

Extracellular Vesicles as Natural Delivery Carriers Regulate Oxidative Stress Under Pathological Conditions

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Extracellular vesicles are cellular secretory particles that can be used as natural drug delivery carriers. They have successfully delivered drugs including chemotherapeutics, proteins, and genes to treat various diseases. Oxidative stress is an abnormal physiological phenomenon, and it is associated with nearly all diseases. In this short review, we summarize the regulation of EVs on oxidative stress. There are direct effects and indirect effects on the regulation of oxidative stress through EVs. On the one hand, they can deliver antioxidant substances or oxides to recipient cells, directly relieving or aggravating oxidative stress. On the other hand, regulate factors of oxidative stress-related signaling pathways can be delivered to recipient cells by the mediation of EVs, realizing the indirect regulation of oxidative stress. To the best of our knowledge, however, only endogenous drugs have been delivered by EVs to regulate oxidative stress. Therefore, this short review aims to draw more attention to the EVs-based regulation of oxidative stress, and we hope excellent EVs-based delivery carriers that can deliver exogenous drugs to regulate oxidative stress can be exploited.

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INTRODUCTION

Extracellular vesicles (EVs) are phospholipid bilayer-encapsulated vesicles that can be secreted by nearly all cell types (Andaloussi et al., 2013). They can be found in cell culture supernatants as well as biological fluids such as saliva (Winck et al., 2015), blood (Arraud et al., 2014), milk (Zonneveld et al., 2014), cerebrospinal fluids (Akers et al., 2016), and malignant ascites (Li et al., 2019). According to their biogenesis, EVs can be generally divided into three distinct categories: exosomes, microvesicles, and apoptotic bodies (Lässer et al., 2018). Exosomes originate from multivesicular bodies (MVBs) formed by the invagination of the limiting membrane of endosomes (Théry et al., 2002). MVBs can fuse with the cellular membrane to release exosomes into the extracellular space. Different from exosomes, microvesicles and apoptotic bodies are both generated from the cell membrane (Akers et al., 2013). Microvesicles derive from direct outward budding of the normal cells membrane, while apoptotic bodies are only secreted by dying cells during their fragmentation. Besides the origination, size is commonly used as a criterion to distinguish between the three types of EVs (Varga et al., 2014). In general, the size of exosomes, microvesicles, and apoptotic bodies is respectively 30–150 nm, 50–1,000 nm, and 50 nm-2 µm. It should be noted that the size range of the three types of EVs is



overlapped due to their severe heterogeneity (Lőrincz et al., 2015). Furthermore, the heterogeneity of EVs may result in the absence of marker proteins belonging to a certain population. The distinguish of different types of EVs, therefore, cannot rely on a single standard, and it needs to comprehensively consider multiple criterions such as size, marker proteins or genes, and specific lipids (Théry et al., 2018). EVs have been recognized as a kind of messengers of intercellular communication during the past several decades. They can transport cargoes including DNA, RNA, proteins, and lipids between neighboring or distant cells (Tkach and Théry 2016). Taking advantage of their cargoes, EVs can perform numerous physiological functions. As an example, EVs can regulate immune responses to participate in the occurrence and development of many diseases such as cancers, cardiovascular diseases, osteoporosis, and central nervous system diseases (Robbins and Morelli, 2014; Yin et al., 2017).

From a material science point of view, EVs are natural drug delivery carriers. Their phospholipid bilayers similar in structure to liposomes endow EVs with the ability of hydrophilic and hydrophobic drugs loading (van der Meel et al., 2014). And the loaded drugs can be effectively protected from degradation. Compared with liposomes, EVs have intrinsic stability owing to their negatively charged surface and the support of skeletal proteins (Frank et al., 2018). Importantly, EVs have many unique properties superior to synthetic drug delivery carriers. They have low immunogenicity and cytotoxicity due to their endogenous source (Elsharkasy et al., 2020). And their lipid composition and protein content endow them with inherent active targeting properties and the capacity to cross multiple biological barriers such as the blood-brain barrier and mucosal barrier (Ashrafian et al., 2019; Matsumoto et al., 2017). Besides endogenous compositions, exogenous drugs can be loaded in EVs and delivered into target cells (Vader et al., 2016). In our previous

researches, we have exploited a series of blood exosome-based delivery systems to efficiently deliver chemotherapeutics for tumor therapy (Qi et al., 2017; Qi et al., 2016; Qi et al., 2018; Yang et al., 2019; Zhan et al., 2020). As shown in Figure 1, we take advantage of the structure and properties of the blood exosome membrane to load nucleic acid drugs and chemotherapeutics. The system with the tumor-targeting ability and endosomeescaping capacity realizes the combined antitumor therapy. In general, EVs have been used as carriers to deliver endogenous or exogenous drugs for disease treatment. Oxidative stress is an abnormal physiological phenomenon, and it is defined as "an imbalance between antioxidants and oxidants, resulting in the disruption of redox signaling and the corresponding molecular damage." To the best of our knowledge, it is associated with nearly all diseases such as cancers (Hayes et al., 2020), cardiovascular diseases (Dubois-Deruy et al., 2020), aortic dissection (Zong et al., 2021), atherosclerosis (Li et al., 2021; Xue et al., 2021), COVID-19 (Li et al., 2021), drug-induced injuries (Li et al., 2021), and diabetes (Zhang et al., 2020). The effective regulation or relief of oxidative stress can potentially prevent many diseases, and EVs play an important role in this process. In this mini-review, we will mainly summarize how oxidative stress under pathological conditions is modulated by EV-mediated drug delivery, and also discuss the influence of oxidative stress on EVs.

