



Review

# Innovative Approaches in the Synthesis and Optimization of Copper Complexes for Antitumor Therapies: A Comprehensive Review

Clara Maria Faria Silva <sup>1,†</sup>, Ricardo Campos Lino <sup>1,†</sup>, Mariana Cristina Teixeira de Moura <sup>1</sup>, Anna Paula de Sá Borges <sup>2</sup> and Robson José de Oliveira Júnior <sup>1,\*</sup>

- Laboratory of Cytogenetics, Institute of Biotechnology, Federal University of Uberlândia, Campus Umuarama, St. Piaui s/n, Uberlândia 38405-320, MG, Brazil; ricardo.lino@ufu.br (R.C.L.)
- Academic Institute of health and biological Sciencies, State University of Goiás, UnU Itumbiara, Av. Modesto de Carvalho, s/n, District Agro. Industrial, Itumbiara 75536-100, GO, Brazil
- \* Correspondence: oliveirajunior@ufu.br
- <sup>†</sup> These authors contributed equally to this work.

Abstract: Cancer is the second leading cause of death worldwide. Late diagnosis, low drug selectivity, high toxicity, and treatment resistance are challenges associated with pharmacological interventions. The commonly used therapies include surgery, radiotherapy, hormonal therapy, immunotherapy, and chemotherapy. Recently, Cu complexes have been studied owing to their biological functions and effects on tumor angiogenesis. In this review, we examined 23 types of cancer and revealed the use of cell lines. The synthesis of Cu complexes with ligands such as phenanthroline and thiosemicarbazones has also been reported. Such co-ligation is promising because of its high cytotoxicity and selectivity. Compared with cisplatin, Cu complexes, especially mixed complexes, showed better interactions with DNA, generating reactive oxygen species and inducing apoptosis. Nanoformulations have also been adopted to improve the pharmacological activity of compounds. They enhance the efficacy of complexes by targeting them to the tumor tissue, thereby improving their safety. Studies have also explored Cu complexes with clinically relevant pharmacophores, suggesting a "hybrid chemotherapy" against resistant tumors. Overall, Cu complexes have demonstrated therapeutic versatility, antitumor efficacy, and reduced adverse effects, showing great potential as alternatives to conventional chemotherapy and justifying future clinical investigations to validate their use.

Keywords: anticancer; copper complexes; cytotoxic activity



Academic Editors: Franco Bisceglie, Piotr Świątek and Edward Krzyżak

Received: 13 March 2025 Revised: 6 April 2025 Accepted: 23 April 2025 Published: 9 May 2025

Citation: Silva, C.M.F.; Lino, R.C.; de Moura, M.C.T.; de Sá Borges, A.P.; de Oliveira Júnior, R.J. Innovative Approaches in the Synthesis and Optimization of Copper Complexes for Antitumor Therapies: A Comprehensive Review. *Molecules* 2025, 30, 2104. https://doi.org/10.3390/molecules30102104

Copyright: © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

#### 1. Introduction

According to the World Health Organization (WHO), cancer is the second leading cause of death worldwide, second only to cardiovascular diseases [1]. In 2022, there were approximately 20 million new cases and 9.7 million deaths from cancers, including non-melanoma skin cancers (NMSCs). Estimates suggest that approximately 1 in 5 men or women develop cancer in their lifetime, while approximately 1 in 9 men and 1 in 12 women die from it [2].

High mortality is related to factors such as late diagnosis and factors related to the drugs used, such as reduced cytotoxic selectivity, high toxicity, and acquired tumor chemoresistance, which leads to low adherence and high evasion of treatment [3,4]. The primary antitumor treatment options include surgery, radiotherapy, hormone therapy, immunotherapy, and chemotherapy. On the other hand, emerging therapies have been showing good

results, such as hormonal therapy, molecular targeting, genetics, cellular, biological, photo-dynamic, proton and nanomedicine.

Much research has been conducted using complexes based on metal ions in chemotherapy because the vital activities of cells and enzymes are organized by inherently existing metals [5]. Among these ions, Cu has been gaining attention in cancer research because it is an essential trace element for mammals and a component of enzymes and proteins, where it plays an important role in a range of biological processes. In addition, Cu plays an essential role in tumor angiogenesis, which is fundamental to tumor growth and metastasis, as demonstrated by the high concentration of Cu in tumor tissues [6].

Cu is an element sensitive to oxygen concentration and forms a rich variety of complexes with oxidation states +1 and +2, with a varied number of coordination and geometries [7,8]. In addition, it also has the properties of ligands, which allows the design of new metal complexes coordinated by Cu in the search for better antitumor drugs. The appropriate choice of ligands is the fundamental step in planning new syntheses. Tetrahedral geometry dominates in Cu(I) complexes, while square planar, trigonal bipyramidal, and octahedral arrangements are more frequently observed in the crystal structure of Cu(II) complexes [9].

Cu complexes have emerged over the years as an attractive chemotype for the treatment of human cancer. Several studies have shown that these complexes exhibit greater cytotoxicity, selectivity, and an ability to interfere with cell growth differently in normal and tumor cells, and they are even safer when compared to platinum-based complexes [10–19].

Therefore, the study of different types of cancer and their respective experimental methodologies has been of great relevance to the advancement of clinical science [12]. This study aimed to identify the main strategies used to develop and improve Cu complexes with antitumor activity.

# 2. Methodology

A comprehensive literature search was performed using PubMed and ScienceDirect. The keywords were applied in several different combinations such as "copper compounds" AND "antineoplastic agents" AND "pharmacological activity" NOT "clinical trial" NOT "review" in PubMed. A search was also performed using the descriptors "copper compounds" AND "cancer" in the ScienceDirect database.

The inclusion criteria for this review focused on past studies using Cu complexes as the metal center and involving biological analyses of tumor cells in vitro and/or in vivo, published between October 2019 and September 2024. Studies that tested compounds for diseases other than neoplasms, clinical studies, and literature reviews were excluded.

A total of 456 articles were identified, including 262 from the PubMed database and 194 from ScienceDirect. After reading the titles and abstracts, 280 were excluded, and 177 were selected (116 from PubMed and 61 from Science Direct). An important fact that was observed is that six articles were present in both databases, and 171 articles were counted for the subsequent analysis. The following data were collected: chemical formula of the Cu complex, type of cancer, cell line, strategies for improving the efficacy of Cu complexes (ligands, co-ligands, and formulations), type of tests (in vivo, in vitro and in silico), mechanistic action of the compounds, and efficacy and safety of the compounds in relation to cisplatin. Furthermore, all figures were created using ChemDraw software (version 12.0).

Molecules **2025**, 30, 2104 3 of 25

# 3. Types of Cancer and Main Cell Lines Studied to Verify the Antitumor Action of Cu Complexes

This review analyzed 23 different types of cancers, reflecting the diversity of neoplasms affecting the global population. Breast, colon, and lung cancers were the most studied types (Table 1). These findings are consistent with the high prevalence and mortality rates associated with these diseases. The widespread attention given to these neoplasms can be explained by their high incidence, which ranges from 34 to 212 times higher in countries in transition [1] as well as the ongoing search for more effective treatments and early diagnosis methods.

**Table 1.** Cancer types and cell lines studied. Other types include urinary bladder cancer (RT-4), human tongue squamous cell carcinoma (TCA8113), thyroid carcinoma (BCPAP), extrahepatic bile duct carcinoma (TFK-1), esophageal carcinoma (Eca-109), and murine mastocytoma (P815).

Type of Cancer	Cell Lines	Total
Breast cancer	MDA-MB-231 [11,14,20–55], MMT06056 [56], MCF-7 [3,6,11,14,17,19–22,25,26,29,33,35,40,43,45,50,51,54,55,57–98], T47D [29], JC [56], ZR-75-1 [33], SUM-159 [99], BT-549 [49], HS-578 [49], and 4T1 [30,31,45].	114
Colon cancer	HCT 116 [12,13,52,63,65,77,89,100–105], CT-26 [51,61,98,106], LoVo [10,88], Colo-205 [78,80,100,104,107–110], LS-180 [111], Colo-320 [70,100,107–109], HT29 [39,61,68,112,113], SW620 [9,15], SW1116 [97], LS174T [52,75,112,114,115], SW480 [9,15,77,116], CaCo2 [17,112], HuTu80 [117], HCT-15 [10,11,59,67,69,86,91,118,119], T-24 [42], HCT-8 [64], and DLD-1 [30,33,40].	68
Lung cancer	A549 [6,17,19–21,23–25,40,52,54,60,62–64,71,73,78,80,83,87,88,90,92,97,100,114–116,120–135], Sk-lu-1 [59], Hop-62 [136], A-427 [137], LCLC-103 [137], H157 [119], NCI-H460 [68,110,138,139], NCIH1975 [77], NCI-H23 [139], and M109 [56].	57
Uterine cancer	HELLA [4,6,16,17,19,20,32,35,39,47,60,62,69,71,76,78,87,88,114,115,117,124,132,140–146], MES-SA [147], SISO [137], and ECC-1 [39].	33
Prostate cancer	PC-3 [9,10,15,17,19,20,32,39,42–44,47,53,59,68,69,71,80,86,92,148–151] DU-145 [39,151], LNCaP [148,150], Myc-CaP [148], and 22Rv1 [17].	28
Liver cancer	HepG2 [6,16,37,44,58,66,79,81,89,93,94,111,113,132,139,145,152–154], HL-7702 [124,136], SMMC7721 [155], Bel-7402 [60,72], Bel-7404 [77], H22 [156], and HCCLM3 [37,157].	30
Ovarian cancer	SK-OV-3/DDP [136,158], 2008 [10,11,56,67,118,119], A2780 [13,17,24,35,65,67,86,121,159], CH1/PA [100,116], OVCAR3 [68,149], and NCI-ADR/RES [68].	22
Leukemia	THP-1 [85,110], K562 [29,49,59,68,86,128,160–162], KG1a [161], HEL [49], Jurkat [29,85,135,163], L1210FR [56], HL60 [49], and Molt-4 [110,138].	21
Human melanoma	B16-F10 [83,113,147,160,164], A375 [10,19,33,46,65,71,92,111,165], G361 [165], MV3 [43], SK-MEL-28 [165], and UACC62 [55,68].	19
Glioblastoma glioma	A172 [32,47], LN229 [32,33,35,47,166], U-87 [32,47,53,167], U-251 [59,63,68] U373-MG [167], T98G [111], and C6 [68,168].	19
Pancreatic cancer	PANC-1 [17,28,63,73,169], PaCa-2 [170], DAN-G [137], Capan-2 [132], BxPc-3 [10,92,107,169], ASPC-1 [169], and PSN-1 [10,11,118].	16
Neuroblastoma	SH-SY5Y [139,167] and IMR-32 [78,79,91,145].	6
Osteosarcoma	HOS [17,121], MG-63 [21,23,171], and 143B [170].	6
Squamous cell carcinoma	A431 [11,67,118,119,172].	5
Gastric cancer	MGC803 [77,124,132] and BGC823 [155].	4

Molecules **2025**, 30, 2104 4 of 25

Table 1. Cont.

Type of Cancer	Cell Lines	Total
Renal cancer	RD0995 [56], TK10 [55], RENCA [56], and 786-0 [68].	4
Lymphoma	Ragi [135], DL [18], and U937 [29,110].	4
Others	RT-4 [137], TCA8113 [152], BCPAP [10], TFK-1 [155], Eca-109 [60], and P815 [56].	6
	Total strains studied	110

Breast cancer, for example, is the most common type of cancer among women world-wide and is one of the most studied [173]. Its molecular heterogeneity and different responses to treatment justify the large number of studies. Similarly, colon and lung cancers are frequently studied owing to their high mortality and the complexity of treatment [174], which calls for continuous research into new therapeutic compounds and prevention approaches. As shown in Table 1, the distribution of the types of cancer reinforces this tendency to concentrate research efforts on a few specific types of cancer.

Other types of cancer are also relevant, but there are few studies that have conducted tests with such cell types. This indicates a possible gap in the analyses conducted, as they may have specific biological characteristics and require greater scientific attention. To advance scientific knowledge [175] and search for more effective Cu metallocomplex therapies for the treatment of neoplasms, it is important to highlight the need to expand studies to other types of cancer, which are equally devastating.

In addition, in the 171 articles analyzed in this review, 108 cell lines were studied and used to investigate the effect of Cu compounds. In several studies, more than one cell line was used (Table 1).

The most frequently used strains are MCF-7 and MDA-MB-231, both derived from breast tumors. The predominance of these strains can be explained by the high prevalence of breast cancer, as well as the heterogeneity of the molecular subtypes of this cancer [176]. This makes these cells relevant models for investigating the therapeutic potential of new compounds. MCF-7 cells are often used to study the estrogen-dependent forms of breast cancer [177]. In contrast, MDA-MB-231 cells are a model of triple-negative breast cancer that tends to be more aggressive and difficult to treat [178].

The A549 strain, derived from lung cancer [179], also appears frequently, in line with the high incidence of lung cancer worldwide, with approximately 2 million new cases and 1.76 million deaths per year [180]. The HeLa strain, which comes from cervical cancer, is another widely used model because of its robustness and replication capacity, and it is a reference in cancer studies [181]. In addition, this was the first established immortal human cell line [182].

The PC-3 strain, derived from prostate cancer [183] is often used because of the increasing prevalence of this neoplasm, especially in older men [184]. The investigation of Cu compounds in this lineage is particularly important, given that prostate cancer is proving resistant to conventional treatments [185] and new therapeutic alternatives are necessary.

The HepG2 strain, originating from hepatocellular carcinoma [186], also features prominently, as liver cancer is one of the leading causes of cancer-related deaths worldwide [187]. HepG2 cells are a relevant model for evaluating the impact of therapeutic compounds on liver tumors [178]. This will enable a better understanding of the mechanisms involved in liver cancer and the effects of Cu compounds on these cells.

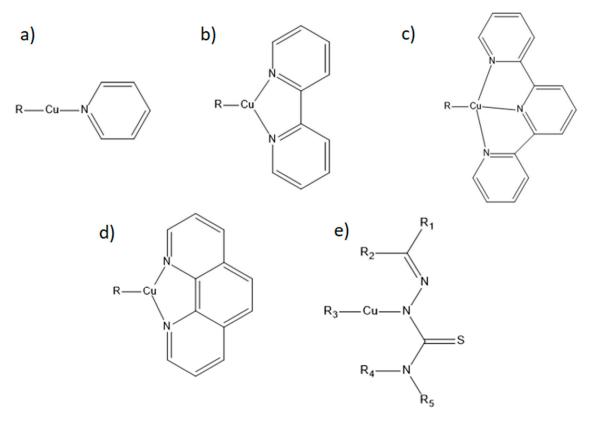
These cell lines allow for a controlled investigation of the effects of Cu compounds, with comparable results between different studies. Therefore, the results of this review

Molecules **2025**, 30, 2104 5 of 25

offer valuable insights into the predominant use of cell lines as study models for exploring the therapeutic potential of Cu compounds in different biological contexts.

# 4. Strategies Used to Improve the Antitumor Efficacy of Cu Complexes (Ligand, Linker and Pharmacophore)

In this review, the commonly used ligands in Cu complexes were phenanthroline (PHEN), followed by pyridines (PYR), bipyridines (BYP), terpyridines (TERP), and thiosemicarbazones (TSC) (Figure 1).



**Figure 1.** Chemical structure of the main copper-coupled ligands. (a)—pyridine (PYR), (b)—bipyridine (BYP), (c)—terpyridine (TERP), (d)—phenanthroline (PHEN), and (e)—thiosemicarbazone (TSC). Source: Own authorship.

The PHEN (Figure 1d) ligand and its substituted derivatives, in both metal-free and coordinated states, disrupt the functioning of a wide variety of biological systems. Cu complexes are more active in the presence of heterocyclic ligands because they exhibit potent antitumor activity against various cancer cell lines through the generation of reactive oxygen species (ROS), which results in the induction of apoptosis [112].

Various studies have been conducted to improve the effectiveness of Cu-PHENs by adding binding agents. Mixed Cu compounds have attracted great interest owing to their ability to interact with DNA and their consequent nuclease actions [140]. In the present review, it was possible to observe the use of mixed Cu-PHEN as a pharmacological strategy to improve antitumor activity [20,21,57,58,120,121,136,163,167,171,188].

