# **iScience**

## Review



# Function and mechanism of exosomes derived from different cells as communication mediators in colorectal cancer metastasis

Yimin E,<sup>2,6</sup> Chen Lu,<sup>1,6</sup> Kuixuan Zhu,<sup>3,6</sup> Wenyuan Li,<sup>4</sup> Jing Sun,<sup>2</sup> Pengcheng Ji,<sup>1</sup> Minjie Meng,<sup>2</sup> Zhengxia Liu,<sup>5</sup> and Chunzhao Yu<sup>1,2,\*</sup>

#### SUMMARY

Colorectal cancer (CRC) ranks as the second leading cause of cancer-related mortality, with metastasis being the primary determinant of poor prognosis in patients. Investigating the molecular mechanisms underlying CRC metastasis is currently a prominent and challenging area of research. Exosomes, as crucial intercellular communication mediators, facilitate the transfer of metabolic and genetic information from cells of origin to recipient cells. Their roles in mediating information exchange between CRC cells and immune cells, fibroblasts, and other cell types are pivotal in reshaping the tumor microenvironment, regulating key biological processes such as invasion, migration, and formation of pre-metastatic niche. This article comprehensively examines the communication function and mechanism of exosomes derived from different cells in cancer metastasis, while also presenting an outlook on current research advancements and future application prospects. The aim is to offer a distinctive perspective that contributes to accurate diagnosis and rational treatment strategies for CRC.

#### INTRODUCTION

Colorectal cancer (CRC) ranks as the second leading cause of cancer-related mortality globally, contributing to an estimated annual death toll of approximately 910,000 individuals.<sup>1</sup> Upon diagnosis of CRC, approximately 70.4% of patients develop metastasis, primarily to the lymph nodes, while 20–25% of patients experience distant metastasis, with liver metastasis being the most prevalent.<sup>2,3</sup> Tumor metastasis serves as the primary determinant of an unfavorable prognosis in patients, with a 5-year survival rate of 71% observed for CRC patients exhibiting peripheral metastasis and merely 14% for those presenting distant organ metastasis.<sup>2</sup> Despite significant advancements in surgical techniques and targeted therapies that have improved long-term survival rates for patients, the prognosis for CRC patients with metastasis remains unfavorable.<sup>4,5</sup> Therefore, the current focus of research on CRC is primarily directed toward the exploration of more effective and safer diagnostic and therapeutic approaches.

The metastasis of CRC is a complex and dynamic biological process involving multiple genes and links.<sup>6</sup> Currently, the underlying mechanisms of tumor metastasis remain poorly understood. Metastasis of tumor requires a series of sequential steps known as the metastatic cascade.<sup>7</sup> This sequence includes the initiation of invasion by tumor cells into the surrounding tissues of the primary tumor. Subsequently, these cells enter the bloodstream and manage to survive in circulation. They then adhere to vessel walls and extravasate into the parenchyma of distant tissues. Micrometastatic colonies form within the parenchyma, followed by reactivation of proliferation, leading to clinically detectable overt metastatic lesions. This last process being termed colonization.<sup>7,8</sup> Importantly, interactions between tumor cells and non-tumor stromal cells play a crucial role throughout the invasion-metastasis cascade. Exosome-mediated intercellular communication plays a crucial role in various critical steps of metastatic colorectal cancer (mCRC) development and progression, including enhancing tumor cell invasion and migration capacity, altering vascular permeability, facilitating immune escape, and promoting pre-metastatic niche formation.<sup>9–11</sup> Exosomes also act as triggers or drivers for tumor metastasis occurrence when interacting with other factors.<sup>6,12</sup>

During the progression of the disease, intercellular communication occurs not only among cancer cells within the tumor but also between diverse cell types in the tumor microenvironment (TME).<sup>13,14</sup> TME is a complex network of diverse cell types embedded within the extracellular matrix (ECM), including stromal cells such as fibroblasts, pericytes, and adipocytes, as well as immune cells such as T lymphocytes, B lymphocytes, natural killer cells (NKs), and tumor-associated macrophages (TAMs).<sup>15,16</sup> Cellular communication encompasses diverse