EVS MODULATE OXIDATIVE STRESS

As we have mentioned above, EVs derived from different cells have heterogeneity. They may have different sizes, shapes, and compositions. Therefore, EVs may exhibit diverse regulations of oxidative stress, and the modulation mechanism is shown in



Figure 2. On the one hand, they can deliver reactive oxygen species (ROS) scavengers to relieve oxidative stress. On the other hand, they may produce ROS by their detrimental contents. Below we will discuss these two aspects in great detail.

EVs Relieve Oxidative Stress

Under normal physiological conditions, the production and elimination of ROS are balanced, while oxidative stress is produced when this balance is disturbed (Betteridge, 2000). In eukaryotic cells, antioxidant enzymes are commonly used as ROS scavengers to maintain the balance. EVs could inherently carry antioxidant enzymes, and numerous proteomic analyses have proved that superoxide dismutases (SOD), catalase (CAT), peroxiredoxin (PRDX), glutathione peroxidase (GPX), glutathione S-transferase (GST), and thioredoxin (TRX), et al. can be secreted into EVs (Bodega et al., 2019). These antioxidant enzymes loaded in EVs can directly relieve oxidative stress under various pathological conditions. For example, EVs released from T lymphocytes can deliver antioxidant enzymes, such as SOD isoforms and CAT, to human umbilical vein endothelial cells (HUVECs) and scavenge ROS (Soleti et al., 2012). Compared with other cells, stem cells are one of the main sources of antioxidant enzyme-loaded EVs. EVs derived from human mesenchymal stem cells (hMSCs) can protect hippocampal neurons from oxidative stress by the mediation of CAT (Bodart-Santos et al., 2019). hMSCs can also secret manganese-SOD (Mn-SOD) into EVs that can reduce oxidative stress in the hepatic ischemia-reperfusion injury (Yao et al., 2018). In addition to antioxidant enzymes, other proteins can also be riched in EVs to relieve oxidative stress. EVs secreted by astroglial cells can transport apolipoprotein D to neurons and reduce oxidative stress damage (Pascua-Maestro et al., 2019). Besides, endothelial EVs carry functional endothelial nitric oxide synthase (eNOS) to regulate eNOS/Akt signaling pathway, protecting against oxidative stress in endothelial cells (Mahmoud et al., 2017). These antioxidant substance-loaded EVs are more like ROS scavengers and they can directly eliminate both intracellular and extracellular ROS to relieve oxidative stress.

Besides the above direct effects, indirect effects on ROS or oxidative stress can be carried out by EVs. They can deliver drugs to target cells and regulate oxidative stress-associated signaling pathways. For example, EVs secreted by H293T cells can deliver microRNA-126 to H2O2-treated neonatal rat ventricular cardiomyocytes, attenuating the ROS elevation and reducing the apoptosis by targeting ERRFI1 (Wang et al., 2019). By comparison, stem cell-derived EVs possess a greater regulatory capacity owing to their versatility. EVs derived from hMSCs can mediate the nuclear factor erythroid 2-related factor-2 (Nrf2) signaling pathway to reduce ROS generation in H₂O₂-stimulated keratinocytes or UV-irradiated mice skin (Wang et al., 2020). They can also resist cisplatin-induced oxidative stress by increasing GSH levels and suppressing activation of the p38MAPK pathway (Zhou et al., 2013). Adipose-derived mesenchymal stem cells (ADMSCs)-derived EVs can alleviate LPS induced accumulation of ROS and inflammatory cytokines in macrophages via regulating the Nrf2/heme oxygenase-1 (HO-1) signaling pathway (Shen et al., 2021). Besides, they can reduce ischemia/reperfusion injury as well as lung injury by suppressing oxidative stress (Liu et al., 2019; Gao et al., 2021). EVs derived from human placental mesenchymal stem cells (hP-MSCs) can eliminate ROS and reduce the activity of myeloperoxidase (MPO) effectively suppressing oxidative stress (Duan et al., 2020). Human Wharton Jelly mesenchymal stem cells (hWJMSCs)released EVs can protect the kidney against ischemia/ reperfusion injury by suppressing nicotinamide adenine dinucleotide phosphate oxidases 2 (NOX2) expression or enhancing Nrf2/ARE activation and the subsequent alleviation of the oxidative stress (Zhang et al., 2014; Zhang et al., 2016a). These indirect regulation effects may be more lasting. That is because antioxidant substances loaded in EVs for the direct relief of oxidative stress are readily destroyed.

Compound types of EVs such as EVs separated from body fluids also show the capacity of oxidative stress relief. Human milk-derived EVs protect intestinal stem cells (ISCs) from oxidative stress injury mediated *via* the Wnt/ β -catenin signaling pathway (Dong et al., 2020). Blood-derived EVs from healthy individuals can restore the homeostasis of oxidative stress in the mice model of Parkinson's disease possessing neuroprotective effects (Sun et al., 2020). Camel milk EVs and their related genes ameliorate oxidative stress, normalize the antioxidant status, regulate the inflammatory pattern and improve the immune response in the cyclophosphamide (CTX)-treated animals (Ibrahim et al., 2019). The regulation mechanisms of compound types of EVs are more complicated because different types of EVs may have distinct functions. Furthermore, the ratio of different types of EVs is diverse at each separation resulting in the unstable regulation efficiency of compound types of EVs.

