For reprint orders, please contact: reprints@future-science.com

The enigmatic helicase DHX9 and its

association with the hallmarks of cancer

¹Institute of Cardiovascular & Medical Science, College of Medical, Veterinary & Life Sciences, University of Glasgow, Glasgow, UK

²Philips Research Europe, High Tech Campus, Eindhoven, The Netherlands

*Author for correspondence: Tel.: +44 141 330 1662; George.Baillie@glasgow.ac.uk

Chloe Gulliver¹, Ralf Hoffmann^{1,2} & George S Baillie*,¹

Much interest has been expended lately in characterizing the association between DExH-Box helicase 9 (DHX9) dysregulation and malignant development, however, the enigmatic nature of DHX9 has caused conflict as to whether it regularly functions as an oncogene or tumor suppressor. The impact of DHX9 on malignancy appears to be cell-type specific, dependent upon the availability of binding partners and activation of inter-connected signaling pathways. Realization of DHX9's pivotal role in the development of several hallmarks of cancer has boosted the enzyme's potential as a cancer biomarker and therapeutic target, opening up novel avenues for exploring DHX9 in precision medicine applications. Our review discusses the ascribed functions of DHX9 in cancer, explores its enigmatic nature and potential as an antineoplastic target.

Lay abstract: DExH-Box helicase 9 (DHX9) is an enzyme with multiple important functions in cells, therefore, its deregulated activity can cause alterations in cellular growth and subsequent formation of tumors. In particular, irregular DHX9 activity is attributed to the development of several hallmarks of cancer, however, it can have both pro- and anti-cancer effects, thus leading to conflicting views on DHX9's role in cancer development. The diverse implications of DHX9 in malignancy exposes the protein as a potential marker for cancer detection and intervention. Our review discusses the vast array of DHX9's functions in cancer development, and its potential as a target for cancer treatment.

First draft submitted: 18 August 2020; Accepted for publication: 20 October 2020; Published online: 2 November 2020

Keywords: biomarker • cancer • DHX9 • helicase • medicine • target • tumor

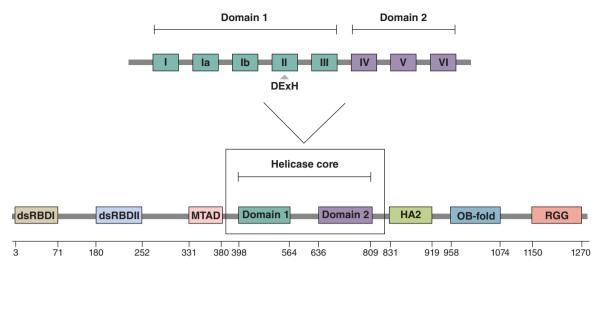
Helicase proteins have a diverse range of cellular functions that stem from their ability to resolve and remodel DNA and RNA structures. DExH-Box helicase 9 (DHX9), also known as RNA helicase A (RHA), belongs to the DExD/H-Box superfamily II of helicases [1]. The profile of DHX9 has rapidly increased in the oncology field lately because of two main factors. Firstly, the enzyme has been implicated in the control of many cellular processes which are often deregulated in cancer, such as genomic stability, transcription and DNA replication [2,3]. Secondly, DHX9 is overexpressed in many tumors including lung and colorectal cancers and for this reason, it has recently been proposed as a potential therapeutic target [4–6]. This review focuses on the role of DHX9 in the development and progression of cancer, highlights DHX9 as a potential biomarker for the evaluation of cancer progression and discusses strategies for targeting DHX9 as novel therapeutic avenues for cancer therapy.

DExD/H-box RNA helicases

The unwinding and remodeling of DNA/RNA structures is ATP dependent and involves the catalytic action of helicases [7]. DExD/H-box RNA helicases are categorized as members of the helicase superfamily II by virtue of their conserved motif DExD/H amino acid signature sequence, which represents the core domain at which NTP binding and hydrolysis occur. While structurally similar, the DExD and DExH families contain slight diversity within the conserved DExD/H motifs [8]. Another difference between the families is that DExH helicases are able to hydrolyze all NTPs, while DExD helicases are specifically ATP driven [9]. It should be noted that although both types are best known for their ability to unwind RNA, they also have important roles in RNA



Future Science



Amino acid

Figure 1. Human DExH-Box helicase 9 protein structure, domains and motifs. The entire protein is 1280 amino acids in length, with amino acid positions of domains and motifs shown as determined by UNIPROT. The helicase core domain consists of two distinct regions – the helicase ATP-binding domain (domain 1) in which the DExH motif is located, and the helicase C-terminal (domain 2).

dsRBD I/II: Double-stranded RNA-binding domain I/II; Domain 1: Helicase ATP-binding domain; Domain 2: Helicase C-terminal domain; HA2: Helicase-associated domain 2; MTAD: Minimal transactivation domain; OB-fold: Oligonucleotide- or oligosaccharide-binding fold; RGG: Repeated arginine and glycine-glycine.

metabolism via remodeling ribonucleoprotein complexes, processing pre-mRNA and catalyzing ribosomal RNA biogenesis [10,11]. These functions are essential in the regulation of transcription and translation, and coordination of gene expression [1,12].

DHX9 general features & functions

The *DHX9* gene, a DExH-box helicase family member, maps to chromosome 1q25 in humans, a location that has been identified as the major prostate cancer (PCa) susceptibility locus [13]. The encoded protein has a molecular weight of 140 kDa and is characterized by eight domains, consisting of dsRNA-binding domains (dsRBD) I and II at the N-terminus, a core helicase domain in the center and a repeated arginine and glycine-glycine-rich region at the C-terminus (Figure 1) [14]. DHX9 is an abundant, nuclear protein with a variety of putative roles including maintenance of genomic stability, DNA replication, transcription and translation [15]. It shuttles between the nucleus and cytoplasm and is capable of binding both ssDNA and ssRNA, as well as unwinding double-stranded (ds) nucleic acids and complex polynucleotide structures. While it can act on multiple nucleotide structures, it appears to have a preference for RNA displacement loops (R-loops), and both RNA and DNA G-quadruplexes [16].

DHX9 & maintenance of genomic stability

Genomic integrity is maintained through the DNA damage response, whereby cells undergo cell cycle arrest to initiate repair pathways or, if irreparable, induce cell death [17]. DHX9 mediates genomic stability via resolution of R-loop-associated DNA damage [3]. R-loops form during transcription when nascent RNA leaves RNA Polymerase II (RNA Pol II) and forms an RNA/DNA hybrid with the template DNA strand, subsequently displacing the nontemplate DNA strand [18,19]. While R-loops facilitate regulation of gene expression, their presence can contribute to genomic instability in several ways. For example, they can cause transcription/replication conflicts leading to stalled replication forks and DNA breaks, and their presence also leaves the displaced nontemplate strand susceptible to DNA damage [20,21]. DHX9 plays a fundamental role in maintaining genomic stability by interacting with RNA/DNA hybrids, resolving these structures and facilitating R-loop suppression [3,16,22]. Interestingly, however, a recent study revealed a conflicting ability of DHX9 to promote R-loop formation in cells with defective

splicing machinery. In the absence of splicing factors to stabilize unwound nascent RNA, DHX9 activity promotes development of an RNA/DNA hybrid and subsequent generation of an R-loop (as shown in Figure 2A), thus highlighting the cellular-context-dependent mechanisms of DHX9 in regulation of genomic stability [23].

An important point to note is that DHX9's ability to interact with DNA repair proteins allows the helicase to mediate their recruitment to R-loops in order to promote repair of R-loop-associated DNA damage. For example, DHX9 links BRCA1 to the RNA Pol II complex [24], which can explain the recruitment of BRCA1 to R-loops where it has been proposed to interact with senataxin to promote DNA repair and R-loop suppression [25,26]. Furthermore, BRCA1 is associated with stalled replication forks whereby it functions in fork protection, R-loop resolution at transcription/replication conflicts and DNA repair (Figure 2B) [27]. Interestingly, R-loops have also been shown to facilitate DNA damage repair following their formation near double-stranded breaks (DSBs) [28]. Accumulation of R-loops at these sites is essential for recruitment of BRCA1 to mediate homologous recombination (HR) [29]. Upon recognition of DNA damage, DHX9 forms a complex with BRCA1 in order to facilitate translocation to sites of DSBs. Recruitment of this complex promotes DNA end resection and subsequent RPA binding required for HR progression and DSB repair (Figure 2C) [30]. Conceptually, it is likely that DHX9-mediated recruitment of BRCA1 to RNA Pol II at transcription pause sites or DSBs functions as a complex to prevent/resolve R-loops, alleviate replication stress and mediate DNA damage repair [25,30]. Crucially, a fine balance is required between R-loop formation and resolution in order to regulate gene transcription without threatening genomic stability, and DHX9 plays a fundamental role in maintaining this balance [3,31]. Taken together, these studies highlight the critical role of DHX9 in the maintenance of genomic stability.

