





## First Insights into the Draft Genome Sequence of the Endophyte *Paenibacillus amylolyticus* Strain GM1FR, Isolated from *Festuca rubra* L.

📵 Anja Poehlein, a Jacqueline Hollensteiner, a Sandra Granzow, b Bernd Wemheuer, a Stefan Vidal, b Franziska Wemheuer a, b

<sup>a</sup>Genomic and Applied Microbiology and Göttingen Genomics Laboratory, Institute of Microbiology and Genetics, University of Göttingen, Göttingen, Germany

**ABSTRACT** Paenibacillus amylolyticus strain GM1FR is an endophyte isolated from aerial plant tissues of Festuca rubra L. Here, we report the draft genome sequence (7.3 Mb) of GM1FR containing 6,241 protein-coding genes, some of which are potentially involved in plant growth promotion and biocontrol.

Several strains of the genus *Paenibacillus* from the plant endosphere are known as plant growth-promoting bacteria (1, 2). They are able to produce plant growth-regulating substances such as cytokinin (3) and indole-3-acetic acid (4). In addition, some *Paenibacillus* species act as biocontrol agents against various important phytopathogens and pests (1, 2). We sequenced the genome of the endophyte *Paenibacillus* sp. GM1FR to determine its potential as a biocontrol agent.

Paenibacillus amylolyticus strain GM1FR was isolated from surface-sterilized aerial tissues of healthy Festuca rubra L. plants. Genomic DNA was extracted using the MasterPure complete DNA purification kit (Epicentre, Madison, WI, USA). The obtained DNA was used to generate Illumina shotgun paired-end sequencing libraries. Sequencing was performed employing the MiSeq system with the MiSeq reagent kit version 3 (600 cycles) as recommended by the manufacturer (Illumina, San Diego, CA, USA). Quality filtering using Trimmomatic version 0.32 (5) resulted in 3,268,102 paired-end reads. The de novo genome assembly was performed with the SPAdes genome assembler version 3.8.0 (6). The assembly resulted in 67 contigs (>500 bp) and an average coverage of 92-fold. The assembly was validated and the read coverage was determined with QualiMap version 2.1 (7). Gene prediction and annotation were performed using Prokka (rapid prokaryotic genome annotation) version 1.11 (8).

The draft genome of strain GM1FR consisted of 7,281,281 bp with an overall GC content of 45.47%. It harbored 11 rRNAs, 99 tRNAs, and 6,241 protein-coding genes, including 2,454 genes with functional annotation. A phylogenetic analysis based on multilocus sequence typing using four genes (*gapA*, *groEL*, *gyrA*, and *pgi*) (9) revealed that strain GM1FR clusters with the species *P. amylolyticus* (10).

A total of 58 potential gene clusters involved in secondary metabolite production were identified using antiSMASH version 3.0.5 (11). The majority of these clusters showed no or weak similarity to known clusters. Three putative nonribosomal peptide synthetase (NRPS) gene clusters were identified. One cluster with 62% of the genes exhibited similarity to a pelgipeptin biosynthetic gene cluster. Pelgipeptin exhibits antimicrobial activity against many pathogenic fungi and bacteria (12, 13). A lassopeptide gene cluster with 40% of the genes sharing similarity to a paeninodin biosynthetic gene cluster was detected. Paeninodin is pharmacologically relevant, as it provides a wide range of antimicrobial and antiviral activities (14, 15). Finally, a transAT polyketide synthase-NRPS gene cluster was identified with orthologous genes for each of the genes of a

**Received** 6 December 2017 **Accepted** 15 December 2017 **Published** 25 January 2018

**Citation** Poehlein A, Hollensteiner J, Granzow S, Wemheuer B, Vidal S, Wemheuer F. 2018. First insights into the draft genome sequence of the endophyte *Paenibacillus amylolyticus* strain GM1FR, isolated from *Festuca rubra* L. Genome Announc 6:e01516-17. https://doi.org/10.1128/genomeA.01516-17.

Copyright © 2018 Poehlein et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0

Address correspondence to Franziska Wemheuer, fwemheu@gwdg.de.

<sup>&</sup>lt;sup>b</sup>Department of Crop Sciences, Agricultural Entomology, University of Göttingen, Göttingen, Germany

Poehlein et al.

paenilarvins biosynthetic gene cluster. Paenilarvins, which are known for having strong antifungal activities, are produced by the honey bee pathogen *P. larvae* (16). However, it has not been determined if strain GM1FR has antifungal activities. The strain *Paenibacillus amylolyticus* GM1FR contains multiple gene clusters assigned to plant growth and protection as well as health promotion.