EVs Aggravate Oxidative Stress

In addition to being therapeutics, EVs may also be accomplices of oxidative stress. They can directly load ROS or other oxides and deliver them to recipient cells. EVs derived from hypoxia/ reoxygenation-treated HUVECs can deliver ROS into H9C2 cardiomyocytes resulting in ROS overload and subsequent oxidative stress (Zhang et al., 2016b). However, this direct effect mediated by EVs-loaded ROS is rarely reported owing to the extremely short life span of ROS. Similar to indirect relief of oxidative stress, the indirect aggravation of oxidative stress depends on the contents of EVs. On the one hand, EVs may directly produce ROS by the loaded enzymes to aggravate oxidative stress. As an example, endothelial EVs containing NOX subunits produce ROS to involve in endothelial damage (Burger et al., 2016). On the other hand, EVs can regulate target cells to indirectly aggravate oxidative stress. Gene drugs loaded in EVs have multi-target control ability, and they may induce numerous target genes to involve in oxidative stress. For example, EVs from ionizing irradiated mouse embryo can fibroblasts deliver microRNA-34c (MEFs) into unirradiated cells, potentially triggering a cascade of gene expression alterations to increase ROS and inducing bystander oxidative stress (Rastogi et al., 2018). And this radiation-induced bystander effect is also present in the body. EVs originated from mice irradiated with 2-Gy X-rays are intravenously injected into bystander mice, and the circulating reactive oxygen metabolite (ROM) level is increased (Hargitai et al., 2021). Ketamineinjured human uroepithelial cells (SV-HUC-1) cells can secrete EVs containing specific miRNAs. They enhance oxidative stress by mediating P38/NF-KB pathway (Xi et al., 2020). Besides genes, proteins or other small molecules can also be delivered by EVs. Polycyclic aromatic hydrocarbons-treated hepatocytes release EVs that are enriched in proteins, such as NOX and ferritin, as well as iron (van Meteren et al., 2020). These EVs could induce oxidative stress in recipient cells, thereby resulting in apoptosis.

It should be noted EVs that can aggravate oxidative stress are mainly derived from stimulated cells. The stimulating factors usually cause changes in the redox state of parent cells, and cells would secrete EVs to maintain their redox homeostasis. These EVs contain genes, proteins, and even lipids that can activate or regulate signaling pathways of oxidative stress aggravation. Therefore, these EVs can be thought of as products of the stress response of cells.

This stress response also occurs in the body. EVs derived from body fluids such as blood potentially aggravate oxidative stress. Platelet-derived EVs are the main type of EVs in the blood, and they are readily instigated to increase oxidative stress. In severe sepsis patients, platelet-derived EVs can produce ROS by the mediation of NOX and subsequently induce vascular cell apoptosis (Janiszewski et al., 2004). However, it is also reported that EVs derived from septic shock patients show protective effects on vascular function (Mostefai et al., 2008). This difference is potentially due to different sources of EVs. Besides, EVs from mice with the acetaminophen-induced liver injury can cause the allogeneic mice plasma ROS elevation inducing in vivo toxicity (Cho et al., 2018). Body fluids of different individuals are significant heterogeneity, and corresponding EVs may differ in origin, composition, and structure. Different from EVs from a specific type of cells, therefore, EVs separated from body fluids may show diverse functions.

OXIDATIVE STRESS MODULATE EVS

As drug delivery carriers, EVs can modulate oxidative stress by their contents. While the contents of EVs are readily changed under different pathological conditions. Cells with or without the treatment of oxidative stress secrete EVs with heterogeneity, and these diverse EVs may possess positive or negative effects on their adjacent or distant cells and tissues.

Oxidative Stress Educates EVs

Under oxidative stress, cells may secrete EVs with positive effects. Firstly, oxidative stress-educated EVs may have the anti-oxidative ability. These EVs can relieve the cell's oxidative stress as well as the oxidative stress of other cells or tissues. EVs derived from the non-pigmented ciliary epithelium (NPCE) which are exposed to oxidative stress can deliver protecting messages to activate major antioxidant genes and enhance SOD and CAT activity, protecting the trabecular meshwork (TM). Mouse mast cells exposed to oxidative stress release EVs, and these EVs transport mRNA to recipient cells providing them with resistance against oxidative stress (Tailleux et al., 2010). Besides relieving oxidative stress, other beneficial physiological effects can also be realized through oxidative stress-educated EVs. Retinal pigment epithelium (RPE) cells release higher amounts of EVs when they are exposed to oxidative stress. These EVs have a higher expression of vascular endothelial growth factor receptors (VEGFRs) in the surface membrane and have VEGFR mRNA in their internal cavity, promoting angiogenesis in endothelial cells (Atienzar-Aroca et al., 2016). Besides, oxidative stress-educated EVs can also involve in the muscle regeneration process. EVs released from oxidatively challenged myotubes have an increased loading of nucleic acid. They can decrease the diameter of the myotube, the mRNA levels of myogenin, and the expression of myosin heavy chain, leading to the proliferation of recipient myoblast (Guescini et al., 2017). EVs derived from stem cells often show beneficial effects, and oxidative stress may enhance these effects. The angiogenic and antimicrobial protein content of EVs derived

from H_2O_2 -treated ADMSCs was altered (Mayo et al., 2019). These EVs can enhance the viability of flaps and increase capillary density, improving tissue recovery. Taken together, these oxidative stress-educated EVs can be used as compensatory measures to minimize damage caused by oxidative stress.

