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ORIGINAL RESEARCH

Design and synthesis of chroman derivatives with dual anti-breast cancer and antiepileptic activities

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Abstract: A series of chroman derivatives was designed, prepared, and examined for their anti-breast cancer and antiepileptic activities. All synthesized compounds yielded results that were in good agreement with spectral data. The bioassay showed that some of the resultant compounds exerted remarkable inhibitory effects on growth of human breast cancer cell line MCF-7. In particular, compound 6i (the concentration required for 50% inhibition of cell growth $[GI_{\text{sol}}] = 34.7 \mu M$) exerted promising anticancer activity toward MCF-7 cell line. Additionally, compounds 6b, 6c, 6d, 6e, 6g, 6i, and 6l showed advanced antiepileptic activity than reference drugs. None of the compounds showed neurotoxicity, as determined by the rotarod test. The obtained results proved that these distinctive compounds could be relevant as models for future discovery and research, as well as for the production of more number of active derivatives. **Keywords:** anti-breast cancer, antiepileptic, chroman, isatin, Schiff base

Introduction

Breast cancer is one of the most commonly diagnosed cancers and is the second leading cause of death among women.¹ Currently, a large number of chemotherapeutic agents are available, but toxicity and resistance to standard drug treatments limit the effectiveness of chemotherapy. Therefore, the search for new chemotherapeutic agents is an important task for researchers.

Heterocyclic compounds containing oxygen are presently considered very interesting due to their physicochemical properties being relevant as far as the design of new drugs is concerned. Literature reveals that chromans are an important chemical synthon, associated with a broad range of biological effects, neuroprotective,² antiestrogens,³ antioxidant, 4 anti-HIV, 5 and anti-breast cancer agents. $6-8$

Chromans have attracted a great deal of attention due to their anticancer activities since the discovery of Ormeloxifene (I) , KBU2046 (Phase II),¹⁰ and B43-genistein (II) (Phase II; Figure 1).¹¹ Furthermore, many chromans have contributed to the search for new anticancer agents. It has also been reported that Schiff base (III and IV) and isatin (V) moieties enhanced the anticancer activity of the chroman core.^{12,13}

Epilepsy is a common neurological disorder affecting approximately 45–100 million people globally.14 Though many antiepileptic drugs are available in clinical use, neurotoxicity and distinctive adverse effects restrict their clinical use.15 Therefore, it is necessary to discover new chemical pharmacophores as more effective antiepileptics with less neurotoxicity. Literature has revealed that the presence of a Schiff base in a chroman nucleus (VI–VIII) is beneficial for the antiepileptic activity.16 Similarly, isatin (IX, X) being associated with a Schiff base accounts for its antiepileptic activity.17

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Figure 1 Reported and designed chromans and isatins as anticancer and antiepileptic agents (6a–q).

Reports also suggest that some anticancer compounds have been studied as antiepileptics.¹⁸ The anti-breast cancer potential of the voltage-gated sodium channel-blocking antiepileptic drug phenytoin has also been discussed by researchers.19 Chroman derivatives XI and XII have been investigated as voltage-gated sodium channels.20 Therefore, chroman analogs designed as anticancer agents may have antiepileptic activity also.

In this study, advances are made in designing chroman derivatives by adding Schiff base and isatin moieties at the second position. Seven compounds with antimicrobial activity have been reported in a previous work.²¹ A series of chroman compounds (6a–q) have been designed, synthesized, and their anti-breast cancer and antiepileptic activities evaluated.

Materials and methods Materials

All commercial chemicals were purchased from Sigma-Aldrich (St Louis, MO, USA) and Spectrochem (Mumbai, India). Thin layer chromatography was performed on plates precoated with silica gel GF-254 (Merck Millipore, Billerica, MA, USA), and the identification was done under UV light and spraying with charring solution, followed by heating. Melting points were determined in open glass capillaries and the uncorrected. NMR spectra were recorded on ECX-500 spectrometer (JEOL, Toko, Japan) in dimethyl sulfoxide (DMSO)- d_6 and deuterated chloroform (CDCl₃) at 400 MHz and 500 MHz for 1 H and 125 MHz for 13 C. The chemical shifts were recorded in (δ, ppm) relative to tetramethylsilane as an internal standard. The mass spectra

were recorded with a Waters-Q-Tof Premier-HAB213 spectrometer and Microscopic II triple-quadrupole mass spectrometer using electron ionization, and the *m/z* values are indicated in daltons. The infrared (IR) spectra (KBr) were recorded on an FTIR Vector 22 spectrophotometer (Bruker Corporation, Billerica, MA, USA). [Supplementary](https://www.dropbox.com/s/v7nkn2ib91k69if/SUPPLEMENTARY%20-%20new.docx?dl=0) [materials](https://www.dropbox.com/s/v7nkn2ib91k69if/SUPPLEMENTARY%20-%20new.docx?dl=0) show ¹H and ¹³C NMR spectra, FT-IR spectra, and mass spectra of compounds **6a**–**q** associated with this article.

Synthesis of compounds

6-Hydroxy-2,5,7,8-tetramethyl-chroman-2-carboxylic acid methyl ester (3a)

6-Hydroxy-2,5,7,8-tetramethyl-chroman-2-carboxylic acid methyl ester (3a) was prepared by the method provided in the literature.²²

6-Methoxy-2,5,7,8-tetramethyl-chroman-2-carboxylic acid methyl ester (3b)

To a solution of 3a (100 mg, 40.0 mmol) in 10 mL anhydrous acetone, 2 g anhydrous K_2CO_3 was added and the mixture was heated at 50°C. Dimethyl sulfate (0.17 mL, 1.80 mmol) was then added, and the mixture was refluxed for 24 hours. After completion of the reaction, HCl 10% was added until pH 6 and then extracted with diethylether. The organic layer was washed with saturated aqueous NaCl and dried, and the solvent was evaporated in vacuo. Purification by column chromatography (petroleum ether/EtOAc: 90/10) yielded 111 mg (100%) white solid.

