

MMP-8 single-nucleotide polymorphisms are related to ankylosing spondylitis in Chinese Han population

Chenyang Meng, Master^{a,b}, Rui Bai, MD^b, Zhenqun Zhao, Master^b, Guimei Huang, Master^c, Tianbo Jin, PhD^d, Wei Feng, MD^{e,*}, Wanlin Liu, Master^{b,*}

Abstract

Ankylosing spondylitis (AS) is an extreme form of inflammatory arthritis which always leads to bony fusion of vertebral and chronic pain of back. A lot of genes including interleukin, matrix metalloproteinases (MMPs), and endoplasmic reticulum aminopeptidase were found associated with AS. MMP family members were involved in the autoimmune disease and orthopedic diseases such as rheumatoid arthritis and osteoarthritis, while few studies concentrated on the correlation between single-nucleotide polymorphisms (SNPs) in MMP and AS. In addition, there is no report on the relationship between *MMP-8* and AS. To investigate the association between SNPs in *MMP-8* and AS, we recruited 268 patients with AS and 654 healthy people to conduct a case–control study. Five SNPs including rs3740938, rs2012390, rs1940475, rs11225394, and rs11225395 of *MMP-8* gene were genotyped. It was found rs3740938 of *MMP-8* was associated with an increased risk of AS under the dominant model and additive model after adjustment for gender and age by performing logistic regression analysis (odds ratio [OR] = 1.49, 95% confidence interval [CI] = 1.02-2.18, P = .038; OR = 1.37, 95% CI = 1.01-1.87, P = .042, respectively). Moreover, haplotype "GGTCA" was associated with an increased risk of AS without adjustment for age and gender (OR = 1.75, 95% CI = 1.05-2.92, P = .032), while no positive result was found after adjustment for age and gender. Based on our results, our study indicates significant association between SNPs of *MMP-8* and AS.

Abbreviations: AS = ankylosing spondylitis, BMI = body mass index (weight [kg]/height [m]²), 95% CI = 95% confidence interval, ERAP = endoplasmic reticulum aminopeptidase, HWE = Hardy–Weinberg equilibrium, IL = interleukin, MAF = minor allele frequency, MMPs = matrix metalloproteinases, OA = osteoarthritis, OR = odds ratio, PsA = psoriaticarthritis, RA = rheumatoid arthritis, SNP = single-nucleotide polymorphism.

Keywords: ankylosing spondylitis, association study, genetic, MMP-8, single-nucleotide polymorphism

1. Introduction

Ankylosing spondylitis (AS) is an extreme form of inflammatory arthritis which always leads to bony fusion of vertebral and chronic pain of back.^[1] The prevalence of AS in Chinese Han

Editor: Wael Alkhiary.

This work was supported by National Natural Science Foundation of China grants (no: 81460331).

The authors have no conflicts of interest to disclose.

^a Department of Graduate School, Inner Mongolia Medical University, Hohhot, Inner Mongolia, China, ^b Department of Pediatric Orthopedics, The Second Affiliated Hospital of Inner Mongolia Medical University, Hohhot, Inner Mongolia, China, ^c Department of Administrative Affairs Office, The Second Affiliated Hospital of Guangxi Medical University, Nanning, Guangxi, China, ^d School of Life Sciences, Northwest University, Xi'an, Shaanxi, China, ^e Department of Pelvic and Acetabular Surgery, The Second Affiliated Hospital of Inner Mongolia Medical University, Hohhot, Inner Mongolia, China.

^{*} Correspondence: Wanlin Liu, Department of the 2nd Affiliated Hospital of Inner Mongolia Medical University, Hohhot, Inner Mongolia 010030, China (e-mails: 1015327682@qq.com, 603261696@qq.com); Wei Feng, Department of Pelvic and Acetabular Surgery, The Second Affiliated Hospital of Inner Mongolia Medical University, Hohhot, Inner Mongolia, China.