Oxidative Stress Miseducates EVs

Whether *in vitro* or *in vivo*, the cells can interact with each other, and EVs are important media of the interaction. Under pathological conditions, oxidative stress cause damage to cells, while these cells would expand the damage to other cells by their miseducated EVs. To date, immune cells are the main victims of oxidative stress-miseducated EVs, and immunosuppressive EVs are often released under oxidative stress. It has been reported that Jurkat and Raji cells release EVs containing NKG2D relative mRNA and proteins under thermal or oxidative stress. These EVs can reduce the cytotoxicity mediated by natural killer cell group 2D (NKG2D) receptor-ligand system and thus impair the functions of natural killer (NK) cells (Zimmer et al., 2011). Besides the specific type of cells, the body fluids of patients also contain immunosuppressive EVs. EVs separated from the blood of patients with rheumatoid arthritis contain oxidized phospholipids. These EVs can stimulate cells expressing Tolllike receptor 4 (TLR4), involving in inflammatory diseases (Manček-Keber et al., 2015). In addition, EVs have an important role in feto-maternal communication. Under oxidative stress, fetal cells-derived EVs can deliver unique cargos to parturition-associated uterine cells, inducing inflammation (Shahin et al., 2021). The effects of oxidative stress-miseducated EVs on the immune system need to be taken seriously since the immune system involves in nearly all pathological diseases. Furthermore, how to effectively avoid the effects is also worth study.

FUTURE AND PROSPECT

Genes, proteins, and even lipids loaded in EVs can all be used as regulatory factors. In recent researches, however, EVs used to regulate oxidative stress are undecorated products, and it means that regulatory factors are endogenous components of EVs. These crude EVs have low regulation efficiency of oxidative stress. That is because EVs have heterogeneity, and parts of EVs may not have the regulation ability because of their lack of endogenous regulatory factors. Furthermore, it is complicated to adjust endogenous components of EVs for the regulation of oxidative stress. To improve the regulation efficiency, therefore, exogenous drugs possessing the efficient

REFERENCES

regulation ability of oxidative stress should be loaded into EVs. Numerous methods, such as electroporation and surfactant incubation, have been applied to load exogenous drugs into EVs. But it is still difficult to efficiently load macromolecular drugs into EVs, and there is a great need to exploit new drug loading methods. Moreover, it should be noted that EVs should not be oxidated during the drug loading process because oxidative stress potentially miseducates these EVs rendering opposite functions.

The sources of EVs applied for oxidative stress regulation should be carefully selected. As we have mentioned above, once parent cells are subjected to oxidative stress, EVs are potentially miseducated, and they may have side effects. To improve the safety of EVs, an appropriate source of EVs should be identified, and parent cells should not be under pathological conditions. In vitro cells are readily influenced by culture conditions, and corresponding EVs can be miseducated to a large extent. Therefore, the body fluids of healthy individuals are outstanding sources of EVs. As an example, EVs separated from the blood of healthy individuals show excellent biosafety and can be used for the delivery of chemotherapeutics. However, EVs separated from body fluids are often compound. Homogeneous types of EVs should be separated from the source, potentially minimizing the heterogeneity. Furthermore, we should make full use of the characteristics of EVs for different diseases. For example, specific types of EVs can target specific types of diseases, facilitating their loading of exogenous drugs to regulate oxidative stress.

In conclusion, EVs can be used as carriers to deliver endogenous or exogenous drugs for the modulation of oxidative, but the source and loading drugs of EVs should be charily selected. And how to reduce the impact of the heterogeneity of EVs should also be carefully considered.

AUTHOR CONTRIBUTIONS

HQ and LY: conceptualization. HQ and YW: original draft preparation. SF and CY: review and editing. LY: supervision. All authors contributed to the manuscript and approved the submitted version.

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Akers, J. C., Gonda, D., Kim, R., Carter, B. S., and Chen, C. C. (2013). Biogenesis of Extracellular Vesicles (EV): Exosomes, Microvesicles, Retrovirus-Like Vesicles, and Apoptotic Bodies. J. Neurooncol. 113, 1–11. doi:10.1007/s11060-013-1084-8

Akers, J. C., Ramakrishnan, V., Nolan, J. P., Duggan, E., Fu, C.-C., Hochberg, F. H., et al. (2016). Comparative Analysis of Technologies for Quantifying Extracellular Vesicles (EVs) in Clinical Cerebrospinal Fluids (CSF). *PLoS One* 11, e0149866. doi:10.1371/journal.pone.0149866

Andaloussi, S., Mäger, I., Breakefield, X. O., and Wood, M. J. A. (2013). Extracellular Vesicles: Biology and Emerging Therapeutic Opportunities. *Nat. Rev. Drug Discov.* 12, 347–357. doi:10.1038/nrd3978

