

Sperm DNA fragmentation: laboratory and clinical aspects

Ahmad Majzoub¹, Ashok Agarwal², Sandro C. Esteves³

¹Department of Urology, Hamad Medical Corporation, Doha, Qatar; ²American Center for Reproductive Medicine, Cleveland Clinic, Cleveland, OH, USA; ³ANDROFERT, Andrology and Human Reproduction Clinic, Referral Center for Male Reproduction, Campinas, SP, Brazil

Correspondence to: Ahmad Majzoub. Associate Consultant, Department of Urology, Hamad Medical Corporation, Doha, Po Box 3050, Qatar. Email: dr.amajzoub@gmail.com.

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The commentary by Drs. Khalili and Agha-Rahimi on the sperm DNA fragmentation (SDF) clinical guideline article by Agarwal *et al.* (1) recognized the presence of a negative association between high seminal SDF levels and fertility outcome specifically with assisted reproductive techniques (ART). Moreover, they have raised important concerns related to the lack of standardized SDF cutoff values and the influence of sperm preparation techniques during the course of ART on SDF levels. Furthermore, they brought into question, the exact treatment strategies that can be used for patients with high SDF. We certainly agree with these concerns which deserve further elaboration as seen below.

Many of the sperm preparation techniques that are performed during semen handling may indeed impose a risk of sperm DNA damage. This risk is believed to be mediated by the state of oxidative stress (OS) that occurs secondary to the exposure of sperm to exaggerated levels of reactive oxygen species (ROS) during sperm preparation. An understanding of the exogenous sources of ROS that take place during the course of ART is hence vital in order to prevent the development of SDF. Variables such as atmospheric oxygen concentration, illumination, centrifugation force, pH and temperature, culture media and cryopreservation are all implicated (2). Drs. Khalili and Agha-Rahimi have particularly tackled the influence of incubation time/temperature and culture media on SDF levels. It is believed that long incubation periods can influence sperm quality, measures of OS and SDF (3,4). It is therefore recommended that the time between sample production and the start of preparation is kept as

short as possible and begin immediately after liquefaction. Incubation temperature is an important variable which could also influence sperm quality with increasing temperatures eliciting ROS-induced cellular damage (5). Recent studies have demonstrated an improvement in sperm quality (6) and a reduction in SDF measures (7) with samples stored at room temperature (23–25 °C) in comparison to those stored at body temperature (37 °C). Lastly, the sperm preparation medium used prior to ART may influence the quality of sperm and consequently the overall treatment outcome (8). The presence of metallic ions in culture media can potentiate increased ROS generation (9,10). Therefore, the use of antioxidants in culture media has been investigated and found to carry favorable effects on sperm quality (11).

While enquiring about treatment options available for patients with high SDF levels, Drs. Khalili and Agha-Rahimi have offered the implementation of lifestyle changes, use of sperm selection methods or testicular sperm for intracytoplasmic sperm injection (ICSI). Indeed, we have highlighted in the clinical guideline article (1), the importance of SDF testing in patients exposed to pollutants or who have modifiable lifestyle risk factors recommending it to reinforce these changes, predict fertility and as a monitoring tool for patient's response to intervention. Furthermore, patients with high SDF levels could also benefit from treatment with antioxidant supplements (12). Antioxidants have been thoroughly investigated in the treatment of male infertility. While a number of studies have confirmed a beneficial effect for antioxidants in reversing OS induced sperm dysfunction, fewer reports did investigate the effects of dietary antioxidant intake on

sperm DNA integrity (13-16). Specifically, vitamins C and E, beta-carotene, zinc, selenium and folic acid were found to be significantly helpful in this regard (14,15,17). One systemic review of 20 trials examining the effect of oral antioxidants on measures of sperm OS and SDF revealed that a significant reduction in these measures was reported in 19 out of the 20 studies selected. (18).

A variety of interventions have been proposed for patients with high SDF undergoing ART. Shortening of the abstinence time with frequent ejaculations (19,20), sperm selection using density centrifugation combined with swim-up technique (7,21), magnetic-activated cell sorting (MACS), physiological ICSI (PICSI) or intracytoplasmic morphologically selected sperm (IMSI) (22,23) and the use of testicular sperm have all been documented (24,25). Nevertheless, studies comparing the efficacy of these techniques in selecting sperm with better DNA integrity for ICSI are sparse. One retrospective analysis by Bradley *et al.* (26) investigated ICSI outcomes of 1,924 infertile men comparing results of patients with low SDF to those with high SDF who did or did not receive an intervention such as PICSI, IMSI or testicular sperm extraction/aspiration. In general, patients with high SDF who did not have an intervention had lower fertilization and poorer clinical outcomes compared with low SDF patients. In contrast, patients with high SDF who had an intervention had significantly improved clinical outcomes, similar to those with low SDF. Comparing the three interventions, the authors revealed that the highest live birth rate was detected in the testicular sperm subgroup (49.8%), followed by PICSI (38.3%) and IMSI (28.7%).

