

Supplementary Information File.

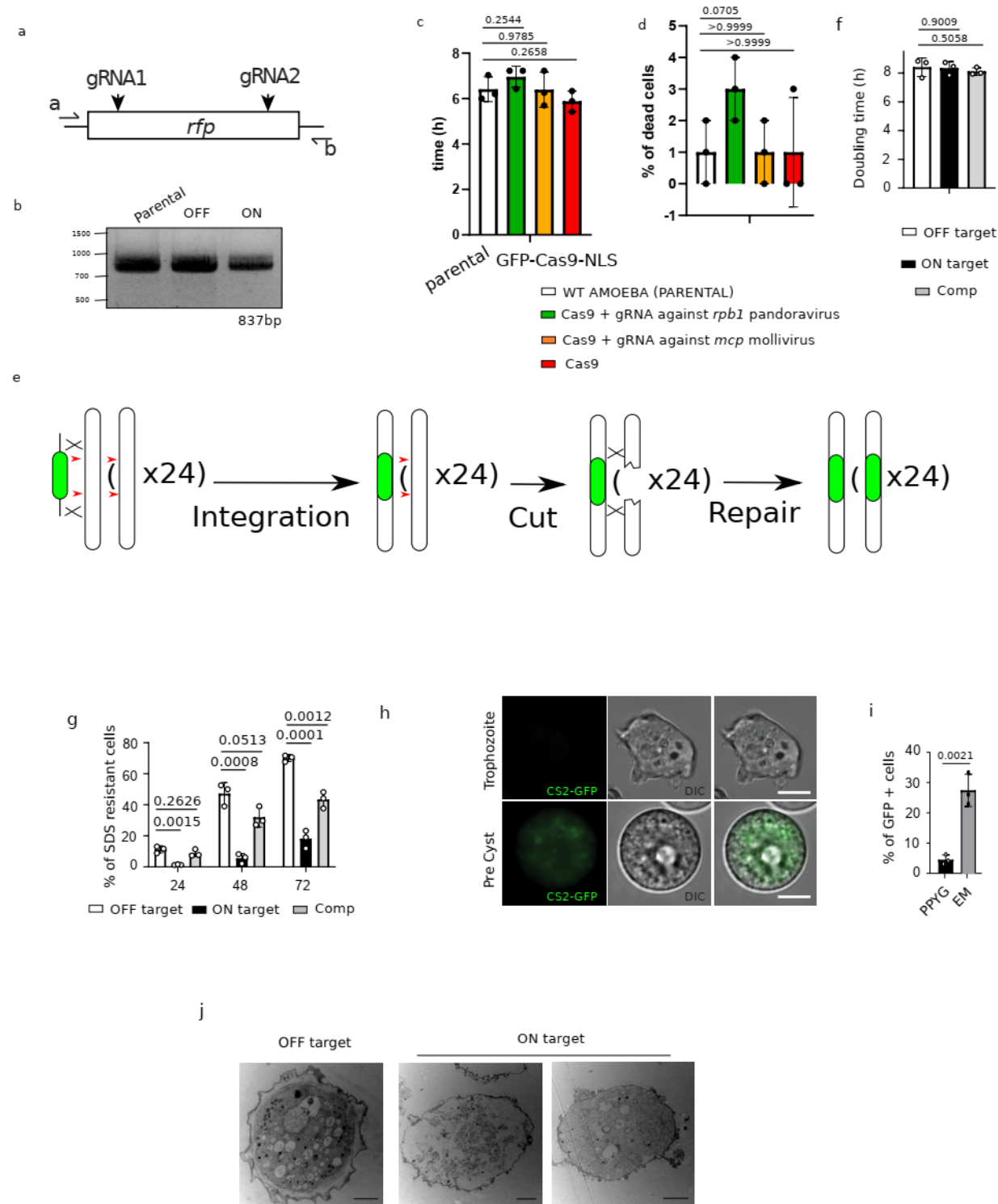


Figure S1. CRISPR/Cas9 allows manipulation of *A. castellanii*.

- (a) Schematic representation of the *rfp* locus, guide targeting location and primer annealing sites.
- (b) PCR of the genomic locus of the *rfp* shows no large deletions.
- (c) Doubling time of the wild type amoeba (parental) and amoebas harboring GFP-Cas9-NLS was calculated. Presence or absence of gRNA targeting different viruses is shown. Data correspond to the mean $\pm$ SD of 3 independent experiments.
- (d) Expression of CRISPR/Cas9 does not affect the percentage of death of *A. castellanii*. Presence or absence of gRNA targeting different viruses is shown. Data correspond to the mean $\pm$ SD of 3 independent experiments.
- (e) Schematic representation of "mutagenic chain reaction" strategy²⁰. Cas9 cleavage of the DNA at the site determined by the gRNA would allow insertion of the Cas9-gRNA cassette by homologous recombination. Continuous expression of Cas9-gRNA would allow continuous cleavage of unmodified alleles, which would propagate the Cas9-gRNA cassette in companion chromosomes if repair is performed by homologous recombination. Such strategy could facilitate the modification of the highly polyploid genome of *A. castellanii* (25n approximately)²¹.
- (f) Depletion of Cellulose synthase (CS) 1-3 does not affect the doubling time of *A. castellanii* trophozoites. Comp: complemented strain. Data correspond to the mean $\pm$ SD of 3 independent experiments.
- (g) Reduced resistance to SDS treatment was shown upon downregulation of CS. Resistance to SDS is associated to encystment and generation of a productive cyst wall. This data represents the mean and SD of 3 independent replicates. Comp: complemented strain.
- (h) The CS1-GFP is expressed upon encystment. A representative image of trophozoite and pre-cyst is shown. Scale bar: 10 μ m.
- (i) Quantification of representative pictures shown in (h). The mean $\pm$ SD of at least 200 amoebas (3 independent experiments (n=3)) is shown. PPYG: trophozoite media. EM: encystment media.
- (j) Electron microscopy shows aberrant cyst formation upon downregulation of CS. Scale bar: 2 μ m. Micrographs are representative of 3 independent experiments.

Source data are provided as a Source Data file. Micrographs are representative of 3 independent experiments.

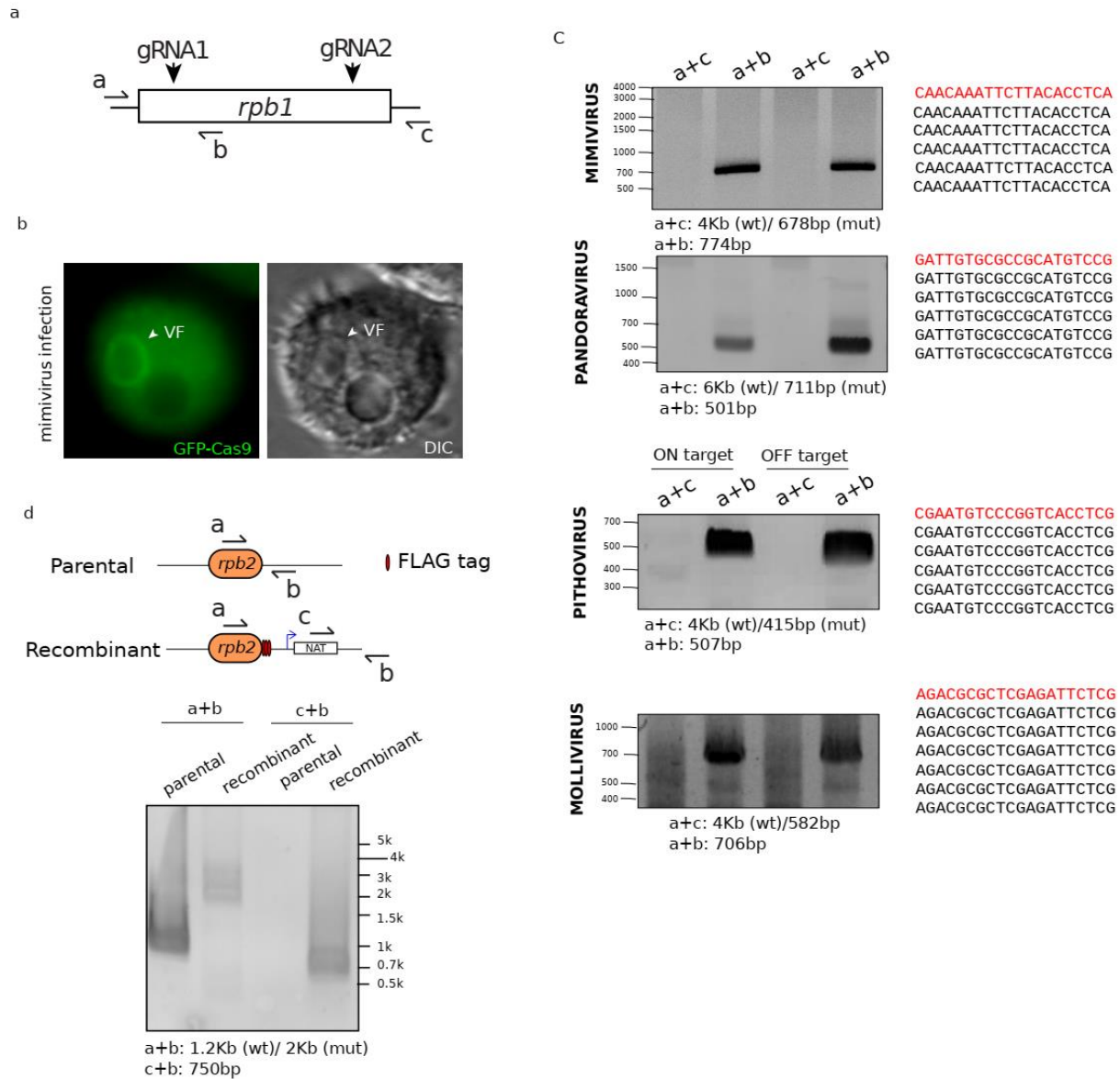


Figure S2. CRISPR/Cas9 allows manipulation of nuclear GVs.