DNA replication

The aforementioned role of DXH9 in the resolution of transcription/replication conflicts prevents replication stress by allowing the replication machinery to function efficiently (Figure 2B) and this highlights a role for DHX9 in regulation of DNA replication [30]. Consistent with this notion, DHX9 binds to, and enhances the activity of, an enzyme called WRN helicase that is involved in both DNA replication and the maintenance of genomic integrity [32,33]. It is suggested that the DHX9-WRN complex functions in DNA replication and/or repair to resolve structures that could impede DNA replication machinery [7]. While the exact function of DHX9 in DNA replication, subsequently inducing p53-dependent senescence in human diploid fibroblasts. Such data frank the importance of DHX9 in this process [34].

Transcription & translation

As noted above, R-loops modulate the regulation of gene expression and their coformation with G quadruplexes (G4s) further stabilizes structures, which can impair transcription elongation [16,35]. While DHX9 is capable of unwinding many types of nucleotide structures, it shows preference for R-loops and DNA G4s, resolving these structures to promote unhindered transcription [16]. However, it is important to note that DHX9's conflicting ability to promote R-loop formation in the absence of splicing factors [23] exposes the dual abilities of this helicase in regulating transcription in a cell-type-specific manner (Figure 2A).

DHX9 can also influence transcriptional activation via binding to the promoter regions of numerous genes, including *p16^{INK4a}* [36] and *MDR1* [37]. Likewise, DHX9 forms complexes with nuclear EGFR to mediate binding to target gene promoters following EGF-stimulated EGFR nuclear translocation [38]. While DHX9 bridging of BRCA1 to RNA Pol II [24] has been discussed above, BRCA1 is thought to function as both a transcriptional coactivator and repressor, thereby also implicating this complex in transcriptional regulation [39]. Similarly, DHX9 acts as a scaffold to bridge the transcriptional coactivator CBP (CREB-binding protein) to RNA Pol II, stimulating CREB-dependent transcriptional activation of target genes [40]. This interaction with CBP was also shown to facilitate recruitment of CBP to the p65 subunit of NF-kB following DHX9-p65 binding, stimulating NF-kB-mediated transcriptional activation [41]. However, a recent study revealed that p65 is recruited to preloaded CBP/p300 on p65-target gene promoters [42], leading to speculation that DHX9 binds p65 to facilitate recruitment to CBP at target promoter sites, rather than recruiting CBP to p65.

A major function of DHX9 is processing of secondary structures on mRNA, and an inability to resolve these structures can lead to hindered translation initiation [43]. One example is the formation of RNA G4s (rG4s) at the 5' untranslated region (5' UTR) of mRNA, which can cause stalling of the preinitiation complex and repress translation [44]. DHX9 regulates translational efficiency via binding and unwinding rG4 structures to stimulate the

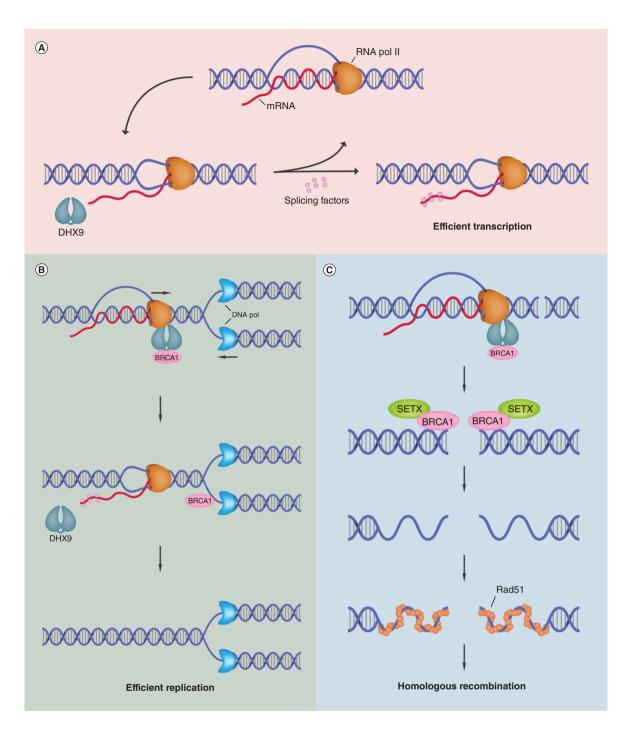


Figure 2. DExH-Box helicase 9-mediated R-loop resolution in the maintenance of genomic stability, transcription and DNA replication. (A) DHX9 associates with RNA Pol II and functions in the resolution of R-loops by unwinding nascent RNA. The unwound mRNA is then bound by splicing factors to prevent strand invasion and thus transcription continues efficiently. In the absence of splicing factors, the unwound mRNA can invade duplex DNA to re-form an RNA/DNA hybrid and generate an R-loop. (B) DHX9-BRCA1 interaction recruits BRCA1 to sites of transcription/replication conflicts. Upon collision of transcription and replication machinery, DHX9 functions in R-loop resolution while BRCA1 functions in protection and repair of stalled replication forks and associated DNA damage. Upon removal of R-loops, RNA Pol II can dissociate allowing efficient DNA replication. (C) DHX9 recruits BRCA1 to sites of states of dsDNA breaks. Formation of R-loops near double-stranded break sites recruits BRCA1 where it interacts with SETX in order to mediate end resection and initiate homologous recombination, resulting in repaired DNA and maintenance of genomic stability.

DHX9: DExH-Box helicase 9; DNA Pol: DNA polymerase; mRNA: Messenger RNA; RNA Pol II: RNA polymerase II; SETX: Senataxin.

initiation of translation [44,45]. Conversely, DHX9 also participates in post-transcriptional repression via a role in RNA interference. DHX9 can promote the assembly of the RNA-induced silencing complex (RISC) to drive gene silencing [46,47]. Similarly, DHX9's interaction with BRCA1 promotes maturation of primary transcripts of miRNA (pri-miRNA), thus participating in miRNA processing, from which miRNAs mediate translational suppression and mRNA cleavage [12,48].

Innate immunity

The enigmatic nature of DHX9 also extends to the field of innate immunity where the helicase has conflicting roles in both the defense against viral infections and contribution to viral replication [49]. As previously stated, DHX9 mediates NF- κ B-dependent gene transcription [41]. In response to DNA virus infection, DHX9 induces NF- κ B-mediated transcription of antiviral cytokines in fibroblasts and epithelial cells, thus enhancing the antiviral immune response of permissive cells against DNA viruses [50]. In addition to this, DHX9 has been identified as a cytosolic viral DNA sensor in plasmacytoid dendritic cells, dependent on interaction with MyD88 [51], as well as a viral dsRNA sensor in myeloid dendritic cells [52], actions that are required to trigger NF- κ B activation and subsequent induction of the antiviral cytokine response. In apparent opposition to these protective actions, DHX9 also facilitates the replication of numerous viruses, including hepatitis B virus (HBV) [53,54], hepatitis C virus [55], human immunodeficiency virus [56] and chikungunya virus [57]. In particular regarding HBV, it has recently been revealed that the HBV protein X promotes upregulated DHX9 expression, while DHX9 facilitates HBV DNA replication dependent on an interaction with Nup98 to stimulate DHX9's helicase activity [53]. Another recent study also revealed that DHX9 promotes HBV DNA replication via inhibiting the apolipoprotein B mRNA-editing enzyme catalytic polypeptide-like 3B (A3B), which usually functions to impede HBV DNA replication [54].

DHX9 in cancer

The hallmarks of cancer refer to biological traits acquired by cells during the development of cancer, which include enhanced proliferative signaling, evasion of growth suppression, replicative immortality, resistance to cell death, and promotion of angiogenesis, invasion and metastasis [58]. DHX9 plays a fundamental role in mediating genomic stability as well as regulating cellular processes preceding the development of numerous cancer hallmarks [16,22]. Aberrant DHX9 activity leading to genomic instability and dysregulation of molecular events can have profound pathological consequences including malignant development, and it is clear that overexpression of DHX9 is a characteristic of many cancer types [5,6,59]. From this point onward, this review will focus on the implications of DHX9 in malignant development and discuss the potential exploitation of DHX9 as an antineoplastic therapeutic target.

