

# Role of Bile Acid Receptors in the Development and Function of Diabetic Nephropathy

Check for updates

Yuanyuan Fang<sup>[1](#page-0-0),[2](#page-0-1)</sup>, Minjing Qin<sup>[1,](#page-0-0)2</sup>, Qitong Zheng<sup>1</sup>, Kuilong Wang<sup>1</sup>, Xin Han<sup>1</sup>, Qiao Yang<sup>1</sup>, Xia'nan Sang $<sup>1</sup>$  $<sup>1</sup>$  $<sup>1</sup>$  and Gang Cao<sup>1</sup></sup>

<span id="page-0-0"></span><sup>1</sup>School of Pharmacy, Zhejiang Chinese Medical University, Hangzhou, China

Diabetic nephropathy (DN) is a prevalent microvascular complication that occurs often in individuals with diabetes. It significantly raises the mortality rate of affected patients. Therefore, there is an urgent need to identify therapeutic targets for controlling and preventing the occurrence and development of DN. Bile acids (BAs) are now recognized as intricate metabolic integrators and signaling molecules. The activation of BAs has great promise as a therapeutic approach for preventing DN, renal damage caused by obesity, and nephrosclerosis. The nuclear receptors (NRs), farnesoid X receptor (FXR), pregnane X receptor (PXR), vitamin D receptor (VDR); and the G protein-coupled BA receptor, Takeda G-protein-coupled receptor 5 (TGR5) have important functions in controlling lipid, glucose, and energy metabolism, inflammation, as well as drug metabolism and detoxification. Over the past 10 years, there has been advancement in comprehending the biology and processes of BA receptors in the kidney, as well as in the creation of targeted BA receptor agonists. In this review, we discuss the role of BA receptors, FXR, PXR, VDR, and TGR5 in DN and their role in renal physiology, as well as the development and application of agonists that activate BA receptors for the treatment of kidney diseases.

Kidney Int Rep (2024) 9, 3116–3133; <https://doi.org/10.1016/j.ekir.2024.08.002> KEYWORDS: bile acids; diabetic nephropathy; membrane receptor TGR5; nuclear receptor FXR PXR VDR ª 2024 International Society of Nephrology. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license [\(http://creativecommons.org/licenses/by-nc-nd/4.0/](http://creativecommons.org/licenses/by-nc-nd/4.0/)).

he increase in sugar and fat-rich diets due to economic progress and improved quality of life has led to a rise in obesity and diabetes mellitus, a signifi-cant global public health concern.<sup>[1](#page-12-0)</sup> The International Diabetes Federation predicts that diabetes mellitus will reach 10.5% in [2](#page-12-1)021 and 12.2% by 2045.<sup>2</sup> In addition, 30% to 40% of individuals with diabetes will develop  $DN$ , a prevalent microvascular complication, contributing to end-stage renal disease. $4.5$  $4.5$  DN is characterized by structural alterations in the kidney, such as glomerulosclerosis, tubule interstitial fibrosis, thickening of the glomerular basement membrane, loss of podocytes, glomerular hypertrophy, and expansion of the mesangial matrix. The pathogenesis and progression of DN involve several key factors, including hypertension, abnormal carbohydrate and fat

metabolism, lipid accumulation, increased glycation end product, increased oxidative stress, and upregula-tion of profibrotic growth factors.<sup>[6](#page-12-5)</sup> The development of albuminuria and a decrease in glomerular filtration rate are the results of these alterations taken together.<sup>7</sup>

The kidney is a crucial organ in the context of  $DN$ , $\degree$ and a potential therapeutic strategy for kidney illness could involve targeting the expression and activation of BA receptors. BA receptors play a vital role in renal physiology and disease,<sup>[9](#page-12-8)</sup> activating various BA receptors such as FXR, G-protein-coupled receptor-1 / TGR5, PXR, and VDR. $10,11$  $10,11$  The kidney shows a high expression level of FXR and TGR5 and their target genes. BAs also regulate metabolism by activating NRs and G-protein-coupled receptor signaling pathways,<sup>12,[13](#page-12-12)</sup> regulating kidney lipid, glucose, and energy balance. <sup>14-16</sup> Disruptions in BA metabolism can lead to cholestatic liver disorders, dyslipidemia, fatty liver illnesses, cardiovascular diseases, and diabetes. $17$  Activating these receptors has great potential for treating acute and chronic kidney illnesses by reducing lipid buildup, inflammation, oxidative stress, and fibrosis in the kidneys.<sup>18</sup>

In recent times, the significance of BAs in renal pathophysiology has grown due to their activation of

Correspondence: Xia'nan Sang or Gang Cao, School of Pharmacy, Zhejiang Chinese Medical University, 548 Binwen Road, Hangzhou, 310053, China. E-mail: [xnsang881014@163.com](mailto:xnsang881014@163.com) or [caogang33@163.com](mailto:caogang33@163.com)

<span id="page-0-1"></span><sup>&</sup>lt;sup>2</sup>YYF and MJQ are the primary writers of this manuscript

Received 27 March 2024; revised 25 July 2024; accepted 4 August 2024; published online 10 August 2024

FXR and TGR5, as well as transcription factors associated with lipid, cholesterol, and glucose metabolism. In addition, BAs have been found to influence genes implicated in inflammation and renal fibrosis. $19,20$  $19,20$ Therefore, the activation of BA receptors potentially contribute to the therapeutic management of renal disease. This study aims to examine the existing knowledge on the role and functionality of BAs and BA receptors, namely FXR and TGR5, in the context of kidney disease. Specifically, we will emphasize their involvement in diabetic kidney-related inflammation, lipid buildup, and fibrosis.

Furthermore, this review examines different agents that activate BA receptors and their impact on DN upon activation. It also discusses the current utilization and prospects of these agents, offering novel targets and therapeutic approaches for the clinical management and successful regulation of DN.

### DN

DN is a common microvascular consequence in individuals with diabetes, leading to kidney cell dysfunction and renal failure.<sup>[21-25](#page-12-18)</sup> It is the primary contributor to end-stage nephropathy<sup>[26](#page-12-19)</sup> and is responsible for most cases of chronic and end-stage renal disease globally.[27,](#page-12-20)[28](#page-12-21) DN represents a renal manifestation of the same glucose-driven pathological process, $29,30$  $29,30$  which also occurs in other susceptible areas of the body. $31$ 

The pathological characteristics of DN encompass many changes, such as the thickening of basement membranes in glomeruli and tubules, mesangial dilatation, interstitial inflammation, hypertrophy of glomeruli and tubules, glomerular sclerosis, and fibrosis in the tubule interstitium<sup>[28](#page-12-21)[,32](#page-13-1)</sup> [\(Figure 1](#page-1-0)). During the initial phase of DN, there is a presence of subtle pathological alterations, and the manifestation of clinical symptoms is challenging to identify. The observed manifestations mostly consist of glomerular mesangial hyperplasia, basal membrane thickening, and glomerular sclerosis.<sup>33</sup> As the disease advances, glomerular nodular degeneration undergoes a slow transition into diffuse lesions, encompassing interstitial fibrosis, renal tubule atrophy, and hyaline degeneration of the arterioles within and outside the bulb. These lesions exhibit a limited capacity for reversal. Ultimately, they advance to the terminal stage of renal disease, occurring approximately 20 to 25

<span id="page-1-0"></span>

Figure 1. Pathological lesions of DN. The normal healthy glomerulus includes afferent arterioles, capillary loops, endothelial cells, basement membrane, podocytes, parietal epithelial cells, and tubule epithelial cells and is impermeable to albumin. In contrast, the diabetic glomerulus displays arterial hyalinosis, mesangial expansion, collagen deposition, basement membrane thickening, podocyte loss and hypertrophy, albuminuria, tubular epithelial atrophy, accumulation of activated myofibroblasts and matrix, influx of inflammatory cells, and capillary rarefaction. DN, diabetic nephropathy.

years after the first signs of diabetes. This stage is characterized by reduced glomerular filtration rate, arterial hypertension, low serum renin levels, decreased protein excretion, and gradual renal function decline.<sup>[34](#page-13-3)</sup> The etiology of DN is intricate and encompasses the activation of various pathways that result in renal damage. These pathways include the polyol pathways, advanced glycosylation end products, oxidative stress, proinflammatory cytokines, and profibrotic growth factors. The dysregulation of the signaling cascade is a consequence of these pathways, ultimately leading to the development of DN.

Strict glycemic control, blood pressure management, and the use of renin-angiotensin system (RAS) blockers have all been shown in clinical studies to prevent the development and progression of diabetic neuropathy. Regrettably, a sizable portion of patients continue to be uncontrolled. Therefore, we need novel strategies for better therapies.

#### BA ANABOLISM AND ITS RECEPTORS

BAs, a class of steroid acids, are crucial signaling molecules that facilitate precise intratissue communication. They are primarily synthesized in the liver and undergo metabolic transformations in the intestine by the gut microbiota. $35$  These transformations include deconjugation, dehydroxylation, oxidation, epimerization, and reconjugation.  $36,37$  $36,37$  BAs play a vital role in maintaining a healthy gut microbiome, regulating lipid and carbohydrate metabolism, enhancing insulin sensitivity, and supporting innate immunity.<sup>3</sup>

The composition of the human BA pool comprises the principal Bas, cholic acid (CA) and chenodeoxycholic acid (CDCA), along with the secondary BA, deoxycholic acid and a minimal quantity of lithocholic acid. The synthesis of primary BAs occurs in the liver through the conversion of cholesterol. Enzymatic processes facilitated by gut bacteria lead to the conversion of certain primary BAs into secondary BAs. Firmicutes, Bacteroides, Lactobacillus, Bifidobacterium, and Clostridium all contribute significantly to the production of secondary BAs. $^{39}$  $^{39}$  $^{39}$  Within the hepatic organ, the synthesis of primary BAs occurs via 2 prominent pathways: the conventional pathway (often referred to as the neutral pathway) and the alternate pathway (known as the acidic pathway owing to the generation of acidic intermediates). The classical pathway involves the conversion of cholesterol into the primary BAs, CA and CDCA through the actions of cholesterol 7a-hydroxylase and sterol 12a-hydroxylase. In contrast, the alternative pathway is initiated by sterol 27 hydroxylase and primarily results in the production of CDCA.<sup>[40](#page-13-9)</sup> In humans, the primary BAs consist of CA

and CDCA; however, in mice, the primary BAs are CA, CDCA, ursodeoxycholic acid (UDCA), and muricholic acid (MCA). $41$  Within the gastrointestinal tract, the process of deconjugation occurs, whereby conjugated CA and CDCA are enzymatically modified by the bile salt hydrolase (BSH) enzyme produced by gut bacteria. Subsequently, the deconjugated CA and CDCA undergo a transformation mediated by the 7a-dehydroxylase enzyme in humans, resulting in the production of secondary BAs such as deoxycholic acid, lithocholic acid, and ursodeoxycholic acid. $42,43$  $42,43$  ([Figure 2](#page-3-0)).

Abnormalities in the gut flora can cause metabolic abnormalities in people with  $DN.^{39,44,45}$  $DN.^{39,44,45}$  $DN.^{39,44,45}$  $DN.^{39,44,45}$  $DN.^{39,44,45}$  Metabolites produced by gut microorganisms facilitate many interactions between the gut microbiota and the human body are facilitated by metabolites produced by gut microorganisms. Gut microorganisms' metabolic products mostly consist of SCFAs (formic acid, acetic acid, propionic acid, isobutyric acid, butyric acid, isovaleric acid, and valeric acid), BAs, TMAO (a metabolite of choline, L-carnitine), and betaine, uremic toxins, and hydrogen sulfide.<sup>46[,47](#page-13-16)</sup> These metabolites alter the intestinal epithelium's barrier function by modulating receptor expression or activating transcription factors, playing an important role in the onset and progression of DN. This study focuses on the relationship between DN and receptors for metabolites of gut bacteria, BAs.

BA ligands are endogenous ligands that activate NRs and a G-protein-coupled receptor, allowing them to modulate metabolic pathways in various tissues.<sup>[48](#page-13-17)</sup> They can act as agonists or antagonists of signaling molecules on FXR and G protein-coupled BA receptor  $1.^{10,48}$  $1.^{10,48}$  $1.^{10,48}$  $1.^{10,48}$  Abnormal BA metabolism and receptor expression have been linked to liver injury, $49$  metabolic disorders, cardiovascular diseases, digestive system diseases,  $50$  colorectal cancer,  $51$ and kidney disease. $52,53$  $52,53$  Using synthetic and natural receptor agonists or antagonists has shown potential in managing metabolic disturbances and inflammation.

# BA Receptors in DN

BA receptors, FXR, PXR, VDR, and TGR5 play an important role in the pathophysiology of the kidney. BA receptors are important targets for the treatment and control of DN. Many studies have shown the importance of BA receptors in DN.

# The Function of NRs in the Kidney

NRs are crucial regulators of systemic homeostasis and play a significant role in various illnesses. They regulate various metabolic processes, including renal lipid metabolism, medication clearance, inflammation, fibrosis, cell differentiation, and oxidative stress. Dysregulation of NRs, often caused by obesity, can lead to metabolic syndrome, end-stage renal disease, and

<span id="page-3-0"></span>

Figure 2. Bile acid synthesis and bile acids receptors. In the classic bile acid synthesis pathway, cholesterol is converted to cholic acid (CA, 3a, <sup>7</sup>a, 12a) and chenodeoxycholic acid (CDCA, 3a, 7a) CYP7A1 is the rate-limiting enzyme and CYP8B1 catalyzes the synthesis of CA. In mouse liver, CDCA is converted to  $\alpha$ -muricholic acid ( $\alpha$ -MCA, 3 $\alpha$ , 6 $\beta$ , 7 $\alpha$ ) and  $\beta$ -MCA (3 $\alpha$ , 6 $\beta$ , 7 $\beta$ ) Most bile acids in mice are taurine (T)-conjugated and secreted into bile. In the intestine, gut bacteria de-conjugate bile acids and then remove the 7a-hydroxyl group from CA and CDCA to form secondary bile acids deoxycholic acid (3α, 12α) and lithocholic acid (3α), respectively. T-α-MCA and T-β-MCA are converted to T-hyodeoxycholic acid (THDCA, 3 $\alpha$ , 6 $\alpha$ ), T-ursodeoxycholic acid (TUDCA, 3 $\alpha$ , 7 $\beta$ ), T-hyocholic acid (THCA, 3 $\alpha$ , 6 $\alpha$ , 7 $\alpha$ ) and T-murideoxycholic acid (TMDCA, 3 $\alpha$ , 6 $\beta$ ).

chronic renal disease progression. NRs are at the forefront of innovative therapeutic approaches for kidney diseases such as glomerulosclerosis, tubulointerstitial disease, renal lipotoxicity, kidney fibrosis, and hypertension. The kidney expresses NRs, FXR, PXR, and VDR, which are essential in renal disease.