- Arraud, N., Linares, R., Tan, S., Gounou, C., Pasquet, J.-M., Mornet, S., et al. (2014). Extracellular Vesicles from Blood Plasma: Determination of Their Morphology, Size, Phenotype and Concentration. *J. Thromb. Haemost.* 12, 614–627. doi:10.1111/jth.12554
- Ashrafian, F., Shahriary, A., Behrouzi, A., Moradi, H. R., Keshavarz Azizi Raftar, S., Lari, A., et al. (2019). Akkermansia Muciniphila-Derived Extracellular Vesicles as a Mucosal Delivery Vector for Amelioration of Obesity in Mice. *Front. Microbiol.* 10, 2155. doi:10.3389/fmicb.2019.02155
- Atienzar-Aroca, S., Flores-Bellver, M., Serrano-Heras, G., Martinez-Gil, N., Barcia, J. M., Aparicio, S., et al. (2016). Oxidative Stress in Retinal Pigment Epithelium Cells Increases Exosome Secretion and Promotes Angiogenesis in Endothelial Cells. J. Cel. Mol. Med. 20, 1457–1466. doi:10.1111/jcmm.12834
- Betteridge, D. J. (2000). What Is Oxidative Stress? *Metabolism* 49, 3–8. doi:10.1016/ S0026-0495(00)80077-3
- Bodart-Santos, V., de Carvalho, L. R. P., de Godoy, M. A., Batista, A. F., Saraiva, L. M., Lima, L. G., et al. (2019). Extracellular Vesicles Derived from Human Wharton's Jelly Mesenchymal Stem Cells Protect Hippocampal Neurons from Oxidative Stress and Synapse Damage Induced by Amyloid-β Oligomers. Stem Cel Res. Ther. 10, 1–13. doi:10.1186/s13287-019-1432-5
- Bodega, G., Alique, M., Puebla, L., Carracedo, J., and Ramírez, R. M. (2019). Microvesicles: ROS Scavengers and ROS Producers. J. Extracellular Vesicles 8, 1626654. doi:10.1080/20013078.2019.1626654
- Burger, D., Turner, M., Munkonda, M. N., and Touyz, R. M. (2016). Endothelial Microparticle-Derived Reactive Oxygen Species: Role in Endothelial Signaling and Vascular Function. Oxidative Med. Cell Longevity 2016, 1–10. doi:10.1155/ 2016/5047954
- Cho, Y.-E., Seo, W., Kim, D.-K., Moon, P.-G., Kim, S.-H., Lee, B.-H., et al. (2018). Exogenous Exosomes from Mice with Acetaminophen-Induced Liver Injury Promote Toxicity in the Recipient Hepatocytes and Mice. *Sci. Rep.* 8, 1–13. doi:10.1038/s41598-018-34309-7
- Dong, P., Zhang, Y., Yan, D.-y., Wang, Y., Xu, X., Zhao, Y.-c., et al. (2020). Protective Effects of Human Milk-Derived Exosomes on Intestinal Stem Cells Damaged by Oxidative Stress. *Cel Transpl.* 29, 096368972091269. doi:10.1177/ 0963689720912690
- Duan, L., Huang, H., Zhao, X., Zhou, M., Chen, S., Wang, C., et al. (2020). Extracellular Vesicles Derived from Human Placental Mesenchymal Stem Cells Alleviate Experimental Colitis in Mice by Inhibiting Inflammation and Oxidative Stress. Int. J. Mol. Med. 46, 1551–1561. doi:10.3892/ijmm.2020.4679
- Dubois-Deruy, E., Peugnet, V., Turkieh, A., and Pinet, F. (2020). Oxidative Stress in Cardiovascular Diseases. Antioxidants 9, 864. doi:10.3390/antiox9090864
- Eldh, M., Ekström, K., Valadi, H., Sjöstrand, M., Olsson, B., Jernås, M., et al. (2010). Exosomes Communicate Protective Messages during Oxidative Stress; Possible Role of Exosomal Shuttle RNA. *PLoS One* 5, e15353. doi:10.1371/ journal.pone.0015353
- Elsharkasy, O. M., Nordin, J. Z., Hagey, D. W., de Jong, O. G., Schiffelers, R. M., Andaloussi, S. E., et al. (2020). Extracellular Vesicles as Drug Delivery Systems: Why and How? *Adv. Drug Deliv. Rev.* 159, 332–343. doi:10.1016/ j.addr.2020.04.004
- Frank, J., Richter, M., de Rossi, C., Lehr, C.-M., Fuhrmann, K., and Fuhrmann, G. (2018). Extracellular Vesicles Protect Glucuronidase Model Enzymes during Freeze-Drying. Sci. Rep. 8, 1–8. doi:10.1038/s41598-018-30786-y
- Gao, Y., Huang, X., Lin, H., Zhao, M., Liu, W., Li, W., et al. (2021). Adipose Mesenchymal Stem Cell-Derived Antioxidative Extracellular Vesicles Exhibit Anti-Oxidative Stress and Immunomodulatory Effects under PM2.5 Exposure. *Toxicology* 447, 152627. doi:10.1016/j.tox.2020.152627
- Guescini, M., Maggio, S., Ceccaroli, P., Battistelli, M., Annibalini, G., Piccoli, G., et al. (2017). Extracellular Vesicles Released by Oxidatively Injured or Intact C2C12 Myotubes Promote Distinct Responses Converging toward Myogenesis. *Int. J. Mol. Sci.* 18, 2488. doi:10.3390/ijms18112488
- Hargitai, R., Kis, D., Persa, E., Szatmári, T., Sáfrány, G., and Lumniczky, K. (2021). Oxidative Stress and Gene Expression Modifications Mediated by Extracellular Vesicles: An *In Vivo* Study of the Radiation-Induced Bystander Effect. *Antioxidants* 10, 156. doi:10.3390/antiox10020156
- Hayes, J. D., Dinkova-Kostova, A. T., and Tew, K. D. (2020). Oxidative Stress in Cancer. Cancer cell 38, 167–197. doi:10.1016/j.ccell.2020.06.001
- Hedlund, M., Nagaeva, O., Kargl, D., Baranov, V., Mincheva-Nilsson, L., and Mincheva-Nilsson, L. (2011). Thermal- and Oxidative Stress Causes Enhanced Release of NKG2D Ligand-Bearing Immunosuppressive Exosomes in