DHX9 in oncogene & tumor suppressor regulation

An accumulation of genetic and epigenetic changes drives malignant development and this manifests via dysregulation of oncogenes and tumor suppressor genes in most cancers. Proto-oncogenes stimulate processes such as growth, proliferation and survival, while tumor suppressor genes promote DNA repair, cell cycle arrest, growth suppression and cell death pathways [60]. Aberrant activation of proto-oncogenes into oncogenes, and inactivation of tumor suppressor genes, drives oncogenic signaling pathways while diminishing tumor suppressive mechanisms, thus conferring a selective advantage to the tumor [61]. DHX9 activity has been identified as a factor in the transcription of both oncogenes and tumor suppressor genes in human cancers. Additionally, R-loop perturbations that occur in most cancers induce aberrant transcription of oncogenes and tumor suppressor genes, suggesting that DHX9's function in R-loop resolution may be defective in cancer cells [62]. Furthermore, DHX9 transcriptionally regulates both oncogenes and tumor suppressor genes, or modulates the activity of their encoded proteins (see below), however, whether DHX9 functions as an oncoprotein or tumor suppressor is a current topic of debate.

DHX9 in sustained proliferative signaling

Sustained proliferation of cancer cells can be achieved through the upregulation of a variety of proliferative signaling pathways [58]. While there has been little work characterizing DHX9's role in proliferative signaling, the helicase's ability to regulate transcription of oncogenes and tumor suppressor genes could promote the activation of proliferative signaling transduction routes. A recent multiomics approach identified notable activation and connection between DHX9, EED and AURKA genomic, proteomic and metabolomic networks in breast tumors, which correlated with activation of proliferative signaling pathways, such as RAS, PI3K and Rb/E2F [63]. Furthermore, DHX9 lies downstream of the transcription factor SOX4, which suggests that DHX9 may be a SOX4 transcriptional target [64]. *SOX4* has been proposed as an oncogene because of its frequent overexpression in cancers, including prostate and breast cancer. SOX4 also has involvement in proliferative signaling pathways such as Wnt and PI3K [65–67]. In PCa, SOX4 forms a nuclear complex with plankoglobin following induction of Wnt signaling. This complex affects SOX4-DHX9 binding causing DHX9 suppression, highlighting SOX4's role in modulating DHX9 expression in PCa progression and the SOX4-DHX9 complex's role in Wnt signaling [68].

DHX9 in evasion of growth suppressors & replicative immortality

Evasion of growth suppression and dysregulated cell cycle control not only leads to replicative immortality but can also contribute to cell proliferation [58]. Overexpression of the EGFR oncogene has been identified in many cancers that are a product of aberrant EGFR signaling [69]. As previously mentioned, DHX9 mediates EGFR binding to target gene promoters, including the CCND1 proto-oncogene that stimulates cyclin D1 transcriptional activation (Figure 3A), as observed in human breast cancer cells [38,70]. Cyclin D1 is a cell cycle regulator that promotes G1/S transition upon interaction with cyclin-dependent kinase-4/-6 (CDK4/6). The retinoblastoma protein (Rb) functions as a growth suppressor, by binding E2F transcription factors to repress E2F-target gene transcription. The cyclin D1-CDK4/6 complex catalyzes hyperphosphorylation of Rb to inhibit its activity in order to stimulate cell cycle progression and cell proliferation [71]. Overexpression of cyclin D1 is frequently observed in cancers, including breast [72], pancreatic [73] and lung cancer [74]. It therefore appears that DHX9 may contribute to the overexpression of cyclin D1 in cancer following oncogenic EGFR upregulation, thus promoting proliferation and replicative immortality. DHX9 also forms a cocomplex with RNA Pol II and EWS-FLI1 to augment EWS-FLI1-dependent CCND1 transcription (Figure 3B) [15]. Interestingly, however, upon inhibition of this interaction, DHX9 associates with Sam68, recruiting it to *pncCCND1_B* (a *CCND1* promoter-associated noncoding RNA), ultimately repressing CCND1 transcription. This work highlights DHX9's ability to form alternative complexes with either EWS-FLI1 or pncCCND1_B/Sam68 in modulating CCND1 expression in Ewing sarcoma cells, exhibiting both oncogenic and tumor suppressive functions [75]. In this context, it should also be noted that DHX9 binds to the p16^{INK4a} promoter and functions as a transcriptional coactivator (Figure 3C) [36]. p16^{INK4a} is a tumor suppressor that functions to inhibit the cyclin D1-CDK4/6 complex thus, promoting cell cycle regulation, however, in cancer cells this interaction is often suppressed, leading to transcriptional silencing of $p16^{INK4a}$ that results in uncontrolled G1/S transition and replicative immortality [36,76]. This highlights the tumor suppressive role of DHX9 in modulating p16^{INK4a} transcription on cell cycle control. In a similar vein, DHX9 can also mediate CDK6 downregulation, thereby supporting the helicase's tumor suppressive functions that are compromised in hepatocellular carcinoma (HCC) due to the overexpressed long noncoding (lnc) RNA lnc-UCID. CDK6 binds to lnc-UCID sequestering the kinase from DHX9, preventing DHX9-mediated CDK6 downregulation and leading to dysregulated G1/S transition and uncontrolled cell proliferation [59].

DHX9 is also capable of suppressing activity of the tumor suppressor BRCA1, which functions in the DNA damage response and cell cycle arrest [77,78]. BRCA1 interacts with hypophosphorylated Rb, which is suspected to maintain Rb in its E2F-bound activated state, preventing E2F-target gene transcription and inhibiting cell proliferation [79]. As previously discussed, DHX9 interacts with BRCA1 to coordinate HR and maintain genomic integrity [30], however, overexpression of a DHX9 peptide fragment containing the region for BRCA1 interaction was shown to inhibit BRCA1 activity in breast epithelial cells and promote breast cancer progression [77]. Additionally, truncated DHX9 mutants affect the interaction between BRCA1 and RNA pol II, subsequently leading to decreased BRCA1 transcription [80]. Decreased BRCA1 transcription can result in defective cell cycle checkpoints and enhanced proliferation [78,81]. Taken together, these studies reveal that DHX9 can contribute to BRCA1 suppression and subsequent Rb dysregulation, promoting genomic instability, replicative immortality and proliferation.

DHX9 in evasion of cell death

p53 is a tumor suppressor that has been named 'the guardian of the genome' as it promotes cell cycle arrest or induction of apoptosis upon detection of DNA damage via enhanced production of proapoptotic proteins. p53 is commonly mutated or deficient in cancer, therefore allowing damaged cells to evade apoptosis, accumulate and promote cancer progression [82]. As previously noted, DHX9 is overexpressed in many cancers and inhibition of DHX9 evokes p53-mediated apoptosis in numerous human cancer cell lines, highlighting DHX9's involvement in this pathway [83]. Interestingly, a follow-up study also identified p53-independent induction of cell death following *DHX9* inhibition in p53-deficient mouse lymphomas and colon cancer cells, highlighting the dependency of both

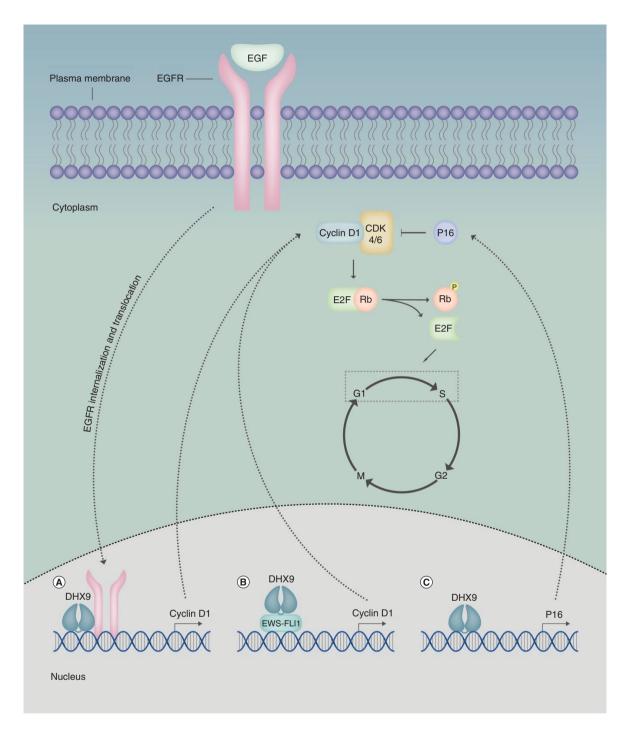


Figure 3. DExH-Box helicase 9 participates in the transcription of both cyclin D1 and p16^{INK4A}, exhibiting both oncogenic and tumor suppressive functions. (A) Following EGF-stimulated nuclear translocation of *EGFR*, DHX9 facilitates EGFR-mediated cyclin D1 transcription. **(B)** DHX9 interacts with EWS-FL11 in Ewing sarcoma cells, promoting EWS-FL11-mediated cyclin D1 transcription. **(C)** DHX9 binds to the p16^{INK4A} promoter, functioning as a transcriptional coactivator to induce *p16* transcriptional. Cyclin D1 binds CDK4/6 to induce dissociation of Rb and E2F, stimulating G1/S transition to promote replicative immortality. P16^{INK4A} functions as a tumor suppressor by inhibiting the cyclin D1-CDK4/6 complex, thus allowing Rb to inhibit E2F-mediated transcription of genes promoting G1/S transition. In cancer cells, the interaction between DHX9 and p16^{INK4A} is often suppressed, thus diminishing its tumor suppressive abilities.

