

invertiaDB: a database of inverted repeats across organismal genomes

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Abstract

Inverted repeats are repetitive elements that can form hairpin and cruciform structures. They are linked to genomic instability; however, they also have various biological functions. Their distribution differs markedly across taxonomic groups in the tree of life, and they exhibit high polymorphism due to their inherent genomic instability. Advances in sequencing technologies and declined costs have enabled the generation of an ever-growing number of complete genomes for organisms across taxonomic groups in the tree of life. However, a comprehensive database encompassing inverted repeats across diverse organismal genomes has been lacking. We present invertiaDB, the first comprehensive database of inverted repeats spanning multiple taxa, featuring repeats identified in the genomes of 118 101 organisms across all major taxonomic groups. For each organism, we derived inverted repeats with arm lengths of at least 10 bp, spacer lengths up to 8 bp, and no mismatches in the arms. The database currently hosts 34 330 450 inverted repeat sequences, serving as a centralized, user-friendly repository to perform searches and interactive visualizations, and download existing inverted repeat data for independent analysis. invertiaDB is implemented as a web portal for browsing, analyzing, and downloading inverted repeat data. invertiaDB is publicly available at <https://invertiadb.netlify.app/homepage.html>.

Graphical abstract



Introduction

The right-handed double helix DNA structure, also known as the canonical B-DNA structure, was originally described by Watson, Crick, Wilkins, and Franklin. Several alternative

DNA conformations have since been identified [1–3]. These noncanonical DNA structures that do not conform to the standard B-DNA form are termed non-B DNA. Such conformations include hairpin and cruciform structures, which can

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form at inverted repeat (IR) sequences [4–7]. IRs are composed of two sequence parts, one of which is the reverse complement of the other, separated by an intervening spacer sequence. IRs undergo intrastrand base pairing to form hairpin or cruciform structures, in which the two complementary arms hybridize, and the spacer loop remains single-stranded.

The formation of hairpin and cruciform structures can be facilitated by negative supercoiling, often associated with DNA transcription and replication [8, 9]. The biophysical properties of IRs, including the spacer and arm lengths and their GC content, influence the likelihood of structure formation. Increased GC content in the IR arms is linked to increased hairpin stability [5, 10, 11], whereas mismatches in the arms reduce the likelihood of structure formation [12–16]. Additionally, longer arms are linked to increased hairpin formation likelihood and stability [10], while shorter spacer lengths are favorable to hairpin and cruciform structure formation [10]. Specifically, cruciforms favor shorter spacer lengths, whereas hairpins favor short spacers but with spacer lengths above 4 bp due to steric constraints [10, 17].

IRs are over-represented in organismal genomes relative to the expected random distribution and show an inhomogeneous genomic distribution with enrichment hotspots [18]. They are particularly enriched in promoters and near transcription termination sites and can regulate gene expression [19, 20]. In prokaryotes, IRs can drive rho-independent transcription termination [21, 22]. At replication origins, IRs are involved in replication initiation [23], and in certain transposons, they are found flanking the internal sequence [24]. Additionally, several proteins can bind preferentially to hairpin and cruciform structures [25].

Hairpins and cruciforms are associated with increased genomic instability in both prokaryotes and eukaryotes [6, 15, 19, 26–35]. The human genome is depleted of perfect IRs with long arms, which are highly unstable [5, 36, 37]. IRs are recombination and rearrangement hotspots across organisms [29, 30], and artificial introduction of IRs with long arms in eukaryotic cells leads to thousands of times higher recombination and deletion rates than expected [15]. In human cancers, IRs are highly enriched in somatic mutations across mutation categories, and this excess can confound statistical models aimed at identifying driver mutations [35, 38, 39]. Specifically, off-target APOBEC mutagenesis is often directed at hairpin structures, causing an excess of mutagenesis at IRs, particularly at the single-stranded loop [5].

Multiple bioinformatic tools have been developed to systematically identify IRs [40–43]. Additionally, a previous database named Non-B DB encompasses IRs found in five organisms [44]. The recently published LIRBase is a comprehensive long IR database across 424 eukaryotic genomes and also provides information about expression levels, and serves as a valuable resource for exploring long IRs [45]. msRepDB is a database dedicated to microsatellite repeats that provides detailed annotations of repetitive elements across genomes, offering insights into the broader repeat landscapes within which IRs occur [46]. Additional resources, such as Repbase and Microsatellites Explorer, focus on repetitive DNA elements associated with transposable elements and satellite DNA across organismal genomes [47, 48].

Here, we present invertiaDB, an IR database across organismal genomes. The database currently hosts 34 330 450 IR sequences found across 118 101 organismal genomes and serves as a centralized, user-friendly web portal to perform searches

and interactive visualizations, and download existing IR data for further analysis. This resource will be valuable for researchers across various disciplines, studying the functional roles and genomic instability of IRs.