<sup>6</sup>These authors contributed equally

<sup>&</sup>lt;sup>1</sup>Department of General Surgery, Sir Run Run Hospital of Nanjing Medical University, Long Mian Avenue 109 Jiangning, Nanjing 211112, Jiangsu, China <sup>2</sup>Department of General Surgery, The Second Affiliated Hospital of Nanjing Medical University, 121 Jiang Jia Yuan Road, Nanjing 210011, Jiangsu, China <sup>3</sup>Department of Radiotherapy, The Third Affiliated Hospital of Kunming Medical University (Yunnan Cancer Hospital, Yunnan Cancer Center), Kunming 650100, Yunan, China <sup>4</sup>Department of Anesthesiology, The Second Affiliated Hospital of Nanjing Medical University, 121 Jiang Jia Yuan Road, Nanjing 210011, Jiangsu, China <sup>5</sup>Department of Geriatrics, The Second Affiliated Hospital of Nanjing Medical University, 121 Jiang Jia Yuan Road, Nanjing 210011, Jiangsu, China

<sup>\*</sup>Correspondence: chunzhaoyu@njmu.edu.cn

https://doi.org/10.1016/j.isci.2024.109350





mechanisms, including direct cell-to-cell interactions and indirect regulation mediated by secreted bioactive molecule or extracellular vesicle (EV), thereby reshaping TME and exerting multifaceted influences on tumor development, encompassing cancer cell proliferation, invasion, metastasis, immune escape, angiogenesis, and chemotherapy resistance.<sup>17,18</sup>

In recent years, extensive researches on EVs have revealed their pivotal role in mediating and regulating intercellular communication associated with physiological and pathological processes.<sup>19–21</sup> Consequently, they have garnered significant attention in the biomedical field for their potential applications in cancer diagnosis and treatment. EV is a collective term, encompassing a diverse range of cell-released subtypes of membrane structures.<sup>22</sup> Exosome, a specific subtype of EV, possesses a diameter ranging from approximately 40 to 160 nm (with an average size of about 100 nm). They consist of various membrane-associated higher-order oligomeric protein complexes, nucleic acids, lipids, ECM components, transcription factors, intracellular and extracellular receptors, enzymes as well as other protein complexes.<sup>23,24</sup> The biogenesis and release of exosomes constitute a series of intricate, multi-step processes. Intracellular endosomes undergo continuous invagination of the plasma membrane, leading to the formation of multivesicular bodies. These dynamic structures can engage in interactions with other intracellular vesicles and organelles before ultimately fusing with the plasma membrane for secretion into the extracellular space.<sup>25</sup> Exosomes can be secreted by a wide range of cell types, encompassing tumor cells, macrophages, T lymphocytes, B lymphocytes, fibroblasts, dendritic cells (DCs), astrocytes, and mesenchymal stem cells (MSCs). These exosomes exhibit stability in diverse bodily fluids such as blood, urine, saliva, tears and cerebrospinal fluid.<sup>24,26</sup> In the humoral milieu, exosomes exist as distinct subpopulations characterized by variations in size, morphology, composition, and biogenesis mechanisms. Primarily functioning as intercellular communicators, exosomes facilitate the exchange of molecular cargo between cells, thereby modulating the biological attributes of recipient cells.<sup>27</sup> The intercellular communication by exosomes can be summarized as follows: (1) Exosomal membrane proteins directly engage recipient cells, initiating intracellular signaling cascades; (2) Their cargo to recipient cells through fusion with the cellular membrane; (3) Recipient cells internalize exosomes and release their signaling molecules following lysosomal degradation.<sup>24</sup> Furthermore, mediators facilitating interactions between exosomes and recipient cells have been identified, encompassing tetraspanins (i.e., CD81), integrins, lipids, lectins, heparan sulfate proteoglycans, and ECM constituents. The interplay between integrins and ECM proteins (primarily fibronectin and laminin) has demonstrated its indispensable role in the binding of exosomes to recipient cells.<sup>28,29</sup> Within the TME, diverse cellular components, including exosomes and other communication mediators, coordinate a range of biological effects to facilitate tumor cell invasion and metastasis. These effects encompass migratory potential acquisition from the primary tumor, promotion of angiogenesis, evasion of immune surveillance, organ-specific metastasis patterns, establishment of pre-metastatic niches, and growth initiation at secondary sites. 18,30 Tumor-derived exosomes (TDEs) play a pivotal role in creating a conducive microenvironment at future metastatic sites while also mediating non-random patterns of metastasis by selectively targeting organs such as lung or liver based on their integrin composition.<sup>28</sup> Other cells within the TME, including cancer-associated fibroblasts (CAFs) and macrophages, also play crucial roles in different stages of cancer progression and metastasis. LINC00659, originating from CAFs in the TME, is transferred to CRC cells through exosomes, thereby promoting cancer cell proliferation, invasion, and migration. Additionally, it facilitates epithelial-mesenchymal transition (EMT).<sup>31</sup> Exosomes derived from M2 macrophages, carrying miR-21-5p and miR-155-5p, exerted a downregulatory effect on the expression of Brahma-related gene 1 (BRG1) receptor in CRC cells, thereby enhancing their metastatic potential.<sup>32</sup> Exosomes derived from tumor cells or other cells play a crucial role in various stages of cancer development by delivering proteins, metabolites, or nucleic acids to recipient cells, thereby modulating the transcriptome and/or cell phenotype of the recipients. In CRC, this phenomenon manifests as biological processes that either promote or inhibit biological processes such as tumor proliferation, invasion, migration and metastasis.<sup>30</sup>

