





# Understanding Efflux-Mediated Multidrug Resistance in *Botrytis cinerea* for Improved Management of Fungicide Resistance

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### **ABSTRACT**

Botrytis cinerea is a major fungal pathogen infecting over 1400 plant species. It poses a significant threat to agriculture due to multiple fungicide resistance and multidrug resistance, involves resistance to fungicides with different modes of action. Multiple fungicide resistance is mostly due to an accumulation of point mutations in target genes over time, and MDR is result from efflux (e-MDR) and metabolism (m-MDR). This review introduces the occurrence of e-MDR of *B. cinerea*, the key mechanisms, origins and management strategies of e-MDR in fields. New materials such as nanomaterials become a strategy to overcoming MDR via inhibition of ABC transporter. A deeper understanding of efflux-mediated MDR will provide a support for the MDR management of *B. cinerea* and the efficient utilization of fungicides.

### 1 | Problem Statement

Botrytis cinerea is a fungal pathogen that can infect more than 1400 plant species (Figure 1). It ranks as the second most important plant pathogen (Dean et al. 2012). Chemical control remains the most effective and efficient method in managing grey mould (Alzohairy et al. 2023). However, the development of resistance to different classes of fungicides by B. cinerea presents a significant threat to modern agriculture. Among them, one type of resistance has been described as multidrug resistance (MDR) (Sun et al. 2010), which is indicated by resistance to two or more fungicides with different modes of action (MOAs). From the perspective of mechanisms, MDR can be further categorised as that caused by efflux (e-MDR) and metabolism (m-DMR) (Kretschmer et al. 2009; Zhang and Yang 2021; Cheng et al. 2022, 2023; Wang et al. 2024), while MFR is related to Several studies have shown that B. cinerea strains, isolated from vineyards, tomato farms and strawberry farms, exhibit resistance to three or more fungicides, with a resistance frequency as high as 70% and an MDR rate reaching up to 30% (Nielsen et al. 2022; Sofianos, Samaras, and Karaoglanidis 2023). The high risk of resistance development has resulted in a growing number of previously efficacious fungicides becoming ineffective against grey mould, necessitating higher application rates and dosages, which in turn raises concerns regarding human health, environmental safety and food security.

### 2 | The Development of MDR due to Fungicide Use

Alarmingly, the occurrence of MDR in the field is often observed shortly after the introduction of some new fungicides for the control of diseases in various crops (Ishii et al. 2009; Rupp et al. 2016), such as dollar spot of lawn grasses, wheat leaf blight and apple blue mould (Song et al. 2021; Sofianos, Samaras, and Karaoglanidis 2023). *Botrytis cinerea* has developed MDR against several commonly used fungicides belonging

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**FIGURE 1** | Infection of *Botrytis cinerea* in (a) strawberry, (b) grape, and (c) tomato fruit or plant. (a) Symptoms of *Botrytis cinerea* infection in strawberry fruit grown in plantation; (b) Symptoms of disease caused by artificial inoculation of *B. cinerea* spores on the leaves of grapes grown in greenhouse; (c) Symptoms of tomato calyx infection with *B. cinerea* grown in plantation.