Methyl-6-(benzyloxy)-2,5,7,8-tetramethyl-3,4 dihydro-2H-chromene-2-carboxylate (3c)

To a mixture of 3a (2 g, 7.566 mmol) and K_2CO_3 (1.568 g, 11.349 mmol) was added with stirring 30 mL of dimethylformamide at 0°C for 20 minutes. Benzyl bromide (1.553 g, 9.080 mmol) was added to the reaction solution and stirred at room temperature overnight. The reaction mixture was diluted with 50 mL cold water, filtered, and washed with cold water to give a white solid (81.67%).

General method for the synthesis of 6-substituted-2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (4a–c)

Compounds 3a–c (1 mmol) and 80% hydrazine hydrate (5 mmol) in 10 mL ethanol were refluxed at 80°C for 10 hours. While the reaction proceeded, the ethanol was evaporated. The separated solid was filtered, washed with water, and dried to obtain a dull white solid.

General method for the synthesis of the final compounds (6a–q)

The compounds were prepared through the condensation reaction between 5a–q (1 mmol) and 4a–c (1 mmol) in 10 mL acetic acid under reflux at 120°C for 3–12 hours. Then, 20 mL distilled water was added into the reaction medium. The compounds were filtered and recrystallized in ethanol.

6-Hydroxy-2,5,7,8-tetramethyl-N′-(2 oxoindolin-3-ylidene)-3,4-dihydro-2H-chromene-2 carbohydrazide (6a)

Yield: Yellow solid (74%); melting point (MP): 240°C; IR (KBr, ν, cm⁻¹): 3,426 (NH), 3,227 (OH), 1,694 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 11.17 (s, 1H, -NH), 7.52 (s, 1H, OH), 7.48 (d, *J*=*7*.*3 Hz*, 1H), 7.32 (t, *J*=*7*.*6 Hz*, 1H), 7.03 (t, *J*=*7*.*6 Hz*, 1H), 6.86 (d, *J*=*7*.*6 Hz*, 1H), 2.59–2.54 (m, 1H), 2.47–2.45 (m, 1H), 2.28–2.25 (m, 1H), 2.18 (s, 3H), 2.04 (s, 3H), 1.96 (s, 3H), 1.86–1.81 (m, 1H), 1.47 (s, 3H); ¹³C-NMR (125 MHz, DMSO-d₆) 12.3 (-CH₃), 12.5 (-CH₃), 13.3 (-CH₃), 20.4 (-CH₂), 24.7 (-CH₃), 29.6 (-CH₂), 78.1 $(*SC*, 111.6, 117.2, 120.2, 120.8, 121.4, 122.2, 123.1,$ $123.5, 132.3, 139.1, 143.1, 143.8, 146.7, 162.9 (>C=0),$ 171.9 ($>$ C=O); high-resolution mass spectrometry (HR-MS): 394.1700 (M+H)⁺ , calculated: 394.1766.

6-hydroxy-2,5,7,8-tetramethyl-N′-(1-methyl-2-oxoindolin-3-ylidene)-3,4-dihydro-2H-chromene-2-

carbohydrazide (6b)

Yield: Yellow solid (72%) , MP: 237° C; IR (KBr, v, cm⁻¹): 3,406 (OH), 3,261 (NH), 1,683 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 7.54 (s, 1H, OH), 7.50 (d, *J*=7.3 *Hz*, 1H), 7.40 (t, *J*=*7*.*6 Hz*, 1H), 7.10–7.05 (m, 2H), 3.13 (s, 3H), 2.59–2.54 (m, 1H), 2.50–2.44 (m, 1H), 2.29–2.23 (m, 1H), 2.21 (s, 3H), 2.05 (s, 3H), 1.96 (s, 3H), 1.87–1.81 (m, 1H), 1.47 (s, 3H); ¹³C-NMR (125 MHz, DMSO-d₆) 12.3 (-CH₃), 12.5 (-CH₃), 13.3 (-CH₃), 20.4 (-CH₂), 24.6 (-CH₃), 26.1 $($ >N–CH₃), 29.6 (–CH₂), 78.1 (>C<), 110.2, 117.3, 119.5, 120.8, 121.0, 122.2, 123.5, 123.6, 132.2, 138.2, 143.8, 144.3, 146.83, 161.0 ($>$ C=O), 171.9 ($>$ C=O); HR-MS: 408.1926 (M+H)⁺ , calculated, 408.4702.

N′-(5-chloro-2-oxoindolin-3-ylidene)-6-hydroxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2-

carbohydrazide (6c)

Yield: Light Yellow solid (87%), MP: 255°C; IR (KBr, ν, cm-¹): 3,598 (NH), 3,151 (OH), 1,680 (CO); 1 H-NMR $(500 \text{ MHz}, \text{ DMSO-d}_6, \text{ ppm})$: 11.28 (s, 1H, -NH), 7.54 (s,

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1H, OH), 7.46 (d, *J*=*2*.*1 Hz*, 1H), 7.35 (dd, *J*=*2*.*1, 8*.*2 Hz*, 1H), 6.88 (d, *J*=*8*.*2 Hz*, 1H), 2.60–2.54 (m, 1H), 2.47–2.43 (m, 1H), 2.30–2.24 (m, 1H), 2.17 (s, 3H), 2.04 (s, 3H), 1.96 (s, 3H), 1.86–1.80 (m, 1H), 1.47 (s, 3H); 13C-NMR (125 MHz, DMSO- d_6) 12.3 (-CH₃), 12.4 (-CH₃), 13.3 (-CH₃), 20.3 (-CH₂), 24.6 (-CH₃), 29.5 (-CH₂), 78.1 (>C<), 113.1, 117.2, 120.8, 120.9, 122.0, 122.2, 123.5, 127.2, 131.7, 138.2, 141.8, 143.8, 146.8, 162.7 ($>C=O$), 172.0 ($-C=O$); HR-MS: 450.1377 (M+H)⁺ , calculated, 450.1196.