Copyright © 2018 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Medicine (2018) 97:35(e12136)

Received: 19 March 2018 / Accepted: 4 August 2018 http://dx.doi.org/10.1097/MD.000000000012136 ethnic population was about 0.2% to 0.5%, which is similar to white Europeans and American.^[2-4] In addition, it often occurs in people who aged before 40, with male predominance.^[3] There were 3 factors involved in the pathogenesis of AS. They were environmental triggers such as microbiota and mechanical stress, autoimmune factor such as autoreactive T cells, genetic risk.^[5] AS is one of the most common genetic diseases and it has high monozygotic twin concordance (63%); besides, familial aggregation studies indicate a heritability of over 90%.^[6] It was well known that HLA-B27 played a significant role in pathogenesis of AS. However, some reports argued that it only accounted for 20% to 25% of the total heritability and 40% of the genetic risk. Fewer than 5% of HLA-B27 carriers in the general population develop disease.^[6,7] It suggested that there existed other genetic susceptibility. A lot of genes including interleukin (IL), matrix metalloproteinases (MMPs), and endoplasmic reticulum aminopeptidase (ERAP) were found associated with AS.^[8-10]

The MMPs are zinc-dependent enzymes. MMPs are a family consisting of 23 protein members that could be involved in the degradation of osseous tissue and other extracellular matrices in the human body.^[11] It was reported that MMPs were key regulators of tissue degradation and remodeling.^[11] They were revealed as participating in the nosogenesis of several diseases of synovial joints such as rheumatoid arthritis (RA), osteoarthritis (OA), and AS.^[12] Several studies showed that AS was associated with high levels of MMP-3.^[13,14]*MMP-8* is the member of MMPs. Just like types I, II, and III collagens, it could degrade the cartilage proteoglycan, aggrecan, and involved in tissue remodeling.^[15,16]*MMP-8* was found to be correlated with RA and

CM and RB authors have contributed equally to this work.

osteonecrosis of the femoral head.^[17,18] Mattey et al found that bath AS disease activity index (BASDAI) was correlated with a group of clustered biomarkers consisting of MMP-8, MMP-9, hepatocyte growth factor, the chemokine, and CXCL8. However, there was no study revealed the association between MMP-8 and AS.^[19] In addition, the expression of MMP-8 is related to inflammatory cytokines, growth factors, and hormones.^[20] As we all know, AS is inflammatory arthritis, MMP family members were involved in the autoimmune disease and some kinds of orthopedic diseases such as RA, OA, psoriaticarthritis, osteonecrosis of femoral head, and so on.^[12,21] The expression of MMP genes were found to be associated with several proinflammatory cytokines, such as tumor necrosis factor-alfa (TNF- α) and IL-17.^[22,23] Recent studies have indicated that single-nucleotide polymorphisms (SNPs) in IL and ERAP are associated with an increased risk of AS,^[24,25] while few studies concentrated on the correlation between SNPs in MMP and AS. In addition, there is no report on the relationship between MMP-8 and AS. Based on those studies, we conducted a case-control study and genotyped 5 SNPs in MMP genes to investigate the association between SNPs in MMP-8 and AS.

2. Materials and methods

2.1. Research objects

A total of 268 patients with AS and 654 healthy people were recruited among Shaanxi Province. All the subjects we recruited were the Han nationality. All patients were treated by the Xi'an Honghui Hospital and were newly diagnosed AS by clinical features and examination of laboratory and radiology. Patients who had not yet received any treatment were included for the case group. People who suffered chronic metabolic disorder of the heart, kidney, or liver and other bone diseases were excluded. Individuals with other immune or inflammatory diseases were also excluded from our study. About 654 healthy unrelated subjects were recruited randomly as control group. Individuals are Han Chinese living Xi'an. Moreover, people with chronic disease involving bone, brain, liver, heart, and lung were excluded from our study. All samples were collected with informed consent and the study was approved by the regional ethics committee.