**TABLE 1** | Fungicides contributing to multidrug resistance in *Botrytis cinerea* in the field.

Years of release	Representative fungicides	Groups	Target protein	Years resistance reported	References
1960s	Benomyl, carbendazim	Methyl benzimidazole carbamates (MBCs)	β-tubulin	1970s	Bollen and Scholten (1971); Wang et al. (2022)
1970s	Imazalil, prochloraz	Demethylation inhibitors (DMIs)	CYP51	1970s	Stehmann and De Waard (1995); Zhang et al. (2020)
1970s	Procymidone, iprodione	Dicarboximides (DCFs)	BcOs1	1980s	Katan (1982); Adnan et al. (2018)
1980s	Diethofencarb	N-phenylcarbamate (NPC)	_	1980s	Katan, Elad, and Yunis (1989); Fan et al. (2017)
1990s	Pyrimethanil	Anilinopyrimidine (APs)	Protein kinase	1990s	Chapeland et al. (1999); Nielsen et al. (2022)
1960s	Fludioxonil	Phenylpyrroles (PPs)	Protein kinase	1990s	Faretra and Pollastro (1993); Liu et al. (2023)
1990s	Azoxystrobin	Quinone outside inhibitors (QoIs)	Cytochrome b	2000s	Markoglou et al. (2006); Harper et al. (2022)
2000s	Boscalid, fluxapyroxa	Succinate dehydrogenase inhibitors (SDHIs)	Succinate dehydrogenase	2000s	De Miccolis Angelini et al. (2010); Malandrakis et al. (2022)

to benzimidazoles, dicarboximides, demethylation inhibitors (DMIs), quinone outside inhibitors (QoIs) and succinate dehydrogenase inhibitors (SDHIs) across cultivation areas of grape, tomato and strawberry as shown in Table 1 (Kretschmer et al. 2009; Adnan et al. 2019).

### 3 | Overexpression of ABC and MFS Transporters Mediates e-MDR

The overexpression of key efflux proteins in fungi, specifically those in the ATP-binding cassette (ABC) and the major facilitator superfamily (MFS), is considered the primary cause for the emergence of low-level resistance populations in the field (Kretschmer et al. 2009; Walker et al. 2013; Hahn 2014; Sun et al. 2022). Field populations of *B. cinerea* predominantly consist of strains overexpressing these two types of

protein families Hayashi, Schoonbeek, and De Waard (2002); Kretschmer et al. (2009); Nielsen et al. (2022).

ABC efflux transporters can expel various toxic substances from the cell, playing an important role in the e-MDR of *B. cinerea* (Schoonbeek, van Nistelrooy, and de Waard 2003; Sun et al. 2022). *B. cinerea* contains 14 ABC efflux proteins (BcatrA-BcatrN), four of which are related to the MDR phenotype, namely BcatrB, BcatrD, BcatrK and BcatrO. The BcatrB protein primarily interacts with aromatic compounds (Vermeulen, Schoonbeek, and De Waard 2001; Schoonbeek, van Nistelrooy, and de Waard 2003) and has been confirmed to be involved in the resistance of *B. cinerea* to fludioxonil, dicarboximide fungicides and the phytoalexin resveratrol secreted by the host plant. Complementation of the *BcatrB* gene in *B. cinerea* mutants leads to a reduction in the accumulation of fungicides, concomitantly diminishing fenpiclonil

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and fludioxonil sensitivity compared to the knockout mutant Schoonbeek, Del Sorbo, and De Waard (2001; Kretschmer et al. (2009). The sensitivity of  $B.\ cinerea$  to oximidazole, pyrrole and DMI fungicides increases when the BcatrD gene is knocked out (Hayashi et al. 2001). Additionally, the BcatrK (BMR1) knockout strain exhibits increased sensitivity to polyoxin and iprobenfos, and the BcatrO knockout strain has an enhanced sensitivity to  $H_2O_2$  and fenpiclonil (Pane et al. 2008). Furthermore, the MDR mutants of  $B.\ cinerea$ , obtained by procymidone adaptation, exhibit a significant upregulation of BcatrG (Wu et al. 2024c). The ABC transporter genes, including BcATRO, BMR1, BMR3 and BcNMT1, are upregulated in fludioxonil-resistant strains, indicating that the overexpression of these efflux transporters may be responsible for the fludioxonil resistance in  $B.\ cinerea$  (Liu et al. 2023).