N′-(5-bromo-2-oxoindolin-3-ylidene)-6-hydroxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6d)

Yield: Yellow solid (56%) , MP: 260°C; IR (KBr, v, cm⁻¹): 3,594 (NH), 3,149 (OH), 1,680 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 11.30 (s, 1H, -NH), 7.56 (s, 1H,), 7.54 (s, 1H, OH), 7.48 (d, *J*=*8*.*0 Hz*, 1H), 6.83 (d, *J*=*8*.*0 Hz*, 1H), 2.60–2.54 (m, 1H), 2.47–2.43 (m, 1H), 2.29–2.24 (m, 1H), 2.17 (s, 3H), 2.03 (s, 3H), 1.96 (s, 3H), 1.86–1.80 (m, 1H), 1.47 (s, 3H); ¹³C-NMR (125 MHz, DMSO- d_6) 12.3 $(-CH₃), 12.4 (-CH₃), 13.3 (-CH₃), 20.3 (-CH₂), 24.6 (-CH₃),$ 29.5 (–CH₂), 78.1 (>C<), 113.5, 114.8, 117.2, 120.8, 122.2, 122.4, 123.5, 123.7, 134.5, 138.1, 142.1, 143.8, 146.8, 162.6 $(>=0)$, 172.0 $(>=0)$; HR-MS: 494.0690 $(M+Na)^{+}$, calculated, 494.0691.

N′-(6-chloro-2-oxoindolin-3-ylidene)-6-hydroxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2-

carbohydrazide (6e)

Yield: Yellow solid (57%) , MP: 290°C; IR (KBr, v, cm⁻¹): 3,443 (NH), 3,141 (OH), 1,678 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 11.20 (s, 1H, -NH), 7.54 (s, 1H, OH), 7.29 (dd, *J*=*2*.*5, 8*.*0 Hz*, 1H), 7.19–7.14 (m, 1H), 6.87–6.85 (dd, *J*=*4*.*3, 8*.*5 Hz*, 1H), 2.60–2.54 (m, 1H), 2.50–2.40 (m, 1H), 2.29–2.24 (m, 1H), 2.17 (s, 3H), 2.03 (s, 3H), 1.96 (s, 3H), 1.85–1.80 (m, 1H), 1.47 (s, 3H); HR-MS: 428.1372 (M+H)⁺ , calculated, 428.1377.

N′-(5-fluoro-2-oxoindolin-3-ylidene)-6-hydroxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2-

carbohydrazide (6f)

Yield: Yellow solid (51%) , MP: 290°C; IR (KBr, v, cm⁻¹): 3,566 (NH), 3,206 (OH), 1,686 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 11.18 (s, 1H, -NH), 7.52 (s, 1H, OH), 7.29 (dd, *J*=*2*.*5, 8*.*0 Hz*, 1H), 7.18–7.14 (m, 1H), 6.86 (dd, *J*=*4*.*3, 8*.*6 Hz*, 1H), 2.59–2.54 (m, 1H), 2.50–2.42 (m, 1H), 2.30–2.25 (m, 1H), 2.18 (s, 3H), 2.04 (s, 3H), 1.96 (s, 3H), 1.86–1.80 (m, 1H), 1.47 (s, 3H); 13C-NMR (125 MHz, DMSO- d_6) 12.3 (-CH₃), 12.4 (-CH₃), 13.3 (-CH₃),

 20.4 (-CH₂), 24.6 (-CH₃), 29.6 (-CH₂), 78.1 (>C<), 108.4, 108.6, 112.6, 112.7, 117.2, 118.6, 118.8, 120.8, 121.5, 121.6, 122.2, 123.5, 138.7, 139.4, 143.8, 146.8, 157.8, 159.7, 163.1 $(>=0)$, 172.0 $(>=0)$; HR-MS: 412.1681 $(M+H)^{+}$, calculated, 412.1672.

N′-(6-chloro-1-methyl-2-oxoindolin-3-ylidene)-6 hydroxy-2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6g)

Yield: Yellow solid (59%) , MP: 257°C; IR (KBr, v, cm⁻¹): 3,479 (NH), 3,223 (OH), 1,697 (CO); 1 H-NMR (500 MHz, DMSO-d₆, ppm): 7.54 (s, 1H, OH), 7.47–7.44 (m, 2H), 7.10 (d, *J*=*8*.*2 Hz*, 1H), 3.13 (s, 3H), 2.60–2.54 (m, 1H), 2.50–2.41 (m, 1H), 2.29–2.24 (m, 1H), 2.20 (s, 3H), 2.04 (s, 3H), 1.96 (s, 3H), 1.87–1.81 (m, 1H), 1.47 (s, 3H); 13C-NMR (125 MHz, DMSO-d₆) 12.2 (–CH₃), 12.5 (–CH₃), 13.3 $(-CH₃), 20.3 (-CH₂), 24.6 (-CH₃), 26.3 (>N-CH₃), 29.5$ $(-CH₂), 78.1$ (>C<), 111.8, 117.2, 120.5, 120.8, 121.2, 122.2, 123.5, 127.7, 131.5, 137.3, 143.0, 143.7, 146.8, 160.8 $($ >C=O), 172.0 (>C=O); HR-MS: 442.1537 (M+H)⁺, calculated, 442.1530.