(a) Schematic representation of the *rpb1* locus, guide targeting location and primer annealing sites.

(b) Representative micrograph showing exclusion of GFP-Cas9 from the viral factory of mimivirus. VF: viral factory, white arrow head. An image of a non-infected cell is shown in Fig. 1a. Micrographs are representative of 3 independent experiments.

(c) PCR of the genomic locus of the *rpb1* shows no noticeable deletions. Due to the length of the PCR product from primers a+c, when deletions are not present, wt PCR products are not detected with the current PCR setting. Sequencing results from a library from the PCR products demonstrate the lack of mutant viruses. The wild type sequence is shown in red. PCRs are representative of 3 independent experiments.

(d) PCR of the genomic locus of the *rpb2* demonstrate correct integration and clonality. A schematic representation of the wild-type and recombinant *rpb2* locus, and primer annealing sites are also shown. PCRs are representative of 2 independent experiments.

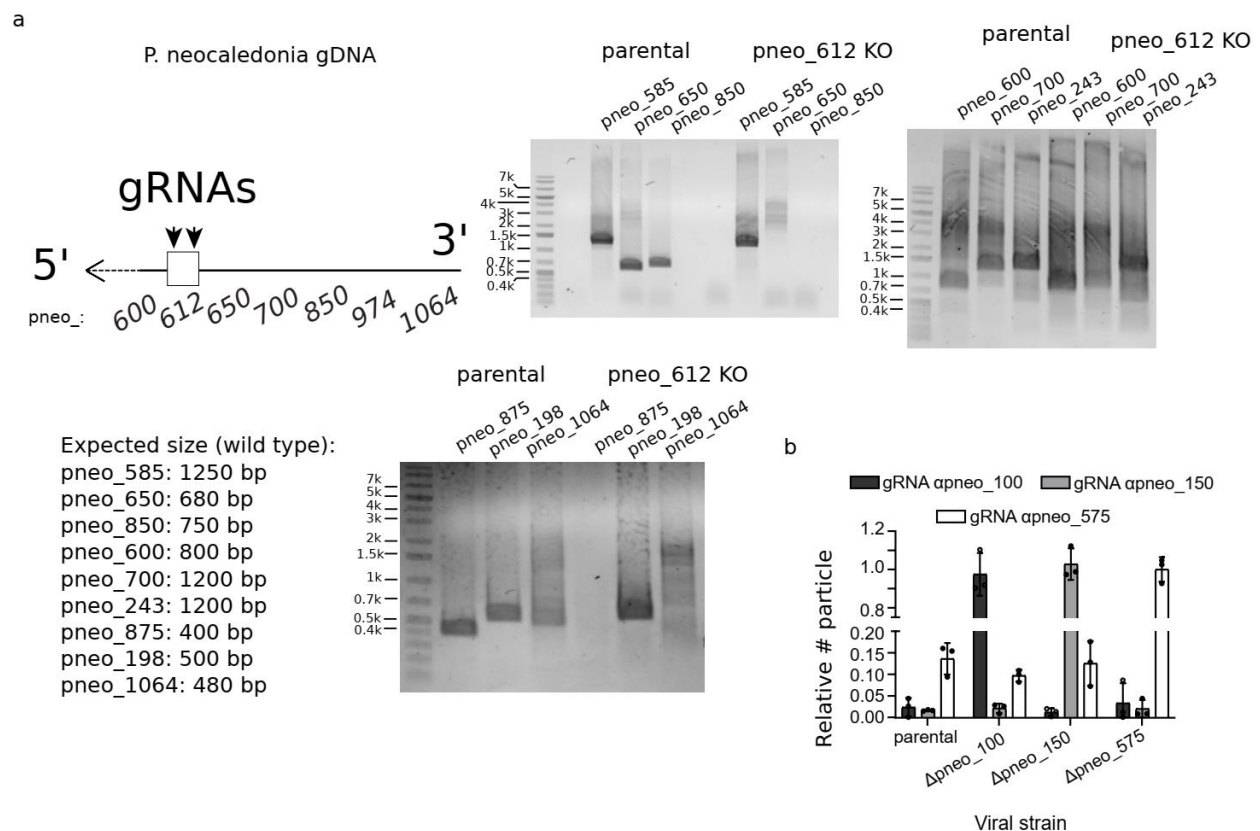


Figure S3. CRISPR/Cas9 targeting of pandoravirus genome demonstrates the presence of a core essential genome.

(a) PCR of different genomic locus shows large deletion at the 3' end of pandoravirus. Lack of specific PCR product indicates loss of the gene. Arrows indicate the gRNA targeting site.

(b) Relative quantification of the number of viral particles by parental (wild-type virus) or CRISPR/Cas9 modified viral strains after 24h of infection of CRISPR/Cas9 expressing

Acanthamoeba cells. Modified viral strains were generated by continuous passage in CRISPR/Cas9 expressing cells until resistant viruses were obtained by each specific gRNA. Data is expressed relative to the growth of the strains in an off-target strain (gRNA against the mcp of mollivirus). Data correspond to the mean $\pm$ SD of 3 independent experiments. α indicates the target of the gRNA.

Source data are provided as a Source Data file.