CDK4/6: Cyclin-dependent kinase-4/-6; DHX9: DExH-Box helicase 9; EGFR: EGF receptor; *p16*: p16INK4a; Rb: Retinoblastoma protein.

p53-wildtype and p53-deficient tumors on DHX9 for survival [83,84]. Conversely, nuclear DHX9 is essential for KIF1B isoform β (KIF1B β)-mediated neuronal apoptosis. In cells with aberrant nerve growth factor signaling, the tumor suppressor KIF1B β mediates nuclear localization and accumulation of DHX9, which is necessary for KIF1B β -induced apoptosis of neuronal cells. However, *KIF1B\beta* is often deleted in neuroblastoma tumors, thus impairing nuclear DHX9 localization and subsequently allowing neuroblastoma cells to evade apoptosis [85]. This work further highlights the context-dependent manner of DHX9 in promoting either tumor suppression or tumorigenesis and adds another layer of contradiction to this enigmatic helicase.

DHX9 in angiogenesis, metastasis & invasion

One method by which DHX9 activity has been shown to influence cancer cell invasion is via interactions with IncRNAs leading to modulation of their activity. Lnc-cervical cancer DHX9 suppressive transcript (Inc-CCDST) binds DHX9, acts as a scaffold to enhance MDM2-DHX9 binding and thus induces DHX9 degradation. In cervical cancer (CC) cells, Inc-CCDST is downregulated via the human papillomavirus-encoded E6 and E7 oncoproteins, allowing DHX9 to accumulate and result in enhanced invasion and angiogenesis [86]. DHX9 is also implicated in the production and regulation of circRNAs, another type of noncoding RNA which can serve as miRNA sponges in order to regulate gene expression [87,88]. Recently, it has been discovered that DHX9 phosphorylation can promote upregulation of oncogenic circCCDC66, which correlates with colorectal cancer (CRC) growth, invasion and metastasis [89,90]. However, circRNAs can also possess tumor suppressive functions, for example, in the case of cSMARCA5 that inhibits growth and migration of HCC cells. DHX9 upregulation induces cSMARCA5 downregulation, suppressing its tumor suppressive functions to promote HCC progression and metastasis [91]. Overexpression of DHX9 has previously been identified in osteosarcoma cells exhibiting high metastatic ability [92], and these studies highlight the ability of DHX9 to both promote oncogenic and inhibit tumor suppressive circRNAs to promote invasion, migration and metastasis in CRC and HCC. Additionally, circ0005276 has been associated with proliferation, invasion and migration in PCa. circ0005276 regulates XIAP transcription via interaction with FUS, with both circ0005276 and XIAP overexpressed in PCa cells and tissues [93]. Interestingly, DHX9 has been shown to interact with FUS in HEK293 cells [94], which positions DHX9 to mediate FUS-circ0005276-mediated *XIAP* expression, and subsequent PCa invasion and migration.

Epithelial-mesenchymal transition (EMT) is a critical cellular process occurring during development that is characterized by a shift from epithelial to mesenchymal phenotype, however, cancer cells can hijack this event to promote transition to the more invasive mesenchymal phenotype, permitting escape from the primary tumor, invasion into the bloodstream and metastasis to distant locations [95]. Many different circRNAs have been associated with EMT in cancer cells [96], and DHX9 plays fundamental roles in modulating circRNAs during EMT. For example, DHX9 expression in bladder cancer (BC) tissue suppresses circPICALM action, which usually functions as a tumor suppressor by 'sponging' for miR-1265, preventing FAK phosphorylation and EMT. This is a prime example of how DHX9 overexpression can overcome tumor suppressive protection in BC and negatively influence EMT to promote BC invasion and metastasis [97]. Furthermore, as previously noted, DHX9 phosphorylation augments circCCDC66 expression in CRC [90], and circCCDC66 levels have been associated with the mesenchymal phenotype in lung cancer, which is likely regulated by FAK [98]. Therefore, DHX9's involvement in circCCDC66 regulation, and putative cross-talk with FAK phosphorylation and EMT upon DHX9-mediated circPICALM suppression, further implicates it in cancer cells' acquisition of a mesenchymal phenotype and enhanced invasive features. As with most functions of DHX9, there is a converse role for the helicase in lung adenocarcinoma where DHX9 functions as a tumor suppressor and inhibits EMT. DHX9 knockdown in these cells induced STAT3 phosphorylation and subsequent proliferation, migration and invasion in vitro [5]. Upregulated STAT3 has been shown to promote EMT in non-small-cell lung cancer [99], therefore, it is speculated that DHX9 mediates EMT via STAT3 regulation in lung adenocarcinoma [5].

DHX9 as a biomarker & therapeutic target

DHX9 as a biomarker for chemotherapy resistance

While the literature franking the involvement of DHX9 in the development and progression of cancer is robust and plentiful, a newly discovered feature of DHX9 revolves around resistance to chemotherapeutic agents. The majority of chemotherapy drugs, such as platinum-derived agents, target highly proliferative cells and induce DNA damage that triggers cellular DNA damage responses and repair pathways, however, upon irreversible DNA damage cell death is programmed via apoptosis [100]. Resistance to DNA-damage inducing chemotherapeutics can occur

via upregulated DNA repair responses leading to sufficient repair and oncogenic cell survival, therefore, DHX9's aforementioned involvement in regulating these processes position the helicase as an influencer of chemotherapy [3,30,101]. It is thought that post-translational modification of DHX9 occurs in drug-resistant cells, with a higher molecular weight observed in drug-resistant than drug-sensitive leukemic cells [102]. DNA-PK-mediated DHX9 phosphorylation upregulates MDR1 expression, and subsequently drives multidrug resistance in leukemia cells [37]. Additionally in CRC, DHX9-mediated circCCDC66 upregulation promotes resistance to oxaliplatin, while inhibiting DHX9 phosphorylation suppressed circCCDC66 expression and chemoresistance, highlighting DHX9 activity in the development of oxaliplatin-resistant CRC cells [90]. Contrary to this, a proteomics-based study identified DHX9 as a key protein downregulated in temozolomide-resistant glioblastoma, and that the spliceosome signaling pathway was affected in these cells [103]. Given that DHX9 is regulated via alternative splicing [15], dysregulation of this pathway may aberrantly affect DHX9 synthesis and promote temozolomide-resistance. This study recognized DHX9's potential as a biomarker in glioblastoma, with low expression correlating with a less favorable prognosis. Another study identified DHX9 as a binding partner of Schlafen-11 (SLFN11), a proposed predictive biomarker of resistance to platinum-based chemotherapeutics in ovarian and lung cancers. It is speculated that diminished SLFN11 expression compromises the DHX9-BRCA1 interaction, altering the regular DNA damage repair pathway and promoting chemoresistance [101].

Targeting DHX9 to mediate chemosensitivity

Analysis of a cadre of RNA/DNA hybrid-binding proteins has revealed that DHX9 has one of the highest levels of drug sensitivity interactions with the US FDA-approved chemotherapeutic agents, exhibiting both positive and negative correlations, highlighting the fact that DHX9 expression can variably affect chemotherapy response [104]. p53-induced apoptosis is an essential mechanism for cancer cell death induction following chemotherapy-induced DNA damage and this is the reason why P53 has been strongly liked with chemosensitivity [105]. DHX9's involvement in the p53 pathway has previously been discussed, with DHX9 inhibition evoking p53-mediated apoptosis in numerous cancer cell lines [83]. Similarly, silencing DHX9 induces activation of the p53 signaling pathway in a mouse lymphoma model. In this context, the activated replicative stress response induced apoptosis and sensitized resistant lymphoma cells to ABT-737 [106]. Conceptually, DHX9 could be targeted alongside ABT-737 treatment as a feasible strategy to overcoming resistant lymphoma cells. Additionally, DHX9 expression can be tuned to improve chemosensitivity of Ewing sarcoma cells. As previously discussed, DHX9 binds EWS-FLI1 in Ewing sarcoma, enhancing EWS-FLI1-dependent transcription [107]. Inclusion of poison-exon 6A in DHX9 would usually target it to nonsense-mediated mRNA decay, however, in Ewing sarcoma DHX9 expression is upregulated following repression of this exon and correlates with a poor prognosis [15]. A different study revealed that SRSF3 and hnRNPM can bind to DHX9 exon 6A to suppress its inclusion and promote DHX9 expression. Downregulation of these proteins inhibits DHX9 expression, suppresses proliferation and sensitizes Ewing sarcoma cells to doxorubicin treatment, revealing the importance of DHX9 alternative splicing in conferring chemosensitivity or chemoresistance [108].