It is worth noting that the binding agents generally comprise molecules with biological characteristics (antioxidant, anti-inflammatory, and antitumor activities) [57]. The main mechanisms of the antitumor action of Cu-PHEN as a single ligand or mixed ligand identified in this review are described in Table 2.

Molecules **2025**, 30, 2104 6 of 25

**Table 2.** Main ligands and co-ligands of the Cu complex and the pharmacological action.

#### Single Ligand Pharmacological Action

PHEN: Delayed apoptosis and release of interleukin 6 (IL-6) [23]; inhibition of aquaglyceroporins, which reduced glycerol permeation and impaired cell migration [106]; induction of ROS and apoptosis [112]; effective binding with antiapoptotic proteins of the BCL family [76]; not investigated [147].

BYP: Cell cycle arrest in the G0/G1 phase and inhibited signaling pathways regulated by Cathepsin D [148].

TERP: ROS production, inhibition in the G0/G1 phase and subsequent apoptosis [63]; ROS production and apoptosis [61]; strong affinity of the compounds to bind to DNA as intercalators and induce DNA conformational transitions [60]; not demonstrated [64].

TSC: inhibition of protein disulfide isomerase [10,11]; inhibition of EGFR protein [66]; activation of caspases 3 and 7 leading to apoptosis [100]; G2/M phase cell cycle arrest and DNA damage [165]; inhibition of OCT1-3 [107]; accumulation of cells in the sub-G1 fraction, as well as reversible arrest in the G0/G1 and G2/M phases in K562 and KG1a cells [161], externalization of phosphatidylserine and activation of caspase-3 leading to apoptosis, increased formation of ROS in K562 and KG1a cells [161]; reduction in catalase activity [25]; inhibition of glutathione synthesis [25,152]; inhibition of topoisomerase [152]; not demonstrated [3,19,65,129].

#### Pharmacological Action of Complexes with Ligands

PHEN + Schiff base: ROS [167]; DNA and FGRF receptor binding; increased planarity, chelation [58]. PHEN + diplacone: G2/M cell cycle arrest; ROS [121]. PHEN + biotin: DIP inhibition together with nuclear DNA and apoptosis [52]. PHEN + phenylmethyltriazolol: Methanuclease activity and DNA degradation [164]. PHEN + hydrazide: Reduced expression of the Ki-67 substance and the Cyclin D1 protein [156]. PHEN + hydroxyphenylimino: Apoptosis induced by intrinsic and extrinsic pathways [57]. PHEN + 8-hydroxyquinolone: Apoptosis via mitophagy and ATP depletion [136]. PHEN + hydrazone: Apoptosis and DNA cleavage [21,171]. PHEN + quercetin: Apoptosis; caspase 3/7 activity; mitochondrial depolarization [185]; PHEN + hydrocyananthracene: inhibition of topoisomerase I, ROS, DNA cleavage [188]; PHEN + phenolate; PHEN + naphthenolate:  $\pi$ -stacking interaction; DNA cleavage [140]. PHEN + TERP: Not demonstrated [120]. BYP + 8-hydroxyquinoline: Apoptosis via mitophagy and ATP depletion [136]. BYP + Schiff base: ROS [167] BYP + phenylmethyltriazolol: Elimination of superoxide; methanuclease activity [164]. BYP + quercetin: Apoptosis; caspase 3/7 activity; mitochondrial depolarization [185]. BYP + hydrazone: apoptosis and DNA cleavage [21,171]. BYP + hydrocyananthracene: Inhibition of topoisomerase I, ROS, DNA cleavage [188]. BYP + phenolate; BYP + naphthenolate:  $\pi$ -stacking interaction; DNA cleavage [140]. TERP + phosphine: Not demonstrated [62]; TERP + BYP: Not demonstrated [120]. TSC + phosphane: Apoptosis [149].

PHEN: phenanthroline; BYP: bipyridine; TERP: terpyridine; PYR: pyridine; ROS: reactive oxygen species; DNA: deoxyribonucleic acid; TSC: thiosemicarbazones; EGFR: epidermal growth factor inhibitor; OCT: organic cation transporter.

Another common ligand in Cu complexation is the heterocyclic compounds BYP and TERP (Figure 1b,c). These two compounds have characteristic optical and electrochemical properties, such as the ability to form complexes with metals. The BYP ligand has been tested as a simple ligand [148] and in combination with other ligands [21,136,140,164,167,171].

Several studies on the Cu-BYP complex have also evaluated the Cu-PHEN complex [20,21,59,140,164,171,188]. In general, Cu-BYP mixed ligands show excellent cytotoxic activity, with IC $_{50}$  values close to 10  $\mu$ M. However, when compared to Cu-PHEN mixed ligands, the latter showed greater cytotoxic activity [20,59,140,167,171,188].

Only one study has shown greater specificity of the Cu-BYP/phenylmethyltriazole mixed heterocycle ligands in the B16F10 cell line compared to that of the Cu-PHEN/phenylmethyltriazole [164]. Another study showed that the Cu-BYP/Schiff base mixed ligands were more potent in the gliobastoma cell lines U87 MG and U373 MG, while the Cu-PHEN/Schiff base mixed ligands were more potent in the neuroblastoma cell line Sh-SY5Y [167].

Regarding the TERP ligand (Figure 1c), of the seven studies identified in this review, Cu-TERP complexes have been shown to be highly cytotoxic [60–62], to demonstrate good selectivity [62,63], and to have better anticancer effects than cisplatin [60,62,64]. Notably, only one study [120] added a linker (PHEN, BYP, or methylimidazole), all of which were

Molecules **2025**, 30, 2104 7 of 25

found to be more cytotoxic and selective than cisplatin. These data revealed that Cu-TERP complexes are promising antitumor agents.

Another prevalent ligand in this review is thiosemicarbazides, which comprise a group of sulfur derivatives of semicarbazones obtained by the condensation of appropriate aldehydes or ketones and thiosemicarbazides in an acidic environment. The structure of thiosemicarbazides has a significant influence on their biological activity [165] and has therefore been used as ligands in metal complexes.

Recently, TSC (Figure 1e) has been studied for their promising anticarcinogenic properties. These ligands and their transition metal complexes are biologically active compounds and anticancer agents with versatile structural properties [65]. Metal complexes with this ligand may have different cellular targets than cisplatin, raising the prospect of increasing the spectrum of action and obtaining better selectivity [10].

Fourteen articles used TSCas as ligand. In four of these articles, the results showed greater cytotoxic effects than those of cisplatin [10,66,107,149]. Two articles showed a good selectivity index [11,22], and two that carried out in vivo studies observed improved safety [10,11].

Different studies report varying mechanisms of action for Cu-TSC complexes (Table 2). Four studies used salicylaldehyde group [10,11,100,107]. In these studies, three distinct mechanistic actions were identified: inhibition of disulfide isomerase [10,11], activation of caspases 3 and 7 [100], and inhibition of organic cation transporters [107].

As shown in Table 2, the pharmacological actions of the Cu complexes differed between studies. This difference may be related to the different actions of the ligands complexed with Cu and the experimental design of the studies, which differ in terms of the methods used to investigate the pharmacological actions of the compounds.

Some past studies have presented ligand-improvement techniques to improve the binding of Cu metal complexes to DNA and increase their cytotoxicity in cancer cells. Romo et al. [23] used the ligand 1,3-bis(1,10-phenanthrolin-2-yloxy)-*N*-(4-(methylthio)benzylidene)propan-2-amine. According to the authors, the distorted trigonal-bipyramidal geometry of the Cu complex with this ligand allowed PHEN fragments to be easily accommodated in the DNA double helix. In addition, the aromaticity of these fragments improves their local hydrophobicity, thereby increasing their affinity for the hydrophobic domains of DNA. The Cu complex showed superior cytotoxicity in the cell lines studied when compared with cisplatin.

The tetramethylphenanthroline ligand is a virtually flat molecule that intercalates DNA and is highly cytotoxic. Alvarez et al. [24] evaluated the cytotoxicity of Cu complexes in various cancer cell lines. They found that Cu complexes with the L-dipeptide ligand tetramethylphenanthroline were highly cytotoxic compared to Cu complexes with the L-dipeptide ligands PHEN and cisplatin.

Another ligand, bathophenanthroline (BPHEN), has shown promising results. Du et al. [136] and Trávníček et al. [121] compared this ligand to the PHEN ligand and showed inferior activity. By contrast, Cu-L-dipeptide complexes and BPHEN showed superior activity when compared with that of Cu-L-dipeptide complexes, PHEN, and cisplatin [25]. Therefore, these ligands are potential candidates for studying in vivo activity in the treatment of aggressive tumors for which there is no curative pharmacological treatment. In addition, the Cu-naringenin/BPHEN ternary complex showed greater selectivity against cancer cells than the Cu-naringenin complex. Cell death was found to be related to the generation of ROS, loss of mitochondrial membrane potential, depletion of GSH, and GSH/GSSG ratio [122].

As can be seen, in addition to modifications to the structure of the ligand, peptide additions have been used to improve the activity of the compounds and are intended to

Molecules **2025**, 30, 2104 8 of 25

cover a range of different side chains and increase lipophilicity. The results indicated that the use of L-dipeptides increased cytotoxic activity compared to other Cu complexes [24,25].

In the search for more effective anticancer drugs aimed at specific molecular targets, the use of a "hybrid" medicinal chemistry approach, which exploits the unique behavior of transition metal complexes conjugated to pharmacophores, can provide an advance in therapy. In the present review, the main pharmacological classes and their representatives found were selective estrogen receptor modulators (tamoxifen) [26], sex hormones and steroids (estradiol) [67], antibiotics (pefloxacin, ciprofloxacin, nalidixic acid, doxycycline, tetracycline) [12,68,101,160], anti-inflammatory drugs (indomethacin) [141], antiparasitics (albendazole) [69], and disulfiram [123].

Regarding the tamoxifen-derived ligand (TAML), Cu-TAML retained its estrogen receptor-binding activity. Its pharmacological action is related to redox imbalance, characterized by a reduction in the total thiol content and an increase in ROS production. In addition, it promotes mitochondrial swelling and the release of pro-apoptotic factors, probably due to extensive oxidative damage. The compound exhibited cytotoxic activity toward estrogen-sensitive and -insensitive breast cancer cells and overcame cancer resistance [26].

Barrett et al. [67] in their study complexed Cu-PHEN with estradiol. These complexes showed strong in vitro intercalatory interactions with nuclear DNA, ROS production, and DNA cleavage. This series of compounds showed reduced and submicromolar  $IC_{50}$  values and cellular uptake followed the order of lipophilicity, indicating that internalization occurred mainly by passive diffusion.

Several studies have investigated Cu-complexed fluoroquinolones as antibiotic-derived ligands. The wide applications of Cu ions and fluoroquinolones, such as antimicrobial, anti-inflammatory, and antiviral activities, have stimulated research into mixed Cu-ciprofloxacin/isatin and Cu-pefloxacin/isatin complexes. The antitumoral actions of these compounds are as follows: antioxidant action, inhibition of clonogenic capacity, and activation of apoptotic pathways in HCT116 cells [12,101].

Another complex synthesized using antibiotics is hydrazone/nalidixic acid. Notably, nalidixic acid has shown promising antimicrobial and antitumor activity. The complexes showed prominent methanuclease activity with cytotoxic activity against U251, UACC-62, MCF-7, and HT-29 cells. These findings stimulated new studies on this mixed compound [68].

Another synthesized metallopharmaceutical is a Cu ternary complex with doxycycline and phenanthroline (Cu-Dox-PHEN), which is highly cytotoxic and has great potential for DNA cleavage. The mechanistic actions include the generation of free radicals, ROS generation, intercalation, and DNA cleavage. The compound exhibited moderate genotoxicity, selective inhibition of B16F10 murine melanoma tumor cell growth, in vivo chemotherapeutic potential against S180 and Ehrlich sarcoma tumors [160].

Ligands derived from anti-inflammatory drugs can improve the action of antitumor compounds by reducing inflammation at the tumor site. In a study by Godínez-Loyola et al. [141] the action of the Cu-indomethacin/diimines/Schiff base complex was associated with tumor reduction due to cyclooxygenase inhibition. The release of the compound exhibited a burst effect in acidic media, which is characteristic of the tumor site.

Owing to its antiparasitic effects and recent evidence of its antitumor activity, albendazole has been used in the synthesis of coordination compounds. Cu-albenzazole complexes show cytotoxic activity against HeLa, MCF-7, PC3, and HCT-15 cell lines. Although the mechanism of action has not yet been elucidated, these findings indicated cell death by apoptosis [69].

Another drug with potential antitumoral activity is disulfiram, whose activity is enhanced in the presence of Cu ions. The Cu-Disulfiram complex easily crossed the membrane

Molecules **2025**, 30, 2104 9 of 25

of A549 cells and accumulated intracellularly. This process triggered cell morphological changes, increased ROS, cell cycle arrest in the G0/G1 phase and apoptosis [77,123].

# 5. Mechanistic Actions of Cu Complexes

Of the 171 articles reviewed, 18 (10.5%) did not present synthesis studies in conjunction with biological studies, and 129 (75%) investigated the classic mechanisms of action of Cu complexes that are already well established in the scientific community, including the ability to cleave DNA by releasing reactive oxygen species, causing cell cycle arrest and consequently apoptosis.

Of these biological tests, 90 studies carried out cytotoxicity, DNA interaction, and cleavage tests, as well as flow cytometry marker tests to elucidate their mechanisms of action. Of these, 39 showed only studies on cytotoxic capacity, DNA interactions, and cleavage capacity. Of these, 4 presented data exclusively on DNA cleavage, while another 5 carried out only interaction tests with macromolecules and/or in silico simulations. Another 4 presented only cytotoxicity studies, and another 26 presented cytotoxicity, cleavage, or docking studies. Of the total number of selected articles, 38 did not present mechanistic studies and focused only on synthesis and proliferation tests. Another five studies conducted phototoxicity tests and in vivo experiments.

Of all the studies selected for this review, 72 (41.9%) studied other mechanisms of action and other potential targets for the antitumor activity of Cu complexes or investigated the expression of enzymes that corroborate the classic mechanism of programmed cell death.

Carcelli et al. [10] and Pellei et al. [118] proposed a new target, disulfide isomerase proteins (DPIs), which catalyzes the reduction in disulfide bonds and the oxidation of thiols. DPIs are abundant oxidoreductases that reside in the endoplasmic reticulum (ER) and play crucial roles in protein folding. However, because of the major interference between the ER and mitochondria in redox homeostasis, the inhibition of IPRs has been shown to strongly affect mitochondrial pathophysiological stability [11].

In addition, studies have evaluated the levels of reduced thiols in cells treated with metal complexes. These results supported the hypothesis that Cu(I) derivatives linked to pyrazoyl or Cu(II) linked to TSC can effectively target DPIs in colon cancer cells. This causes an imbalance in the cellular redox homeostasis, shifting it to a reduced state. In addition, morphological analysis revealed that both complexes induced a slight increase in mitochondrial size, reduced the electronic density of the inner membrane and matrix regions, and altered cristae characteristics [10,118].

Compounds that inhibit mitochondria-resident IPRs induce non-oxidative stress-mediated cancer cell death. In addition, IPRs have a binding affinity for Cu and they have been shown to play an essential role in regulating the intracellular redox state of Cu ions, which catalyzes the formation of disulfide bonds [189].

Another mechanistic action is the externalization of phosphatidylserine and the activation of endogenous caspases, both via extrinsic and intrinsic pathways, resulting in apoptosis [189]. Parsa et al. [161], Reheman et al. [142], and Vitomirov et al. [71] demonstrated that activation of caspase-3, caspase-8 and caspase-9 induced apoptosis in HeLa cells with the Cu-PHEN compound, in HeLa cells with the Cu-pyrazolone compound and in leukemia cells with the Cu-TSC compound, respectively. Two of these past studies also showed an increase in BAX protein expressions in relation to BCL-2 protein transcripts, which are pro-apoptotic and antiapoptotic regulators [142,161]. Changes in the expression of these proteins may be associated with increased cancer risk and treatment resistance.

Additionally, Reheman et al. [142] demonstrated cleavage of poly (ADP-ribose) polymerase (PARP) protein. PARP is a polymerase that participates in the DNA repair process,

and its inhibition is an efficient approach for treating various types of cancers. The success of this approach has led to the approval of four different PARP inhibitors for the treatment of various types of cancer; seven different compounds are currently under clinical investigation for various indications [190].

