**Supplemental table 1.** Demographic data of donors, clinical information and assays applied to each tissue/donor are listed.

ID	Location	Age	Gender	Autoimmune Manifestations	Used for
Control1(E)	Axillary	71	M	-	GeoMx, Panel1,
					Panel2, Panel3,
					Panel4, Panel5
Control2(F)	Cervical	39	M	-	Panel1, Panel2,
					Panel3, Panel4
Control3(G)	Inguinal	49	M	-	GeoMx, Panel1,
					Panel2, Panel3,
					Panel4, Panel5
Control4(U)	Inguinal	78	M	-	GeoMx, Panel1,
					Panel3, Panel4,
					Panel5
SLE1(A)	Inguinal	35	M	ANA (+), anti-SSA-52 (+), anti-	GeoMx, Panel1,
				SSA-60(+), C4(low), C3(low),	Panel2, Panel3,
				isolated hemolysis, IgG=30.1 g/l	Panel4, Panel5
SLE2(B)	Axillary	56	F	ANA (+), anti-nucleosome	Panel1, Panel2,
				(borderline), SLE GN class V,	Panel3, Panel4
				IgG=10.7 g/l	
SLE3(C)	Inguinal	21	M	ANA (+), anti-SSA-60 (+), anti-	GeoMx, Panel1,
				RNP (+), anti-Sm (+), anti-	Panel2, Panel3,
				dsDNA (+), anti-nucleosome	Panel4, Panel5
				(+), C4(low), C3(low),	
				leukopenia-anemia, IgG=47.5 g/l	
SLE4(D)	Inguinal	25	F	ANA (+), anti-SSA-52	GeoMx, Panel1,
				(borderline), (+), anti-RNP (+),	Panel2, Panel3,
				anti-Sm (+), anti- dsDNA (+),	Panel4, Panel5
				anti-nucleosome (+), C4(low),	
				C3(low), leukopenia, IgG=23.1	
				g/l	

SLE5(I)	Cervical	21	F	ANA (+), anti-SSA-52 (+), anti-	GeoMx, Panel1,
				SSA-60(+), anti- dsDNA (+),	Panel3, Panel4,
				anti-nucleosome (+), C4(low),	Panel5
				C3(low), leukopenia-anemia,	
				SLE GN class IV, IgG=14.78 g/l	

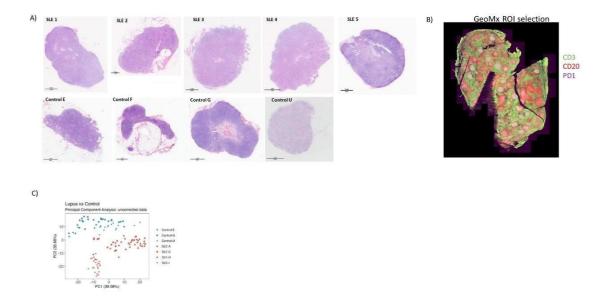
**Supplemental table 2.** The antibody clones, catalogue numbers and fluorochrome used are shown.

Epitope	Clone	Cat No	Fluorophore
CD4	SP35	790-4423	Opal690
PD-1	NAT 105	ACI 3137	Opal620
Ki67	MIB-1	M7240	Opal780(1), Opal520(2)
CD20	L26	NCL-L-CD20-L26	Opal520
GATA-3	L50-823	760-4897	Opal570
Bcl-6	GI191E/A8	760-4241	Opal480
CD8	C8/144B	M710301-2	Opal780
PAX5	SP34	790-4420	Opal690
GRZb	GrB-7	MON7029C	Opal480
Perforin	5B10	Mob555	Opal570
CD57	NK-1	Mob 163	Opal480
CD14	EPR3653	114R-15	Opal520
CD16	2H7	CD16-L-CE	Opal570
CD11c	EP1347Y	ab52632	Opal780(4), Alexa555(5)
CD19	BT51E	NCL-L-CD19-163	Opal690

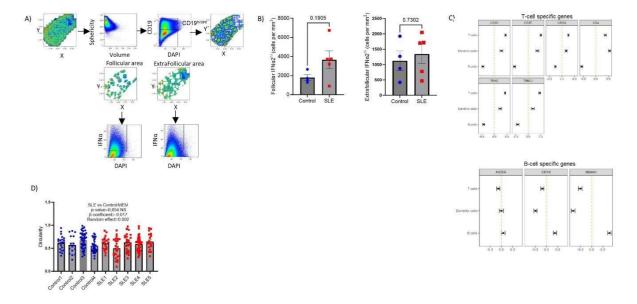
Tbet	MRQ-46	760-4598	Opal480
IFN-α2	Polyclonal	ab198914	Opal620
FDC	CNA.42	14-99-68-82	Alexa488
CXCL-13	Polyclonal	PA5-28827	Opal780
IL-21	Polyclonal	AHP1845	BV421

