



Effect of Important Food Sources of Fructose-Containing Sugars on Inflammatory Biomarkers: A Systematic Review and Meta-Analysis of Controlled Feeding Trials

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Abstract: Background: Fructose-containing sugars as sugar-sweetened beverages (SSBs) may increase inflammatory biomarkers. Whether this effect is mediated by the food matrix at different levels of energy is unknown. To investigate the role of food source and energy, we conducted a systematic review and meta-analysis of controlled trials on the effect of different food sources of fructose-containing sugars on inflammatory markers at different levels of energy control. Methods: MEDLINE, Embase, and the Cochrane Library were searched through March 2022 for controlled feeding trials \geq 7 days. Four trial designs were prespecified by energy control: substitution (energy matched replacement of sugars); addition (excess energy from sugars added to diets); subtraction (energy from sugars subtracted from diets); and ad libitum (energy from sugars freely replaced). The primary outcome was C-reactive protein (CRP). Secondary outcomes were tumour necrosis factor-alpha (TNF- α) and interleukin-6 (IL-6). Independent reviewers extracted data and assessed risk of bias. GRADE assessed certainty of evidence. Results: We identified 64 controlled trials (91 trial comparisons, n = 4094) assessing 12 food sources (SSB; sweetened dairy; sweetened dairy alternative [soy]; 100% fruit juice; fruit; dried fruit; mixed fruit forms; sweetened cereal grains and bars; sweets and desserts; added nutritive [caloric] sweetener; mixed sources [with SSBs]; and mixed sources [without SSBs]) at 4 levels of energy control over a median 6-weeks in predominantly healthy mixed weight or overweight/obese adults. Total fructose-containing sugars decreased CRP in addition trials and had no effect in substitution, subtraction or ad libitum trials. No effect was observed on other outcomes at any level of energy control. There was evidence of interaction/influence by food source: substitution trials (sweetened dairy alternative (soy) and 100% fruit juice decreased, and mixed sources (with SSBs) increased CRP); and addition trials (fruit decreased CRP and TNF- α ; sweets and desserts (dark chocolate) decreased IL-6). The certainty of evidence was moderate-to-low for the majority of analyses. Conclusions: Food source appears to mediate the effect of fructose-containing sugars on inflammatory markers over the short-to-medium term. The evidence provides good indication that mixed sources that contain SSBs increase CRP, while most other food sources have no effect with some sources (fruit, 100% fruit juice, sweetened soy beverage or dark chocolate) showing decreases, which may be dependent on energy control. Clinicaltrials.gov: (NCT02716870).



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Keywords: inflammation; CRP; fructose; sugars; food sources; fruit; fruit juice; sugar-sweetened beverages; systematic review; meta-analysis

1. Introduction

Chronic inflammation resulting in elevated levels of inflammatory biomarkers have been associated with a higher risk for the development of cardiovascular disease (CVD), diabetes, and other non-communicable diseases [1]. Randomized controlled trials have shown that agents that decrease inflammation (e.g., canakinumab, colchicine) also decrease CVD risk [2]. Low-grade inflammation can be quantified with acute phase proteins, including *C*-reactive protein (CRP), tumour necrosis factor-alpha (TNF- α), and interleukin-6 (IL-6). Although there are various potential factors that affect low-grade inflammation (e.g., physical activity, smoking, weight loss), diet quality has been show to influence inflammation [3] and relate to the risk of developing CVD [4].

The World Health Organization, among other international organizations, recommend limiting sugars intake, thus sugars remain an important public health focus [5]. There is a particular focus on fructose due to its unique metabolism, and implied contribution towards obesity and the related downstream cardiometabolic implications. Fructose is thought to act as an unregulated substrate for de novo lipogenesis, bypassing negative feedback control, unlike its glucose counterpart. This mechanism is postulated to impair other metabolic signaling and lead to increased adiposity [6,7]. However, the harmful effects of fructose on some cardiometabolic outcomes, including body weight, are only observed when fructosecontaining sugars are consumed as excess energy [8-12]. There is some evidence that a diet high in fructose may increase interstitial inflammation, although this comes from animal models [13]. Observational studies have shown that fructose-containing sugar sweetened beverages (SSB) may be associated with increased pro-inflammatory proteins [14–16]. Other observational studies have found that dietary patterns play an important role in mediating pro-inflammatory biomarkers where foods high in antioxidants, fiber or long chain-polyunsaturated fatty acids, many of which may be a source of fructose-containing sugars, are associated with lower pro-inflammatory biomarker levels [4].

Whether the evidence linking fructose from SSB to inflammation in humans holds for other commonly consumed food sources of fructose-containing sugars which are sources of anti-inflammatory nutrients/constituents, such as fruit, 100% fruit juice, sweetened cereal grains and sweetened dairy and dairy alternatives, at different levels of energy control is unclear. Therefore, we conducted a systematic review and meta-analysis of controlled trials of the effect of different food sources of fructose-containing sugars at different levels of energy control on biomarkers of inflammation with an assessment of the certainty of evidence using Grading of Recommendations, Assessment, Development, and Evaluation (GRADE).

2. Materials and Methods

We followed the Cochrane Handbook for Systematic Reviews of Interventions (version 6.1) [17] for the conduct of our systematic review and meta-analysis and reported our results following the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines (Supplemental Table S1) [18]. The study protocol was registered at ClinicalTrials.gov (NCT02716870).

2.1. Data Sources and Search Strategy

We conducted a systematic search in MEDLINE, Embase, and the Cochrane Central Register of Controlled Studies databases through 6 March 2022. Supplemental Tables S2 and S3 present the search strategy. There were no language restrictions. Validated filters were applied [19]. The searches were supplemented with manual searches.

2.2. Study Selection

We included randomized and non-randomized controlled feeding trials in humans of all health backgrounds and ages, with intervention periods \geq 7 days [20] investigating the effect of orally consumed fructose-containing sugars from various food sources compared with control diets free of or lower in fructose-containing sugars on CRP, TNF- α , and IL-6. We excluded studies of liquid meal replacement interventions and studies of interventions or comparators of rare sugars that contain fructose (e.g., isomaltulose, melezitose, turanose) or were low calorie epimers of fructose (e.g., allulose, tagatose, sorbose). Reports were initially excluded based on review of their titles and abstracts by a single reviewer. Those reports that remained were then excluded based on review of the full text reports by at least two reviewers (X.Q., L.C., D.L., S.A.-C., F.A.Y., A.A., A.C., Q.L.), leaving the final set of reports to be included in our syntheses. We prespecified four study designs based on energy control: (1) 'substitution' trials, in which energy from the food sources of fructose-containing sugars was substituted for other non-fructose-containing macronutrients under energy matched conditions; (2) 'addition' trials, in which excess energy from the food sources of fructose-containing sugars was added to the background diet compared to the same diet alone without the excess energy (with or without the use of non-nutritive/low-calorie sweeteners to match sweetness); (3) 'subtraction' trials, in which energy from the food sources of fructose-containing sugars was subtracted from background diets compared with the original background diets through displacement by water or low-calorie sweeteners or elimination altogether; and (4) 'ad libitum' trials, in which energy from the food sources of fructose-containing sugars was freely replaced (usually within reasonable limits, e.g., intake required to be between 75 and 125% of predicted daily energy requirements) with other non-fructose-containing macronutrients without any strict control of either the study foods or the background diets, allowing for free replacement of energy. In reports containing more than one eligible trial comparison, we included each available trial comparison separately.

2.3. Data Extraction

At least two reviewers independently extracted data from eligible studies. Relevant information included food source of fructose-containing sugars, number of participants, setting, participant health status, study design, level of feeding control, randomization, comparator, fructose-containing sugars type, macronutrient profile of the diets, follow-up duration, energy balance, funding source and outcome data. Supplemental Table S4 shows the definitions for the different food sources of fructose-containing sugars. Authors were contacted for missing outcome data when it was indicated that an inflammatory outcome was measured but not reported. Graphically presented data were extracted from figures using Plot Digitizer [21].

2.4. Risk of Bias Assessment

Included studies were assessed for risk of bias independently and in duplicate by ≥ 2 reviewers using the Cochrane Risk of Bias Tool [17]. Assessment was done across six domains of bias (sequence generation, allocation concealment, blinding, incomplete outcome data, selective outcome reporting and other). Risk of bias for each domain was assessed as either "low" (proper methods taken to reduce bias), "high" (improper methods creating bias) or "unclear" (insufficient information provided). The "other" domain applied only to crossover trials; "high" risk of bias was given when there was no washout between interventions, otherwise the trial was rated as "low". Reviewer discrepancies were resolved by consensus or arbitration by the senior author (J.L.S.).

2.5. Outcomes

The primary outcome was CRP. Secondary outcomes included TNF- α and IL-6. Mean differences (MDs) between the intervention and control arm and their standard errors (SEs) were extracted for each eligible trial comparison. If unavailable, they were derived from available data using published formulas [17]. Mean pairwise difference in change-from-baseline values were preferred over end values. When median data was provided, they were converted to mean data with corresponding variances using methods developed by Luo et al. (2018) [22] and Wan et al. (2014) [23]. When no variance data was available, the standard deviation (SD) was borrowed from a trial similar in size, participants and nature of intervention [24].

2.6. Data Syntheses and Analyses

We used Stata software, version 16.1 (StataCorp, College Station, TX, USA) for all analyses. As our primary research question was to assess the effect of different food sources of fructose-containing sugars at different energy control levels, we performed separate pairwise meta-analyses for each of the four prespecified designs by energy control level (substitution, addition, subtraction and ad libitum trials) and assessed the interaction between food sources of fructose-containing sugars within each energy control level using the Cochrane Handbook's recommended standard Q-test for subgroup differences (significance at p < 0.10) [25–27].

The principal effect measures were the mean pair-wise differences in change-frombaseline (or alternatively, end differences) between the food sources of fructose-containing sugars arm and the comparator arm (significance at p < 0.05). Data were analyzed using the generic inverse variance method with DerSimonian and Laird random-effects model [17,28]. A fixed effects model was used when <5 trial comparisons were available [29]. Paired analyses were applied to all crossover trials with the use of a within-individual correlation coefficient between treatment of 0.5 as described by Elbourne et al. to calculate SEs [30–32]. Data were expressed as MDs with 95% confidence intervals (CIs) for all outcomes. To mitigate a unit-of-analysis error, when arms of trials with multiple intervention or control arms were used more than once, the corresponding sample size was divided by the number of times it was used for calculation of the standard error [17].

Heterogeneity was assessed by visual inspection of the forest plots and using the Cochrane Q statistic and quantified using the I^2 statistic [17]. We considered an $I^2 \ge 50\%$ and $P_Q < 0.10$ as evidence of substantial heterogeneity [17]. Sources of heterogeneity were explored by sensitivity analyses, including individual trial influence, altering pairwise comparison correlation coefficient and subgroup analyses. The influence analysis systematically removed each trial comparison from the meta-analysis with recalculation of the summary effect estimate. A trial whose removal explained the heterogeneity or changed the significance, direction, or magnitude of the effect by more than the minimally important difference (MID) for each outcome (0.5 mg/L for CRP [33-35], 0.28 pg/mL for TNF- α [36], 0.18 pg/mL for IL-6 [37]) was considered an influential trial. To determine whether the overall results were robust to the use of different correlation coefficients in crossover trials, we also conducted sensitivity analyses using correlation coefficients of 0.25 and 0.75. If ≥ 10 trials were available [26,38], we conduced subgroup analyses to explore sources of heterogeneity using meta-regression (significance at $P_O < 0.05$). A priori subgroup analyses were conducted by participant health status, age, anti-inflammatory medication use, baseline outcome level, randomization, energy balance, fructose sugars type, comparator, study design, follow-up, feeding control, fructose-containing sugars dose, sugars regulatory designation, funding and risk of bias. Post hoc subgroup analyses were conducted by type of imputation done for deriving variances (data used assessed change from baseline vs. end differences) and type of CRP analysis (CRP vs. high-sensitivity CRP, for CRP analyses). Meta-regression analyses were used to assess the significance of each subgroup categorically and, when applicable, continuously.

If ≥ 6 trial comparisons were available [39], then we assessed linear and non-linear (restricted cubic splines) dose–response relationships (significance at p < 0.05) using meta-regression. We also assessed non-linear dose–response threshold effects with three prespecified spline knots at important public health thresholds of 5% [5,40], 10% [40,41], and 25% [42] total energy (%E).

If ≥ 10 trials were available, then we assessed publication bias by visual inspection of contour-enhanced funnel plots and formal testing with Egger's [43] and Begg's [44] tests (significance at p < 0.10) [45]. If there was evidence of publication bias, then we adjusted for funnel plot asymmetry and assessed for small-study effects by imputing the missing trial data using the Duval and Tweedie trim-and-fill method [46].

2.7. Certainty of the Evidence

The certainty of the evidence was assessed using the GRADE approach and software (GRADEpro GDT, McMaster University and Evidence Prime Inc., Hamilton, Canada) [47]. The assessments were conducted by two independent reviewers (X.Q., L.C.) and discrepancies were resolved by consensus or arbitration by the senior author (J.L.S.). The evidence was rated as high, moderate, low, or very low certainty. The included controlled trials were initially rated as high certainty by default and then downgraded or upgraded based on pre-specified criteria. Reasons for downgrading the evidence included risk of bias (assessed by the Cochrane Risk of Bias Tool [17]), inconsistency (substantial unexplained inter-study heterogeneity, $I^2 > 50\%$ and $P_O < 0.10$), indirectness (presence of factors that limit the generalizability of the results), imprecision (the 95% CI for effect estimates overlap the MID for benefit or harm), and publication bias (significant evidence of small study effects). The reason for upgrading the evidence was presence of a significant dose–response gradient [48–53]. The importance of the magnitude of the pooled estimates were assessed using our prespecified MIDs and the effect size categories according GRADE guidance [47,54–56] as follows: large effect (\geq 5× MID); moderate effect (\geq 2× MID); small important effect $(\geq 1 \times \text{MID})$; and trivial/unimportant effect (<1 MID).

3. Results

3.1. Search Results

Figure 1 shows the flow of the literature. We retrieved 2850 reports from databases and manual searches, 2698 of which were excluded based on the title or abstract. Of the 152 reports reviewed in full text, 64 reports of controlled feeding trials (91 trial comparisons, n = 4094) met the eligibility criteria [57–120]. These trials included 12 different food sources of fructose-containing sugars (SSB; sweetened dairy; sweetened dairy alternative [soy]; 100% fruit juice; fruit; dried fruit; mixed fruit forms; added nutritive [caloric] sweetener; sweetened cereal grains and bars; sweets and desserts; mixed sources [with SSBs], and mixed sources [without SSBs]) across four energy control levels: substitution (39 trial comparisons); addition (45 trial comparisons); subtraction (4 trial comparisons); and ad libitum (3 trial comparisons). The mixed sources (without SSBs) food category includes those trials in which the intervention included more than one of the food sources, excluding SSBs (e.g., sweets and desserts and fruits).



Figure 1. Flow of literature for the effect of food sources of fructose-containing sugars and inflammation; CRP = *C*-reactive protein; IL-6 = interleukin-6; TNF- α = tumor necrosis factor-alpha.

3.2. Trial Characteristics

Table 1 and Supplemental Table S5 show the trial characteristics. Trial sizes ranged from a median of 15 participants (range 12-120) in subtraction trials to 40 participants (range 12–192) in addition trials. Participants were predominantly adults with and without overweight/obesity, some of whom had a diagnosed chronic condition (e.g., diabetes) or at elevated risk for cardiovascular disease (e.g., dyslipidemia, metabolic syndrome). There were slightly more females in most trial categories with the exception of subtraction trials where there were slightly more males. Most participants were middle-aged adults with ages ranging from a median of 27 (range 26–29) years in subtraction trials to 48 (range 8–72) years in addition trials. Most trials were conducted in an outpatient setting (85–100%), performed in American and European countries, and were parallel in design (62% in substitution and addition, 100% in subtraction, and 67% in ad libitum trials). Feeding control was mostly supplemented for substitution (77%), addition (96%), subtraction (100%), and ad libitum (100%) trials. Most studies were randomized (82–100%). The dose of fructose-containing sugars ranged from a median of 8% (range 1–35%) in addition trials to 19% (range 6–19%) of total energy intake in ad libitum trials. The follow-up duration ranged from a median of 5 weeks in addition trials (range 1-24weeks) to 30 weeks in subtraction trials (range 12–48weeks). The most common source of funding was by agency sources (government, not-for-profit health agency, or university sources) for substitution (41%), addition (45%), and subtraction (50%) trials, with agency and industry sources for ad libitum trials (67%). The comparators for substitution trials were mostly mixed comparator (13/39, 33%) or glucose (12/39, 31%), diet alone for addition trials (31/45, 69%), non-nutritive sweetener for subtraction (3/4, 75%) and mixed for ad libitum trials (2/3, 67%). The main food sources in substitution trials were SSB (10/37, 27%) and mixed sources with (6/37) or without (4/37)SSBs; for addition trials, 100% fruit juice (13/45, 29%), SSB (11/45, 24%) and fruit (9/45, 20%); for subtraction trials, SSB (4/4, 100%); and for ad libitum trials mixed sources (with SSBs) (3/3, 100%).

