



## Complete Genome Sequence of the *Campylobacter ureolyticus* Clinical Isolate RIGS 9880

## William G. Miller,<sup>a</sup> Emma Yee,<sup>a</sup> Stephen L. W. On,<sup>b</sup> Leif P. Andersen,<sup>c</sup> James L. Bono<sup>d</sup>

Produce Safety and Microbiology Research Unit, Agricultural Research Service, U.S. Department of Agriculture, Albany, California, USA<sup>a</sup>; Institute of Environmental Science and Research Limited, Christchurch Science Centre, Christchurch, New Zealand<sup>b</sup>; Department of Clinical Microbiology, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark<sup>c</sup>; Meat Safety and Quality Research Unit, Agricultural Research Service, U.S. Department of Agriculture, Clay Center, Nebraska, USA<sup>d</sup>

The emerging pathogen *Campylobacter ureolyticus* has been isolated from human and animal genital infections, human periodontal disease, domestic and food animals, and from cases of human gastroenteritis. We report the whole-genome sequence of the human clinical isolate RIGS 9880, which is the first closed genome for *C. ureolyticus*.

Received 18 September 2015 Accepted 25 September 2015 Published 5 November 2015

Citation Miller WG, Yee E, On SLW, Andersen LP, Bono JL. 2015. Complete genome sequence of the *Campylobacter ureolyticus* clinical isolate RIGS 9880. Genome Announc 3(6):e01291-15. doi:10.1128/genomeA.01291-15.

Copyright © 2015 Miller et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to William G. Miller, william.miller@ars.usda.gov.

The emerging organism *Campylobacter ureolyticus* (formerly *Bacteroides ureolyticus* [1]) was initially isolated from superficial soft tissue lesions, and genital and periodontal infections (2–4). However, *C. ureolyticus* has been increasingly isolated from cases of human gastroenteritis (5), including patients with ulcerative colitis (6) and Crohn's disease (7). *C. ureolyticus* strain RIGS 9880 was isolated from a fecal sample from an immunocompromised patient with diarrhea at a tertiary hospital in Denmark. In this study, we report the whole-genome sequence of strain RIGS 9880, which represents the first closed *C. ureolyticus* genome.

Three next-generation sequencing platforms were used to complete the *C. ureolyticus* strain RIGS 9880 genome: Roche GS-FLX, Illumina HiSeq, and PacBio RS. A total of 533,763 shotgun and paired-end Roche 454 reads (101× coverage) were assembled, using the Roche Newbler assembler version 2.6, into a single scaffold of 12 contigs. Sanger DNA sequencing of contig-bridging amplicons was used to close the scaffold into a single contig. All 454 base calls were validated using 20,908,072 Illumina HiSeq reads (SeqWright, Houston, TX, USA), adding a coverage of 1,237×. An optical restriction map (OpGen, Gaithersburg, MD, USA) with restriction enzyme XbaI was used to validate the assembly. Finally, the domain structure of a large hypervariable adhesin-encoding gene was confirmed with PacBio Continuous Long Reads, adding a further  $502 \times$  coverage (1,840× total coverage).

*C. ureolyticus* strain RIGS 9880 has a circular genome of 1,642 kb with a GC content of 29.23%. Protein-, rRNA- and tRNA-encoding genes were identified as described previously (8). The genome encodes 1,595 putative protein-coding genes, 24 pseudogenes, and 3 rRNA operons. No plasmids or hypervariable homopolymeric GC tracts ( $\geq$ 8 bp) were identified.

The RIGS 9880 genome contains several genes and features identified previously in *C. ureolyticus* (9), including a zonula occludens toxin genomic island (10, 11) and a large YadA-domain adhesin (9). In strain RIGS 9880, this adhesin gene comprises unique N- and C-terminal regions flanking a central region com-

posed of tandemly repeated 203- and 460-bp domains. PCR amplification and PacBio sequencing indicates that this central region is hypervariable: within a population, genes are present containing a range of repeats, from a single 203-bp domain to 14½ 203-bp/460-bp domain pairs.