Meanwhile, MFS transporters can also expel chemical compounds from the cell. These fungal transporters are involved in the efflux of natural toxic compounds such as camptothecin, cercosporin and DMIs fungicides (Hayashi, Schoonbeek, and De Waard 2002). MFS transporter genes, including BcAMF1, BcTOP1, BcVBA2 and BcYHK8, are also upregulated in fludioxonil-resistant B. cinerea (Liu et al. 2023). MfsG transporter is a virulence factor that increases tolerance to glucosinolates (Vela-Corcia et al. 2019). A comprehensive and systematic investigation of these transporter genes and their interactions is needed in future studies.

### 4 | Regulation of ABC Expression

The expression of ABC transporter genes is modulated by factors such as promoter regions of ABC transporters and their transcription factors (Kretschmer et al. 2009). In B. cinerea, the transcription factor Mrr1 has been reported to positively regulate BcatrB expression (Kretschmer et al. 2009). The point mutations V575M in Mrr1 resulted in upregulation of BcatrB in the MDR1 strain. Additionally, the transcription factor XDR found from Sclerotinia homoeocarpa has been proved to engage the ability to regulate the expression of BcatrD and BcCYP65 in B. cinerea (Sang et al. 2018). Meanwhile, a rearrangement in the upstream region of mfsM2, caused by the insertion of a foreign gene fragment and concurrent deletion of a portion of the putative mfsM2 promoter, leads to the overexpression of mfsM2 (Kretschmer et al. 2009). Resent research has found procymidone-induced mutations on target gene BcBos1 linked to the overexpression of BcatrG in B. cinerea. Although no direct association has been identified between BcBos1 belonged to the MAPK pathway and expression of ABC transporters, the overexpressed of BcatrG expression was speculatively regulated by this regulatory pathway through BcBos1 mutation (Wu et al. 2024a, 2024c). The identification of more novel regulators or mechanisms of MDR through the xenobiotic efflux pathway in filamentous fungi may facilitate the discovery of new antifungal fungicides to control pathogenic fungi.

## 5 | Tracking the Origin of Multidrug Resistance in the Fields

To the best of our knowledge, the current understanding does not fully explain the field origin of MDR in plant pathogens. One of the laboratory techniques for inducing resistant strains involves selection of pathogens on agar media amended with fungicides, utilising a stepwise cultivation process with increasing fungicide concentrations. Our recent studies have demonstrated that the uncoupler fungicide SYP-14288 can induce MDR of Rhizoctonia solani, resulting in resistance to fluazinam, chlorothalonil, fludioxonil, difenoconazole, azoxystrobin, fentin chloride, cyazofamid and 2,4-dinitrophenol. Similarly, MDR strains of Phytophthora capsici were obtained through selection with SYP-14288, showing resistance to fluazinam, chlorothalonil and oxathiapiprolin (Cheng et al. 2020; Dai et al. 2022). Furthermore, UV irradiation-produced Phytophthora infestans mutants showed a reduced sensitivity to metalaxyl, enoylpyrazole, carbendazim, ether fungicide and antimycin A (Ziogas et al. 2006). In vitro induction of DMI fungicides led to the development of MDRs in Penicillium digitatum to DMIs, cycloheximide, 4-nitroquinoline-N-oxide and acriflavin (Nakaune et al. 1998). These studies suggest that the occurrence of MDR in plant pathogens is associated with the use of DMIs, uncouplers and UV irradiation.

Research has demonstrated that MDR in B. cinerea is not solely induced by fungicides like procymidone (Wu et al. 2024b, 2024c), but also induced by plant secondary metabolites, such as rishitin, capsidiol, camptothecin, flavanone and resveratrol (Kuroyanagi et al. 2022; Bulasag et al. 2023; Wu et al. 2024b, 2024c). Our recent work provides insight into the mechanisms underlying the role of ABC transporters in facilitating MDR induced by plant secondary metabolites (Wu et al. 2024b, 2024c). Overexpression of the ABC transporter genes, namely BcatrB, BcatrD and BcatrK, enables B. cinerea to pre-adapt to the pressure of antifungal plant secondary metabolites (Wu et al. 2024b, 2024c). This pre-adaptation leads to enhanced efflux activities for fungicides, including sterol synthesis inhibitors, osmotic signal inhibitors and methionine inhibitors. These findings suggest that ABC transporters can be stimulated by exogenous chemicals, resulting in an adjusted ability to efflux fungicides with different MOAs, playing a crucial role in the development of MDR.