2.2. SNP selection and genotyping

We reviewed the literatures related to association between MMP-8 polymorphisms and orthopedic diseases, especially for the diseases with the similar pathologic changes as AS. In addition, SNPs associated with inflammatory response were also considered in our study. Of course, the selection of SNPs also depended on their location, allele frequencies, and disease relevance determined by use of the Hapmap public databases (dbSNP, http://www.ncbi.nlm.nih.gov/SNP/; HAPMAP, http://www.Hap map.org/index.html.en). Finally, selected SNPs in MMP-8 with the minor allele frequencies (MAFs) $\geq 5\%$ in Asian by using HapMap database. [17,26-28] In addition, the relationship between chosen SNPs and AS in Chinese Han population has not been reported before. Genomic DNA was extracted from whole blood samples using the Gold Mag-Mini Whole Blood Genomic DNA Purification Kit (version 3.0; TaKaRa, Tokyo, Japan). The DNA concentration was measured by spectrometry (DU530UV/VIS spectrophotometer; Beckman Instruments, Fullerton, CA). The Sequenom MassARRAY Assay Design 3.0 software (Sequenom, Inc, San Diego, CA) was used to design the multiplexed SNP Mass EXTEND assay. Genotyping was performed using a Sequenom MassARRAY RS1000 (Sequenom, Inc) in accordance with the manufacturer's protocol. Sequenom Typer 4.0 software was used to perform data management and analyses.^[29] Based on these results, 5 SNPs including rs3740938, rs2012390, rs1940475, rs11225394, and rs11225395 were selected.

2.3. Statistical analysis

The differences of gender and age between 2 groups were analyzed by 2-sided Chi-squared test and independent samples t test, respectively. We performed an exact test to examine Hardy-Weinberg equilibrium (HWE) in case and control groups. Minor alleles of SNPs were seemed as risk alleles for AS susceptibility. The differences in frequency distributions of alleles were compared between cases and controls by Pearson Chi-squared test. Odds ratios (ORs), 95% confidence intervals (CIs), and Pvalue were used for logistic regression analysis and we performed the Wald test by unconditional logistic regression analysis so that the adjustment for age and sex were done for the dominant, recessive, codominant, and log-additive models. We used the Haploview software package (version 4.2) and the SHEsi software platform to analyze the linkage disequilibrium and SNP haplotypes.^[30,31] A logistic regression analysis was performed to assess haplotype association with response. SPSS version 22.0 statistical package (SPSS, Chicago, IL) and Microsoft Excel were used for all statistical analyses. P < .05 was considered statistically significant.

3. Result

3.1. Participant characteristics

In our study, we recruited 268 patients with AS and 654 healthy people. Basic characteristics of the control individuals and patients with AS are shown in Table 1. There were statistical significance differences in age between groups of case and control while no significant difference in gender.

3.2. Hardy-Weinberg equilibrium test

Our study reveals that genotype distributions in cases and controls accorded with HWE for *MMP-8* gene rs3740938, rs2012390, rs1940475, rs11225394, and rs11225395 sites at Table 2, indicating that samples were representative.

3.3. Association between genetic polymorphisms of MMP-8 and AS risk

The detail information including position, band, MAF of candidate SNPs is summarized in Table 2. As we can see, the

_	
	68
1.5.1	

Basic characteristics of patients with AS and control individuals.								
Characteristic	Case (N = 268)	Controls (N=654)	Р					
Mean age \pm SD Gender	$31.64 \pm 12.251 (N = 268)$	$49.72 \pm 10.256 (N = 654)$.001*					
Male Female	200 68	464 190	.259†					

AS = ankylosing spondylitis, BMI=body mass index (weight [kg]/height [m]²), SD=standard deviation.

P < .05 indicates statistical significance.

^{*} P-value was calculated by Welch t test and Levene Variance Equality Test.

[†] P-value was calculated by Pearson Chi-squared test.

Table 2

Candidate SN	IPs examined in I	MMP-8 gene.			Ν	/AF		
SNP ID	Position	Band	Alleles A/B	HWE, <i>P</i> [*]	Case	Control	OR (95% CI)	P [†]
rs3740938	102587062	11q22.2	A/G	.3618	0.244	0.221	1.14 (0.9–1.45)	.267
rs2012390	102590777	11g22.2	G/A	.3564	0.274	0.255	1.10 (0.87–1.38)	.401
rs1940475	102593248	11g22.2	T/C	.5458	0.384	0.346	1.18 (0.96–1.45)	.122
rs11225394	102595413	11g22.2	T/C	.3481	0.108	0.093	1.17 (0.84–1.64)	.343
rs11225395	102596480	11q22.2	A/G	.6011	0.381	0.340	1.19 (0.97–1.47)	.099

95% CI = 95% confidence interval, HWE = Hardy-Weinberg equilibrium, MAF = minor allele frequency, OR = odds ratio, SNP = single-nucleotide polymorphism.