Leukemia/lymphoma T and B Cells. PLoS One 6, e16899. doi:10.1371/journal.pone.0016899

- Ibrahim, H. M., Mohammed-Geba, K., Tawfic, A. A., and El-Magd, M. A. (2019). Camel Milk Exosomes Modulate Cyclophosphamide-Induced Oxidative Stress and Immuno-Toxicity in Rats. *Food Funct.* 10, 7523–7532. doi:10.1039/ c9fo01914f
- Janiszewski, M., Do Carmo, A. O., Pedro, M. A., Silva, E., Knobel, E., and Laurindo, F. R. M. (2004). Platelet-derived Exosomes of Septic Individuals Possess Proapoptotic NAD(P)H Oxidase Activity: A Novel Vascular Redox Pathway*. Crit. Care Med. 32, 818–825. doi:10.1097/ 01.ccm.0000114829.17746.19
- Lässer, C., Jang, S. C., and Lötvall, J. (2018). Subpopulations of Extracellular Vesicles and Their Therapeutic Potential. *Mol. Aspects Med.* 60, 1–14. doi:10.1016/j.mam.2018.02.002
- Li, Y., Zhou, J., Zhuo, Q., Zhang, J., Xie, J., Han, S., et al. (2019). Malignant Ascite-Derived Extracellular Vesicles Inhibit T Cell Activity by Upregulating Siglec-10 Expression. *Cancer. Manag. Res.* 11, 7123–7134. doi:10.2147/CMAR.S210568
- Li, D., Yang, Y., Wang, S., He, X., Liu, M., Bai, B., et al. (2021). Role of Acetylation in Doxorubicin-Induced Cardiotoxicity. *Redox Biol.* 46, 102089. doi:10.1016/ j.redox.2021.102089
- Li, X., Yang, Y., Wang, Z., Jiang, S., Meng, Y., Song, X., et al. (2021). Targeting Noncoding RNAs in Unstable Atherosclerotic Plaques: Mechanism, Regulation, Possibilities, and Limitations. *Int. J. Biol. Sci.* 17, 3413–3427. doi:10.7150/ ijbs.62506
- Li, X., Yin, D., Yang, Y., Bi, C., Wang, Z., Ma, G., et al. (2021). Eosinophil: A Nonnegligible Predictor in COVID-19 Re-Positive Patients. *Front. Immunol.* 12, 690653. doi:10.3389/fimmu.2021.690653
- Liu, Z., Xu, Y., Wan, Y., Gao, J., Chu, Y., and Li, J. (2019). Exosomes from Adipose-Derived Mesenchymal Stem Cells Prevent Cardiomyocyte Apoptosis Induced by Oxidative Stress. *Cell Death Discov.* 5, 1–7. doi:10.1038/s41420-019-0159-5
- Lorincz, A. M., Schütte, M., Timár, C. I., Veres, D. S., Kittel, A., McLeish, K. R., et al. (2015). Functionally and Morphologically Distinct Populations of Extracellular Vesicles Produced by Human Neutrophilic Granulocytes. J. Leukoc. Biol. 98, 583–589. doi:10.1189/jlb.3VMA1014-514R
- Mahmoud, A. M., Wilkinson, F. L., McCarthy, E. M., Moreno-Martinez, D. M., Langford-Smith, A., Romero, M., et al. (2017). Endothelial Microparticles Prevent Lipid-Induced Endothelial Damage via Akt/eNOS Signaling and Reduced Oxidative Stress. *FASEB j.* 31, 4636–4648. doi:10.1096/ fj.201601244RR
- Manček-Keber, M., Frank-Bertoncelj, M., Hafner-Bratkovič, I., Smole, A., Zorko, M., Pirher, N., et al. (2015). Toll-Like Receptor 4 Senses Oxidative Stress Mediated by the Oxidation of Phospholipids in Extracellular Vesicles. *Sci. Signal.* 8, ra60. doi:10.1126/scisignal.2005860
- Matsumoto, J., Stewart, T., Banks, W. A., and Zhang, J. (2018). The Transport Mechanism of Extracellular Vesicles at the Blood-Brain Barrier. *Curr. Pharm. Des.* 23, 6206–6214. doi:10.2174/1381612823666170913164738
- Mayo, J. S., Kurata, W. E., O'Connor, K. M., and Pierce, L. M. (2019). Oxidative Stress Alters Angiogenic and Antimicrobial Content of Extracellular Vesicles and Improves Flap Survival. *Plast. Reconstr. Surg. - Glob. Open* 7, e2588. doi:10.1097/gox.00000000002588
- Mostefai, H. A., Meziani, F., Mastronardi, M. L., Agouni, A., Heymes, C., Sargentini, C., et al. (2008). Circulating Microparticles from Patients with Septic Shock Exert Protective Role in Vascular Function. Am. J. Respir. Crit. Care Med. 178, 1148–1155. doi:10.1164/rccm.200712-1835OC
- Pascua-Maestro, R., González, E., Lillo, C., Ganfornina, M. D., Falcón-Pérez, J. M., and Sanchez, D. (2019). Extracellular Vesicles Secreted by Astroglial Cells Transport Apolipoprotein D to Neurons and Mediate Neuronal Survival upon Oxidative Stress. *Front. Cel. Neurosci.* 12, 526. doi:10.3389/ fncel.2018.00526
- Qi, H., Liu, C., Long, L., Ren, Y., Zhang, S., Chang, X., et al. (2016). Blood Exosomes Endowed with Magnetic and Targeting Properties for Cancer Therapy. ACS Nano 10, 3323–3333. doi:10.1021/acsnano.5b06939
- Qi, H., Jia, H., Sang, J., Ren, Y., Zhao, J., Hou, X., et al. (2017). Using Endogenous Ligands for Direct Superparamagnetic Nanoparticle Cluster-Based Body Fluid Exosome Separation. RSC Adv. 7, 2926–2933. doi:10.1039/C6RA24937J
- Qi, H., Yang, L., Li, X., Zhan, Q., Han, D., Zhao, J., et al. (2018). Exosomes Separated Based on the "STOP" Criteria for Tumor-Targeted Drug Delivery. J. Mater. Chem. B 6, 2758–2768. doi:10.1039/C8TB00355F