## Supplemental table 3. The antibody panels and dilutions used are shown.

Panel1	Panel2	Panel3	Panel4	Panel5
Bcl-6 (Ready to use)	CD8 (1/100)	CD4 (Ready to use)	CD11c (1/200)	CD20 (1/400)
CD4 (Ready to use)	PD-1 (1/100)	PD-1 (1/100)	CD19 (1/200)	FDC (1/350)
PD-1 (1/100)	Perforin (1/10)	Ki67 (1/300)	Tbet (Ready to use)	IL-21 (1/80)
Ki67 (1/300)	GRZb (1/40)	CD20 (1/400)	IFN-α2 (1/200)	CXCL-13 (1/300)
CD20 (1/400)	PAX-5 (Ready to use)	GATA-3 (1/10)	CD14 (1/100)	CD11c (1/20)
GATA-3 (1/10)	Ki67 (1/300)	CD57 (1/200)	CD16 (1/100)	
DAPI	DAPI	DAPI	DAPI	SYTO45

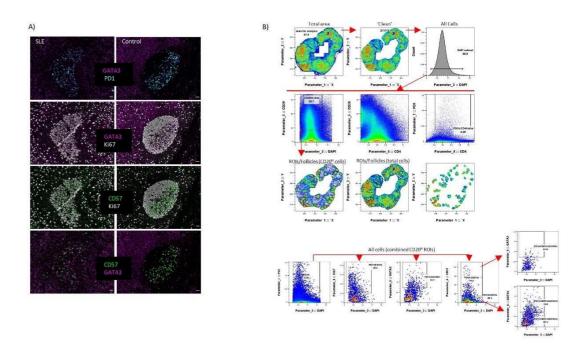


**Supplemental Figure 1.** (**A**) Hematoxylin and eosin (H&E) staining for all the LN tissues used in this study (scale bar: 1.5 mm). (**B**) Representative staining of one LN tissue using antibodies against CD3, CD20 and PD1 before GeoMx ROI selection.CD20-dense areas populated by CD3<sup>hi</sup>PD1<sup>hi</sup> cells were selected as ROIs (secondary mature follicles). (**C**) The PCA plot shows the distribution of spatial transcriptomic uncorrected data from 111 Regions of Interest (ROIs, follicles) collected from SLE (N=64, 4 donors) and control LNs (N=47, 3 donors). Each point represents an individual ROI, with the colour indicating the different cohort (Red=SLE, Blue=Control).

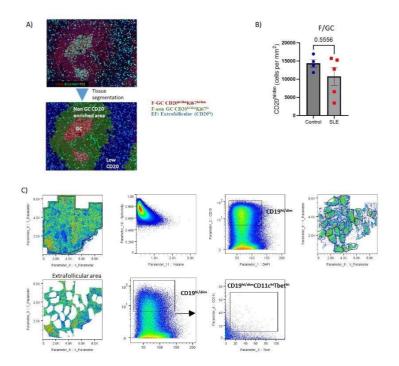


**Supplemental Figure 2. (A)** Histo-cytometry gating scheme used for the quantification of IFN $\alpha$ 2<sup>hi</sup> cells. F and EF areas were manually identified based on the density of the CD19 signal

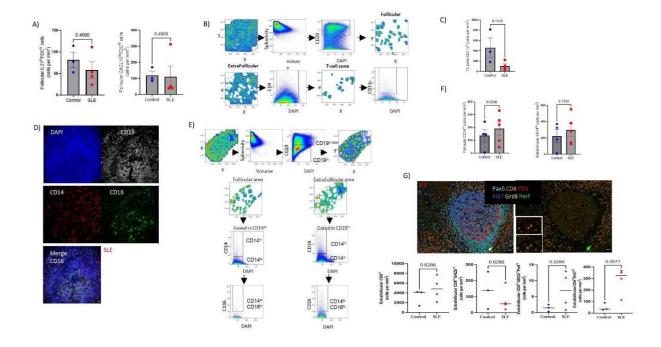
and gated back to the X, Y dot plot. Representative data from a SLE LN are plotted. (**B**) Bar graphs demonstrating the cell densities of extrafollicular (right) and follicular (left) IFN $\alpha$ 2<sup>hi</sup> cells in SLE (N = 5) and control LNs (N = 4). Each dot/square represents one donor. The p values were calculated using the Mann–Whitney test. Data represent mean  $\pm$  SEM. (**C**) Graphs representing the relative correlation of deconvoluted cell subsets proportion (T, B, dendritic cells) from all the ROIs with T (upper) and B (lower) cell related genes. (**D**) Bar graphs demonstrating the quantification of Circularity of follicles (identified as CD19<sup>hi/dim</sup> areas. Area and solidity of ROIs were calculated using FIJI. Each dot/square represents a different follicle. The p values were calculated using the mixed effects model (MEM). Data represent mean  $\pm$  SEM.