* P-value was calculated by exact test.

[†] *P*-value was calculated by Pearson Chi-squared test.

P < .05 indicates statistical significance.

MAF of each SNP was >0.05. However, there was no significant difference between case group and control group in the allele model.

We further assessed the association between each chosen SNP and AS risk under 4 models including codominant, dominant, recessive, and additive model a (Tables 3 and 4). We found rs3740938 of MMP-8 was associated with an increased risk of AS under the dominant model and additive model after adjustment for gender and age by performing logistic regression analysis (OR=1.49, 95% CI=1.02–2.18, P=.038; OR=1.37, 95% CI=1.01–1.87, P=.042, respectively). Unfortunately, there was no positive result showed statistically significant difference in other SNPs we chose.

Furthermore, there was a strong linkage between the chosen SNPs in the *MMP-8* gene (Fig. 1). We further assess the correlation between haplotype and risk of AS. As shown in Table 5, haplotype "GGTCA" was associated with an increased risk of AS without adjustment for age and gender (OR=1.75, 95% CI=1.05–2.92, P=.032). Moreover, we performed an unconditional logistic regression adjusted for age and gender while no positive result found.

Table 3

The association	between	the	SNPs	and	AS	risk	in	codominant
model.								

SNP ID	Genetype	Case	Control	OR (95% CI)	Р
rs3740938	GG	156	401	1.00 [Ref]	.11
	AG	93	216	1.47 (0.99-2.19)	
	AA	19	36	1.65 (0.74-3.67)	
rs2012390	AA	144	367	1.00 [Ref]	.23
	AG	101	240	1.39 (0.94-2.06)	
	GG	23	47	1.36 (0.66-2.83)	
rs1940475	CC	99	283	1.00 [Ref]	.25
	CT	132	289	1.36 (0.91-2.03)	
	TT	37	82	1.45 (0.79-2.64)	
rs11225394	CC	211	532	1.00 [Ref]	.58
	CT	51	115	0.92 (0.57-1.49)	
	TT	3	3	2.87 (0.34-24.09)	
rs11225395	GG	101	288	1.00 [Ref]	.22
	AG	130	287	1.37 (0.92-2.04)	
	AA	37	79	1.48 (0.81–2.69)	

95% CI=95% confidence interval, MAF=minor allele frequency, OR=odds ratio, SNP=singlenucleotide polymorphism.

P-values were calculated by Wald test by unconditional logistic regression adjusted for age and gender.

P<.05 indicates statistical significance.

4. Discussion

As we know, HLA-B27 is strongly correlated with AS. HLA-B27 allele was found to be a classic genetic marker for predicting the development of AS.^[32] Just like HLA-B27, important biomarkers such as C-reactive protein and erythrocyte sedimentation rate were associated with AS.^[32] Interestingly, several studies showed MMP family was evidence of biomarkers associated with AS. The study of Maksymowych et al showed high levels of serum MMP-3 could be able to predict the degree of structural damage in patients with AS.^[33] In addition, the gene polymorphisms on MMP-2 and MMP-1 were found to be correlated with AS.^[34,35] Although the associations between MMP-3, MMP-1, MMP-2, and AS were reported in many studies, [36-38] there were few studies reported the correlation between MMP-8 and AS. In our study, we found rs3740938 in MMP-8 was associated with an increased risk of AS under the dominant model and log-additive model. We were the first study revealed the relationship between MMP-8 and AS. Although the previous study suggested BASDAI was found correlated strongly with a component consisting of MMP-8, MMP-9, hepatocyte growth factor, and CXCL8, there was no research on the association between MMP-8 and AS alone.^[19]MMP-8 might play a significant role in the pathogenesis of AS with or without the other MMPs involving based on our result.