Furthermore, Reheman et al. [142] observed three pathways, namely: PI3K/AKT, P38/MAPK, and JNK/MAPK. The study proves that the copper complex induced apoptosis by blocking the cell cycle in the S phase. Furthermore, the compound in question promoted the activation of caspase-3 and caspase-9 in HeLa cells. It inhibited the PI3K/AKT pathway and activated the P38/MAPK and JNK/MAPK pathways. It also inhibited the phosphorylation of Ik-B $\alpha$  in the NF-kB pathway activated by TNF- $\alpha$ , thus restricting the routine of HeLa cells.

The MAPK signaling pathway is involved in the regulation of biological mechanisms such as cell proliferation and apoptosis. In the PI3K/AKT signaling pathway, the balance between cell proliferation and apoptosis can be modulated by regulating AKT expression to inhibit tumor cell growth. The P38/MAPK pathway has been associated with the induction of apoptosis through various cellular stress signals, such as TNF- $\alpha$ , interleukin-1, ultraviolet radiation, hyperosmotic stress and chemotherapy, while activation of the JNK pathway can increase caspase-3 activity, which culminates in apoptosis [191,192].

Zhang et al. [124] studied Cu(II) complexes with 5-pyridin-2-yl-[1,3]dioxolo[4,5-g]isoquinoline derivatives and monitored the expression of caspase-3 and caspase-9 in seven compounds by flow cytometry, two of which showed an increase in the expression of caspases-3 and caspase-9, causing apoptosis via the intrinsic pathway.

Mutlu Gençkal et al. [185], when investigating copper compounds and other transition metals, linked to quercetin and diimines, carried out tests to evaluate the behavior of caspases-3 and caspase-7. Strong activation of this pathway has been observed in human oral squamous cell carcinoma cells (KBv200), demonstrating the induction of apoptosis via caspases-3 and caspase-7 in the Cu-PHEN/quercetin complex. This same complex showed an increase in the level of the M30 antigen, which was detected in an ELISA test for cytokeratin-18 breakdown, which refers to the action of caspases at a specific aspartate 396 (Asp396) cleavage site. This is an important indicator of apoptosis [193].

Another mechanism that has been studied is the inhibition of proteasomal activity. Modified cells that become tumors positively modulate the ubiquitin-proteasome system (UPS) and use it to degrade tumor suppressor proteins and prevent apoptosis [27]. Balsa et al. [82] showed that the Cu-acylhydrazone complex inhibited proteasomal activity by 30% in MCF7 cells and 47% in MDA-MB-231 cells, which helped prevent the antiapoptotic mechanism developed by tumor cells. In addition, the complex was shown to bind to the 20S proteasome activation site via molecular docking.

With regards to miRNA, they comprise a class of small non-coding RNA molecules that can silence up to 60% of coding genes. They can act as tumor suppressors or oncogenes (oncomiR). Studies have shown that oncomiR suppression therapy or increased expression of tumor suppressor genes is effective in treating cancer. Overexpression of miR-21 and miR-155 is associated with apoptosis, proliferation, and invasive potential via silencing of programmed cell death 4 (PDCD4) and liver kinase B1 (LKB1), respectively [194].

miR-206 and miR-133b are members of the miR-bicistronic cluster. Higher levels of miR-206 increase apoptosis and prevent cancer formation, while miR-133b is an oncomiR that activates the ERK and AKT1 pathways, which are important for oncogenesis [114]. Measurement of miRNA levels via RT-PCR represents an additional level of investigation into the antitumorigenic effects of the selected compounds. Petronijević et al. [195] analyzed the RNA of cells treated with three Cu-quinoxalinone complexes. Among them, miR-21, which has high oncogenic potential, showed no changes. By contrast, two compounds

increased the levels of miR-206, and all three increased the levels of miR-155. These data show that the compounds exert better anticancer activity by stimulating suppressor genes and inhibiting tumor-promoting genes.

Tubulin is one of the main targets in cancer treatment because it plays essential roles in cell division and intracellular transport. Inhibiting the formation of microtubules induces cell death by apoptosis, making them a new target for chemotherapy studies aimed at inhibiting the polymerization or depolymerization of microtubules [70].

Tubulin disassembly experiments showed that the two Cu-benzazozine complexes are effective microtubule-destabilizing agents that bind to the colchicine site, which was also confirmed by molecular modeling tests [28]. The binding of a metal complex to the tubulin-colchicine pocket is unprecedented. Hossan et al. [196] treated cells for 24 h with a semi-synthetic Cu-cardamonin compound, stained them with DRAQ5 and anti- $\alpha$ -tubulin monoclonal antibody. and investigated them morphologically using confocal microscopy. The images revealed the induction of a microtubule-disrupting agent (MDA) in cancer cells, such as multinucleation, nuclear fragmentation, and disruption of the microtubule network.

Topoisomerases are proteins that are essential for cellular processes such as replication, transcription, DNA duplication, chromatin assembly, and chromosome segregation, and are capables of modifying the topological properties of DNA. For example, they regulate the level of supercoiling in double helices. These inhibitors act mainly through interactions with topoisomerase I and tumor DNA, thus inhibiting the replication of tumor cells [108].

Wittmann et al. [109] studied complexes of Cu-latonduines and Cu-quinolines, which proved to be DNA-intercalating agents and subsequently caused poisoning of topoisomerases I and II, which is considered a mechanism of action of quinolines. By contrast, Pósa et al. [152] demonstrated the inhibition of topoisomerase by Cu-TSC complexes using electrophoresis. This process leads to cell cycle blockage in many tumors cell lines and consequently generates numerous DNA breaks during non-homologous recombination. The result of this is lethal chromosomal damage, such as breaks, which subsequently lead to nonhomologous recombination, and the accumulation of errors in the genome causes cell apoptosis.

# 6. Nanoformulations as a Strategy to Improve the Effectiveness and Safety of Cu Complexes Used as Antitumor Agents

The use of nanoformulations is a therapeutic strategy that uses nanoparticles to transport drugs or other active ingredients, thereby improving quality, safety, and efficacy. In the present review, six studies used the nanoencapsulation of Cu compounds as an improved strategy to reach the tumor environment more effectively and safely.

Encapsulation involves the modification of different physicochemical and biochemical characteristics, including solubility, stability, and rapid release. This method prevents drug degradation, increases therapeutic efficacy, and also reduces side effects. Nanoparticles can increase the permeability and retention effect, so they are able to increase the concentration of the drug inside the tumor, controlling drug release and specific targeting [141,197]. These characteristics give nanocomplexes an important role in cancer therapy and make them promising candidates for replacing conventional chemotherapy.

This was observed in a study using Casiopein<sup>®</sup>, CasIII-ia nanoencapsulated in niosomes designed and optimized with Quality by Design (QbD) tools for intravenous (IV) administration. In vivo analyses showed good efficiency and lower toxicity compared with the compound and free cisplatin. The MDA-MB-231 strain showed greater CasIII-ia activity in the period of up to 48 h absorbed by a diffusion process, demonstrating better distribution of the drug and low toxicity [37].

In a combination of Cu and cetyltrimethylammonium bromide (Cu-CTAB) in gallium oxide nanoparticles (Cu-CTAB + GaONPs), the formulation showed an IC $_{50}$  of 0.2 µg/mL in hepatocellular carcinoma cells (HepG-2). This showed greater antiproliferative power when compared to Cu-CTAB and GaO-NPs administered alone. In Wistar rats, the recovery of damaged tissue was observed histologically, and biochemical tests showed a reduction in liver function markers alanine aminotransferase (ALT), aspartate aminotransferase (AST) and a decrease in the transcription of tumor markers such as alpha-fetoprotein (AFP), transforming growth factor beta (TGF- $\beta$ 1),  $\alpha$ -L-fucosidase. Meanwhile antioxidant markers (SOD), apoptosis markers (caspase-3 mRNA) and arginase showed high values in real-time polymerase chain reaction (RT-qPCR) [158]. Gallium oxide nanoparticles can be used as multifunctional drug carriers because they easily penetrate cell membranes and, owing to their luminescent properties, facilitate their distribution within cells [198].

Pinho et al. [106] used a pH-sensitive liposome with Cu-PHEN.CL<sub>2</sub>. Its use in the treatment of rectal colon cancer cells maintained the cytotoxic properties of unformulated Cu-PHENs. This treatment caused a reduction in glycerol permeability and impaired cell migration, probably owing to the inhibition of aquaglyceroporins. In a syngeneic murine colon cancer model, the nanocomposite significantly reduced tumor progression compared to the control group and those treated with unformulated Cu-PHENs. No toxic side effects were observed, leading to a 50% reduction in the rectal colon cancer tumor volume. This study highlighted the maximization of biological activity using a lipid-based nanosystem [106,153].

To overcome this deficiency in the solubility of a Cu diethyldithiocarbamate (Cu-ET) complexes, the complexes were prepared with nanoparticles that were dispersed in bovine serum albumin (BSA) (Cu-ET-NPs). Such therapy generates free hydroxyl radicals and increases ROS, mediated by glutathione, which is crucial for the inhibitory role of human hepatocellular carcinoma cells (HepG2). Consequently, they induce polyubiquitination of proteins from cancer cells inoculated into mice [199].

The third generation Casiopein<sup>®</sup>, [Cu(N-N)(Indo)]NO<sub>3</sub>, was synthesized and nanoencapsulated, where Indo is deprotonated indomethacin and N-N is BYP or PHEN. The complexes showed high cytotoxicity compared to cisplatin and demonstrated anticancer performance with the characteristic and multifunctional mechanisms of action of their peers. The formulation used chitosan hydrogels, which are responsive to changes in the environment such as pH and temperature. These characteristics make them suitable as anticancer drugs, as tumor microenvironments are characterized by an acidic pH and increased temperature. The release of the compound showed a burst effect and was faster under acidic conditions in the first six hours, with a cellular absorption mechanism by passive diffusion; however, the study did not carry out in vivo tests after encapsulation [141].

The synthesis of the Cu-hydrazone nanocomplex resulted in high inhibition of HepG-2. The antitumor activity of the Cu-hydrazone nanocomplex is discussed in relation to its chemical structure, Cu (II)-reducing capacity, and potential inhibitory effect on the cyclin-dependent kinase 2 (CDK2) enzyme, as visualized by molecular docking tests [197]. A similar therapy was presented in a previous study that identified cell death via apoptosis [200].

## 7. Effectiveness and Safety of Cu Complexes Compared to Cisplatin

Cisplatin is one of the best-known and most widely used metal-based chemotherapeutics for cancer [201,202]. However, because of their high toxicity, poor tolerance, and high cost, other transition metal complexes, including Cu complexes, have been synthesized and tested as chemotherapeutic drugs [29].

In this review, 68 studies found one or more Cu compounds with a high cytotoxicity for some cell lines and with  $IC_{50}$  values lower than that for cisplatin. In 34 ar-

ticles [6,9,13–19,22,25,65,72–76,82,99,101–103,110,122,124,136,137,142,150,159,160,171] one or more compounds showed good selectivity for tumor cells and in twelve [10,11,31,37,77,106,138,156,157,160,169,171], the compounds showed more favorable safety data than cisplatin, making these promising antitumor agents. However, some cases have shown that cisplatin proved to be more effective than the tested compound [19,58,137,145].

Some in vivo studies have demonstrated the greater safety of Cu complexes, Casiopein<sup>®</sup> [37,169], TSC [10,11], dithiocarbazate [138], hydrazide [30,171], and triphenylphosphine [77], than cisplatin. In these studies, mice that were treated with the Cu complexes and afore mentioned ligands showed lower body weight loss than those treated with cisplatin, which can be inferred from the lower incidence of anorexia.

In addition, other safety data related to Cu complexes have been reported, such as the non-induction of mutagenicity, recombinogenicity, and carcinogenicity in *D. melanogaster* with the Cu-PHEN/hydrazide compound [156]; an improvement in the survival rate of mice with Ehrlich ascites tumors with the Cu-Dox/PHEN compound [160]; and an absence of toxic side effects with the Cu-PHEN compound [106].

Although copper compounds may exhibit lower systemic toxicity and greater selectivity compared to cisplatin, their efficacy can vary significantly depending on the tested compound and the type of cancer [62,189,203]. Such compounds can induce cell death through mechanisms distinct from cisplatin's DNA-binding meanism. This difference may affect their efficacy and selectivity in certain types of cancer. However, it does not diminish the therapeutic potential of copper compounds, opening the possibility for further studies that will strengthen their use as a potential chemotherapeutic agent [204,205].

However, some studies have shown unsatisfactory safety data, such as signs of behavioral toxicity and mortality with Cu-thioxoimidazolone complexes [16], systemic toxicity with significant histopathological changes in the liver of mice with Cu-pyridine with halogen substitution [14], and a significant decrease in monocyte count with Cu-acylhydrazone [171].

## 8. Conclusions

In this review, we highlighted the diversity of neoplasms investigated in recent studies covering 23 types of cancer, with special attention paid to breast, colon, and lung cancers, which are the most studied. However, the limited amount of research dedicated to other types of cancer reveals a gap that demands greater scientific attention, given the potential for specific biological characteristics in these cases. Thus, among the 171 articles analyzed, there was a predominance of studies with cell lines, such as MCF-7, MDA-MB-231, A549, PC-3, HepG2, and HeLa, which served as the main models for evaluating the effects of Cu compounds and their impact in different tumor contexts.

In addition, this review has highlighted the versatility of Cu complexes, especially those with ligands such as phenanthroline (PHEN), pyridines (BYP and TERP) and thiosemicarbazones (TSC), for potential antitumor applications. The Cu-PHEN, Cu-BYP, and Cu-TERP complexes showed high cytotoxic activity, with emphasis on the mixed complexes and the use of linkers, which increased the ability to interact with DNA and the action of nucleases, thereby increasing the effectiveness and selectivity compared to cisplatin. In particular, the Cu-TSC complexes have various mechanisms of action, including the inhibition of specific enzymes and activation of apoptotic pathways. These data suggest that Cu complexes with different ligands have promising potential as therapeutic alternatives against cancer, especially for tumors that are resistant to conventional treatments.

The impact of structural modifications and the use of ligands to improve the cytotoxic activity and selectivity of compounds toward DNA and cancer cells has also been explored. Recent studies have shown that the use of ligands such as PHEN, tetramethylphenan-

throline, bathophenanthroline (BPHEN), and dipeptides increases the affinity of Cu for DNA and thus improves cytotoxicity when compared to conventional treatments such as cisplatin. In addition, complexes derived from clinically applied drugs, such as tamoxifen, estradiol, fluoroquinolones, anti-inflammatories, albendazole and disulfiram, showed promising antitumor actions when combined with Cu ions, indicating a possible "hybrid" chemotherapy approach capable of tackling tumors more effectively.

A review of 171 articles revealed that most studies on Cu complexes focused on classic mechanisms of action, such as DNA cleavage, caspase activation and induction of apoptosis by reactive oxygen species (ROS). Some studies have explored new targets, such as protein disulfide isomerases (DPIs) and specific signaling pathways (PI3K/AKT, P38/MAPK, and JNK/MAPK), which are essential for redox homeostasis and apoptosis. In addition, proteasome inhibitors and miRNA modulators have emerged as potential antitumor agents, whereas the inhibition of tubulin and topoisomerases has shown efficacy in interrupting cell proliferation and inducing apoptosis. These studies showed that Cu complexes offer multiple routes of action against tumor cells.

Nanoformulation of Cu compounds are emerging as a promising approach for cancer treatment, using nanoparticles to increase effectiveness and safety by concentrating the drug in the tumor tissue and thus reducing adverse effects. Compounds such as Casiopein® and combinations of Cu with other agents have shown significant antiproliferative effects, especially in types of cancer such as hepatocellular carcinoma. In comparative studies, Cu compounds have been shown to be safer than cisplatin with lower toxicity in experimental models. However, these findings reinforce the potential of Cu complexes as therapeutic agents and justify the need for future research to validate and improve their clinical use.

**Author Contributions:** C.M.F.S., R.C.L., M.C.T.d.M. and A.P.d.S.B. were responsible for the review of the scientific literature and writing the original manuscript. A.P.d.S.B. and R.J.d.O.J. were responsible for the manuscript review. R.J.d.O.J. was also responsible for the supervision and funding acquisition of this project. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Fundação de Amparo à Pesquisa de Minas Gerais (FAPEMIG-APQ-01087-21 and APQ-00704-21), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), National Institute of Science, Technology in Theranostics and Nanobiotechnology (INCT—Teranano), Rede Mineira de Pesquisa Translacional em Imunobiológicos e Biofármacos no Câncer (REMITRIBIC), Universidade Federal de Ubelândia (UFU) and Universidade Estadual de Goiás (UEG).