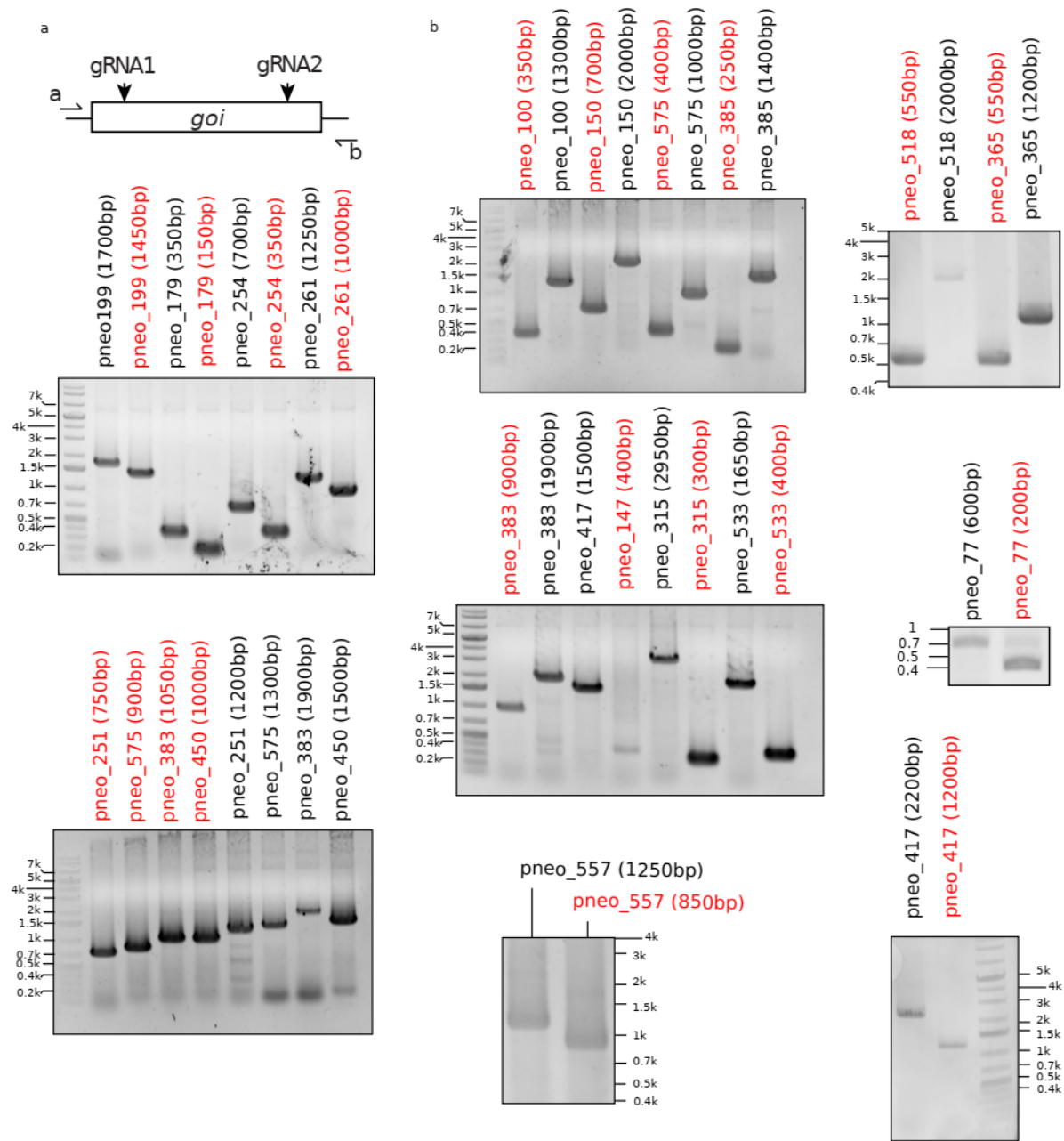


Figure S4. Pandoravirus genome organization keeps traces of their smaller ancestors.

(a) Schematic representation of the *gene of interest (goi)* locus, guide targeting location and primer annealing sites.

(b) PCR of the genomic locus of the *goi*. Genotyping on parental viruses is shown in black while genotyping in recombinant viruses is marked in red. Expected PCR product size is indicated between parenthesis.

(c) Quantification of the infectious dose (ID50) per particle number was assessed as described in materials and methods. Values were normalized to wild type ID50/particle number (in grey). Data correspond to the mean $\pm$ SD of 3 independent experiments.

Source data are provided as a Source Data file.

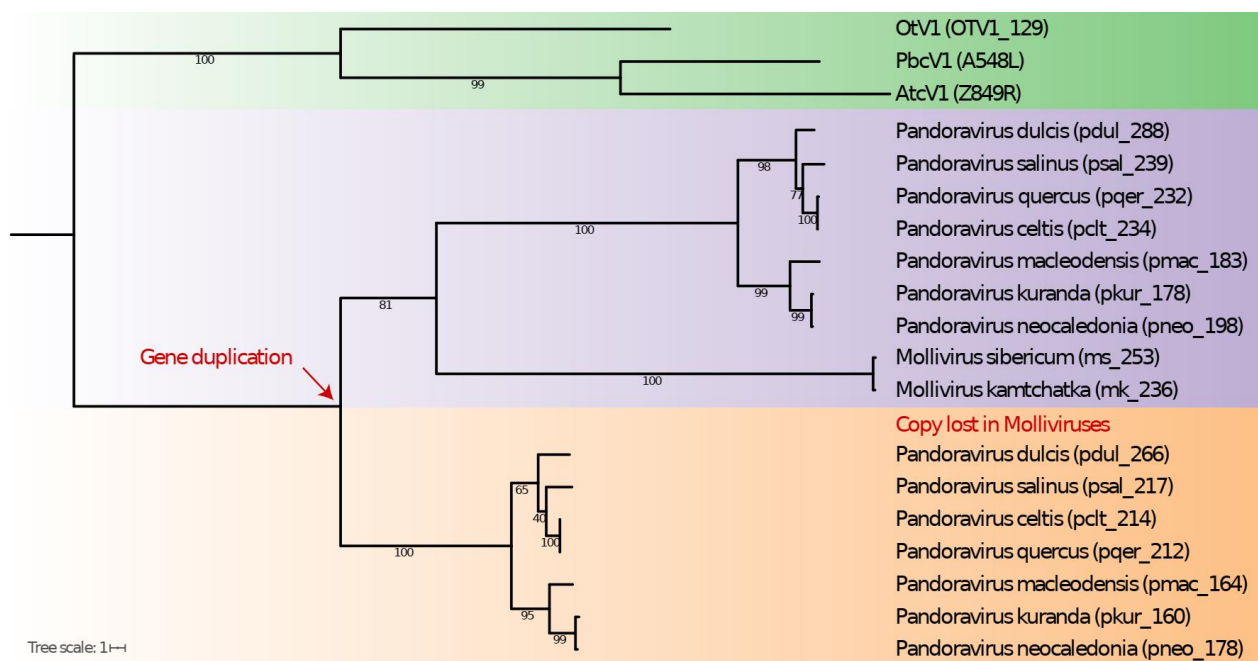


Figure S5. Phylogenetic analysis of the SNF2-like helicases.

(A) Phylogenetic analysis of the SNF2-like helicases in selected members of the *Phycodnaviridae*, mollivirus and pandoravirus. *Emiliana huxleyi* virus 86 (NC_007346.1), *Feldmannia species* virus (NC_011183.1), *Ectocarpus siliculosus* virus 1 (NC_002687.1), *Ostreococcus tauri* virus 1 (NC_013288.1), *Paramecium bursaria* Chlorella virus 1 (NC_000852.5), *Mollivirus sibericum* (NC_02 7867.1), *Mollivirus kamchatka* (MN812837.1), *Pandoravirus dulcis* (NC_021858.1), *Pandoravirus salinus*

(NC_022098.1), *Pandoravirus quercus* (NC_037667.1), *Pandoravirus macleodensis* (NC_037665.1). and *Pandoravirus kuranda* (ON887157).

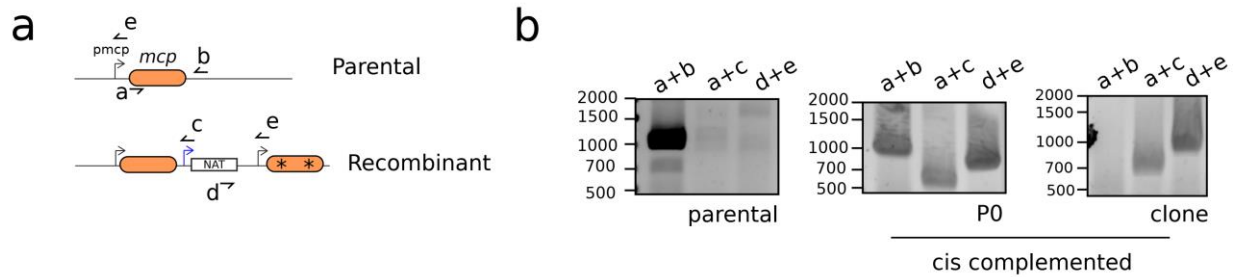


Figure S6. Mollivirus MCP acts as a scaffolding protein for tegument biosynthesis.

(A) Schematic representation of the *mcp* locus and primer annealing sites.

(B) PCR of the genomic locus of the *mcp* demonstrate correct integration and clonality. Integration in the population at the generation 0 (P0, 24hours post transfection) is also shown. Expected size for PCR products: a+b (parental): 1000bp, a+c (recombinant): 650bp, d+e (recombinant): 800bp. PCRs are representative of 2 independent experiments.

Table S1: List of primers used in this study.

ID	Purpose	Sequence
HB17	3'UTR-pU6-(tRNA-guide-scaffold)	CTGGGCGGTGACTAActag
HB18	3'UTR-pU6-(tRNA-guide-scaffold)	gttggtccgggtcgactctaG
HB19	convert guide to NotI for cloning	CTGGCCAgcggccgcGtttagagctagaaatagcaag
HB20	convert guide to NotI for cloning	ctaaaacgcggccgcTGGCCAGTATGGGGATC
HB21	guide RBP1 pandoravirus neocal	TCCCCATACTGGCCAgTGACGGCCACCACCGAACCGgttttagagctagaaatagc
HB22	guide RBP1 pandoravirus neocal	ttctagctctaaaacGATTGTGCGCCGCATGTCCGTGGCCAGTATGGGG
HB23	guide RBP1 mollivirus kamch	TCCCCATACTGGCCAAGACGCGCTCGAGATTCTCGgttttagagctagaaatagc
HB24	guide RBP1 mollivirus kamch	ttctagctctaaaacGCTCGAGGTCCACATACCTGTGGCCAGTATGGGG
HB25	guide RBP1 pithovirus siber	TCCCCATACTGGCCAgCGAGGTGACGGGGACATTTCgttttagagctagaaatagc
HB26	guide RBP1 pithovirus siber	ttctagctctaaaacGATGGGTGAGATCATCACCGTGGCCAGTATGGGG
HB27	convert guide to NotI for cloning	gtcctgcgggttcgcc