Materials and methods

Data collection

The organismal genomes present in the database were downloaded from the GenBank and RefSeq databases on 21 March 2024 [49, 50] and included all complete organismal genomes. We did not consider incomplete genomes, scaffolds, and contigs in order to increase the confidence in our identified sequences. Duplicate assembly accessions were filtered out. The organism names and accession IDs are provided in [Supplementary Data S1](#). Gene annotation files in the form of GFF files were downloaded for each genome from the same source using genome_updater version v0.6.4 from https://github.com/pirovc/genome_updater [51]. Coordinates for genic regions were derived using bash and Python scripts. The intersection with IR coordinates and the subsequent determination of the nearest genic region were extracted using BEDTools intersect and BEDTools closest commands [52]. IR densities were calculated as the ratio of total length of IR sequences to the genome size of the assembly accession, multiplied by 1 Mb. These coordinates were then examined for IR densities across organismal genomes. The organismal genome database consists of 49 197 bacteria, 67 742 viruses, 492 Eukaryota, and 687 Archaea.

Identification of IRs in organismal genomes

IRs with arm lengths ≥ 10 bp, spacer lengths < 9 bp, and without mismatches in the arms were obtained from [53]. For IR detection, a modified version of the non-B gfa package was developed and wrapped in a Python program [44]. The subprocess call to the C script utilized all necessary skip flags to extract only the IR sequences present in each organismal genome, and we provide the command below:

```
./non-B_gfa/gfa -seq < input_fasta> -out < output> -
skipAPR -skipSlipped -skipCruciform -skipTriplex -
skipWGET -minIRrep 10 -maxIRspacer 8 -skipMR -
skipDR -skipZ -skipGQ -skipSTR
```

A custom script was used to manually transform the data into a processable tabular format and extract the corresponding arm and spacer sequences along with their corresponding lengths based on the coordinates of the IR sequence. The IR spacer and arm thresholds align with previous observations that shorter arms and longer spacers are less prone to forming IRs and that mismatches in the hairpin arms and increased spacer lengths further reduce the probability of stable structure formation [12–16]. We systematically validated the loci and sequences of the detected IRs in order to ensure their correctness.

IR density across temperature classes

We separated bacteria strains into four mutually exclusive groups, as a function of their average optimal growth temperature (Topt_ave): psychrophiles, mesophiles, thermophiles, and hyperthermophiles, in accordance with the TEMPURA database [54]. In particular, we classified a bacterial strain as psychrophile, if the optimal growth temperature

is below 20°C; mesophile, if the optimal growth temperature is at least 20°C and at most 44°C; thermophile, if it is at least 45°C and at most 79°C; and finally, hyperthermophile, if the optimal growth temperature is at least 80°C. In order to utilize the results from TEMPURA DB, we mapped the NCBI Taxonomy ID corresponding to the average optimal growth temperature to the RefSeq or GenBank assembly accession IDs, as derived from the assembly summaries of the latter genomic databases, using the species taxonomy ID column. After removing duplicated assembly accessions between RefSeq and GenBank databases, a total of 3894 bacterial genomes were analyzed using the TEMPURA database. Subsequently, we compared the derived IR densities for each individual bacterial assembly accession to the respective temperature classes. The IR density for each individual genome was calculated as the ratio of the total number of IRs to the genome size multiplied by 1 Mb. If a genome lacked IRs, we set an IR density of 0. Finally, to calculate the IR arm GC-fold enrichment for each individual bacterial strain, we computed the GC proportion of all IR arms in each strain and compared it to the GC content of the underlying genome, as derived from the assembly summaries in RefSeq and GenBank databases. For all the pairwise comparisons between the four temperature groups, we used independent *t*-tests and one-way ANOVA as provided from the SciPy Python module. All the tests were corrected for multiple hypothesis testing using the Benjamini–Hochberg procedure.

Database design and data pipelines

The backend of the application is built using Flask, a Python-based web framework, and is served through a reverse proxy to manage internal port access. The data layer leverages DuckDB in read-only mode, optimizing for fast, highly compressed, and secure online analytical processing. The DuckDB database driver operates in-process within the Flask app's memory, with all data consolidated into a single DuckDB file. Currently, the stored IRs occupy 3 GB, organized as a vectorized columnar database, accessible through SQL syntax. As of database design, metadata were extracted from the NCBI database for each of the 118 070 assembly genomes that were analyzed for IRs, and enriched with the IR filename to facilitate joint operations between the two. The term “enriched” in this context refers to the process of combining data from two separate tables, the “inverted_repeats” table and the “metadata” table, to create a single, more comprehensive dataset. Then, using joins between the metadata and the IR genomic data, the metadata table was enriched with the IR statistics (Supplementary Fig. S1). We used various scripts to curate the data and to convert them to multiple formats (Parquet, CSV, JSON, and BED), while grouping them into the three domains of life and viruses and providing them for user download.

Web interface

The frontend of invertiaDB is built using HTML, CSS, and JavaScript. The application is deployed on a standard Google Cloud Compute Engine, featuring a JavaScript-based frontend app. A comprehensive full-stack web application was developed to enable user-friendly access, in-depth analysis features, and visualization of the integrated data. To provide in-depth understanding of the IR data, dynamic and interactive graphs were also created using custom HTML and Chart.js. Addi-

tionally, pre-built components from the Bootstrap 5 and Bootstrap 5 DataTables libraries were utilized.