DHX9 as an antineoplastic target for inhibition

One of the main pitfalls of chemotherapy is lack of specificity due to the general targeting of highly proliferative cells as opposed to only bona fide cancer cells, which can result in highly toxic cellular conditions that inadvertently damage healthy tissue [109]. Targeted inhibition of proteins implicated in tumorigenesis is a promising anticancer therapy because of the heightened selectivity of the approach. Given that DHX9 is overexpressed in many cancers and contributes to the development of numerous hallmarks of cancer (as previously discussed), the helicase should represent an attractive antineoplastic target. While Lee et al. revealed that DHX9 depletion in mice induced embryonic lethality, thus proving essential for embryonic development [110], several studies exploring inhibition of DHX9 in cancer cells have discovered that it is not only effective but also safe in adult mice. For example, shRNA-mediated DHX9 knockdown was lethal to a number of cancer cell lines and lymphoma cells of a mouse model while imposing no detrimental effects on healthy tissue in vivo. Additionally, adult mice seemed to tolerate long-term genetic DHX9 suppression. Of particular importance in this regard is the fact that in vivo DHX9 knockdown did not negatively affect other highly proliferative cells, ensuring high specificity while exhibiting low toxicity to healthy tissue [83]. A follow-up study then highlighted the potential of targeting DHX9 in p53-deficient tumors, whereby DHX9 suppression in p53-null tumors also induced cell death or cell cycle arrest [84]. Given that p53 is commonly inactivated in cancer cells [111], this validates the approach of targeting DHX9, which could be broadly applied to many cancers. With this in mind, an aptamer-targeting DHX9 has been identified for use in CRC tumor cells *in vivo*. The aptamer binds DHX9 with high affinity and specificity allowing preferential action in cancer cells over normal tissue due to DHX9 overexpression in CRC cells. Further specificity is achieved by localizing the aptamer to the nucleus allowing targeted delivery and this approach did not negatively affect normal cells. Given that this work was performed in human xenografts, it is highly likely that such a method could translate to humans, further establishing DHX9 as an effective and safe molecular target [6]. Another study investigating the overexpression of DHX9 in lung cancer highlighted the potential use of enoxacin, a fluoroquinolone, for targeting DHX9. It is speculated that enoxacin enhances RNA interference dependent on DHX9 expression in non-small-cell lung cancer cells, inhibiting their proliferation [4]. Further research into combining enoxacin with other chemotherapeutic agents may be a future avenue to explore in cancer treatment.

DHX9 in oncolytic viral therapy

Cancer immunotherapy can offer more selective and tailored treatment than chemotherapy by hijacking the immune system to recognize and destroy cancer cells [109]. JP Allison and T Honjo won the 2018 Nobel Prize for their discovery of immune checkpoint blockade, which has redefined treatment options for many cancers, including melanoma, through the use of monoclonal antibodies targeting CTLA-4 and PD-1/PD-L1 [112]. Oncolytic viruses (OVs) are a novel type of immuno-oncotherapy, which can selectively replicate in tumor cells with high specificity to elicit an effective antitumor immune response [113]. Clinical trials combining immune checkpoint blockade with OVs have achieved markedly higher efficacy than monotherapy and appear tolerable without unexpected additional adverse events in patients with advanced melanoma [114,115], therefore, combined therapy has the potential to minimize off-target toxicity, as well as being applicable to a larger range of cancers.

DHX9's aforementioned roles in innate immunity with respect to antiviral immune responses and viral replication places the helicase in a pivotal position to hone cancer cells' responses to oncolytic viral therapies. Myxoma virus (MYXV) is a potential candidate for oncolytic viral therapy because of its ability to infect human cancer cells both efficiently and safely [116,117]. Rahman et al. identified that DHX9, along with several other DExD/H-box helicases, provokes antiviral responses by attenuating MYXV viral replication in human cancer cells. siRNA-mediated DHX9 knockdown enhanced MYXV replication in cancer cells, showing that DHX9's regulatory role in the antiviral immune response is highly relevant in this context [118]. However, as with all aspects of DHX9 functionality, there are two sides to every coin. DHX9 can elicit both pro- and anti-viral effects. For example, an earlier study revealed proviral effects of DHX9 following its interaction with MYXV-encoded protein M029 in several human cancer cell lines, an action which promoted viral replication. M029 also binds PKR, leading to inhibition of antiviral signaling [119]. Interestingly, DHX9 is a substrate of PKR-mediated phosphorylation [120], therefore, DHX9 can also promote the MYXV viral life cycle upon repression of PKR, highlighting the dual functions of the enzyme in MYXV regulation. DHX9 also facilitates HBV replication via Nup98 regulation, and it was shown that cells infected with HBV contained upregulated DHX9 expression, with levels correlating to viral load [53]. It was then later revealed that DHX9 interacts with A3B in order to negatively regulate anti-HBV function, therefore contributing to HBV viral DNA replication [54]. Overall, given DHX9's regulatory role in the life cycle of many viruses used as OVs, DHX9 could be targeted to enhance cancer cells' response to OVs, thereby widening the applications of targeting DHX9 in cancer therapy to modulate cancer immunotherapy.

Conclusion

It is clear from the literature cited above that DHX9 has well-defined roles as an oncogene and a tumor suppressor in different tissues and cancer cell types. The versatility of the helicase is a product of its multiple functions in resolving nucleotide structures and also of the numerous protein–protein interactions it makes with binding partners that transduce vital signals for fundamental cellular processes such as gene expression and DNA replication. DHX9's multitude of functions in the development of numerous hallmarks of cancer highlight a pivotal role in malignancy and potential as both a biomarker and selective target for cancer therapy.

Future perspective

There is growing evidence to suggest that DHX9 would make an effective therapeutic target and possibly a diagnostic gene for some cancers, however, careful thought is needed to hone approaches that selectively focus on the oncogenic potential of the helicase rather than the plethora of vital 'housekeeping' functions it undertakes to maintain the stability of the human genome. The proposal of DHX9 as a potential target for enhancing oncolytic viral therapy is an avenue to be further explored and given the recent advances in cancer immunotherapy, it can be

assumed that this field will continue to evolve rapidly. Ultimately, targeting DHX9 for inhibition appears to offer a safe and selective approach for cancer treatment, and combined with current cancer therapies could lead to more effective and specific treatments applicable to a broad range of cancers, however, clinical trials are lacking in this area, therefore further research is required to fully ascertain its full potential.

Executive summary

- The hallmarks of cancer are features of cancer cells that promote/support the development and progression of cancer.
- DExH-Box helicase 9 (DHX9)'s enigmatic functions include maintenance of genomic stability, DNA replication and gene expression. Dysregulated DHX9 activity leads to a lack of coordination of these important cellular processes, which in turn can lead to the development of numerous cancer hallmarks and malignancy.
- DHX9 can function as either an oncogene or tumor suppressor dependent on its interaction with downstream partners and activation state of inter-connected signaling pathways.
- The involvement of DHX9 in malignancy positions it as an attractive biomarker and target for cancer therapy.

Author contributions

C Gulliver contributed to the initial conception/design and writing of the manuscript. GS Baillie performed the substantial review, editing and final approval of the manuscript for publishing. R Hoffmann was responsible for the review, editing and final approval of the manuscript publishing.

Acknowledgments

The authors thank Jonathan Paterson for design of diagram figures.

Financial & competing interests disclosure

C Gulliver and GS Baillie are funded by the College of Medical, Veterinary and Life Sciences PhD scheme at the University of Glasgow. GS Baillie is a member of the journal's Editorial Board and a director of Portage Glasgow Ltd. R Hoffmann is an employee of Philips Research. GS Baillie and R Hofmann hold patent rights related to DHX9. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

Open access

This work is licensed under the Creative Commons Attribution 4.0 License. To view a copy of this license, visit http://creativecomm ons.org/licenses/by/4.0/

References

Papers of special note have been highlighted as: • of interest; •• of considerable interest

- Fuller-Pace FV. DExD/H box RNA helicases: multifunctional proteins with important roles in transcriptional regulation. *Nucleic Acids* Res. 34(15), 4206–4215 (2006).
- Jain A, Bacolla A, Del Mundo IM, Zhao J, Wang G, Vasquez KM. DHX9 helicase is involved in preventing genomic instability induced by alternatively structured DNA in human cells. *Nucleic Acids Res.* 41(22), 10345–10357 (2013).
- 3. Cristini A, Groh M, Kristiansen MS, Gromak N. RNA/DNA hybrid interactome identifies DXH9 as a molecular player in transcriptional termination and R-loop-associated DNA damage. *Cell Rep.* 23(6), 1891–1905 (2018).
- Cao S, Sun R, Wang W et al. RNA helicase DHX9 may be a therapeutic target in lung cancer and inhibited by enoxacin. Am. J. Transl. Res. 9(2), 674–682 (2017).
- Yan X, Chang J, Sun R *et al.* DHX9 inhibits epithelial-mesenchymal transition in human lung adenocarcinoma cells by regulating STAT3. *Am. J. Transl. Res.* 11(8), 4881–4894 (2019).
- 6. Mi J, Ray P, Liu J *et al. In vivo* selection against human colorectal cancer xenografts identifies an aptamer that targets RNA helicase protein DHX9. *Mol. Ther. Nucleic Acids* 5, e315 (2016).
- 7. Lee T, Pelletier J. The biology of DHX9 and its potential as a therapeutic target. Oncotarget 7(27), 42716–42739 (2016).
- 8. Umate P, Tuteja N, Tuteja R. Genome-wide comprehensive analysis of human helicases. Commun. Integr. Biol. 4(1), 118 (2011).
- 9. Gilman B, Tijerina P, Russell R. Distinct RNA-unwinding mechanisms of DEAD-box and DEAH-box RNA helicase proteins in remodeling structured RNAs and RNPs. *Biochem. Soc. Trans.* 45(6), 1313–1321 (2017).