Elucidation of the regulatory mechanism underlying intercellular communication mediated by exosomes in metastasis of CRC is pivotal for gaining deeper insights into tumorigenesis and development, as well as for future clinical applications.<sup>30</sup> Exosomes enable the transmission of biological signals between different types of cells or tissues, and their regulation of intracellular signaling pathways enhances their potential diagnostic value and therapeutic efficacy in various diseases, including cancer.<sup>18,23</sup> Exosomes can be detected in a diverse range of human bodily fluids, and their isolation through liquid biopsies provides a convenient approach to acquire comprehensive genetic and metabolic information using various detection methodologies.<sup>23</sup> The primary potential of exosome-based liquid biopsy lies in its capability for disease status determination and stage diagnosis.<sup>33</sup> Furthermore, comprehensive analysis of exosomes enables determination of disease progression and treatment response.<sup>23</sup> The application of exosomes lies not only in their diagnostic value but also in their great potential in the therapeutic management of tumors and other diseases.<sup>34</sup> The lipid and protein composition of exosomes can influence the pharmacokinetic properties of drugs while their inherent components may contribute to enhanced bioavailability and reduced adverse reactions. Exosomes can be engineered to serve as efficient carriers for delivering a diverse range of therapeutic drugs, such as short interfering RNAs (siRNAs), antisense oligonucleotides (ASOs), chemotherapy drugs, and immunomodulators, effectively targeting specific sites.<sup>23</sup> Especially in the field of nanomedicine, EVs have broad application prospects as a new type of drug carrier.<sup>35,36</sup> This review critically examines the functions and underlying mechanisms of exosomes derived from diverse cellular sources, encompassing tumor cells, fibroblasts, immune cells, and other cell types, as pivotal mediators in mCRC. Simultaneously, this article elucidates the potential applications of exosomes in CRC diagnosis and treatment along with their future development prospects to offer valuable insights for rational disease management.