Database overview and functionality

Database contents and usage

invertiaDB features comprehensive integration and curation of IRs in 118 101 assemblies of organisms in the domains of life and viruses, in a user-friendly, interactive web interface (Fig. 1A and B). Out of all the examined viral genomes, 32 874 did not contain any IRs, while only 1 bacterial genome did not contain any IR sequences. Interestingly, viral genomes displayed greater variability in their genome-wide IR density, while bacterial genomes were found to have the highest average IR density. IR sequences can be filtered based on their biophysical properties, including their spacer and arm lengths, sequence motifs that the user searches for, and IR metadata such as density, the total IR length, their nucleotide composition, and genic overlaps (Fig. 1A and B). The database includes multiple features to explore, search, and download IRs from each organism in Parquet, CSV, JSON, and BED formats.

The top navigation bar is composed of six interactive pages, namely the Homepage, the Explore page, the Downloads page, the Help page, the Motif page, and the About/Contact page tabs, which enable the navigation across the different parts of the database. The Explore page is further split into the invertiaDB dataset, the Domains page, and the Organisms page, with each of these options navigating the user to subsequent pages for IR explorations. The Domains page provides aggregations of IRs by the domain of life, in Archaea, Eukaryota, bacteria, and viruses.

invertiaDB dataset page

Upon accessing the Explore → invertiaDB Dataset page, users are presented with a table of assemblies featuring advanced querying capabilities for exploring IRs across various genomes (Fig. 2A). Quick searches can be performed on NCBI Taxonomy IDs, GenBank/Reference genome accessions, or species names. Complex queries can be performed using combinations of the available metadata columns provided by the Advanced Filters feature, and users may project all columns from the NCBI database metadata. Multiple genomes can be selected for downloading IR annotations in Parquet, CSV, JSON, or BED formats. Upon clicking the assembly button that is shown as the first column, the user navigates to an analysis of the assembly displaying the IR and genome metadata (Fig. 2B). The metadata include links to the established publicly available databases of the ENA Browser [55] and the NCBI Genome Browser [56]. We generated a visualization depicting the occurrences of IRs in the organism, focusing on the arm-to-spacer ratio. This allows for an assessment of whether the structure is arm- or spacer-dominant, as some organisms exhibit longer spacer/loop lengths relative to arms/stems.

Analysis and visualization pages

The invertiaDB website features interactive bar plots, doughnut charts, tables, and drop-down menus, allowing users to select and analyze IRs across various assemblies. On the assembly analysis page on inverted repeats, the user also encounters a line chart displaying the length distribution of IRs per arm, spacer, and sequence (Fig. 3A), along with three doughnut graphs that showcase respective nucleotide compositions for the three sequences, offering insight into the structural