HB28	convert guide to NotI for cloning	cgaaacccgacaggac
HB29	Check genome of RBP1 Queen	GACATTCGGCTATTGTAGATCCT
HB30	Check genome of RBP1 Queen	CAAAGGTCCAAGACCCTC
HB31	Check genome of RBP1 Queen	GAGAACCATCGCCTTTAGTG
HB32	Check genome of RBP1 Pitho siber	CTTGGAGGTGGTATCCGTC
HB33	Check genome of RBP1 Pitho siber	TTCCTCGTAATCAGCCTCC
HB34	Check genome of RBP1 Pitho siber	CAGATAAGGGCACGAACAAC
HB35	Check genome of RBP1 Pandora neocal	ATGACATCGCCGGTCATCTAC
HB36	Check genome of RBP1 Pandora neocal	GTCCGCTGCCGAGGTG
HB37	Check genome of RBP1 Pandora neocal	TGCCTTGCCGCGCTTG
HB38	Check genome of RBP1 Molli kamtch	GAGGCCTCAACGACCAC
HB39	Check genome of RBP1 Molli kamtch	GATGCCGGACACCACCAC
HB40	Check genome of RBP1 Molli kamtch	CACGGCATGCAGCAGGC
HB49	guide against RFP	TCCCCATACTGGCCAgCTTGGTCACCTTCAGCTTGggttttagagctagaaatagc
HB50	guide against RFP	ttctagctctaaacCGATGGTGTAGTCCTCGTTGcTGGCCAGTATGGGG
HB60	Guide NLF3 pando	TCCCCATACTGGCCAg CCTGGCTCGCTGCACTCTGT gttttagagctagaaatagc
HB61	Guide NLF3 pando	ttctagctctaaacCCGAGCGCGTGCGCCAGATCTGGCCAGTATGGGG
HB62	guide MCP molli	TCCCCATACTGGCCAgCAACGACACCGCCGTGAGCCgttttagagctagaaatagc
HB63	guide MCP molli	ttctagctctaaacTGCGCCGCAGCATCCACATGcTGGCCAGTATGGGG
HB64	Guide Pneo-177	TCCCCATACTGGCCAg TTGGGCGAGGACATtgacg gttttagagctagaaatagc
HB65	Guide Pneo-177	ttctagctctaaacACACCAAGTCCTCGTCGTGCTGGCCAGTATGGGG
HB66	Guide pneo-585	TCCCCATACTGGCCAg CGTTGGCGTTGGTCGTATAG gttttagagctagaaatagc
HB67	Guide pneo-585	ttctagctctaaacGTGGACGCTCGCCGACCAGTcTGGCCAGTATGGGG
HB68	guide pneo-972	TCCCCATACTGGCCAg CATTGGCCTTGGTCCAGTAG gttttagagctagaaatagc
HB69	guide pneo-972	ttctagctctaaacCCGACGTGTCGGTGACGTTGcTGGCCAGTATGGGG
HB76	To generate Cas9 to integrate in RFP locus	AGAATTCACGCGTCCGGAGGCATCGTGGAACAGTACGAGCG
HB77	To generate Cas9 to integrate in RFP locus	ATATAGATCTaggcctGCTGACTAATTGAGATGCATGC

HB78	To generate Cas9 to integrate in RFP locus	ATTAGTCAGCaggcctAGATCTATATGCAACCTGTGACG
HB79	To generate Cas9 to integrate in RFP locus	CGCTGCACCTAGGGGAGAGGCTTGGCGGTCTGGGTGCC
HB84	MCP1-RFP	TCTACTAAGACATAATGACGGACCACCTTCTGGC
HB85	MCP1-RFP	ccTCGGAGGAGGCCATCGCTCGGCACACGTGGTTG
HB86	RFP-MCP1	CACTCCACCGGCGCCATGACGGACCACCTTCTGGC
HB87	RFP-MCP1	CCGGGTCGACTCTAGTTACGCTCGGCACACGTG
HB88	Check genome of MCP Molli	cggcttctgttctgtccatcgtgc
HB89	Check genome of MCP Molli	CGACGGCCGTCGCGTCG
HB90	Check genome of NLF3 Pandora neocal	gtctgccctgtcctctgttgc
HB91	Check genome of NLF3 Pandora neocal	GGTCCCGACAGGCCGGTG
HB92	Check genome of Pneo-177 Pandora neocal	ctcccagatgcgtcgtgtcc
HB93	Check genome of Pneo-177 Pandora neocal	ccggcgaggcagcgacc
HB94	Check genome of pneo-585 Pandora neocal	ATGCAACGACCTGTAGCGACCG
HB95	Check genome of pneo-585 Pandora neocal	ggcgcccgcctatggac
HB96	Check genome of pneo-972 Pandora neocal	gtccagATGGGTATGCGTGCTCG
HB97	Check genome of pneo-972 Pandora neocal	gtccggtgtcagcactaccaagc
HB98	Mutations MCP molli for complementation	tGGgGctAGaCCAGCGTCGACCAGAAC
HB99	Mutations MCP molli for complementation	gAGaCTgACcGCGGTGTCGTTGATGGC
HB100	Mutations MCP molli for complementation	ACCGCgGTcAGtCTcGCCGACATCTCGCCGGCG
HB101	Mutations MCP molli for complementation	GCTGGTCTAGCCCCACAGCATCCACATGTAGAAGATGATG
HB102	guide CS1-3 A cast	TCCCCATACTGGCCAGAGAACGCGTGCGCCTTCGCgttttagagctagaataagc
HB103	guide CS1-3 A cast	ttctagctctaaaacgccgtcaagatctggccacTGGCCAGTATGGGG
HB104	guide pneo182 pandora neocal	TCCCCATACTGGCCAgACTCGCCGGGCACGTGATAGgttttagagctagaataagc
HB105	guide pneo182 pandora neocal	ttctagctctaaaacCTAACGGCGCCGCAGCCAGCTGGCCAGTATGGGG
HB126	HR MCP molli	TCTACTAAGACATAtgCGAAGCCATGCCAGGACAG
HB127	HR MCP molli	CCCTTGCTCACCATAATGGCATTGTTGAAGCGGTGGg
HB128	HR MCP molli	GCGTGGTACCTCTAGCCAGCGTCGACCAGAACTC

HB129	HR MCP molli	CCGGGTCGACTCTAGaGGACACGGCAAGGCAAAAC
HB134	reverse primer in RFP (check clonality)	CCAGCCCATGGTCTTCTTCTGC
HB135	CS1-genotyping	GTTCGTGCAGACGCCGAG
HB136	CS1-genotyping	gcTCGGCGAAGAAGACGAAGAC
HB137	CS1-genotyping	GCGAGGAGGACCGCATCGAG
HB138	CS1-genotyping	GCAGATGCGGTGGGTGAGG
HB139	CS2-genotyping	Agcagcgcgacaagatgctc
HB140	CS2-genotyping	Agatgcggtggcacaggtgg
HB141	CS2-genotyping	gtgctgcgaaggccag
HB142	CS2-genotyping	tgtagaactgggccaccag
HB143	CS3-genotyping	TGCCCTGGTTCTACAACGAG
HB144	CS3-genotyping	TGAAATTGGACGGGCGCTGG
HB173	qPCR 5 end pneo genome	TCCTTTTCCCAACCATCCC
HB174	qPCR 3 end pneo genome	TTCTCCCATTCATTCAAACCC
HB175	qPCR 3 end pneo genome	TAGCATAGCATCACCGTCC
HB182	genotyping MCP KO	cgacccgtggctctgtcaac
HB183	genotyping MCP KO	caccaccaacatcatcactatc
HB184	genotyping pneo182 KO	gtctcctcacacatattg
HB185	genotyping pneo182 KO	cagaagagacaccaccattg
HB199	Promoter pneo650 in for NAT	CTTTTGCAAAAAGCTtCGGCTCTGCAGTTGATCTC
HB200	Promoter pneo650 in for NAT	CAGAAGAATCAAGCTGATGATGGTTGCGGTTGC
HB201	3-UTRP in NAT pneo640	gcggccgcGATATCGCGGGACTCTGGGG
HB202	3-UTRP in NAT pneo640	TTAGGGGCAGGGCATGCTC
HB203	3-UTRP in NAT pneo640	ATGCCCTGCCCTAATAAGCGGCGCCGTGCCGC
HB204	3-UTRP in NAT pneo640	CGATATCgcggccgcTGTGCCTTAGAGACATTTTGTG
HB213	guide pneo62	TCCCCATACTGGCCAGGACCGCAGCGGGTCCATGGgttttagagctagaaatagc
HB214	guide pneo62	ttctagctctaaaacCGACACTGCAACGCTCGTTTcTGGCCAGTATGGGG
HB215	guide pneo350	TCCCCATACTGGCCAgTGACGTCGTGCGCCCCCTCCGgttttagagctagaaatagc
HB216	guide pneo350	ttctagctctaaaacAGGCCGACCAGTACCCGATCTGGCCAGTATGGGG
HB217	guide pneo450	TCCCCATACTGGCCAgTGTCTACGCCCATCTGGACGgttttagagctagaaatagc
HB218	guide pneo450	ttctagctctaaaacTCGCAAATGGCTGCGACATGcTGGCCAGTATGGGG
HB219	guide pneo518	TCCCCATACTGGCCAgCCC GTGAGTGCTTACGTGgttttagagctagaaatagc
HB220	guide pneo518	ttctagctctaaaacCCAGCGAGCAGCGCAAGCGCTGGCCAGTATGGGG