#### **COLORECTAL CANCER-DERIVED EXOSOMES**

Exosomes are actively secreted by tumor cells, serving as carriers of genetic and metabolic information between cells of origin and neighboring healthy or abnormal cells, thus playing a crucial role in the TME. Notably, compared to the healthy population, tumor patients exhibit increased production and release of exosomes with distinct sources and molecular contents. For instance, significant differences at the





#### Figure 1. Exosomes derived from CRC promote tumor invasion and migration

For instance, exosomal circCOG2 released by CRC cells in normoxic conditions regulated the invasion and migration abilities of tumor cells by activating the miR-1305/TGF-β2/SMAD3 signaling pathway.

protein level have been observed among exosomes isolated from plasma samples of healthy controls, patients with primary CRC, and those with colorectal cancer liver metastasis (CRLM).<sup>37</sup> Additionally, previous research has revealed a higher proportion of microRNA(miRNA) in exosomes compared to parent cells.<sup>38</sup> Exosomes derived from primary or metastatic cancer cells exert regulatory control over cancer cell metastasis through diverse signaling mechanisms, including the reprogramming of protein composition and secretion via autocrine or paracrine pathways. This phenomenon enhances the migratory capacity of primary tumors, promotes angiogenesis, facilitates evasion from the immune system, supports the establishment of pre-metastatic niches, and further fosters the growth of metastatic tumors at secondary sites.<sup>39</sup> Tumor cells regulate metastasis of cancer by orchestrating intercellular communication through the secretion of exosomes, which facilitate the exchange of critical information and coordinate diverse aspects of the metastatic cascade.

#### Invasion and migration of tumor cells

The initial step in the formation of cancer metastasis entails the local dissemination of cancer cells derived from the primary tumor, which is instigated by modulating the TME.<sup>9</sup> Tumor cells enhance their invasive and migratory capabilities by secreting diverse exosomes, thereby facilitating the EMT and ECM remodeling, ultimately promoting tumor metastasis. Exosomal cargo, including RNA, proteins, lipids, or metabolites, mediates autocrine or paracrine signaling to augment invasion and migration of tumor cells (Figure 1).<sup>30</sup>

The genetic information or metabolic substances carried by exosomes derived from CRC cells can modulate the invasion and migration of tumor cells through diverse mechanisms upon transfer to recipient cells.<sup>40</sup> MiRNAs are evolutionarily conserved, short, non-coding RNAs (ncRNAs) that play a pivotal role in the post-transcriptional regulation of gene expression.<sup>41</sup> As the most biologically active molecules found within exosomes, miRNAs have garnered increasing attention due to their crucial involvement in regulating the expression of genes associated with metastasis of CRC.<sup>42</sup> A previous study demonstrated a significant elevation of exosomal miR-106b-3p in the plasma of patients with mCRC, which was associated with advanced TNM staging and larger tumor volume. Additionally, both CRC cells and exosomes derived from CRC cell supernatant exhibited significantly increased expression levels of miR-106bp compared to NCM 460 cells. Furthermore, it was discovered that exosomal miR-106b-3p promotes EMT by inhibiting dynein light chain 1 (DLC-1), thereby enhancing the invasive and metastatic potential of CRC cells.<sup>43</sup> Bigagli et al. discovered exosomes derived from HCT-8 cells carrying miR-210 are implicated in anoikis resistance and EMT markers, thereby preserving the conducive microenvironment for local cancer growth and guiding metastatic cells toward new dissemination sites.<sup>44</sup> Moreover, exosomal miR-193a derived from CRC cells exhibited inhibitory effects on tumor cell invasiveness and