The MMP-8 gene was localized to chromosome 11q22.2. It was known as neutrophil collagenase and collagenase-2. MMP-8 was found to be associated with RA and osteonecrosis of the femoral head.^[17,18] It is well known that the pathologic changes of RA and osteonecrosis of the femoral head include cartilage destruction and tissue remodeling as well as AS. In addition, MMP-8 was found to be as an important role in degrading the cartilage proteoglycan and aggrecan.^[15] In addition, MMP-8 was revealed as a significant role in tissue remodeling.^[16,39] AS is manifested as attachment point inflammation with abnormal ossification and fibrosis so that the spine become rigid. A study on the acute allergic rhinitis showed MMP-8 may contribute to osteogenesis and fibrosis while no study revealed this result on the research of AS.^[39] Our study suggested gene polymorphism MMP-8 did correlated with AS, the detail interaction between MMP-8 and AS was not clear vet.

The MMP-8 is a potent collagenolytic enzyme which is involved in the pathogenesis of several inflammatory conditions. Thirkettle and colleagues found that MMP-8 induces the expression of IL-6 and IL-8 in breast cancer cells and García et al's research showed deficiency of MMP-8 increases joint inflammation and bone erosion in the K/BxN serum-transfer arthritis model.^[18,40] It could be noticed that MMP-8 did affect several inflammatory factors. In fact, several cytokines such as

Table 4

Single loci association with AS risk.

		Dominant model		Recessive mod	el	Additive model			
SNP ID	Minor allele	OR (95% CI)	Р	P'	OR (95% CI)	Р	OR (95% CI)	Р	P'
rs3740938	А	1.49 (1.02-2.18)	.038*	.19	1.43 (0.66-3.12)	.37	1.37 (1.01–1.87)	.042*	.21
rs2012390	G	1.39 (0.95-2.02)	.086	.43	1.19 (0.58-2.40)	.64	1.26 (0.94-1.70)	.12	.6
rs1940475	Т	1.38 (0.94-2.01)	.097	.485	1.22 (0.70-2.14)	.48	1.25 (0.95-1.64)	.12	.6
rs11225394	Т	0.97 (0.60-1.55)	.89	NA	2.91 (0.35-24.35)	.32	1.02 (0.65-1.59)	.94	NA
rs11225395	А	1.39 (0.95-2.04)	.085	.425	1.25 (0.71-2.18)	.44	1.26 (0.95-1.66)	.1	.5

95% CI=95% confidence interval, OR=odds ratio, SNP=single nucleotide polymorphism.

P-values were calculated by Wald test by unconditional logistic regression adjusted for age and gender.

P < .05 indicates statistical significance.

P'-values were calculated by performing the Bonferroni correction.



Figure 1. Haplotype block map and the locations for chosen single-nucleotide polymorphisms of the *MMP-8* gene.

TNF- α , interleukin-1 β (IL-1 β), IL-6, IL-7, and IL-8 activate signal transduction pathways to regulate MMP gene expression.^[41,42] A review reported by Malemud suggested high level of IL-6 in the sera and synovial fluid of patients with RA is likely to be responsible for the upregulation of the MMP-9 gene as well as other MMPs that are relevant to RA as well as the degradation of cartilage proteins which is characteristic of RA pathology.^[12] AS is deemed as disease of immune system and the pathogenic mechanisms of AS involve several cytokines including IL-17, TNF- α , and so on.^[43,44] Gonzalez-Lopez et al's study reported patients with AS had higher serum TNF- α , while there was no significant difference on serum IL-6 between 2 groups.^[43] In addition, it was found TNF blockers inhibit spinal radiographic progression in AS by reducing disease activity.^[45] Our result agreed with the previous studies. Based on our result, it could be assume that cytokines regulate MMP-8 gene expression by activating signal transduction pathways and expression of MMP-8 reacts up on the cytokines to involve in the pathogenesis of AS. However, it should be further explored.

Just like all studies, our study has some potential limitations. First of all, this study is limited by its sample size, the further association should be confirmed finally by performing a large sample size meta-analysis. Secondly, we just collected the basic characteristics of the individuals such as age and sex. The environmental and life style factors were not included in our study but we will add those factors in further research. Furthermore, clinical characteristics including the stage of disease and detail pathologic changes were not included in our study, and it is needed to be further analyzed through additional studies. Of course, we did not perform the function study of MMP-8 due to

Table 5

Haplotype association with response (n=922).