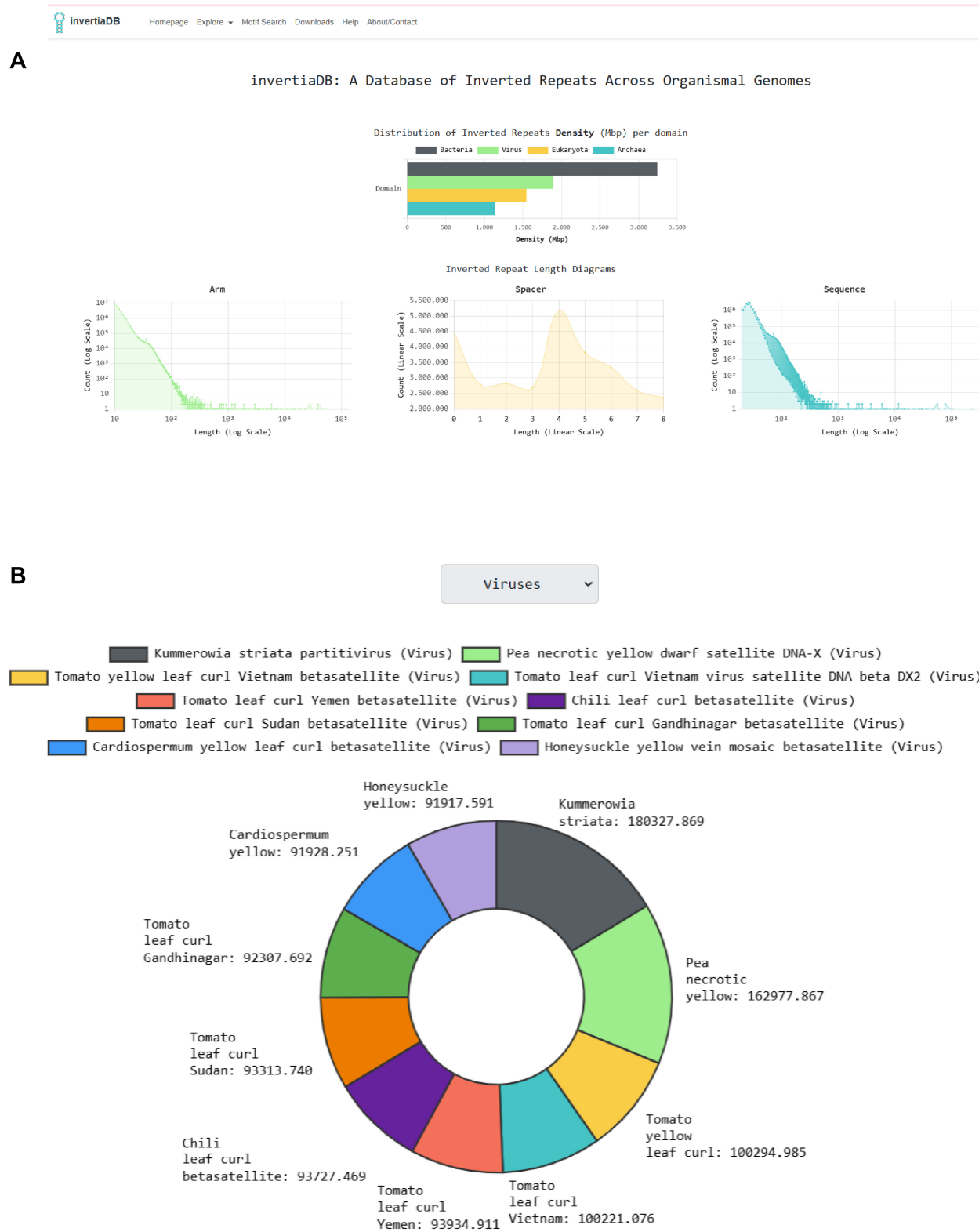
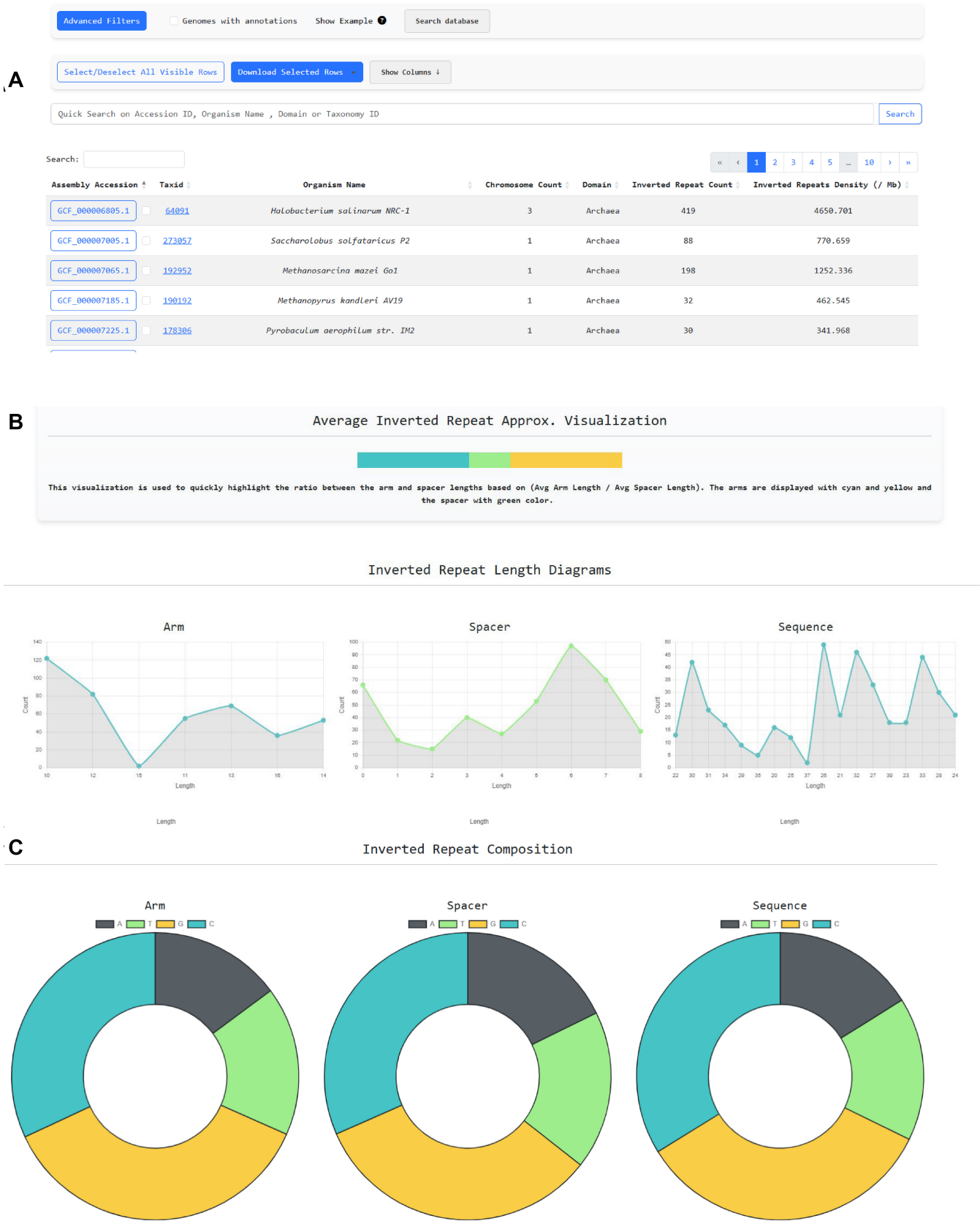


Figure 1. invertiaDB Homepage, navigation bar, and domain selection. **(A)** invertiaDB Homepage showing domains of life bar chart in decreasing order of IR density, and arm, spacer, and sequence length distribution in line charts. IR densities were calculated as the ratio of the total length of IR sequences to the genome size of the assembly accession, multiplied by 1 Mb. **(B)** Doughnut chart of top 10 most dense organisms in IRs for each domain of life.