Consistent with the aflagellate nature of this species, strain RIGS 9880 does not contain genes encoding flagellar, flagellar modification, or chemotaxis proteins. Another noteworthy feature of strain RIGS 9880 is the absence of key glutamine amidotransferases (HisF, CarA, TrpG) related to amino acid (histidine, tryptophan, arginine) and pyrimidine biosynthesis. It is possible that C. ureolyticus uses ammonia derived from urease activity directly in these reactions. The pyrimidine biosynthetic pathway of C. ureolyticus strain RIGS 9880 is also unusual in that the PyrE and PyrF enzymes form a clade that is evolutionarily distinct from orthologs encoded by the remainder of Campylobacter. The most similar orthologs to C. ureolyticus PyrE and PyrF are those encoded by members of the Veillonellaceae. These features were identified also in the draft genome of the C. ureolyticus strain ACS-301-V-Sch3b (data not shown), suggesting that they are general characteristics of C. ureolyticus.

**Nucleotide sequence accession number.** The complete genome sequence of *C. ureolyticus* strain RIGS 9880 has been deposited in GenBank under the accession number CP012195.

## ACKNOWLEDGMENT

This work was supported in part by USDA-ARS CRIS project 2030-42000-047-00D.

## REFERENCES

- 1. Vandamme P, Debruyne L, De Brandt E, Falsen E. 2010. Reclassification of *Bacteroides ureolyticus* as *Campylobacter ureolyticus* comb. nov., and emended description of the genus *Campylobacter*. Int J Syst Evol Microbiol 60:2016–2022. http://dx.doi.org/10.1099/ijs.0.017152-0.
- Duerden B, Bennet KW, Faulkner J. 1982. Isolation of *Bacteroides ureolyticus* (*B corrodens*) from clinical infections. J Clin Pathol 35:309–312. http://dx.doi.org/10.1136/jcp.35.3.309.

- Duerden BI, Eley A, Goodwin L, Magee JT, Hindmarch JM, Bennett KW. 1989. A comparison of *Bacteroides ureolyticus* isolates from different clinical sources. J Med Microbiol 29:63–73. http://dx.doi.org/10.1099/ 00222615-29-1-63.
- 4. Woolley PD, Kinghorn GR, Bennett KW, Eley A. 1992. Significance of *Bacteroides ureolyticus* in the lower genital tract. Int J STD AIDS 3:107–110.
- Bullman S, Corcoran D, O'Leary J, Lucey B, Byrne D, Sleator RD. 2011. Campylobacter ureolyticus: an emerging gastrointestinal pathogen? FEMS Immunol Med Microbiol 61:228–230. http://dx.doi.org/10.1111/j.1574 -695X.2010.00760.x.
- Mukhopadhya I, Thomson JM, Hansen R, Berry SH, El-Omar EM, Hold GL. 2011. Detection of *Campylobacter concisus* and other *Campylobacter* species in colonic biopsies from adults with ulcerative colitis. PLoS One 6:e21490. http://dx.doi.org/10.1371/journal.pone.0021490.
- Man SM, Zhang L, Day AS, Leach ST, Lemberg DA, Mitchell H. 2010. Campylobacter concisus and other Campylobacter species in children with newly diagnosed Crohn's disease. Inflamm Bowel Dis 16:1008–1016. http://dx.doi.org/10.1002/ibd.21157.

- Miller WG, Yee E, Chapman MH, Smith TPL, Bono JL, Huynh S, Parker CT, Vandamme P, Luong K, Korlach J. 2014. Comparative genomics of the *Campylobacter lari* group. Genome Biol Evol 6:3252–3266. http://dx.doi.org/10.1093/gbe/evu249.
- Bullman S, Lucid A, Corcoran D, Sleator RD, Lucey B. 2013. Genomic investigation into strain heterogeneity and pathogenic potential of the emerging gastrointestinal pathogen *Campylobacter ureolyticus*. PLoS One 8:e71515. http://dx.doi.org/10.1371/journal.pone.0071515.
- Mahendran V, Tan YS, Riordan SM, Grimm MC, Day AS, Lemberg DA, Octavia S, Lan R, Zhang L. 2013. The prevalence and polymorphisms of zonula occluden toxin gene in multiple *Campylobacter concisus* strains isolated from saliva of patients with inflammatory bowel disease and controls. PLoS One 8:e75525. http://dx.doi.org/10.1371/ journal.pone.0075525.
- Yap K, Gan H, Teh CS, Chai L, Thong K. 2014. Comparative genomics of closely related *Salmonella enterica* serovar typhi strains reveals genome dynamics and the acquisition of novel pathogenic elements. BMC Genomics 15:1007. http://dx.doi.org/10.1186/1471-2164-15-1007.