Conflicts of Interest: The authors declare no conflicts of interest.

### References

- 1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 2021, 71, 209–249. [CrossRef]
- 2. Bray, F.; Laversanne, M.; Sung, H.; Ferlay, J.; Siegel, R.L.; Soerjomataram, I.; Jemal, A. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 2024, 74, 229–263. [CrossRef]
- 3. Adhikari, H.S.; Garai, A.; Yadav, P.N. Synthesis, characterization, and anticancer activity of chitosan functionalized isatin based thiosemicarbazones, and their copper(II) complexes. *Carbohydr. Res.* **2023**, *526*, 108796. [CrossRef] [PubMed]
- 4. Khamidullina, L.A.; Puzyrev, I.S.; Burygin, G.L.; Dorovatovskii, P.V.; Zubavichus, Y.V.; Mitrofanova, A.V.; Khrustalev, V.N.; Timofeeva, T.V.; Slepukhin, P.A.; Tobysheva, P.D.; et al. Unsymmetrical trifluoromethyl methoxyphenyl β-diketones: Effect of the position of methoxy group and coordination at Cu(II) on biological activity. *Molecules* **2021**, *26*, 6466. [CrossRef] [PubMed]
- 5. Ge, E.J.; Bush, A.I.; Casini, A.; Cobine, P.A.; Cross, J.R.; DeNicola, G.M.; Dou, Q.P.; Franz, K.J.; Gohil, V.M.; Gupta, S.; et al. Connecting copper and cancer: From transition metal signalling to metalloplasia. *Nat. Rev. Cancer* 2022, 22, 102–113. [CrossRef]
- 6. Lu, W.; Tang, J.; Gu, Z.; Sun, L.; Wei, H.; Wang, Y.; Yang, S.; Chi, X.; Xu, L. Crystal structure, in vitro cytotoxicity, DNA binding and DFT calculations of new copper (II) complexes with coumarin-amide ligand. *J. Inorg. Biochem.* 2023, 238, 112030. [CrossRef] [PubMed]

Molecules **2025**, 30, 2104 15 of 25

7. Stanojević, I.M.; Glišić, B.Đ.; Radanović, D.D.; Djuran, M.I. Copper(II) complexes of aminopolycarboxylate ligands with N<sub>2</sub>O<sub>2</sub>, N<sub>2</sub>O<sub>3</sub> and N<sub>2</sub>O<sub>4</sub> donor sets. The relationship between the ligand structure and molecular geometry of the complex. *J. Mol. Struct.* **2021**, 1232, 130001. [CrossRef]

- 8. Lelièvre, P.; Sancey, L.; Coll, J.-L.; Deniaud, A.; Busser, B. The Multifaceted Roles of Copper in Cancer: A Trace Metal Element with Dysregulated Metabolism, but Also a Target or a Bullet for Therapy. *Cancers* **2020**, *12*, 3594. [CrossRef]
- 9. Drzewiecka-Antonik, A.; Rejmak, P.; Klepka, M.; Wolska, A.; Chrzanowska, A.; Struga, M. Structure and anticancer activity of Cu(II) complexes with (bromophenyl)thiourea moiety attached to the polycyclic imide. *J. Inorg. Biochem.* **2020**, 212, 111234. [CrossRef]
- 10. Carcelli, M.; Tegoni, M.; Bartoli, J.; Marzano, C.; Pelosi, G.; Salvalaio, M.; Rogolino, D.; Gandin, V. In vitro and in vivo anticancer activity of tridentate thiosemicarbazone copper complexes: Unravelling an unexplored pharmacological target. *Eur. J. Med. Chem.* 2020, 194, 112266. [CrossRef]
- 11. Miglioli, F.; De Franco, M.; Bartoli, J.; Scaccaglia, M.; Pelosi, G.; Marzano, C.; Rogolino, D.; Gandin, V.; Carcelli, M. Anticancer activity of new water-soluble sulfonated thiosemicarbazone copper(II) complexes targeting disulfide isomerase. *Eur. J. Med. Chem.* 2024, 276, 116697. [CrossRef]
- 12. Bhatt, B.S.; Gandhi, D.H.; Vaidya, F.U.; Pathak, C.; Patel, T.N. Cell apoptosis induced by ciprofloxacin based Cu(II) complexes: Cytotoxicity, SOD mimic and antibacterial studies. *J. Biomol. Struct. Dyn.* **2021**, *39*, 4555–4562. [CrossRef] [PubMed]
- 13. Akhmetova, V.R.; Galimova, E.; Mescheryakova, E.S.; Dzhemileva, L.U.; Dzhemilev, U.M.; D'yakonov, V.A. Mono- and binuclear complexes of copper(II) with dimethylaminomethyl derivatives of 2-naphthol and 6-quinolinol: Synthesis and in vitro study of antitumor properties. *Metallomics* 2023, 15, mfad037. [CrossRef] [PubMed]
- 14. Amiri Rudbari, H.; Saadati, A.; Aryaeifar, M.; Blacque, O.; Cuevas-Vicario, J.V.; Cabral, R.; Raposo, L.R.; Fernandes, A.R. Platinum(II) and copper(II) complexes of asymmetric halogen-substituted [NN'O] ligands: Synthesis, characterization, structural investigations and antiproliferative activity. *Bioorg. Chem.* **2022**, *119*, 105556. [CrossRef] [PubMed]
- 15. Basaran, E.; Gamze Sogukomerogullari, H.; Cakmak, R.; Akkoc, S.; Taskin-Tok, T.; Köse, A. Novel chiral Schiff base palladium(II), nickel(II), copper(II) and iron(II) complexes: Synthesis, characterization, anticancer activity and molecular docking studies. *Bioorg Chem.* 2022, 129, 106176. [CrossRef]
- 16. Krasnovskaya, O.O.; Guk, D.A.; Naumov, A.E.; Nikitina, V.N.; Semkina, A.S.; Vlasova, K.Y.; Pokrovsky, V.; Ryabaya, O.O.; Karshieva, S.S.; Skvortsov, D.A.; et al. Novel copper-containing cytotoxic agents based on 2-Thioxoimidazolones. *J. Med. Chem.* **2020**, *63*, 13031–13063. [CrossRef]
- 17. Makowska, A.; Sączewski, F.; Bednarski, P.J.; Gdaniec, M.; Balewski, Ł.; Warmbier, M.; Kornicka, A. Synthesis, structure and cytotoxic properties of copper(II) complexes of 2-Iminocoumarins bearing a 1,3,5-triazine or benzoxazole/benzothiazole moiety. *Molecules* 2022, 27, 7155. [CrossRef]
- 18. Mijatović, A.; Gligorijević, N.; Ćoćić, D.; Spasić, S.; Lolić, A.; Aranđelović, S.; Nikolić, M.; Baošić, R. In vitro and in silico study of the biological activity of tetradentate Schiff base copper(II) complexes with ethylenediamine-bridge. *J. Inorg. Biochem.* 2023, 244, 112224. [CrossRef]
- 19. Ohui, K.; Stepanenko, I.; Besleaga, I.; Babak, M.V.; Stafi, R.; Darvasiova, D.; Giester, G.; Pósa, V.; Enyedy, E.A.; Vegh, D.; et al. Triapine derivatives act as copper delivery vehicles to induce deadly metal overload in cancer cells. *Biomolecules* **2020**, *10*, 1336. [CrossRef]
- 20. Mutlu Gençkal, H.; Erkisa, M.; Alper, P.; Sahin, S.; Ulukaya, E.; Ari, F. Mixed ligand complexes of Co(II), Ni(II) and Cu(II) with quercetin and diimine ligands: Synthesis, characterization, anti-cancer and anti-oxidant activity. *J. Biol. Inorg. Chem.* **2020**, 25, 161–177. [CrossRef]
- 21. Burgos-López, Y.; Balsa, L.M.; Piro, O.E.; León, I.E.; García-Tojal, J.; Echeverría, G.A.; González-Baró, A.C.; Parajón-Costa, B.S. Tridentate acylhydrazone copper(II) complexes with heterocyclic bases as coligands. Synthesis, spectroscopic studies, crystal structure and cytotoxicity assays. *Polyhedron* **2022**, *213*, 115621. [CrossRef]
- 22. Banti, C.N.; Tsiatouras, V.; Karanicolas, K.; Panagiotou, N.; Tasiopoulos, A.J.; Kourkoumelis, N.; Hadjikakou, S.K. Antiproliferative activity and apoptosis induction, of organo-antimony (III)-copper(I) conjugates, against human breast cancer cells. *Mol. Divers.* **2020**, *24*, 1095–1106. [CrossRef] [PubMed]
- 23. Romo, A.I.B.; Carepo, M.P.; Levín, P.; Nascimento, O.R.; Díaz, D.E.; Rodríguez-López, J.; León, I.E.; Bezerra, L.F.; Lemus, L.; Diógenes, I.C.N. Synergy of DNA intercalation and catalytic activity of a copper complex towards improved polymerase inhibition and cancer cell cytotoxicity. *Dalton Trans.* **2021**, *50*, 11931–11940. [CrossRef] [PubMed]
- 24. Alvarez, N.; Leite, C.M.; Napoleone, A.; Mendes, L.F.S.; Fernández, C.Y.; Ribeiro, R.R.; Ellena, J.; Batista, A.A.; Costa-Filho, A.J.; Facchin, G. Tetramethyl-phenanthroline copper complexes in the development of drugs to treat cancer: Synthesis, characterization and cytotoxicity studies of a series of copper(II)-l-dipeptide-3,4,7,8-tetramethyl-phenanthroline complexes. *J. Biol. Inorg. Chem.* 2022, 27, 431–441. [CrossRef] [PubMed]

25. Fernández, C.Y.; Alvarez, N.; Rocha, A.; Ellena, J.; Costa-Filho, A.J.; Batista, A.A.; Facchin, G. New copper(II)-L-dipeptide-bathophenanthroline complexes as potential anticancer agents-synthesis, characterization and cytotoxicity studies-and comparative DNA-binding study of related Phen complexes. *Molecules* **2023**, *28*, 896. [CrossRef]

- 26. Scalcon, V.; Bonsignore, R.; Aupič, J.; Thomas, S.R.; Folda, A.; Heidecker, A.A.; Pöthig, A.; Magistrato, A.; Casini, A.; Rigobello, M.P. Exploring the anticancer activity of tamoxifen-based metal complexes targeting mitochondria. *J. Med. Chem.* **2023**, *66*, 9823–9841. [CrossRef]
- 27. Aliabadi, F.; Sohrabi, B.; Mostafavi, E.; Pazoki-Toroudi, H.; Webster, T.J. Ubiquitin-proteasome system and the role of its inhib itors in cancer therapy. *Open Biol.* **2021**, *11*, 200390. [CrossRef]
- 28. Wittmann, C.; Dömötör, O.; Kuznetcova, I.; Spengler, G.; Reynisson, J.; Holder, L.; Miller, G.J.; Enyedy, E.A.; Bai, R.; Hamel, E.; et al. Indolo [2,3-e]benzazocines and indolo [2,3-f]benzazonines and their copper(II) complexes as microtubule destabilizing agents. *Dalton Trans.* 2023, 52, 9964–9982. [CrossRef]
- 29. Li, H.; Wang, J.; Wu, C.; Wang, L.; Chen, Z.S.; Cui, W. The combination of disulfiram and copper for cancer treatment. *Drug Discov. Today* **2020**, 25, 1099–1108. [CrossRef]
- 30. Balsa, L.M.; Ruiz, M.C.; Santa Maria de la Parra, L.; Baran, E.J.; León, I.E. Anticancer and antimetastatic activity of copper(II)-tropolone complex against human breast cancer cells, breast multicellular spheroids and mammospheres. *J. Inorg. Biochem.* 2020, 204, 110975. [CrossRef]
- 31. Chen, Y.; Li, H.; Liu, N.; Feng, D.; Wu, W.; Gu, K.; Wu, A.; Li, C.; Wang, X. Multi-mechanism antitumor/antibacterial effects of Cu-EGCG self-assembling nanocomposite in tumor nanotherapy and drug-resistant bacterial wound infections. *J. Colloid Interface Sci.* 2024, 671, 751–769. [CrossRef]
- 32. Abdolmaleki, S.; Panjehpour, A.; Khaksar, S.; Ghadermazi, M.; Rostamnia, S. Evaluation of central-metal effect on anticancer activity and mechanism of action of isostructural Cu(II) and Ni(II) complexes containing pyridine-2,6-dicarboxylate. *Eur. J. Med. Chem.* 2023, 245, 114897. [CrossRef] [PubMed]
- 33. El-Beshti, H.S.; Gercek, Z.; Kayi, H.; Yildizhan, Y.; Cetin, Y.; Adigüzel, Z.; Güngör, G.; Özalp-Yaman, Ş. Antiproliferative Activity of Platinum(II) and Copper(II) Complexes Containing Novel Biquinoxaline Ligands. *Metallomics* **2024**, *16*, mfae001. [CrossRef]
- 34. Jabłońska-Trypuć, A.; Wydro, U.; Wołejko, E.; Świderski, G.; Lewandowski, W. Biological Activity of New Cichoric Acid–Metal Complexes in Bacterial Strains, Yeast-Like Fungi, and Human Cell Cultures In Vitro. *Nutrients* 2020, 12, 154. [CrossRef] [PubMed]
- 35. Emami, F.; Aliomrani, M.; Tangestaninejad, S.; Kazemian, H.; Moradi, M.; Rostami, M. Copper-Curcumin-Bipyridine Dicarboxy-late Complexes as Anticancer Candidates. *Chem. Biodivers.* **2022**, *19*, e202200202. [CrossRef] [PubMed]
- 36. Nunes, P.; Yildizhan, Y.; Adiguzel, Z.; Marques, F.; Costa Pessoa, J.; Acilan, C.; Correia, I. Copper(II) and oxidovanadium(IV) complexes of chromone Schiff bases as potential anticancer agents. *JBIC J. Biol. Inorg. Chem.* **2022**, 27, 89–109. [CrossRef]
- Aguilar-Jiménez, Z.; González-Ballesteros, M.; Dávila-Manzanilla, S.G.; Espinoza-Guillén, A.; Ruiz-Azuara, L. Development and In Vitro and In Vivo Evaluation of an Antineoplastic Copper(II) Compound (Casiopeina III-ia) Loaded in Nonionic Vesicles Using Quality by Design. *Int. J. Mol. Sci.* 2022, 23, 12756. [CrossRef]
- 38. Kuznetcova, I.; Bacher, F.; Alfadul, S.M.; Tham, M.J.R.; Ang, W.H.; Babak, M.V.; Rapta, P.; Arion, V.B. Elucidation of Structure–Activity Relationships in Indolobenzazepine-Derived Ligands and Their Copper(II) Complexes: The Role of Key Structural Components and Insight into the Mechanism of Action. *Inorg. Chem.* 2022, *61*, 10167–10181. [CrossRef]
- Fernández, C.Y.; Alvarez, N.; Rocha, A.; Mendes, L.F.S.; Costa-Filho, A.J.; Ellena, J.; Batista, A.A.; Facchin, G. Phenanthroline and phenyl carboxylate mixed ligand copper complexes in developing drugs to treat cancer. *J. Inorg. Biochem.* 2024, 260, 112700. [CrossRef]
- 40. Rada, J.P.; Forté, J.; Gontard, G.; Corcé, V.; Salmain, M.; Rey, N.A. Isoxazole-Derived Aroylhydrazones and Their Dinuclear Copper(II) Complexes Show Antiproliferative Activity on Breast Cancer Cells with a Potentially Alternative Mechanism of Action. *ChemBioChem* 2020, 21, 2474–2486. [CrossRef]
- 41. Canakci, D.; Koyuncu, I.; Lolak, N.; Durgun, M.; Akocak, S.; Supuran, C.T. Synthesis and cytotoxic activities of novel copper and silver complexes of 1,3-diaryltriazene-substituted sulfonamides. *J. Enzyme Inhib. Med. Chem.* **2019**, *34*, 110–116. [CrossRef] [PubMed]
- 42. Al-Farraj, E.S.; Younis, A.M.; El-Reash, G.M.I.A. Synthesis, characterization, biological potency, and molecular docking of Co<sup>2+</sup>, Ni<sup>2+</sup> and Cu<sup>2+</sup> complexes of a benzoyl isothiocyanate based ligand. *Sci Rep.* **2024**, *14*, 10032. [CrossRef] [PubMed]
- Aly, A.A.M.; Zidan, A.S.A.; Ibrahim, A.B.M.; Mosbah, H.K.; Mayer, P.; Saber, S.H. Binuclear Cu(II) complex based on N-acetylanthranilic acid induces significant cytotoxic effect on three cancer cell lines. J. Mol. Struct. 2022, 1249, 131634.
  [CrossRef]
- 44. Rodrigues, J.A.O.; Oliveira Neto, J.G.; da Silva de Barros, A.O.; Ayala, A.P.; Santos-Oliveira, R.; de Menezes, A.S.; de Sousa, F.F. Copper(II):phenanthroline complexes with l-asparagine and l-methionine: Synthesis, crystal structure and in-vitro cytotoxic effects on prostate, breast and melanoma cancer cells. *Polyhedron* **2020**, *191*, 114807. [CrossRef]