### **6** | New Strategy for Management of Multidrug Resistance

Addressing the emergence of resistance in the field presents a significant challenge for future research. Various strategies based on nanomaterials, as well as physical approaches, highlight their pivotal roles in overcoming MDR (Duan et al. 2023). Specifically designed nanoparticles can bypass P-glycoprotein (P-gp)-mediated cancer MDRs by increasing surface area, encapsulating a variety of guests, and attaching targeting ligands to enhance therapeutic delivery (Xia et al. 2022; Duan et al. 2023). Previous studies have proposed innovative strategies for fungicide formulation, emphasising the rapid and targeted release of nano-carriers at the site of infestation. These approaches aim to minimise adverse effects on non-target organisms (Gao et al. 2023).

Concurrently, plant-derived phytochemicals capable of modulating various signal transduction pathways and inhibiting transcription factor translocation have been used to reverse Pgp-mediated MDRs in the medical field (Ganesan et al. 2021). These studies have explored various available opportunities for therapeutic design in human pathogens, including altered therapeutic regimens through drug combinations and P-gp inhibitors such as verapamil, dexverapamil, dexniguldipine and tariquidar (Palmeira et al. 2012; Abdallah et al. 2015; Dong et al. 2020). This concept can also be applied to the control of plant pathogens in previous research (Liu et al. 2020; Liu, Sun, Cheng, et al. 2021; Liu, Sun, Liang, et al. 2021; Prigigallo et al. 2023; Wu et al. 2024a, 2024b, 2024c). A new perspective on synergistic effects via efflux inhibition with fungicides may open opportunities to overcome MDR in plant pathogens.

### 7 | Conclusion

In conclusion, efflux pumps in *B. cinerea* play a crucial role in the development of e-MDR. Certain exogenous substances, such as plant secondary metabolites and fungicides, can upregulate the expression of these pumps through pre-adaptation or a specific pathway, subsequently triggering e-MDR to fungicides with different modes of action. Consequently, the effective regulation of these pumps represents a promising strategy to manage e-MDR in *B. cinerea*.

#### **Author Contributions**

**Zhaochen Wu:** conceptualization, writing – original draft. **Junting Zhang:** writing – original draft. **Jianjun Hao:** writing – review and editing. **Pengfei Liu:** writing – review and editing, supervision, funding acquisition. **Xili Liu:** writing – review and editing.

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#### **Conflicts of Interest**

The authors declare no conflicts of interest.

### **Data Availability Statement**

The authors have nothing to report.

### References

Abdallah, H. M., A. M. Al-Abd, R. S. El-Dine, and A. M. El-Halawany. 2015. "P-Glycoprotein Inhibitors of Natural Origin as Potential Tumor Chemo-Sensitizers: A Review." *Journal of Advanced Research* 6: 45–62.

Adnan, M., M. S. Hamada, M. Hahn, G. Li, and C. Luo. 2019. "Fungicide Resistance of *Botrytis Cinerea* From Strawberry to Procymidone and Zoxamide in Hubei, China." *Phytopathology Research* 1: 1–12.

Adnan, M., M. S. Hamada, G. Q. Li, and C. X. Luo. 2018. "Detection and Molecular Characterization of Resistance to the Dicarboximide and Benzamide Fungicides in *Botrytis cinerea* From Tomato in Hubei Province, China." *Plant Disease* 102: 1299–1306.

Alzohairy, S. A., L. Heger, N. Nikzainalalam, and T. D. Miles. 2023. "Cross-Resistance of Succinate Dehydrogenase Inhibitors (SDHI) in *Botrytis cinerea* and Development of Molecular Diagnostic Tools for SDHI Resistance Detection." *Phytopathology* 113: 998–1009.