Trial Characteristics	Substitution Trials	Addition Trials	Subtraction Trials	Ad libitum Trials			
Trials (N)	39	45	4	3			
Participants (median <i>n</i> (range))	38 (21–267)	40 (12–192)	15 (12–120)	40 (29–50)			
Underlying disease status (N trials)	healthy mixed weight = 13, overweight or obese = 11, type 2 diabetes mellitus = 4, metabolic syndrome = 3, other = 8	healthy mixed weight = 17, overweight or obese = 8, type 2 diabetes mellitus = 3, metabolic syndrome = 2, other = 15	healthy mixed weight = 2, overweight or obese = 2	healthy normal weight = 1, overweight or obese = 2			
Age (median years (range)) ^b	46 (14–70)	48 (8–72)	27 (26–29)	38 (32–39)			
Sex ratio (% Male:Female)	36:64	42:58	60:40	38:62			
Randomization (%)	90	82	100	100			
Setting ratio (% N = IP:OP:IP + OP)	0:97:3	0:100:0	0:100:0	0:100:0			
Country (N trials)	USA = 14, Iran = 5, Finland = 4, Brazil = 3, Greece = 3, Switzerland = 3, Sweden = 2, UK = 2, Poland = 2, Netherlands = 1	USA = 10, Denmark = 6, Iran = 5, Spain = 4, Switzerland = 3, Thailand = 3, Brazil = 3, India = 2, Italy = 1, Canada = 2, Mexico = 2, Malaysia = 1, Norway = 1, Israel= 1, UK = 1	USA = 2, Switzerland = 2	Netherlands = 2, UK = 1			
Baseline CRP (median mg/L (range)) ^c	2.2 (0.2–8.1)	1.5 (0.2–55.5)	2.2 (0.9–3.5)	3.0 (1.0–3)			
Baseline TNF-α(median pg/mL (range)) ^d	2.4 (1-6.8)	5.4 (1.2–29.2)	Not reported	Not reported			
Baseline IL-6 (median pg/mL (range)) ^e	2.0 (0.8–27.4)	3.1 (0.6–16.4)	Not reported	Not reported			
Fructose-containing sugars dose (median %E (range))	9 (1–45)	8 (1–35)	15 (15–15)	19 (6–19)			
Study design (%; crossover:parallel)	38:62	38:62	0:100	33:67			
Feeding control (%; met:supp:DA:met,supp:supp,DA)	2.5:77:2.5:18	0:96:2:2	0:100:0:0	0:100:0:0			

Table 1. Summary of trial characteristics ^a.

Table 1. Cont.

Trial Characteristics	Substitution Trials	Addition Trials	Subtraction Trials	Ad libitum Trials 24 (8–24)		
Follow-up duration (median weeks (range))	6 (1–24)	5 (1–24)	30 (12–48)			
Fructose-containing sugars type (N trials)	fructose = 8, sucrose = 6, honey =1, fruit = 14, HFCS = 3, mixed type = 7	fructose = 3, sucrose = 13, honey = 3, fruit = 25, mixed type = 1	sucrose = 2, HFCS = 2	sucrose = 1, mixed type = 2		
Comparator (N trials)	mixed = 13, glucose = 12, starch = 4, fat = 4, lactose = 3, maltodextrin = 2, protein= 1	diet alone= 31, non-nutritive sweetener = 5, other = 5, water = 4	non-nutritive sweetener = 3, water = 1	mixed = 2, non-nutritive sweetener = 1		
Food sources of fructose-containing sugars (N trials)	SSB = 10, sweetened dairy = 3, sweetened dairy alternative (soy) = 1, 100% fruit juice = 2, fruit = 6, dried fruit = 5, mixed fruit forms = 1, added nutritive (caloric) sweeteners = 1, mixed sources (with SSBs) = 6, mixed sources (without SSBs) = 4	SSB = 11, sweetened dairy = 2, 100% fruit juice = 13, fruit = 9, dried fruit = 3, sweetened cereal grains and bars = 1, sweets and desserts = 3, added nutritive sweeteners = 3	SSB = 4	mixed sources = 3		
Funding sources ratio (% n = A:I:A,I:NR)	41:23:33:3	45:11:39:5	50:0:50:0	67:33:0:0		

A = agency, A,I = agency and industry, CRP = C-reactive protein, E = Energy, HFCS = high fructose corn syrup, I = industry, IL-6 = interleukin 6, IP = inpatient, NR = not reported, OP = outpatient, SSB = sugar sweetened beverages, TNF- α = tumour necrosis factor alpha, N = number, UK = United Kingdom, USA = United States of America; ^a Values are rounded to nearest whole number except for baseline outcomes. ^b Based on trials which report data. ^c Based on trial comparisons that reported baseline data (N = 4 trials missing baseline CRP substitution trials and N = 3 trials missing baseline CRP addition trials). ^d Based on trial comparisons that reported baseline data (N = 3 trials missing baseline TNF- α substitution trials; N = 3 trials missing for baseline TNF- α addition trials). ^e Based on trial comparisons that reported baseline data (N = 3 trials missing baseline IL-6 addition trials).

3.3. Risk of Bias

Supplemental Figures S1–S8 show a summary of the risk of bias assessments of the included trials. Across energy designs, 49–69% of trials were assessed as having unclear risk of bias in random sequence generation 50–70% as having unclear allocation concealment domains due to poor reporting, 47–69% were assessed as unclear for incomplete outcome, with 22–53% of trials being assessed as low for blinding (22–53% low) and 53–81% as low for selective outcome reporting. Most cross-over trials were assessed as having low risk of bias in the "other" (carry-over effects) domain (95% in substitution, 89% in addition, 100% in subtraction and ad libitum trials). Fewer studies were assessed as having high risk of bias, for random sequence generation (6–24%), allocation concealment (6–24%), blinding of participants and personnel (0–5%), incomplete outcome data (0%), selective outcome reporting (0–12%), and other (carry-over effects) (0–29%) risk of bias domains. Thus, there was no overall serious risk of bias in most trial comparisons except for in addition trials of sweetened cereal grains and bars for CRP, where there was only one trial that was not randomized, and thus sequence generation and allocation concealment were high risk of bias.

3.4. Primary Outcome

Figure 2 and Supplemental Figures S9–S12 present the effect of different food sources of fructose-containing sugars on the primary outcome, CRP, at four levels of energy control. Total fructose-containing sugars resulted in a reduction in CRP for addition trials (37 trials; MD: -0.18mg/L; 95% CI: -0.33, -0.03mg/L, P_{MD} = 0.020; no substantial heterogeneity, $I^2 = 43.7\%$, P_Q = 0.003) but no effect in substitution (37 trials; MD: 0.07mg/L; 95% CI: -0.08, 0.22mg/L, P_{MD} = 0.336; substantial heterogeneity, $I^2 = 53.7\%$, P_Q < 0.001), subtraction (4 trials; MD: 0.14mg/L; 95% CI: -0.29, 0.56mg/L; P_{MD} = 0.522; no heterogeneity, $I^2 = 0.0\%$, P_Q = 0.877), or ad libitum (3 trials; MD: -0.09mg/L; 95% CI: -0.44, 0.25mg/L; P_{MD} = 0.604; no heterogeneity, $I^2 = 0.0\%$, P_Q = 0.910) trials.

An interaction by food source was detected in the substitution trials (p = 0.010), where sweetened dairy alternative from soy (1 trial; MD: -0.96mg/L; 95% CI: -1.67, -0.25mg/L; P_{MD} = 0.008) and 100% fruit juice (2 trials; MD: -1.09mg/L; 95% CI: -2.01, -0.17mg/L; P_{MD} = 0.021; no heterogeneity, $I^2 = 0.0\%$, P_Q = 0.590) resulted in decreased CRP, while mixed sources (with SSBs) (6 trials; MD: 0.64mg/L; 95% CI: 0.12, 1.17mg/L; P_{MD} = 0.016; substantial heterogeneity, $I^2 = 82.9\%$, P_Q < 0.001) increased CRP. No other food sources showed an effect with variable directions of effect. Although the interaction by food source in addition trials was not significant, we assessed an influence by food source as the reduction in CRP was driven by a sole food source: fruit (9 trials; MD: -0.50mg/L; 95% CI: -0.75, -0.25mg/L; P_{MD} < 0.001; no heterogeneity, $I^2 = 0.0\%$, P_Q = 0.960). There was no overall effect in subtraction or ad libitum trials and although there was a significant influence of food source since there were only 1 food source in each analysis, neither had any effect.

					Summary effect estimates for CRP (mg/L)		Heterog	eneity		GRADE							
Energy design and food source	N trials	n	Weight (%)	MD (mg/L) [95%CI]		P _{MD}	I ²	P _Q	Pinteraction series gases Reises Seri		Up grade e-Lesbouse	Certaint	y of e vide nce ^a	Interpretation of magnitude of effect ^b			
Substitution Trials					I												
SSB	10	281	25.4	0.07 [-0.16, 0.30]	-	0.543	0.0%	0.450	Г			⊕⊕⊕⊕	HIGH	No effect			
Sweetened dairy	3	268	8.0	-0.06 [-0.51, 0.39]		0.802	0.0%	0.630				⊕⊕00	LOW	No effect			
Sweetened dairy alternative (soy)	1	64	3.0	-0.96 [-1.67, -0.25]		0.008	-	-				⊕⊕00	LOW	Small important			
100% Fruit juice	2	150	2.2	-0.90 [-1.07, -0.25]		0.000	0.0%	0.590				⊕⊕ 00	LOW	Moderate			
Fruit	5	210	12.7	-0.43 [-0.87, 0.01]	- 	0.021	34.3%	0.193				$\oplus \oplus \oplus \odot$	MODERATE	No effect			
Dried fruit	4	205	9.4	0.21 [-0.14, 0.55]		0.035	0.0%	0.683				$\oplus \oplus \oplus \odot \bigcirc$	MODERATE	No effect			
Mixed fruit forms	1	50	1.6	-0.10 [-1.20, 1.00]		0.859	- 0.070	- 0.005				$\oplus \oplus \bigcirc \bigcirc$	LOW	No effect			
Added nutritive (caloric) sweeteners	: 1	42	8.9	-0.03 [-0.11, 0.05]		0.464	-	-				€€€O	MODERATE	No effect			
Mixed sources (with SSB)	6	277	23.7	0.64 [0.12, 1.17]	Τ	0.016	82.9%	0.000				$\oplus \oplus \oplus \odot$	MODERATE	Small important			
Mixed sources (without SSB)	4	132	6.3	0.28 [-0.21, 0.78]		0.260	0.00%	0.780				$\oplus \oplus \oplus \odot$	MODERATE	No effect			
All food sources	37	1,679	100.0	0.07 [-0.08, 0.22]	*	0.336	53.7%	0.000				⊕⊕00	LOW *	No effect			
Addition Trials																	
SSB	7	540	35.2	0.02 [-0.15, 0.19]	- - -	0.790	30.9%	0.190				$\oplus \oplus \oplus \oplus$	HIGH	No effect			
Sweetened dairy alternative (soy)	2	48	4.0	0.20 [-0.78, 1.18]	_	0.689	52.0%	0.149				$\oplus \oplus \bigcirc \bigcirc$	LOW	No effect			
100% Fruit juice	12	383	23.3	-0.12 [-0.53, 0.30]		0.580	49.3%	0.030				$\oplus \oplus \oplus \odot$	MODERATE	No effect			
Fruit	9	390	21.4	-0.50 [-0.75, -0.25]	-8-	< 0.001	0.0%	0.960				$\oplus \oplus \oplus \bigcirc$	MODERATE	Small important			
Dried fruit	3	130	3.2	0.02 [-0.75, 0.79]		0.962	0.0%	0.575				⊕⊕00	LOW	No effect			
Sweetened cereal grains & bars	1	50	4.8	-0.30 [-0.79, 0.19]	_	0.228	_	_				0000	VERY LOW	No effect			
Sweets and desserts	2	86	8.1	-0.67 [-1.85, 0.50]		0.260	86.8%	0.006				0000	VERY LOW	No effect			
Added nutritive (caloric) sweetener	1	64	0.1	0.56 [-4.19, 5.31]		- 0.817		0.000	F			$\oplus \oplus \bigcirc \bigcirc$	LOW	No effect			
All food sources	37	1,691		-0.18 [-0.33, -0.03]	◆	0.020	43.7%	0.003				$\oplus \oplus \bigcirc \bigcirc \bigcirc$	LOW *	Trivial			
Subtraction Trials																	
SSB	4	273	100.0	0.14 [-0.29, 0.56]		0.522	0.0%	0.877				$\oplus \oplus \bigcirc \bigcirc \bigcirc$	LOW	No effect			
All food sources	4	273	100.0	0.14 [-0.29, 0.56]	- •	0.522	0.0%	0.877	_ 🗆	2		⊕000	VERY LOW *	No effect			
Ad Libitum Trials											_						
Mixed sources (with SSB)	3	111	100.0	-0.09 [-0.44, 0.25]		0.605	0.0%	0.910				$\oplus \oplus \oplus \bigcirc$	MODERATE	No effect			
All food sources	3	111	100.0	-0.09 [-0.44, 0.25]	- - -	0.604	0.0%	0.910	- 🗆	2		⊕⊕⊙⊖	LOW *	No effect			
					-3.0 -2.0 -1.0 0.0 1.0 2.0	3.0											
					Benefit Harm Mean difference for CRP (mg/L)												

Figure 2. Summary plot for the effect of different food sources of fructose-containing sugars on CRP; Data are weighted mean differences (95% confidence intervals) for summary effects of individual food sources and total food sources on CRP. Analyses conducted by generic, inverse variance random effects models (at least five trials available) or fixed effects models (fewer than five trials available). Between-study heterogeneity was assessed by the Cochrane Q statistic, where $P_Q < 0.100$ is considered statistically significant, and quantified by the I^2 statistic, where $I^2 \ge 50\%$ is considered evidence of substantial heterogeneity. The effects of total

fructose-containing sugars are denoted by the bolded lines with the effect estimates as diamonds. The effects of individual food sources are denoted by the non-bolded lines with the effect estimates as squares. Any statistically significant reductions are highlighted in green and significant increases in red. The Grading of Recommendations, Assessment, Development and Evaluation (GRADE) of randomized controlled trials are rated as "High" certainty of evidence and can be downgraded by five domains and upgraded by one domain. The white squares represent no downgrades, while filled black squares indicate a single downgrade or upgrades for each outcome, and the black square with a white "2" indicates a double downgrade for each outcome. CI = confidence interval; GRADE = Grading of Recommendations, Assessment, Development and Evaluation; MD= mean difference; N = number; SSB = sugar-sweetened beverages; ^a Since all included trials were randomized or non-randomized controlled trials, the certainty of the evidence was graded as high for all outcomes by default and then downgraded or upgraded based on pre-specified criteria. Criteria for downgrades included risk of bias (ROB) (downgraded if the majority of trials were considered to be at high ROB); inconsistency (downgraded if there was substantial unexplained heterogeneity $[I^2 > 50\%, P_{\Omega} < 0.10]$; indirectness (downgraded if there were factors absent or present relating to the participants, interventions, or outcomes that limited the generalizability of the results); imprecision (downgraded if the 95% confidence interval crossed the minimally important difference [MID] for harm or benefit set 0.5 mg/L for CRP [33–35]; and publication bias (downgraded if there is evidence of publication bias based on funnel plot asymmetry and/or significant Egger's or Begg's tests (p < 0.10) with confirmation by adjustment by Duval and Tweedie trim-and-fill analysis). Criteria for upgrades included a significant dose-response gradient; ^b For the interpretation of the magnitude, we used the MIDs (see a above) to assess the importance of magnitude of our point estimate using the effect size categories according to new GRADE guidance. We then used the MIDs to assess the importance of the magnitude of our point estimates using the effect size categories according GRADE guidance [47,54,56] as follows: large effect ($\geq 5 \times$ MID); moderate effect ($\geq 2 \times$ MID); small important effect ($\geq 1 \times$ MID); and trivial/unimportant effect (<1 MID); * Where there was a significant interaction by food source in substitution trials, an influence of fruit in addition trials, and SSBs and/or mixed sources (with SSBs) were the sole food sources in subtraction and ad libitum trials, we performed the GRADE analysis for each individual food source.