- Rastogi, S., Hwang, A., Chan, J., Wang, J. Y. J., and Luo, K. (2018). Extracellular Vesicles Transfer Nuclear Abl-Dependent and Radiation-Induced miR-34c into Unirradiated Cells to Cause Bystander Effects. *Mol. Biol. Cell.* 29, 2228–2242. doi:10.1091/mbc.E18-02-0130
- Robbins, P. D., and Morelli, A. E. (2014). Regulation of Immune Responses by Extracellular Vesicles. *Nat. Rev. Immunol.* 14, 195–208. doi:10.1038/nri3622
- Shahin, H. I., Radnaa, E., Tantengco, O. A. G., Kechichian, T., Kammala, A. K., Sheller-Miller, S., et al. (2021). Microvesicles and Exosomes Released by Amnion Epithelial Cells under Oxidative Stress Cause Inflammatory Changes in Uterine Cells[†]. *Biol. Reprod.* 105, 464–480. doi:10.1093/biolre/ioab088
- Shen, K., Jia, Y., Wang, X., Zhang, J., Liu, K., Wang, J., et al. (2021). Exosomes from Adipose-Derived Stem Cells Alleviate the Inflammation and Oxidative Stress via Regulating Nrf2/HO-1 axis in Macrophages. Free Radic. Biol. Med. 165, 54–66. doi:10.1016/j.freeradbiomed.2021.01.023
- Soleti, R., Lauret, E., Andriantsitohaina, R., and Carmen Martínez, M. (2012). Internalization and Induction of Antioxidant Messages by Microvesicles Contribute to the Antiapoptotic Effects on Human Endothelial Cells. *Free Radic. Biol. Med.* 53, 2159–2170. doi:10.1016/j.freeradbiomed.2012.09.021
- Sun, T., Ding, Z.-X., Luo, X., Liu, Q.-S., Cheng, Y., and Hassanzadeh, K. (2020). Blood Exosomes Have Neuroprotective Effects in a Mouse Model of Parkinson's Disease. Oxidative Med. Cell Longevity 2020, 1–14. doi:10.1155/2020/3807476
- Théry, C., Zitvogel, L., and Amigorena, S. (2002). Exosomes: Composition, Biogenesis and Function. Nat. Rev. Immunol. 2, 569–579. doi:10.1038/nri855
- Théry, C., Witwer, K. W., Aikawa, E., Alcaraz, M. J., Anderson, J. D., Andriantsitohaina, R., et al. (2018). Minimal Information for Studies of Extracellular Vesicles 2018 (MISEV2018): A Position Statement of the International Society for Extracellular Vesicles and Update of the MISEV2014 Guidelines. J. Extracell. Vesicles 7, 1535750. doi:10.1080/20013078.2018.1535750
- Tkach, M., and Théry, C. (2016). Communication by Extracellular Vesicles: Where We Are and where We Need to Go. Cell 164, 1226–1232. doi:10.1016/ j.cell.2016.01.043
- Vader, P., Mol, E. A., Pasterkamp, G., and Schiffelers, R. M. (2016). Extracellular Vesicles for Drug Delivery. Adv. Drug Deliv. Rev. 106, 148–156. doi:10.1016/ j.addr.2016.02.006
- van der Meel, R., Fens, M. H. A. M., Vader, P., Van Solinge, W. W., Eniola-Adefeso, O., and Schiffelers, R. M. (2014). Extracellular Vesicles as Drug Delivery Systems: Lessons from the Liposome Field. *J. Controlled Release* 195, 72–85. doi:10.1016/j.jconrel.2014.07.049
- van Meteren, N., Lagadic-Gossmann, D., Podechard, N., Gobart, D., Gallais, I., Chevanne, M., et al. (2020). Extracellular Vesicles Released by Polycyclic Aromatic Hydrocarbons-Treated Hepatocytes Trigger Oxidative Stress in Recipient Hepatocytes by Delivering Iron. *Free Radic. Biol. Med.* 160, 246–262. doi:10.1016/j.freeradbiomed.2020.08.001
- Varga, Z., Yuana, Y., Grootemaat, A. E., Van der Pol, E., Gollwitzer, C., Krumrey, M., et al. (2014). Towards Traceable Size Determination of Extracellular Vesicles. J. Extracellular Vesicles 3, 23298. doi:10.3402/jev.v3.23298
- Wang, W., Zheng, Y., Wang, M., Yan, M., Jiang, J., and Li, Z. (2019). Exosomes Derived miR-126 Attenuates Oxidative Stress and Apoptosis from Ischemia and Reperfusion Injury by Targeting ERRFI1. Gene 690, 75–80. doi:10.1016/j.gene.2018.12.044
- Wang, T., Jian, Z., Baskys, A., Yang, J., Li, J., Guo, H., et al. (2020). MSC-derived Exosomes Protect against Oxidative Stress-Induced Skin Injury via Adaptive Regulation of the NRF2 Defense System. *Biomaterials* 257, 120264. doi:10.1016/ j.biomaterials.2020.120264
- Winck, F. V., Prado Ribeiro, A. C., Ramos Domingues, R., Ling, L. Y., Riaño-Pachón, D. M., Rivera, C., et al. (2015). Insights into Immune Responses in Oral Cancer through Proteomic Analysis of Saliva and Salivary Extracellular Vesicles. Sci. Rep. 5, 1–13. doi:10.1038/srep16305
- Xi, X. J., Zeng, J. J., Lu, Y., Chen, S. H., Jiang, Z. W., He, P. J., et al. (2020). Extracellular Vesicles Enhance Oxidative Stress through P38/NF-kB Pathway in Ketamine-induced Ulcerative Cystitis. J. Cel. Mol. Med. 24, 7609–7624. doi:10.1111/jcmm.15397
- Xue, Q., He, N., Wang, Z., Fu, X., Aung, L. H. H., Liu, Y., et al. (2021). Functional Roles and Mechanisms of Ginsenosides from Panax Ginseng in Atherosclerosis. *J. Ginseng Res.* 45, 22–31. doi:10.1016/j.jgr.2020.07.002