#### The Role of FXR in DN

The NR, FXR was discovered in  $1995^{54,55}$  $1995^{54,55}$  $1995^{54,55}$  $1995^{54,55}$  $1995^{54,55}$  and plays a crucial role as the primary detector of BAs and regulator of their synthesis, secretion, and metabolism in the liver, ileum, and colon.<sup>[56](#page-13-25)</sup> It is a member of the NRs superfamily, which regulates the transcription of specific target genes. FXR maintains BAs, glucose, and lipid equilibrium by regulating various target genes. It is highly expressed in various organs, including the liver,  $57$  gallbladder,  $58$  kidney,  $59$  heart,  $60$  intestine, vasculature, atherosclerotic plaque, $61,62$  $61,62$  $61,62$  and adrenal glands.<sup>[63](#page-14-2)</sup> It is involved in inflammation, immunological responses, and fibrosis. The kidney is vital for regulating water and solute balance, and any impairment or malfunction is linked to increased illness and death rates.<sup>[64](#page-14-3)</sup> FXR plays a significant role in renal water reabsorption and is believed to have protective activities in cases of acute kidney disease, chronic kidney disease, and diabetic kidney disease.<sup>[18](#page-12-15)</sup>

Inhibiting increasing proteinuria, podocyte loss, mesangial expansion, and renal lipid accumulation, the renoprotective impact of FXR is primarily mediated by modulating lipid metabolism, oxidative stress, proinflammatory cytokines, and profibrotic factors.<sup>65</sup> In STZinduced diabetic mice, FXR deletion accelerated fibrosis, increased plasma lipid levels, and worsened diabetic kidney injury compared to diabetic wild-type (WT) mice. $^{66}$  $^{66}$  $^{66}$  The FXR agonist, GW4064, in contrast, decreased visfatin in HG-induced HMCs. This stopped inflammation, fibrosis, and cell division. In animal models of type 2 diabetes,  $66$  OCA and GW4064 were effective in preventing the progression of DN by ameliorating proteinuria, podocyte injury, profibrotic and proinflammatory gene expression, and mesangial cell proliferation.<sup>[67-69](#page-14-6)</sup> In db/db mice, GW4064 reduced glomerular damage and fibrosis, as well as blood glucose, albuminuria, blood urea nitrogen, and serum creatinine. This suggested that FXR activation stopped the progression of  $DN$ .<sup>[69](#page-14-7)</sup>

#### FXR Regulates Glucose Metabolism

FXR is a key player in glucose metabolism, controlling glucose levels<sup>70-74</sup> and increasing glycogen synthe- $\sin^{75,76}$  $\sin^{75,76}$  $\sin^{75,76}$  $\sin^{75,76}$  It inhibits the expression of glycogen synthase kinase-3 beta, a transcription factor that

phosphorylates and inactivates glycogen synthase, leading to increased glycogen synthesis. FXR reduces glycolysis<sup>77</sup> by suppressing the transcriptional activity of carbohydrate response element-binding protein, a transcription factor that promotes glycolysis. This inhibits the expression of glycolytic enzymes and reduces glycolysis. FXR downregulates genes involved in gluconeogenesis, such as phosphoenolpyruvate carboxykinase and glucose-6-phosphatase, which are key enzymes in the gluconeogenic pathway. These downregulations lead to a decrease in gluconeogenesis and serum glucose levels. $75,78$  $75,78$ 

Insulin resistance and hyperglycemia were observed in FXR-null mice, indicating disruption of normal glucose homeostasis.<sup>[79](#page-14-13)</sup> Elevated fatty acid levels likely caused these issues. When activated by an agonist, FXR repressed gluconeogenic genes, phosphoenolpyruvate carboxykinase, and glucose-6-phosphatase, indicating that the FXR/SHP system regulates glucose synthesis in the liver.<sup>[79](#page-14-13)</sup> Activating FXR enhanced insulin sensitivity and tolerance in diabetic obese mice. When FXR-VP16 or FXR agonists were administered to diabetic mice, plasma glucose levels were significantly lowered and insulin sensitivity increased.<sup>[76](#page-14-10)</sup> In summary, FXR activation modulates glucose metabolism by increasing glycogen synthesis, reducing glycolysis, and decreasing gluconeogenesis, ultimately reducing serum glucose levels.

#### FXR Regulates Lipid Metabolism

FXR is recognized as a crucial controller of cholesterol and lipid balance in the body. Numerous studies have established a link between kidney dysfunction and the buildup of lipids in the kidneys in different disease models, such as metabolic disease (obesity, metabolic syndrome, and diabetes mellitus), acute kidney injury, and chronic kidney disease.<sup>80</sup>

In mice models of diabetes and diet-induced obesity, there is an elevation in triglyceride and cholesterol levels as compared to healthy controls. The elevated levels of triglycerides in the kidneys are linked to an increase in the levels of 2 transcription factors, sterol regulatory element-binding protein 1c and carbohydrate response element-binding protein. These transcription factors are known to enhance the synthesis of fatty acids.<sup>[81](#page-14-15)</sup> Sterol regulatory element-binding protein 1c is a crucial factor that promotes the synthesis of fatty acids $82$  and the accumulation of lipids in the kidneys. In these 2 models, treatment with FXR agonists demonstrates a protective effect on the kidneys by reducing the expression of genes involved in the fatty acid synthesis, such as sterol regulatory elementbinding protein 1c, stearoyl CoA desaturase-1, and acetyl CoA carboxylase, and increasing the expression

<span id="page-4-0"></span>

Figure 3. FXR inhibits the synthesis of FA and TG, promotes the oxidative decomposition of FA, and plays a role in lipid lowering through various pathways. ACSL, long chain fatty acyl CoA synthetase; CoA, coenzyme A; CPT1, carnitine palmitoyl transferase 1; FA, fatty acid; FAS, fatty acid synthase; FFA, free fatty acids; FXR, farnesol X receptor; Glu, glucose; PPAR alpha, peroxisome proliferator activates receptor alpha; SHP, small molecule heterodimer; SREBP-1c, sterol element-binding protein-1c; TG, triglyceride.

of genes involved in fatty acid oxidation and lipid breakdown, such as PPARa, CPT1a, UCP-2, PGC-1a, and LPL. FXR suppresses the production of fatty acids and triglycerides, enhances the breakdown of fatty acids by oxidation, and contributes to the reduction of lipid levels by engaging a variety of pathways ([Figure 3\)](#page-4-0).

FXR reduces the concentration of low-density lipoprotein cholesterol in the blood plasma $^{83}$  $^{83}$  $^{83}$  by promoting the uptake and breakdown of low-density lipoprotein particles.<sup>[76](#page-14-10)</sup> FXR knockout mice show increased levels of high-density lipoprotein cholesterol due to a decrease in the expression of genes involved in reverse cholesterol transport, such as SCARB1 and ATP-binding cassette transporters G5 and G8, which remove highdensity lipoprotein cholesterol from the blood-stream.<sup>[84](#page-14-18)</sup> FXR can also decrease cholesterol buildup in hepatocytes and renal epithelial cells by suppressing cholesterol production<sup>[85](#page-14-19)</sup> and increasing ATP-binding cassette transporter A1 expression. $86$  In models of insulin resistance, FXR can decrease triglyceride buildup in the liver and kidneys by lowering the expression of fatty acid synthase and acetyl CoA carboxylase, inhibiting the activity of sterol regulatory elementbinding protein 1c and carbohydrate response element-binding protein, which regulate the response to carbohydrates.<sup>[87](#page-14-21)</sup> This inhibition promotes the clearance of triglycerides by increasing the oxidation of fatty acids through the PPAR $\alpha/\gamma$ -CPT1 pathway<sup>88[,89](#page-14-23)</sup> and reduces the uptake of fatty acids by decreasing CD36 expression. FXR also causes the whitening of

brown adipose tissue in adipocytes by activating SCD expression through PPAR $\gamma$  activation.<sup>90,[91](#page-15-1)</sup>

#### FXR Regulates Inflammation

In the course of diabetes and renal disease, inflammatory infiltrates are a key characteristic of DN develop-ment.<sup>[92](#page-15-2)</sup> NF-KB serves as a crucial signaling pathway for initiating the inflammatory response, facilitating the release of inflammatory mediators such as leukin and tumor necrosis factors such as MCP-1, TNF- $\alpha$ , and<br>IL-6,<sup>[93,](#page-15-3)[94](#page-15-4)</sup> thereby inducing inflammation,<sup>95</sup> which are closely associated with DN development. Consequently, effectively controlling the duration of the inflammatory response can significantly prevent DN ([Figure 4\)](#page-5-0).

BAs treatment in diabetic mice reduced proin-flammatory factors in their kidneys.<sup>[96](#page-15-6)</sup> FXR activation has been shown to inhibit inflammation in various diseases, including nonalcoholic fatty liver disease, pulmonary fibrosis,<sup>[97](#page-15-7)</sup> atherosclerosis, inflammatory bowel disease, chronic pancreatitis, and DN. Macrophages are key mediators of renal vascular inflammation, and DN is characterized by an increase in these cells. High glucose, AGEs (are stable covalent compounds that are derived from the amino groups of proteins, fats and nucleic acids and reducing sugars [glucose, fructose, pentose, etc.] in physiological environments through non-enzymatic catalyzed reactions), and oxidized low-density lipoprotein contribute to

macrophage accumulation and activation. When FXR deficiency is treated, NF-kB activity is linked to increased macrophage infiltration. However, when FXR agonist INT-747 is administered, this trend re-verses.<sup>98,[99](#page-15-9)</sup> INT-747 reduces inflammatory response in hepatocytes and vascular smooth muscle cells by blocking the NF-kB signaling pathway. These findings suggest that FXR activation suppresses inflammatory characteristics in diabetic kidneys and macrophages.

#### FXR Regulates Oxidative Stress

Oxidative stress, the excessive production of reactive oxygen species and weakening of antioxidant capacity, is a significant factor in the development of DN. Redox homeostasis is crucial for cellular functionality and survival, $100$  whereas oxidative stress is a state of imbalance between oxidizing agents and antioxidant substances in the human body. Oxidative stress is a major cause of kidney damage, because it leads to the production of reactive oxygen species in mesangial cells of the glomerulus when exposed to high glucose levels. This intensifies oxidative stress, exacerbating kidney injury. However, mitigating the excessive production of reactive oxygen species can alleviate the negative effects of oxidative stress.<sup>[101](#page-15-11)</sup>

Treatment with GW4064 and choleric acid can induce the glutathione metabolism gene to increase, eliminate excess reactive oxygen species, reduce oxidative stress, restore mitochondrial and endoplasmic reticulum

<span id="page-5-0"></span>

Figure 4. FXR regulates inflammation as well as the kidney fibrosis. The activation of FXR by bile acids can inhibit the NF-KB pathway to reduce inflammation, thereby reducing the production of mitochondrial ROS and alleviating oxidative stress. FXR activation also reduces renal fibrosis by inhibiting the TGF-smad pathway. α-sma, alpha-smooth muscle actin; ECM, extra cellular matrix; IKK, inhibitor of KappaB kinase; IL-1β, interleukin-1 $\beta$ ; IL-6, interleukin-6; MCP-1, monocyte chemotactic protein-1; NAPDH, nicotinamide adenine dinucleotide phosphate; NF-kB, nuclear factor-k-gene binding; ROS, reactive oxygen species; TGF-ß1, transforming growth factor-ß; TLR, toll-like receptor; TNF- $\alpha$ , tumor necrosis factor.

function, and prevent apoptosis, thus protecting proximal renal tubule cells. Activation of FXR can reduce inflammatory factors, alleviate apoptosis, downregulate reactive oxygen species levels, and prevent oxidative stress induced mitochondrial damage. NADPH oxidase 2 is a major source of reactive oxygen species in vivo. CDCA can activate FXR, downregulate NADPH oxidase 2 expression and alleviate oxidative stress in rats fed high fructose. Changes in the protein folding pathway regulated by the endoplasmic reticulum can cause imbalance of reactive oxygen species and increase the production of active oxides, potentially leading to kidney injury by mediating oxidative stress. FXR agonist, Tauro ursodeoxycholic acid can protect renal tubular epithelial cells from hyperglycemia damage and improve endoplasmic reticulum stress signal transduction.

#### FXR Regulates Renal Fibrosis

Renal fibrosis is a significant characteristic of chronic kidney disease,  $96$  involving DN, and is a crucial phase in the progression of all renal diseases toward end-stage renal disease.<sup>102</sup> It is primarily caused by the activation of renal fibroblasts and myofibroblasts in response to factors such as inflammation, hypoxia, and hyperglycemia. The main harmful effects of increasing renal fibrosis include the gathering of fibroblasts, increased expression of extracellular matrix (ECM) proteins, and issues with nephron function. TGF- $\beta$  is a crucial player in the development of renal fibrosis, because it both increases the expression of ECM proteins and stops them from breaking down, leading to the buildup of ECM.