Bollen, G. J., and G. Scholten. 1971. "Acquired Resistance to Benomyl and Some Other Systemic Fungicides in a Strain of *Botrytis cinerea* in Cyclamen." *Netherlands Journal of Plant Pathology* 77: 83–90.

Bulasag, A. S., M. Camagna, T. Kuroyanagi, et al. 2023. "*Botrytis cinerea* Tolerates Phytoalexins Produced by Solanaceae and Fabaceae Plants Through an Efflux Transporter *BcatrB* and Metabolizing Enzymes." *Frontiers in Plant Science* 14: 1177060.

Chapeland, F., R. Fritz, C. Lanen, M. Gredt, and P. Leroux. 1999. "Inheritance and Mechanisms of Resistance to Anilinopyrimidine Fungicides in *Botrytis cinerea* (*Botryotinia fuckeliana*)." *Pesticide Biochemistry and Physiology* 64: 85–100.

Cheng, X. K., T. Dai, Z. H. Hu, et al. 2022. "Cytochrome P450 and Glutathione S-Transferase Confer Metabolic Resistance to SYP-14288 and Multi-Drug Resistance in *Rhizoctonia solani." Frontiers in Microbiology* 13: 806339.

Cheng, X. K., X. J. Man, Z. T. Wang, et al. 2020. "Fungicide SYP-14288 Inducing Multidrug Resistance in *Rhizoctonia solani*." *Plant Disease* 104: 2563–2570.

Cheng, X. K., J. T. Zhang, Z. Y. Liang, et al. 2023. "Multidrug Resistance of *Rhizoctonia solani* Determined by Enhanced Efflux for Fungicides." *Pesticide Biochemistry and Physiology* 195: 105525.

Dai, T., Z. W. Wang, X. K. Cheng, et al. 2022. "Uncoupler SYP-14288 Inducing Multidrug Resistance of *Phytophthora capsici* Through Overexpression of Cytochrome P450 Monooxygenases and P-Glycoprotein." *Pest Management Science* 78: 2240–2249.

De Miccolis Angelini, R. M., W. Habib, C. Rotolo, S. Pollastro, and F. Faretra. 2010. "Selection, Characterization and Genetic Analysis of Laboratory Mutants of *Botryotinia fuckeliana (Botrytis cinerea)* Resistant to the Fungicide Boscalid." *European Journal of Plant Pathology* 128: 185–199.

Dean, R., J. A. Van Kan, Z. A. Pretorius, et al. 2012. "The Top 10 Fungal Pathogens in Molecular Plant Pathology." *Molecular Plant Pathology* 13: 414–430.

Dong, J. Y., Z. D. Qin, W. D. Zhang, et al. 2020. "Medicinal Chemistry Strategies to Discover P-Glycoprotein Inhibitors: An Update." *Drug Resistance Updates* 49: 100681.

Duan, C., M. Yu, J. Xu, B. Li, Y. Zhao, and R. K. Kankala. 2023. "Overcoming Cancer Multi-Drug Resistance (MDR): Reasons, Mechanisms, Nanotherapeutic Solutions, and Challenges." *Biomedicine & Pharmacotherapy* 162: 114643.

Fan, F., M. S. Hamada, N. Li, G. Q. Li, and C. X. Luo. 2017. "Multiple Fungicide Resistance in *Botrytis Cinerea* From Greenhouse Strawberries in Hubei Province, China." *Plant Diseases* 101: 601–606.

Faretra, F., and S. Pollastro. 1993. "Isolation, Characterization and Genetic Analysis of Laboratory Mutants of *Botryotinia fuckeliana* Resistant to the Phenylpyrrole." *Mycological Research* 97: 620–624.

Ganesan, M., G. Kanimozhi, B. Pradhapsingh, et al. 2021. "Phytochemicals Reverse P-Glycoprotein Mediated Multidrug Resistance via Signal Transduction Pathways." *Biomedicine & Pharmacotherapy* 139: 111632.