Inverted Repeat Composition

Arm

Spacer

Sequence

Figure 2. invertiaDB dataset Explore page along with example usage and assembly analysis pages. **(A)** invertiaDB dataset Explore page showcasing Advanced Filtering, Downloading, Projecting NCBI Columns, Quick Search, and Row Filtering features. **(B, C)** Assembly accession analysis page, containing metadata and visualizations of genome and IR patterns.

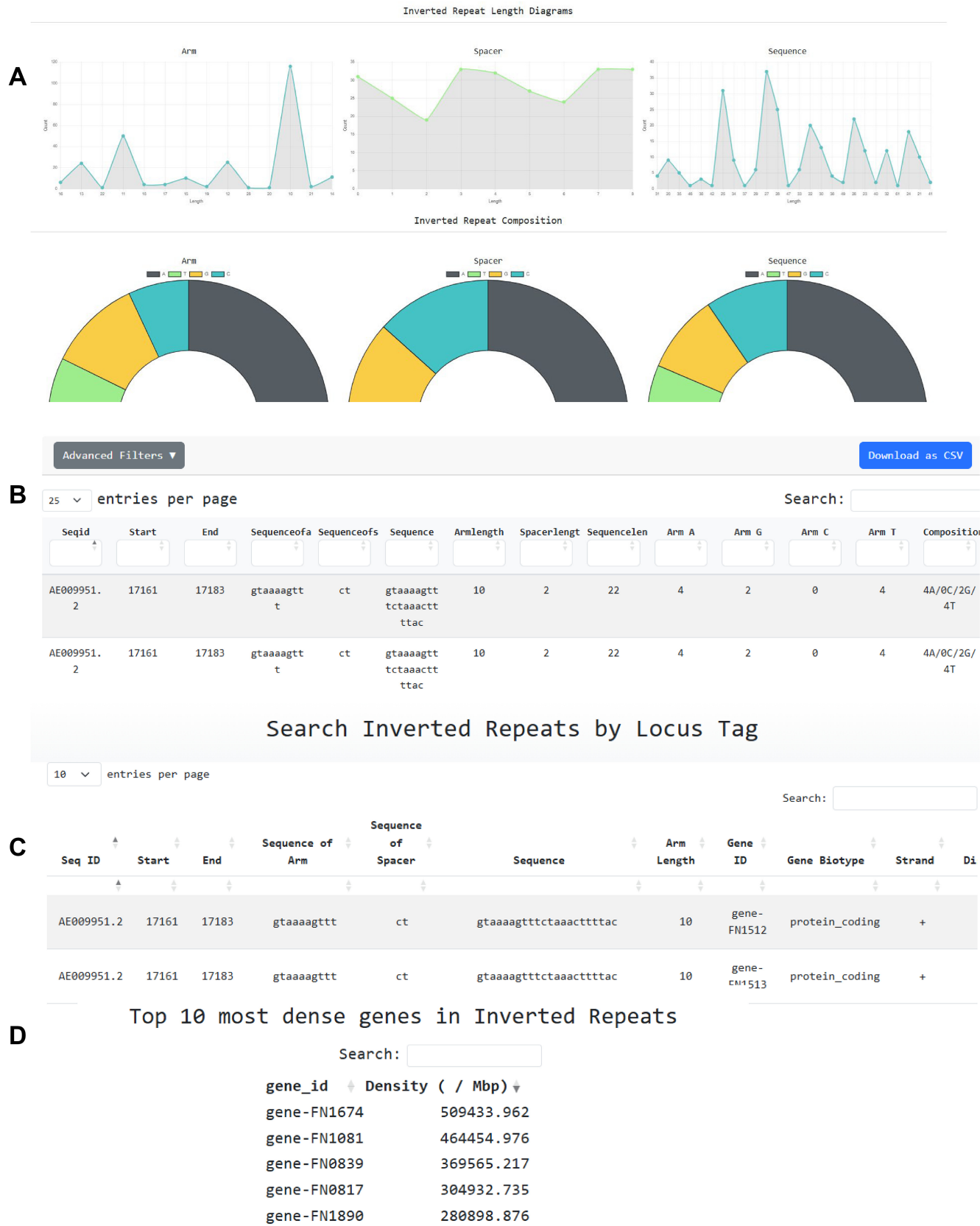


Figure 3. Assembly accession analysis page showing IR graphs, raw data tables, and gene functionalities. **(A)** Line chart of length distribution of arm, spacer, and sequence along with nucleotide composition of arm, spacer, and sequence. **(B)** Table of IRs contained in the organism's assembly with the ability to fuzzy search each column. **(C)** Search IRs overlapping specific genes by gene locus tag. **(D)** Table of most dense genes in IRs by formula: total IR base pairs overlapping a gene over gene size in Mbp.