HB221	guide pneo959	TCCCCATACTGGCCAgAATGTCGGGCCTGAGACTCGgttttagagctagaaatagc
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HB266	3 UTR MCP cis-complementation	CGCGATATCGCGGCCgcCGGCCACATCGTGCAGCAG
HB267	5 UTR MCP cis-complementation	TTTGTTGCGCGAATTcCAGGTGCAGCACCCTCC
HB268	5 UTR MCP cis-complementation	aaaatAAGAACAAGAgggcccTTACGCTCGGCACACGTG
HB295	Check integration MCP 3' end	CGGCAACCAGATCATGGTCATC
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HB334	guide pneo 700	TCCCCATACTGGCCAGCGAGGGCCATCATCGCGTCGgttttagagctagaaatagc
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HB390	genotype pneo100	GGCGCTATCGGCCAC
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HB392	genotype pneo150	TGACGAGTGCCTGGCAGC
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HB414	genotyping pneo243	GCGCCTGCGATCTGACGACG
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HB416	genotyping pneo289	GCCGACGGTCAGCGAG
HB417	genotyping pneo289	TAGGTGGCAGGCGCGTC

HB418	genotyping pneo523	TGCCGCGGCGTGAGCGTG
HB419	genotyping pneo523	CCACACGCTGGCCAGGTTGG
HB420	genotyping pneo700	GACAACAACGACGCCAAAG
HB421	genotyping pneo700	TCGACATCCTCAATGTCGGC
HB422	genotyping pneo801	Ctctccgtgtgcgctacc
HB423	genotyping pneo801	GGCCATGGATGTGCAACGCG
HB424	genotyping pneo600	CCGCTGCTCGATCGGGTCG
HB425	genotyping pneo600	GCCATGTGCCCCGAGATGCGG
HB426	genotyping pneo750	GCGGCCACTGGTCGAC
HB427	genotyping pneo750	CACGGACATCAGCCATCG
HB428	genotyping pneo850	CCCAAGTAGCGTCAAAGGG
HB429	genotyping pneo850	TCGTGTCATCATCAACGTC
HB430	genotyping pneo900	GGCGCGCTGGTCTCTGAG
HB431	genotyping pneo900	GTACAGGCAGTCAAGACTGC
HB432	genotyping pneo950	GTGTGAGGAGAGATGCGGTC
HB433	genotyping pneo950	CTCGGCGTTGCGCAGG
HB434	genotyping pneo533	TTTGCCACGGCGTGGGAG
HB435	genotyping pneo533	AGCGTGTGTCGCGAAGATAG
HB436	genotyping pneo518	Tctcgaggctctgcccgcac
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HB438	genotyping pneo450	aatccacattgccgaggag
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HB440	genotyping pneo557	GCTCACCGCCGAGGCCTACC
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HB450	guide pneo 153	TCCCCATACTGGCCAgCGTGCCAGCGGCACCAACGgttttagagctagaaatagc
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HB454	guide pneo 238	TCCCCATACTGGCCAgCGACGGGACGCTCATCATCGgttttagagctagaaatagc
HB455	guide pneo 238	ttctagctctaaaacCAGTTGGACCTGCGCAAGTTTGGCCAGTATGGGG

HB456	guide pneo 248	TCCCCATACTGGCCAGGTGGTGTGTACACAACAGgttttagagctagaaatagc
HB457	guide pneo 248	ttctagctctaaaacCAACAAGCACGAGACCGAGGcTGGCCAGTATGGGG
HB458	guide pneo 251	TCCCCATACTGGCCAgCGTGTGGTAATACCGCCCGTgttttagagctagaaatagc
HB459	guide pneo 251	ttctagctctaaaacCGAACCGACGGTGCGCGCCAcTGGCCAGTATGGGG
HB460	guide pneo 254	TCCCCATACTGGCCAGGACTATACGGTCTCGACGGgttttagagctagaaatagc
HB461	guide pneo 254	ttctagctctaaaacCTCATGGGCGCGCTGCCTTGcTGGCCAGTATGGGG
HB462	guide pneo 264	TCCCCATACTGGCCAgTGACGCGGCGCTTTGTTCATGgttttagagctagaaatagc
HB463	guide pneo 264	ttctagctctaaaacCGACGATGGCCTCGCCGTGGcTGGCCAGTATGGGG
HB464	guide pneo 301	TCCCCATACTGGCCAGTTGGAGAATGGGATGATGGgttttagagctagaaatagc
HB465	guide pneo 301	ttctagctctaaaacCTGTGATGCCCTTACGACGcTGGCCAGTATGGGG
HB466	guide pneo 315	TCCCCATACTGGCCAAAAGGACGTCGCCTGGCCGGgttttagagctagaaatagc
HB467	guide pneo 315	ttctagctctaaaacCTGCCCCAGAGAAAACACATTTGGCCAGTATGGGG
HB468	guide pneo 321	TCCCCATACTGGCCAgTATCGGCAGATCATCATCGGgttttagagctagaaatagc
HB469	guide pneo 321	ttctagctctaaaacCCAGTCACGGGCGCGCCTTAcTGGCCAGTATGGGG
HB470	guide pneo 325	TCCCCATACTGGCCAgTGACGTGGGCCTCCACTTGgttttagagctagaaatagc
HB471	guide pneo 325	ttctagctctaaaacCATGGGACCGCGGGGCGTCTTGGCCAGTATGGGG
HB472	guide pneo 365	TCCCCATACTGGCCAGAATGGGCCGGACATTGCCGgttttagagctagaaatagc
HB473	guide pneo 365	ttctagctctaaaacCGATGAACCAGCGGTAGTTGcTGGCCAGTATGGGG
HB474	guide pneo 383	TCCCCATACTGGCCAgTGATTGCGAGGGCACGTCCGgttttagagctagaaatagc
HB475	guide pneo 383	ttctagctctaaaacCGAATGTGCGACGAGCAACAcTGGCCAGTATGGGG
HB476	guide pneo 385	TCCCCATACTGGCCAGTGCGCCAGCGAGCGCATAGgttttagagctagaaatagc
HB477	guide pneo 385	ttctagctctaaaacCCTGGCGCTCTCTGGCCAACTGGCCAGTATGGGG
HB478	guide pneo 417	TCCCCATACTGGCCAGATGGCCAACGCGCTCGTTGgttttagagctagaaatagc
HB479	guide pneo 417	ttctagctctaaaacCCAAAGGACCGACGCCAAAGcTGGCCAGTATGGGG
HB480	guide pneo 459	TCCCCATACTGGCCAgTGCTCCAAGGCTATCTGGgttttagagctagaaatagc
HB481	guide pneo 459	ttctagctctaaaacCCAACCTCCTCGCAGACACTTGGCCAGTATGGGG
HB482	genotype 178	GCACTGTGCAGACGCGCGTC
HB483	genotype 178	CAACACGACTCGTGTCTGTC
HB484	genotype 179	ATGCAGTGCCGCAAGAACC
HB485	genotype 179	ATTGGCGCGTAACCTGCGG
HB486	genotype 181	CCATGATCTTCTAGACGCG
HB487	genotype 181	gcaaCTAGAGGGGCGAGTC
HB488	genotype 350	GGCGCTACCCCATACCGTC