- 10. Pyle AM. Translocation and unwinding mechanisms of RNA and DNA helicases. Annu. Rev. Biophys. 37(1), 317-336 (2008).
- 11. Schütz P, Karlberg T, van den Berg S *et al.* Comparative structural analysis of human DEAD-Box RNA helicases. *PLoS ONE* 5(9), e12791 (2010).
- 12. Bourgeois CF, Mortreux F, Auboeuf D. The multiple functions of RNA helicases as drivers and regulators of gene expression. *Nat. Rev. Mol. Cell Biol.* 17(7), 426–438 (2016).
- 13. Lee C-G, Eki T, Okumura K *et al.* The human RNA Helicase A (DDX9) gene maps to the prostate cancer susceptibility locus at chromosome band 1q25 and its pseudogene (DDX9P) to 13q22, respectively. *Somat Cell Mol Genet* 25(1), 33–39 (1999).
- 14. Zhang S, Grosse F. Domain structure of human nuclear DNA helicase II (RNA helicase A). J. Biol. Chem. 272(17), 11487–11494 (1997).
- Fidaleo M, Svetoni F, Volpe E, Miñana B, Caporossi D, Paronetto MP. Genotoxic stress inhibits ewing sarcoma cell growth by modulating alternative pre-mRNA processing of the RNA helicase DHX9. *Oncotarget* 6(31), 31740–31757 (2015).
- 16. Chakraborty P, Grosse F. Human DHX9 helicase preferentially unwinds RNA-containing displacement loops (R-loops) and G-quadruplexes. *DNA Repair (Amst).* 10(6), 654–665 (2011).
- 17. Mikolaskova B, Jurcik M, Cipakova I, Kretova M, Chovanec M, Cipak L. Maintenance of genome stability: the unifying role of interconnections between the DNA damage response and RNA-processing pathways. *Curr. Genet.* 64(5), 971–983 (2018).
- Roy D, Zhang Z, Lu Z, Hsieh C-L, Lieber MR. Competition between the RNA transcript and the nontemplate DNA strand during R-loop formation *in vitro*: a nick can serve as a strong R-loop initiation site. *Mol. Cell. Biol.* 30(1), 146–159 (2010).
- 19. Allison DF, Wang GG. R-loops: formation, function, and relevance to cell stress. Cell Stress. 3(2), 38-46 (2019).
- Hamperl S, Bocek MJ, Saldivar JC, Swigut T, Cimprich KA. Transcription-replication conflict orientation modulates R-loop levels and activates distinct DNA damage responses. *Cell* 170(4), 774 (2017).
- 21. Gan W, Guan Z, Liu J *et al.* R-loop-mediated genomic instability is caused by impairment of replication fork progression. *Genes Dev.* 25(19), 2041–2056 (2011).
- 22. Fidaleo M, De Paola E, Paronetto MP. The RNA helicase A in malignant transformation. Oncotarget 7(19), 28711-28723 (2016).
- 23. Chakraborty P, Huang JTJ, Hiom K. DHX9 helicase promotes R-loop formation in cells with impaired RNA splicing. *Nat. Commun.* 9(1), 4346–4359 (2018).
- 24. Anderson SF, Schlegel BP, Nakajima T, Wolpin ES, Parvin JD. BRCA1 protein is linked to the RNA polymerase II holoenzyme complex via RNA helicase A. *Nat. Genet.* 19(3), 254–256 (1998).
- 25. Hatchi E, Skourti-Stathaki K, Ventz S et al. BRCA1 recruitment to transcriptional pause sites is required for R-loop-driven DNA damage repair. Mol. Cell. 57(4), 636–647 (2015).
- 26. Skourti-Stathaki K, Proudfoot NJ, Gromak N. Human senataxin resolves RNA/DNA hybrids formed at transcriptional pause sites to promote Xrn2-dependent termination. *Mol. Cell.* 42(6), 794–805 (2011).
- Schlacher K, Wu H, Jasin M. A distinct replication fork protection pathway connects fanconi anemia tumor suppressors to RAD51-BRCA1/2. *Cancer Cell* 22(1), 106 (2012).
- Ohle C, Tesorero R, Schermann G, Dobrev N, Sinning I, Fischer T. Transient RNA-DNA hybrids are required for efficient double-strand break repair. *Cell* 167(4), 1001–1013.e7 (2016).
- 29. Yasuhara T, Kato R, Hagiwara Y *et al.* Human Rad52 promotes XPG-mediated R-loop processing to initiate transcription-associated homologous recombination repair. *Cell* 175(2), 558–570.e11 (2018).
- Chakraborty P, Hiom K. DHX9-dependent recruitment of BRCA1 to RNA is required to promote DNA end resection in homologous recombination. *bioRxiv.* doi: https://doi.org/10.1101/2019.12.20.884593 (2019).
- Discusses linking DExH-Box helicase 9 (DHX9)'s role in R-loop resolution and BRCA1 recruitment to mediating DNA damage repair and genomic stability.
- Skourti-Stathaki K, Proudfoot NJ. A double-edged sword: R loops as threats to genome integrity and powerful regulators of gene expression. *Genes Dev.* 28(13), 1384–1396 (2014).
- 32. Friedemann J, Grosse F, Zhang S. Nuclear DNA helicase II (RNA helicase A) interacts with Werner syndrome helicase and stimulates its exonuclease activity. J. Biol. Chem. 280(35), 31303–31313 (2005).
- Chakraborty P, Grosse F. WRN helicase unwinds Okazaki fragment-like hybrids in a reaction stimulated by the human DHX9 helicase. *Nucleic Acids Res.* 38(14), 4722–4730 (2010).
- 34. Lee T, Di Paola D, Malina A *et al.* Suppression of the DHX9 helicase induces premature senescence in human diploid fibroblasts in a p53-dependent manner. *J. Biol. Chem.* 289(33), 22798–22814 (2014).
- 35. Duquette ML, Handa P, Vincent JA, Taylor AF, Maizels N. Intracellular transcription of G-rich DNAs induces formation of G-loops, novel structures containing G4 DNA. *Genes Dev.* 18(13), 1618 (2004).
- 36. Myöhänen S, Baylin SB. Sequence-specific DNA binding activity of RNA helicase A to the p16INK4a promoter. *J. Biol. Chem.* 276(2), 1634–1642 (2001).