Motif search

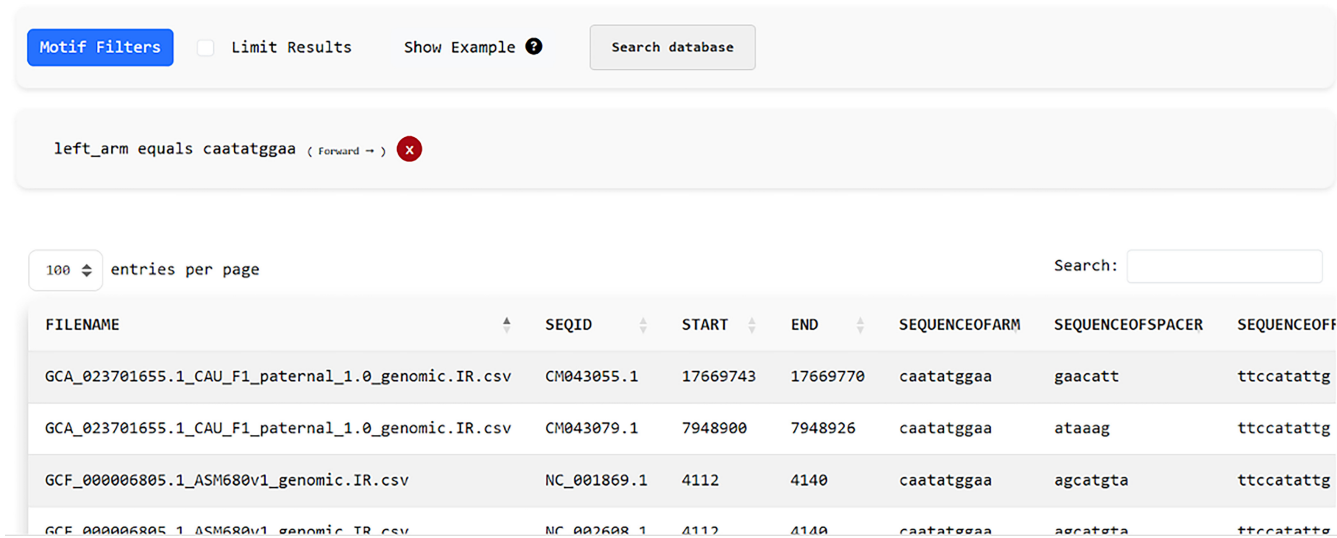


Figure 5. Motif search capabilities. Example of motif search in the left arm using a single equality filter.

characteristics (Fig. 3A). A table listing the IRs can be found within the organism’s assembly, with the ability to apply fuzzy search across various columns, enabling filtering of the IR data. The user can search by exact chromosome and genomic coordinate ranges, as well as by arm, spacer, sequence length, and GC percentage (Fig. 3B). Finally, there are gene-specific analyses with two functionalities: (i) the ability to search for IRs that overlap specific genes by gene locus tag (Fig. 3C) and (ii) a table identifying genes with the highest IR density, calculated by dividing the total size of IRs overlapping the gene by the gene’s size in megabase pairs (Mbp) (Fig. 3D). By default, the top 10 genes with the highest intragenic IR density are displayed in IR bp per Mb.

Domains and Organisms pages

By accessing the Explore → Domains page, the user is redirected to the aggregated IR data in the three domains and viruses. When the user selects a specific domain or virus, they are presented with the statistics of average IR count and density across the assemblies, the average spacer, arm, and sequence length across assemblies and for individual assemblies in a searchable table (Fig. 4A). Upon accessing the Explore → Organisms page, the results of the invertiaDB dataset are arranged by organism name instead of assembly ID (Fig. 4B). The user can analyze and compare the IR data for multiple assemblies of the same organism when multiple assemblies are present (Fig. 4B and C). Additionally, by navigating to the individual organism’s page, aggregated IR statistics and metadata are provided, and the user has the option to download these in different file formats (Parquet, CSV, JSON, or BED formats) (Fig. 4C).

Motif search options

The Motif search page enables the search of motifs within IRs or their sub-compartments. There are multiple options to customize the search; these include the following: (i) String sequence search using standard string filters. The standard

string operations that can be used are equals, contains, starts with, and ends with, and they can be applied sequence-wise or granularly on specific regions such as left/right arm, spacer, and both arms. (ii) String length search implementing different comparators, including “equals,” “greater than,” “less than,” “greater or equal,” and “less or equal.” These filters can also be applied on different IR regions (Fig. 5). After the search filters are applied, a metadata search is performed to fetch the different files and the unique organisms that the returned results belong to. (iii) Users can also search on the forward, reverse, and both strands of the sequences, as well for advanced filtering. (iv) There is also the added option to limit the number of results displayed.

Results

High optimal growth temperature of bacteria is linked to low IR density

We investigated the relationship between the optimal growth temperature of bacterial species and differences in IR density as well as their biophysical properties. We separated bacteria into four groups, psychrophiles, mesophiles, thermophiles, and hyperthermophiles, in accordance with the TEMPURA database [54, 57] and examined the IR density in each group. We reported that the IR density is negatively correlated with the optimal growth temperature of bacterial species, with psychrophilic bacteria having the largest number of IR sequences per Mb, with a median of 139.20 IRs per Mb, followed by mesophilic, thermophilic, and hyperthermophilic bacteria (Fig. 6A and Supplementary Table S1). To further investigate this hypothesis, we performed one-way ANOVA comparison of hairpin IR densities across the four temperature classes, which yielded statistically significant differences between the temperature groups ($P < .0001$). This result indicates that low temperatures result in increased frequency of IRs.