Molecules **2025**, 30, 2104 17 of 25

45. Brustolin, L.; Pettenuzzo, N.; Nardon, C.; Quarta, S.; Montagner, I.; Pontisso, P.; Rosato, A.; Conte, P.; Merigliano, S.; Fregona, D. Labelled micelles for the delivery of cytotoxic Cu(II) and Ru(III) compounds in the treatment of aggressive orphan cancers: Design and biological in vitro data. *J. Inorg. Biochem.* **2020**, 213, 111259. [CrossRef]

- 46. Alvarez, N.; Velluti, F.; Guidali, F.; Serra, G.; Gabriela Kramer, M.; Ellena, J.; Facchin, G.; Scarone, L.; Torre, M.H. New BI and TRI-Thiazole copper (II) complexes in the search of new cytotoxic drugs against breast cancer cells. *Inorg. Chim. Acta* **2020**, 508, 119622. [CrossRef]
- 47. Chowdhury, M.; Biswas, N.; Saha, S.; Rahaman, A.; Gupta, P.S.; Banerjee, A.; Mandal, D.P.; Bhattacharjee, S.; Zangrando, E.; Sciortino, G.; et al. Interaction with CT-DNA and in vitro cytotoxicity of two new copper(II)-based potential drugs derived from octanoic hydrazide ligands. *J. Inorg. Biochem.* 2024, 256, 112546. [CrossRef]
- 48. El–Beshti, H.S.; Yildizhan, Y.; Kayi, H.; Cetin, Y.; Adigüzel, Z.; Gungor-Topcu, G.; Gercek, Z.; Özalp-Yaman, Ş. Anticancer investigation of platinum and copper-based complexes containing quinoxaline ligands. *J. Mol. Struct.* **2022**, 1250, 131928. [CrossRef]
- 49. Mathews, N.A.; Kurup, M.R.P. Copper(II) complexes as novel anticancer drug: Synthesis, spectral studies, crystal structures, in silico molecular docking and cytotoxicity. *J. Mol. Struct.* **2022**, *1258*, 132672. [CrossRef]
- 50. Aguirrechu-Comerón, A.; Oramas-Royo, S.; Pérez-Acosta, R.; Hernández-Molina, R.; Gonzalez-Platas, J.; Estévez-Braun, A. Preparation of new metallic complexes from 2-hydroxy-3-((5-methylfuran-2-yl)methyl)-1,4-naphthoquinone. *Polyhedron* 2020, 177, 114280. [CrossRef]
- 51. do Couto Almeida, J.; Silva, R.T.C.; Zanetti, R.D.; Moreira, M.B.; Portes, M.C.; Polloni, L.; Azevedo, F.V.d.V.; Von Poelhsitz, G.; Pivatto, M.; Netto, A.V.; et al. DNA interactions, antitubercular and cytotoxic activity of heteroleptic CuII complexes containing 1,10-phenanthroline. *J. Mol. Struct.* 2021, 1235, 130234. [CrossRef]
- 52. Barrett, S.; De Franco, M.; Donati, C.; Marzano, C.; Gandin, V.; Montagner, D. Novel Biotinylated Cu(II)-Phenanthroline Complexes: 2D and 3D Cytotoxic Activity and Mechanistic Insight. *Molecules* **2023**, *28*, 4112. [CrossRef] [PubMed]
- 53. Kuznetcova, I.; Ostojić, M.; Gligorijević, N.; Aranđelović, S.; Arion, V.B. Enriching Chemical Space of Bioactive Scaffolds by New Ring Systems: Benzazocines and Their Metal Complexes as Potential Anticancer Drugs. *Inorg. Chem.* **2022**, *61*, 20445–20460. [CrossRef]
- 54. Rigamonti, L.; Reginato, F.; Ferrari, E.; Pigani, L.; Gigli, L.; Demitri, N.; Kopel, P.; Tesarova, B.; Heger, Z. From solid state to in vitro anticancer activity of copper(II) compounds with electronically-modulated NNO Schiff base ligands. *Dalton Trans.* **2020**, 49, 14626–14639. [CrossRef] [PubMed]
- 55. Al-Asbahy, W.M.; Shamsi, M. Synthesis and characterization of copper-based anticancer compound; in vitro interaction studies with DNA/HSA, SOD mimetic, cytotoxic activity and molecular docking investigation. *J. Biomol. Struct. Dyn.* **2021**, *39*, 1431–1446. [CrossRef] [PubMed]
- 56. Konakanchi, R.; Pamidimalla, G.S.; Prashanth, J.; Naveen, T.; Kotha, L.R. Structural elucidation, theoretical investigation, biological screening and molecular docking studies of metal(II) complexes of NN donor ligand derived from 4-(2-aminopyridin-3-methylene)aminobenzoic acid. *BioMetals* **2021**, *34*, 529–556. [CrossRef]
- 57. Mohammadizadeh, F.; Mahmoodi, M.; Rezaei, A.; Mohamadi, M.; Hajizadeh, M.R.; Mirzaei, M.R.; Falahati-pour, S.K. A new copper complex enhanced apoptosis in human breast cancerous cells without considerable effects on normal cells. *Gene Rep.* **2019**, 17, 100475. [CrossRef]
- 58. Syed Ali Fathima, S.; Mohamed Sahul Meeran, M.; Nagarajan, E.R. Design and synthesis of novel pyrazolone based coordination compounds: DNA synergy, biological screening, apoptosis, molecular docking and in-silico ADMET profile. *J. Mol. Struct.* **2019**, 1197, 292–307. [CrossRef]
- 59. Sánchez-Lara, E.; Favela, R.; Tzian, K.; Monroy-Torres, B.; Romo-Pérez, A.; Ramírez-Apan, M.T.; Flores-Alamo, M.; Rodríguez-Diéguez, A.; Cepeda, J.; Castillo, I. Effects of the tetravanadate [V4O12]4- anion on the structural, magnetic, and biological properties of copper/phenanthroline complexes. *J. Biol. Inorg. Chem.* **2024**, *29*, 139–158. [CrossRef]
- 60. Li, J.; Yan, H.; Wang, Z.; Liu, R.; Luo, B.; Yang, D.; Chen, H.; Pan, L.; Ma, Z. Copper chloride complexes with substituted 4'-phenyl-terpyridine ligands: Synthesis, characterization, antiproliferative activities and DNA interactions. *Dalton Trans.* **2021**, 50, 8243–8257. [CrossRef]
- 61. Ni, K.; Montesdeoca, N.; Karges, J. Highly cytotoxic Cu(II) terpyridine complexes as chemotherapeutic agents. *Dalton Trans.* **2024**, 53, 8223–8228. [CrossRef] [PubMed]
- 62. Smoleński, P.; Śliwińska-Hill, U.; Kwiecień, A.; Wolińska, J.; Poradowski, D. Design, synthesis, and anti-cancer evaluation of novel water-soluble copper(I) complexes bearing terpyridine and PTA ligands. *Molecules* **2024**, *29*, 945. [CrossRef]
- 63. Malarz, K.; Zych, D.; Kuczak, M.; Musioł, R.; Mrozek-Wilczkiewicz, A. Anticancer activity of 4'-phenyl-2,2':6. *Eur. J. Med. Chem.* **2020**, *189*, 112039. [CrossRef]
- 64. Liu, C.; Jiang, J.; Li, J.; Liang, X.; Zhou, Y.; Chen, H.; Ma, Z. Synthesis, structural characterization and antiproliferative potential of copper 4'-phenyl-terpyridine complexes constructed from building block reaction. *Polyhedron* **2020**, *182*, 114465. [CrossRef]

Molecules **2025**, 30, 2104 18 of 25

65. Jevtovic, V.; Alshamari, A.K.; Milenković, D.; Dimitrić Marković, J.; Marković, Z.; Dimić, D. The effect of metal ions (Fe, Co, Ni, and Cu) on the molecular-structural, protein binding, and cytotoxic properties of metal pyridoxal-thiosemicarbazone complexes. *Int. J. Mol. Sci.* 2023, 24, 11910. [CrossRef] [PubMed]

- 66. Manakkadan, V.; Haribabu, J.; Palakkeezhillam, V.N.V.; Rasin, P.; Vediyappan, R.; Kumar, V.S.; Garg, M.; Bhuvanesh, N.; Sreekanth, A. Copper-mediated cyclization of thiosemicarbazones leading to 1,3,4-thiadiazoles: Structural elucidation, DFT calculations, in vitro biological evaluation and in silico evaluation studies. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 2024, 313, 124117. [CrossRef]
- 67. Barrett, S.; De Franco, M.; Kellett, A.; Dempsey, E.; Marzano, C.; Erxleben, A.; Gandin, V.; Montagner, D. Anticancer activity, DNA binding and cell mechanistic studies of estrogen-functionalised Cu(II) complexes. J. Biol. Inorg. Chem. 2020, 25, 49–60. [CrossRef]
- 68. Bergamini, F.R.G.; Nunes, J.H.B.; Manzano, C.M.; de Carvalho, M.A.; Ribeiro, M.A.; Ruiz, A.L.T.G.; de Carvalho, J.E.; Lustri, W.R.; de Paiva, R.E.F.; Portes, M.C.; et al. Investigating the antiproliferative activities of new CuII complexes with pyridine hydrazone derivatives of nalidixic acid. *J. Inorg. Biochem.* 2022, 234, 111881. [CrossRef]
- 69. Navarro-Peñaloza, R.; Vázquez-Palma, A.B.; López-Sandoval, H.; Sánchez-Bartéz, F.; Gracia-Mora, I.; Barba-Behrens, N. Coordination compounds with heterocyclic ester derivatives. Structural characterization and anti-proliferative activity. *J. Inorg. Biochem.* **2021**, 219, 111432. [CrossRef]
- 70. Hawash, M. Recent advances of tubulin inhibitors targeting the colchicine Binding Site for cancer therapy. *Biomolecules* **2022**, 12, 1843. [CrossRef]
- 71. Vitomirov, T.; Dimiza, F.; Matić, I.Z.; Stanojković, T.; Pirković, A.; Živković, L.; Spremo-Potparević, B.; Novaković, I.; Anđelković, K.; Milčić, M.; et al. Copper(II) complexes with 4-(diethylamino)salicylaldehyde and α-diimines: Anticancer, antioxidant, antigenotoxic effects and interaction with DNA and albumins. *J. Inorg. Biochem.* **2022**, 235, 111942. [CrossRef]
- 72. Dömötör, O.; Kiss, M.A.; Gál, G.T.; May, N.V.; Spengler, G.; Nové, M.; Gašparović, A.Č.; Frank, É.; Enyedy, É.A. Solution equilibrium, structural and cytotoxicity studies on Ru(η6-p-cymene) and copper complexes of pyrazolyl thiosemicarbazones. *J. Inorg. Biochem.* **2020**, 202, 110883. [CrossRef]
- 73. Hou, L.; Jia, X.; Wu, Y.; Li, J.; Yao, D.; Gou, Y.; Huang, G. Aroylhydrazone Cu(II) complexes: Syntheses, crystal structures, and anticancer properties. *J. Mol. Struct.* **2021**, 1239, 130469. [CrossRef]
- 74. Komarnicka, U.K.; Kozieł, S.; Zabierowski, P.; Kruszyński, R.; Lesiów, M.K.; Tisato, F.; Porchia, M.; Kyzioł, A. Copper(I) complexes with phosphines P(p-OCH3-Ph)2CH2OH and P(p-OCH3-Ph)2CH2SarGly. Synthesis, multimodal DNA interactions, and prooxidative and in vitro antiproliferative activity. *J. Inorg. Biochem.* **2020**, 203, 110926. [CrossRef]
- Massoud, S.S.; Louka, F.R.; Salem, N.M.H.; Fischer, R.C.; Torvisco, A.; Mautner, F.A.; Vančo, J.; Belza, J.; Dvořák, Z.; Trávníček, Z. Dinuclear doubly bridged phenoxido copper(II) complexes as efficient anticancer agents. Eur. J. Med. Chem. 2023, 246, 114992. [CrossRef] [PubMed]
- 76. Nath, H.; Sharma, P.; Gomila, R.M.; Frontera, A.; Barceló-Oliver, M.; Verma, A.K.; Dutta, K.; Bhattacharyya, M.K. Unconventional enclathration of guest adipic acid and energetically significant antiparallel π-stacked ternary assemblies involving unusual regium-π(chelate) contacts in phenanthroline-based Ni(II) and Cu(II) compounds-antiproliferative evaluation and theoretical studies. *J. Mol. Struct.* **2021**, *1245*, 131038. [CrossRef]
- 77. Shao, J.; Li, M.; Guo, Z.; Jin, C.; Zhang, F.; Ou, C.; Xie, Y.; Tan, S.; Wang, Z.; Zheng, S.; et al. TPP-related mitochondrial targeting copper (II) complex induces p53-dependent apoptosis in hepatoma cells through ROS-mediated activation of Drp1. *Cell Commun. Signal* 2019, 17, 149. [CrossRef]
- 78. Zarei, L.; Asadi, Z.; Samolova, E.; Dusek, M.; Amirghofran, Z. Pyrazolate as bridging ligand in stabilization of self-assemble Cu(II) Schiff base complexes: Synthesis, structural investigations, DNA/protein (BSA) binding and growth inhibitory effects on the MCF7, CT-26, MDA-MB-231 cell lines. *Inorganica Chim. Acta* 2020, 509, 119674. [CrossRef]
- 79. Aranda, E.E.; da Luz, J.S.; Oliveira, C.C.; Divina Petersen, P.A.; Petrilli, H.M.; da Costa Ferreira, A.M. Heterobinuclear copper(II) platinum(II) complexes with oxindolimine ligands: Interactions with DNA, and inhibition of kinase and alkaline phosphatase proteins. *J. Inorg. Biochem.* **2020**, 203, 110863. [CrossRef]
- 80. Varna, D.; Geromichalos, G.; Gioftsidou, D.K.; Tzimopoulos, D.; Hatzidimitriou, A.G.; Dalezis, P.; Papi, R.; Trafalis, D.; Angaridis, P.A. N-heterocyclic-carbene vs diphosphine auxiliary ligands in thioamidato Cu(I) and Ag(I) complexes towards the development of potent and dual-activity antibacterial and apoptosis-inducing anticancer agents. *J. Inorg. Biochem.* 2024, 252, 112472. [CrossRef]
- 81. Varna, D.; Geromichalos, G.D.; Dalezis, P.; Hatzidimitriou, A.G.; Psomas, G.; Zachariadis, G.; Psatha, K.; Aivaliotis, M.; Papi, R.; Trafalis, D.; et al. Amine-substituted heterocyclic thioamide Cu(I) and Ag(I) complexes as effective anticancer and antibacterial agents targeting the periplasm of E. coli bacteria. *Eur. J. Med. Chem.* **2024**, 277, 116746. [CrossRef] [PubMed]
- 82. Balsa, L.M.; Rodriguez, M.R.; Parajón-Costa, B.S.; González-Baró, A.C.; Lavecchia, M.J.; León, I.E. Anticancer Activity and Mechanism of Action Evaluation of an Acylhydrazone Cu(II) Complex toward Breast Cancer Cells, Spheroids, and Mammospheres. *ChemMedChem* 2022, 16, 17. [CrossRef] [PubMed]