HB489	genotype 350	GTGCGGACGGTCGGCGTC
HB490	cis complementation MCP	ACTCGTCTCCCGTTTCC
HB491	cis complementation MCP	TGTGCCTTAGAGACATTTTGTG
HB492	cis complementation MCP	CAAAATGTCTCTAAGGCACATTTGTCCGTCTGTCCACTTTGG
HB493	cis complementation MCP	GGTCCGTCATCGTTTCTTTGCTCGCTGGGG
HB494	cis complementation MCP	CAAAGAAACGATGACGGACCACCTTCTGGC
HB495	cis complementation MCP	GGGGAAACGGGAGGACGAGTTTACGCTCGGCACACGTG
HB502	genotype pneo153	GCTGCGCACACGCATCGATC
HB503	genotype pneo153	GTCGACGCGCCTTGC
HB504	genotype pneo228	TTCGGCCTACCTGGCCG
HB505	genotype pneo228	GCCGGCATGCAATGCCAC
HB506	genotype pneo238	CCTGCACCAGGGCACCTC
HB507	genotype pneo238	CAGTGGACGATGCCCCACG
HB508	genotype pneo248	CTCGTGGCACACCCGCC
HB509	genotype pneo248	CAGCGCCTTCGAGCGCC
HB510	genotype pneo251	GAGCAGCGGAGCGATGCC
HB511	genotype pneo251	CGTCCCTTGTGCGTCGGAC
HB512	genotype pneo254	GCGTGCGCCTGCGTCATTTT
HB513	genotype pneo254	GTGCGATAGTCGGGCGCAC
HB514	genotype pneo264	GCCTACTATGGCGTCGAGAG
HB515	genotype pneo264	ACTGCTCGGGAAAGTCCATG
HB516	genotype pneo301	GATGGACGAGGACGACGATG
HB517	genotype pneo301	CGAACTCGAGCACGAACC
HB518	genotype pneo315	GAGTTTGCCAGCGTCCGG
HB519	genotype pneo315	GCGCATTGATTGAGCACGG
HB520	genotype pneo321	CCCAACGACACGCGGCTC
HB521	genotype pneo321	CGAGCCGCAAGGCGTTG
HB522	genotype pneo325	CGACCGATGTCAACCGCG
HB523	genotype pneo325	CATGTTGAGGCACGCGCAC

HB524	genotype pneo365	CCCGCATCTCAACACGCAG
HB525	genotype pneo365	TGGCGTGGTGCATGACGATG
HB526	genotype pneo383	CTGCAGACTCTGCGTTGGCG
HB527	genotype pneo383	CGTCGTGCACCACCTTGTC
HB528	genotype pneo385	GGAGACCACCTAATGGCCG
HB529	genotype pneo385	ACGTCACATGGCATCGATCC
HB530	genotype pneo417	GGAACCGGATAGCACCACG
HB531	genotype pneo417	GGTCGCGCTCAGAGAGTAC
HB532	genotype pneo459	CATCGTCGAGCGCCTCAC
HB533	genotype pneo459	CGACAGGTCGGATCCGAC
HB534	guide pneo 187	TCCCCATACTGGCCAgCACGTGCCTCGGCATTTGGGgttttagagctagaaatagc
HB535	guide pneo 187	ttctagctctaaaacCGATAGTAGTGCCGACGAAAcTGGCCAGTATGGGG
HB536	guide pneo 198	TCCCCATACTGGCCAgCAAGGTACTGCGCCTCTTGGgttttagagctagaaatagc
HB537	guide pneo 198	ttctagctctaaaacCTGCTATCGCCATCCCCAGCTGGCCAGTATGGGG
HB538	guide pneo 221	TCCCCATACTGGCCAGTAGGGAAAGCTGCTCACGGgttttagagctagaaatagc
HB539	guide pneo 221	ttctagctctaaaacCCGCCAACCAGTACCGAGCTGGCCAGTATGGGG
HB540	guide pneo 231	TCCCCATACTGGCCAgCGAGCGCATGGGGAGCGTTGgttttagagctagaaatagc
HB541	guide pneo 231	ttctagctctaaaacCAAGAATGCGTCGGTACGTGcTGGCCAGTATGGGG
HB542	guide pneo 250	TCCCCATACTGGCCAgTGGCGACGGCGGTCGCGCTGgttttagagctagaaatagc
HB543	guide pneo 250	ttctagctctaaaacCCATCGATCCCACCAACATCTGGCCAGTATGGGG
HB544	guide pneo 261	TCCCCATACTGGCCAgAATCTCATTGCGACCCATGGgttttagagctagaaatagc
HB545	guide pneo 261	ttctagctctaaaacCACCGCGCCCGACCGCGTATcTGGCCAGTATGGGG
HB546	guide pneo 276	TCCCCATACTGGCCAgCAAGGGCGTGGCGATCGTCGgttttagagctagaaatagc
HB547	guide pneo 276	ttctagctctaaaacCCAGCGAGCGGCCGCACCATcTGGCCAGTATGGGG
HB548	guide pneo 304	TCCCCATACTGGCCAgCATGTGACCCACGCCGTCGGgttttagagctagaaatagc
HB549	guide pneo 304	ttctagctctaaaacAGATGATGTGCCTGGCCACGcTGGCCAGTATGGGG
HB550	guide pneo 313	TCCCCATACTGGCCAgTGAGCCCGACTGGAGGTCA TGtttttagagctagaaatagc
HB551	guide pneo 313	ttctagctctaaaacCAAAGCAGCACGCACGCTCCTGGCCAGTATGGGG
HB552	guide pneo 322	TCCCCATACTGGCCAgCCACTTTGCGGCCCTGTTAGgttttagagctagaaatagc
HB553	guide pneo 322	ttctagctctaaaacCTGCCGATCGGCCATCGTCCTGGCCAGTATGGGG
HB554	guide pneo 342	TCCCCATACTGGCCAGGTCGCTCACGCGCTCGTCGgttttagagctagaaatagc

HB555	guide pneo 342	ttctagctctaaaacCCATGCACACCGACGAGATCTGGCCAGTATGGGG
HB556	guide pneo 344	TCCCCATACTGGCCAgCGCCGGGCGCAAGTTTGAAGgttttagagctagaaatagc
HB557	guide pneo 344	ttctagctctaaaacCGATGAGATCGTCGGCCTCGcTGGCCAGTATGGGG
HB558	guide pneo 407	TCCCCATACTGGCCAgTTTCGATCCAGCGTTCGGGGgttttagagctagaaatagc
HB559	guide pneo 407	ttctagctctaaaacCCGACCTGCTCGACAAGATAcTGGCCAGTATGGGG
HB560	guide pneo 414	TCCCCATACTGGCCAgAGAGTGCGTGGCCTTTATGGgttttagagctagaaatagc
HB561	guide pneo 414	ttctagctctaaaacCGAACAGCTGCACGCCGAGCTGGCCAGTATGGGG
HB562	guide pneo 425	TCCCCATACTGGCCAgATACTCTTGGCGGATGCCGGgttttagagctagaaatagc
HB563	guide pneo 425	ttctagctctaaaacCAGACGATCAGAGCCAGACAcTGGCCAGTATGGGG
HB564	guide pneo 429	TCCCCATACTGGCCAgCGAATGGACGGCCACCCTCGgttttagagctagaaatagc
HB565	guide pneo 429	ttctagctctaaaacCCAATATGGCCGAGGCCATCTGGCCAGTATGGGG
HB566	guide pneo 443	TCCCCATACTGGCCAgCGTGCGGGTTCTACGCTTGgttttagagctagaaatagc
HB567	guide pneo 443	ttctagctctaaaacCAAGTATGCAGCGGCACGCGcTGGCCAGTATGGGG
HB568	guide pneo 544	TCCCCATACTGGCCAgAAAAGAATCGCGTCTGTTGGgttttagagctagaaatagc
HB569	guide pneo 544	ttctagctctaaaacCCATTGATCGCGTGGCGTCGcTGGCCAGTATGGGG
HB572	qPCR molli	TGATGGATGGATACAGCGAG
HB573	qPCR molli	GAGAAGGAGGAGAAGGAAGAG
HB576	genotype 187	GCCAACACTCGACGCGAC
HB577	genotype 187	GAGGTCAAGCGCATTTGTG
HB578	genotype 198	GACGCAACGGACGAGCAG
HB579	genotype 198	CACAACTGGCGTGTCATC
HB580	genotype 221	CGCGCACTGCCATACTCG
HB581	genotype 221	GCCTGCGCCATCATTTGC
HB582	genotype 231	GCCCAGTCAACACACAG
HB583	genotype 231	ACAGTTCTGCGACCATGTG
HB584	genotype 250	TCAGGCCTCACGCGAGC
HB585	genotype 250	TTGCCGCTGATCGGATG
HB586	genotype 261	GGCAACAATGACAATGAGCATG
HB587	genotype 261	CATGCTGCTTATGCGGTCG
HB588	genotype 276	CAGTGCAACGCGTCGAC
HB589	genotype 276	TATGTAGTCGGCCACGAG
HB590	genotype 304	CATGCACTTTGACATGGGC
HB591	genotype 304	ACACAGCTGGTAGGTGAC
HB592	genotype 313	TGGCGGTGGCGATGATG
HB593	genotype 313	CAAACATCGCACGACGATCG
HB594	genotype 322	CTTTCAGTGCCCGGAGC
HB595	genotype 322	CGCGTATAATAGAGCCAGGC