Gao, T., Zhang, Z. Wu, et al. 2023. "Fabrication of ROS-Responsive Nanoparticles by Modifying the Interior Pore-Wall of Mesoporous Silica for Smart Delivery of Azoxystrobin." *Journal of Materials Chemistry B* 11: 11496–11504.

Hahn, M. 2014. "The Rising Threat of Fungicide Resistance in Plant Pathogenic Fungi: *Botrytis* as a Case Study." *Journal of Chemical Biology* 7: 133–141.

Harper, L. A., S. Paton, B. Hall, S. McKay, R. P. Oliver, and F. J. Lopez Ruiz. 2022. "Fungicide Resistance Characterized Across Seven Modes of Action in *Botrytis cinerea* Isolated From Australian Vineyards." *Pest Management Science* 78: 1326–1340.

Hayashi, K., H. J. Schoonbeek, and M. A. De Waard. 2002. "Bcmfs1, a Novel Major Facilitator Superfamily Transporter From *Botrytis cinerea*, Provides Tolerance Towards the Natural Toxic Compounds Camptothecin and Cercosporin and Towards Fungicides." *Applied and Environmental Microbiology* 68: 4996–5004.

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- Hayashi, K., H. Schoonbeek, H. Sugiura, and M. A. De Waard. 2001. "Multidrug Resistance in *Botrytis cinerea* Associated With Decreased Accumulation of the Azole Fungicide Oxpoconazole and Increased Transcription of the ABC Transporter Gene *BcatrD*." *Pesticide Biochemistry and Physiology* 70: 168–179.
- Ishii, H., J. Fountaine, W. H. Chung, et al. 2009. "Characterisation of QoI-Resistant Field Isolates of *Botrytis cinerea* From Citrus and Strawberry." *Pest Management Science* 65: 916–922.
- Katan, T. 1982. "Resistance to 3,5-Dichlorophenyl-N-Cyclic Imide ('Dicarboximide') Fungicides in the Grey Mould Pathogen *Botrytis cine*rea on Protected Crops." *Plant Pathology* 31: 133–141.
- Katan, T., Y. Elad, and H. Yunis. 1989. "Resistance to Diethofencarb (NPC) in Benomyl-Resistant Field Isolates of *Botrytis cinerea*." *Plant Pathology* 38: 86–92.
- Kretschmer, M., M. Leroch, A. Mosbach, et al. 2009. "Fungicide-Driven Evolution and Molecular Basis of Multidrug Resistance in Field Populations of the Grey Mould Fungus *Botrytis cinerea*." *PLoS Pathogens* 5: e1000696.
- Kuroyanagi, T., A. S. Bulasag, K. Fukushima, et al. 2022. "Botrytis cinerea Identifies Host Plants via the Recognition of Antifungal Capsidiol to Induce Expression of a Specific Detoxification Gene." PNAS Nexus 1: c274.