6-Methoxy-2,5,7,8-tetramethyl-N′-(2-oxoindolin-3-

ylidene)-3,4-dihydro-2H-chromene-2-carbohydrazide (6h) Yield: Light Yellow solid (58%), MP: 256°C; IR (KBr, ν, cm-¹): 3,184 (NH), 1,683 (CO); 1 H-NMR (500 MHz, DMSO-d6 , ppm): 11.19 (s, 1H, –NH), 7.49 (d, *J*=*7*.*3 Hz*, 1H), 7.32 (t, *J*=*7*.*6 Hz*, 1H), 7.03 (t, *J*=*7*.*6 Hz*, 1H), 6.87 (d, *J*=8.0 Hz, 1H), 3.49 (s, 3H, -OCH₃), 2.62-2.58 (m, 1H), 2.51–2.46 (m, 1H), 2.29–2.23 (m, 1H), 2.19 (s, 3H), 2.08 (s, 3H), 2.01 (s, 3H), 1.89–1.84 (m, 1H), 1.48 (s, 3H); ¹³C-NMR (125 MHz, DMSO-d₆) 11.9 (-CH₃), 12.4 (-CH₃), 12.9 (-CH₃), 20.1 (-CH₂), 24.5 (-CH₃), 29.3 (-CH₂), 60.3 $(-OCH₃), 78.4 (>C<), 111.6, 118.0, 120.2, 121.5, 123.1,$ 123.1, 126.0, 128.0, 132.4, 139.2, 143.1, 146.4, 150.5, 162.9 $($ >C=O), 171.6 (>C=O); HR-MS: 408.1924 (M+H)⁺, calculated, 408.1923.

N′-(5-bromo-2-oxoindolin-3-ylidene)-6-methoxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6i)

Yield: Yellow solid (51%) , MP: 290°C; IR (KBr, v, cm⁻¹): 3,194 (NH), 1,681 (CO); ¹H-NMR (500 MHz, DMSO-d₆, ppm): 11.31 (s, 1H, –NH), 7.57 (s, 1H), 7.49 (d, *J*=*8*.*2 Hz*, 1H), 6.84 (d, *J*=8.2 *Hz*, 1H), 3.48 (s, 3H, -OCH₃), 2.61-2.57 (m, 1H), 2.50–2.45 (m, 1H), 2.30–2.23 (m, 1H), 2.19 (s, 3H), 2.08 (s, 3H), 2.01 (s, 3H), 1.88–1.86 (m, 1H), 1.48 (s, 3H); ¹³C-NMR (125 MHz, DMSO-d₆) 11.9 (–CH₃), 12.4 (–CH₃), 12.9 (-CH₃), 20.1 (-CH₂), 24.5 (-CH₃), 29.2 (-CH₂), 60.3

 $(-OCH₃), 78.4 (>C<), 113.6, 114.8, 118.0, 122.4, 123.0,$ 123.7, 126.1, 128.0, 134.5, 138.2, 142.1, 146.3, 150.6, 162.6 ($>C=O$), 171.7 ($>C=O$); HR-MS: 486.1027 (M+H)⁺, calculated, 486.1028.

N′-(5-fluoro-2-oxoindolin-3-ylidene)-6-methoxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2-

carbohydrazide (6j)

Yield: Yellow solid (66%), MP: $>$ 300°C; IR (KBr, v, cm⁻¹): 3,487 (NH), 1,684 (CO); ¹H-NMR (500 MHz, DMSO- d_6 , ppm): 11.21 (s, 1H, –NH), 7.30 (dd, *J*=*2*.*8, 8*.*0 Hz*, 1H), 7.19–7.15 (m, 1H), 6.87 (dd, *J*=*4*.*3, 8*.*5 Hz*, 1H, 1H), 3.49 (s, 3H, -OCH₃), 2.62-2.57 (m, 1H), 2.50-2.45 (m, 1H), 2.27–2.22 (m, 1H), 2.19 (s, 3H), 2.08 (s, 3H), 2.01 (s, 3H), 1.88–1.85 (m, 1H), 1.48 (s, 3H); 13C-NMR (125 MHz, $DMSO-d₆$) 11.9(-CH₃), 12.4 (-CH₃), 12.9 (-CH₃), 20.1 $(-CH₂)$, 24.4 $(-CH₃)$, 29.3 $(-CH₂)$, 60.3 $(-OCH₃)$, 78.4 $($ >C \lt), 108.5, 108.7, 112.7, 112.8, 118.0, 118.6, 118.8, 121.5, 121.6, 123.1, 126.1, 128.0, 138.9, 139.4, 146.3, 150.6 , 157.8 , 159.7 , 163.1 ($>C=O$), 171.7 ($>C=O$); HR-MS: 426.1822 (M+H)⁺ , calculated, 426.1829.

N′-(6-chloro-2-oxoindolin-3-ylidene)-6-methoxy-2,5,7,8 tetramethyl-3,4-dihydro-2H-chromene-2-

carbohydrazide (6k)

Yield: Yellow solid (52%) , MP: 270° C; IR (KBr, v, cm⁻¹): 3,207 (NH), 1,683 (CO); ¹H-NMR (500 MHz, DMSO- d_6 , ppm): 11.32 (s, 1H, –NH), 7.48 (d, *J*=*8*.*0 Hz*, 1H), 7.07 (d, *J*=8.0 Hz, 1H), 6.90 (s, 1H), 3.49 (s, 3H, -OCH₃), 2.63–2.57 (m, 1H), 2.51–2.42 (m, 1H), 2.29–2.24 (m, 1H), 2.19 (s, 3H), 2.08 (s, 3H), 2.01 (s, 3H), 1.90–1.84 (m, 1H), 1.48 (s, 3H); HR-MS: 442.1539 (M+H)⁺ , calculated, 442.1539.

6-(benzyloxy)-2,5,7,8-tetramethyl-N′-(2-oxoindolin-3-

ylidene)-3,4-dihydro-2H-chromene-2-carbohydrazide (6l) Yield: Light Yellow solid (74%), MP: 258°C; IR (KBr, ν, cm-¹): 3,186 (NH), 1,679 (CO); 1 H-NMR (500 MHz, DMSO-d6 , ppm): 11.20 (s, 1H, –NH), 7.50 (d, *J*=*8*.*0 Hz*, 1H), 7.43 (d, *J*=*7*.*3 Hz*, 2H), 7.36 (t, *J*=*7*.*3 Hz*, 2H), 7.33–7.30 (m, 2H), 7.04 (t, *J*=*7*.*3 Hz*, 1H), 6.87 (d, *J*=*8*.*0 Hz*, 1H), 4.60 (s, 2H), 2.65–2.60 (m, 1H), 2.54–2.47 (m, 1H), 2.30–2.26 (m, 1H), 2.22 (s, 3H), 2.13 (s, 3H), 2.06 (s, 3H), 1.92–1.86 $(m, 1H), 1.49$ (s, 3H); ¹³C-NMR (125 MHz, DMSO- d_6) 12.3 (-CH₃), 12.5 (-CH₃), 13.2 (-CH₃), 20.3 (-CH₂), 24.5 $(-CH₃), 29.4 (-CH₂), 74.6 (-CH₂), 78.5 (>CH₂), 111.6,$ 118.1, 120.3, 121.5, 123.2, 126.4, 128.3, 128.9, 132.4, 138.1, 139.2, 143.1, 146.6, 149.3, 163.0 (\geq C=O), 171.7 (\geq C=O); HR-MS: 484.2235 (M+H)⁺ , calculated, 484.2236.