83. Fahim, A.M.; Dacrory, S.; Hashem, A.H.; Kamel, S. Antimicrobial, anticancer activities, molecular docking, and DFT/B3LYP/LANL2DZ analysis of heterocyclic cellulose derivative and their Cu-complexes. *Int. J. Biol. Macromol.* 2024, 269, 132027. [CrossRef]

- 84. Mariani, D.; Ghasemishahrestani, Z.; Freitas, W.; Pezzuto, P.; Costa-da-Silva, A.C.; Tanuri, A.; Kanashiro, M.; Fernandes, C.; Horn, A.; Pereira, M.D. Antitumoral synergism between a copper(II) complex and cisplatin improves in vitro and in vivo anticancer activity against melanoma, lung and breast cancer cells. *Biochim. Biophys. Acta (BBA)-Gen. Subj.* 2021, 1865, 129963. [CrossRef]
- 85. Ghorbanpour, M.; Soltani, B.; Mota, A.; Jahanbin Sardroodi, J.; Mehdizadeh Aghdam, E.; Shayanfar, A.; Molavi, O.; Mohammad-Rezaei, R.; Ebadi-Nahari, M.; Ziegler, C.J. Copper (II) complexes with N, S donor pyrazole-based ligands as anticancer agents. *BioMetals* 2022, 35, 1095–1111. [CrossRef] [PubMed]
- 86. Akhmetova, V.R.; Akhmadiev, N.S.; Gubaidullin, A.T.; Samigullina, A.I.; Glazyrin, A.B.; Sadykov, R.A.; Ishmetova, D.V.; Vakhitova, Y.V. Novel binuclear copper(II) complexes with sulfanylpyrazole ligands: Synthesis, crystal structure, fungicidal, cytostatic, and cytotoxic activity. *Metallomics* 2024, 16, mfae024. [CrossRef]
- 87. Ajibade, P.A.; Andrew, F.P.; Botha, N.L.; Solomane, N. Synthesis, Crystal Structures and Anticancer Studies of Morpholinyldithio-carbamato Cu(II) and Zn(II) Complexes. *Molecules* **2020**, *25*, 3584. [CrossRef]
- 88. Shao, J.; Zhang, Q.; Wei, J.; Yuchi, Z.; Cao, P.; Li, S.Q.; Wang, S.; Xu, J.-Y.; Yang, S.; Zhang, Y.; et al. Synthesis, crystal structures, anticancer activities and molecular docking studies of novel thiazolidinone Cu(II) and Fe(II) complexes targeting lysosomes: Special emphasis on their binding to DNA/BSA. *Dalton Trans.* 2021, 50, 13387–13398. [CrossRef]
- 89. Śliwa, E.I.; Śliwińska-Hill, U.; Bażanów, B.; Siczek, M.; Kłak, J.; Smoleński, P. Synthesis, Structural, and Cytotoxic Properties of New Water-Soluble Copper(II) Complexes Based on 2,9-Dimethyl-1,10-Phenanthroline and Their One Derivative Containing 1,3,5-Triaza-7-Phosphaadamantane-7-Oxide. *Molecules* 2020, 25, 741. [CrossRef]
- 90. Al-Harbi, S.A.; Al-Saidi, H.M.; Debbabi, K.F.; Allehyani, E.S.; Alqorashi, A.A.; Emara, A.A.A. Design and anti-tumor evaluation of new platinum(II) and copper(II) complexes of nitrogen compounds containing selenium moieties. *J. Saudi Chem. Soc.* **2020**, 24, 982–995. [CrossRef]
- 91. Al-Noaimi, M.; Awwadi, F.F.; Al-Wahaib, D.; Bardaweel, S.; Alhmaideen, A.; Alshammari, M. Competition between Cu-Br semi-coordinate bond and C-H···Br, C-H···S and S···S interactions; new two thioalkylazothiophenol (SNS) copper (II) dimers [Cu(L)(μ-Br)]2. *Polyhedron* **2024**, 250, 116827. [CrossRef]
- 92. Patel, A.K.; Jadeja, R.N.; Roy, H.; Patel, R.N.; Patel, S.K.; Butcher, R.J.; Cortijo, M.; Herrero, S. Copper(II) hydrazone complexes with different nuclearities and geometries: Synthesis, structural characterization, antioxidant SOD activity and antiproliferative properties. *Polyhedron* 2020, 186, 114624. [CrossRef]
- 93. Porchia, M.; Tisato, F.; Zancato, M.; Gandin, V.; Marzano, C. In vitro antitumor activity of water-soluble copper(I) complexes with diimine and monodentate phosphine ligands. *Arab. J. Chem.* **2020**, *13*, 998–1010. [CrossRef]
- 94. Kiwaan, H.A.; El-Mowafy, A.S.; El-Bindary, A.A. Synthesis, spectral characterization, DNA binding, catalytic and in vitro cytotoxicity of some metal complexes. *J. Mol. Liq.* **2021**, *326*, 115381. [CrossRef]
- 95. Babgi, B.A.; Mashat, K.H.; Abdellattif, M.H.; Arshad, M.N.; Alzahrani, K.A.; Asiri, A.M.; Du, J.; Humphrey, M.G.; Hussien, M.A. Synthesis, structures, DNA-binding, cytotoxicity and molecular docking of CuBr(PPh3)(diimine). *Polyhedron* **2020**, *192*, 114847. [CrossRef]
- 96. Fatemikia, H.; Keypour, H.; Zeynali, H.; Karamian, R.; Ranjbar, N.; Gable, R.W. The X-ray crystal structures, molecular docking and biological activities of two novel Cu(II) and Zn(II) complexes with a ligand having a potentially N4O2 donor set and two nitro phenyl rings as pendant arms. *J. Inorg. Biochem.* 2022, 235, 111910. [CrossRef] [PubMed]
- 97. Shahabadi, N.; Shiri, F.; Hadidi, S.; Farshadfar, K.; Sajadimajd, S.; Roe, S.M. Equilibrium and site selective analysis for DNA threading intercalation of a new phosphine copper(I) complex: Insights from X-ray analysis, spectroscopic and molecular modeling studies. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2020**, 235, 118280. [CrossRef]
- 98. Memišević, M.; Zahirović, A.; Višnjevac, A.; Osmanović, A.; Žilić, D.; Kralj, M.; Muratović, S.; Martin-Kleiner, I.; Završnik, D.; Kahrović, E. Copper(II) salicylideneimine complexes revisited: From a novel derivative and extended characterization of two homologues to interaction with BSA and antiproliferative activity. *Inorganica Chim. Acta* 2021, 525, 120460. [CrossRef]
- 99. Dimitrijević, J.; Arsenijević, A.N.; Milovanović, M.Z.; Arsenijević, N.N.; Milovanović, J.Z.; Stanković, A.S.; Bukonjić, A.M.; Tomović, D.L.; Ratković, Z.R.; Potočňák, I.; et al. Synthesis, characterization and cytotoxic activity of binuclear copper(II)-complexes with some S-isoalkyl derivatives of thiosalicylic acid. Crystal structure of the binuclear copper(II)-complex with S-isopropyl derivative of thiosalicylic acid. *J. Inorg. Biochem.* 2020, 208, 111078. [CrossRef]
- 100. Petrasheuskaya, T.V.; Wernitznig, D.; Kiss, M.A.; May, N.V.; Wenisch, D.; Keppler, B.K.; Frank, É.; Enyedy, É.A. Estrone-salicylaldehyde N-methylated thiosemicarbazone hybrids and their copper complexes: Solution structure, stability and anticancer activity in tumour spheroids. *J. Biol. Inorg. Chem.* **2021**, *26*, 775–791. [CrossRef] [PubMed]
- 101. Gandhi, D.H.; Vaidya, F.U.; Pathak, C.; Patel, T.N.; Bhatt, B.S. Mechanistic insight of cell anti-proliferative activity of fluoro-quinolone drug-based Cu(II) complexes. *Mol. Divers.* **2022**, *26*, 869–878. [CrossRef]

Molecules **2025**, 30, 2104 20 of 25

102. Chrzanowska, A.; Drzewiecka-Antonik, A.; Dobrzyńska, K.; Stefańska, J.; Pietrzyk, P.; Struga, M.; Bielenica, A. The Ccytotoxic effect of copper (II) complexes with halogenated 1,3-Disubstituted arylthioureas on cancer and bacterial cells. *Int. J. Mol. Sci.* **2021**, 22, 11415. [CrossRef] [PubMed]

- 103. De Oliveira Neto, J.G.; Filho, J.G.S.; Bittar, E.M.; Silva, L.M.; de Sousa, F.F.; Domingos, H.V.; Costa-Lotufo, L.V.; Reis, A.S.; Dos Santos, A.O. Structural, thermal, electronic, vibrational, magnetic, and cytotoxic properties of chloro(glycinato-N,O)(1,10-phenanthroline-N,N')-copper(II) trihydrate coordination complex. *J. Inorg. Biochem.* 2022, 226, 111658. [CrossRef] [PubMed]
- 104. Faghih, Z.; Neshat, A.; Mastrorilli, P.; Gallo, V.; Faghih, Z.; Gilanchi, S. Cu(II), Ni(II) and Co(II) complexes with homoscorpionate Bis(2-Mercaptobenzimidazolyl) and Bis(2-Mercaptobenzothiazolyl)borate ligands: Synthesis and in vitro cytotoxicity studies. *Inorganica Chim. Acta.* 2020, 512, 119896. [CrossRef]
- 105. Abousaty, A.I.; Reda, F.M.; Hassanin, W.A.; Felifel, W.M.; El-Shwiniy, W.H.; Selim, H.M.R.M.; Bendary, M.M. Sorbate metal complexes as newer antibacterial, antibiofilm, and anticancer compounds. *BMC Microbiol.* **2024**, 24, 262. [CrossRef] [PubMed]
- 106. Pinho, J.O.; da Silva, I.V.; Amaral, J.D.; Rodrigues, C.M.P.; Casini, A.; Soveral, G.; Gaspar, M.M. Therapeutic potential of a copper complex loaded in pH-sensitive long circulating liposomes for colon cancer management. *Int. J. Pharm.* **2021**, 599, 120463. [CrossRef]
- 107. Milunović, M.N.M.; Palamarciuc, O.; Sirbu, A.; Shova, S.; Dumitrescu, D.; Dvoranová, D.; Rapta, P.; Petrasheuskaya, T.V.; Enyedy, E.A.; Spengler, G.; et al. Insight into the anticancer activity of copper(II) 5-Methylenetrimethylammonium-Thiosemicarbazonates and their interaction with organic cation transporters. *Biomolecules* **2020**, *10*, 1213. [CrossRef]
- 108. Mastrangelo, S.; Attina, G.; Triarico, S.; Romano, A.; Maurizi, P.; Ruggiero, A. The DNA-topoisomerase inhibitors in cancer therapy. *Biomed. Pharmacol. J.* **2022**, *15*, 553–562. [CrossRef]
- 109. Wittmann, C.; Bacher, F.; Enyedy, E.A.; Dömötör, O.; Spengler, G.; Madejski, C.; Reynisson, J.; Arion, V.B. Highly antiproliferative Latonduine and indolo [2,3-c]quinoline Derivatives: Complex Formation with copper(II) Markedly Changes the kinase Inhibitory Profile. *J. Med. Chem.* 2022, 65, 2238–2261. [CrossRef] [PubMed]
- 110. Stevanović, N.; Zlatar, M.; Novaković, I.; Pevec, A.; Radanović, D.; Matić, I.Z.; Đorđić Crnogorac, M.; Stanojković, T.; Vujčić, M.; Gruden, M.; et al. Cu(ii), Mn(ii) and Zn(ii) complexes of hydrazones with a quaternary ammonium moiety: Synthesis, experimental and theoretical characterization and cytotoxic activity. *Dalton Trans.* 2022, 51, 185–196. [CrossRef] [PubMed]
- 111. Balewski, Ł.; Plech, T.; Korona-Głowniak, I.; Hering, A.; Szczesio, M.; Olczak, A.; Bednarski, P.J.; Kokoszka, J.; Kornicka, A. Copper(II) complexes with 1-(Isoquinolin-3-yl)heteroalkyl-2-ones: Synthesis, structure and evaluation of anticancer, antimicrobial and antioxidant potential. *Int. J. Mol. Sci.* 2023, 25, 8. [CrossRef] [PubMed]
- 112. Ruiz, M.C.; Perelmulter, K.; Levín, P.; Romo, A.I.B.; Lemus, L.; -Fogolín, M.B.; León, I.E.; Di Virgilio, A.L. Antiproliferative activity of two copper (II) complexes on colorectal cancer cell models: Impact on ROS production, apoptosis induction and NF-κB inhibition. *Eur. J. Pharm. Sci.* **2022**, *169*, 106092. [CrossRef]
- 113. Gourdon-Grünewaldt, L.; Blacque, O.; Gasser, G.; Cariou, K. Towards Copper(I) Clusters for Photo-Induced Oxidation of Biological Thiols in Living Cells. *ChemBioChem.* **2024**, *24*, e202300496. [CrossRef] [PubMed]
- 114. Klockner, I.; Schutt, C.; Gerhardt, T.; Boettger, T.; Braun, T. Control of CRK-RAC1 activity by the miR-1/206/133 miRNA family is essential for neuromuscular junction function. *Nat. Commun.* 2022, *13*, 3180. [CrossRef]
- 115. García-Valdivia, A.A.; Cepeda, J.; Fernández, B.; Medina-O'donnell, M.; Oyarzabal, I.; Parra, J.; Jannus, F.; Choquesillo-Lazarte, D.; García, J.A.; Lupiáñez, J.A.; et al. 5-Aminopyridine-2-carboxylic acid as appropriate ligand for constructing coordination polymers with luminescence, slow magnetic relaxation and anti-cancer properties. *J. Inorg. Biochem.* 2020, 207, 111051. [CrossRef]
- 116. Joksimović, N.; Petronijević, J.; Radisavljević, S.; Petrović, B.; Mihajlović, K.; Janković, N.; Milović, E.; Milivojević, D.; Ilić, B.; Djurić, A. Synthesis, characterization, antitumor potential, and investigation of mechanism of action of copper(II) complexes with acylpyruvates as ligands: Interactions with biomolecules and kinetic study. *RSC Adv.* 2022, 12, 30501–30513. [CrossRef] [PubMed]
- 117. Enyedy, É.A.; Petrasheuskaya, T.V.; Kiss, M.A.; Wernitznig, D.; Wenisch, D.; Keppler, B.K.; Spengler, G.; May, N.V.; Frank, É.; Dömötör, O. Complex formation of an estrone-salicylaldehyde semicarbazone hybrid with copper(II) and gallium(III): Solution equilibria and biological activity. *J. Inorg. Biochem.* **2021**, 220, 111468. [CrossRef]
- 118. Pellei, M.; Santini, C.; Bagnarelli, L.; Battocchio, C.; Iucci, G.; Venditti, I.; Meneghini, C.; Amatori, S.; Sgarbossa, P.; Marzano, C.; et al. Exploring the antitumor potential of copper complexes based on ester derivatives of Bis(pyrazol-1-yl)acetate ligands. *Int. J. Mol. Sci.* 2022, 23, 9397. [CrossRef]
- 119. Wu, Y.; Hou, L.; Lan, J.; Yang, F.; Huang, G.; Liu, W.; Gou, Y. Mixed-ligand copper(II) hydrazone complexes: Synthesis, structure, and anti-lung cancer properties. *J. Mol. Struct.* **2023**, 1279, 134986. [CrossRef]
- 120. Jain, S.; Bhar, K.; Kumar, S.; Bandyopadhyaya, S.; Tapryal, S.; Mandal, C.C.; Sharma, A.K. Homo- and heteroleptic trimethoxy terpyridine-Cu(ii) complexes: Synthesis, characterization, DNA/BSA binding, DNA cleavage and cytotoxicity studies. *Dalton Trans.* 2020, 49, 4100–4113. [CrossRef]
- 121. Trávníček, Z.; Vančo, J.; Belza, J.; Zoppellaro, G.; Dvořák, Z.; Beláková, B.; Schmid, J.A.; Molčanová, L.; Šmejkal, K. C-Geranylated flavanone diplacone enhances in vitro antiproliferative and anti-inflammatory effects in its copper(II) complexes. *J. Inorg. Biochem.* **2024**, 258, 112639. [CrossRef] [PubMed]