In animal models of obesity and insulin resistance induced by high-fat diet,<sup>[103](#page-15-13)</sup> type  $1^{104}$  $1^{104}$  $1^{104}$  or type 2 diabetes, and aging models,<sup>12</sup> the downregulation of FXR and its target genes in the kidney is closely related to renal fibrosis and the degree of renal insufficiency.<sup>105</sup> FXR agonists can regulate ECM production through the TGF- $\beta$ -Smad pathway and FXR-SHP pathway, improving liver, myocardial, and kidney fibrosis. When FXR is turned on, many markers related to ECM, myofibroblast activation, profiber signaling proteins, and histochemical fibrosis go down.<sup>[106](#page-15-16)</sup> The FXR protein controls the TGF- $\beta$  pathway by decreasing the expression of SREBP-1, which can enhance the expression of TGF- $\beta$ . When FXR is turned on, it activates the  $\alpha/\beta$  hydrolase NDRG2, stopping the production of TGF- $\beta$ .<sup>[107](#page-15-17)</sup><br>The downregulation

The downregulation of FXR prevents renal fibrosis. It is important to note that inflammatory mediators can induce organ fibrosis by promoting inflammatory response; and FXR, as an anti-inflammatory NR, can delay the progression of renal fibrosis by inhibiting inflammation.

# FXR Reduces Proteinuria

Proteinuria is a key indicator of DN, with microalbuminuria being a key predictor of disease progres-sion.<sup>[59](#page-13-28)</sup> Hemodynamic abnormalities, podocyte dysfunction, and renal tubular reabsorption damage contribute to the progression of nephropathy. $108-110$ Wang et  $al.^{66}$  $al.^{66}$  $al.^{66}$  treated DBA/2J mice fed with a Western diet by daily gavage of INT-747, which significantly reduced urinary protein, improved podocyte loss and mesangial expansion, and considered that activation of FXR could regulate fatty acid synthesis and oxidation, improve triglyceride accumulation and kidney structure. In Wistar rats,<sup>[111](#page-15-19)</sup> intragastric administration of CDCA alleviated kidney damage, and urine protein was significantly reduced. FXR KO diabetic mice exhibited typical DN pathological changes, including podocyte loss,<sup>[66](#page-14-5)</sup> glomerular lobulation, mesangial matrix expansion, tubular injury, and protein cast. Treatment with CA for 12 weeks improved proteinuria, glomerulosclerosis, and tubulointerstitial fibrosis, $81$  further confirming FXR's role in reducing urinary protein in DN. Urinary microalbumin is a clinically recognized early biomarker of DN. The combination of oral enalapril and intraperitoneal Tauro ursodeoxycholic acid can effectively reduce urinary albumin in 16-week-old db/db mice, and Tauro ursodeoxycholic acid only can improve renal tubular damage. FXR activation can protect podocytes, tubular cells, and mesangial cells, thus reducing DN urinary protein.

# The Role of PXR in DN

PXR, a NR, is highly expressed in the human liver, intestinal tissues, and kidneys[.112](#page-15-20) It regulates various physiological and pathological processes, including metabolism, transport, combination, excretion, and exogenous and endogenous substances.<sup>113</sup> PXR also regulates glucose, $114$  lipids, steroids, CA, bilirubin, and bone minerals, and immune response.<sup>[115](#page-15-23)[,116](#page-15-24)</sup> It affects liver, gastrointestinal, and tumor development and plays a crucial role in renal physiology and pathology.<sup>[117-119](#page-15-25)</sup> This study demonstrates that xenobiotic NR, PXR is activated and epigenetically changed, potentially playing a role in diabetic kidneys.<sup>[120](#page-16-0)</sup> PXR is essential in metabolic alterations associated with diabetes and obesity. $121$ 

#### PXR Regulates Cholesterol Homeostasis

Endogenous cholesterol metabolites can activate PXR, suggesting that PXR plays a role in the removal of potentially hazardous intermediates. Cholesterol catabolism is known to be inhibited by the signaling molecule known as cholecalciferol acid. When mice lacking PXR were given a diet supplemented with CA in research by Sonoda et  $al.$ <sup>[122](#page-16-2)</sup> adding cholesterol to their food caused rapid mortality accompanied by symptoms

<span id="page-7-0"></span>

Figure 5. PXR regulates glucose and cholesterol metabolism. (a) PXR, a xenobiotic sensing regulator, is critical for glucose metabolism. (b) PXR regulates cholesterol metabolism. CD36, cluster determinant 36; CPT1A, carnitine palmitoyl transferase 1A; CREB, cAMP -response element binding protein; FA, fatty acid; FAT, fatty acid translocase; FFA, free fatty acid; FoxA2, forkhead box A2; FOXO1, forkhead box O1; G6Pase, glucose-6-phosphatase; HNF4, hepatocyte nuclear factor 4; PEPCK, phosphoenol pyruvate; carboxykinase; PGC-1a, peroxisome proliferatorsactivated receptor  $\gamma$  coactivator I alpha; PXR, pregnane X receptor; SCD1, stearoyl-CoA desaturase-1; TG, triglyceride.

of hepatorenal failure. Renal failure was suggested as the possible direct cause of death. These findings highlight the special and vital role that PXR plays in defending against cholesterol and its metabolites.

Carnitine palmitoyltransferase 1A (CPT1A) and mitochondrial 3-hydroxy-3-methylglutaryl-coenzyme A synthase 2 (HMGCS2) are key enzymes for  $\beta$ -oxidation and ketogenesis. FoxA2, a winged helix/forkhead transcription factor, speeds up the transcription of CPT1A and HMGCS2 when insulin is not present. $123$ Insulin stimulates FoxA2 by phosphorylating and exonucleating it while suppressing the transcription of CPT1A and HMGCS2.<sup>[124](#page-16-4)</sup> Nakamura et al.<sup>[125](#page-16-5)</sup> discovered that pregnenolone-16 $\alpha$ -carbonitrile suppresses transcription of CPT1A and HMGCS2 in WT mice but not in Pxr knockout mice. PXR may bind directly to FoxA2, inhibiting the activation of the CPT1A and HMGCS2 genes. In [Figure 5a](#page-7-0), we depict PXR's entire mechanism in cholesterol metabolism.

#### PXR Regulates Glucose Metabolism

PXR regulates gluconeogenesis by modulating the expression of key enzymes; and is involved in the oxidative uptake of glucose by regulating glucose transporter 2 and glucokinase. PXR also regulates

cellular processes such as sterol regulatory element binding protein 1c, fatty acid translocation enzyme CD36, carnitine palmitoyl transferase 1A, mitochondrial hydroxyl-3-methylglutaryl CoA synthetase 2, mitochondrial sterol 27 hydroxylase, and cholesterol effluent transporter A1, G1.

Increasing evidence suggests that PXR activation has a function in glucose homeostasis. Hormones (insulin, glucagon, glucocorticoids, etc.) regulate the activity of key enzymes involved in various glucose metabolism processes, allowing blood glucose concentrations to remain relatively constant. When PXR is turned on, it lowers the amounts of 2 important enzymes that con-trol gluconeogenesis.<sup>[126](#page-16-6)[,127](#page-16-7)</sup> These are glucose-6phosphatase and phosphoenolpyruvate carboxy kinase. Kodama et  $al.^{127}$  $al.^{127}$  $al.^{127}$  discovered that PXR regulates gluconeogenesis by interacting with FOXO1, CREB, and HNF4. HNF4, together with PGC-1 $\alpha$ , stimulates gluconeogenesis. Bhalla et  $al.^{126}$  $al.^{126}$  $al.^{126}$  found that PXR competes with HNF4 for PGC-1 $\alpha$  and suppresses gluconeogenesis. In vivo studies confirmed the validity of these findings. When insulin levels are low, FOXO1 activates glucose-6-phosphatase and phosphoenolpyruvate carboxykinase. In contrast, PXR stops CREB from attaching to similar sites, which stops the transcription

of glucagon-activated glucose-6-phosphatase and stops the process of making glucose.<sup>[128](#page-16-8)</sup> In a nutshell, PXR is an important xenobiotic sensing regulator in glucose metabolism ([Figure 5](#page-7-0)b).

#### PXR Regulates Renal Fibrosis

PXR is expressed in many parts of the kidney, indicating that it plays an important role in regulating kidney function. In the mouse model of cisplatininduced acute kidney injury, the expression of PXR in the kidney also decreased with the aggravation of acute kidney injury. When pregnenolone-16 $\alpha$ -carbonitrile, a specific PXR agonist, was used, renal tissue damage and functional decline got a lot better. This protective mechanism may have something to do with PI3K being turned on. Renal fibrosis is the common pathological feature and final manifestation of DN, and PXR plays an important role in renal fibrosis. $129,130$  $129,130$ Pregnenolone-160-carbonitrile treated a mouse model of chronic kidney disease, and they found significant improvements in renal function decline and kidney fibrosis. On the contrary, when the PXR gene is knocked out, the degree of renal fibrosis is significantly aggravated.<sup>131</sup> By stopping the Smad3 signaling pathway and the Wnt7a/ $\beta$ -catenin signaling pathway from working, PXR helps treat renal fibrosis. PXR controls Wnt7a, which is linked to p53. PXR can bind to p53 and stop p53 from increasing Wnt7a gene transcription, which lowers ECM production. $132$ 

#### The role of VDR in DN

VDRs are nucleophilic proteins within the thyroid hormone superfamily.<sup>[133](#page-16-13)</sup> VDRs form a heterodimer with retinol receptors, necessary for high-affinity DNA binding. They are part of the ligand-activated transcription factor family.  $1,25-(OH)_{2}D_{3}$  is the primary ligand that activates VDRs, binding to the VDRretinoid X receptor complex and a specific cis-DNA sequence of VDRE.<sup>[53](#page-13-22)</sup> This modulates transcription by activating the target motif.  $1,25-(OH)_2D_3$  has both active and inhibitory sexual properties and has a wide range of physiological effects,<sup>[134](#page-16-14)</sup> including maintaining calcium and phosphorus homeostasis, regulating the immune response, participating in inflammatory responses, inhibiting cell proliferation, causing cell apoptosis, and promoting differentiation.<sup>[135](#page-16-15)</sup>

VDRs are found in various cells, including skin, vascular, immunological, colon, pancreatic, and kidney cells.<sup>136</sup> In kidney tissues, they produce the highest  $1,25-(OH)<sub>2</sub>D<sub>3</sub>$  due to high selectivity and affinity binding. $137$  This results in a heterodimer between VDR and retinoid X receptor, facilitating transcription of VDR-targeted genes. VDRs are involved in mineral metabolism, renal and cardiovascular control, $^{137}$  $^{137}$  $^{137}$  and have various implications in renal illness.

Recent studies have shown that VDRs play a crucial role in the development of DN by controlling inflammatory reactions, reducing proteinuria, stopping renal fibrosis, and inhibiting the RAS. $138,139$  $138,139$  Treatment with a VDR agonist can reduce proteinuria, podocyte damage, mesangial enlargement, and ECM protein buildup in a mouse model of diet-induced obesity. The VDR agonist also decreases macrophage infiltration, oxidative stress, proinflammatory cytokines, and profibrotic growth factors. In addition, VDR activation reduces the accumulation of neutral lipids and adipophilin in the kidney, inhibiting renal diet-induced obesity symptoms.

#### VDR and RAS

The RAS is a regulatory process where angiotensin (Ang) II acts as the main mediator. Ang II is produced through 2 enzymatic cleavages. The first step involves the cleavage of angiotensinogen by renin, resulting in the formation of Ang I. This is then converted to Ang II by the angiotensin-converting enzyme.<sup>[140](#page-16-20)</sup> Clinical research $141$  shows that Ang II type 1 receptor blockers or angiotensin-converting enzyme inhibitors slow the development of tubulointerstitial fibrosis, glomerulosclerosis, and proteinuria in diabetic patients. This suggests that the RAS plays a key role in the progressive renal injury seen in  $DN$ .<sup>142</sup>

According to a study by Zhang et al.,<sup>[143](#page-16-23)</sup> diabetic VDR KO mice experienced more severe nephropathy than WT mice because of increased RAS activation in the kidney. This finding confirms that VDR protects against renal injury caused by hyperglycemia by suppressing RAS. A model of type 1 diabetes produced by STZ was employed in this investigation. Diabetes was established in VDR KO mice and WT mice by administering a low-dose injection of STZ. The mice were then observed for 19 weeks. Both diabetic WT and VDR KO mice had a similar level of increasing hyperglycemia. VDR KO animals exhibited accelerated and more pronounced albuminuria compared to WT mice. The electron microscopy analysis of VDR KO mice indicates that there is a notable thickening of the glomerular basement membrane and an increase in podocyte foot process effacement, which is consistent with the more severe albumin uric phenotype. In mice that do not have the VDR gene, the production of renin in the kidney was significantly increased. This resulted in a significant increase in plasma renin and Ang II levels, leading to hypertension and cardiac hypertrophy. However, when the human VDR gene was specifically expressed in the juxtaglomerular cells of transgenic mice using the renin gene promoter, the expression of renin was suppressed. This inhibitory effect was not influenced by parathyroid hormone or calcium levels, indicating that the VDR receptor directly regulates renin production.<sup>[144](#page-16-24)</sup>

#### VDR is Involved in Inflammation and Fibrosis

NF-kB is a key regulator of genes involved in inflammation and fibrogenesis, including TNF- $\alpha$ , TGF- $\beta$ 1, monocyte chemoattractant protein-1, and plasminogen activator inhibitor-1. These genes are crucial in the progression of kidney disease and are linked to the functioning of endothelium.  $TNF-\alpha$  activates the c-Jun N-terminal kinase pathway, which hinders the activity of endothelial NO synthase (eNOS), reducing the availability of nitric oxide. This leads to the formation of the superoxide anion  $O_2$  by upregulating xanthine oxidase activity, further impacting endothelial function. TNF-a-bound TNFR triggers NF-kB translocation to the nucleus, promoting the expression of proinflammatory mediators [\(Figure 6](#page-9-0)).

NF-kB activation has been reported in patients with DN.<sup>[145](#page-16-25)</sup> In a mouse model, TNF- $\alpha$  stimulates renal tubular cells to produce the RANTES chemokine, facilitating the infiltration of monocytes and T cells into the kidneys, contributing to kidney damage. Paricalcitol injection disrupts this harmful cycle by increasing the expression of VDR in renal tubule cells, which binds to NF-kB and inhibits the transcription of RANTES.