6-(benzyloxy)-2,5,7,8-tetramethyl-N′-(1-methyl-2 oxoindolin-3-ylidene)-3,4-dihydro-2H-chromene-2 carbohydrazide (6m)

Yield: Yellow-orange solid (92%), MP: 170°C; IR (KBr, v, cm⁻¹): 3,236 (NH), 1,696 (CO); ¹H-NMR (500 MHz, CDCl₃, ppm): 7.77 (d, *J*=*7*.*3 Hz*, 1H), 7.47 (d, *J*=*6*.*8 Hz*, 2H), 7.39–7.36 (m, 2H), 7.35–7.30 (m, 2H), 7.10 (t, *J*=*7*.*3 Hz*, 1H), 6.82 (d, *J*=*8*.*0 Hz*, 1H), 4.68 (s, 2H), 3.23 (s, 3H), 2.70–2.62 (m, 2H), 2.53–2.49 (m, 1H), 2.37 (s, 3H), 2.25 (s, 3H), 2.13 (s, 3H), 2.03–1.99 (m, 1H); ¹³C-NMR (125 MHz, CHCl₃) 12.1 (-CH₃), 12.2 $(-CH₃), 13.0 (-CH₃), 20.4 (-CH₂), 24.6 (-CH₃), 25.7 (>N-CH₃),$ 29.5 (-CH₂), 74.7 (-CH₂), 78.5 (>C<), 108.8, 117.9, 119.6, 112.0, 123.4, 123.7, 126.3, 127.7, 127.9, 128.5, 128.6, 131.7, 137.9, 138.2, 143.8, 146.6, 149.3, 161.0 (\geq C=O), 172.9 (\geq C=O); HR-MS: 498.2399 (M+H)⁺, calculated, 498.2392.

6-(benzyloxy)-N′-(5-bromo-2-oxoindolin-3-ylidene)- 2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6n)

Yield: Yellow solid (68%), MP: 270°C; IR (KBr, v, cm⁻¹): 3,189 (NH), 1,677 (CO); ¹H-NMR (500 MHz, DMSO-d₆, ppm): 11.32 (s, 1H, –NH), 7.57 (s, 1H), 7.50 (d, *J*=*8*.*2 Hz*, 1H), 7.43 (d, *J*=*7*.*3 Hz*, 2H), 7.37 (t, *J*=*7*.*3 Hz*, 2H), 7.33–7.30 (m, 1H), 6.84 (d, *J*=*8*.*2 Hz*, 1H), 4.59 (s, 2H), 2.66–2.58 (m, 1H), 2.54– 2.44 (m, 1H), 2.32–2.25 (m, 1H), 2.22 (s, 3H), 2.13 (s, 3H), 2.06 (s, 3H), 1.94–1.86 (m, 1H), 1.50 (s, 3H); 13C-NMR (125 MHz, DMSO-d₆): 12.2 (-CH₃), 12.4 (-CH₃), 13.2 (-CH₃), 20.2 $(-CH₂), 24.4 (-CH₃), 29.3 (-CH₂), 74.6 (-CH₂), 78.5 ([>]C<),$ 113.6, 118.1, 121.5, 123.1, 123.7, 126.4, 128.3, 128.9, 135.0, 138.2, 142.2, 146.5, 149.3, 162.7 ($>C=0$), 171.7 ($>C=0$); HR-MS: 562.1349 (M+H)⁺ , calculated, 562.1341.

6-(benzyloxy)-N′-(5-fluoro-2-oxoindolin-3-ylidene)- 2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6o)

Yield: Yellow solid (85%), MP: $>$ 300°C; IR (KBr, v, cm⁻¹): 3,195 (NH), 1,681 (CO); ¹H-NMR (500 MHz, DMSO- d_6 , ppm): 11.21 (s, 1H, –NH), 7.43 (d, *J*=*7*.*4 Hz*, 2H), 7.37 (t, *J*=*7*.*0 Hz*, 2H), 7.31–7.30 (m, 2H), 7.18 (t, *J*=*9*.*0 Hz*, 1H), 6.87 (dd, *J*=*4, 8*.*6 Hz*, 1H), 4.59 (s, 2H), 2.65–2.60 (m, 1H), 2.53–2.47 (m, 1H), 2.31–2.25 (m, 1H), 2.22 (s, 3H), 2.12 (s, 3H), 2.06 (s, 3H), 1.92–1.87 (m, 1H), 1.50 (s, 3H); HR-MS: 502.2146 (M+H)⁺ , calculated, 502.2142.

6-(benzyloxy)-N′-(5-chloro-1-methyl-2-oxoindolin-3 ylidene)-2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6p)

Yield: Dark Yellow solid (81%), MP: 220°C; IR (KBr, ν, cm-¹): 3,260 (NH), 1,712 (CO); 1 H-NMR (500 MHz,

DMSO-d₆, ppm): 7.49–7.46 (m, 2H), 7.44–7.43 (m, 2H), 7.37 (t, *J*=*7*.*3 Hz*, 2H), 7.33–7.29 (m, 1H), 7.12 (d, *J*=*8*.*5 Hz*, 1H), 4.59 (s, 2H), 3.14 (s, 3H), 2.65–2.60 (m, 1H), 2.54–2.47 (m, 1H), 2.30–2.25 (m, 1H), 2.24 (s, 3H), 2.13 (s, 3H), 2.06 (s, 3H), 1.93–1.89 (m, 1H), 1.50 (s, 3H); HR-MS: 532.1992 (M+H)⁺ , calculated, 532.2003.