							Without adjustment		With adjustment		
	rs3740938	rs2012390	rs1940475	rs11225394	rs11225395	Freq	OR (95% CI) *	P [*]	OR (95% CI) †	P [†]	
1	G	А	С	С	G	0.634	1	NA	1	NA	
2	А	G	Т	С	А	0.2196	1.12 (0.87-1.43)	0.38	1.33 (0.96-1.84)	.084	
3	G	А	Т	Т	А	0.0965	1.36 (0.96-1.92)	0.086	1.16 (0.73–1.83)	.52	
4	G	G	Т	С	А	0.0345	1.75 (1.05–2.92)	0.032*	1.39 (0.71–2.72)	.34	

Global haplotype association ^aP-value: .035, ^bP-value: .26.

95% CI=95% confidence interval, Freq=frequency, OR=odds ratio, SNP=single-nucleotide polymorphism.

^a*P* value was without adjustment for age and gender which was the same meaning as ^{*}*P*-value.

^bP value was adjusted by age and gender which was the meaning as [†]P-value.

P<.05 indicates statistical significance.

the purpose of our study was to explore whether gene polymorphism of *MMP-8* was associated with AS. However, the detail relationship between expression of *MMP-8* gene and other cytokines will be investigated in our future study based on our present result. Last, although our result showed only rs3740938 in *MMP-8* was associated with an increased risk of AS, we revealed a relationship between *MMP-8* and AS which was not reported in previous study.

To sum up, we have demonstrated that rs3740938 in *MMP-8* gene was associated with risk of AS in Chinese Han population for the first time. Our study may provide new data for screening of AS in Han population and could be used as diagnostic and prognostic markers in clinical studies of patients with AS.

Acknowledgments

The authors are grateful to the patients and control individuals for their participation in the study. The authors also thank the clinicians and hospital staff who contributed to sample and data collection for this study.

Author contributions

Conceptualization: Chenyang Meng, Wanlin Liu, Rui Bai.

Data curation: Chenyang Meng, Wei Feng, Rui Bai, Zhenqun Zhao.

Formal analysis: Chenyang Meng, Zhenqun Zhao.

- Funding acquisition: Rui Bai.
- Investigation: Chenyang Meng.
- Methodology: Chenyang Meng, Wei Feng, Zhenqun Zhao, Guimei Huang.

Project administration: Chenyang Meng, Wei Feng, Tianbo Jin. Resources: Chenyang Meng, Rui Bai, Tianbo Jin, Guimei Huang. Software: Chenyang Meng.

Supervision: Chenyang Meng, Tianbo Jin.

Validation: Chenyang Meng, Tianbo Jin.

Visualization: Chenyang Meng.

Writing - original draft: Chenyang Meng, Rui Bai.

Writing – review & editing: Chenyang Meng, Wanlin Liu, Wei Feng.

References

- Campochiaro C, Caruso PF. Ankylosing spondylitis and axial spondyloarthritis. N Engl J Med 2016;375:1302.
- [2] Zeng QY, Chen R, Darmawan J, et al. Rheumatic diseases in China. Arthritis Res Ther 2008;10:R17.
- [3] Reveille JD, Weisman MH. The epidemiology of back pain, axial spondyloarthritis and HLA-B27 in the United States. Am J Med Sci 2013;345:431.
- [4] Thomas GP, Brown MA. Genetics and genomics of ankylosing spondylitis. Immunol Rev 2010;233:162.
- [5] Smith JA. Update on ankylosing spondylitis: current concepts in pathogenesis. Curr Allergy Asthma Rep 2015;15:489.
- [6] Brown MA, Kennedy LG, Macgregor AJ, et al. Susceptibility to ankylosing spondylitis in twins the role of genes, HLA, and the environment. Arthritis Rheum 1997;40:1823.
- [7] Cortes A, Hadler J, Pointon JP, et al. Identification of multiple risk variants for ankylosing spondylitis through high-density genotyping of immune-related loci. Nat Genet 2013;45:730–8.
- [8] Sarin R, Wu X, Abraham C. Inflammatory disease protective R381Q IL23 receptor polymorphism results in decreased primary CD4+ and CD8+ human T-cell functional responses. Proc Natl Acad Sci U S A 2011;108:9560–5.
- [9] Consortium AAAS, Evans DM, Spencer CCA, et al. Interaction between ERAP1 and HLA-B27 in ankylosing spondylitis implicates peptide handling in the mechanism for HLA-B27 in disease susceptibility. Nat Genet 2011;43:761.