TGF- $\beta$ 1 is synthesized in an inactive state and stored in the endothelium. $146$  When activated, it attaches to its receptor and enters the nucleus to initiate the transcription of genes necessary for myofibroblast development, ECM components, and connective tissue growth factor.<sup>147</sup> In addition to the aforementioned traditional TGF- $\beta$ 1 signal pathways, numerous addi-<br>tional routes exist.<sup>148</sup> Monocyte chemoattractant protein-1 promotes macrophage infiltration in the kidney, a problem seen in a number of kidney diseases. Macrophages release many factors that promote kidney disease progression.<sup>149</sup> By inhibiting the activation of NF-KB, 1,25- $(OH)_{2}D_{3}$  at the molecular level reduces the expressions of angiotensinogen and monocyte chemoattractant protein-1, as well as plasminogen activator inhibitor-1, which is generated by inflammation and high hyperglycemia.

# VDR Reduces Proteinuria

Podocytes are essential for controlling the kidney's glomerular filtration rate. The glomerular filtration barrier, which keeps proteins and other big molecules from being filtered into the urine, is largely composed

<span id="page-9-0"></span>

Figure 6. VDR regulates inflammation and oxidative stress. Ligand bound vitamin D receptor (VDR) activation suppresses gene expression of nuclear factor-κB (NF-κB) and tumor necrosis factor (TNF)-α receptors 2 and 4 (TNFRs). Among these proinflammatory cytokines, TNF-α activates the c-Jun N-terminal kinase (JNK) pathway that inhibits endothelial NO synthase endothelial NO synthase activity, consequently resulting in reduction of nitric oxide (NO) bioavailability. In addition, the upregulated JNK pathway induces the formation of superoxide anion  $O<sub>2</sub>$  from oxygen ( $O<sub>2</sub>$ ) through upregulating xanthine oxidase (XO) activity, which further impairs endothelial function via inducing oxidative stress in the cell. Furthermore, TNF-a bound TNFRs trigger the translocation of NF-kB to the nucleus to promote the expression of pro-inflammatory mediators.

of the foot processes of podocytes. Strong evidence from Wang's work<sup>[150](#page-17-0)</sup> suggests that vitamin D/VDR signaling in podocytes is essential for protecting the kidney from diabetic damage. In this work, DBA/2J mice that overexpress the human VDR were employed. Following streptozotocin-induced diabetes development, transgenic mice exhibited reduced levels of albuminuria in comparison to WT controls. A low dosage of the vitamin D analog doxercalciferol inhibited podocyte loss and death, decreased glomerular fibrosis, and avoided the development of albuminuria in transgenic mice. However, in WT mice, the same dosage of vitamin D had minimal impact on the development of DN. Furthermore, significant diabetesrelated kidney damage was prevented in VDR-null mice by restoration of the mice using the human VDR transgene in podocytes.<sup>[151](#page-17-1)</sup>

# The Function of Membrane Receptor in the Kidney

The G-protein-coupled receptor, TGR5, abundantly expressed on eukaryotic cell membranes, serves as a highly prominent membrane receptor in the kidney. It exerts its cellular functions by binding to agonists such as hormones, neurotransmitters, or external stimuli upon activation. The activation of G-protein-coupled receptors subsequently triggers the activation of G proteins located on the cell membrane. These activated G proteins then bind to effector enzymes present on the cell membrane, leading to the generation of second messengers that initiate cytoplasmic cellular responses. Research has demonstrated that TGR5 plays a crucial role in kidney pathophysiology.

#### The Role of TGR5 in DN

TGR5 serves as a cell membrane receptor for BAs and belongs to the G-protein-coupled receptor family. $11,152$  $11,152$ It is composed of 7 transmembrane domains and facilitates the transmission of extracellular signals by interacting with a heterotrimeric G protein structure. In contrast to other BA receptors, TGR5 exhibits selectivity toward the structural characteristics of BAs, remaining unaffected by the derivation pattern and binding state of BAs. Different BAs can be ranked by how well they activate TGR5. Lithocholic acid is the most effective, followed by deoxycholic acid, CDCA, CA, and UDCA. The TGR5 receptor exhibits broad expression patterns in various human and animal tissues, including but not limited to the spleen, kidney, lung, liver, gallbladder, intestine, and skeletal muscle. In the kidney, TGR5 is highly expressed in tubular cells but also in glomerular cells including podocytes and mesangial cells. $153,154$  $153,154$  Possible mechanisms by which TGR5 activation is beneficial to the kidney

include increased mitochondrial oxidative phosphorylation, mitochondrial fatty acid  $\beta$ -oxidation and mitochondrial superoxide dismutase activity, inhibition of mitochondrial ROS production, and anti-inflammatory effects.

# TGR5 Improves the Lesion of DN

On an HFD, TGR5 knockout mice gain weight. In standard rodent chow diet, TGR5 knockout mice do not exhibit any kidney disease phenotype and remain viable. In contrast to their male WT littermates or female TGR5 knockout mice, TGR5 knockout mice exhibited more obvious hepatic steatosis when subjected to an HFD or diabetes. When INT-777 or another TGR5 agonist, oleanolic acid, is administered to WT mice, it also increases fat burning and energy expenditure, which mitigates HFD-induced kidney impairment.<sup>[155](#page-17-5)</sup>

TGR5, a membrane sensor that is turned on by BAs, can directly control how podocytes work. In addition, activating TGR5 led to the release of ileac glucagon-like peptide-1, $156$  which suggests that BAs might be useful as a treatment for DN. To treat diabetic db/db mice with the TGR5 agonist INT-777,  $152,157,158$  $152,157,158$  $152,157,158$  proteinuria, glomerular mesangial expansion, glomerular podocyte damage, and the buildup of ECM proteins and macrophages were all greatly reduced.<sup>13</sup> Consistently, Xiao et al.<sup>[159](#page-17-9)</sup> revealed that activating TGR5 stopped the disease from getting worse by reducing inflammation through the NF-kB pathway. Studies done in the laboratory also revealed that turning on TGR5 greatly reduced the levels of TGF- $\beta$ 1 and fibronectin in HGinduced GMCs, both of which can help cause kidney fibrosis.<sup>[153,](#page-17-3)[154](#page-17-4)</sup> Studies<sup>[13](#page-12-12)</sup> show that lowering the high levels of acetylation of SOD2 and isocitrate dehydrogenase 2 in mitochondria from db/db animals that had not been treated exhibited higher SIRT3 activity. Therefore, INT-777 increased SOD2 activity and decreased the production of  $H_2O_2$  in mitochondria. This was connected to a decrease in  $H_2O_2$  and thiobarbituric acid reactive chemicals in the urine. TGR5 activation leads to increased energy metabolism, mitochondrial biogenesis, and fatty acid oxidation in the kidneys by activating AMPK, SIRT1, PGC-1a, ERRa, and SIRT3. $<sup>13</sup>$  $<sup>13</sup>$  $<sup>13</sup>$  This is in line with several studies in</sup> humans and animal models that link impaired mitochondrial function and oxidative phosphorylation to the development of type 2 diabetes.<sup>[160](#page-17-10),[161](#page-17-11)</sup> This stops oxidative stress and lipid buildup, firmly establishing TGR5's important role in preventing kidney disease in people with diabetes and obesity. Overall, TGR5 has a significant impact on controlling the formation and function of kidney mitochondria, the breakdown of fatty acids by beta-oxidation, the reduction of

<span id="page-11-0"></span>

Figure 7. TGR5 regulates renal mitochondrial biogenesis, fatty acid  $\beta$ -oxidation and oxidative stress, alleviates renal inflammation and fibrosis. Activation of TGR5 by bile acids or TGR5 agonists leads to rapid intracellular cAMP production and activation of PKA, which affects downstream cellular events. TGR5 activation in inducing energy metabolism, mitochondrial biogenesis, and fatty acid oxidation in the kidney by activating AMPK, SIRT1, PGC-1a, ERRa, and SIRT3, which lead to prevention of oxidative stress and lipid accumulation. TGR5 activation inhibits NF-kB signaling pathway to reduce renal inflammation and inhibits HG-induced up-regulation of GMCs fibrosis markers, thereby alleviating fibrosis. Activation of TGR5 can regulate glycolipid metabolism through downstream cAMP and inhibit inflammatory response through PKA pathway. AMPK, adenosine 5'-monophosphate (AMP)-activated protein kinase; cAMP, Cyclic adenosine monophosphate; ERRa, estrogenrelated receptor alpha; GMCs, glomerular mesangial cell; HG, high glucose; PGC-1 $\alpha$ , peroxisome proliferators-activated receptor  $\gamma$  coactivator l alpha; PKA, protein kinase A; SIRT1, Silent information regulator 1; SIRT3, Silent information regulator 1; TGR5, G protein-coupled receptor.

oxidative stress, and the prevention of kidney inflammation and fibrosis ([Figure 7\)](#page-11-0).

### **CONCLUSION**

Kidney diseases, particularly DN, are a major global health issue with increasing morbidity and mortality. DN is a microvascular complication, particularly prevalent in patients with diabetes, and significantly increases mortality rates. It poses a threat to public health, peripheral vascular disease, and cerebrovascular disease. BA receptors play a crucial role in renal pathophysiology, activating transcription factors for lipid, cholesterol, and glucose metabolism, as well as genes linked to inflammation and renal fibrosis. Advances in understanding kidney BA receptor biology and creating agonists have shown promising therapeutic potential for preventing DN and obesityinduced kidney damage.

Studies suggest that FXR, PXR, VDR, and TGR5 play a role in the pathogenesis of kidney disease (e.g., DN), suggesting that manipulating these receptors may offer new treatments. FXR activation reduces proteinuria, podocellular loss, mesangial dilation, fibrosis, and glomerular basement membrane thickening in patients with DN. PXR activation inhibits inflammation and liver fibrosis, plays a crucial role in cholesterol and lipid homeostasis, and shows promise in slowing DN progression. VDR activation has numerous positive

effects on RAS, inflammation, fibrosis, proteinuria, and lipid metabolism. TGR5 activation increases mitochondrial oxidative phosphorylation, fatty acid <sup>b</sup> oxidation, mitochondrial superoxide dismutase activity, inhibits mitochondrial ROS production, reduces proteinuria, and inhibits inflammation and liver fibrosis, making it an important target for DN treatment.

Nuclear hormone receptors have been understudied in the context of DN; however, ongoing clinical tri-als<sup>69[,162](#page-17-12)[,163](#page-17-13)</sup> and animal research<sup>[164-166](#page-17-14)</sup> are exploring the development of drugs that specifically alter the activity of these receptors. This could lead to increased application of nuclear hormone receptors modification for DN treatment and prevention. TGR5 modulators, which release glucagon-like peptide-1 and impact metabolic pathways, are expected to be helpful in treating diabetes and obesity.<sup>167-169</sup> The combined action of both FXR and TGR5 can reduce inflammation, lipid buildup, and fibrosis. Dual agonists such as INT-767, which activate both FXR and TGR5, can provide additional protective effects in the kidney and for in-juries related to diabetes and obesity.<sup>[12](#page-12-11),[61](#page-14-0)</sup> BA receptors FXR and TGR5 are prominent subjects of investigation in translational and therapeutic studies related to metabolic illnesses. Understanding the effects of BA metabolism and their interactions could provide a solid foundation for understanding preclinical data.

# **DISCLOSURE**

All the authors declared no conflict of interests.

#### ACKNOWLEDGMENTS

This study was supported by The Natural Science Foundation of Zhejiang Province (LQ23H280004).

Figures were created with [BioRender.com](http://BioRender.com)

# AUTHOR CONTRIBUTIONS

YYF and MJQ are the primary writers of this manuscript. QTZ, KLW, XH, and QY participated in the writing of this manuscript. XNS and GC designed and revised the whole manuscript and wrote the manuscript.