6-(benzyloxy)-N′-(5-chloro-2-oxoindolin-3-ylidene)- 2,5,7,8-tetramethyl-3,4-dihydro-2H-chromene-2 carbohydrazide (6q)

Yield: Yellow solid (94%), MP: 280° C; IR (KBr, v, cm⁻¹): 3,188 (NH), 1,678 (CO); ¹H-NMR (500 MHz, DMSO- d_6 , ppm): 11.31 (s, 1H, –NH), 7.46 (d, *J*=*2*.*5 Hz*, 1H), 7.44–7.42 (m, 2H), 7.38–7.35 (m, 3H), 7.32–7.29 (m, 1H), 6.89 (d, *J*=*8*.*0 Hz*, 1H), 4.60 (s, 2H), 2.66–2.60 (m, 1H), 2.53–2.47 (m, 1H), 2.31–2.27 (m, 1H), 2.21 (s, 3H), 2.12 (s, 3H), 2.06 (s, 3H), 1.92–1.87 (m, 1H), 1.50 (s, 3H); HR-MS: 516.1685 (M-H)- , calculated, 516.1690.

In vitro anticancer activity determination by sulforhodamine B assay

The anticancer activity was measured in vitro in the human breast cancer cell line (MCF-7) using sulforhodamine B stain assay applying the method of Skehan et al.²³ Human breast cancer cell line MCF-7 was obtained from the Advanced Centre for Treatment Research and Education in Cancer (ACTREC), Navi Mumbai, Maharashtra, India, and were grown in RPMI-1640 medium supplemented with 10% heatinactivated fetal calf serum, 100 µg/mL streptomycin, and 100 U/mL penicillin at 37°C in a humidified atmosphere with 5% CO₂. All compounds (6a–q) were dissolved in DMSO to give a stock solution of 10 µg/mL, from which further dilutions (10⁻⁷ M, 10⁻⁶ M, 10⁻⁵ M, and 10⁻⁴ M) in culture medium were prepared. Control cultures were treated with DMSO alone. Cells were plated in 96-well plates (10⁴ cells per well) for 24 hours before treatment with the compounds. Different concentrations of the test compounds $(10^{-7}$ M, 10^{-6} M, 10^{-5} M, and 10^{-4} M) were added to the cell monolayer. Triplicate wells were prepared for each individual dose. Monolayer cells were then incubated with test compounds for 48 hours at 37°C in an atmosphere of 5% CO_2 . After 48 hours, cells were fixed, washed, and stained with sulforhodamine B stain. Excess stain was washed with acetic acid and the attached stain was recovered with Tris-EDTA buffer. Color intensity was measured using an enzyme-linked immunosorbent assay reader. The relation between surviving fraction and drug concentration was plotted and the $GI₅₀$ was calculated for each compound.

Antiepileptic activity

The synthesized compounds (6a–q) obtained from the reaction sequence were dissolved in polyethyleneglycol (PEG-400) administered intraperitoneally into male albino mice (CF-1 strain, $18-25$ g) and tested in the MES,²⁴ PTZ seizure,²⁵ and neurotoxicity screens,²⁶ using 30 mg/kg, 100 mg/kg, and 300 mg/kg doses; and observations were recorded at two different time intervals. All experimental protocols were carried out with permission from the Institutional Animal Ethics Committee (IAEC), Birla Institute of Technology, Mesra, Ranchi (CPCSEA approval no: BIT/PH/IAEC/13/2014, dated April 30, 2014), and the experiments were in compliance with the animal care guidelines issued by the IAEC.

The MES test

Albino mice were divided into groups of six animals each. Maximal seizures were induced by the application of alternating current of 60 Hz (50 mA) for 0.2 seconds using corneal electrodes primed with an electrolyte solution containing an anesthetic agent (0.5% tetracaine HCl). During the preliminary screening, the test compounds were administered by intraperitoneal injection at 0.01 mL/kg body weight at doses of 30 mg/kg, 100 mg/kg, and 300 mg/kg prior to testing, and anticonvulsant activity was evaluated after 0.5 hours and 4 hours of administration. Abolition of the hind limb tonic extensor spasm was recorded as anticonvulsant activity in the MES method.²⁷

The PTZ seizure test

The synthesized compounds were administered to mice (n=6) by intraperitoneal injection. After 30 minutes, mice were treated with a subcutaneous injection of PTZ (60 mg/kg). Then, the mice were placed singly in isolated plastic cages and observed for 60 minutes. Protection was referred to as the failure to detect an episode of clonic spasms of at least 5-second duration during this time period.

Neurotoxicity screening

Neurotoxicity was measured by the rotarod test. Mice (n=6) were given intraperitoneal injection of the synthesized compounds at dose levels of 30 mg/kg, 100 mg/kg, and 300 mg/kg. Then, after 30 minutes, the mice were placed on the rotating rod. Neurotoxicity was determined by the inability of the mice to maintain equilibrium on the rod for 1 minute in each of the three trials.16

Results and discussion Chemistry

The chroman derivatives (6a–q) described in this study were prepared as depicted in Figure 2. The synthesis of

Figure 2 Synthesis pathway for compounds 6a–q. <code>Notes:</code> Reactions a and b carried out under the following conditions: NH₂NH₂H₂O, C₂H₃OH, reflux, CP₃COOH, CH₃COOH, reflux, 4–12 hours.

6-substituted-2,5,7,8-tetramethyl-chroman-2-carboxylic acid methyl ester (3a–c) as a key intermediate was accomplished by the reaction of trimethylhydroquinone and methyl methacrylate to produce $3a$,²² followed by methylation (yielding $3b)^{28}$ and benzylation (to yield $3c$).¹¹ The ester moieties of 3a–c endured the nucleophilic attack by hydrazine hydrate, resulting in $4a-c$.²⁹ The latter compounds were subjected to condensation reaction with different substituted isatins to produce 6a–q with 51% –94% yield.²¹ The properties of the synthesized compounds 6a–q are shown in Table 1.