- Zhong X, Safa AR. Phosphorylation of RNA helicase a by DNA-dependent protein kinase is indispensable for expression of the MDR1 gene product P-glycoprotein in multidrag-resistant human leukemia cells. *Biochemistry* 46(19), 5766–5775 (2007).
- Huo L, Wang YN, Xia W et al. RNA helicase A is a DNA-binding partner for EGFR-mediated transcriptional activation in the nucleus. Proc. Natl Acad. Sci. USA 107(37), 16125–16130 (2010).
- 39. Monteiro AN. BRCA1: exploring the links to transcription. Trends Biochem. Sci. 25(10), 469-474 (2000).
- 40. Nakajima T, Uchida C, Anderson SF *et al.* RNA helicase A mediates association of CBP with RNA polymerase II. *Cell* 90(6), 1107–1112 (1997).
- Tetsuka T, Uranishi H, Sanda T *et al.* RNA helicase A interacts with nuclear factor κB p65 and functions as a transcriptional coactivator. *Eur. J. Biochem.* 271(18), 3741–3751 (2004).
- 42. Mukherjee SP, Behar M, Birnbaum HA, Hoffmann A, Wright PE, Ghosh G. Analysis of the RelA:CBP/p300 interaction reveals its involvement in NF-κB-driven transcription. *PLoS Biol.* 11(9), e1001647 (2013).
- Parsyan A, Svitkin Y, Shahbazian D et al. mRNA helicases: the tacticians of translational control. Nat. Rev. Mol. Cell Biol. 12(4), 235–245 (2011).
- 44. Murat P, Marsico G, Herdy B, Ghanbarian A, Portella G, Balasubramanian S. RNA G-quadruplexes at upstream open reading frames cause DHX36- and DHX9-dependent translation of human mRNAs. *Genome Biol.* 19(1), 229 (2018).
- 45. Hartman TR, Qian S, Bolinger C, Fernandez S, Schoenberg DR, Boris-Lawrie K. RNA helicase A is necessary for translation of selected messenger RNAs. *Nat. Struct. Mol. Biol.* 13(6), 509–516 (2006).
- Robb GB, Rana TM. RNA helicase A interacts with RISC in human cells and functions in RISC loading. *Mol. Cell* 26(4), 523–537 (2007).
- Fu Q, Yuan YA. Structural insights into RISC assembly facilitated by dsRNA-binding domains of human RNA helicase A (DHX9). *Nucleic Acids Res.* 41(5), 3457 (2013).
- 48. Kawai S, Amano A. BRCA1 regulates microRNA biogenesis via the DROSHA microprocessor complex. J. Cell Biol. 197(2), 201–208 (2012).
- Boeras I, Song Z, Moran A et al. DHX9/RHA binding to the PBS-segment of the genomic RNA during HIV-1 assembly bolsters virion infectivity. J. Mol. Biol. 428(11), 2418–2429 (2016).
- Ng YC, Chung WC, Kang HR et al. A DNA-sensing-independent role of a nuclear RNA helicase, DHX9, in stimulation of NF-B-mediated innate immunity against DNA virus infection. *Nucleic Acids Res.* 46(17), 9011–9026 (2018).
- Kim T, Pazhoor S, Bao M et al. Aspartate-glutamate-alanine-histidine box motif (DEAH)/RNA helicase A helicases sense microbial DNA in human plasmacytoid dendritic cells. Proc. Natl Acad. Sci. USA 107(34), 15181–15186 (2010).
- Zhang Z, Yuan B, Lu N, Facchinetti V, Liu Y-J. DHX9 pairs with IPS-1 to sense double-stranded RNA in myeloid dendritic cells. J. Immunol. 187(9), 4501 (2011).
- Shen B, Chen Y, Hu J et al. Hepatitis B virus X protein modulates upregulation of DHX9 to promote viral DNA replication. Cell. Microbiol. 22(3), e13148–57 (2020).
- 54. Chen Y, Shen B, Zheng X *et al.* DHX9 interacts with APOBEC3B and attenuates the anti-HBV effect of APOBEC3B. *Emerg. Microbes Infect.* 9(1), 366–377 (2020).
- 55. He QS, Tang H, Zhang J, Truong K, Wong-Staal F, Zhou D. Comparisons of RNAi approaches for validation of human RNA helicase A as an essential factor in hepatitis C virus replication. *J. Virol. Methods* 154(1–2), 216–219 (2008).
- Bolinger C, Sharma A, Singh D, Yu L, Boris-Lawrie K. RNA helicase A modulates translation of HIV-1 and infectivity of progeny virions. *Nucleic Acids Res.* 38(5), 1686–1696 (2010).
- 57. Matkovic R, Bernard E, Fontanel S *et al.* The host DHX9 DExH-box helicase is recruited to chikungunya virus replication complexes for optimal genomic RNA translation. *J. Virol.* 93(4), e01764–18 (2019).
- 58. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 144(5), 646-674 (2011).
- Describes the hallmarks of cancer and being the focus for this review article.
- Wang YL, Liu JY, Yang JE *et al.* Lnc-UCID promotes G1/S transition and hepatoma growth by preventing DHX9-mediated CDK6 down-regulation. *Hepatology* 70(1), 259–275 (2019).
- 60. Lee EYHP, Muller WJ. Oncogenes and tumor suppressor genes. Cold Spring Harb. Perspect. Biol. 2(10), a003236-53 (2010).
- 61. Pearson PL, Van der Luijt RB. The genetic analysis of cancer. J. Intern. Med. 243(6), 413-417 (1998).
- 62. Wells JP, White J, Stirling PC. R loops and their composite cancer connections. Trends in Cancer 5(10), 619-631 (2019).
- 63. Turanli B, Karagoz K, Bidkhori G *et al.* Multi-omic data interpretation to repurpose subtype specific drug candidates for breast cancer. *Front. Genet.* 10, 420–32 (2019).
- 64. Scharer CD, McCabe CD, Ali-Seyed M, Berger MF, Bulyk ML, Moreno CS. Genome-wide promoter analysis of the *SOX4* transcriptional network in prostate cancer cells. *Cancer Res.* 69(2), 709 (2009).

- 65. Mehta GA, Parker JS, Silva GO, Hoadley KA, Perou CM, Gatza ML. Amplification of *SOX4* promotes PI3K/Akt signaling in human breast cancer. *Breast Cancer Res. Treat.* 162(3), 439 (2017).
- 66. Liu P, Ramachandran S, Ali Seyed M *et al.* Sex-determining region Y box 4 is a transforming oncogene in human prostate cancer cells. *Cancer Res.* 66(8), 4011–4019 (2006).
- 67. Moreno CS. SOX4: the unappreciated oncogene. Semin. Cancer Biol. (18), 30145-7 (2019).
- Lai YH, Cheng J, Cheng D et al. SOX4 interacts with plakoglobin in a Wnt3a-dependent manner in prostate cancer cells. BMC Cell Biol. 12, 50–57 (2011).
- 69. Zandi R, Larsen AB, Andersen P, Stockhausen M-T, Poulsen HS. Mechanisms for oncogenic activation of the epidermal growth factor receptor. *Cell. Signal.* 19(10), 2013–2023 (2007).
- 70. Lin S-Y, Makino K, Xia W *et al.* Nuclear localization of EGF receptor and its potential new role as a transcription factor. *Nat. Cell Biol.* 3(9), 802–808 (2001).
- 71. Harbour JW, Dean DC. The Rb/E2F pathway: expanding roles and emerging paradigms. Genes Dev. 14(19), 2393–2409 (2000).
- 72. Ormandy CJ, Musgrove EA, Hui R, Daly RJ, Sutherland RL. Cyclin D1, EMS1 and 11q13 amplification in breast cancer. *Breast Cancer Res. Treat.* 78(3), 323–35 (2003).
- 73. Garcea G, Neal C, Pattenden C, Steward W, Berry D. Molecular prognostic markers in pancreatic cancer: a systematic review. *Eur. J. Cancer* 41(15), 2213–2236 (2005).
- 74. Gautschi O, Ratschiller D, Gugger M, Betticher D, Heighway J. Cyclin D1 in non-small cell lung cancer: a key driver of malignant transformation. *Lung Cancer* 55(1), 1–14 (2007).
- 75. Palombo R, Frisone P, Fidaleo M, Mercatelli N, Sette C, Paronetto MP. The promoter-associated noncoding RNA *PNCCCND1_B* assembles a protein–RNA complex to regulate cyclin D1 transcription in Ewing sarcoma. *Cancer Res.* 79(14), 3570–3582 (2019).
- 76. Li J, Poi MJ, Tsai M-D. The regulatory mechanisms of tumor supressor p16INK4 and relevance to cancer. *Biochemistry* 50(25), 5566–5582 (2011).
- 77. Schlegel BP, Starita LM, Parvin JD. Overexpression of a protein fragment of rna helicase a causes inhibition of endogenous brca1 function and defects in ploidy and cytokinesis in mammary epithelial cells. *Oncogene* 22(7), 983–991 (2003).
- 78. Deng C-X. BRCA1: cell cycle checkpoint, genetic instability, DNA damage response and cancer evolution. *Nucleic Acids Res.* 34(5), 1416 (2006).
- 79. Aprelikova ON, Fang BS, Meissner EG et al. BRCA1-associated growth arrest is RB-dependent. Proc. Natl Acad. Sci. USA 96(21), 11866 (1999).
- Guénard F, Labrie Y, Ouellette G et al. Genetic sequence variations of BRCA1-interacting genes AURKA, BAP1, BARD1 and DHX9 in French Canadian Families with high risk of breast cancer. J. Hum. Genet. 54(3), 152–161 (2009).
- Furuta S, Jiang X, Gu B, Cheng E, Chen P, Lee W. Depletion of BRCA1 impairs differentiation but enhances proliferation of mammary epithelial cells. *Proc. Natl Acad. Sci. USA* 102(26), 9176–9181 (2005).
- 82. Ozaki T, Nakagawara A. Role of p53 in cell death and human cancers. Cancers (Basel) 3(1), 994-1013 (2011).
- Lee T, Paquet M, Larsson O, Pelletier J. Tumor cell survival dependence on the DHX9 DExH-box helicase. Oncogene 35(39), 5093–5105 (2016).
- •• Reveals safety and efficacy of targeting DHX9.
- 84. Lee T, Pelletier J. Dependence of p53-deficient cells on the DHX9 DExH-box helicase. Oncotarget 8(19), 30908-30921 (2017).
- Chen ZX, Wallis K, Fell SM *et al.* RNA helicase a is a downstream mediator of *KIF1Bβ* tumor-suppressor function in neuroblastoma. *Cancer Discov.* 4(4), 434–451 (2014).
- Ding X, Jia X, Wang C, Xu J, Gao S-J, Lu C. A DHX9-IncRNA-MDM2 interaction regulates cell invasion and angiogenesis of cervical cancer. *Cell Death Differ*. 26(9), 1750–1765 (2019).
- 87. Hansen T, Jensen T, Clausen B et al. Natural RNA circles function as efficient microRNA sponges. Nature 495(7441), 384-388 (2013).
- Aktaş T, Ilik IA, Maticzka D *et al.* DHX9 suppresses RNA processing defects originating from the Alu invasion of the human genome. *Nature* 544(7648), 115–119 (2017).
- Hsiao K-Y, Lin Y-C, Gupta SK *et al.* Non-coding effects of circular RNA CCDC66 promote colon cancer growth and metastasis. *Cancer Res.* 77(9), 2339 (2017).
- 90. Lin Y, Yu Y, Lin H, Hsiao K. Oxaliplatin-induced DHX9 phosphorylation promotes oncogenic circular RNA CCDC66 expression and development of chemoresistance. *Cancers (Basel)* 12(3), 1–14 (2020).
- 91. Yu J, Xu Q-G, Wang Z-G et al. Circular RNA cSMARCA5 inhibits growth and metastasis in hepatocellular carcinoma. J. Hepatol. 68(6), 1214–1227 (2018).
- Zucchini C, Rocchi A, Manara MC *et al.* Apoptotic genes as potential markers of metastatic phenotype in human osteosarcoma cell lines. *Int. J. Oncol.* 32(1), 17–31 (2008).