Finally, we agree that the presence of different SDF testing methods with lack of standardized cutoff values may still impose an obstacle to its routine use in clinical practice. Recent reports have raised this issue advising andrology laboratories to evaluate the type of SDF test they use and determine their own threshold values (27). Nevertheless, it is arguably rational to suggest that with the intended purpose of the clinical guideline article (1) to identify the proper indications of SDF testing, meaningful refinements of its practical methods and diagnostic thresholds could be achieved.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

References

1. Agarwal A, Majzoub A, Esteves SC, et al. Clinical utility of sperm DNA fragmentation testing: practice recommendations based on clinical scenarios. *Transl Androl Urol* 2016;5:935-50.
2. Agarwal A, Majzoub A. Role of Antioxidants in Assisted Reproductive Techniques. *World J Mens Health* 2017;35:77-93.
3. Nabi A, Khalili MA, Halvaei I, et al. Prolonged incubation of processed human spermatozoa will increase DNA fragmentation. *Andrologia* 2014;46:374-9.
4. Banihani S, Sharma R, Bayachou M, et al. Human sperm DNA oxidation, motility and viability in the presence of L-carnitine during in vitro incubation and centrifugation. *Andrologia* 2012;44 Suppl 1:505-12.
5. Larkindale J, Knight MR. Protection against heat stress-induced oxidative damage in Arabidopsis involves calcium, abscisic acid, ethylene, and salicylic acid. *Plant Physiol* 2002;128:682-95.
6. Thijssen A, Klerkx E, Huyser C, et al. Influence of temperature and sperm preparation on the quality of spermatozoa. *Reprod Biomed Online* 2014;28:436-42.
7. Matsuura R, Takeuchi T, Yoshida A. Preparation and incubation conditions affect the DNA integrity of ejaculated human spermatozoa. *Asian J Androl* 2010;12:753-9.
8. Mortimer D. Sperm preparation techniques and iatrogenic failures of in-vitro fertilization. *Hum Reprod* 1991;6:173-6.
9. Guerin P, El Mouatassim S, Menezo Y. Oxidative stress and protection against reactive oxygen species in the pre-implantation embryo and its surroundings. *Hum Reprod Update* 2001;7:175-89.
10. Kehrer JP. The Haber-Weiss reaction and mechanisms of toxicity. *Toxicology* 2000;149:43-50.
11. Agarwal A, Durairajanayagam D, du Plessis SS. Utility of antioxidants during assisted reproductive techniques: an evidence based review. *Reprod Biol Endocrinol* 2014;12:112.
12. Agarwal A, Majzoub A. Role of antioxidants in male infertility. *BJUI Knowledge* 2016. [Epub ahead of print].

13. Greco E, Iacobelli M, Rienzi L, et al. Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment. *J Androl* 2005;26:349-53.
14. Menezo YJ, Hazout A, Panteix G, et al. Antioxidants to reduce sperm DNA fragmentation: an unexpected adverse effect. *Reprod Biomed Online* 2007;14:418-21.
15. Tremellen K, Miari G, Froiland D, et al. A randomised control trial examining the effect of an antioxidant (Menevit) on pregnancy outcome during IVF-ICSI treatment. *Aust N Z J Obstet Gynaecol* 2007;47:216-21.
16. Gil-Villa AM, Cardona-Maya W, Agarwal A, et al. Role of male factor in early recurrent embryo loss: do antioxidants have any effect? *Fertil Steril* 2009;92:565-71.
17. Omu AE, Al-Azemi MK, Kehinde EO, et al. Indications of the mechanisms involved in improved sperm parameters by zinc therapy. *Med Princ Pract* 2008;17:108-16.
18. Gharagozloo P, Aitken RJ. The role of sperm oxidative stress in male infertility and the significance of oral antioxidant therapy. *Hum Reprod* 2011;26:1628-40.
19. Gosalvez J, Gonzalez-Martinez M, Lopez-Fernandez C, et al. Shorter abstinence decreases sperm deoxyribonucleic acid fragmentation in ejaculate. *Fertil Steril* 2011;96:1083-6.
20. Agarwal A, Gupta S, Du Plessis S, et al. Abstinence Time and Its Impact on Basic and Advanced Semen Parameters. *Urology* 2016;94:102-10.
21. Xue X, Wang WS, Shi JZ, et al. Efficacy of swim-up versus density gradient centrifugation in improving sperm deformity rate and DNA fragmentation index in semen samples from teratozoospermic patients. *J Assist Reprod Genet* 2014;31:1161-6.
22. Lee TH, Liu CH, Shih YT, et al. Magnetic-activated cell sorting for sperm preparation reduces spermatozoa with apoptotic markers and improves the acrosome reaction in couples with unexplained infertility. *Hum Reprod* 2010;25:839-46.
23. Hammoud I, Boitrelle F, Ferfourri F, et al. Selection of normal spermatozoa with a vacuole-free head (x6300) improves selection of spermatozoa with intact DNA in patients with high sperm DNA fragmentation rates. *Andrologia* 2013;45:163-70.
24. Esteves SC, Sanchez-Martin F, Sanchez-Martin P, et al. Comparison of reproductive outcome in oligozoospermic men with high sperm DNA fragmentation undergoing intracytoplasmic sperm injection with ejaculated and testicular sperm. *Fertil Steril* 2015;104:1398-405.
25. Arafa M, Elbardisi H, AlMalki A, et al. ICSI outcome in patients with high DNA fragmentation: testicular Vs ejaculated sperm. *Andrologia* 2017. [Epub ahead of print].
26. Bradley CK, McArthur SJ, Gee AJ, et al. Intervention improves assisted conception intracytoplasmic sperm injection outcomes for patients with high levels of sperm DNA fragmentation: a retrospective analysis. *Andrology* 2016;4:903-10.
27. Vandekerckhove FW, De Croo I, Gerris J, et al. Sperm Chromatin Dispersion Test before Sperm Preparation Is Predictive of Clinical Pregnancy in Cases of Unexplained Infertility Treated with Intrauterine Insemination and Induction with Clomiphene Citrate. *Front Med (Lausanne)* 2016;3:63.

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