Molecules **2025**, 30, 2104 21 of 25

122. Caro-Ramírez, J.Y.; Rivas, M.G.; Gonzalez, P.J.; Williams, P.A.M.; Naso, L.G.; Ferrer, E.G. Copper(II) cation and bathophenanthroline coordination enhance therapeutic effects of naringenin against lung tumor cells. *BioMetals* **2022**, *35*, 1059–1076. [CrossRef]

- 123. Lu, X.; Lin, B.; Xu, N.; Huang, H.; Wang, Y.; Lin, J.M. Evaluation of the accumulation of disulfiram and its copper complex in A549 cells using mass spectrometry. *Talanta* **2020**, 211, 120732. [CrossRef]
- 124. Zhang, Y.-L.; Deng, C.-X.; Zhou, W.-F.; Zhou, L.-Y.; Cao, Q.-Q.; Shen, W.-Y.; Liang, H.; Chen, Z.-F. Synthesis and in vitro antitumor activity evaluation of copper(II) complexes with 5-pyridin-2-yl-[1,3]dioxolo [4,5-g]isoquinoline derivatives. *J. Inorg. Biochem.* 2019, 201, 110820. [CrossRef]
- 125. Enikeeva, K.R.; Shamsieva, A.V.; Kasimov, A.I.; Litvinov, I.A.; Lyubina, A.P.; Voloshina, A.D.; Musina, E.I.; Karasik, A.A. Pyridyl-containing dialkylphosphine oxides and their chelate copper(II) complexes. *Inorganica Chim. Acta* 2023, 545, 121286. [CrossRef]
- 126. Adhikari, S.; Bhattacharjee, T.; Butcher, R.J.; Porchia, M.; De Franco, M.; Marzano, C.; Gandin, V.; Tisato, F. Synthesis and characterization of mixed-ligand Zn(II) and Cu(II) complexes including polyamines and dicyano-dithiolate(2-): In vitro cytotoxic activity of Cu(II) compounds. *Inorganica Chim. Acta.* 2019, 498, 119098. [CrossRef]
- 127. Pantelic, L.; Skaro Bogojevic, S.; Andrejević, T.P.; Pantović, B.V.; Marković, V.R.; Ašanin, D.P.; Milanović, Ž.; Ilic-Tomic, T.; Nikodinovic-Runic, J.; Glišić, B.D.; et al. Copper(II) and Zinc(II) Complexes with Bacterial Prodigiosin Are Targeting Site III of Bovine Serum Albumin and Acting as DNA Minor Groove Binders. *Int. J. Mol. Sci.* 2024, 25, 8395. [CrossRef]
- 128. Komarnicka, U.K.; Pucelik, B.; Wojtala, D.; Lesiów, M.K.; Stochel, G.; Kyzioł, A. Evaluation of anticancer activity in vitro of a stable copper(I) complex with phosphine-peptide conjugate. *Sci. Rep.* **2021**, *11*, 23943. [CrossRef]
- 129. Pradhan, R.; Tiwari, L.; Groner, V.M.; Leach, C.; Lusk, K.; Harrison, N.S.; Cornell, K.A.; Waynant, K.V. Evaluation of azothioformamides and their copper(I) and silver(I) complexes for biological activity. *J. Inorg. Biochem.* **2023**, 246, 112294. [CrossRef]
- 130. Jozefíková, F.; Perontsis, S.; Koňáriková, K.; Švorc, L'.; Mazúr, M.; Psomas, G.; Moncol, J. In vitro biological activity of copper(II) complexes with NSAIDs and nicotinamide: Characterization, DNA- and BSA-interaction study and anticancer activity. *J. Inorg. Biochem.* **2022**, 228, 111696. [CrossRef]
- 131. Wojciechowska, A.; Bregier–Jarzębowska, R.; Komarnicka, U.K.; Kozieł, S.; Szuster–Ciesielska, A.; Sztandera–Tymoczek, M.; Jarząb, A.; Staszak, Z.; Witkowska, D.; Bojarska–Junak, A.; et al. Isothiocyanate l–argininato copper(II) complexes—Solution structure, DNA interaction, anticancer and antimicrobial activity. *Chem. Biol. Interact.* 2021, 348, 109636. [CrossRef] [PubMed]
- 132. Bansal, A.; Saleh-E-In, M.d.M.; Kar, P.; Roy, A.; Sharma, N.R. Synthesis of Carvacrol Derivatives as Potential New Anticancer Agent against Lung Cancer. *Molecules* **2022**, 27, 4597. [CrossRef] [PubMed]
- 133. Qiao, Y.; Chen, Y.; Zhang, S.; Huang, Q.; Zhang, Y.; Li, G. Six novel complexes based on 5-Acetoxy-1-(6-chloro-pyridin-2-yl)-1H-pyrazole-3-carboxylic acid methyl ester derivatives: Syntheses, crystal structures, and anti-cancer activity. *Arab. J. Chem.* **2021**, 14, 103237. [CrossRef]
- 134. Jain, S.; Bhar, K.; Bandyopadhayaya, S.; Singh, V.K.; Mandal, C.C.; Tapryal, S.; Sharma, A.K. Development, evaluation and effect of anionic co-ligand on the biological activity of benzothiazole derived copper(II) complexes. *J. Inorg. Biochem.* **2020**, 210, 111174. [CrossRef]
- 135. Umar, Q.; Huang, Y.H.; Nazeer, A.; Yin, H.; Zhang, J.C.; Luo, M.; Meng, X.G. Synthesis, characterization and anticancer activities of Zn<sup>2+</sup>, Cu<sup>2+</sup>, Co<sup>2+</sup> and Ni<sup>2+</sup> complexes involving chiral amino alcohols. *RSC Adv.* **2022**, *12*, 32119–32128. [CrossRef]
- 136. Du, L.-Q.; Zeng, C.-J.; Mo, D.-Y.; Qin, Q.-P.; Tan, M.-X.; Liang, H. 8-hydroxyquinoline-N-oxide copper(II)- and zinc(II)- phenanthroline and bipyridine coordination compounds: Design, synthesis, structures, and antitumor evaluation. *J. Inorg. Biochem.* **2024**, 251, 112443. [CrossRef]
- 137. Liu, M.; Song, X.Q.; Wu, Y.D.; Qian, J.; Xu, J.Y. Cu(ii)-TACN complexes selectively induce antitumor activity in HepG-2 cells via DNA damage and mitochondrial-ROS-mediated apoptosis. *Dalton Trans.* **2020**, *49*, 114–123. [CrossRef]
- 138. Gou, Y.; Chen, M.; Li, S.; Deng, J.; Li, J.; Fang, G.; Yang, F.; Huang, G. Dithiocarbazate-copper complexes for bioimaging and treatment of pancreatic cancer. *J. Med. Chem.* **2021**, *64*, 5485–5499. [CrossRef]
- 139. Asadi, Z.; Zarei, L.; Golchin, M.; Skorepova, E.; Eigner, V.; Amirghofran, Z. A novel Cu(II) distorted cubane complex containing Cu4O4 core as the first tetranuclear catalyst for temperature dependent oxidation of 3,5-di-tert-butyl catechol and in interaction with DNA & protein (BSA). *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 2020, 227, 117593. [CrossRef]
- 140. Novoa-Ramírez, C.S.; Silva-Becerril, A.; González-Ballesteros, M.M.; Gomez-Vidal, V.; Flores-Álamo, M.; Ortiz-Frade, L.; Gracia-Mora, J.; Ruiz-Azuara, L. Biological activity of mixed chelate copper(II) complexes, with substituted diimine and tridentate Schiff bases (NNO) and their hydrogenated derivatives as secondary ligands: Casiopeina's fourth generation. *J. Inorg. Biochem.* 2023, 242, 112097. [CrossRef]
- 141. Godínez-Loyola, Y.; Gracia-Mora, J.; Rojas-Montoya, I.D.; Hernández-Ayala, L.F.; Reina, M.; Ortiz-Frade, L.A.; Rascón-Valenzuela, L.A.; Robles-Zepeda, R.E.; Gómez-Vidales, V.; Bernad-Bernad, M.J.; et al. Casiopeinas<sup>®</sup> third generation, with indomethacin: Synthesis, characterization, DFT studies, antiproliferative activity, and nanoencapsulation. *RSC Adv.* **2022**, *12*, 21662–21673. [CrossRef] [PubMed]

Molecules **2025**, 30, 2104 22 of 25

142. Reheman, D.; Zhao, J.; Guan, S.; Xu, G.C.; Li, Y.J.; Sun, S.R. Apoptotic effect of novel pyrazolone-based derivative [Cu(PMPP-SAL)(EtOH)] on HeLa cells and its mechanism. *Sci. Rep.* **2020**, *10*, 18235. [CrossRef] [PubMed]

- 143. Barad, S.; Chaudhari, K.; Jadeja, R.N.; Roy, H.; Choquesillo-Lazarte, D. Square pyramidal Cu(II) acylpyrazolone complex: Synthesis, characterization, crystal structure, DFT and Hirshfeld analysis, in-vitro anti-cancer evaluation. *J. Mol. Struct.* **2023**, 1294, 136345. [CrossRef]
- 144. Yang, P.; Zhang, D.D.; Wang, Z.Z.; Liu, H.Z.; Shi, Q.S.; Xie, X.B. Copper(II) complexes with NNO ligands: Synthesis, crystal structures, DNA cleavage, and anticancer activities. *Dalton Trans.* **2019**, *48*, 17925–17935. [CrossRef]
- 145. Jana, A.; Aher, A.; Brandao, P.; Sharda, S.; Bera, P.; Phadikar, U.; Manna, S.K.; Mahapatra, A.K.; Bera, P. Dissociation of a tripodal pyridyl-pyrazole ligand and assortment of metal complex: Synthesis, structure, DFT, thermal stability, cytotoxicity, DNA cleavage, and molecular docking studies. *J. Mol. Struct.* 2022, 1256, 132479. [CrossRef]
- 146. Parsekar, S.U.; Velankanni, P.; Sridhar, S.; Haldar, P.; Mate, N.A.; Banerjee, A.; Sudhadevi Antharjanam, P.K.; Koley, A.P.; Kumar, M. Protein binding studies with human serum albumin, molecular docking and in vitro cytotoxicity studies using HeLa cervical carcinoma cells of Cu(II)/Zn(II) complexes containing a carbohydrazone ligand. *Dalton Trans.* **2020**, *49*, 2947–2965. [CrossRef] [PubMed]
- 147. Alvarez, N.; Viña, D.; Leite, C.M.; Mendes, L.F.S.; Batista, A.A.; Ellena, J.; Costa-Filho, A.J.; Facchin, G. Synthesis and structural characterization of a series of ternary copper(II)-L-dipeptide-neocuproine complexes. Study of their cytotoxicity against cancer cells including MDA-MB-231, triple negative breast cancer cells. *J. Inorg. Biochem.* **2020**, 203, 110930. [CrossRef]
- 148. Qu, J.-J.; Bai, P.; Liu, W.-N.; Liu, Z.-L.; Gong, J.-F.; Wang, J.-X.; Zhu, X.; Song, B.; Hao, X.-Q. New NNN pincer copper complexes as potential anti-prostate cancer agents. *Eur. J. Med. Chem.* 2022, 244, 114859. [CrossRef]
- 149. Machado, J.F.; Marques, F.; Pinheiro, T.; Villa de Brito, M.J.; Scalese, G.; Pérez-Díaz, L.; Otero, L.; António, J.P.M.; Gambino, D.; Morais, T.S. Copper(I)-thiosemicarbazone complexes with dual anticancer and antiparasitic activity. *ChemMedChem* 2023, 18, e202300074. [CrossRef]
- 150. Terra, W.D.S.; Bull, É.S.; Morcelli, S.R.; Moreira, R.R.; Maciel, L.L.F.; Almeida, J.C.A.; Kanashiro, M.M.; Fernandes, C.; Horn, A. Antitumor activity via apoptotic cell death pathway of water soluble copper(II) complexes: Effect of the diamino unit on selectivity against lung cancer NCI-H460 cell line. *BioMetals* 2021, 34, 661–674. [CrossRef]
- 151. Biswas, S.; Wasai, A.; Ghosh, M.; Rizzoli, C.; Roy, A.; Saha, S.; Mandal, S. A mononuclear N,N,N,O donor schiff base Cu(II) complex inhibits bacterial biofilm formation and promotes apoptosis and cell cycle arrest in prostate cancer cells. *J. Inorg. Biochem.* **2023**, 247, 112314. [CrossRef] [PubMed]
- 152. Pósa, V.; Hajdu, B.; Tóth, G.; Dömötör, O.; Kowol, C.R.; Keppler, B.K.; Spengler, G.; Gyurcsik, B.; Enyedy, É.A. The coordination modes of (thio)semicarbazone copper(II) complexes strongly modulate the solution chemical properties and mechanism of anticancer activity. *J. Inorg. Biochem.* **2022**, *231*, 111786. [CrossRef]
- 153. Cruz, N.; Pinho, J.O.; Soveral, G.; Ascensão, L.; Matela, N.; Reis, C.; Gaspar, M.M. A Novel hybrid nanosystem integrating cytotoxic and magnetic properties as a tool to potentiate melanoma therapy. *Nanomaterials* **2020**, *10*, 693. [CrossRef] [PubMed]
- 154. Sk, S.; Majumder, A.; Sow, P.; Samadder, A.; Bera, M. Exploring a new family of designer copper(II) complexes of anthracene-appended polyfunctional organic assembly displaying potential anticancer activity via cytochrome c mediated mitochondrial apoptotic pathway. *J. Inorg. Biochem.* 2023, 243, 112182. [CrossRef]
- 155. Li, S.; Zhao, J.; Yuan, B.; Wang, X.; Zhang, J.; Yue, L.; Hou, H.; Hu, J.; Chen, S. Crystal structure, DNA interaction and in vitro anticancer activity of Cu(II) and Pt(II) compounds based on benzimidazole-quinoline derivative. *Polyhedron* **2020**, *179*, 114369. [CrossRef]
- 156. Bontempo, N.J.S.; Paixão, D.A.; Lima, P.M.A.P.; Barros, D.C.T.; Borges, D.S.; Orsolin, P.C.; Martins, I.C.; Machado, P.H.A.; Lino, R.C.; Souza, T.R.; et al. Copper(II) complex containing 4-Fluorophenoxyacetic acid hydrazide and 1,10-phenanthroline: A prostate cancer cell-selective and low-toxic copper(II) compound. *Molecules* 2022, 27, 7097. [CrossRef]
- 157. Maciel, L.L.F.; de Freitas, W.R.; Bull, E.S.; Fernandes, C.; Horn, A.; de Aquino Almeida, J.C.; Kanashiro, M.M. In vitro and in vivo anti-proliferative activity and ultrastructure investigations of a copper(II) complex toward human lung cancer cell NCI-H460. *J. Inorg. Biochem.* 2020, 210, 111166. [CrossRef]
- 158. Moawed, F.S.; Haroun, R.A.H.; Abou Zaid, E.S.; Mansour, S.Z.; Badawi, A.M.; Kandil, E.I. In vitro and in vivo studies of a newly synthesized copper-cetyl tri-methyl ammonium bromide combined with gallium oxide nanoparticles complex as an antitumor agent against hepatocellular carcinoma. *Int. J. Immunopathol. Pharmacol.* 2023, 37, 3946320231180708. [CrossRef]
- 159. Nonkuntod, P.; Senawong, T.; Soikum, C.; Chaveerach, P.; Watwiangkham, A.; Suthirakun, S.; Chaveerach, U. Copper(II) compounds of 4-nitrobenzohydrazide with different anions (ClO<sub>4</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup> and Br<sup>-</sup>): Synthesis, characterization, DFT calculations, DNA interactions and cytotoxic properties. *Chem. Biodivers.* **2022**, *19*, e202100708. [CrossRef]
- 160. Lopes, J.C.; Botelho, F.V.; Barbosa Silva, M.J.; Silva, S.F.; Polloni, L.; Alves Machado, P.H.; Rodrigues de Souza, T.; Goulart, L.R.; Silva Caldeira, P.P.; Pereira Maia, E.C.; et al. In vitro and in vivo antitumoral activity of a ternary copper (II) complex. *Biophys. Res. Commun.* 2020, 533, 1021–1026. [CrossRef]