#### <span id="page-12-0"></span>**REFERENCES**

- 1. Valencia WM, Florez H. How to prevent the microvascular complications of type 2 diabetes beyond glucose control. BMJ. 2017;356:i6505. <https://doi.org/10.1136/bmj.i6505>
- <span id="page-12-1"></span>2. Sun H, Saeedi P, Karuranga S, et al. IDF Diabetes Atlas: global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. Diabetes Res Clin Pract. 2022;183:109119. [https://doi.org/10.1016/j.diabres.2021.](https://doi.org/10.1016/j.diabres.2021.109119) [109119](https://doi.org/10.1016/j.diabres.2021.109119)
- <span id="page-12-2"></span>3. Umanath K, Lewis JB. Update on diabetic nephropathy: core curriculum 2018. Am J Kidney Dis. 2018;71:884–895. [https://](https://doi.org/10.1053/j.ajkd.2017.10.026) [doi.org/10.1053/j.ajkd.2017.10.026](https://doi.org/10.1053/j.ajkd.2017.10.026)
- <span id="page-12-3"></span>4. Hostetter TH. Diabetic nephropathy. N Engl J Med. 1985;312: 642–644. <https://doi.org/10.1056/nejm198503073121008>
- <span id="page-12-5"></span><span id="page-12-4"></span>5. [DIABETIC nephropathy.](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref5) Lancet. 1951;2:974.
- 6. Forbes JM, Coughlan MT, Cooper ME. Oxidative stress as a major culprit in kidney disease in diabetes. Diabetes. 2008;57:1446–1454. <https://doi.org/10.2337/db08-0057>
- <span id="page-12-6"></span>7. Mauer SM. Structural-functional correlations of diabetic nephropathy. Kidney Int. 1994;45:612–622. [https://doi.org/10.](https://doi.org/10.1038/ki.1994.80) [1038/ki.1994.80](https://doi.org/10.1038/ki.1994.80)
- <span id="page-12-7"></span>8. Raparia K, Usman I, Kanwar YS. Renal morphologic lesions reminiscent of diabetic nephropathy. Arch Pathol Lab Med. 2013;137:351–359. <https://doi.org/10.5858/arpa.2012-0243-RA>
- <span id="page-12-8"></span>9. Li S, Li C, Wang W. Bile acid signaling in renal water regulation. Am J Physiol Ren Physiol. 2019;317:F73–F76. [https://](https://doi.org/10.1152/ajprenal.00563.2018) [doi.org/10.1152/ajprenal.00563.2018](https://doi.org/10.1152/ajprenal.00563.2018)
- <span id="page-12-9"></span>10. Fiorucci S, Distrutti E. Bile acid-activated receptors, intestinal microbiota, and the treatment of metabolic disorders. Trends Mol Med. 2015;21:702–714. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.molmed.2015.09.001) [molmed.2015.09.001](https://doi.org/10.1016/j.molmed.2015.09.001)
- <span id="page-12-10"></span>11. Herman-Edelstein M, Weinstein T, Levi M. Bile acid receptors and the kidney. Curr Opin Nephrol Hypertens. 2018;27:56–62. <https://doi.org/10.1097/mnh.0000000000000374>
- <span id="page-12-11"></span>12. Wang XX, Luo Y, Wang D, et al. A dual agonist of farnesoid X receptor (FXR) and the G protein-coupled receptor TGR5, INT-767, reverses age-related kidney disease in mice. J Biol Chem. 2017;292:12018–12024. [https://doi.org/10.1074/jbc.](https://doi.org/10.1074/jbc.C117.794982) [C117.794982](https://doi.org/10.1074/jbc.C117.794982)
- <span id="page-12-12"></span>13. Wang XX, Edelstein MH, Gafter U, et al. G protein-coupled bile acid receptor TGR5 activation inhibits kidney disease in obesity and diabetes. J Am Soc Nephrol. 2016;27:1362– 1378. <https://doi.org/10.1681/asn.2014121271>
- <span id="page-12-13"></span>14. Levi M. Role of bile acid-regulated nuclear receptor FXR and G protein-coupled receptor TGR5 in regulation of cardiorenal syndrome (cardiovascular disease and chronic kidney disease). Hypertension. 2016;67:1080–1084. [https://](https://doi.org/10.1161/hypertensionaha.115.06417) [doi.org/10.1161/hypertensionaha.115.06417](https://doi.org/10.1161/hypertensionaha.115.06417)
- 15. Yang G, Wei J, Liu P, et al. Role of the gut microbiota in type 2 diabetes and related diseases. Metabolism. 2021;117:154712. [https://doi.org/10.1016/j.metabol.2021.](https://doi.org/10.1016/j.metabol.2021.154712) [154712](https://doi.org/10.1016/j.metabol.2021.154712)
- 16. Qi Y, Jiang C, Cheng J, et al. Bile acid signaling in lipid metabolism: metabolomic and lipidomic analysis of lipid and bile acid markers linked to anti-obesity and anti-diabetes in mice. Biochim Biophys Acta. 2015;1851:19–29. [https://doi.](https://doi.org/10.1016/j.bbalip.2014.04.008) [org/10.1016/j.bbalip.2014.04.008](https://doi.org/10.1016/j.bbalip.2014.04.008)
- <span id="page-12-14"></span>17. Lefebvre P, Cariou B, Lien F, Kuipers F, Staels B. Role of bile acids and bile acid receptors in metabolic regulation. Physiol Rev. 2009;89:147–191. [https://doi.org/10.1152/physrev.](https://doi.org/10.1152/physrev.00010.2008) [00010.2008](https://doi.org/10.1152/physrev.00010.2008)
- <span id="page-12-15"></span>18. Guo Y, Luo T, Xie G, Zhang X. Bile acid receptors and renal regulation of water homeostasis. Front Physiol. 2023;14: 1322288. <https://doi.org/10.3390/ijms24032408>
- <span id="page-12-16"></span>19. Tanase DM, Gosav EM, Neculae E, et al. Role of gut microbiota on onset and progression of microvascular complications of type 2 diabetes (T2DM). Nutrients. 2020;12. [https://](https://doi.org/10.3390/nu12123719) [doi.org/10.3390/nu12123719](https://doi.org/10.3390/nu12123719)
- <span id="page-12-17"></span>20. Hylemon PB, Zhou H, Pandak WM, Ren S, Gil G, Dent P. Bile acids as regulatory molecules. J Lipid Res. 2009;50:1509– 1520. <https://doi.org/10.1194/jlr.R900007-JLR200>
- <span id="page-12-18"></span>21. Warren AM, Knudsen ST, Cooper ME. Diabetic nephropathy: an insight into molecular mechanisms and emerging therapies. Expert Opin Ther Targets. 2019;23:579–591. [https://](https://doi.org/10.1080/14728222.2019.1624721) [doi.org/10.1080/14728222.2019.1624721](https://doi.org/10.1080/14728222.2019.1624721)
- 22. Wolf G. New insights into the pathophysiology of diabetic nephropathy: from haemodynamics to molecular pathology. Eur J Clin Investig. 2004;34:785–796. [https://doi.org/10.1111/](https://doi.org/10.1111/j.1365-2362.2004.01429.x) [j.1365-2362.2004.01429.x](https://doi.org/10.1111/j.1365-2362.2004.01429.x)
- 23. Schena FP, Gesualdo L. Pathogenetic mechanisms of diabetic nephropathy. J Am Soc Nephrol. 2005;16:S30-S33. <https://doi.org/10.1111/j.1365-2362.2004.01429.x>
- 24. Susztak K, Böttinger EP. Diabetic nephropathy: a frontier for personalized medicine. J Am Soc Nephrol. 2006;17:361–367. <https://doi.org/10.1681/asn.2005101109>
- 25. [Giunti S, Barit D, Cooper ME. Diabetic nephropathy: from](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref25) [mechanisms to rational therapies.](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref25) Minerva Med. 2006;97: [241](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref25)–262.
- <span id="page-12-19"></span>26. Russell DW. The enzymes, regulation, and genetics of bile acid synthesis. Annu Rev Biochem. 2003;72:137-174. [https://](https://doi.org/10.1146/annurev.biochem.72.121801.161712) [doi.org/10.1146/annurev.biochem.72.121801.161712](https://doi.org/10.1146/annurev.biochem.72.121801.161712)
- <span id="page-12-20"></span>27. Samsu N. Diabetic nephropathy: challenges in pathogenesis, diagnosis, and treatment. BioMed Res Int. 2021;2021: 1497449. <https://doi.org/10.1155/2021/1497449>
- <span id="page-12-21"></span>28. Reidy K, Kang HM, Hostetter T, Susztak K. Molecular mechanisms of diabetic kidney disease. J Clin Invest. 2014;124:2333–2340. <https://doi.org/10.1172/jci72271>
- <span id="page-12-22"></span>29. Lundbaek K. Diabetic angiopathy: a specific vascular disease. Lancet. 1954;266:377–379. [https://doi.org/10.1016/](https://doi.org/10.1016/s0140-6736(54)90924-1) [s0140-6736\(54\)90924-1](https://doi.org/10.1016/s0140-6736(54)90924-1)
- <span id="page-12-23"></span>30. Kaiser N, Sasson S, Feener EP, et al. Differential regulation of glucose transport and transporters by glucose in vascular

endothelial and smooth muscle cells. Diabetes. 1993;42:80– 89. <https://doi.org/10.2337/diab.42.1.80>

- <span id="page-13-0"></span>31. Zheng C, Huang L, Luo W, et al. Inhibition of STAT3 in tubular epithelial cells prevents kidney fibrosis and nephropathy in STZ-induced diabetic mice. Cell Death Dis. 2019;10:848. <https://doi.org/10.1038/s41419-019-2085-0>
- <span id="page-13-1"></span>32. Thomas MC, Brownlee M, Susztak K, et al. Diabetic kidney disease. Nat Rev Dis Primers. 2015;1:15018. [https://doi.org/](https://doi.org/10.1038/nrdp.2015.18) [10.1038/nrdp.2015.18](https://doi.org/10.1038/nrdp.2015.18)
- <span id="page-13-2"></span>33. Tervaert TW, Mooyaart AL, Amann K, et al. Pathologic classification of diabetic nephropathy. J Am Soc Nephrol. 2010;21:556–563. <https://doi.org/10.1681/asn.2010010010>
- <span id="page-13-3"></span>34. Thomas MC. Targeting the pathobiology of diabetic kidney disease. Adv Chronic Kidney Dis. 2021;28:282-289. [https://](https://doi.org/10.1053/j.ackd.2021.07.001) [doi.org/10.1053/j.ackd.2021.07.001](https://doi.org/10.1053/j.ackd.2021.07.001)
- <span id="page-13-4"></span>35. Perino A, Demagny H, Velazquez-Villegas L, Schoonjans K. Molecular physiology of bile acid signaling in health, disease, and aging. Physiol Rev. 2021;101:683–731. [https://doi.](https://doi.org/10.1152/physrev.00049.2019) [org/10.1152/physrev.00049.2019](https://doi.org/10.1152/physrev.00049.2019)
- <span id="page-13-5"></span>36. Li T, Chiang JY. Bile acid signaling in metabolic disease and drug therapy. Pharmacol Rev. 2014;66:948–983. [https://doi.](https://doi.org/10.1124/pr.113.008201) [org/10.1124/pr.113.008201](https://doi.org/10.1124/pr.113.008201)
- <span id="page-13-6"></span>37. Lucas LN, Barrett K, Kerby RL, et al. Dominant bacterial phyla from the human gut show widespread ability to transform and conjugate bile acids. mSystems. 2021: e0080521. <https://doi.org/10.1128/mSystems.00805-21>
- <span id="page-13-7"></span>38. Jia W, Xie G, Jia W. Bile acid-microbiota crosstalk in gastrointestinal inflammation and carcinogenesis. Nat Rev Gastroenterol Hepatol. 2018;15:111–128. [https://doi.org/10.](https://doi.org/10.1038/nrgastro.2017.119) [1038/nrgastro.2017.119](https://doi.org/10.1038/nrgastro.2017.119)
- <span id="page-13-8"></span>39. Duseja A, Chawla YK. Obesity and NAFLD: the role of bacteria and microbiota. Clin Liver Dis. 2014;18:59–71. [https://](https://doi.org/10.1016/j.cld.2013.09.002) [doi.org/10.1016/j.cld.2013.09.002](https://doi.org/10.1016/j.cld.2013.09.002)
- <span id="page-13-9"></span>40. Wang K, Xu X, Maimaiti A, et al. Gut microbiota disorder caused by diterpenoids extracted from Euphorbia pekinensis aggravates intestinal mucosal damage. Pharmacol Res Perspect. 2021;9:e00765. [https://doi.org/10.3389/fendo.](https://doi.org/10.3389/fendo.2021.799648) [2021.799648](https://doi.org/10.3389/fendo.2021.799648)
- <span id="page-13-10"></span>41. Hofmann AF, Hagey LR. Bile acids: chemistry, pathochemistry, biology, pathobiology, and therapeutics. Cell Mol Life Sci. 2008;65:2461–2483. [https://doi.org/10.1007/s00018-008-](https://doi.org/10.1007/s00018-008-7568-6) [7568-6](https://doi.org/10.1007/s00018-008-7568-6)
- <span id="page-13-11"></span>42. Ridlon JM, Harris SC, Bhowmik S, Kang DJ, Hylemon PB. Consequences of bile salt biotransformations by intestinal bacteria. Gut Microbes. 2016;7:22–39. [https://doi.org/10.](https://doi.org/10.1080/19490976.2015.1127483) [1080/19490976.2015.1127483](https://doi.org/10.1080/19490976.2015.1127483)
- <span id="page-13-12"></span>43. Staley C, Weingarden AR, Khoruts A, Sadowsky MJ. Interaction of gut microbiota with bile acid metabolism and its influence on disease states. Appl Microbiol Biotechnol. 2017;101:47–64. [https://doi.org/10.1007/s00253-](https://doi.org/10.1007/s00253-016-8006-6) [016-8006-6](https://doi.org/10.1007/s00253-016-8006-6)
- <span id="page-13-13"></span>44. Kakiyama G, Pandak WM, Gillevet PM, et al. Modulation of the fecal bile acid profile by gut microbiota in cirrhosis. J Hepatol. 2013;58:949–955. [https://doi.org/10.1016/j.jhep.](https://doi.org/10.1016/j.jhep.2013.01.003) [2013.01.003](https://doi.org/10.1016/j.jhep.2013.01.003)
- <span id="page-13-14"></span>45. Ridlon JM, Alves JM, Hylemon PB, Bajaj JS. Cirrhosis, bile acids and gut microbiota: unraveling a complex relationship. Gut Microbes. 2013;4:382–387. [https://doi.org/10.1080/](https://doi.org/10.1080/19490976.2015.1127483) [19490976.2015.1127483](https://doi.org/10.1080/19490976.2015.1127483)
- <span id="page-13-15"></span>46. Koh A, Bäckhed F. From association to causality: the role of the gut microbiota and its functional products on host metabolism. Mol Cell. 2020;78:584–596. [https://doi.org/10.](https://doi.org/10.1016/j.molcel.2020.03.005) [1016/j.molcel.2020.03.005](https://doi.org/10.1016/j.molcel.2020.03.005)
- <span id="page-13-16"></span>47. Zhang L, Wang Z, Zhang X, et al. Alterations of the gut microbiota in patients with diabetic nephropathy. Microbiol Spectr. 2022;10:e0032422. [https://doi.org/10.1128/spectrum.](https://doi.org/10.1128/spectrum.00324-22) [00324-22](https://doi.org/10.1128/spectrum.00324-22)
- <span id="page-13-17"></span>48. de Aguiar Vallim TQ, Tarling EJ, Edwards PA. Pleiotropic roles of bile acids in metabolism. Cell Metab. 2013;17:657– 669. <https://doi.org/10.1016/j.cmet.2013.03.013>
- <span id="page-13-18"></span>49. McMahan RH, Wang XX, Cheng LL, et al. Bile acid receptor activation modulates hepatic monocyte activity and improves nonalcoholic fatty liver disease. J Biol Chem. 2013;288:11761–11770. [https://doi.org/10.1074/jbc.M112.](https://doi.org/10.1074/jbc.M112.446575) [446575](https://doi.org/10.1074/jbc.M112.446575)
- <span id="page-13-19"></span>50. Biagioli M, Carino A, Cipriani S, et al. The bile acid receptor GPBAR1 regulates the M1/M2 phenotype of intestinal macrophages and activation of GPBAR1 rescues mice from murine colitis. J Immunol. 2017;199:718–733. [https://doi.org/](https://doi.org/10.4049/jimmunol.1700183) [10.4049/jimmunol.1700183](https://doi.org/10.4049/jimmunol.1700183)
- <span id="page-13-20"></span>51. Gadaleta RM, Garcia-Irigoyen O, Moschetta A. Bile acids and colon cancer: is FXR the solution of the conundrum? Mol Aspects Med. 2017;56:66–74. [https://doi.org/10.1016/j.mam.](https://doi.org/10.1016/j.mam.2017.04.002) [2017.04.002](https://doi.org/10.1016/j.mam.2017.04.002)
- <span id="page-13-21"></span>52. Gai Z, Gui T, Hiller C, Kullak-Ublick GA. Farnesoid X receptor protects against kidney injury in uninephrectomized obese mice. J Biol Chem. 2016;291:2397–2411. [https://doi.org/10.](https://doi.org/10.1074/jbc.M115.694323) [1074/jbc.M115.694323](https://doi.org/10.1074/jbc.M115.694323)
- <span id="page-13-22"></span>53. Levi M. Nuclear receptors in renal disease. Biochim Biophys Acta. 2011;1812:1061–1067. [https://doi.org/10.1016/j.bbadis.](https://doi.org/10.1016/j.bbadis.2011.04.003) [2011.04.003](https://doi.org/10.1016/j.bbadis.2011.04.003)
- <span id="page-13-23"></span>54. Forman BM, Goode E, Chen J, et al. Identification of a nuclear receptor that is activated by farnesol metabolites. Cell. 1995;81:687–693. [https://doi.org/10.1016/0092-8674\(95\)](https://doi.org/10.1016/0092-8674(95)90530-8) [90530-8](https://doi.org/10.1016/0092-8674(95)90530-8)
- <span id="page-13-24"></span>55. Seol W, Choi HS, Moore DD. Isolation of proteins that interact specifically with the retinoid X receptor: two novel orphan receptors. Mol Endocrinol. 1995;9:72–85. [https://doi.](https://doi.org/10.1210/mend.9.1.7760852) [org/10.1210/mend.9.1.7760852](https://doi.org/10.1210/mend.9.1.7760852)
- <span id="page-13-25"></span>56. Cai J, Rimal B, Jiang C, Chiang JYL, Patterson AD. Bile acid metabolism and signaling, the microbiota, and metabolic disease. Pharmacol Ther. 2022;237:108238. [https://doi.org/](https://doi.org/10.1016/j.pharmthera.2022.108238) [10.1016/j.pharmthera.2022.108238](https://doi.org/10.1016/j.pharmthera.2022.108238)
- <span id="page-13-26"></span>57. Kim SG, Kim BK, Kim K, Fang S. Bile acid nuclear receptor farnesoid X receptor: therapeutic target for nonalcoholic fatty liver disease. Endocrinol Metab (Seoul). 2016;31:500– 504. <https://doi.org/10.3803/EnM.2016.31.4.500>
- <span id="page-13-27"></span>58. Juran BD, Lazaridis KN. Is the FXR the fix for cholesterol gallstone disease? Hepatology. 2005;42:218-221. [https://doi.](https://doi.org/10.1002/hep.20776) [org/10.1002/hep.20776](https://doi.org/10.1002/hep.20776)
- <span id="page-13-28"></span>59. Marquardt A, Al-Dabet MM, Ghosh S, et al. Farnesoid X receptor agonism protects against diabetic Tubulopathy: potential add-on therapy for diabetic nephropathy. J Am Soc Nephrol. 2017;28:3182–3189. [https://doi.org/10.1681/asn.](https://doi.org/10.1681/asn.2016101123) [2016101123](https://doi.org/10.1681/asn.2016101123)
- <span id="page-13-29"></span>60. Gao J, Liu X, Wang B, et al. Farnesoid X receptor deletion improves cardiac function, structure and remodeling following myocardial infarction in mice. Mol Med Rep. 2017;16:673–679. <https://doi.org/10.3892/mmr.2017.6643>