3.5. Secondary Outcomes

Figures 3 and 4 and Supplemental Figures S13–S16 present the effect of different food sources of fructose-containing sugars on our secondary outcomes, TNF- α and IL-6, at four levels of energy control. In substitution trials, there was no overall effect on either outcome, with no significant interaction by food source (p > 0.05). In addition trials, there was no overall effect on either outcome. However, there was a significant interaction by food source for IL-6 (p = 0.020) where sweets and desserts coming from dark chocolate (1 trial; MD: -8.79pg/mL; 95% CI: -14.26, -3.32pg/mL; P_{MD} = 0.002) resulted in a decrease in IL-6. An influence by food source was determined for TNF- α in addition trials since there was a significant reduction for fruit (3 trials; MD: -0.89pg/mL; 95% CI: -1.58, -0.20pg/mL; P_{MD} = 0.012; no substantial heterogeneity, $I^2 = 14.3\%$, P_Q = 0.311), similar to what was observed for CRP.

3.6. Sensitivity and Subgroup Analyses

Supplemental Figures S17–S20 present the influence analyses for the effect of total fructose-containing sugars at the 4 levels of energy control on the primary outcome, CRP. Removal of single trial comparisons provided a partial explanation of the evidence of substantial heterogeneity [81,89] in substitution trials and resulted in a loss of significance for CRP [80] in addition trials.

Supplemental Figures S21–S33 present the influence analyses for the effect of individual food sources, for those analyses that showed evidence of an interaction or influence by food source, on the primary outcome, CRP. Removal of single trial comparisons resulted in loss of significance for the increase in CRP with mixed sources (with SSBs) in substitution trials [81,89] and for the decrease in CRP with 100% fruit juice in substitution trials [109]; a gain of significance for a reduction in CRP with fruit in substitution trials [94] and with sweets and desserts in addition trials [59]; and a partial explanation of heterogeneity for mixed sources (with SSBs) [114] in substitution trials.

Supplemental Table S5 shows sensitivity analyses for the different correlation coefficients (0.25 and 0.75) used in paired analyses of crossover trials for CRP. The use of these different correlation coefficients did not alter the direction, magnitude, or significance of the effect or evidence for heterogeneity with the following exception: a gain of significance for a reduction in CRP with fruit (MD: -0.43mg/L; 95% CI: -0.85, -0.01mg/L; P_{MD} = 0.045) in substitution trials with the use of 0.75.

Supplemental Figures S34–S47 present the sensitivity analyses for the secondary outcomes. For total fructose-containing sugars, removal of single trial comparisons resulted in a gain of significance for a reduction in TNF- α in addition trials [87,111] and partial explanation of heterogeneity for TNF- α in substitution [84,116] and addition [92] trials. For individual food sources for those analyses that showed evidence of an interaction or influence by food source for secondary outcomes, removal of single trial comparison resulted in: a loss of significance for the reduction in TNF- α with fruit [88] in addition trials; and a gain of significance for a reduction in TNF- α with sweets and desserts (98) and in IL-6 with sweetened dairy [71] and 100% fruit juice [113] in addition trials; and a partial explanation of heterogeneity for SSBs [111] and sweets and desserts [80] on TNF- α and 100% fruit juice [113] on IL-6 in addition trials.

Supplemental Table S6 shows sensitivity analyses for the different correlation coefficients (0.25 and 0.75) used in paired analyses of crossover trials for secondary outcomes. The use of these different correlation coefficients did not alter the direction, magnitude, or significance of the effect or evidence for heterogeneity for any outcomes across food sources and levels of energy control, with the following exception: gain of significance for a reduction in TNF- α with total fructose-containing sugars with the use of 0.25.

					Summary effect estimates for	·TNF-α (pg/mL)		Heterog	eneity			GRAI	ЭE	
Energy design and food source	N trials	n	Weight (%)	MD (pg/mL) [95%C1]			P _{MD}	I ²	P _Q	Downgrade Pinteraction Reint of the set of	Publication bias Dose-response	_	y of evidence ⁸	Interpretation of magnitude of effect ^b
Substitution Trials														
SSB	2	64	28.9	-0.03 [-0.18, 0.13]		+	0.720	45.7%	0.175					
Fruit	3	159	15.1	-0.15 [-0.39, 0.08]		-8-	0.206	0.0%	0.455					
Dried fruit	5	196	23.5	0.03 [-0.49, 0.55]			0.920	65.5%	0.021					
Mixed fruit forms	1	50	5.4	0.20 [-0.34, 0.74]			0.467	-	-					
Mixed sources (with SSB)	4	204	7.4	0.35 [-0.69, 1.39]			0.511	69.2%	0.020					
Mixed sources (without SSB)	2	69	19.9	0.15 [-0.01, 0.31]			0.057	0.00%	0.940					N. 00
Total food sources	17	742	100.0	0.04 [-0.11, 0.19]		•	0.610	52.6%	0.006	0.830		$\oplus \oplus \oplus \oplus$	HIGH	No effect
Addition Trials														
SSB	3	82	15.3	0.79 [-1.23, 2.80]	—		0.444	58.6%	0.089			⊕⊕⊙⊙	LOW	No effect
Sweetened Dairy	2	44	21.8	-0.10 [-0.30, 0.10]		-8-	0.336	0.0%	1.000			$\oplus \oplus \bigcirc \bigcirc$	LOW	No effect
100% Fruit juice	3	116	28.6	-0.66 [-2.66, 1.35]		■	0.507	95.6%	0.000			$\oplus \oplus \bigcirc \bigcirc$	LOW	No effect
Fruit	3	65	20.6	-0.89 [-1.58, -0.20]			0.012	14.3%	0.311			$\oplus \oplus \oplus \odot$	MODERATE	Small important
Sweets and desserts	2	65		-4.66 [-16.21, 6.90]			0.439	77.3%	0.040			€000	VERY LOW	No effect
Added nutritive (caloric) sweeteners		77		-0.91 [-2.17, 0.36]			0.162	2.00%	0.361			⊕⊕00	LOW	No effect
Total food sources	16	449	100.0	-0.48 [-0.99, 0.04]	-	•	0.069	86.8% ·	<0.001	0.430		⊕000	VERY LOW *	No effect
					-5.0 -4.0 -3.0 -2.0 -1.0	0.0 1.0 2.0)							
					Benefit Mean difference for T	Harm								

Figure 3. Summary plot for the effect of different food sources of fructose-containing sugars on TNF- α ; Data are weighted mean differences (95% confidence intervals) for summary effects of individual food sources and total food sources on TNF- α . Analyses conducted by generic, inverse variance random effects models (at least five trials available) or fixed effects models (fewer than five trials available). Between-study heterogeneity was assessed by the Cochrane Q statistic, where $P_Q < 0.100$ is considered statistically significant, and quantified by the l^2 statistic, where $l^2 \ge 50\%$ is considered evidence of substantial heterogeneity. The effects of total fructose-containing sugars are denoted by the bolded lines with the effect estimates as diamonds. The effects of individual food sources are denoted by the non-bolded lines with the effect estimates as squares. Any statistically significant reductions are highlighted in green and significant increases in red. The Grading of Recommendations, Assessment, Development and Evaluation (GRADE) of randomized controlled trials are rated as "High" certainty of evidence and can be downgraded by five domains and upgraded by one domain. The white squares represent no downgrades, while filled black squares indicate a single downgrade or upgrades for each outcome, and the black square with a white "2" indicates a double downgrade for each outcome. CI = confidence interval; GRADE = Grading of Recommendations, Assessment, Development and Evaluation; MD= mean difference; N = number; SSB = sugar-sweetened beverages; TNF- α = tumour necrosis factor-alpha; ^a Since all included trials were randomized or non-randomized controlled trials, the certainty of the evidence was graded as high for all outcomes by default and then downgraded or upgraded based on pre-specified criteria. Criteria for downgrades included risk of bias (ROB) (downgraded if the majority of trials were considered to be at high ROB); inconsistency (downgraded if there was substantial unexplained het

(downgraded if the 95% confidence interval crossed the minimally important difference [MID] for harm or benefit set at 0.28 pg/mL for TNF- α [36]; and publication bias (downgraded if there is evidence of publication bias based on funnel plot asymmetry and/or significant Egger's or Begg's tests (p < 0.10) with confirmation by adjustment by Duval and Tweedie trim-and-fill analysis). Criteria for upgrades included a significant dose–response gradient; ^b For the interpretation of the magnitude, we used the MIDs (see a above) to assess the importance of magnitude of our point estimate using the effect size categories according to new GRADE guidance. We then used the MIDs to assess the importance of the magnitude of our point estimates using the effect size categories according GRADE guidance [47,54,56] as follows: large effect ($\geq 5 \times$ MID); moderate effect ($\geq 2 \times$ MID); small important effect ($\geq 1 \times$ MID); and trivial/unimportant effect (<1 MID); * Where there was an influence of fruit in addition trials, we performed the GRADE analysis for each individual food source.

					Summary effect	t estimates t	for IL-6 (pg	/mL)		Heterog	geneity				GRAI	DE	
Energy design and food source	N trials	n	Weight (%)	MD (pg/mL) [95%C1]					P _{MD}	I ²		Pinteraction	Risk of bias Inconsistency Indirectness Imprecision Publication bias	Up grade estication or state of the state of	Certaint	y of evidence	a Interpretation of magnitude of effect ^b
Substitution Trials																	
SSB	5	143	18.7	-0.10 [-0.49, 0.30]					0.634	0.0%	0.798						
Fruit	4	183	19.2	0.16 [-0.83, 1.16]				-	0.748	46.1%	0.135						
Dried fruit	3	172	25.8	0.30 [-0.73, 1.34]				_	0.566	71.9%	0.028						
Mixed sources (with SSBs)	1	105	17.8	0.02 [-0.30, 0.34]			+		0.903	-	-						
Mixed sources (without SSB)	3	102	18.6	0.18 [-0.20, 0.57]			₽		0.350	0.0%	0.660						
Total food sources	16	705	100.0	-0.04 [-0.24, 0.15]			•		0.663	22.2%	0.202	0.930			⊕⊕⊕C	MODERATE	No effect
Addition Trials																	
SSB	5	253	31.8	0.06 [-0.34, 0.46]			-		0.776	29.0%	0.229				⊕⊕⊕С	MODERATE	No effect
Sweetened dairy	2	44	21.3	-0.52 [-1.12, 0.08]			-8-		0.087	46.6%	0.171				⊕⊕OC		No effect
100% Fruit juice	4	149	5.4	-3.01 [-6.91, 0.88]					0.130	74.1%	0.001				$\oplus \oplus \oplus \bigcirc \bigcirc$	MODERATE	No effect
Fruit	3	102	31.5	-0.19 [-0.59, 0.21]			-#-		0.352	86.5%	0.001				⊕⊕OC	LOW	No effect
Sweet and desserts (dark chocolate)	1	44	0.3	-8.79 [-14.26, -3.32]					0.002	-	-				$\oplus \oplus \oplus \bigcirc \bigcirc$		Large
Added nutritive (caloric) sweetener	1	64	9.8	0.40 [-0.22, 1.02]			-8-		0.207	-	-				€⊕OC		No effect
Total food sources	16	656	100.0	-0.15 [-0.45, 0.16]			•		0.352	82.9%	0.000	0.020	2		⊕00C	VERY LOW	No effect
						-6.0 -4.0 enefit fference for		Harm	4.0								

Figure 4. Summary plot for the effect of different food sources of fructose-containing sugars on IL-6; Data are weighted mean differences (95% confidence intervals) for summary effects of individual food sources and total food sources on IL-6. Analyses conducted by generic, inverse variance random effects models (at least five trials available) or fixed effects models (fewer than five trials available). Between-study heterogeneity was assessed by the Cochrane Q statistic, where $P_Q < 0.100$ is considered statistically significant, and quantified by the I^2 statistic, where $I^2 \ge 50\%$ is considered evidence of substantial heterogeneity. The effects of total fructose-containing sugars are denoted by the bolded lines with the effect estimates as diamonds. The effects of individual food sources are denoted by the non-bolded lines with the effect estimates as squares. Any statistically significant reductions are highlighted in green and significant increases in red. The Grading of

Recommendations, Assessment, Development and Evaluation (GRADE) of randomized controlled trials are rated as "High" certainty of evidence and can be downgraded by five domains and upgraded by one domain. The white squares represent no downgrades, while filled black squares indicate a single downgrade or upgrades for each outcome, and the black square with a white "2" indicates a double downgrade for each outcome. CI = confidence interval; GRADE = Grading of Recommendations, Assessment, Development and Evaluation; MD = mean difference; N = number; SSB = sugar-sweetened beverages; ^a Since all included trials were randomized or non-randomized controlled trials, the certainty of the evidence was graded as high for all outcomes by default and then downgraded or upgraded based on pre-specified criteria. Criteria for downgrades included risk of bias (ROB) (downgraded if the majority of trials were considered to be at high ROB); inconsistency (downgraded if there was substantial unexplained heterogeneity $|I^2 \ge 50\%$, $P_Q < 0.10]$; indirectness (downgraded if there were factors absent or present relating to the participants, interventions, or outcomes that limited the generalizability of the results); imprecision (downgraded if there is evidence of publication bias based on funnel plot asymmetry and/or significant Egger's or Begg's tests (p < 0.10) with confirmation by adjustment by Duval and Tweedie trim-and-fill analysis). Criteria for upgrades included a significant tage response gradient; ^b For the interpretation of the magnitude, we used the MIDs (see a above) to assess the importance of magnitude of our point estimate using the effect size categories according to new GRADE guidance. We then used the MIDs to assess the importance of the magnitude of our point estimate using the effect size categories according GRADE guidance [47,54,56] as follows: large effect ($\geq 5 \times$ MID); moderate effect ($\geq 2 \times$ MID); small important effect ($\geq 1 \times$ MID); and trivial/unimportant effect (<1 MID); * Where there was a sig

Supplemental Figures S48–S53 present the subgroup analyses and continuous metaregression analyses for the effect of total fructose-containing sugars, where there were at least 10 trial comparisons, on the primary outcome, CRP. There was significant effect modification by health status (trials of participants with other chronic conditions, such as chronic kidney disease, non-alcoholic fatty liver disease and irritable bowel syndrome, showed increases while trials with other participant types showed no effect), fructosecontaining sugars type (trials providing mixed type showed increases while those with fruit showed a tendency for reductions, and others showed no effect in substitution trials), randomization (trials without randomization showed increases while those randomized showed no effect in substitution trials), energy balance (trials with neutral energy balance showed increases while those with positive or negative showed a tendency for reductions in substitution trials), feeding control (metabolic and metabolic with supplementation showed reductions while others showed no effect in substitution and addition trials), other risk of bias (trials with high risk of bias showed increases while those with low showed no effect in substitution trials), and baseline CRP (trials above the median baseline CRP showed reductions while those below the median baseline CRP showed no effect in addition trials).

Supplemental Figures S54–S59 present the subgroup analyses and continuous meta regression analyses for the effect of individual food sources of fructose-containing sugars on the primary outcome, CRP. There was significant effect modification by baseline CRP (trials with baseline CRP greater than the median showed a tendency for increases for SSB in substitution trials, however they showed a tendency for reductions for 100% fruit juice in addition trials), follow up (trials with greater than 8-weeks duration showed a tendency for reductions while those \geq 8-weeks showed no effect for 100% fruit juice in addition trials), and selective outcome reporting (low risk of bias trials showed a tendency for reductions for 100% fruit juice in addition trials while unclear risk of bias trials showed no effect).