Next, we examined the biophysical properties of the IRs across the four temperature groups. Longer IR arm length

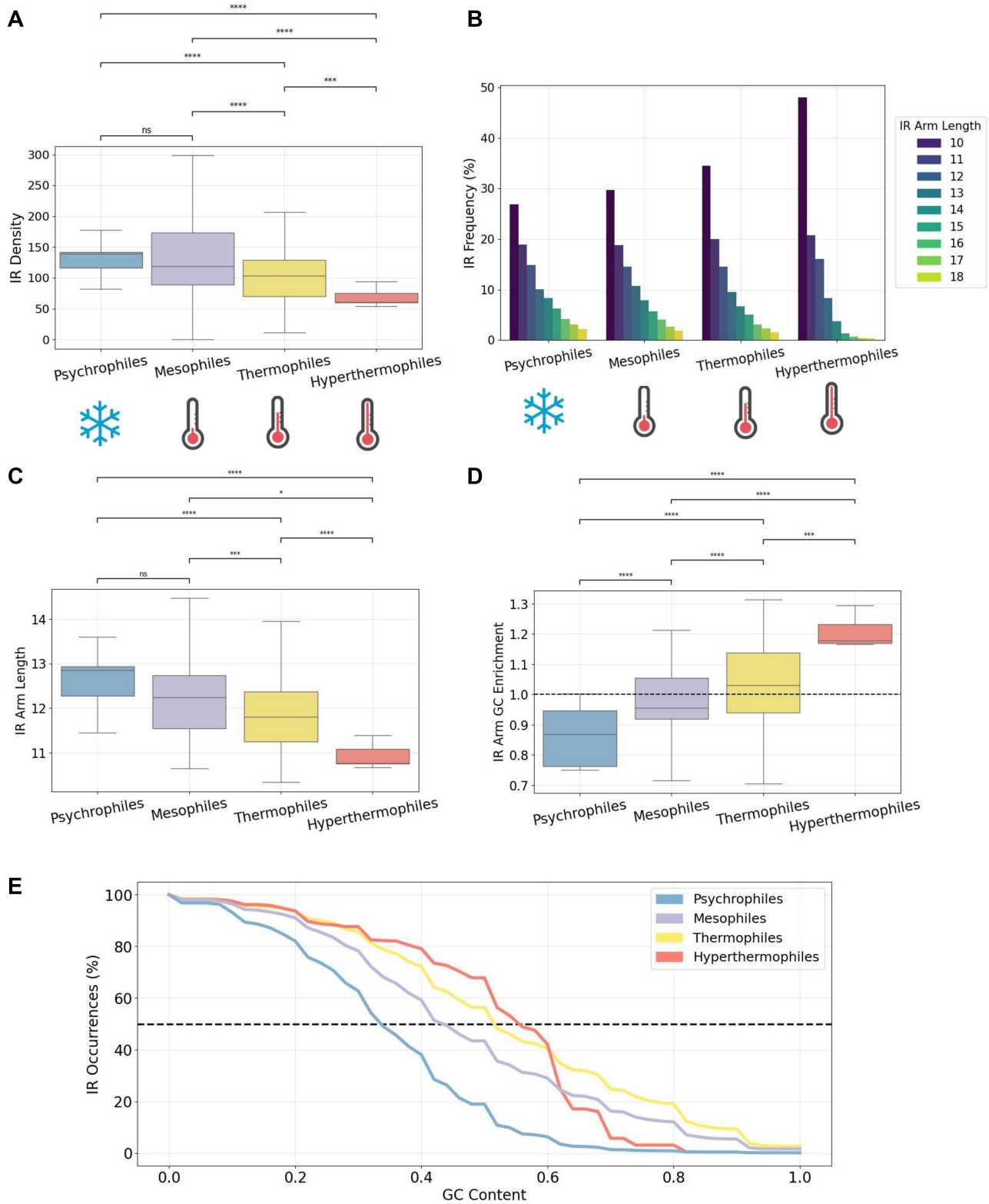


Figure 6. Relationship of hairpins to bacterial optimal growth temperature. **(A)** IR density per Mb as a function of the optimal growth temperature class of bacterial growth. **(B)** IR frequency distribution (%) across each optimal growth temperature class partitioned on IR spacer length. **(C)** Average IR arm length as a function of the optimal growth temperature class of bacterial growth. **(D)** GC enrichment of IR arm as a function of the optimal growth temperature class of bacterial growth. **(E)** Drop rate (%) of total IR sequences present in each growth temperature partition class as a function of the IR arm total GC content.