The compounds were characterized by Fourier transform infrared spectroscopy (FTIR), ¹H-nuclear magnetic resonance (NMR), 13C-NMR, mass spectroscopy, and melting point analysis. The analytical data were fully consistent with the proposed structures. The retention factor (Rf) value of all synthesized compounds (6a–q) was calculated in ethyl acetate (EtOAc) and hexane (40:60) and ranged between 0.4 and 0.7.

In the FTIR spectrum, bands at $3,284-3,598$ cm⁻¹ and $1,600-1,700$ cm⁻¹ confirm the presence of $-NH$ and $-CO$

Compound ^{a,b}	$\mathbf R$	R ¹	R ²	Time (hours)	Yield $(\%)^c$	Melting point $(^{\circ}C)$	Molecular formula	Molecular weight	Rf ^d
6b	Н	н	CH,	6	72	237	$C_{23}H_{25}N_{3}O_{4}$	407.46	0.7
6с	H	5-CI	н	6	87	255	$C_{22}H_{22}N_{3}O_{4}Cl$	427.88	0.6
6d	H	5-Br	н	6	56	260	$C_{22}H_{22}N_{3}O_{4}Br$	472.33	0.4
6e	н	$6-CI$	H	5	57	290	$C_{22}H_{22}N_{3}O_{4}Cl$	427.88	0.5
6f	H	$5-F$	H	4	51	290	$C_{22}H_{22}N_{3}O_{4}F$	411.43	0.4
6g	н	$5-CI$	CH,	5	59	257	$C_{23}H_{24}N_{3}O_{4}Cl$	441.91	0.5
6h	CH,	H.	н	3	58	256	$C_{23}H_{25}N_{3}O_{4}$	407.46	0.6
6i	CH,	5-Br	H	5	51	290	$C_{23}H_{24}N_{3}O_{4}Br$	486.36	0.6
6j	CH,	5-F	н	5	66	$<$ 300	$C_{23}H_{24}N_{3}O_{4}F$	425.45	0.5
6k	CH,	$6-C$	н	5	52	270	$C_{23}H_{24}N_{3}O_{4}Cl$	441.91	0.5
6 ¹	$C_{6}H_{5}$	H	н	12	74	258	$C_{29}H_{29}N_{3}O_{4}$	483.56	0.7
6 _m	$C_{6}H_{5}$	н	CH,	12	92	170	$C_{31}H_{33}N_{3}O_{4}$	511.61	0.6
6n	$C_{6}H_{5}$	5-Br	н	12	68	270	$C_{29}H_{28}N_{3}O_{4}Br$	562.45	0.6
60	$C_{6}H_{5}$	5-F	н	12	85	$<$ 300	$C_{29}H_{28}N_{3}O_{4}F$	501.55	0.6
6 _p	$C_{6}H_{5}$	5-CI	CH,	12	81	220	$C_{30}H_{30}N_{3}O_{4}Cl$	532.03	0.7
6q	$C_{6}H_{5}$	5-CI	н	12	94	280	$C_{29}H_{28}N_{3}O_{4}Cl$	518.00	0.6

Table 1 Some properties of the compounds 6a–q

Notes: ^aReagents and conditions: 5a-q (0.001 mol), 4a-c (0.001 mol), and acetic acid (10 mL). ^bRatio of the product was determined by the analysis of crude FTIR, ¹ Notes: ªReagents and conditions: 5a–q (0.001 mol), 4a–c (0.001 mol), and acetic acid (10 mL). ^bRatio of the product was determined by the analysis of crude FTIR, 'H-NMR,
¹³C-NMR, and mass spectra. Yield refers to pure halogen; and R^2 = hydrogen and methyl.

Abbreviations: EtOAc, ethyl acetate; FTIR, Fourier transform infrared spectroscopy; NMR, nuclear magnetic resonance; Rf, retention factor.

Table 2 Anti-breast cancer and antiepileptic activities of the compounds 6a–q

Compound $GI_{50} (\mu M)^a$		MES		PTZ ^b	
		0.5 hours	4 hours	0.5 hours	4 hours
6a	37.1			300	300
6b	>100	30	30	100	100
6с	76.4	30	30	100	300
6d	57.3			30	30
6е	46.9	100	300	30	30
6f	62.7			300	300
6g	>100	30	30	100	100
6h	45.1	300	300	100	100
6i	34.7			30	100
6j	>100	100	100	100	300
6k	38.2	100	300	100	100
61	45.9			30	30
6 _m	> 100	100	100	100	100
6n	76.6			100	100
60	64.1	300	300	300	300
6 _p	77.9	100	100	100	100
6q	> 100	100	100	100	100
Adriamycin	$<$ 0.1				
Phenytoin		30	30		
Sodium valproate				200	200

Notes: ${}^{3}G_{50}$ is the concentration of drug causing 50% inhibition of cell growth.
bDoses of 30 mg/kg 100 mg/kg and 300 mg/kg were administered. The figure in the **bDoses of 30 mg/kg, 100 mg/kg, and 300 mg/kg were administered. The figure in the** table indicates the minimum dose required for bioactivity in half or more of the mice examined at 0.5 hours and 4 hours. Dash indicates the absence of anticonvulsant activity and neurotoxicity at the maximum dose administered (300 mg/kg). **Abbreviations:** MES, maximal electroshock seizure; PTZ, pentylenetetrazole.

stretching vibrations, respectively. Compounds 6a–g showed broad absorption bands at $3,141-3,406$ cm⁻¹, indicative of a hydroxy group (-OH stretch). The ¹H-NMR spectrum revealed a singlet at δ =11.19–11.32 ppm due to NH₂ protons (- CONH_2). The presence of a hydroxy group (-OH) in the compounds 6a–g was confirmed by the sharp singlet at δ =7.52–7.55 ppm. The methoxy group (–OCH₃) showed a singlet at $\delta = 3.48 - 3.49$ ppm in compounds 6h–k. Compounds 6l–q displayed a singlet peak at δ =4.59–4.67 ppm, corresponding to benzylic methyl $(-CH₂-)$. In ¹³C-NMR, the carbonyl group of amide (C=O) was observed at approximately δ =160–172 ppm. The mass spectra showed (M+H)⁺ and $(M+Na^{+})^{+}$ (m/z) peaks, respectively confirming their purity and molecular weight.