Supplemental Figures S60–S71 present the subgroup analyses and continuous meta regression analyses for the effect of total fructose-containing sugars, where there were at least 10 trial comparisons, on secondary outcomes. There was significant effect modification involving both TNF- α and IL-6 by incomplete outcome (trials with low risk of bias showed tendency for reductions for TNF- α , yet increases for IL-6) in substitution trials, and randomization (trials without randomization tended to show reductions, while those randomized showed no effect) and design (crossover trials showed reductions while parallel trials showed no effect) in addition trials. A few of other subgroup analyses showed subgroup differences for individual outcomes across levels of energy control, without any discernable pattern. There were no subgroup analyses for the effect of individual food sources on secondary outcomes as there was no interaction or influence by food source or there were <10 trial comparisons available.

3.7. Dose Response Analyses

Supplemental Figures S72–S81 present linear and non-linear dose–response analyses for the primary outcome, CRP. In substitution trials, there was no dose response for the effect of total fructose-containing sugars nor for any food source with ≥ 6 trial comparisons. In addition trials, there was a non-linear dose response for total fructose-containing sugars ($P_{non-linear} < 0.001$) and dose threshold relationships at 5% (p = 0.001) and 10% (p = 0.002). There was also a dose threshold relationship at 5% for 100% fruit juice (p = 0.043). In subtraction and ad libitum trials, there were too few trials to assess dose responses for total fructose-containing sugars or any food source.

Supplemental Figures S82–S89 present linear and non-linear dose–response analyses for secondary outcomes. There was a dose threshold relationship at 5% for TNF- α in addition trials (p = 0.002) where greater reductions are seen with lower doses. There was also a dose threshold relationship at 5% for IL-6 in substitution trials (p = 0.025), where the increase at low doses was driven by only 1 study (2 trial comparisons) [94]. Although this study found increases in IL-6, they found reductions for TNF- α . There were too few trials with dose data to assess dose responses for secondary outcomes by food source.

3.8. Publication Bias

Supplemental Figures S90–S99 present the publication bias and trim-and-fill (where applicable) assessments for all outcomes where there were ≥ 10 trials available. There was no evidence of funnel plot asymmetry in any analysis of the primary outcome, CRP. There was evidence of funnel plot asymmetry for the effect of total fructose-containing sugars on IL-6 in substitution (Egger's test, p = 0.004) and addition (Egger's test, p = 0.015) trials. Adjustment for funnel plot asymmetry with the imputation of 4 missing trials by the Duval and Tweedie trim-and-fill method, however, did not alter the direction, magnitude or significance of the effect, suggesting that there was no meaningful influence of publication bias on the results (Original MD in substitution trials: -0.04pg/mL; 95% CI: -0.24 to 0.15, p = 0.664; imputed MD: -0.05; 95% CI: -0.28 to 0.18, p = 0.677; Original MD in addition trials: -0.15pg/mL; 95% CI: -0.45 to 0.16, p = 0.349; imputed MD: -0.06pg/mL; 95% CI: -0.39 to 0.27, p = 0.718).

3.9. GRADE Assessment

Figures 2–4 and Supplemental Tables S7 and S8 present the GRADE assessments. The certainty of evidence for the effect of total fructose-containing sugars on the primary outcome, CRP, was low in substitution (no effect), addition (trivial reduction), and ad libitum (no effect) trials and very low for subtraction trials (no effect), owing to double downgrades for indirectness across the 4 levels of energy control and a single downgrade for imprecision in subtraction trials.

As there was evidence of significant interaction or influence by food source, the certainty of evidence was assessed for the individual food sources. The certainty of evidence was low for sweetened dairy alternative (soy) (small important reduction) and 100% fruit juice (moderate reduction), and moderate for mixed sources (with SSBs) (small important increase) in substitution trials owing to downgrades for indirectness and/or imprecision; and moderate for fruit (small important reduction) in addition trials owing to a downgrade for imprecision. The certainty of evidence for the remaining food sources which showed no effect, was generally moderate, ranging from high to very low, owing to downgrades for risk of bias, inconsistency, indirectness, and/or imprecision.

The certainty of evidence for the effect of total fructose-containing sugars on secondary outcomes was high for TNF- α and moderate for IL-6 in substitution trials, due to a downgrade for imprecision, and very low for both outcomes in addition trials owing to double downgrades for indirectness and at least one single downgrade for inconsistency and/or imprecision.

As there was evidence of influence by food source in addition trials for TNF- α and IL-6, the certainty of evidence was assessed for individual food sources. The certainty of evidence was moderate for the effect of fruit on TNF- α (small important reduction) and sweets and desserts (dark chocolate) on IL-6 (large reductions) owing to a downgrade for imprecision and indirectness, respectively. The certainty of evidence for the remaining food sources which showed no effect, was moderate or low, owing to downgrades for inconsistency, indirectness, and/or imprecision.

4. Discussion

We conducted a systematic review and meta-analysis of 64 reports (91 trial comparisons) in 4094 generally healthy participants with or without obesity, with few trials of participants who have or are at risk for cardiometabolic diseases, of the effects of 12 different food sources of fructose-containing sugars (SSB; sweetened dairy; sweetened dairy alternative [soy]; 100% fruit juice; fruit; dried fruit; mixed fruit forms; sweetened cereal grains and bars; sweets and desserts; added nutritive [caloric] sweetener; mixed sources [with SSBs]; and mixed sources [without SSBs]) with a median dose of 8% to 19% of total energy across four different levels of energy control over median follow-up of 5–30 weeks. Total fructose-containing sugars led to a trivial reduction in CRP (-0.18 mg/L) in addition trials. There was no effect of total fructose-containing sugars at the other levels of energy control or on secondary outcomes. There was evidence of interaction or influence by food source in most analyses. In substitution trials, sweetened dairy alternatives as a soy beverage at a dose of 1%E (5 g sugar) led to small important reductions in CRP (-0.96 mg/L) and 100% fruit juice at doses of 8.8%E and 12%E led to a moderate reduction in CRP (-1.09 mg/L), while mixed sources (with SSBs) at a median dose of 6.4%E (ranging from 6.3%E to 27%E) led to a small important increase in CRP (0.64 mg/L). In addition trials, fruit at a median dose of 3.8%E (ranging from 1.6%E to 10%E) led to a small important reduction in CRP (-0.50 mg/L) and TNF- α (-0.89 pg/mL), while sweets and desserts as dark chocolate at a dose of 1.1%E led to a large reduction in IL-6 (-8.79 pg/mL). Other food sources of fructose-containing sugars showed no effect on markers of inflammation.

4.1. Findings in Relation to the Literature

Our results for total fructose-containing sugars are similar to a previous systematic review and meta-analysis of the effects of fructose-containing sugars on CRP which included 6 controlled trials (n = 403) and found no significant difference between fructose and glucose interventions (MD: -0.03 mg/L; 95% CI: -0.52, 0.46 mg/L; $I^2 = 44\%$) [121]. The present analyses build on the previous study, as it identified many more reports, including on additional inflammatory biomarkers (TNF- α , IL-6), prespecified 4 energy designs in order to separate the effect of energy control, and explored the interaction between food sources of fructose-containing sugars.

The benefits or lack of harm observed for certain food sources of fructose-containing sugars is in agreement with previous observations. The reduction in CRP and TNF- α observed in addition trials for fruit at a median intake of 3.8%E (range of 1.6%E to 10%E), where the predominant type was berries, is supported by a systematic review and metaanalysis of controlled trials which showed similar reductions in inflammation (TNF- α , MD: -0.99 pg/mL; 95% CI: -1.96, -0.02 pg/mL; p = 0.04), as well as reductions in adiposity, glycemic control, blood lipids, and blood pressure [122]. Inflammatory benefits have also been observed in a recent systematic review and meta-analysis of fruit and vegetable intake where 10 observational studies found an inverse association between intakes of fruit or vegetables and inflammatory biomarkers and 71 clinical trials showed significant reductions in both CRP (23 trial comparisons, MD: -0.34 mg/L; 95% CI: -0.58, -0.11 mg/L; p < 0.01) and TNF- α (16 trial comparisons, MD: -0.87 pg/mL; 95% CI: -1.59, -0.15 pg/mL; p = 0.02) [123]. Improvements in cardiovascular risk factors were demonstrated for fruit in our systematic reviews and meta-analyses of the effect of food sources of fructose-containing sugars on adiposity, blood pressure, and glycemic control [11,124,125].

The reduction in CRP by 100% fruit juice, specifically 100% orange juice, at doses of 9%E and 12%E, in substitution trials is in agreement with improvements in inflammatory markers observed in a systematic reviews and meta-analyses of clinical trials on 100% orange juice (IL-6: 5 trial comparisons, MD: -1.51 pg/mL; 95% CI: -2.31, -0.70 pg/mL; hs-CRP; 9 trial comparisons, MD: -0.58 mg/L; 95% CI: -1.22, 0.05 mg/L) [126]. This result is also supported by systematic reviews and meta-analyses of prospective cohort studies which have demonstrated U-shaped associations between 100% fruit juice intake and cardiometabolic outcomes such as incident hypertension [127], metabolic syndrome [128], and cardiovascular event risk [129] and our recent systematic review and meta-analysis of fructose-containing sugars showing improvements in markers of adiposity at doses $\leq 10\%$ E [124]. Evidence in the literature generally shows improvement in risk factors at low to moderate doses of 100% fruit juice [130].

The reduction in CRP in the substitution trial of a sweetened soy beverage, which included participants with non-alcoholic fatty liver disease, is supported by systematic reviews and meta-analyses demonstrating improvements in inflammatory biomarkers from the consumption of soy [131,132], including one specifically showing benefit of soy protein [133]. A subgroup analysis in one of these studies demonstrated the reduction was stronger in participants who were affected various chronic diseases [131]. The sweetened soy beverage reduction in CRP observed in the present study is also reflected in the CRP

reduction observed with the dietary portfolio, a cholesterol-lowering dietary pattern that involves a relatively high soy milk consumption [134,135]. Therefore, the CRP reduction in sweetened soy beverage may be generalizable to those at higher risk of or with a chronic disease.

The reduction in IL-6 observed in the one addition trial of dark chocolate, which provided 1.1%E (5 g sugar/day) as 84% dark chocolate, is supported by systematic reviews and meta-analyses showing associations between chocolate intake and lower risk of CVD incidence and mortality [136].

In substitution trials with mixed sources with SSBs there was an increase in CRP. For these trials, in the comparator arm, there was a specific focus on restricting SSBs and added sugars in the diet, replacing them predominantly with starch. It is possible that this resulted in an increase in whole grain and dietary fibre intake on the comparator, both of which have been demonstrated to reduce inflammatory markers in systematic reviews and meta-analyses of controlled trials, particularly in participants with chronic diseases [137,138], which was the participant type in all of the included trials. The lack of harm observed for SSBs alone at all levels of energy control is supported by a previous systematic review and meta-analysis including 7 trial comparisons of fructose versus glucose which showed no overall effect [121]. Previous systematic reviews and metaanalyses exploring the effect of different food sources of fructose containing sugars on cardiometabolic outcomes in controlled trials have demonstrated harm when SSBs are consumed as a source of excess calories, including on glycemic control, adiposity, blood pressure and uric acid [8,11,139,140]. It is possible that the lack of harm observed in the present analysis may be the result of fewer trials and those which only comprised of healthy participants free of chronic diseases and with low baseline CRP levels (median 0.4 mg/L, range 0.2–1.22 mg/L).

Baseline CRP level may be an important consideration since it was the only factor that was significant in subgroup analyses for the effect of SSB on CRP in substitution trials and of 100% fruit juice on CRP in addition trials; the only 2 food sources in all energy designs where there were ≥ 10 trial comparisons allowing for subgroup analyses to be performed. In categorical subgroup analyses of substitution trials, SSB showed a tendency to increase CRP when baseline CRP was greater than the median. There was also a positive continuous relationship where SSB showed a greater effect on CRP with greater baseline CRP. Conversely, 100% fruit juice tended to reduce CRP to a greater extent when baseline CRP was greater than the median. This effect is supported by the significant reduction in CRP found with 100% fruit juice in substitution trials in which the 2 trials included had higher baseline CRP levels. Thus, the potential effects of difference food sources of fructose-containing sugars may be more prominent in populations with higher baseline inflammation.

4.2. Potential Mechanisms

These advantages seen for some foods may be partly explained by the food's content of antioxidants, flavonoids, and/or polyphenols. Fruit, especially berries and apples, which were the predominant source in the included trials, as well as oranges in 100% orange juice, are rich sources of antioxidants, while soy milk is a source of isoflavones and dark chocolate is a rich source of flavonoids, all of which have evidence to support an explanation for cardiovascular improvements [126,141,142]. Conversely, some of the food sources of fructose-containing sugars which showed no effect (e.g., SSBs, sweetened dairy, sweets and desserts, sweetened cereal grains and bars, added nutritive sweeteners) would be expected to be lower in or devoid of these bioactives. Dried fruit would be expected to have similar level of bioactives as fruit, as indicated in the few included trials of dried fruit in the present analysis, however they showed no overall effect. Our similar systematic review and meta-analysis on markers of adiposity included more controlled trials and showed improvements in body weight and BMI for dried fruit [124], thus it remains uncertain whether additional trial data may affect the conclusion on inflammation. Bioactives, including antioxidants and flavonoids, may also interfere with fructose metabolism. For example, antioxidants and flavonoids may reduce oxidative stress and thus fructose-induced uric acid production [143], which is supported by the lack of harm observed for fruit and fruit juice in a systematic review and meta-analysis of food sources of fructose-containing sugars on uric acid which contrasts the significant increases in uric acid that were observed for SSBs [8]. In addition to bioactives as mechanisms through which various foods may influence inflammation, these food sources of fructose-containing sugars can be higher in dietary fibre and lower in glycemic index (GI). Fruit which showed reductions in markers of inflammation have higher fibre (e.g., apples 4 g/medium, berries 4 g/cup,) and lower GI (e.g., apples 38, berries 28) [144], and soy beverages, orange juice and dark chocolate are low GI foods [144], whereas the SSBs, added nutritive sweeteners, and mixed sources (with SSBs) would be expected to be lower in fibre and higher in GI. Low GI diets may improve inflammation as demonstrated in a recent systematic review and meta-analysis of low glycemic index/load diets which showed similar reductions in CRP resulting from low GI compared to higher GI diets [145]. Circulating insulin and related incretin hormones may be reduced, increasing satiety and decreasing subsequent energy intake with the consumption of lower GI and higher fibre foods [146–150].

4.3. Strengths and Limitations

Our systematic review and meta-analysis has several strengths. First, we conducted a comprehensive and reproducible search and selection process of the literature examining the effect of food sources of fructose-containing sugars on markers of inflammation. Second, we collated and synthesized the totality of available evidence from a large body (64 reports, 89 trial comparisons, n = 3958) of controlled intervention studies, which give the greatest protection against bias. Third, we had comprehensive exploration of possible sources of heterogeneity. Fourth, we evaluated the shape and strength of the dose–response relationships. Fifth, we assessed the overall quality of evidence using the GRADE assessment approach.

Our analyses also presented limitations. First, there was evidence for serious risk of bias in one analysis of sweetened cereal grains and bars on CRP in addition trials due to the lack of randomization of the one trial resulting in high risk of bias for sequence generation and allocation concealment. Second, there was evidence of indirectness. The significant interaction or influence of food source in substitution trials for CRP and addition trials for all outcomes and the limited number of food sources of fructose-containing sugars available in the subtraction and ad libitum trials for CRP (only one or two food sources available [SSB and/or mixed sources with SSBs]) in the pooled analyses for total fructose-containing sugars meant the results could not be generalized to all food sources. We therefore double downgraded for very serious indirectness in these analyses and rated the evidence separately for individual food sources. The downgrades for indirectness of individual food sources were related to insufficient trial comparisons which limited generalizability related to population, intervention or comparator. The absence of longterm trials (>1-year diet duration), might be another reason to downgrade for serious indirectness, however we concluded based on short term intake. Third, there was evidence of inconsistency in a few of the pooled estimates. In the addition trial analyses for IL-6 of total fructose-containing sugars, for CRP of sweets and desserts, and for TNF- α of 100% fruit juice, we downgraded for serious inconsistency due to substantial unexplained heterogeneity. Finally, there was evidence of imprecision in almost all of the pooled analyses. We downgraded for serious imprecision due to the crossing of the prespecified MID, which meant that clinically important benefit and/or harm could not be ruled out.