is linked to more likely hairpin formation [4, 16]. Upon examining the IR arm length proportions within each respective temperature class, we noticed that IRs with shorter arms tend to be more abundant in bacterial strains thriving in higher growth temperatures (Fig. 6B). To further evaluate this observation, we compared the distributions of IR arm length as a function of the bacteria temperature class. We report that psychrophilic bacteria exhibited a propensity for larger IR arms, with a median of 12.9 bp IR arm length, whereas hyperthermophilic bacteria had a median of 10.8 bp IR arm length, indicating that the average IR arm length is anticorrelated with optimal growth temperature (Fig. 6C). IR arms with high GC content are more likely to form stable hairpin structures [5]. We investigated the GC-content enrichment of IR arms in relationship to the GC-content background ratio in the genome of each individual bacterial species. Our results indicate that IR sequences have higher arm GC content than expected in bacteria thriving in higher temperatures, with hyperthermophilic bacteria exhibiting an average 1.15-fold enrichment, whereas mesophilic and psychrophilic bacteria displayed a below-average enrichment in GC-content arm composition, with the latter displaying an average of 0.86-fold enrichment (two-tailed *t*-test with multiple testing adjusted *P*-value; *P*-value < .001 in all comparisons) (Fig. 6D and Supplementary Fig. S2B). We conclude that higher optimal growth temperature is linked to lower IR density, and when these IRs are present in organisms that prefer higher growth temperature, they have shorter arms with higher GC content.

Discussion

Here, we have developed invertiaDB, the first centralized and comprehensive IR database spanning all major taxa across the tree of life. The website is user-friendly and interactive, and provides various features, including search and filtering tools, dynamic tables that allow for querying and sorting, visualizations, and data downloads for in-depth, independent analysis. As the number of available organismal genomes continues to grow, we plan to incorporate them in our database with regular updates.

IRs are a highly dynamic DNA element that can form hairpin and cruciform structures. They are associated with a plethora of functions, but are also linked to increased genomic instability [15, 19, 26–35]. The invertiaDB database enables the systematic examination of the functional roles of IRs, including those in gene regulation, replication, transposition, genome organization, and gene and genome evolution, among others [20, 58, 59]. The integration of biophysical properties, including the spacer and arm lengths, and the nucleotide composition of each IR allows for the study of these parameters and the impact of IR stability on its functional roles and genomic instability. Finally, our database could be utilized by researchers who are interested in examining DNA repair systems associated with hairpin and cruciform structures, which often differ between organisms belonging to different taxonomies [60, 61].

Our findings highlight a relationship between bacterial growth temperature and IR density across the studied bacterial genomes. Specifically, the reduced IR density in thermophiles and hyperthermophiles suggests a selective pressure to minimize DNA secondary structures that could destabilize genomic integrity under thermal stress. The higher GC

content and shorter arm lengths observed in the IRs of high-temperature bacteria further underscore the role of sequence composition and structure in maintaining DNA stability in extreme environments. These adaptations may represent evolutionary strategies to mitigate the risk of DNA damage or replication hindrance at elevated temperatures. In contrast, the higher IR density and longer arms with lower GC enrichment in psychrophiles and mesophiles may reflect a lower selective pressure against IR formation, potentially enabling diverse regulatory roles, such as Rho-independent transcription termination [62], for IRs at lower temperatures. Overall, these findings provide new insights into how environmental conditions, such as temperature, shape genomic architecture through selective pressures on repetitive DNA elements.

One limitation of invertiaDB is excluding imperfect IRs. There are two reasons for this: (i) the search space, computational resources, and number of detected imperfect IRs across 118 101 are prohibitively large and would require large computational resources, while imperfect IRs are less likely to form hairpins and cruciforms [12, 16]. Another limitation of our study is its reliance on computational predictions without experimental validation, which may introduce biases or inaccuracies, including sequencing errors. Experimental research will be essential to validate the biological relevance and functionality of the identified IRs when utilizing our database across research projects.

Maintaining invertiaDB in sync with the continual updates to genome assemblies at NCBI poses significant challenges, including managing the integration of new assemblies and accommodating revisions to existing ones. To address these challenges, we plan to implement pipelines that retrieve and analyze updated genome data every 12 months, ensuring that the database remains comprehensive and current. Future efforts will also focus on developing a version-controlled framework to track changes, improve scalability, and support community-driven contributions to enhance the database's utility for diverse research applications.

We envision that invertiaDB will be adopted by researchers around the world to explore the roles and applications of IRs. invertiaDB will enable the systematic exploration of IRs and advance our understanding of their broader roles and impact on organismal evolution and biological functions.

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ing [equal]), Georgios A. Pavlopoulos (Project administration [equal], Supervision [equal], Writing—review & editing [equal]), and Ilias Georgakopoulos-Soares (Conceptualization [lead], Funding acquisition [lead], Investigation [lead], Methodology [lead], Project administration [lead], Resources [lead], Software [supporting], Supervision [lead], Visualization [supporting], Writing—original draft [lead], Writing—review & editing [lead])

Supplementary data

Supplementary data is available at NAR online.

Conflict of interest

None declared.

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Data availability

The invertiaDB dataset can be found in Zenodo with a stable version at <https://zenodo.org/records/13856709>. The GitHub code for the extraction of IRs, filtering, and analyses is found at <https://github.com/Georgakopoulos-Soares-lab/invertiaDB-analysis>.

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