HB596	genotype 342	TACGCGCTCATGGAGACC
HB597	genotype 342	GAACCACAAGAGCAGGTAGG
HB598	genotype 344	CAACCCGGTGAGCGTGAG
HB599	genotype 344	CGCAGCGGCGTGAGAATG
HB600	genotype 407	CGTCGACGACATTGCGCG
HB601	genotype 407	CTGCCGTAGCCGCAGTG
HB602	genotype 414	CAAGGAACGCCAACAGACC
HB603	genotype 414	GGTGGGGTCGACCGAAGC
HB604	genotype 425	ATGACCGAGACGCATCGC
HB605	genotype 425	TGCGGCGAGTTGAGACCG
HB606	genotype 429	CTCGACAATGTGCACCACG
HB607	genotype 429	GGCTTGACGCCAGGTTT
HB608	genotype 443	ACGCCGAGCGCATCGTG
HB609	genotype 443	CGCCATTCTGCCAGCGTC
HB610	genotype 544	CGTCGACGCGCCAGACAG
HB611	genotype 544	GTAGGCGGCCTCTTCAACG
HB612	genotype mk236	GAGCCCCAAGGTCAGTCC
HB613	genotype mk236	CCAGTCGTTGCATGCCTG
HB614	genotype 875	ACGCACCTCCACTATGAGG
HB615	genotype 875	GAGGATGCCGATCTGGACG
HB616	genotype 925	CGTGCCTTTCCAAGTGCG
HB617	genotype 925	CGTTGCTGGTCGGTTCAG
HB618	genotype 1064	AGCCTCTGCGTGGTCGAC
HB619	genotype 1064	CAGTGCGTGCCGGTAGTG
HB740	Endogenous taging Rbp2 pneo add HA	TTTGTTGCGCGAATTcTACCCATACGATGTTCTCTGAC
HB741	Endogenous taging Rbp2 pneo add HA	aaaatAAGAACAAGAGTTAAGCGTAATCTGGAACGTC
HB742	Endogenous taging Rbp2 pneo add FLAG	TTTGTTGCGCGAATTCCATATGTCAGACTACAAAGACCATG
HB743	Endogenous taging Rbp2 pneo add FLAG	aaaatAAGAACAAGATTACTTGTCTATCGTCATCCTTG
HB744	5HR FLAG+HA	TTTGTTGCGCGAATTCAAGAACGCGCCCGTCGTCG
HB745	5HR FLAG	GTAGTCTGACATATGATTGTCGGCATTGTCGTGATCG
HB746	5HR HA	AACATCGTATGGGTAATTGTCGGCATTGTCGTGATCG
HB747	3HR	tgtctcTAAGGCACACCGCCGGTGCTCTCCGAATG
HB748	3HR	CGCGATATCGCGGCCgcCTCTGCGCCGCCACGCC
HB749	gRNA pneoRbp1	TCCCCTACTGGCCAGGTGGTGGCGATGATTACGGgttttagagctagaaatagc
HB750	gRNA pneoRbp1	ttctagctctaaaacCCACGCGGCCGACGCCATCTGGCCAGTATGGGG
HB751	gRNA pneoRbp1	TCCCCTACTGGCCAGGTGCGGGTGGCGCTTCTCTAGgttttagagctagaaatagc
HB752	gRNA pneoRbp1	ttctagctctaaaacATGTCAACGACCGCCACTTTcTGGCCAGTATGGGG

Table S2: List of constructs generated in this study.

ID	Purpose	Addgene ID
vHB4	GFP-Cas9/guidesRBP1 queen	
vHB5	GFP-Cas9-NLS/guidesRBP1 queen	
vHB6	lowGFP-Cas9/guidesRBP1 queen	
vHB7	GFP-Cas9/NotI site	
vHB8	GFP-Cas9-NLS/NotI site	
vHB9	lowGFP-Cas9/NotI site	193454
vHB11	GFP-Cas9-NLS/guidesRBP1 Molli kamtch	
vHB12	GFP-Cas9/guidesRBP1 Pitho siber	
vHB13	lowGFP-Cas9/guidesRBP1 Pitho siber	
vHB16	GFP-Cas9-NLS/guidesRBP1 Pandora neocal	
vHB17	GFP-Cas9-NLS/guides Red Fluor Prot	
vHB18	GFP-Cas9-NLS/guides NLF3 Pandora neocal	193455
vHB19	GFP-Cas9-NLS/guides MCP Molli kamtch	
vHB20	GFP-Cas9-NLS/guides Pneo-177 Pandora neocal	
vHB21	GFP-Cas9-NLS/guides pneo-585 Pandora neocal	
vHB22	GFP-Cas9-NLS/guides pneo-972 Pandora neocal	
vHB27	GFP-Cas9-NLS/guidesRFP for homologous recombination in RFP	
vHB29	MCP-RFP nouseo selection (molli)	
vHB30	RFP-MCP nouseo selection (molli)	
vHB32	GFP-Cas9-NLS/guides pneo-182 Pandora neocal	
vHB55	GFP-Cas9-NLS/guides pneo-62 Pandora neocal	
vHB56	GFP-Cas9-NLS/guides pneo-350 Pandora neocal	
vHB57	GFP-Cas9-NLS/guides pneo-450 Pandora neocal	
vHB58	GFP-Cas9-NLS/guides pneo-518 Pandora neocal	
vHB59	GFP-Cas9-NLS/guides pneo-959 Pandora neocal	
vHB66	3'UTRpneo480-pneo650-vc241 KO	193456
vHB72	GFP-Cas9-NLS/guides pneo-178 Pandora neocal	
vHB73	GFP-Cas9-NLS/guides pneo-179 Pandora neocal	
vHB74	GFP-Cas9-NLS/guides pneo-181 Pandora neocal	
vHB75	GFP-Cas9-NLS/guides pneo-537 Pandora neocal	
vHB76	GFP-Cas9-NLS/guides pneo-557 Pandora neocal	
vHB77	GFP-Cas9-NLS/guides pneo-575 Pandora neocal	
vHB93	GFP-Cas9-NLS/guides pneo-523 Pandora neocal	
vHB94	GFP-Cas9-NLS/guides pneo-528 Pandora neocal	
vHB95	GFP-Cas9-NLS/guides pneo-600 Pandora neocal	
vHB96	GFP-Cas9-NLS/guides pneo-700 Pandora neocal	
vHB97	GFP-Cas9-NLS/guides pneo-750 Pandora neocal	
vHB98	GFP-Cas9-NLS/guides pneo-850 Pandora neocal	

vHB99	GFP-Cas9-NLS/guides pneo-900 Pandora neocal
vHB100	GFP-Cas9-NLS/guides pneo-243 Pandora neocal
vHB101	GFP-Cas9-NLS/guides pneo-950 Pandora neocal
vHB102	GFP-Cas9-NLS/guides pneo-533 Pandora neocal
vHB103	GFP-Cas9-NLS/guides pneo-100 Pandora neocal
vHB104	GFP-Cas9-NLS/guides pneo-650 Pandora neocal
vHB105	GFP-Cas9-NLS/guides pneo-150 Pandora neocal
vHB106	GFP-Cas9-NLS/guides pneo-289 Pandora neocal
vHB107	GFP-Cas9-NLS/guides pneo-177 Pandora neocal
vHB108	GFP-Cas9-NLS/guides pneo-801 Pandora neocal
vHB113	GFP-Cas9-NLS/guides pneo-77 Pandora neocal
vHB114	GFP-Cas9-NLS/guides pneo-153 Pandora neocal
vHB115	GFP-Cas9-NLS/guides pneo-228 Pandora neocal
vHB116	GFP-Cas9-NLS/guides pneo-238 Pandora neocal
vHB117	GFP-Cas9-NLS/guides pneo-248 Pandora neocal
vHB118	GFP-Cas9-NLS/guides pneo-251 Pandora neocal
vHB119	GFP-Cas9-NLS/guides pneo-254 Pandora neocal
vHB120	GFP-Cas9-NLS/guides pneo-264 Pandora neocal
vHB121	GFP-Cas9-NLS/guides pneo-301 Pandora neocal
vHB122	GFP-Cas9-NLS/guides pneo-315 Pandora neocal
vHB123	GFP-Cas9-NLS/guides pneo-321 Pandora neocal
vHB124	GFP-Cas9-NLS/guides pneo-325 Pandora neocal
vHB125	GFP-Cas9-NLS/guides pneo-365 Pandora neocal
vHB126	GFP-Cas9-NLS/guides pneo-383 Pandora neocal
vHB127	GFP-Cas9-NLS/guides pneo-385 Pandora neocal
vHB128	GFP-Cas9-NLS/guides pneo-417 Pandora neocal
vHB129	GFP-Cas9-NLS/guides pneo-459 Pandora neocal
vHB136	GFP-Cas9-NLS/guides pneo-187 Pandora neocal
vHB137	GFP-Cas9-NLS/guides pneo-198 Pandora neocal
vHB138	GFP-Cas9-NLS/guides pneo-221 Pandora neocal
vHB139	GFP-Cas9-NLS/guides pneo-231 Pandora neocal
vHB140	GFP-Cas9-NLS/guides pneo-250 Pandora neocal
vHB141	GFP-Cas9-NLS/guides pneo-261 Pandora neocal
vHB142	GFP-Cas9-NLS/guides pneo-276 Pandora neocal
vHB143	GFP-Cas9-NLS/guides pneo-304 Pandora neocal
vHB144	GFP-Cas9-NLS/guides pneo-313 Pandora neocal
vHB145	GFP-Cas9-NLS/guides pneo-322 Pandora neocal
vHB146	GFP-Cas9-NLS/guides pneo-342 Pandora neocal
vHB147	GFP-Cas9-NLS/guides pneo-344 Pandora neocal
vHB148	GFP-Cas9-NLS/guides pneo-407 Pandora neocal
vHB149	GFP-Cas9-NLS/guides pneo-414 Pandora neocal
vHB150	GFP-Cas9-NLS/guides pneo-425 Pandora neocal
vHB151	GFP-Cas9-NLS/guides pneo-429 Pandora neocal