Weighing the strengths and limitations, the certainty of evidence was low for the decreasing effect of sweetened dairy alternative as soy and 100% fruit juice and moderate for the increasing effect of mixed sources (with SSBs) on CRP in substitution trials, moderate for the decreasing effect of fruit on CRP and TNF- α and sweets and desserts as dark chocolate on IL-6 in addition trials, and generally low (very low to high) for the effect of all other comparisons on markers of inflammation.

4.4. Implications

Our findings, similar to our previous systematic reviews and meta-analyses on the importance of food sources of fructose-containing sugars on other cardiometabolic outcomes [8,11,124,125,139], suggest that the focus of dietary guidelines [151] should be on dietary patterns, recognizing the importance and complex interactions of the food matrix and the energy conditions under which foods are consumed, rather than limited to single nutrients, like total fructose-containing sugars. Our present results demonstrating the benefit of fruit on inflammation are also supported by the Global Burden of Disease Study which showed the most important contributors to the global burden of morbidity and mortality are foods we should increase intake of, include increased intake of fruit [152]. Fruits are an important source of dietary fibre and key nutrients, such as potassium, both of which are heavily under-consumed in many populations [153,154]. Thus, a focus on policies encouraging the intake of important foods like fruit can help improve nutrient intakes in the general population in addition to improving health outcomes. In addition to fruit, the present evidence to support sweetened dairy alternatives, specifically soy beverages, and limiting sugars coming from mixed sources, including sugar-sweetened beverages, are supported by obesity, diabetes and cardiovascular guidelines which recommend following plant-based dietary patterns (Mediterranean, vegetarian, Portfolio, dietary approaches to stop hypertension (DASH), low-GI dietary patterns), which encourage food sources of fructose-containing sugars like fruits, vegetables, and whole grains, and reducing intake of others like sugar-sweetened beverages [155–160].

5. Conclusions

Overall food source appears to mediate the effect of fructose-containing sugars on inflammatory markers in predominantly adults with or without obesity, some of whom have or are at risk for cardiometabolic diseases over the short-to-medium term. The evidence provides good indication that mixed sources that contain SSBs increase CRP, while most other food sources have no effect with some sources (fruit, 100% fruit juice, sweetened soy beverage or dark chocolate) showing decreases, which may be dependent on energy control. The main sources of uncertainty across the analyses were imprecision and indirectness with a particular lack of food sources assessed and data available for subtraction and ad libitum trials. Although there remains a need for larger, longer, high-quality randomized trials assessing a broader variety of food sources of fructose-containing sugars on inflammatory biomarkers, clinical practice guidelines should consider the role of food source of fructose-containing sugars in the management of inflammation.

Supplementary Materials: Supplementary materials are available online at https://www.mdpi.com/ article/10.3390/nu14193986/s1, Table S1: PRISMA Checklist, Table S2: Search strategy for controlled trials assessing the effect of important food sources of fructose-containing sugars on biomarkers of inflammation, Table S3: PICOTS framework of the search strategy, Table S4: Food source definitions, Table S5: Trial characteristics, Table S6: Sensitivity analyses of the use of correlation coefficients of 0.25and 0.75 for crossover trials in the primary analysis of the effect of important food sources of fructosecontaining sugars on biomarkers of inflammation outcomes, Table S7: GRADE certainty of evidence assessment* for the effect of fructose-containing sugars on biomarkers of inflammation by energy control, Table S8: GRADE certainty of evidence assessment* for the effect of fructose-containing sugars on biomarkers of inflammation by important food source of fructose-containing sugars, Figure S1: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on CRP (mg/L) in substitution trials, Figure S2: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on CRP (mg/L) in addition trials, Figure S3: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on CRP (mg/L) in subtraction trials, Figure S4: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on CRP (mg/L) in ad libitum trials, Figure S5: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on TNF- a(pg/mL) trials in substitution trials, Figure S6: Risk of bias proportion graph for the effect of important food sources of fructose-containing sugars on TNF-a(pg/mL) in addition trials, Figure

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for the effect of important food sources of fructose-containing sugars on IL-6 (pg/mL) in substitution trials, Figure S97: Publication bias funnel plots for the effect of important food sources of fructose-containing sugars on IL-6 (pg/mL) in addition trials, Figure S98: Trim and Fill funnel plot for the effect of important food sources of fructose-containing sugars IL-6 (pg/mL) in substitution trials, Figure S99: Trim and Fill funnel plot for the effect of important food sources of fructose-containing sugars IL-6 (pg/mL) in addition trials. **Author Contributions:** Research design (project conception, development of overall research plan, and study oversight), J.L.S., L.C., V.L.C., S.B.M., R.J.d.S., T.M.S.W., L.A.L., C.W.C.K., and D.J.A.J.; research conduct (hands-on conduct of the experiments and data collection), X.Q., L.C., S.A.-C., A.A., D.L., F.A.-Y., Q.L., and A.C.; data analysis and statistical analysis, X.Q., L.C., D.L., A.A., S.A.-C. and T.A.K.; writing—review and editing, 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Institutes of Health Research. He has served as an independent director of the Helderleigh Foundation (Canada). He serves as a member of the Nutrition Science Advisory Committee to Health Canada (Government of Canada), and a coopted member of the Scientific Advisory Committee on Nutrition (SACN) Subgroup on the Framework for the Evaluation of Evidence (Public Health England). T.M.S.W. was previously part owner, and now is an employee of INQUIS and received an honorarium from Springer/Nature for being an Associate Editor of the European Journal of Clinical Nutrition. C.W.C.K. has received grants or research support from the Advanced Food Materials Network, Agriculture and Agri-Foods Canada (AAFC), Almond Board of California, Barilla, Canadian Institutes of Health Research (CIHR), Canola Council of Canada, International Nut and Dried Fruit Council, International Tree Nut Council Research and Education Foundation, Loblaw Brands Ltd., the Peanut Institute, Pulse Canada and Unilever. 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Diabetes. He received an honorarium from the United States Department of Agriculture to present the 2013 W.O. Atwater Memorial Lecture. He received the 2013 Award for Excellence in Research from the International Nut and Dried Fruit Council. He received funding and travel support from the Canadian Society of Endocrinology and Metabolism to produce mini cases for the Canadian Diabetes Association (CDA). He is a member of the International Carbohydrate Quality Consortium (ICQC). His wife, Alexandra L Jenkins, is a director and partner of INQUIS Clinical Research for the Food Industry, his 2 daughters, Wendy Jenkins and Amy Jenkins, have published a vegetarian book that promotes the use of the foods described here, The Portfolio Diet for Cardiovascular Risk Reduction (Academic Press/Elsevier 2020 ISBN:978-0-12-810510-8) and his sister, Caroline Brydson, received funding through a grant from the St. Michael's Hospital Foundation to develop a cookbook for one of his studies. He is also a vegan. J.L.S. has received research support from the Canadian Foundation for Innovation, Ontario Research Fund, Province of Ontario Ministry of Research and Innovation and Science, Canadian Institutes of health Research (CIHR), Diabetes Canada, American Society for Nutrition (ASN), International Nut and Dried Fruit Council (INC) Foundation, National Honey Board (U.S. Department of Agriculture [USDA] honey "Checkoff" program), Institute for the Advancement of Food and Nutrition Sciences (IAFNS; formerly ILSI North America), Pulse Canada, Quaker Oats Center of Excellence, The United Soybean Board (USDA soy "Checkoff" program), The Tate and Lyle Nutritional Research Fund at the University of Toronto, The Glycemic Control and Cardiovascular Disease in Type 2 Diabetes Fund at the University of Toronto (a fund established by the Alberta Pulse Growers), The Plant Protein Fund at the University of Toronto (a fund which has received contributions from IFF), and The Nutrition Trialists Network Fund at the University of Toronto (a fund established by an inaugural donation from the Calorie Control Council). He has received food donations to support randomized controlled trials from the Almond Board of California, California Walnut Commission, Peanut Institute, Barilla, Unilever/Upfield, Unico/Primo, Loblaw Companies, Quaker, Kellogg Canada, WhiteWave Foods/Danone, Nutrartis, and Dairy Farmers of Canada. He has received travel support, speaker fees and/or honoraria from ASN, Danone, Dairy Farmers of Canada, FoodMinds LLC, Nestlé, Abbott, General Mills, Nutrition Communications, International Food Information Council (IFIC), Calorie Control Council, International Sweeteners Association, and International Glutamate Technical Committee. He has or has had ad hoc consulting arrangements with Perkins Coie LLP, Tate & Lyle, Phynova, and Inquis Clinical Research. He is a former member of the European Fruit Juice Association Scientific Expert Panel and former member of the Soy Nutrition Institute (SNI) Scientific Advisory Committee. He is on the Clinical Practice Guidelines Expert Committees of Diabetes Canada, European Association for the study of Diabetes (EASD), Canadian Cardiovascular Society (CCS), and Obesity Canada/Canadian Association of Bariatric Physicians and Surgeons. He serves or has served as an unpaid member of the Board of Trustees and an unpaid scientific advisor for the Carbohydrates Committee of IAFNS. He is a member of the International Carbohydrate Quality Consortium (ICQC), Executive Board Member of the Diabetes and Nutrition Study Group (DNSG) of the EASD, and Director of the Toronto 3D Knowledge Synthesis and Clinical Trials foundation. His spouse is an employee of AB InBev. X.Y.Q., Q.L, S.B.M., V.L.C., and L.A.L. declare no relevant competing interests.

References

- 1. Berg, A.H.; Scherer, P.E. Adipose tissue, inflammation, and cardiovascular disease. *Circ. Res.* 2005, *96*, 939–949. [CrossRef] [PubMed]
- Ridker, P.M. From Cantos to Cirt to Colcot to Clinic: Will All Atherosclerosis Patients Soon Be Treated with Combination Lipid-Lowering and Inflammation-Inhibiting Agents? *Circulation* 2020, 141, 787–789. [CrossRef] [PubMed]
- Calder, P.C.; Ahluwalia, N.; Brouns, F.; Buetler, T.; Clement, K.; Cunningham, K.; Esposito, K.; Jonsson, L.S.; Kolb, H.; Lansink, M.; et al. Dietary factors and low-grade inflammation in relation to overweight and obesity. *Br. J. Nutr.* 2011, 106 (Suppl. 3), S5–S78. [CrossRef] [PubMed]
- Li, J.; Lee, D.H.; Hu, J.; Tabung, F.K.; Li, Y.; Bhupathiraju, S.N.; Rimm, E.B.; Rexrode, K.M.; Manson, J.E.; Willett, W.C.; et al. Dietary Inflammatory Potential and Risk of Cardiovascular Disease Among Men and Women in the U.S. *J. Am. Coll. Cardiol.* 2020, 76, 2181–2193. [CrossRef]
- World Health Organization. *Guideline: Sugars Intake for Adults and Children;* World Health Organization: Geneva, Switzerland, 2015; Available online: http://apps.who.int/iris/bitstream/handle/10665/149782/9789241549028_eng.pdf;jsessionid=F9FAD1 9E165BB45830BA1A484FC6FD93?sequence=1 (accessed on 30 June 2022).
- Campos, V.C.; Tappy, L. Physiological handling of dietary fructose-containing sugars: Implications for health. *Int. J. Obes.* 2016, 40 (Suppl. 1), S6–S11. [CrossRef]
- 7. Sun, S.Z.; Empie, M.W. Fructose metabolism in humans—What isotopic tracer studies tell us. Nutr. Metab. 2012, 9, 89. [CrossRef]

- Ayoub-Charette, S.; Chiavaroli, L.; Liu, Q.; Khan, T.A.; Zurbau, A.; Au-Yeung, F.; Cheung, A.; Ahmed, A.; Lee, D.; Choo, V.L.; et al. Different Food Sources of Fructose-Containing Sugars and Fasting Blood Uric Acid Levels: A Systematic Review and Meta-Analysis of Controlled Feeding Trials. J. Nutr. 2021, 151, 2409–2421. [CrossRef]
- 9. Chiavaroli, L.; de Souza, R.J.; Ha, V.; Cozma, A.I.; Mirrahimi, A.; Wang, D.D.; Yu, M.; Carleton, A.J.; Di Buono, M.; Jenkins, A.L.; et al. Effect of Fructose on Established Lipid Targets: A Systematic Review and Meta-Analysis of Controlled Feeding Trials. J. Am. Heart Assoc. 2015, 4, e001700. [CrossRef]
- Chiu, S.; Sievenpiper, J.L.; de Souza, R.J.; Cozma, A.I.; Mirrahimi, A.; Carleton, A.J.; Ha, V.; Di Buono, M.; Jenkins, A.L.; Leiter, L.A.; et al. Effect of fructose on markers of non-alcoholic fatty liver disease (NAFLD): A systematic review and metaanalysis of controlled feeding trials. *Eur. J. Clin. Nutr.* 2014, *68*, 416–423. [CrossRef]
- 11. Choo, V.L.; Viguiliouk, E.; Blanco Mejia, S.; Cozma, A.I.; Khan, T.A.; Ha, V.; Wolever, T.M.S.; Leiter, L.A.; Vuksan, V.; Kendall, C.W.C.; et al. Food sources of fructose-containing sugars and glycaemic control: Systematic review and meta-analysis of controlled intervention studies. *BMJ* **2018**, *363*, k4644. [CrossRef]
- Sievenpiper, J.L.; de Souza, R.J.; Mirrahimi, A.; Yu, M.E.; Carleton, A.J.; Beyene, J.; Chiavaroli, L.; Di Buono, M.; Jenkins, A.L.; Leiter, L.A.; et al. Effect of fructose on body weight in controlled feeding trials: A systematic review and meta-analysis. *Ann. Intern. Med.* 2012, 156, 291–304. [CrossRef]
- Gersch, M.S.; Mu, W.; Cirillo, P.; Reungjui, S.; Zhang, L.; Roncal, C.; Sautin, Y.Y.; Johnson, R.J.; Nakagawa, T. Fructose, but not dextrose, accelerates the progression of chronic kidney disease. *Am. J. Physiol. Ren. Physiol.* 2007, 293, F1256–F1261. [CrossRef] [PubMed]
- 14. Malik, V.S.; Hu, F.B. The role of sugar-sweetened beverages in the global epidemics of obesity and chronic diseases. *Nat. Rev. Endocrinol.* **2022**, *18*, 205–218. [CrossRef] [PubMed]
- 15. Schulze, M.B.; Hoffmann, K.; Manson, J.E.; Willett, W.C.; Meigs, J.B.; Weikert, C.; Heidemann, C.; Colditz, G.A.; Hu, F.B. Dietary pattern, inflammation, and incidence of type 2 diabetes in women. *Am. J. Clin. Nutr.* **2005**, *82*, 675–684. [CrossRef] [PubMed]
- 16. DeChristopher, L.R.; Uribarri, J.; Tucker, K.L. Intake of high fructose corn syrup sweetened soft drinks, fruit drinks and apple juice is associated with prevalent coronary heart disease, in U.S. adults, ages 45–59 y. *BMC Nutr.* **2017**, *3*, 51. [CrossRef] [PubMed]
- Higgins, J.P.T.T.J.; Chandler, J.; Cumpston, M.; Li, T.; Page, M.J.; Welch, V.A. Cochrane Handbook for Systematic Reviews of Interventions Version 6.3. Cochrane. 2022. Available online: https://training.cochrane.org/handbook/current (accessed on 10 June 2022).
- 18. Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* **2009**, *6*, e1000097. [CrossRef] [PubMed]
- 19. Wilczynski, N.L.; Morgan, D.; Haynes, R.B.; Hedges, T. An overview of the design and methods for retrieving high-quality studies for clinical care. *BMC Med. Inf. Decis. Mak.* 2005, *5*, 20. [CrossRef]
- 20. Te Morenga, L.; Mallard, S.; Mann, J. Dietary sugars and body weight: Systematic review and meta-analyses of randomised controlled trials and cohort studies. *BMJ* **2012**, *346*, e7492. [CrossRef]
- 21. SourceForge. Plot Digitizer. 2001. Available online: http://plotdigitizer.sourceforge.net/ (accessed on 24 October 2015).
- Luo, D.; Wan, X.; Liu, J.; Tong, T. Optimally estimating the sample mean from the sample size, median, mid-range, and/or mid-quartile range. *Stat. Methods Med. Res.* 2018, 27, 1785–1805. [CrossRef]
- 23. Wan, X.; Wang, W.; Liu, J.; Tong, T. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. *BMC Med. Res. Methodol.* **2014**, *14*, 135. [CrossRef]
- 24. Furukawa, T.A.; Barbui, C.; Cipriani, A.; Brambilla, P.; Watanabe, N. Imputing missing standard deviations in meta-analyses can provide accurate results. *J. Clin. Epidemiol.* **2006**, *59*, 7–10. [CrossRef] [PubMed]
- 25. Borenstein, M.; Higgins, J.P. Meta-analysis and subgroups. Prev. Sci. 2013, 14, 134–143. [CrossRef] [PubMed]
- 26. Borenstein, M.; Hedges, L.V.; Higgins, J.P.; Rothstein, H.R. Introduction to Meta-Analysis; John Wiley & Sons: London, UK, 2009.
- 27. Deeks, J.J.; Higgins, J.P.; Altman, D.G.; Cochrane Statistical Methods Group. Analysing data and undertaking meta-analyses. In *Cochrane Handbook for Systematic Reviews of Interventions*; Wiley: New York, NY, USA, 2019; pp. 241–284.
- 28. DerSimonian, R.; Laird, N. Meta-analysis in clinical trials. Control. Clin. Trials 1986, 7, 177–188. [CrossRef]
- 29. Tufanaru, C.; Munn, Z.; Stephenson, M.; Aromataris, E. Fixed or random effects meta-analysis? Common methodological issues in systematic reviews of effectiveness. *Int. J. Evid. Based Healthc.* **2015**, *13*, 196–207. [CrossRef]
- 30. Elbourne, D.R.; Altman, D.G.; Higgins, J.P.T.; Curtin, F.; Worthington, H.V.; Vail, A. Meta-analyses involving cross-over trials: Methodological issues. *Int. J. Epidemiol.* **2002**, *31*, 140–149. [CrossRef]
- 31. Follmann, D.; Elliott, P.; Suh, I.; Cutler, J. Variance imputation for overviews of clinical trials with continuous response. *J. Clin. Epidemiol.* **1992**, 45, 769–773. [CrossRef]
- 32. Balk, E.M.; Earley, A.; Patel, K.; Trikalinos, T.A.; Dahabreh, I.J. AHRQ Methods for Effective Health Care. In *Empirical Assessment* of Within-Arm Correlation Imputation in Trials of Continuous Outcomes; Agency for Healthcare Research and Quality (US): Rockville, MD, USA, 2012.
- 33. Reynolds Risk Score. Available online: http://www.reynoldsriskscore.org/Default.aspx (accessed on 14 March 2020).
- 34. Ridker, P.M.; Buring, J.E.; Rifai, N.; Cook, N.R. Development and validation of improved algorithms for the assessment of global cardiovascular risk in women: The Reynolds Risk Score. *JAMA* 2007, 297, 611–619. [CrossRef]
- 35. Ridker, P.M.; Paynter, N.P.; Rifai, N.; Gaziano, J.M.; Cook, N.R. C-reactive protein and parental history improve global cardiovascular risk prediction: The Reynolds Risk Score for men. *Circulation* **2008**, *118*, 2243–2251. [CrossRef]