- <span id="page-14-0"></span>61. Miyazaki-Anzai S, Masuda M, Levi M, Keenan AL, Miyazaki M. Dual activation of the bile acid nuclear receptor FXR and G-protein-coupled receptor TGR5 protects mice against atherosclerosis. PLoS One. 2014;9:e108270. [https://](https://doi.org/10.1371/journal.pone.0108270) [doi.org/10.1371/journal.pone.0108270](https://doi.org/10.1371/journal.pone.0108270)
- <span id="page-14-1"></span>62. Huang H, Xu Y, Zhu J, Li J. Recent advances in non-steroidal FXR antagonists development for therapeutic applications. Curr Top Med Chem. 2014;14:2175–2187. [https://doi.org/10.](https://doi.org/10.2174/1568026614666141112101840) [2174/1568026614666141112101840](https://doi.org/10.2174/1568026614666141112101840)
- <span id="page-14-2"></span>63. Miyazaki-Anzai S, Levi M, Kratzer A, Ting TC, Lewis LB, Miyazaki M. Farnesoid X receptor activation prevents the development of vascular calcification in ApoE –/– mice with chronic kidney disease. Circ Res. 2010;106:1807–1817. <https://doi.org/10.1161/circresaha.109.212969>
- <span id="page-14-3"></span>64. Jiang L, Zhang H, Xiao D, Wei H, Chen Y. Farnesoid X receptor (FXR): structures and ligands. Comput Struct Biotechnol J. 2021;19:2148–2159. [https://doi.org/10.1016/j.csbj.](https://doi.org/10.1016/j.csbj.2021.04.029) [2021.04.029](https://doi.org/10.1016/j.csbj.2021.04.029)
- <span id="page-14-4"></span>65. Masaoutis C, Theocharis S. The farnesoid X receptor: a potential target for expanding the therapeutic arsenal against kidney disease. Expert Opin Ther Targets. 2019;23:107–116. <https://doi.org/10.1080/14728222.2019.1559825>
- <span id="page-14-5"></span>66. Wang XX, Jiang T, Shen Y, et al. Diabetic nephropathy is accelerated by farnesoid X receptor deficiency and inhibited by farnesoid X receptor activation in a type 1 diabetes model. Diabetes. 2010;59:2916–2927. [https://doi.org/10.2337/](https://doi.org/10.2337/db10-0019) [db10-0019](https://doi.org/10.2337/db10-0019)
- <span id="page-14-6"></span>67. Han SY, Song HK, Cha JJ, Han JY, Kang YS, Cha DR. Farnesoid X receptor (FXR) agonist ameliorates systemic insulin resistance, dysregulation of lipid metabolism, and alterations of various organs in a type 2 diabetic kidney animal model. Acta Diabetol. 2021;58:495–503. [https://doi.](https://doi.org/10.1007/s00592-020-01652-z) [org/10.1007/s00592-020-01652-z](https://doi.org/10.1007/s00592-020-01652-z)
- 68. Wang XX, Wang D, Luo Y, et al. FXR/TGR5 dual agonist prevents progression of nephropathy in diabetes and obesity. J Am Soc Nephrol. 2018;29:118–137. [https://doi.org/](https://doi.org/10.1681/asn.2017020222) [10.1681/asn.2017020222](https://doi.org/10.1681/asn.2017020222)
- <span id="page-14-7"></span>69. Zhou B, Feng B, Qin Z, et al. Activation of farnesoid X receptor downregulates visfatin and attenuates diabetic nephropathy. Mol Cell Endocrinol. 2016;419:72–82. [https://doi.](https://doi.org/10.1016/j.mce.2015.10.001) [org/10.1016/j.mce.2015.10.001](https://doi.org/10.1016/j.mce.2015.10.001)
- <span id="page-14-8"></span>70. Lee FY, Lee H, Hubbert ML, Edwards PA, Zhang Y. FXR, a multipurpose nuclear receptor. Trends Biochem Sci. 2006;31:572–580. <https://doi.org/10.1016/j.tibs.2006.08.002>
- 71. Staels B, Kuipers F. Bile acid sequestrants and the treatment of type 2 diabetes mellitus. Drugs. 2007;67:1383–1392. <https://doi.org/10.2165/00003495-200767100-00001>
- 72. Fiorucci S, Mencarelli A, Palladino G, Cipriani S. Bile-acidactivated receptors: targeting TGR5 and farnesoid-Xreceptor in lipid and glucose disorders. Trends Pharmacol Sci. 2009;30:570–580. [https://doi.org/10.1016/j.tips.2009.08.](https://doi.org/10.1016/j.tips.2009.08.001) [001](https://doi.org/10.1016/j.tips.2009.08.001)
- 73. Fiorucci S, Rizzo G, Donini A, Distrutti E, Santucci L. Targeting farnesoid X receptor for liver and metabolic disorders. Trends Mol Med. 2007;13:298–309. [https://doi.org/10.](https://doi.org/10.1016/j.molmed.2007.06.001) [1016/j.molmed.2007.06.001](https://doi.org/10.1016/j.molmed.2007.06.001)
- 74. Modica S, Moschetta A. Nuclear bile acid receptor FXR as pharmacological target: are we there yet? FEBS Lett. 2006;580:5492–5499. [https://doi.org/10.1016/j.febslet.2006.](https://doi.org/10.1016/j.febslet.2006.07.082) [07.082](https://doi.org/10.1016/j.febslet.2006.07.082)
- <span id="page-14-9"></span>75. Gong J, Zhan H, Li Y, Zhang W, Jin J, He Q. Krüppellike factor 4 ameliorates diabetic kidney disease by activating autophagy via the mTOR pathway. Mol Med Rep. 2019;20: 3240–3248. <https://doi.org/10.3892/mmr.2019.10585>
- <span id="page-14-10"></span>76. Zhang Y, Lee FY, Barrera G, et al. Activation of the nuclear receptor FXR improves hyperglycemia and hyperlipidemia in diabetic mice. Proc Natl Acad Sci U S A. 2006;103:1006– 1011. <https://doi.org/10.1073/pnas.0506982103>
- <span id="page-14-11"></span>77. Gäckler D, Jäkel S, Fricke L, Reinsch B, Fischer F. [Diabetes and kidneys]. Dtsch Med Wochenschr. 2013;138:949–955. <https://doi.org/10.1055/s-0032-1332992>
- <span id="page-14-12"></span>78. Fioretto P, Bruseghin M, Berto I, Gallina P, Manzato E, Mussap M. Renal protection in diabetes: role of glycemic control. J Am Soc Nephrol. 2006;17:S86–S89. [https://doi.org/](https://doi.org/10.1681/asn.2005121343) [10.1681/asn.2005121343](https://doi.org/10.1681/asn.2005121343)
- <span id="page-14-13"></span>79. Ma K, Saha PK, Chan L, Moore DD. Farnesoid X receptor is essential for normal glucose homeostasis. J Clin Invest. 2006;116:1102–1109. <https://doi.org/10.1172/jci25604>
- <span id="page-14-14"></span>80. Bobulescu IA. Renal lipid metabolism and lipotoxicity. Curr Opin Nephrol Hypertens. 2010;19:393–402. [https://doi.org/](https://doi.org/10.1097/MNH.0b013e32833aa4ac) [10.1097/MNH.0b013e32833aa4ac](https://doi.org/10.1097/MNH.0b013e32833aa4ac)
- <span id="page-14-15"></span>81. Jiang T, Wang XX, Scherzer P, et al. Farnesoid X receptor modulates renal lipid metabolism, fibrosis, and diabetic nephropathy. Diabetes. 2007;56:2485–2493. [https://doi.org/](https://doi.org/10.2337/db06-1642) [10.2337/db06-1642](https://doi.org/10.2337/db06-1642)
- <span id="page-14-16"></span>82. Sun L, Halaihel N, Zhang W, Rogers T, Levi M. Role of sterol regulatory element-binding protein 1 in regulation of renal lipid metabolism and glomerulosclerosis in diabetes mellitus. J Biol Chem. 2002;277:18919–18927. [https://doi.org/10.](https://doi.org/10.1074/jbc.M110650200) [1074/jbc.M110650200](https://doi.org/10.1074/jbc.M110650200)
- <span id="page-14-17"></span>83. Singh AB, Dong B, Kraemer FB, Liu J. FXR activation promotes intestinal cholesterol excretion and attenuates hyperlipidemia in SR-B1-deficient mice fed a high-fat and high-cholesterol diet. Physiol Rep. 2020;8:e14387. [https://doi.](https://doi.org/10.14814/phy2.14387) [org/10.14814/phy2.14387](https://doi.org/10.14814/phy2.14387)
- <span id="page-14-18"></span>84. Liu Y, Song A, Yang X, et al. Farnesoid X receptor agonist decreases lipid accumulation by promoting hepatic fatty acid oxidation in db/db mice. Int J Mol Med. 2018;42:1723– 1731. <https://doi.org/10.3892/ijmm.2018.3715>
- <span id="page-14-19"></span>85. Xu S, Jia P, Fang Y, et al. Nuclear farnesoid X receptor attenuates acute kidney injury through fatty acid oxidation. Kidney Int. 2022;101:987–1002. [https://doi.org/10.1016/j.kint.](https://doi.org/10.1016/j.kint.2022.01.029) [2022.01.029](https://doi.org/10.1016/j.kint.2022.01.029)
- <span id="page-14-20"></span>86. Shinohara S, Fujimori K. Promotion of lipogenesis by PPAR<sub>Y</sub>-activated FXR expression in adipocytes. Biochem Biophys Res Commun. 2020;527:49–55. [https://doi.org/10.](https://doi.org/10.1016/j.bbrc.2020.04.075) [1016/j.bbrc.2020.04.075](https://doi.org/10.1016/j.bbrc.2020.04.075)
- <span id="page-14-21"></span>87. Yang J, de Vries HD, Mayeuf-Louchart A, et al. Role of bile acid receptor FXR in development and function of brown adipose tissue. Biochim Biophys Acta Mol Cell Biol Lipids. 2023;1868:159257. [https://doi.org/10.1016/j.bbalip.2022.](https://doi.org/10.1016/j.bbalip.2022.159257) [159257](https://doi.org/10.1016/j.bbalip.2022.159257)
- <span id="page-14-22"></span>88. Lambert G, Amar MJ, Guo G, Brewer HB Jr, Gonzalez FJ, Sinal CJ. The farnesoid X-receptor is an essential regulator of cholesterol homeostasis. J Biol Chem. 2003;278:2563– 2570. <https://doi.org/10.1074/jbc.M209525200>
- <span id="page-14-23"></span>89. Lai CR, Tsai YL, Tsai WC, et al. Farnesoid X receptor overexpression decreases the migration, invasion and angiogenesis of human bladder cancers via AMPK activation and

cholesterol biosynthesis inhibition. Cancers (Basel). 2022;14. <https://doi.org/10.3390/cancers14184398>