**Accession number(s).** This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number MKZL00000000. The version described here is the first version, MKZL01000000.

## **ACKNOWLEDGMENTS**

We thank the Ministry of Science and Culture of Lower Saxony and the "Nieder-sächsisches Vorab" for funding and Melanie Heinemann for technical support.

The research leading to these results has received funding from the Ministry of Science and Culture of Lower Saxony and the "Niedersächsisches Vorab" as part of the Cluster of Excellence "Functional Biodiversity Research." The funders had no role in the study design, data collection, and interpretation, or the decision to submit the work for publication.

## **REFERENCES**

- Son SH, Khan Z, Kim SG, Kim YH. 2009. Plant growth-promoting rhizobacteria, *Paenibacillus polymyxa* and *Paenibacillus lentimorbus* suppress disease complex caused by root-knot nematode and fusarium wilt fungus. J Appl Microbiol 107:524–532. https://doi.org/10.1111/j.1365-2672 .2009.04238.x.
- Grady EN, MacDonald J, Liu L, Richman A, Yuan ZC. 2016. Current knowledge and perspectives of *Paenibacillus*: a review. Microb Cell Fact 15:203. https://doi.org/10.1186/s12934-016-0603-7.
- Timmusk S, Nicander B, Granhall U, Tillberg E. 1999. Cytokinin production by *Paenibacillus polymyxa*. Soil Biol Biochem 31:1847–1852. https://doi.org/10.1016/S0038-0717(99)00113-3.
- Da Mota FF, Gomes EA, Seldin L. 2008. Auxin production and detection of the gene coding for the Auxin Efflux Carrier (AEC) protein in *Paeni-bacillus polymyxa*. J Microbiol 46:257–264. https://doi.org/10.1007/ s12275-007-0245-x.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- García-Alcalde F, Okonechnikov K, Carbonell J, Cruz LM, Götz S, Tarazona S, Dopazo J, Meyer TF, Conesa A. 2012. Qualimap: evaluating next-generation sequencing alignment data. Bioinformatics 28:2678–2679. https://doi.org/ 10.1093/bioinformatics/bts503.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- liyama K, Nishi O, Mon H, Lee JM, Kusakabe T, Asano S-I, Yasunaga-Aoki C, Shimizu S. 2013. Phylogenetic analysis of *Paenibacillus popilliae* and its

- related taxa based on housekeeping genes. J Insect Biotechnol Sericol 82:1–11.
- Akuzawa S, Nagaoka J, Kanekatsu M, Kubota E, Ohtake R, Suzuki T, Kanesaki Y. 2016. Draft genome sequence of *Paenibacillus amylolyticus* Heshi-A3, isolated from fermented rice bran in a Japanese fermented seafood dish. Genome Announc 4(2):e00218-16. https://doi.org/10.1128/ genomeA.00218-16.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–W243. https://doi .org/10.1093/nar/gkv437.
- Ding R, Wu XC, Qian CD, Teng Y, Li O, Zhan ZJ, Zhao YH. 2011. Isolation and identification of lipopeptide antibiotics from *Paenibacillus elgii* B69 with inhibitory activity against methicillin-resistant *Staphylococcus aureus*. J Microbiol 49:942–949. https://doi.org/10.1007/s12275-011-1153-7.
- Wu XC, Shen XB, Ding R, Qian CD, Fang HH, Li O. 2010. Isolation and partial characterization of antibiotics produced by *Paenibacillus elgii* B69. FEMS Microbiol Lett 310:32–38. https://doi.org/10.1111/j.1574-6968.2010 .02040.x.
- 14. Hegemann JD, Zimmermann M, Xie X, Marahiel MA. 2015. Lasso peptides: an intriguing class of bacterial natural products. Acc Chem Res 48: 1909–1919. https://doi.org/10.1021/acs.accounts.5b00156.
- Maksimov MO, Pan SJ, Link AJ. 2012. Lasso peptides: structure, function, biosynthesis, and engineering. Nat Prod Rep 29:996–1006. https://doi .org/10.1039/c2np20070h.
- Sood S, Steinmetz H, Beims H, Mohr KI, Stadler M, Djukic M, von der Ohe W, Steinert M, Daniel R, Müller R. 2014. Paenilarvins: iturin family lipopeptides from the honey bee pathogen *Paenibacillus larvae*. Chembiochem 15:1947–1955. https://doi.org/10.1002/cbic.201402139.

Volume 6 Issue 4 e01516-17 genomea.asm.org **2**