- Mayoclinic. Tumour Necrosis Factor (TNF), Plasma. Available online: https://www.mayocliniclabs.com/test-catalog/Clinical+ and+Interpretive/63022 (accessed on 18 March 2020).
- Mayoclinic. Interleukin 6, Plasma. Available online: https://www.mayocliniclabs.com/test-catalog/Clinical+and+Interpretive/ 63020 (accessed on 18 March 2020).
- 38. Thompson, S.G.; Higgins, J.P. How should meta-regression analyses be undertaken and interpreted? *Stat. Med.* 2002, 21, 1559–1573. [CrossRef]
- Fu, R.; Gartlehner, G.; Grant, M.; Shamliyan, T.; Sedrakyan, A.; Wilt, T.J.; Griffith, L.; Oremus, M.; Raina, P.; Ismaila, A.; et al. Conducting quantitative synthesis when comparing medical interventions: AHRQ and the Effective Health Care Program. *J. Clin. Epidemiol.* 2011, 64, 1187–1197. [CrossRef]
- Carbohydrates and Health: Scientific Advisory Committee on Nutrition. 2015. Available online: https://www.gov.uk/ government/publications/sacn-carbohydrates-and-health-report#:~{}:text=The%20Scientific%20Advisory%20Committee% 20on%20Nutrition%20(%20SACN%20)%20was%20asked%20by,2%20diabetes%2C%20bowel%20health%20and (accessed on 18 March 2020).
- 41. U.S. Department of Health and Human Services; U.S. Department of Agriculture. 2015–2020 Dietary Guidelines for Americans; National Academies Press: Washington, DC, USA, 2015; ISBN 9780160934650.
- 42. Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids; Institute of Medicine, The National Academies Press: Washington, DC, USA, 2005; 1358p.
- 43. Egger, M.; Smith, G.D.; Schneider, M.; Minder, C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* **1997**, *315*, 629–634. [CrossRef] [PubMed]
- 44. Begg, C.B.; Mazumdar, M. Operating Characteristics of a Rank Correlation Test for Publication Bias. *Biometrics* **1994**, *50*, 1088–1101. [CrossRef] [PubMed]
- 45. Sterne, J.A.; Gavaghan, D.; Egger, M. Publication and related bias in meta-analysis: Power of statistical tests and prevalence in the literature. *J. Clin. Epidemiol.* 2000, 53, 1119–1129. [CrossRef]
- 46. Duval, S.; Tweedie, R. Trim and fill: A simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics* **2000**, *56*, 455–463. [CrossRef] [PubMed]
- 47. Higgins, J.P.T.; Green, S. GRADE Handbook. 2013. Available online: https://handbook-5-1.cochrane.org (accessed on 10 November 2018).
- Andrews, J.; Guyatt, G.; Oxman, A.D.; Alderson, P.; Dahm, P.; Falck-Ytter, Y.; Nasser, M.; Meerpohl, J.; Post, P.N.; Kunz, R.; et al. GRADE guidelines: 14. Going from evidence to recommendations: The significance and presentation of recommendations. J. Clin. Epidemiol. 2013, 66, 719–725. [CrossRef] [PubMed]
- 49. Brunetti, M.; Shemilt, I.; Pregno, S.; Vale, L.; Oxman, A.D.; Lord, J.; Sisk, J.; Ruiz, F.; Hill, S.; Guyatt, G.H.; et al. GRADE guidelines: 10. Considering resource use and rating the quality of economic evidence. *J. Clin. Epidemiol.* **2013**, *66*, 140–150. [CrossRef]
- 50. Guyatt, G.H.; Oxman, A.D.; Schünemann, H.J. GRADE guidelines-an introduction to the 10th–13th articles in the series. *J. Clin. Epidemiol.* **2013**, *66*, 121–123. [CrossRef]
- Guyatt, G.H.; Thorlund, K.; Oxman, A.D.; Walter, S.D.; Patrick, D.; Furukawa, T.A.; Johnston, B.C.; Karanicolas, P.; Akl, E.A.; Vist, G.; et al. GRADE guidelines: 13. Preparing summary of findings tables and evidence profiles-continuous outcomes. *J. Clin. Epidemiol.* 2013, 66, 173–183. [CrossRef]
- Guyatt, G.; Oxman, A.D.; Sultan, S.; Brozek, J.; Glasziou, P.; Alonso-Coello, P.; Atkins, D.; Kunz, R.; Montori, V.; Jaeschke, R.; et al. GRADE guidelines: 11. Making an overall rating of confidence in effect estimates for a single outcome and for all outcomes. J. Clin. Epidemiol. 2013, 66, 151–157. [CrossRef]
- 53. Guyatt, G.H.; Oxman, A.D.; Santesso, N.; Helfand, M.; Vist, G.; Kunz, R.; Brozek, J.; Norris, S.; Meerpohl, J.; Djulbegovic, B.; et al. GRADE guidelines: 12. Preparing summary of findings tables-binary outcomes. *J. Clin. Epidemiol.* **2013**, *66*, 158–172. [CrossRef]
- Santesso, N.; Glenton, C.; Dahm, P.; Garner, P.; Akl, E.A.; Alper, B.; Brignardello-Petersen, R.; Carrasco-Labra, A.; De Beer, H.; Hultcrantz, M.; et al. GRADE guidelines 26: Informative statements to communicate the findings of systematic reviews of interventions. J. Clin. Epidemiol. 2020, 119, 126–135. [CrossRef] [PubMed]
- 55. Guyatt, G.H.; Oxman, A.D.; Sultan, S.; Glasziou, P.; Akl, E.A.; Alonso-Coello, P.; Atkins, D.; Kunz, R.; Brozek, J.; Montori, V.; et al. GRADE guidelines: 9. Rating up the quality of evidence. *J. Clin. Epidemiol.* **2011**, *64*, 1311–1316. [CrossRef] [PubMed]
- Balshem, H.; Helfand, M.; Schunemann, H.J.; Oxman, A.D.; Kunz, R.; Brozek, J.; Vist, G.E.; Falck-Ytter, Y.; Meerpohl, J.; Norris, S.; et al. GRADE guidelines: 3. Rating the quality of evidence. *J. Clin. Epidemiol.* 2011, 64, 401–406. [CrossRef] [PubMed]
- 57. Aeberli, I.; Gerber, P.A.; Hochuli, M.; Kohler, S.; Haile, S.R.; Gouni-Berthold, I.; Berthold, H.K.; Spinas, G.A.; Berneis, K. Low to moderate sugar-sweetened beverage consumption impairs glucose and lipid metabolism and promotes inflammation in healthy young men: A randomized controlled trial. *Am. J. Clin. Nutr.* **2011**, *94*, 479–485. [CrossRef]
- Aghababaee, S.K.; Vafa, M.; Shidfar, F.; Tahavorgar, A.; Gohari, M.; Katebi, D.; Mohammadi, V. Effects of blackberry (*Morus nigra* L.) consumption on serum concentration of lipoproteins, apo A-I, apo B, and high-sensitivity-C-reactive protein and blood pressure in dyslipidemic patients. *J. Res. Med. Sci.* 2015, 20, 685–691.
- Alavinejad, P.; Farsi, F.; Rezazadeh, A.; Mahmoodi, M.; Hajiani, E.; Mas-jedizadeh, A.R.; Mard, S.A.; Neisi, N.; Hoseini, H.; Moghaddam, E.K.; et al. The Effects of Dark Chocolate Consumption on Lipid Profile, Fasting Blood Sugar, Liver Enzymes, Inflammation, and Antioxidant Status in Patients with Non-Alcoholic Fatty Liver Disease: A Randomized, Placebo-Controlled, Pilot study. J. Gastroenterol. Hepatol. Res. 2015, 4, 1858–1864. [CrossRef]

- Angelopoulos, T.J.; Lowndes, J.; Sinnett, S.; Rippe, J.M. Fructose Containing Sugars at Normal Levels of Consumption Do Not Effect Adversely Components of the Metabolic Syndrome and Risk Factors for Cardiovascular Disease. *Nutrients* 2016, *8*, 179. [CrossRef]
- Asgary, S.; Sahebkar, A.; Afshani, M.R.; Keshvari, M.; Haghjooyjavanmard, S.; Rafieian-Kopaei, M. Clinical evaluation of blood pressure lowering, endothelial function improving, hypolipidemic and anti-inflammatory effects of pomegranate juice in hypertensive subjects. *Phytother. Res.* 2014, 28, 193–199. [CrossRef]
- 62. Basu, A.; Du, M.; Leyva, M.J.; Sanchez, K.; Betts, N.M.; Wu, M.; Aston, C.E.; Lyons, T.J. Blueberries decrease cardiovascular risk factors in obese men and women with metabolic syndrome. *J. Nutr.* **2010**, *140*, 1582–1587. [CrossRef]
- 63. Blum, A.; Monir, M.; Khazim, K.; Peleg, A.; Blum, N. Tomato-rich (Mediterranean) diet does not modify inflammatory markers. *Clin. Investig. Med.* **2007**, *30*, E70–E74. [CrossRef]
- 64. Brymora, A.; Flisinski, M.; Johnson, R.J.; Goszka, G.; Stefanska, A.; Manitius, J. Low-fructose diet lowers blood pressure and inflammation in patients with chronic kidney disease. *Nephrol. Dial. Transpl.* **2012**, *27*, 608–612. [CrossRef] [PubMed]
- 65. Campos, V.; Despland, C.; Brandejsky, V.; Kreis, R.; Schneiter, P.; Chiolero, A.; Boesch, C.; Tappy, L. Sugar- and artificially sweetened beverages and intrahepatic fat: A randomized controlled trial. *Obesity* **2015**, *23*, 2335–2339. [CrossRef] [PubMed]
- Castilla, P.; Davalos, A.; Teruel, J.L.; Cerrato, F.; Fernandez-Lucas, M.; Merino, J.L.; Sanchez-Martin, C.C.; Ortuno, J.; Lasuncion, M.A. Comparative effects of dietary supplementation with red grape juice and vitamin E on production of superoxide by circulating neutrophil NADPH oxidase in hemodialysis patients. *Am. J. Clin. Nutr.* 2008, *87*, 1053–1061. [CrossRef] [PubMed]
- 67. Chiu, S.; Siri-Tarino, P.; Bergeron, N.; Suh, J.H.; Krauss, R.M. A Randomized Study of the Effect of Replacing Sugar-Sweetened Soda by Reduced Fat Milk on Cardiometabolic Health in Male Adolescent Soda Drinkers. *Nutrients* **2020**, *12*, 405. [CrossRef]
- 68. Cox, C.L.; Stanhope, K.L.; Schwarz, J.M.; Graham, J.L.; Hatcher, B.; Griffen, S.C.; Bremer, A.A.; Berglund, L.; McGahan, J.P.; Keim, N.L.; et al. Circulating concentrations of monocyte chemoattractant protein-1, plasminogen activator inhibitor-1, and soluble leukocyte adhesion molecule-1 in overweight/obese men and women consuming fructose- or glucose-sweetened beverages for 10 weeks. *J. Clin. Endocrinol. Metab.* 2011, *96*, E2034–E2038. [CrossRef]
- 69. Dow, C.A.; Going, S.B.; Chow, H.H.; Patil, B.S.; Thomson, C.A. The effects of daily consumption of grapefruit on body weight, lipids, and blood pressure in healthy, overweight adults. *Metabolism* **2012**, *61*, 1026–1035. [CrossRef]
- 70. Ebbeling, C.B.; Feldman, H.A.; Steltz, S.K.; Quinn, N.L.; Robinson, L.M.; Ludwig, D.S. Effects of Sugar-Sweetened, Artificially Sweetened, and Unsweetened Beverages on Cardiometabolic Risk Factors, Body Composition, and Sweet Taste Preference: A Randomized Controlled Trial. J. Am. Heart Assoc. 2020, 9, e015668. [CrossRef]
- Ellis, C.L.; Edirisinghe, I.; Kappagoda, T.; Burton-Freeman, B. Attenuation of meal-induced inflammatory and thrombotic responses in overweight men and women after 6-week daily strawberry (Fragaria) intake. A randomized placebo-controlled trial. *J. Atheroscler. Thromb.* 2011, *18*, 318–327. [CrossRef]
- 72. Eslami, O.; Shidfar, F.; Maleki, Z.; Jazayeri, S.; Hosseini, A.F.; Agah, S.; Ardiyani, F. Effect of Soy Milk on Metabolic Status of Patients with Nonalcoholic Fatty Liver Disease: A Randomized Clinical Trial. *J. Am. Coll. Nutr.* **2019**, *38*, 51–58. [CrossRef]
- Fatel, E.C.S.; Rosa, F.T.; Alfieri, D.F.; Flauzino, T.; Scavuzzi, B.M.; Lozovoy, M.A.B.; Iriyoda, T.M.V.; Simao, A.N.C.; Dichi, I. Beneficial effects of fish oil and cranberry juice on disease activity and inflammatory biomarkers in people with rheumatoid arthritis. *Nutrition* 2021, *86*, 111183. [CrossRef]
- 74. Franck, M.; de Toro-Martin, J.; Garneau, V.; Guay, V.; Kearney, M.; Pilon, G.; Roy, D.; Couture, P.; Couillard, C.; Marette, A.; et al. Effects of Daily Raspberry Consumption on Immune-Metabolic Health in Subjects at Risk of Metabolic Syndrome: A Randomized Controlled Trial. *Nutrients* 2020, *12*, 3858. [CrossRef] [PubMed]
- 75. Fridell, S.; Strom, E.; Agebratt, C.; Leanderson, P.; Guldbrand, H.; Nystrom, F.H. A randomised study in young subjects of the effects of eating extra fruit or nuts on periodontal inflammation. *BDJ Open* **2018**, *4*, 17022. [CrossRef] [PubMed]
- Ghazali, W.S.; Romli, A.C.; Mohamed, M. Effects of honey supplementation on inflammatory markers among chronic smokers: A randomized controlled trial. BMC Complement. Altern. Med. 2017, 17, 175. [CrossRef] [PubMed]
- 77. Goss, A.M.; Dowla, S.; Pendergrass, M.; Ashraf, A.; Bolding, M.; Morrison, S.; Amerson, A.; Soleymani, T.; Gower, B. Effects of a carbohydrate-restricted diet on hepatic lipid content in adolescents with non-alcoholic fatty liver disease: A pilot, randomized trial. *Pediatr. Obes.* 2020, 15, e12630. [CrossRef]
- 78. Hooshmand, S.; Kern, M.; Metti, D.; Shamloufard, P.; Chai, S.C.; Johnson, S.A.; Payton, M.E.; Arjmandi, B.H. The effect of two doses of dried plum on bone density and bone biomarkers in osteopenic postmenopausal women: A randomized, controlled trial. *Osteoporos. Int.* 2016, 27, 2271–2279. [CrossRef]
- 79. Irannejad Niri, Z.; Shidfar, F.; Jabbari, M.; Zarrati, M.; Hosseini, A.; Malek, M.; Dehnad, A. The effect of dried Ziziphus vulgaris on glycemic control, lipid profile, Apo-proteins and hs-CRP in patients with type 2 diabetes mellitus: A randomized controlled clinical trial. *J. Food Biochem.* **2021**, *45*, e13193. [CrossRef]
- Jafarirad, S.; Ayoobi, N.; Karandish, M.; Jalali, M.T.; Haghighizadeh, M.H.; Jahanshahi, A. Dark Chocolate Effect on Serum Adiponectin, Biochemical and Inflammatory Parameters in Diabetic Patients: A Randomized Clinical Trial. *Int. J. Prev. Med.* 2018, 9, 86. [CrossRef]
- Jalilvand, A.; Behrouz, V.; Nikpayam, O.; Sohrab, G.; Hekmatdoost, A. Effects of low fructose diet on glycemic control, lipid profile and systemic inflammation in patients with type 2 diabetes: A single-blind randomized controlled trial. *Diabetes Metab. Syndr.* 2020, 14, 849–855. [CrossRef]