- Song Z, Zhuo Z, Ma Z, Hou C, Chen G, Xu G. Hsa_Circ_0001206 is downregulated and inhibits cell proliferation, migration and invasion in prostate cancer. Artif. Cells, Nanomedicine, Biotechnol. 47(1), 2449–2464 (2019).
- Kamelgarn M, Chen J, Kuang L et al. Proteomic analysis of FUS interacting proteins provides insights into FUS function and its role in ALS. Biochim. Biophys. Acta 1862(10), 2004–2014 (2016).
- 95. Micalizzi DS, Farabaugh SM, Ford HL. Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. *J. Mammary Gland Biol. Neoplasia* 15(2), 117 (2010).
- 96. Shang B-Q, Li M-L, Quan H *et al.* Functional roles of circular RNAs during epithelial-to-mesenchymal transition. *Mol. Cancer* 18, 138–147 (2019).
- Yan D, Dong W, He Q et al. Circular RNA circPICALM sponges miR-1265 to inhibit bladder cancer metastasis and influence FAK phosphorylation. *EBioMedicine* 48, 316–331 (2019).
- Joseph NA, Chiou S-H, Lung Z et al. The role of HGF-MET pathway and CCDC66 cirRNA expression in EGFR resistance and epithelial-to-mesenchymal transition of lung adenocarcinoma cells. J. Hematol. Oncol. 11, 557–559 (2018).
- Wang W-P, Sun Y, Lu Q *et al.* Gankyrin promotes epithelial-mesenchymal transition and metastasis in NSCLC through forming a closed circle with IL-6/STAT3 and TGF-β/SMAD3 signaling pathway. *Oncotarget* 8(4), 5909–5923 (2017).
- 100. Woods D, Turchi JJ. Chemotherapy induced DNA damage response: convergence of drugs and pathways. *Cancer Biol. Ther.* 14(5), 379–389 (2013).
- 101. Nogales V, Reinhold WC, Varma S *et al.* Epigenetic inactivation of the putative DNA/RNA helicase SLFN11 in human cancer confers resistance to platinum drugs. *Oncotarget* 7(3), 3084–3097 (2016).
- 102. Zhong X, Safa AR. RNA helicase A in the MEF1 transcription factor complex up-regulates the MDR1 gene in multidrug-resistant cancer cells. J. Biol. Chem. 279(17), 17134–17141 (2004).
- 103. Yi G-Z, Xiang W, Feng W-Y et al. Identification of key candidate proteins and pathways associated with temozolomide resistance in glioblastoma based on subcellular proteomics and bioinformatical analysis. *Biomed Res. Int.* 2018, 5238760 (2018).
- 104. Boros-Oláh B, Dobos N, Hornyák L et al. Drugging the R-loop interactome: RNA-DNA hybrid binding proteins as targets for cancer therapy. DNA Repair (Amst). 84, 102642–52 (2019).
- 105. Chesler L, Goldenberg DD, Collins R et al. Chemotherapy-induced apoptosis in a transgenic model of neuroblastoma proceeds through p53 induction. Neoplasia 10(11), 1268–1274 (2008).
- 106. Mills JR, Malina A, Lee T et al. RNAi screening uncovers DHX9 as a modifier of ABT-737 resistance in an Em-myc/BCL-2 mouse model. Blood 121(17), 3402–3412 (2013).
- 107. Toretsky JA, Erkizan V, Levenson A *et al.* Oncoprotein EWS-FLI1 activity is enhanced by RNA helicase A. *Cancer Res.* 66(11), 5574–5581 (2006).
- Palombo R, Verdile V, Paronetto MP. Poison-exon inclusion in DHX9 reduces its expression and sensitizes Ewing sarcoma cells to chemotherapeutic treatment. *Cells* 9(2), 328 (2020).
- 109. Schirrmacher V. From chemotherapy to biological therapy: a review of novel concepts to reduce the side effects of systemic cancer treatment (Review). Int. J. Oncol. 54(2), 407–419 (2019).
- 110. Lee CG, Da Costa Soares V, Newberger C, Manova K, Lacy E, Hurwitz J. RNA helicase A is essential for normal gastrulation. *Proc. Natl Acad. Sci. USA* 95(23), 13709–13713 (1998).
- 111. Rivlin N, Brosh R, Oren M, Rotter V. Mutations in the p53 tumor suppressor gene: important milestones at the various steps of tumorigenesis. *Genes Cancer* 2(4), 466 (2011).
- 112. Guo ZS. The 2018 Nobel Prize in medicine goes to cancer immunotherapy (editorial for BMC cancer). BMC Cancer 18, 1086–1088 (2018).
- 113. Raja J, Ludwig JM, Gettinger SN, Schalper KA, Kim HS. Oncolytic virus immunotherapy: future prospects for oncology. J. Immunother. Cancer 6(1), 140 (2018).
- 114. Chesney J, Puzanov I, Collichio F *et al.* Randomized, open-label Phase II study evaluating the efficacy and safety of talimogene laherparepvec in combination with ipilimumab versus ipilimumab alone in patients with advanced, unresectable melanoma. *J. Clin. Oncol.* 36(17), 1658–1667 (2018).
- Ribas A, Dummer R, Puzanov I et al. Oncolytic virotherapy promotes intratumoral T cell infiltration and improves anti-PD-1 immunotherapy. Cell 170(6), 1109–1119.e10 (2017).
- 116. Chan WM, Rahman MM, McFadden G. Oncolytic myxoma virus: the path to clinic. Vaccine 31(39), 4252-4258 (2013).
- 117. Rahman MM, Madlambayan GJ, Cogle CR, McFadden G. Oncolytic viral purging of leukemic hematopoietic stem and progenitor cells with Myxoma virus. *Cytokine Growth Factor Rev.* 21(2–3), 169–175 (2010).
- 118. Rahman MM, Bagdassarian E, Ali MAM, McFadden G. Identification of host DEAD-box RNA helicases that regulate cellular tropism of oncolytic Myxoma virus in human cancer cells. *Sci. Rep.* 7(1), 1–14 (2017).

- 119. Rahman MM, Liu J, Chan WM, Rothenburg S, McFadden G. Myxoma virus protein M029 is a dual function immunomodulator that inhibits PKR and also conscripts RHA/DHX9 to promote expanded host tropism and viral replication. *PLoS Pathog.* 9(7), e1003465 (2013).
- 120. Sadler AJ, Latchoumanin O, Hawkes D, Mak J, Williams BRG. An antiviral response directed by PKR phosphorylation of the RNA helicase A. *PLoS Pathog.* 5(2), e1000311 (2009).