**Supplemental Figure 3.** (**A**) Representative mIF images of GATA3 (magenta), CD57 (green), Ki67 (grey) and PD1 (cyan) from SLE and control LNs (scale bar:20mm). (**B**) Histo-cytometry gating scheme used for the quantification of T<sub>FH</sub> cell subsets. F and EF areas were manually identified based on the density of the CD20 signal and gated back to the X, Y dot plot. Representative data from a control LN are plotted.



**Supplemental Figure 4.** (**A**) Representative example of tissue segmentation into different anatomical categories (F/GC-CD20<sup>hi/dim</sup>Ki67<sup>hi/dim</sup>- F/ non-GC-CD20<sup>hi/dim</sup>Ki67<sup>lo</sup>-EF CD20<sup>lo</sup>) using the inForm image analysis software, version 2.4.8 (Akoya). (**B**) Bar graphs demonstrating the cell densities of F/GC- CD20<sup>hi/dim</sup> B cells in SLE (N = 5) and control LNs (N = 4). Each dot/square represents one donor. The p values were calculated using the Mann–Whitney test. Data represent mean  $\pm$  SEM. (**C**) Histo-cytometry gating scheme used for the quantification of CD19<sup>hi/dim</sup>CD11c<sup>hi</sup>Tbet<sup>hi</sup> cells. F and EF areas were manually identified based on the density of the CD19 signal and gated back to the X, Y dot plot. Representative data from a SLE LN are plotted.



Supplemental Figure 5. (A) Bar graphs demonstrating the cell densities of follicular IL21<sup>hi</sup>FDC<sup>hi</sup> and CXCL13<sup>hi</sup>FDC<sup>hi</sup> cells in SLE (N = 4) and control LNs (N = 3). Each dot/square represents one donor. The p values were calculated using the Mann-Whitney test. Data represent mean ± SEM. (B) Histo-cytometry gating scheme used for the quantification of Tcell zone CD11chi cells. Real cells were gated based on their sphericity and volume. T-cell zone area was manually identified based on the density of the CD4 signal and gated back to the X, Y dot plot. Representative data from a control LN are plotted. (C) Bar graph demonstrating the cell densities of T-cell zone CD11chi cells in SLE (N = 4) and control LNs (N = 3). Each dot/square represents one donor. The p values were calculated using the Mann-Whitney test. Data represent mean  $\pm$  SEM. (**D**) Representative mIF images of CD14 (red), CD16 (green) CD19 (grey) and DAPI (blue) from a SLE donor (40X, scale bar:30 µm). (E) Histo-cytometry gating scheme used for the quantification of CD14<sup>hi</sup>, CD14<sup>lo</sup>CD16<sup>hi</sup> cell subsets. F and EF areas were manually identified based on the density of the CD19 signal and gated back to the X, Y dot plot. Representative data from a SLE LN are plotted. (F) Bar graphs demonstrating the cell densities of follicular (left) and extrafollicular (right) CD14<sup>hi</sup> cells in SLE (N = 5) and control LNs (N = 4). Each dot/square represents one donor. The p values were calculated using the Mann–Whitney test. Data represent mean  $\pm$  SEM (G) Representative examples of PD1 (red), CD8 (orange), GRZb (yellow), Perforin(green), Ki67 (blue) and Pax5 (cyan) staining patterns from a SLE LN (upper panel, scale bar:20mm). Bar graphs demonstrating the cell densities of extrafollicular CD8hi, CD8hi GRZbhi, CD8hi GRZbhiPerfhi, CD8hi Ki67hi cells in SLE (N = 4)

and control LNs (N = 3). Each dot/square represents one donor. The p values were calculated using the Mann–Whitney test. Data represent mean  $\pm$  SEM.