- Jin, R.; Welsh, J.A.; Le, N.A.; Holzberg, J.; Sharma, P.; Martin, D.R.; Vos, M.B. Dietary fructose reduction improves markers of cardiovascular disease risk in Hispanic-American adolescents with NAFLD. *Nutrients* 2014, 6, 3187–3201. [CrossRef]
- Johnston, R.D.; Stephenson, M.C.; Crossland, H.; Cordon, S.M.; Palcidi, E.; Cox, E.F.; Taylor, M.A.; Aithal, G.P.; Macdonald, I.A. No difference between high-fructose and high-glucose diets on liver triacylglycerol or biochemistry in healthy overweight men. *Gastroenterology* 2013, 145, 1016–1025.e1012. [CrossRef]
- Kaliora, A.C.; Kokkinos, A.; Diolintzi, A.; Stoupaki, M.; Gioxari, A.; Kanellos, P.T.; Dedoussis, G.V.Z.; Vlachogiannakos, J.; Revenas, C.; Ladas, S.D.; et al. The effect of minimal dietary changes with raisins in NAFLD patients with non-significant fibrosis: A randomized controlled intervention. *Food Funct.* 2016, 7, 4533–4544. [CrossRef] [PubMed]
- 85. Kanellos, P.T.; Kaliora, A.C.; Protogerou, A.D.; Tentolouris, N.; Perrea, D.N.; Karathanos, V.T. The effect of raisins on biomarkers of endothelial function and oxidant damage; an open-label and randomized controlled intervention. *Food Res. Int.* **2017**, *102*, 674–680. [CrossRef] [PubMed]
- Kanellos, P.T.; Kaliora, A.C.; Tentolouris, N.K.; Argiana, V.; Perrea, D.; Kalogeropoulos, N.; Kountouri, A.M.; Karathanos, V.T. A pilot, randomized controlled trial to examine the health outcomes of raisin consumption in patients with diabetes. *Nutrition* 2014, 30, 358–364. [CrossRef] [PubMed]
- Karlsen, A.; Paur, I.; Bohn, S.K.; Sakhi, A.K.; Borge, G.I.; Serafini, M.; Erlund, I.; Laake, P.; Tonstad, S.; Blomhoff, R. Bilberry juice modulates plasma concentration of NF-kappaB related inflammatory markers in subjects at increased risk of CVD. *Eur. J. Nutr.* 2010, 49, 345–355. [CrossRef] [PubMed]
- Kelley, D.S.; Adkins, Y.; Reddy, A.; Woodhouse, L.R.; Mackey, B.E.; Erickson, K.L. Sweet bing cherries lower circulating concentrations of markers for chronic inflammatory diseases in healthy humans. J. Nutr. 2013, 143, 340–344. [CrossRef] [PubMed]
- Khodami, B.; Hatami, B.; Yari, Z.; Alavian, S.M.; Sadeghi, A.; Varkaneh, H.K.; Santos, H.O.; Hekmatdoost, A. Effects of a low free sugar diet on the management of nonalcoholic fatty liver disease: A randomized clinical trial. *Eur. J. Clin. Nutr.* 2022, 76, 987–994. [CrossRef] [PubMed]
- Kolehmainen, M.; Mykkanen, O.; Kirjavainen, P.V.; Leppanen, T.; Moilanen, E.; Adriaens, M.; Laaksonen, D.E.; Hallikainen, M.; Puupponen-Pimia, R.; Pulkkinen, L.; et al. Bilberries reduce low-grade inflammation in individuals with features of metabolic syndrome. *Mol. Nutr. Food Res.* 2012, 56, 1501–1510. [CrossRef] [PubMed]
- 91. Kuzma, J.N.; Cromer, G.; Hagman, D.K.; Breymeyer, K.L.; Roth, C.L.; Foster-Schubert, K.E.; Holte, S.E.; Weigle, D.S.; Kratz, M. No differential effect of beverages sweetened with fructose, high-fructose corn syrup, or glucose on systemic or adipose tissue inflammation in normal-weight to obese adults: A randomized controlled trial. *Am. J. Clin. Nutr.* **2016**, *104*, 306–314. [CrossRef]
- 92. Leelarungrayub, J.; Laskin, J.J.; Bloomer, R.J.; Pinkaew, D. Consumption of star fruit juice on pro-inflammatory markers and walking distance in the community dwelling elderly. *Arch. Gerontol. Geriatr.* **2016**, *64*, 6–12. [CrossRef] [PubMed]
- 93. Lehtonen, H.M.; Suomela, J.P.; Tahvonen, R.; Vaarno, J.; Venojärvi, M.; Viikari, J.; Kallio, H. Berry meals and risk factors associated with metabolic syndrome. *Eur. J. Clin. Nutr.* **2010**, *64*, 614–621. [CrossRef]
- Lehtonen, H.M.; Suomela, J.P.; Tahvonen, R.; Yang, B.; Venojärvi, M.; Viikari, J.; Kallio, H. Different berries and berry fractions have various but slightly positive effects on the associated variables of metabolic diseases on overweight and obese women. *Eur. J. Clin. Nutr.* 2011, 65, 394–401. [CrossRef] [PubMed]
- Liddle, D.M.; Lin, X.; Cox, L.C.; Ward, E.M.; Ansari, R.; Wright, A.J.; Robinson, L.E. Daily apple consumption reduces plasma and peripheral blood mononuclear cell-secreted inflammatory biomarkers in adults with overweight and obesity: A 6-week randomized, controlled, parallel-arm trial. *Am. J. Clin. Nutr.* 2021, *114*, 752–763. [CrossRef] [PubMed]
- Maki, K.C.; Palacios, O.M.; Kramer, M.W.; Trivedi, R.; Dicklin, M.R.; Wilcox, M.L.; Maki, C.E. Effects of substituting eggs for high-carbohydrate breakfast foods on the cardiometabolic risk-factor profile in adults at risk for type 2 diabetes mellitus. *Eur. J. Clin. Nutr.* 2020, 74, 784–795. [CrossRef] [PubMed]
- 97. Markey, O.; Le Jeune, J.; Lovegrove, J.A. Energy compensation following consumption of sugar-reduced products: A randomized controlled trial. *Eur. J. Nutr.* 2016, *55*, 2137–2149. [CrossRef] [PubMed]
- Martini, D.; Rosi, A.; Tassotti, M.; Antonini, M.; Dall'Asta, M.; Bresciani, L.; Fantuzzi, F.; Spigoni, V.; Dominguez-Perles, R.; Angelino, D.; et al. Effect of coffee and cocoa-based confectionery containing coffee on markers of cardiometabolic health: Results from the pocket-4-life project. *Eur. J. Nutr.* 2021, 60, 1453–1463. [CrossRef]
- Mietus-Snyder, M.L.; Shigenaga, M.K.; Suh, J.H.; Shenvi, S.V.; Lal, A.; McHugh, T.; Olson, D.; Lilienstein, J.; Krauss, R.M.; Gildengoren, G.; et al. A nutrient-dense, high-fiber, fruit-based supplement bar increases HDL cholesterol, particularly large HDL, lowers homocysteine, and raises glutathione in a 2-wk trial. *FASEB J.* 2012, *26*, 3515–3527. [CrossRef]
- Moazen, S.; Amani, R.; Homayouni Rad, A.; Shahbazian, H.; Ahmadi, K.; Taha Jalali, M. Effects of freeze-dried strawberry supplementation on metabolic biomarkers of atherosclerosis in subjects with type 2 diabetes: A randomized double-blind controlled trial. *Ann. Nutr. Metab.* 2013, 63, 256–264. [CrossRef]
- 101. Munsters, M.J.; Saris, W.H. The effect of sugar-sweetened beverage intake on energy intake in an ad libitum 6-month low-fat high-carbohydrate diet. *Ann. Nutr. Metab.* **2010**, *57*, 116–123. [CrossRef]
- 102. Navaei, N.; Pourafshar, S.; Akhavan, N.S.; Litwin, N.S.; Foley, E.M.; George, K.S.; Hartley, S.C.; Elam, M.L.; Rao, S.; Arjmandi, B.H.; et al. Influence of daily fresh pear consumption on biomarkers of cardiometabolic health in middle-aged/older adults with metabolic syndrome: A randomized controlled trial. *Food Funct.* 2019, 10, 1062–1072. [CrossRef]

- Nilholm, C.; Larsson, E.; Sonestedt, E.; Roth, B.; Ohlsson, B. Assessment of a 4-Week Starch- and Sucrose-Reduced Diet and Its Effects on Gastrointestinal Symptoms and Inflammatory Parameters among Patients with Irritable Bowel Syndrome. *Nutrients* 2021, 13, 416. [CrossRef]
- 104. Njike, V.Y.; Faridi, Z.; Shuval, K.; Dutta, S.; Kay, C.D.; West, S.G.; Kris-Etherton, P.M.; Katz, D.L. Effects of sugar-sweetened and sugar-free cocoa on endothelial function in overweight adults. *Int. J. Cardiol.* 2011, 149, 83–88. [CrossRef] [PubMed]
- 105. Palacios, O.M.; Maki, K.C.; Xiao, D.; Wilcox, M.L.; Dicklin, M.R.; Kramer, M.; Trivedi, R.; Burton-Freeman, B.; Edirisinghe, I. Effects of Consuming Almonds on Insulin Sensitivity and Other Cardiometabolic Health Markers in Adults with Prediabetes. J. Am. Coll. Nutr. 2020, 39, 397–406. [CrossRef]
- 106. Ponce, O.; Benassi, R.; Cesar, T. Orange juice associated with a balanced diet mitigated risk factors of metabolic syndrome: A randomized controlled trial. *J. Nutr. Intermed. Metab.* **2019**, *17*, 100101. [CrossRef]
- 107. Puglisi, M.J.; Vaishnav, U.; Shrestha, S.; Torres-Gonzalez, M.; Wood, R.J.; Volek, J.S.; Fernandez, M.L. Raisins and additional walking have distinct effects on plasma lipids and inflammatory cytokines. *Lipids Health Dis*/ **2008**, *7*, 14. [CrossRef]
- 108. Ravn-Haren, G.; Dragsted, L.O.; Buch-Andersen, T.; Jensen, E.N.; Jensen, R.I.; Németh-Balogh, M.; Paulovicsová, B.; Bergström, A.; Wilcks, A.; Licht, T.R.; et al. Intake of whole apples or clear apple juice has contrasting effects on plasma lipids in healthy volunteers. *Eur. J. Nutr.* 2013, 52, 1875–1889. [CrossRef]
- 109. Ribeiro, C.; Dourado, G.; Cesar, T. Orange juice allied to a reduced-calorie diet results in weight loss and ameliorates obesityrelated biomarkers: A randomized controlled trial. *Nutrition* **2017**, *38*, 13–19. [CrossRef]
- Sadeghi, F.; Akhlaghi, M.; Salehi, S. Adverse effects of honey on low-density lipoprotein cholesterol and adiponectin concentrations in patients with type 2 diabetes: A randomized controlled cross-over trial. J. Diabetes Metab. Disord. 2020, 19, 373–380.
 [CrossRef]
- 111. Sanchez-Delgado, M.; Estrada, J.A.; Paredes-Cervantes, V.; Kaufer-Horwitz, M.; Contreras, I. Changes in nutrient and calorie intake, adipose mass, triglycerides and TNF-alpha concentrations after non-caloric sweetener intake: A pilot study. *Int. J. Vitam Nutr. Res.* 2021, 91, 87–98. [CrossRef]
- 112. Schell, J.; Betts, N.M.; Lyons, T.J.; Basu, A. Raspberries Improve Postprandial Glucose and Acute and Chronic Inflammation in Adults with Type 2 Diabetes. *Ann. Nutr. Metab.* **2019**, *74*, 165–174. [CrossRef]
- 113. Simao, T.N.; Lozovoy, M.A.; Simao, A.N.; Oliveira, S.R.; Venturini, D.; Morimoto, H.K.; Miglioranza, L.H.; Dichi, I. Reducedenergy cranberry juice increases folic acid and adiponectin and reduces homocysteine and oxidative stress in patients with the metabolic syndrome. *Br. J. Nutr.* 2013, *110*, 1885–1894. [CrossRef]
- 114. Souto, D.L.; Zajdenverg, L.; Rodacki, M.; Rosado, E.L. Does sucrose intake affect antropometric variables, glycemia, lipemia and C-reactive protein in subjects with type 1 diabetes?: A controlled-trial. *Diabetol. Metab. Syndr.* **2013**, *5*, 67. [CrossRef] [PubMed]
- 115. Thimoteo, N.S.B.; Iryioda, T.M.V.; Alfieri, D.F.; Rego, B.E.F.; Scavuzzi, B.M.; Fatel, E.; Lozovoy, M.A.B.; Simao, A.N.C.; Dichi, I. Cranberry juice decreases disease activity in women with rheumatoid arthritis. *Nutrition* **2019**, *60*, 112–117. [CrossRef] [PubMed]
- 116. van Meijl, L.E.; Mensink, R.P. Low-fat dairy consumption reduces systolic blood pressure, but does not improve other metabolic risk parameters in overweight and obese subjects. *Nutr. Metab. Cardiovasc. Dis.* **2011**, *21*, 355–361. [CrossRef] [PubMed]
- 117. Vaz, M.; Pauline, M.; Unni, U.S.; Parikh, P.; Thomas, T.; Bharathi, A.V.; Avadhany, S.; Muthayya, S.; Mehra, R.; Kurpad, A.V. Micronutrient supplementation improves physical performance measures in Asian Indian school-age children. *J. Nutr.* 2011, 141, 2017–2023. [CrossRef]
- 118. Zafrilla, P.; Masoodi, H.; Cerdá, B.; García-Viguera, C.; Villaño, D. Biological effects of stevia, sucralose and sucrose in citrus-maqui juices on overweight subjects. *Food Funct.* 2021, *12*, 8535–8543. [CrossRef]
- 119. Du, C.; Smith, A.; Avalos, M.; South, S.; Crabtree, K.; Wang, W.; Kwon, Y.H.; Vijayagopal, P.; Juma, S. Blueberries Improve Pain, Gait Performance, and Inflammation in Individuals with Symptomatic Knee Osteoarthritis. *Nutrients* **2019**, *11*, 290. [CrossRef]
- Pothasak, Y.; Leelarungrayub, J.; Natakankitkul, S.; Singhatong, S. Prototype Star Fruit-Honey Product and Effectiveness on Antixidants, Inflammation and Walking Distance in Participants with Stable Chronic Obstructive Pulmonary Disease (COPD). *Pharm. J.* 2020, 12, 1121–1134. [CrossRef]
- 121. Della Corte, K.W.; Perrar, I.; Penczynski, K.J.; Schwingshackl, L.; Herder, C.; Buyken, A.E. Effect of Dietary Sugar Intake on Biomarkers of Subclinical Inflammation: A Systematic Review and Meta-Analysis of Intervention Studies. *Nutrients* 2018, 10, 606. [CrossRef]
- 122. Huang, H.; Chen, G.; Liao, D.; Zhu, Y.; Xue, X. Effects of Berries Consumption on Cardiovascular Risk Factors: A Meta-analysis with Trial Sequential Analysis of Randomized Controlled Trials. *Sci. Rep.* **2016**, *6*, 23625. [CrossRef]
- 123. Hosseini, B.; Berthon, B.S.; Saedisomeolia, A.; Starkey, M.R.; Collison, A.; Wark, P.A.B.; Wood, L.G. Effects of fruit and vegetable consumption on inflammatory biomarkers and immune cell populations: A systematic literature review and meta-analysis. *Am. J. Clin. Nutr.* 2018, 108, 136–155. [CrossRef]
- 124. Chiavaroli, L.; Cheung, A.; Ayoub-Charette, S.; Ahmed, A.; Lee, D.; Au-Yeung, F.; Qi, X.; Back, S.; McGlynn, N.; Ha, V.; et al. Important food sources of fructose-containing sugars and adiposity: A systematic review and meta-analysis of controlled feeding trials. *Am. J. Clin. Nutr.* **2022**, *in press*.
- 125. Liu, Q.; Chiavaroli, L.; Ayoub-Charette, S.; Ahmed, A.; Khan, T.A.; Au-Yeung, F.; Lee, D.; Cheung, A.; Zurbau, A.; Choo, V.L.; et al. Fructose-containing food sources and blood pressure: A systematic review abnd meta-analysis of controlled feeding trials. *PLoS ONE* 2022, *in press*.