- Yang, L., Han, D., Zhan, Q., Li, X., Shan, P., Hu, Y., et al. (2019). Blood TfR+ Exosomes Separated by a pH-Responsive Method Deliver Chemotherapeutics for Tumor Therapy. *Theranostics* 9, 7680–7696. doi:10.7150/thno.37220
- Yao, J., Zheng, J., Cai, J., Zeng, K., Zhou, C., Zhang, J., et al. (2018). Extracellular Vesicles Derived from Human Umbilical Cord Mesenchymal Stem Cells Alleviate Rat Hepatic Ischemia-Reperfusion Injury by Suppressing Oxidative Stress and Neutrophil Inflammatory Response. *FASEB j.* 33, 1695–1710. doi:10.1096/fj.201800131RR
- Yin, P., Lv, H., Li, Y., Deng, Y., Zhang, L., and Tang, P. (2017). Exosome-mediated Genetic Information Transfer, a Missing Piece of Osteoblast-Osteoclast Communication Puzzle. *Front. Endocrinol.* 8, 336. doi:10.3389/ fendo.2017.00336
- Zhan, Q., Yi, K., Qi, H., Li, S., Li, X., Wang, Q., et al. (2020). Engineering Blood Exosomes for Tumor-Targeting Efficient Gene/chemo Combination Therapy. *Theranostics* 10, 7889–7905. doi:10.7150/thno.45028
- Zhang, G., Zou, X., Miao, S., Chen, J., Du, T., Zhong, L., et al. (2014). The Anti-Oxidative Role of Micro-Vesicles Derived from Human Wharton-Jelly Mesenchymal Stromal Cells through NOX2/gp91(phox) Suppression in Alleviating Renal Ischemia-Reperfusion Injury in Rats. *PLoS One* 9, e92129. doi:10.1371/journal.pone.0092129
- Zhang, G., Zou, X., Huang, Y., Wang, F., Miao, S., Liu, G., et al. (2016a). Mesenchymal Stromal Cell-Derived Extracellular Vesicles Protect against Acute Kidney Injury through Anti-oxidation by Enhancing Nrf2/ARE Activation in Rats. *Kidney Blood Press. Res.* 41, 119–128. doi:10.1159/ 000443413
- Zhang, Q., Shang, M., Zhang, M., Wang, Y., Chen, Y., Wu, Y., et al. (2016b). Microvesicles Derived from Hypoxia/reoxygenation-Treated Human Umbilical Vein Endothelial Cells Promote Apoptosis and Oxidative Stress in H9c2 Cardiomyocytes. *BMC Cel Biol.* 17, 25. doi:10.1186/s12860-016-0100-1
- Zhang, P., Li, T., Wu, X., Nice, E. C., Huang, C., and Zhang, Y. (2020). Oxidative Stress and Diabetes: Antioxidative Strategies. *Front. Med.* 14, 583–600. doi:10.1007/s11684-019-0729-1
- Zhou, Y., Xu, H., Xu, W., Wang, B., Wu, H., Tao, Y., et al. (2013). Exosomes Released by Human Umbilical Cord Mesenchymal Stem Cells Protect against Cisplatin-Induced Renal Oxidative Stress and Apoptosis In Vivo and In Vitro. Stem Cel Res. Ther. 4, 34. doi:10.1186/scrt194
- Zong, T., Yang, Y., Lin, X., Jiang, S., Zhao, H., Liu, M., et al. (2021). 5'-tiRNA-Cys-GCA Regulates VSMC Proliferation and Phenotypic Transition by Targeting STAT4 in Aortic Dissection. *Mol. Ther. - Nucleic Acids.* in press doi:10.1016/ j.omtn.2021.07.013
- Zonneveld, M. I., Brisson, A. R., van Herwijnen, M. J. C., Tan, S., van de Lest, C. H. A., Redegeld, F. A., et al. (2014). Recovery of Extracellular Vesicles from Human Breast Milk Is Influenced by Sample Collection and Vesicle Isolation Procedures. J. Extracellular Vesicles 3, 24215. doi:10.3402/jev.v3.24215

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