vHB152	GFP-Cas9-NLS/guides pneo-443 Pandora neocal	
vHB153	GFP-Cas9-NLS/guides pneo-544 Pandora neocal	
vHB154	GFP-Cas9-NLS/guides mk-236	
vHB155	cis-complementation MCP	
vc241	mRFP	193453
vHB178	vHB66 + 3xHA	193457
vHB179	vHB66 + 3xFLAG	193458

Source supplementary.

Fig S1b

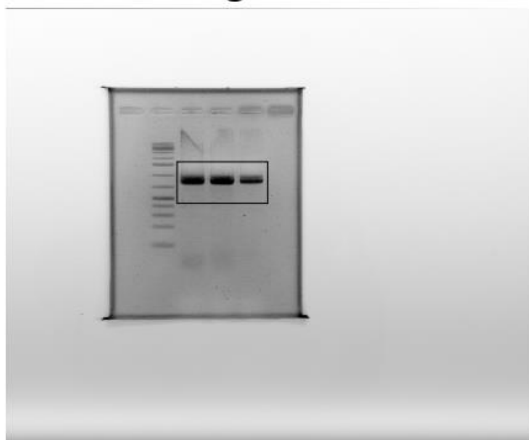


Fig S2c

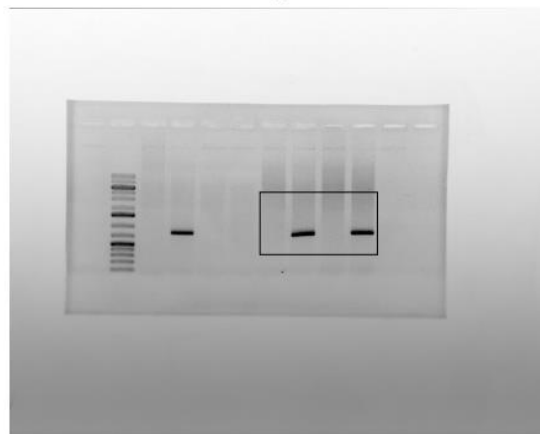


Fig S2d

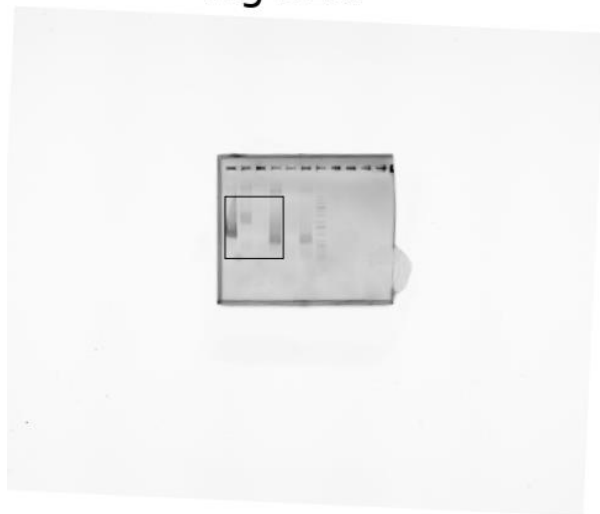


Fig S3a

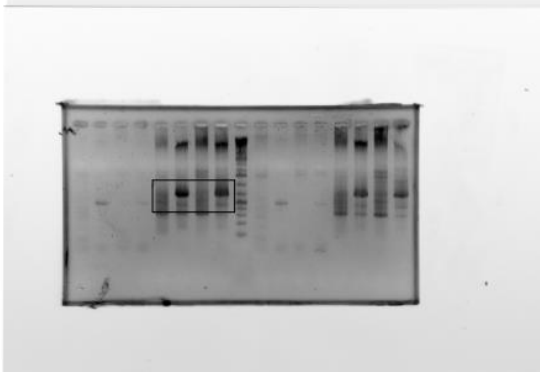


Fig S3a (cont)



Fig S4b

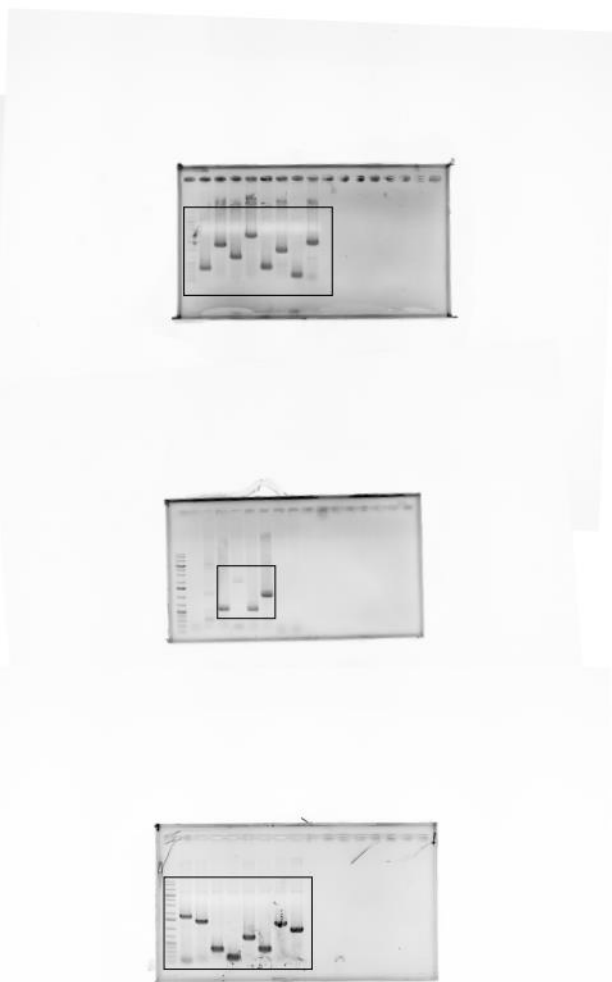


Fig S4b (cont)

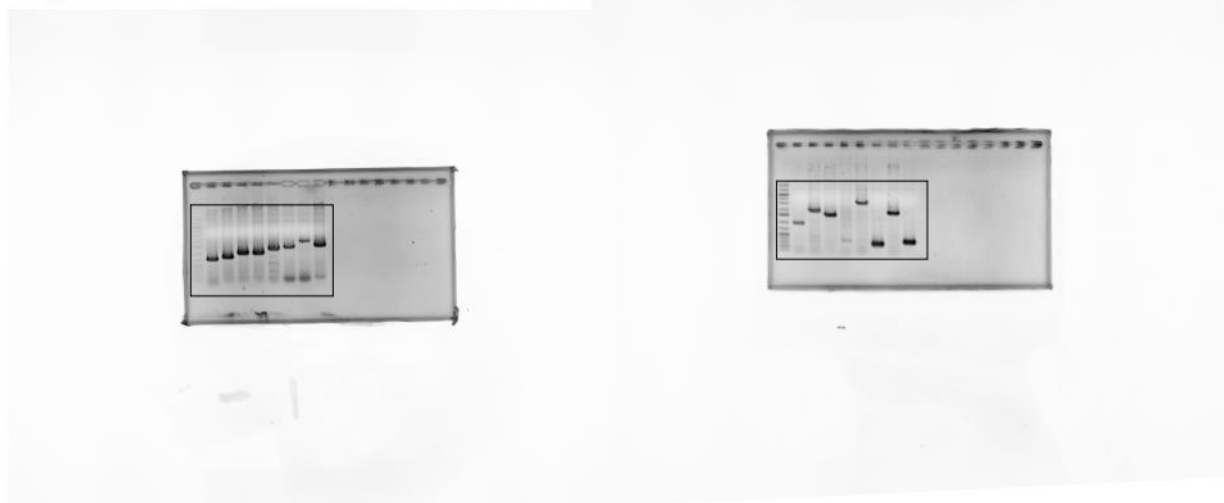


Fig S4b (cont)



Fig S6b