- <span id="page-15-0"></span>90. Langhi C, Le May C, Kourimate S, et al. Activation of the farnesoid X receptor represses PCSK9 expression in human hepatocytes. FEBS Lett. 2008;582:949–955. [https://doi.org/10.](https://doi.org/10.1016/j.febslet.2008.02.038) [1016/j.febslet.2008.02.038](https://doi.org/10.1016/j.febslet.2008.02.038)
- <span id="page-15-1"></span>91. Nakahara M, Fujii H, Maloney PR, Shimizu M, Sato R. Bile acids enhance low density lipoprotein receptor gene expression via a MAPK cascade-mediated stabilization of mRNA. J Biol Chem. 2002;277:37229–37234. [https://doi.org/](https://doi.org/10.1074/jbc.M206749200) [10.1074/jbc.M206749200](https://doi.org/10.1074/jbc.M206749200)
- <span id="page-15-2"></span>92. Galkina E, Ley K. Leukocyte recruitment and vascular injury in diabetic nephropathy. J Am Soc Nephrol. 2006;17:368-377. <https://doi.org/10.1681/asn.2005080859>
- <span id="page-15-3"></span>93. Mason RM, Wahab NA. Extracellular matrix metabolism in diabetic nephropathy. J Am Soc Nephrol. 2003;14:1358– 1373. <https://doi.org/10.1097/01.asn.0000065640.77499.d7>
- <span id="page-15-4"></span>94. Navarro JF, Milena FJ, Mora C, León C, García J. Renal proinflammatory cytokine gene expression in diabetic nephropathy: effect of angiotensin-converting enzyme inhibition and pentoxifylline administration. Am J Nephrol. 2006;26:562–570. <https://doi.org/10.1159/000098004>
- <span id="page-15-5"></span>95. Wada J, Makino H. Inflammation and the pathogenesis of diabetic nephropathy. Clin Sci (Lond). 2013;124:139–152. <https://doi.org/10.1042/cs20120198>
- <span id="page-15-6"></span>96. Calle P, Hotter G. Macrophage phenotype and fibrosis in diabetic nephropathy. Int J Mol Sci. 2020;21. [https://doi.org/](https://doi.org/10.1042/cs20120198) [10.1042/cs20120198](https://doi.org/10.1042/cs20120198)
- <span id="page-15-7"></span>97. Chow FY, Nikolic-Paterson DJ, Atkins RC, Tesch GH. Macrophages in streptozotocin-induced diabetic nephropathy: potential role in renal fibrosis. Nephrol Dial Transplant. 2004;19:2987–2996. <https://doi.org/10.1093/ndt/gfh441>
- <span id="page-15-8"></span>98. Li YT, Swales KE, Thomas GJ, Warner TD, Bishop-Bailey D. Farnesoid X receptor ligands inhibit vascular smooth muscle cell inflammation and migration. Arterioscler Thromb Vasc Biol. 2007;27:2606–2611. [https://doi.org/10.1161/atv](https://doi.org/10.1161/atvbaha.107.152694)[baha.107.152694](https://doi.org/10.1161/atvbaha.107.152694)
- <span id="page-15-9"></span>99. Wang YD, Chen WD, Wang M, Yu D, Forman BM, Huang W. Farnesoid X receptor antagonizes nuclear factor kappaB in hepatic inflammatory response. Hepatology. 2008;48:1632– 1643. <https://doi.org/10.1002/hep.22519>
- <span id="page-15-10"></span>100. Jha JC, Banal C, Chow BS, Cooper ME, Jandeleit-Dahm K. Diabetes and kidney disease: role of oxidative stress. Antioxid Redox Signal. 2016;25:657–684. [https://doi.org/10.1089/](https://doi.org/10.1089/ars.2016.6664) [ars.2016.6664](https://doi.org/10.1089/ars.2016.6664)
- <span id="page-15-11"></span>101. Østergaard JA, Cooper ME, Jandeleit-Dahm KAM. Targeting oxidative stress and anti-oxidant defence in diabetic kidney disease. J Nephrol. 2020;33:917–929. [https://doi.org/10.1007/](https://doi.org/10.1007/s40620-020-00749-6) [s40620-020-00749-6](https://doi.org/10.1007/s40620-020-00749-6)
- <span id="page-15-12"></span>102. Lin YC, Chang YH, Yang SY, Wu KD, Chu TS. Update of pathophysiology and management of diabetic kidney disease. J Formos Med Assoc. 2018;117:662-675. [https://doi.](https://doi.org/10.1016/j.jfma.2018.02.007) [org/10.1016/j.jfma.2018.02.007](https://doi.org/10.1016/j.jfma.2018.02.007)
- <span id="page-15-13"></span>103. Wang XX, Jiang T, Shen Y, et al. The farnesoid X receptor modulates renal lipid metabolism and diet-induced renal inflammation, fibrosis, and proteinuria. Am J Physiol Ren Physiol. 2009;297:F1587–F1596. [https://doi.org/10.1152/](https://doi.org/10.1152/ajprenal.00404.2009) [ajprenal.00404.2009](https://doi.org/10.1152/ajprenal.00404.2009)
- <span id="page-15-14"></span>104. Proctor G, Jiang T, Iwahashi M, Wang Z, Li J, Levi M. Regulation of renal fatty acid and cholesterol metabolism, inflammation, and fibrosis in Akita and OVE26 mice with type 1 diabetes. Diabetes. 2006;55:2502–2509. [https://doi.](https://doi.org/10.2337/db05-0603) [org/10.2337/db05-0603](https://doi.org/10.2337/db05-0603)
- <span id="page-15-15"></span>105. Glastras SJ, Wong MG, Chen H, et al. FXR expression is associated with dysregulated glucose and lipid levels in the offspring kidney induced by maternal obesity. Nutr Metab (Lond). 2015;12:40. [https://doi.org/10.1186/s12986-015-](https://doi.org/10.1186/s12986-015-0032-3) [0032-3](https://doi.org/10.1186/s12986-015-0032-3)
- <span id="page-15-16"></span>106. Zhao K, He J, Zhang Y, et al. Activation of FXR protects against renal fibrosis via suppressing Smad3 expression. Sci Rep. 2016;6:37234. <https://doi.org/10.1038/srep37234>
- <span id="page-15-17"></span>107. Qu X, Li X, Zheng Y, et al. Regulation of renal fibrosis by Smad3 Thr388 phosphorylation. Am J Pathol. 2014;184:944– 952. <https://doi.org/10.1016/j.ajpath.2013.12.003>
- <span id="page-15-18"></span>108. Williams ME. Diabetic nephropathy: the proteinuria hypothesis. Am J Nephrol. 2005;25:77–94. [https://doi.org/10.](https://doi.org/10.1159/000084286) [1159/000084286](https://doi.org/10.1159/000084286)
- 109. Campbell RC, Ruggenenti P, Remuzzi G. Proteinuria in diabetic nephropathy: treatment and evolution. Curr Diab Rep. 2003;3:497–504. [https://doi.org/10.1007/s11892-003-](https://doi.org/10.1007/s11892-003-0014-0) [0014-0](https://doi.org/10.1007/s11892-003-0014-0)
- 110. Heyman SN, Raz I, Dwyer JP, Weinberg Sibony R, Lewis JB, Abassi Z. Diabetic proteinuria revisited: updated physiologic perspectives. Cells. 2022;11. [https://doi.org/10.3390/](https://doi.org/10.3390/cells11182917) [cells11182917](https://doi.org/10.3390/cells11182917)
- <span id="page-15-19"></span>111. Hu Z, Ren L, Wang C, Liu B, Song G. Effect of chenodeoxycholic acid on fibrosis, inflammation and oxidative stress in kidney in high-fructose-fed Wistar rats. Kidney Blood Press Res. 2012;36:85–97. <https://doi.org/10.1159/000341485>
- <span id="page-15-20"></span>112. Zhang B, Xie W, Krasowski MD. PXR: a xenobiotic receptor of diverse function implicated in pharmacogenetics. Pharmacogenomics. 2008;9:1695–1709. [https://doi.org/10.2217/](https://doi.org/10.2217/14622416.9.11.1695) [14622416.9.11.1695](https://doi.org/10.2217/14622416.9.11.1695)
- <span id="page-15-21"></span>113. Kliewer SA, Goodwin B, Willson TM. The nuclear pregnane X receptor: a key regulator of xenobiotic metabolism. Endocr Rev. 2002;23:687–702. [https://doi.org/10.1210/er.](https://doi.org/10.1210/er.2001-0038) [2001-0038](https://doi.org/10.1210/er.2001-0038)
- <span id="page-15-22"></span>114. [Luan ZL, Huo XX, Guan YF, Zhang XY. \[Role of pregnane X](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref114) [receptor \(PXR\) in endobiotic metabolism\].](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref114) Sheng Li Xue Bao[. 2019;71:311](http://refhub.elsevier.com/S2468-0249(24)01873-4/sref114)–318.
- <span id="page-15-23"></span>115. Kliewer SA, Moore JT, Wade L, et al. An orphan nuclear receptor activated by pregnanes defines a novel steroid signaling pathway. Cell. 1998;92:73-82. [https://doi.org/10.](https://doi.org/10.1016/s0092-8674(00)80900-9) [1016/s0092-8674\(00\)80900-9](https://doi.org/10.1016/s0092-8674(00)80900-9)
- <span id="page-15-24"></span>116. Orans J, Teotico DG, Redinbo MR. The nuclear xenobiotic receptor pregnane X receptor: recent insights and new challenges. Mol Endocrinol. 2005;19:2891–2900. [https://doi.](https://doi.org/10.1210/me.2005-0156) [org/10.1210/me.2005-0156](https://doi.org/10.1210/me.2005-0156)
- <span id="page-15-25"></span>117. Wu S, Lu H, Wang W, et al. Prevention of D-GalN/LPSinduced ALI by 18ß-glycyrrhetinic acid through PXRmediated inhibition of autophagy degradation. Cell Death Dis. 2021;12:480. <https://doi.org/10.1038/s41419-021-03768-8>
- 118. Ning L, Lou X, Zhang F, Xu G. Nuclear receptors in the pathogenesis and management of inflammatory bowel disease. Mediators Inflamm. 2019;2019:2624941. [https://doi.](https://doi.org/10.1155/2019/2624941) [org/10.1155/2019/2624941](https://doi.org/10.1155/2019/2624941)

- 119. Xing Y, Yan J, Niu YPXR. a center of transcriptional regulation in cancer. Acta Pharmacol Sin B. 2020;10:197–206. <https://doi.org/10.1016/j.apsb.2019.06.012>
- <span id="page-16-0"></span>120. Lv Y, Luo YY, Ren HW, Li CJ, Xiang ZX, Luan ZL. The role of pregnane X receptor (PXR) in substance metabolism. Front Endocrinol (Lausanne). 2022;13:959902. [https://doi.org/10.](https://doi.org/10.1016/j.apsb.2019.06.012) [1016/j.apsb.2019.06.012](https://doi.org/10.1016/j.apsb.2019.06.012)
- <span id="page-16-1"></span>121. Watanabe A, Marumo T, Kawarazaki W, et al. Aberrant DNA methylation of pregnane X receptor underlies metabolic gene alterations in the diabetic kidney. Am J Physiol Ren Physiol. 2018;314:F551–F560. [https://doi.org/10.1152/ajpre](https://doi.org/10.1152/ajprenal.00390.2017)[nal.00390.2017](https://doi.org/10.1152/ajprenal.00390.2017)
- <span id="page-16-2"></span>122. Sonoda J, Chong LW, Downes M, et al. Pregnane X receptor prevents hepatorenal toxicity from cholesterol metabolites. Proc Natl Acad Sci U S A. 2005;102:2198–2203. [https://doi.](https://doi.org/10.1073/pnas.0409481102) [org/10.1073/pnas.0409481102](https://doi.org/10.1073/pnas.0409481102)
- <span id="page-16-3"></span>123. Shizu R, Ezaki K, Sato T, et al. PXR suppresses PPARadependent HMGCS2 gene transcription by inhibiting the interaction between PPARa and PGC1a. Cells. 2021;10. <https://doi.org/10.3390/cells10123550>
- <span id="page-16-4"></span>124. Wolfrum C, Asilmaz E, Luca E, Friedman JM, Stoffel M. Foxa2 regulates lipid metabolism and ketogenesis in the liver during fasting and in diabetes. Nature. 2004;432:1027– 1032. <https://doi.org/10.1038/nature03047>
- <span id="page-16-5"></span>125. Nakamura K, Moore R, Negishi M, Sueyoshi T. Nuclear pregnane X receptor cross-talk with FoxA2 to mediate druginduced regulation of lipid metabolism in fasting mouse liver. J Biol Chem. 2007;282:9768–9776. [https://doi.org/10.](https://doi.org/10.1074/jbc.M610072200) [1074/jbc.M610072200](https://doi.org/10.1074/jbc.M610072200)
- <span id="page-16-6"></span>126. Bhalla S, Ozalp C, Fang S, Xiang L, Kemper JK. Ligandactivated pregnane X receptor interferes with HNF-4 signaling by targeting a common coactivator PGC-1alpha. Functional implications in hepatic cholesterol and glucose metabolism. J Biol Chem. 2004;279:45139–45147. [https://doi.](https://doi.org/10.1074/jbc.M405423200) [org/10.1074/jbc.M405423200](https://doi.org/10.1074/jbc.M405423200)
- <span id="page-16-7"></span>127. Kodama S, Koike C, Negishi M, Yamamoto Y. Nuclear receptors CAR and PXR cross talk with FOXO1 to regulate genes that encode drug-metabolizing and gluconeogenic enzymes. Mol Cell Biol. 2004;24:7931–7940. [https://doi.org/](https://doi.org/10.1128/mcb.24.18.7931-7940.2004) [10.1128/mcb.24.18.7931-7940.2004](https://doi.org/10.1128/mcb.24.18.7931-7940.2004)
- <span id="page-16-8"></span>128. Kodama S, Moore R, Yamamoto Y, Negishi M. Human nuclear pregnane X receptor cross-talk with CREB to repress cAMP activation of the glucose-6-phosphatase gene. Biochem J. 2007;407:373–381. [https://doi.org/10.1042/](https://doi.org/10.1042/bj20070481) [bj20070481](https://doi.org/10.1042/bj20070481)
- <span id="page-16-9"></span>129. Marek CJ, Tucker SJ, Konstantinou DK, et al. Pregnenolone-16alpha-carbonitrile inhibits rodent liver fibrogenesis via PXR (pregnane X receptor)-dependent and PXRindependent mechanisms. Biochem J. 2005;387:601–608. <https://doi.org/10.1042/bj20041598>
- <span id="page-16-10"></span>130. Wright MC. The impact of pregnane X receptor activation on liver fibrosis. Biochem Soc Trans. 2006;34:1119–1123. <https://doi.org/10.1042/bst0341119>
- <span id="page-16-11"></span>131. Luan Z, Wei Y, Huo X, et al. Pregnane X receptor (PXR) protects against cisplatin-induced acute kidney injury in mice. Biochim Biophys Acta Mol Basis Dis. 2021;1867: 165996. <https://doi.org/10.1016/j.bbadis.2020.165996>
- <span id="page-16-12"></span>132. Ming WH, Luan ZL, Yao Y, et al. Pregnane X receptor activation alleviates renal fibrosis in mice via interacting with p53 and inhibiting the Wnt7a/β-catenin signaling. Acta