- 126. Cara, K.C.; Beauchesne, A.R.; Wallace, T.C.; Chung, M. Effects of 100% Orange Juice on Markers of Inflammation and Oxidation in Healthy and At-Risk Adult Populations: A Scoping Review, Systematic Review, and Meta-analysis. *Adv. Nutr.* 2022, 13, 116–137. [CrossRef] [PubMed]
- 127. Liu, Q.; Ayoub-Charette, S.; Khan, T.A.; Au-Yeung, F.; Blanco Mejia, S.; de Souza, R.J.; Wolever, T.M.S.; Leiter, L.A.; Kendall, C.W.C.; Sievenpiper, J.L. Important Food Sources of Fructose-Containing Sugars and Incident Hypertension: A Systematic Review and Dose-Response Meta-Analysis of Prospective Cohort Studies. J. Am. Heart Assoc. 2019, 8, e010977. [CrossRef] [PubMed]
- 128. Semnani-Azad, Z.; Khan, T.A.; Blanco Mejia, S.; de Souza, R.J.; Leiter, L.A.; Kendall, C.W.C.; Hanley, A.J.; Sievenpiper, J.L. Association of Major Food Sources of Fructose-Containing Sugars with Incident Metabolic Syndrome: A Systematic Review and Meta-analysis. JAMA Netw. Open 2020, 3, e209993. [CrossRef] [PubMed]
- 129. Zurbau, A.; Au-Yeung, F.; Blanco Mejia, S.; Khan, T.A.; Vuksan, V.; Jovanovski, E.; Leiter, L.A.; Kendall, C.W.C.; Jenkins, D.J.A.; Sievenpiper, J.L. Relation of Different Fruit and Vegetable Sources with Incident Cardiovascular Outcomes: A Systematic Review and Meta-Analysis of Prospective Cohort Studies. J. Am. Heart Assoc. 2020, 9, e017728. [CrossRef]
- 130. Pan, A.; Malik, V.S.; Hao, T.; Willett, W.C.; Mozaffarian, D.; Hu, F.B. Changes in water and beverage intake and long-term weight changes: Results from three prospective cohort studies. *Int. J. Obes.* **2013**, *37*, 1378–1385. [CrossRef]
- 131. Khodarahmi, M.; Foroumandi, E.; Asghari Jafarabadi, M. Effects of soy intake on circulating levels of TNF-alpha and interleukin-6: A systematic review and meta-analysis of randomized controlled trials. *Eur. J. Nutr.* **2021**, *60*, 581–601. [CrossRef]
- Bajerska, J.; Lagowska, K.; Mori, M.; Regula, J.; Skoczek-Rubinska, A.; Toda, T.; Mizuno, N.; Yamori, Y. A Meta-Analysis of Randomized Controlled Trials of the Effects of Soy Intake on Inflammatory Markers in Postmenopausal Women. *J. Nutr.* 2022, 152, 5–15. [CrossRef]
- 133. Prokopidis, K.; Mazidi, M.; Sankaranarayanan, R.; Tajik, B.; McArdle, A.; Isanejad, M. Effects of whey and soy protein supplementation on inflammatory cytokines in older adults: A systematic review and meta-analysis. *Br. J. Nutr.* **2022**, 1–12. [CrossRef]
- 134. Chiavaroli, L.; Nishi, S.K.; Khan, T.A.; Braunstein, C.R.; Glenn, A.J.; Mejia, S.B.; Rahelic, D.; Kahleova, H.; Salas-Salvado, J.; Jenkins, D.J.A.; et al. Portfolio Dietary Pattern and Cardiovascular Disease: A Systematic Review and Meta-analysis of Controlled Trials. Prog. Cardiovasc. Dis. 2018, 61, 43–53. [CrossRef] [PubMed]
- 135. Jenkins, D.J.; Kendall, C.W.; Marchie, A.; Faulkner, D.A.; Wong, J.M.; de Souza, R.; Emam, A.; Parker, T.L.; Vidgen, E.; Lapsley, K.G.; et al. Effects of a dietary portfolio of cholesterol-lowering foods vs lovastatin on serum lipids and C-reactive protein. *JAMA* 2003, 290, 502–510. [CrossRef]
- 136. Zhao, B.; Gan, L.; Yu, K.; Mannisto, S.; Huang, J.; Albanes, D. Relationship between chocolate consumption and overall and cause-specific mortality, systematic review and updated meta-analysis. *Eur. J. Epidemiol.* **2022**, *37*, 321–333. [CrossRef] [PubMed]
- Rahmani, S.; Sadeghi, O.; Sadeghian, M.; Sadeghi, N.; Larijani, B.; Esmaillzadeh, A. The Effect of Whole-Grain Intake on Biomarkers of Subclinical Inflammation: A Comprehensive Meta-analysis of Randomized Controlled Trials. *Adv. Nutr.* 2020, 11, 52–65. [CrossRef]
- 138. Reynolds, A.N.; Akerman, A.P.; Mann, J. Dietary fibre and whole grains in diabetes management: Systematic review and meta-analyses. *PLoS Med.* 2020, *17*, e1003053. [CrossRef] [PubMed]
- 139. Lee, D.; Chiavaroli, L.; Ayoub-Charette, S.; Khan, T.A.; Zurbau, A.; Au-Yeung, F.; Cheung, A.; Liu, Q.; Qi, X.; Ahmed, A.; et al. Important Food Sources of Fructose-Containing Sugars and Non-Alcoholic Fatty Liver Disease: A Systematic Review and Meta-Analysis of Controlled Trials. *Nutrients* 2022, 14, 2846. [CrossRef]
- 140. Chiavaroli, L.; Cheung, A.; Ayoub-Charette, S.; Ahmed, A.; Lee, D.; Au-Yeung, F.; McGlynn, N.; Ha, V.; Khan, T.A.; Blanco Mejia, S.; et al. Important Food Sources of Fructose-Containing Sugars and Adiposity: A Systematic Review and Meta-Analysis of Controlled Feeding Trials. *Curr. Dev. Nutr.* 2021, *5*, 1017. [CrossRef]
- 141. Suen, J.; Thomas, J.; Kranz, A.; Vun, S.; Miller, M. Effect of Flavonoids on Oxidative Stress and Inflammation in Adults at Risk of Cardiovascular Disease: A Systematic Review. *Healthcare* **2016**, *4*, 69. [CrossRef]
- 142. Amiot, M.J.; Riva, C.; Vinet, A. Effects of dietary polyphenols on metabolic syndrome features in humans: A systematic review. *Obes. Rev.* **2016**, *17*, 573–586. [CrossRef]
- 143. Nakagawa, T.; Lanaspa, M.A.; Johnson, R.J. The effects of fruit consumption in patients with hyperuricaemia or gout. *Rheumatology* **2019**, *58*, 1133–1141. [CrossRef]
- 144. Atkinson, F.S.; Brand-Miller, J.C.; Foster-Powell, K.; Buyken, A.E.; Goletzke, J. International tables of glycemic index and glycemic load values 2021: A systematic review. *Am. J. Clin. Nutr.* **2021**, *114*, 1625–1632. [CrossRef] [PubMed]
- 145. Chiavaroli, L.; Lee, D.; Ahmed, A.; Cheung, A.; Khan, T.A.; Blanco, S.M.; Mirrahimi, A.; Jenkins, D.J.A.; Livesey, G.; Wolever, T.M.S.; et al. Effect of low glycaemic index or load dietary patterns on glycaemic control and cardiometabolic risk factors in diabetes: Systematic review and meta-analysis of randomised controlled trials. *BMJ* **2021**, *374*, n1651. [CrossRef] [PubMed]
- 146. Colagiuri, S.; Dickinson, S.; Girgis, S.; Colagiuri, R. National Evidence Based Guideline for Blood Glucose Control in Type 2 Diabetes. Diabetes Australia and the NHMRC. Canberra. Available online: https://www.diabetesaustralia.com.au/wpcontent/uploads/National-Evidence-Based-Guideline-for-Blood-Glucose-Control-in-Type-2-Diabetes.pdf2009 (accessed on 18 March 2020).
- 147. Jenkins, D.J.; Kendall, C.W.; Augustin, L.S.; Mitchell, S.; Sahye-Pudaruth, S.; Blanco Mejia, S.; Chiavaroli, L.; Mirrahimi, A.; Ireland, C.; Bashyam, B.; et al. Effect of legumes as part of a low glycemic index diet on glycemic control and cardiovascular risk factors in type 2 diabetes mellitus: A randomized controlled trial. *Arch. Intern. Med.* **2012**, *172*, 1653–1660. [CrossRef]

- 148. Slavin, J.L. Dietary fiber and body weight. Nutrition 2005, 21, 411–418. [CrossRef] [PubMed]
- 149. Ludwig, D.S. Dietary glycemic index and obesity. J. Nutr. 2000, 130 (Suppl. S2), 280S–283S. [CrossRef] [PubMed]
- Viguiliouk, E.; Nishi, S.K.; Wolever, T.M.S.; Sievenpiper, J.L. Point: Glycemic Index—An Important but Oft misunderstood Marker of Carbohydrate Quality. *Cereal Foods World* 2018, 63, 158–164.
- 151. Sievenpiper, J.L.; Dworatzek, P.D. Food and dietary pattern-based recommendations: An emerging approach to clinical practice guidelines for nutrition therapy in diabetes. *Can. J. Diabetes* **2013**, *37*, 51–57. [CrossRef]
- 152. Afshin, A.; Sur, P.J.; Fay, K.A.; Cornaby, L.; Ferrara, G.; Salama, J.S.; Mullany, E.C.; Hassen, K.; Abbafati, A.C.; Murray, C.J.; et al. Health effects of dietary risks in 195 countries, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet* 2019, 393, 1958–1972. [CrossRef]
- 153. Ahmed, M.; Praneet Ng, A.; L'Abbe, M.R. Nutrient intakes of Canadian adults: Results from the Canadian Community Health Survey (CCHS)-2015 Public Use Microdata File. *Am. J. Clin. Nutr.* **2021**, *114*, 1131–1140. [CrossRef]
- 154. Lee, S.H.; Moore, L.V.; Park, S.; Harris, D.M.; Blanck, H.M. Adults Meeting Fruit and Vegetable Intake Recommendations—United States, 2019. *MMWR Morb. Mortal. Wkly. Rep.* 2022, 71, 1–9. [CrossRef]
- 155. Brown, J.; Clarke, C.; Johnson Stoklossa, C.; Sievenpiper, J. Canadian Adult Obesity Clinical Practice Guidelines: Medical Nutrition Therapy in Obesity Management. Available online: <u>https://obesitycanada.ca/guidelines/nutrition</u> (accessed on 4 March 2022).
- 156. Wharton, S.; Lau, D.C.W.; Vallis, M.; Sharma, A.M.; Biertho, L.; Campbell-Scherer, D.; Adamo, K.; Alberga, A.; Bell, R.; Boule, N.; et al. Obesity in adults: A clinical practice guideline. *CMAJ* **2020**, *192*, E875–E891. [CrossRef] [PubMed]
- 157. Canadian Diabetes Association Clinical Practice Guidelines Expert Committee; Sievenpiper, J.L.; Chan, C.B.; Dworatzek, P.D.; Freeze, C.; Williams, S.L. Nutrition Therapy. *Can. J. Diabetes* **2018**, *42* (Suppl. 1), S64–S79. [CrossRef]
- 158. Anderson, T.J.; Gregoire, J.; Pearson, G.J.; Barry, A.R.; Couture, P.; Dawes, M.; Francis, G.A.; Genest, J., Jr.; Grover, S.; Gupta, M.; et al. 2016 Canadian Cardiovascular Society Guidelines for the Management of Dyslipidemia for the Prevention of Cardiovascular Disease in the Adult. *Can. J. Cardiol.* 2016, *32*, 1263–1282. [CrossRef] [PubMed]
- 159. Grundy, S.M.; Stone, N.J.; Bailey, A.L.; Beam, C.; Birtcher, K.K.; Blumenthal, R.S.; Braun, L.T.; de Ferranti, S.; Faiella-Tommasino, J.; Forman, D.E.; et al. 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/ADA/AGS/APhA/ASPC/NLA/PCNA Guideline on the Management of Blood Cholesterol: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *Circulation* 2019, 139, e1082–e1143. [CrossRef]
- Mach, F.; Baigent, C.; Catapano, A.L.; Koskinas, K.C.; Casula, M.; Badimon, L.; Chapman, M.J.; De Backer, G.G.; Delgado, V.; Ference, B.A.; et al. 2019 ESC/EAS Guidelines for the management of dyslipidaemias: Lipid modification to reduce cardiovascular risk. *Eur. Heart J.* 2020, *41*, 111–188. [CrossRef] [PubMed]