- [10] Jadon DR, Sengupta R, Nightingale A, et al. Serum bone-turnover biomarkers are associated with the occurrence of peripheral and axial arthritis in psoriatic disease: a prospective cross-sectional comparative study. Arthritis Res Ther 2017;19:210.
- [11] Nagase H, Visse R, Murphy G. Structure and function of matrix metalloproteinases and TIMPs. Cardiovasc Res 2006;69:562–73.
- [12] Malemud CJ. Matrix Metalloproteinases and Synovial Joint Pathology. Prog Mol Biol Transl Sci 2017;148:305.
- [13] Zhu J, Yu DT. Matrix metalloproteinase expression in the spondyloarthropathies. Curr Opin Rheumatol 2006;18:364–8.
- [14] Chen CH, Yu DTY, Chou CT. Biomarkers in spondyloarthropathies. Adv Exp Med Biol 2009;649:122–32.
- [15] Fosang AJ, Last K, Neame PJ, et al. Neutrophil collagenase (MMP-8) cleaves at the aggrecanase site E373-A374 in the interglobular domain of cartilage aggrecan. Biochem J 1994;304:347–51.
- [16] Gutiérrezfernández A, Fueyo A, Folgueras AR, et al. Matrix metalloproteinase-8 functions as a metastasis suppressor through modulation of tumor cell adhesion and invasion. Cancer Res 2008;68:2755–63.
- [17] An F, Du J, Cao Y, et al. MMP8 polymorphism is associated with susceptibility to osteonecrosis of the femoral head in a Chinese Han population. Oncotarget 2017;8:21561–6.
- [18] García S, Forteza J, López-Otin C, et al. Matrix metalloproteinase-8 deficiency increases joint inflammation and bone erosion in the K/BxN serum-transfer arthritis model. Arthritis Res Ther 2010;12:R224.
- [19] Mattey DL, Packham JC, Nixon NB, et al. Association of cytokine and matrix metalloproteinase profiles with disease activity and function in ankylosing spondylitis. Arthritis Res Ther 2012;14:R127.
- [20] Nagase HH, Woessner JF. Matrix metalloproteinases. J Biol Chem 1999;274:21491–4.
- [21] Yu Y, Xie Z, Wang J, et al. Single-nucleotide polymorphisms of MMP2 in MMP/TIMP pathways associated with the risk of alcohol-induced osteonecrosis of the femoral head in Chinese males: a case–control study. Medicine 2016;95:e5407.
- [22] Meszaros EC, Dahoud W, Mesiano S, et al. Blockade of recombinant human IL-6 by tocilizumab suppresses matrix metalloproteinase-9 production in the C28/I2 immortalized human chondrocyte cell line. Integr Mol Med 2015;2:304.
- [23] Moran EM, Mullan R, Mccormick J, et al. Human rheumatoid arthritis tissue production of IL-17A drives matrix and cartilage degradation: synergy with tumour necrosis factor-Oncostatin M and response to biologic therapies. Arthritis Res Ther 2009;11:R113.
- [24] Yang B, Xu Y, Liu X, et al. IL-23R and IL-17A polymorphisms correlate with susceptibility of ankylosing spondylitis in a southwest Chinese population. Oncotarget 2017;8:70310–6.
- [25] Popa OM, Cherciu M, Cherciu LI, et al. ERAP1 and ERAP2 gene variations influence the risk of psoriatic arthritis in romanian population. Arch Immunol Ther Exp 2017;64:1–7.
- [26] Du J, Jin T, Cao Y, et al. Association between genetic polymorphisms of MMP8 and the risk of steroid-induced osteonecrosis of the femoral head in the population of northern China. Medicine 2016;95:e4794.
- [27] Rella JM, Jilma B, Fabry A, et al. MMP-8 genotypes influence the inflammatory response in human endotoxemia. Inflammation 2014; 37:451–6.
- [28] Chen J, Liu W, Cao Y, et al. MMP-3 and MMP-8 single-nucleotide polymorphisms are related to alcohol-induced osteonecrosis of the femoral head in Chinese males. Oncotarget 2017;8:25177–88.
- [29] Gabriel S, Ziaugra L, Tabbaa D. SNP genotyping using the Sequenom MassARRAY iPLEX platform. Curr Protocols in Human Genetics 2009; Chapter 2(Unit 2):Unit 2.12.
- [30] Barrett JC, Fry B, Maller J, et al. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005;21:263–5.
- [31] Shi YY, He L. SHEsis, a powerful software platform for analyses of linkage disequilibrium, haplotype construction, and genetic association at polymorphism loci. Cell Res 2005;15:97–8.
- [32] Reveille JD. Biomarkers for diagnosis, monitoring of progression, and treatment responses in ankylosing spondylitis and axial spondyloarthritis. Clin Rheumatol 2015;34:1009–18.
- [33] Maksymowych WP, Landewé R, Conner-Spady B, et al. Serum matrix metalloproteinase 3 is an independent predictor of structural damage progression in patients with ankylosing spondylitis. Arthritis Rheum 2007;56:1846–53.
- [34] Cortes A, Danoy P, Wordsworth B, et al. MMP1 polymorphisms are associated with severity of radiographic measures of ankylosing spondylitis. Int Med J 2011;41:22–122.
- [35] Sun R, Huang Y, Zhang H, et al. MMP 2, TNF and NLRP1 polymorphisms in Chinese patients with ankylosing spondylitis and rheumatoid arthritis. Mol Biol Rep 2013;40:6303–8.