Pharmacol Sin. 2023;44:2075–2090. [https://doi.org/10.1038/](https://doi.org/10.1038/s41401-023-01113-7) [s41401-023-01113-7](https://doi.org/10.1038/s41401-023-01113-7)

- <span id="page-16-13"></span>133. Holick MF. Vitamin D deficiency. N Engl J Med. 2007;357: 266–281. <https://doi.org/10.1056/NEJMra070553>
- <span id="page-16-14"></span>134. Haussler MR, Haussler CA, Bartik L, et al. Vitamin D receptor: molecular signaling and actions of nutritional ligands in disease prevention. Nutr Rev. 2008;66(10 suppl 2):S98–S112. <https://doi.org/10.1111/j.1753-4887.2008.00093.x>
- <span id="page-16-15"></span>135. Ordóñez-Morán P, Muñoz A. Nuclear receptors: genomic and non-genomic effects converge. Cell Cycle. 2009;8:1675– 1680. <https://doi.org/10.4161/cc.8.11.8579>
- <span id="page-16-16"></span>136. Chokhandre MK, Mahmoud MI, Hakami T, Jafer M, Inamdar AS. Vitamin D & its analogues in type 2 diabetic nephropathy: a systematic review. J Diabetes Metab Disord. 2015;14:58. <https://doi.org/10.1186/s40200-015-0186-6>
- <span id="page-16-17"></span>137. Nair R, Maseeh A. Vitamin D: the "sunshine" vitamin. J Pharmacol Pharmacother. 2012;3:118–126. [https://doi.org/](https://doi.org/10.4103/0976-500x.95506) [10.4103/0976-500x.95506](https://doi.org/10.4103/0976-500x.95506)
- <span id="page-16-18"></span>138. Hu X, Liu W, Yan Y, et al. Vitamin D protects against diabetic nephropathy: evidence-based effectiveness and mechanism. Eur J Pharmacol. 2019;845:91–98. [https://doi.org/10.](https://doi.org/10.1016/j.ejphar.2018.09.037) [1016/j.ejphar.2018.09.037](https://doi.org/10.1016/j.ejphar.2018.09.037)
- <span id="page-16-19"></span>139. Lei M, Liu Z, Guo J. The emerging role of vitamin D and vitamin D receptor in diabetic nephropathy. BioMed Res Int. 2020;2020:4137268. <https://doi.org/10.1155/2020/4137268>
- <span id="page-16-20"></span>140. Li YC. Vitamin D and diabetic nephropathy. Curr Diab Rep. 2008;8:464–469. <https://doi.org/10.1007/s11892-008-0080-4>
- <span id="page-16-21"></span>141. Lavoie JL, Sigmund CD. Minireview: overview of the reninangiotensin system–an endocrine and paracrine system. Endocrinology. 2003;144:2179–2183. [https://doi.org/10.1210/](https://doi.org/10.1210/en.2003-0150) [en.2003-0150](https://doi.org/10.1210/en.2003-0150)
- <span id="page-16-22"></span>142. Chan JC, Ko GT, Leung DH, et al. Long-term effects of angiotensin-converting enzyme inhibition and metabolic control in hypertensive type 2 diabetic patients. Kidney Int. 2000;57:590–600. [https://doi.org/10.1046/j.1523-1755.2000.](https://doi.org/10.1046/j.1523-1755.2000.00879.x) [00879.x](https://doi.org/10.1046/j.1523-1755.2000.00879.x)
- <span id="page-16-23"></span>143. Zhang Z, Sun L, Wang Y, et al. Renoprotective role of the vitamin D receptor in diabetic nephropathy. Kidney Int. 2008;73:163–171. <https://doi.org/10.1038/sj.ki.5002572>
- <span id="page-16-24"></span>144. Kong J, Qiao G, Zhang Z, Liu SQ, Li YC. Targeted vitamin D receptor expression in juxtaglomerular cells suppresses renin expression independent of parathyroid hormone and calcium. Kidney Int. 2008;74:1577–1581. [https://doi.org/10.](https://doi.org/10.1038/ki.2008.452) [1038/ki.2008.452](https://doi.org/10.1038/ki.2008.452)
- <span id="page-16-25"></span>145. Li YC. Renoprotective effects of vitamin D analogs. Kidney Int. 2010;78:134–139. <https://doi.org/10.1038/ki.2009.175>
- <span id="page-16-26"></span>146. Tan X, Wen X, Liu Y. Paricalcitol inhibits renal inflammation by promoting vitamin D receptor-mediated sequestration of NF-kappaB signaling. J Am Soc Nephrol. 2008;19:1741– 1752. <https://doi.org/10.1681/asn.2007060666>
- <span id="page-16-27"></span>147. Sheppard D. Integrin-mediated activation of latent transforming growth factor beta. Cancer Metastasis Rev. 2005;24: 395–402. <https://doi.org/10.1007/s10555-005-5131-6>
- <span id="page-16-28"></span>148. Gressner OA, Gressner AM. Connective tissue growth factor: a fibrogenic master switch in fibrotic liver diseases. Liver Int. 2008;28:1065–1079. [https://doi.org/10.1111/j.1478-](https://doi.org/10.1111/j.1478-3231.2008.01826.x) [3231.2008.01826.x](https://doi.org/10.1111/j.1478-3231.2008.01826.x)
- <span id="page-16-29"></span>149. Zhang YE. Non-Smad pathways in TGF-beta signaling. Cell Res. 2009;19:128–139. <https://doi.org/10.1038/cr.2008.328>
- <span id="page-17-0"></span>150. Li YC. Vitamin D receptor signaling in renal and cardiovascular protection. Semin Nephrol. 2013;33:433-447. [https://](https://doi.org/10.1016/j.semnephrol.2013.07.005) [doi.org/10.1016/j.semnephrol.2013.07.005](https://doi.org/10.1016/j.semnephrol.2013.07.005)
- <span id="page-17-1"></span>151. Deb DK, Zhang Z, Sun T, et al. Vitamin D receptor signaling in podocytes protects against diabetic nephropathy.  $J$  Am Soc Nephrol. 2012;23:1977–1986. [https://doi.org/10.1681/](https://doi.org/10.1681/asn.2012040383) [asn.2012040383](https://doi.org/10.1681/asn.2012040383)
- <span id="page-17-2"></span>152. Kawamata Y, Fujii R, Hosoya M, et al. A G protein-coupled receptor responsive to bile acids. J Biol Chem. 2003;278: 9435–9440. <https://doi.org/10.1074/jbc.M209706200>
- <span id="page-17-3"></span>153. Yang Z, Xiong F, Wang Y, et al. TGR5 activation suppressed S1P/S1P2 signaling and resisted high glucose-induced fibrosis in glomerular mesangial cells. Pharmacol Res. 2016;111:226–236. <https://doi.org/10.1016/j.phrs.2016.05.035>
- <span id="page-17-4"></span>154. Xiong F, Li X, Yang Z, et al. TGR5 suppresses high glucoseinduced upregulation of fibronectin and transforming growth factor-b1 in rat glomerular mesangial cells by inhibiting RhoA/ROCK signaling. Endocrine. 2016;54:657– 670. <https://doi.org/10.1007/s12020-016-1032-4>
- <span id="page-17-5"></span>155. Sato H, Genet C, Strehle A, et al. Anti-hyperglycemic activity of a TGR5 agonist isolated from Olea europaea. Biochem Biophys Res Commun. 2007;362:793–798. [https://doi.org/10.](https://doi.org/10.1016/j.bbrc.2007.06.130) [1016/j.bbrc.2007.06.130](https://doi.org/10.1016/j.bbrc.2007.06.130)
- <span id="page-17-6"></span>156. Ding L, Yang Q, Zhang E, et al. Notoginsenoside Ft1 acts as a TGR5 agonist but FXR antagonist to alleviate high fat dietinduced obesity and insulin resistance in mice. Acta Pharmacol Sin B. 2021;11:1541–1554. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.apsb.2021.03.038) [apsb.2021.03.038](https://doi.org/10.1016/j.apsb.2021.03.038)
- <span id="page-17-7"></span>157. Katsuma S, Hirasawa A, Tsujimoto G. Bile acids promote glucagon-like peptide-1 secretion through TGR5 in a murine enteroendocrine cell line STC-1. Biochem Biophys Res Commun. 2005;329:386–390. [https://doi.org/10.1016/j.bbrc.](https://doi.org/10.1016/j.bbrc.2005.01.139) [2005.01.139](https://doi.org/10.1016/j.bbrc.2005.01.139)
- <span id="page-17-8"></span>158. Watanabe M, Houten SM, Mataki C, et al. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. Nature. 2006;439:484–489. [https://doi.org/](https://doi.org/10.1038/nature04330) [10.1038/nature04330](https://doi.org/10.1038/nature04330)
- <span id="page-17-9"></span>159. Xiao H, Sun X, Liu R, et al. Gentiopicroside activates the bile acid receptor Gpbar1 (TGR5) to repress NF-kappaB pathway and ameliorate diabetic nephropathy. Pharmacol Res. 2020;151:104559. <https://doi.org/10.1016/j.phrs.2019.104559>
- <span id="page-17-10"></span>160. Hardie DG, Ross FA, Hawley SA. AMPK: a nutrient and energy sensor that maintains energy homeostasis. Nat Rev

Mol Cell Biol. 2012;13:251–262. [https://doi.org/10.1038/](https://doi.org/10.1038/nrm3311) [nrm3311](https://doi.org/10.1038/nrm3311)

- <span id="page-17-11"></span>161. Bause AS, Haigis MC. SIRT3 regulation of mitochondrial oxidative stress. Exp Gerontol. 2013;48:634-639. [https://doi.](https://doi.org/10.1016/j.exger.2012.08.007) [org/10.1016/j.exger.2012.08.007](https://doi.org/10.1016/j.exger.2012.08.007)
- <span id="page-17-12"></span>162. Neuschwander-Tetri BA, Loomba R, Sanyal AJ, et al. Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (flint): a multicentre, randomised, placebo-controlled trial. Lancet. 2015;385:956–965. [https://doi.org/10.1016/s0140-6736\(14\)](https://doi.org/10.1016/s0140-6736(14)61933-4) [61933-4](https://doi.org/10.1016/s0140-6736(14)61933-4)
- <span id="page-17-13"></span>163. Mudaliar S, Henry RR, Sanyal AJ, et al. Efficacy and safety of the farnesoid X receptor agonist obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver disease. Gastroenterology. 2013;145:574–582.e571. [https://doi.org/](https://doi.org/10.1053/j.gastro.2013.05.042) [10.1053/j.gastro.2013.05.042](https://doi.org/10.1053/j.gastro.2013.05.042)
- <span id="page-17-14"></span>164. Armstrong LE, Guo GL. Role of FXR in liver inflammation during nonalcoholic steatohepatitis. Curr Pharmacol Rep. 2017;3:92–100. <https://doi.org/10.1007/s40495-017-0085-2>
- 165. Xu W, Lu C, Zhang F, Shao J, Yao S, Zheng S. Dihydroartemisinin counteracts fibrotic portal hypertension via farnesoid X receptor-dependent inhibition of hepatic stellate cell contraction. FEBS Journal. 2017;284:114-133. [https://](https://doi.org/10.1111/febs.13956) [doi.org/10.1111/febs.13956](https://doi.org/10.1111/febs.13956)
- 166. Gai Z, Visentin M, Gui T, et al. Effects of farnesoid X receptor activation on arachidonic acid metabolism, NF-kB signaling, and hepatic inflammation. Mol Pharmacol. 2018;94:802–811. <https://doi.org/10.1124/mol.117.111047>
- <span id="page-17-15"></span>167. Comeglio P, Filippi S, Sarchielli E, et al. Anti-fibrotic effects of chronic treatment with the selective FXR agonist obeticholic acid in the bleomycin-induced rat model of pulmonary fibrosis. J Steroid Biochem Mol Biol. 2017;168:26–37. <https://doi.org/10.1016/j.jsbmb.2017.01.010>
- 168. Su J, Zhang Q, Qi H, et al. The G-protein-coupled bile acid receptor Gpbar1 (TGR5) protects against renal inflammation and renal cancer cell proliferation and migration through antagonizing NF-KB and STAT3 signaling pathways. Oncotarget. 2017;8:54378–54387. [https://doi.org/10.18632/onco](https://doi.org/10.18632/oncotarget.17533)[target.17533](https://doi.org/10.18632/oncotarget.17533)
- 169. Liu H, Hu C, Zhang X, Jia W. Role of gut microbiota, bile acids and their cross-talk in the effects of bariatric surgery on obesity and type 2 diabetes. J Diabetes Investig. 2018;9: 13–20. <https://doi.org/10.18632/oncotarget.17533>