- Liu, M., J. Peng, X. Wang, et al. 2023. "Transcriptomic Analysis of Resistant and Wild-Type *Botrytis cinerea* Isolates Revealed Fludioxonil-Resistance Mechanisms." *International Journal of Molecular Sciences* 24: 988.
- Liu, P. F., T. Dai, L. Liang, et al. 2020. "Application of Amitriptyline as a Fungicidal Synergist in the Prevention and Control of Plant Epidemics." CN110338191B.
- Liu, P. F., M. Y. Sun, X. K. Cheng, et al. 2021. "The Application of Reserpine as a Fungicidal Synergist in the Control of Plant Gray Mold Disease." CN108739835B.
- Liu, P. F., M. Y. Sun, L. Liang, et al. 2021. "The Application of Berberine as a Fungicidal Synergist in the Control of Plant Gray Mold Disease." CN112970771B.
- Malandrakis, A. A., N. Krasagakis, N. Kavroulakis, et al. 2022. "Fungicide Resistance Frequencies of *Botrytis cinerea* Greenhouse Isolates and Molecular Detection of a Novel SDHI Resistance Mutation." *Pesticide Biochemistry and Physiology* 183: 105058.
- Markoglou, A. N., A. A. Malandrakis, A. G. Vitoratos, and B. N. Ziogas. 2006. "Characterization of Laboratory Mutants of *Botrytis cinerea* Resistant to QoI Fungicides." *European Journal of Plant Pathology* 115: 149–162.
- Nakaune, R., K. Adachi, O. Nawat, M. Tomiyama, K. Akutsu, and T. Hibi. 1998. "A Novel ATP-Binding Cassette Transporter Involved in Multidrug Resistance in the Phytopathogenic Fungus *Penicillium digitatum.*" Applied and Environmental Microbiology 64: 3983–3988.
- Nielsen, K. A. G., M. N. Skårn, G. M. Strømeng, M. B. Brurberg, and A. Stensvand. 2022. "Pervasive Fungicide Resistance in *Botrytis* From Strawberry in Norway: Identification of the Grey Mould Pathogen and Mutations." *Plant Pathology* 71: 1392–1403.
- Palmeira, A., E. Sousa, M. H. Vasconcelos, and M. M. Pinto. 2012. "Three Decades of P-Gp Inhibitors: Skimming Through Several Generations and Scaffolds." *Current Medicinal Chemistry* 19: 1946–2025.
- Pane, C., D. Rekab, G. Firrao, M. Ruocco, and F. Scala. 2008. "A Novel Gene Coding for an ABC Transporter in *Botrytis Cinerea (Botryotinia fuckeliana)* is Involved in Resistance to  $\rm H_2O_2$ ." *Journal of Plant Pathology* 3: 453–462.
- Prigigallo, M. I., A. Staropoli, F. Vinale, and G. Bubici. 2023. "Interactions Between Plant-Beneficial Microorganisms in a Consortium: *Streptomyces microflavus* and *Trichoderma harzianum*." *Microbial Biotechnology* 16: 2292–2312.