- [36] Gao JW, Zhang KF, Lu JS, et al. Serum matrix metalloproteinases-3 levels in patients with ankylosing spondylitis. Genet Mol Res 2015; 14:17068–78.
- [37] Ravani A, Vincenzi F, Bortoluzzi A, et al. Role and function of A2A and A3 adenosine receptors in patients with ankylosing spondylitis, psoriatic arthritis and rheumatoid arthritis. Int J Mol Sci 2017;18:697.
- [38] Sun R, Huang Y, Zhang H, et al. MMP-2, TNF-(and NLRP1 polymorphisms in Chinese patients with ankylosing spondylitis and rheumatoid arthritis. Mol Biol Rep 2013;40:6303–8.
- [39] Sautter NB, Delaney KL, Trune DR. Altered expression of tissue remodeling genes in a mouse model of acute allergic rhinitis. Int Forum Allergy Rhinol 2011;1:262–7.
- [40] Thirkettle S, Decock J, Arnold H, et al. Matrix metalloproteinase 8 (collagenase 2) induces the expression of interleukins 6 and 8 in breast cancer cells*. J Biol Chem 2013;288:16282–94.

- [41] Tanaka T, Kishimoto T. Immunotherapy of tocilizumab for rheumatoid arthritis. J Clin Cell Immunol 2013;1:
- [42] Mertens M, Singh JA. Anakinra for rheumatoid arthritis: a systematic review. J Rheumatol 2009;36:1118–25.
- [43] Gonzalez-Lopez L, Fajardo-Robledo NS, Miriam Saldaña-Cruz A, et al. Association of adipokines, interleukin-6, and tumor necrosis factor-(concentrations with clinical characteristics and presence of spinal syndesmophytes in patients with ankylosing spondylitis: a cross-sectional study. J Int Med Res 2017;45:1024–35.
- [44] Taurog JD, Chhabra A, Colbert RA. Ankylosing spondylitis and axial spondyloarthritis. N Engl J Med 2016;375:1302.
- [45] Molnar C, Scherer A, Baraliakos X, et al. TNF blockers inhibit spinal radiographic progression in ankylosing spondylitis by reducing disease activity: results from the Swiss Clinical Quality Management cohort. Ann Rheum Dis 2018;77:63–9.