Molecules **2025**, 30, 2104 23 of 25

161. Parsa, F.G.; Feizi, M.A.H.; Safaralizadeh, R.; Hosseini-Yazdi, S.A.; Mahdavi, M. Molecular mechanisms of apoptosis induction in K562 and KG1a leukemia cells by a water-soluble copper(II) thiosemicarbazone complex. *J. Biol. Inorg. Chem.* **2020**, 25, 383–394. [CrossRef]

- 162. Camargo, T.P.; Oliveira, J.A.F.; Costa, T.G.; Szpoganicz, B.; Bortoluzzi, A.J.; Marzano, I.M.; Silva-Caldeira, P.P.; Bucciarelli-Rodriguez, M.; Pereira-Maia, E.C.; Castellano, E.E.; et al. New AlIIIZnII and AlIIICuII dinuclear complexes: Phosphatase-like activity and cytotoxicity. *J. Inorg. Biochem.* 2021, 219, 111392. [CrossRef]
- 163. Trofimova, T.P.; Tafeenko, V.A.; Borodkov, A.S.; Proshin, A.N.; Orlova, M.A. New copper complexes with N-(5,6-dihydro-4H-1,3-thiazin-2-yl)benzamide ligand. *Mendeleev. Commun.* **2021**, *31*, 552–554. [CrossRef]
- 164. Rostas, A.M.; Badea, M.; Ruta, L.L.; Farcasanu, I.C.; Maxim, C.; Chifiriuc, M.C.; Popa, M.; Luca, M.; Celan Korosin, N.; Cerc Korosec, R.; et al. Copper(II) complexes with mixed heterocycle ligands as promising antibacterial and antitumor species. *Molecules* 2020, 25, 3777. [CrossRef]
- 165. Pitucha, M.; Korga-Plewko, A.; Czylkowska, A.; Rogalewicz, B.; Drozd, M.; Iwan, M.; Kubik, J.; Humeniuk, E.; Adamczuk, G.; Karczmarzyk, Z.; et al. Influence of complexation of thiosemicarbazone derivatives with Cu (II) ions on their antitumor activity against melanoma cells. *Int. J. Mol. Sci.* 2021, 22, 3104. [CrossRef] [PubMed]
- 166. Climova, A.; Pivovarova, E.; Szczesio, M.; Gobis, K.; Ziembicka, D.; Korga-Plewko, A.; Kubik, J.; Iwan, M.; Antos-Bielska, M.; Krzyżowska, M.; et al. Anticancer and antimicrobial activity of new copper (II) complexes. *J. Inorg. Biochem.* **2023**, 240, 112108. [CrossRef]
- 167. Diz, M.; Durán-Carril, M.L.; Castro, J.; Alvo, S.; Bada, L.; Viña, D.; García-Vázquez, J.A. Antitumor activity of copper(II) complexes with Schiff bases derived from N'-tosylbenzene-1,2-diamine. *J. Inorg. Biochem.* **2022**, 236, 111975. [CrossRef] [PubMed]
- 168. Illán-Cabeza, N.A.; Jiménez-Pulido, S.B.; Hueso-Ureña, F.; Ramírez-Expósito, M.J.; Martínez-Martos, J.M.; Moreno-Carretero, M.N. Relationship between the antiproliferative properties of Cu(II) complexes with the Schiff base derived from pyridine-2-carboxaldehyde and 5,6-diamino-1,3-dimethyluracil and the redox status mediated by antioxidant defense systems on glioma tumoral cells. *J. Inorg. Biochem.* 2020, 207, 111053. [CrossRef]
- 169. González-Ballesteros, M.M.; Sánchez-Sánchez, L.; Espinoza-Guillén, A.; Espinal-Enríquez, J.; Mejía, C.; Hernández-Lemus, E.; Ruiz-Azuara, L. Antitumoral and antimetastatic activity by mixed chelate copper(II) compounds (Casiopeínas®) on triple-negative breast cancer, in vitro and in vivo models. *Int. J. Mol. Sci.* 2024, 25, 8803. [CrossRef]
- 170. Mishra, A.; Djoko, K.Y.; Lee, Y.H.; Lord, R.M.; Kaul, G.; Akhir, A.; Saxena, D.; Chopra, S.; Walton, J.W. Water-soluble copper pyrithione complexes with cytotoxic and antibacterial activity. *Org. Biomol. Chem.* **2023**, *21*, 2539–2544. [CrossRef]
- 171. Balsa, L.M.; Solernó, L.M.; Rodriguez, M.R.; Parajón-Costa, B.S.; Gonzalez-Baró, A.C.; Alonso, D.F.; Garona, J.; León, I.E. Cu(II)-acylhydrazone complex, a potent and selective antitumor agent against human osteosarcoma: Mechanism of action studies over in vitro and in vivo models. *Chem. Biol. Interact.* 2023, 384, 110685. [CrossRef]
- 172. Durigon, D.C.; Glitz, V.A.; Pimenta, B.F.; Guedes, A.M.V.; Silva, J.V.O.; Bella Cruz, C.C.; Andrade, L.M.; Pereira-Maia, E.C.; Mikcha, J.M.G.; Bella Cruz, A.; et al. The influence of thioether-substituted ligands in dicopper(II) complexes: Enhancing oxidation and biological activities. *J. Inorg. Biochem.* 2024, 256, 112573. [CrossRef] [PubMed]
- 173. Łukasiewicz, S.; Czeczelewski, M.; Forma, A.; Baj, J.; Sitarz, R.; Stanisławek, A. Breast cancer-epidemiology, risk factors, classification, prognostic markers, and current treatment strategies-an updated review. *Cancers* 2021, 13, 4287. [CrossRef] [PubMed]
- 174. Pilleron, S.; Gower, H.; Janssen-Heijnen, M.; Signal, V.C.; Gurney, J.K.; Morris, E.J.; Cunningham, R.; Sarfati, D. Patterns of age disparities in colon and lung cancer survival: A systematic narrative literature review. *BMJ Open* **2021**, *11*, e044239. [CrossRef]
- 175. Gallicchio, L.; Daee, D.L.; Rotunno, M.; Barajas, R.; Fagan, S.; Carrick, D.M.; Divi, R.L.; Filipski, K.K.; Freedman, A.N.; Gillanders, E.M.; et al. Epidemiologic research of rare cancers: Trends, resources, and challenges. *Cancer Epidemiol. Biomark. Prev.* 2021, 30, 1305–1311. [CrossRef]
- 176. Swarbrick, A.; Fernandez-Martinez, A.; Perou, C.M. Gene-expression profiling to decipher breast cancer inter- and intratumor heterogeneity. *Cold Spring Harb. Perspect. Med.* **2024**, *14*, a041320. [CrossRef] [PubMed]
- 177. Gonzalez, T.L.; Hancock, M.; Sun, S.; Gersch, C.L.; Larios, J.M.; David, W.; Hu, J.; Hayes, D.F.; Wang, S.; Rae, J.M. Targeted degradation of activating estrogen receptor α ligand-binding domain mutations in human breast cancer. *Breast Cancer Res. Treat.* **2020**, *180*, 611–622. [CrossRef]
- 178. Yu, M.; Sun, Y.; Yang, G.; Wang, Z. An experimental study on [125I]I-pHLIP (Var7) for SPECT/CT imaging of an MDA-MB-231 triple-negative breast cancer mouse model by targeting the tumor microenvironment. *Mol. Imaging* **2021**, 2021, 5565932. [CrossRef]
- 179. Tsuji, K.; Kida, Y.; Koshikawa, N.; Yamamoto, S.; Shinozaki, Y.; Watanabe, T.; Lin, J.; Nagase, H.; Takenaga, K. Suppression of non-small-cell lung cancer A549 tumor growth by an mtDNA mutation-targeting pyrrole-imidazole polyamide-triphenylphosphonium and a senolytic drug. *Cancer Sci.* 2022, 113, 1321–1337. [CrossRef]
- 180. Thai, A.A.; Solomon, B.J.; Sequist, L.V.; Gainor, J.F.; Heist, R.S. Lung cancer. Lancet 2021, 398, 535–554. [CrossRef]

Molecules **2025**, 30, 2104 24 of 25

181. Acevedo-Sánchez, V.; Martínez-Ruiz, R.S.; Aguilar-Ruíz, S.R.; Torres-Aguilar, H.; Chávez-Olmos, P.; Garrido, E.; Baltiérrez-Hoyos, R.; Romero-Tlalolini, M.L.A. Quantitative proteomics for the identification of differentially expressed proteins in the extracellular vesicles of cervical cancer cells. *Viruses* **2023**, *15*, 702. [CrossRef] [PubMed]

- 182. Zhao, C. Cell culture: In vitro model system and a promising path to in vivo applications. *J. Histotechnol.* **2023**, *46*, 1–4. [CrossRef] [PubMed]
- 183. Cheng, S.; Li, L.; Yu, X. Comprehension characterization of prostate cancer cell lines identified JAK-STAT3 signaling in lineage-switched non-neuroendocrine cells. *J. Clin. Oncol.* **2023**, *41* (Suppl. 16), e17005. [CrossRef]
- 184. Zahed, H.; Feng, X.; Sheikh, M.; Bray, F.; Ferlay, J.; Ginsburg, O.; Shiels, M.S.; Robbins, H.A. Age at diagnosis for lung, colon, breast and prostate cancers: An international comparative study. *Int. J. Cancer* **2024**, *154*, 28–40. [CrossRef]
- 185. Moreira-Silva, F.; Henrique, R.; Jerónimo, C. From therapy resistance to targeted therapies in prostate cancer. *Front. Oncol.* **2022**, 12, 877379. [CrossRef]
- 186. Rahman, M.M.; Opo, F.A.D.M.; Asiri, A.M. Cytotoxicity study of cadmium-selenium quantum dots (Cdse QDs) for destroying the human HepG2 liver cancer cell. *J. Biomed. Nanotechnol.* **2021**, *17*, 2153–2164. [CrossRef]
- 187. Lin, L.; Yan, L.; Liu, Y.; Qu, C.; Ni, J.; Li, H. The burden and trends of primary liver cancer caused by specific etiologies from 1990 to 2017 at the global, regional, national, age, and sex level results from the global burden of disease Study 2017. *Liver Cancer* 2020, 9, 563–582. [CrossRef]
- 188. De Souza, Í.P.; de Melo, A.C.C.; Rodrigues, B.L.; Bortoluzzi, A.; Poole, S.; Molphy, Z.; McKee, V.; Kellett, A.; Fazzi, R.B.; da Costa Ferreira, A.M.; et al. Antitumor copper(II) complexes with hydroxyanthraquinones and N,N-heterocyclic ligands. *J. Inorg. Biochem.* 2023, 241, 112121. [CrossRef]
- 189. Zehra, S.; Tabassum, S.; Arjmand, F. Biochemical pathways of copper complexes: Progress over the past 5 years. *Drug Discov. Today* **2021**, *26*, 1086–1096. [CrossRef]
- 190. Dias, M.P.; Moser, S.C.; Ganesan, S.; Jonkers, J. Understanding and overcoming resistance to PARP inhibitors in cancer therapy. *Nat. Rev. Clin. Oncol.* **2021**, *18*, 773–791. [CrossRef]
- 191. He, Y.; Sun, M.M.; Zhang, G.G.; Yang, J.; Chen, K.S.; Xu, W.W.; Li, B. Targeting PI3K/Akt signal transduction for cancer therapy. Signal Transduct. Target. Ther. 2021, 6, 425. [CrossRef] [PubMed]
- 192. Whitaker, R.H.; Cook, J.G. Stress relief techniques: p38 MAPK determines the balance of cell cycle and apoptosis pathways. *Biomolecules* **2021**, *11*, 1444. [CrossRef]
- 193. Bhangu, J.S.; Macher-Beer, A.; Schimek, V.; Garmroudi, B.; Tamandl, D.; Unger, L.W.; Bachleitner-Hofmann, T.; Oehler, R. Circulating caspase-cleaved cytokeratin 18 correlates with tumour burden and response to therapy in patients with colorectal cancer liver metastasis. *Clin. Chim. Acta* 2023, 538, 53–59. [CrossRef] [PubMed]
- 194. Liu, Y.; Munsayac, A.; Hall, I.; Keane, S.C. Solution structure of NPSL2, A regulatory element in the oncomiR-1 RNA. *J. Mol. Biol.* **2022**, 434, 167688. [CrossRef]
- 195. Petronijević, J.; Joksimović, N.; Milović, E.; Crnogorac, M.Đ.; Petrović, N.; Stanojković, T.; Milivojević, D.; Janković, N. Antitumor activity, DNA and BSA interactions of novel copper(II) complexes with 3,4-dihydro-2(1H)-quinoxalinones. *Chem. Biol. Interact.* **2021**, 348, 109647. [CrossRef]
- 196. Hossan, M.S.; Break, M.K.B.; Bradshaw, T.D.; Collins, H.M.; Wiart, C.; Khoo, T.J.; Alafnan, A. Novel semi-synthetic Cu (II)-cardamonin complex exerts potent anticancer activity against triple-negative breast and pancreatic cancer cells via inhibition of the Akt signaling pathway. *Molecules* **2021**, *26*, 2166. [CrossRef] [PubMed]
- 197. Fouad, R.; Adly, O.M.I. Novel Cu<sup>2+</sup> and Zn<sup>2+</sup> nanocomplexes drug based on hydrazone ligand bearings chromone and triazine moieties: Structural, spectral, DFT, molecular docking and cytotoxic studies. *J. Mol. Struct.* **2021**, 1225, 129158. [CrossRef]
- 198. Shi, F.; Qiao, H. Preparations, properties and applications of gallium oxide nanomaterials—A review. *Nano Sel.* **2022**, *3*, 348–373. [CrossRef]
- 199. He, Y.; Yang, M.; Yang, L.; Hao, M.; Wang, F.; Li, X.; Taylor, E.W.; Zhang, X.; Zhang, J. Preparation and anticancer actions of CuET-nanoparticles dispersed by bovine serum albumin. *Colloids Surf. B Biointerfaces* **2023**, 226, 113329. [CrossRef]
- 200. Mandour, A.A.; Nassar, I.F.; Abdel Aal, M.T.; Shahin, M.A.E.; El-Sayed, W.A.; Hegazy, M.; Yehia, A.M.; Ismail, A.; Hagras, M.; Elkaeed, E.B.; et al. Synthesis, biological evaluation, and in silico studies of new CDK2 inhibitors based on pyrazolo pyrimidine and pyrazolo [4,3-e][1,2,4]triazolo [1,5-c]pyrimidine scaffold with apoptotic activity. *J. Enzym. Inhib. Med. Chem.* 2022, 37, 1957–1973. [CrossRef]
- 201. Dasari, S.; Njiki, S.; Mbemi, A.; Yedjou, C.G.; Tchounwou, P.B. Pharmacological effects of cisplatin combination with natural products in cancer chemotherapy. *Int. J. Mol. Sci.* **2022**, *23*, 1532. [CrossRef] [PubMed]
- 202. Romani, A.M.P. Cisplatin in cancer treatment. Biochem. Pharmacol. 2022, 206, 115323. [CrossRef] [PubMed]
- 203. Kapustina, A.; Tupolova, Y.; Popov, L.; Vlasenko, V.; Gishko, K.; Berejnaya, A.; Shcherbatykh, A.; Golubeva, Y.; Klyushova, L.; Lider, E.; et al. Copper(II) coordination compounds based on bis-hydrazones of 2,6-diacetylpyridine: Synthesis, structure, and cytotoxic activity. *Dalton Trans.* 2024, 53, 3330–3347. [CrossRef] [PubMed]

Molecules **2025**, 30, 2104 25 of 25

204. Lu, Y.; Pan, Q.; Gao, W.; Pu, Y.; He, B. Reversal of cisplatin chemotherapy resistance by glutathione-resistant copper-based nanomedicine via cuproptosis. *J. Mater. Chem. B* **2022**, *10*, 6296–6630. [CrossRef]

205. Li, A.; Huang, K.; Pan, W.; Wu, Y.; Liang, Y.; Zhang, Z.; Wu, D.; Ma, L.; Gou, Y. Thiosemicarbazone Mixed-Valence Cu(I/II) Complex against Lung Adenocarcinoma Cells through Multiple Pathways Involving Cuproptosis. *J. Med. Chem.* 2024, 67, 9091–9103. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.