In vitro anticancer activity screening

The synthesized compounds (6a–q) were screened for in vitro anticancer activity against human mammary adenocarcinoma $(MCF-7)$ by using sulforhodamine B assay.²³ The concentration required for 50% inhibition of cell growth $\left(\frac{GI}{50}\right)$ for each compound was calculated with reference to a control sample.

For each compound, $GI₅₀$ was calculated from the sigmoidal dose–response curves and the values are presented in Table 2. The data for adriamycin are included as reference.

The resultant data showed that the chroman derivatives (6a-q) produced significant cytostatic effects against the human breast cancer MCF-7 cell line. The results revealed that compound 6i was the most active member of the series, with GI_{50} =34.7 µM. Among all the compounds, compounds 6a, 6i, and 6k showed promising activity, with GI_{so} values of 37.1 µM, 34.7 µM, and 38.2 µM, respectively; compounds 6d, 6e, 6h, and 6l showed $GI₅₀$ values of 57.3 µM, 46.9 µM, 45.1 µM, and 45.9 µM, respectively; compounds 6c, 6f, 6n, 6o, and 6p showed $GI₅₀$ values of 76.4 μ M, 62.7 µM, 76.6 µM, 64.1 µM, and 77.9 µM, respectively; the remaining five compounds 6b, 6g, 6j, 6m, and 6q showed $GI₅₀$ values $>100 \mu M$ in the MCF-7 breast cancer cells. The activity was attributed to the presence of chlorine or bromine group at the fifth position on the phenyl ring of the isatin ring system, whereas in the presence of fluorine, the phenyl group of the isatin ring system induced reduced activity. However, the chlorine group at the sixth position accounts for more activity than the chlorine group at the fifth position. Benzyl substitution in the chroman ring system appeared to be less active than methoxy and hydroxyl substitution.

Antiepileptic activity

The antiepileptic activity and the acute neurotoxicity of all synthesized compounds (6a–q) were evaluated by the use of standard techniques.24–26 Results are presented in Table 2. Phenytoin and carbamazepine were used as the standard drugs for the comparisons.

The most potent compounds 6b, 6c, and 6g resembled the standard drug phenytoin at the dose level of 30 mg/kg after 0.5 hours and 4 hours by the maximal electroshock seizure (MES) method. On the other hand, compounds 6j, 6m, 6p, and 6q displayed activity at the dose level of 100 mg/kg against seizure spread at both the time intervals, whereas compounds 6e and 6k displayed protection at the dose level of 300 mg/kg after 4 hours, suggesting rapid onset but shorter duration of action. Compounds 6h and 6o exhibited protection at both the time intervals 0.5 hours and 4 hours after the dose level of 300 mg/kg.

In the pentylenetetrazole (PTZ) screen, the most active of these compounds were 6d, 6e, and 6l, which provided protection at the dose level of 30 mg/kg after both the time intervals 0.5 hours and 4 hours, excluding compound 6i, which showed protection at the 100 mg/kg dose level after 4 hours. Compounds 6b, 6g, 6h, 6k, 6m, 6n, 6p, and 6q exhibited protection

at the 100 mg/kg dose level after both time intervals, whereas compounds 6c and 6j showed protection at 100 mg/kg after 4 hours. The remaining compounds had activity at the dose level of 300 mg/kg at both the time intervals.

Compounds that afforded protection against seizures in the MES and PTZ screens were further tested for their neurotoxicity. All compounds (6a–q) were devoid of neurotoxicity, as determined by the rotarod test.

Gamma-aminobutyric acid (GABA) is the principal inhibitory neurotransmitter substance in the brain and it is widely implicated in epilepsy. It has been reported that PTZ produces seizures by inhibiting GABA neurotransmission. In order to inhibit seizures, enhancement of GABAergic neurotransmission is useful. All newly synthesized compounds (6a–q) showed protection against seizure in the PTZ model (Table 2). These findings suggest that the compounds may have inhibited or attenuated PTZ-induced seizure in mice by enhancing GABAergic neurotransmission.30

The antiepileptic activity correlation of the synthesized compounds (6a–q) revealed that compounds containing a hydroxyl group in the chroman ring system, such as 6b, 6c, 6d, 6e, and 6g, were more potent as compared to the compounds containing a methyl group (6h, 6i, and 6k). Compounds having a benzyl group, such as 6l, 6n, and 6o, in the chroman ring system were the least active. The presence of electron-withdrawing substituents (Cl, Br, and F) in the isatin ring system might have increased the activity, as compounds 6c, 6g, 6j, 6m, and 6q were more active than compounds 6a, 6h, and 6l. However, compounds, such as 6f, 6j, and 6o, which have a fluorine substituent, were found to be less active than compounds with the other electron-withdrawing substituents (Cl and Br).

Conclusion

In this study, the design and synthesis of a series of chroman derivatives have been described. All synthesized compounds were evaluated for their anti-breast cancer and antiepileptic activities. The results of the study demonstrated that compound 6i exerted the most potent activity ($GI₅₀=34.7 \mu M$) among the compounds in the series. Additionally, all synthesized compounds were evaluated for their antiepileptic efficacy. In the antiepileptic evaluation, compounds 6b–e, 6g, 6i, and 6l were found to be more active than the reference drugs. This work adds new data to the relationship between chromans and their anti-breast cancer and antiepileptic activities. This provides a platform for further development of related compounds into more active compounds.

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Disclosure

The authors report no conflicts of interest in this work.

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