- Rupp, S., R. W. Weber, D. Rieger, P. Detzel, and M. Hahn. 2016. "Spread of *Botrytis cinerea* Strains With Multiple Fungicide Resistance in German Horticulture." *Frontiers in Microbiology* 7: 2075.
- Sang, H., J. P. Hulvey, R. Green, et al. 2018. "A Xenobiotic Detoxification Pathway Through Transcriptional Regulation in Filamentous Fungi." *MBio* 9: e418–e457.
- Schoonbeek, H., G. Del Sorbo, and M. A. De Waard. 2001. "The ABC Transporter *BcatrB* Affects the Sensitivity of *Botrytis cinerea* to the Phytoalexin Resveratrol and the Fungicide Fenpicionil." *Molecular Plant-Microbe Interactions* 14: 562–571.
- Schoonbeek, H., J. G. M. van Nistelrooy, and M. A. de Waard. 2003. "Functional Analysis of ABC Transporter Genes From *Botrytis cine*rea Identifies *BcatrB* as a Transporter of Eugenol." *European Journal of Plant Pathology* 109: 1003–1011.
- Sofianos, G., A. Samaras, and G. Karaoglanidis. 2023. "Multiple and Multidrug Resistance in *Botrytis cinerea*: Molecular Mechanisms of MLR/MDR Strains in Greece and Effects of Co-Existence of Different Resistance Mechanisms on Fungicide Sensitivity." *Frontiers in Plant Science* 14: 1273193.
- Song, J., X. Cheng, P. Liu, and X. Liu. 2021. "Current Situation of Multidrug Resistance of Plant Pathogens to Fungicides." *Plant Protection* 47: 28–33.
- Stehmann, C., and M. A. De Waard. 1995. "Accumulation of Tebuconazole by Isolates of *Botrytis cinerea* Differing in Sensitivity to Sterol Demethylation Inhibiting Fungicides." *Pesticide Science* 45: 311–318.
- Sun, H. Y., H. C. Wang, Y. Chen, H. X. Li, C. J. Chen, and M. G. Zhou. 2010. "Multiple Resistance of *Botrytis cinerea* From Vegetable Crops to Carbendazim, Diethofencarb, Procymidone, and Pyrimethanil in China." *Plant Disease* 94: 551–556.
- Sun, M., Z. Wu, B. Wang, et al. 2022. "Research Status of ATP-Binding Cassette Transporters and Related Multidrug Resistance." *Journal of Plant Protection* 49: 374–382.
- Vela-Corcia, D., S. D. Aditya, A. Dafa-Berger, N. Rotem, O. Barda, and M. Levy. 2019. "MFS Transporter From *Botrytis cinerea* Provides Tolerance to Glucosinolate-Breakdown Products and Is Required for Pathogenicity." *Nature Communications* 10: 2886.
- Vermeulen, T., H. Schoonbeek, and M. A. De Waard. 2001. "The ABC Transporter *BcatrB* From *Botrytis cinerea* Is a Determinant of the Activity of the Phenylpyrrole Fungicide Fludioxonil." *Pest Management Science* 57: 393–402.
- Walker, A. S., A. Micoud, F. Remuson, J. Grosman, M. Gredt, and P. Leroux. 2013. "French Vineyards Provide Information That Opens Ways for Effective Resistance Management of *Botrytis cinerea* (Grey Mould)." *Pest Management Science* 69: 667–678.
- Wang, R., B. Chen, M. Yue, W. Ding, and Y. Li. 2022. "Multi-Resistance of *Botrytis cinerea* Isolates From Ginseng Growing Regions in China to Carbendazim, Iprodione and Pyrimethanil." *Crop Protection* 156: 105929.
- Wang, T., X. Shi, Z. Wu, et al. 2024. "Carboxylesterase and Cytochrome P450 Confer Metabolic Resistance Simultaneously to Azoxystrobin and Some Other Fungicides in *Botrytis cinerea*." *Journal of Agricultural and Food Chemistry* 72: 9680–9690.
- Wu, J., R. Xin, Y. Jiang, et al. 2024a. "Botrytis cinerea Type II Inhibitor of Apoptosis BcBIR1 Enhances the Biocontrol Capacity of Coniothyrium minitans." Microbial Biotechnology 17: e14402.
- Wu, Z., Y. Bi, J. Zhang, et al. 2024b. "Multidrug Resistance of *Botrytis cinerea* Associated With Its Adaptation to Plant Secondary Metabolites." *MBio* 15: e2223–e2237.
- Wu, Z., C. Yu, Q. Bi, et al. 2024c. "Procymidone Application Contributes to Multidrug Resistance of *Botrytis cinerea*." *Journal of Fungi* 10: 261.

Xia, H., B. Li, Y. Zhao, et al. 2022. "Nanoarchitectured Manganese Dioxide  $(MnO_2)$ -Based Assemblies for Biomedicine." *Coordination Chemistry Reviews* 464: 214540.

Zhang, C., M. Imran, M. Liu, et al. 2020. "Two Point Mutations on CYP51 Combined With Induced Expression of the Target Gene Appeared to Mediate Pyrisoxazole Resistance in *Botrytis cinerea*." *Frontiers in Microbiology* 11: 1396.

Zhang, J. J., and H. Yang. 2021. "Metabolism and Detoxification of Pesticides in Plants." *Science of the Total Environment* 790: 148034.

Ziogas, B. N., A. N. Markoglou, D. I. Theodosiou, A. Anagnostou, and S. Boutopoulou. 2006. "A High Multi-Drug Resistance to Chemically Unrelated Oomycete Fungicides in *Phytophthora infestans*." *European Journal of Plant Pathology* 115: 283–292.

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