proliferation, while let-7g demonstrates contrasting effects. By assessing the expression levels of exosomal miR-193a and let-7g in the plasma of patients with CRC, their functions as diagnostic and prognostic markers for this disease were confirmed.<sup>45</sup> Exosomal miR-548c-5p derived from CRC cells exerted inhibitory effects on the proliferation and invasion of CRC cells *in vitro*, as well as prevents the development of CRC *in vivo* in nude mice. Furthermore, it demonstrated that exosomal miR-548c-5p derived from CRC cells modulates CRC cell growth, migration, and invasion through the hypoxia-inducible factor 1 alpha (HIF-1α)/cell division control protein 42 (CDC42) axis.<sup>46</sup> Rezaei et al. demonstrated that the efficient delivery of miR-375 mimics by TDEs *in vitro* resulted in down-regulation of β-catenin, vimentin, zinc finger E-box binding homeobox 1 (ZEB1), and snail, while significantly reducing the expression of E-cadherin. Furthermore, TDEs inhibited the migratory and invasive capabilities of HT-29 and SW480 cells and induced an upregulation in CD44 and CD133 expression during EMT.<sup>47</sup>

Recent studies have demonstrated that circular RNAs (circRNAs) are highly enriched in exosomes and play a pivotal role in various biological processes within tumors, particularly by functioning as molecular sponges for miRNAs.<sup>48–50</sup> CircRNAs are characterized by their closed structure and single-stranded nature, lacking poly (A) tails or 5'-3' ends.<sup>9</sup> Due to this unique feature, they exhibit enhanced stability and prolonged lifespan within cells as they can withstand degradation caused by exonucleases.<sup>51</sup> Yu et al. demonstrated the upregulation of circFMN2 in both serum and exosomes from CRC patients, as well as CRC cells. Additionally, they discovered that exosomal circFMN2 promotes CRC cell proliferation and metastasis while inhibiting apoptosis through the miR-338-3p/Musashi1 (MSI1) axis. Moreover, overexpression of MSI1 counteracts the suppressive effect of miR-338-p on CRC progression, suggesting that targeting exosomal circFMN2 could be a potential therapeutic strategy for treating CRC.<sup>52</sup> In another study, exosomal circEPB41L2 derived from CRC cells inhibited the progression of CRC by sponging miR-21-5p and miR-942-5p and regulating the phosphatase and tensin homolog (PTEN)/AKT signaling pathway.<sup>49</sup> Furthermore, it has been demonstrated that circ-ABCC1, carried by exosomes derived from CD133+ CRC cells, facilitated cell stemness and metastasis in CRC via activation of the Wnt/ $\beta$ -catenin pathway.<sup>53</sup> Gao et al. reported that exosomes derived from CRC cells containing circCOG2 facilitate the proliferation, migration, and invasion of CRC by activating the miR-1305/transforming growth factor β2 (TGF-β2)/SMAD3 pathway. Moreover, this effect can be transferred from high metastatic potential CRC cells to low metastatic potential CRC cells through exosomal communication.<sup>54</sup> Chen et al. proposed a novel mechanism for the secretion of circRNAs in exosomes, whereby circRHOBTB3 is specifically sorted into exosomes through interaction between ESCRT-II complex member SNF8 and its own specific element, resulting in extracellular release via the "tumor exosome escape mechanism." Additionally, based on regulatory elements for circularization and exosomal secretion, ASOs were designed and synthesized to increase expression of circRHOBTB3 while blocking its exosomal secretion. It was found that these ASOs inhibited growth and metastasis of CRC both in vitro and in vivo.55 In a previous study, aberrant upregulation of circ-PABPC1 expression was observed in both CRC tissues and exosomes. In vitro and in vivo functional experiments provided evidence supporting the oncogene of exosomal circPABPC1. Mechanistically, circPABPC1 recruits KDM4C to the HMGA2 promoter, resulting in reduced H3K9me3 modification and subsequent initiation of transcription within the nucleus. Moreover, cytoplasmic circPABPC1 promoted CRC metastasis by safeguarding A disintegrin and metalloproteinase 19 (ADAM19) and bone morphogenetic protein 4 (BMP4) from degradation mediated by miR-874/miR-1292.<sup>56</sup>

Exosomal long non-coding RNA (IncRNA) regulates intercellular communication between tumor cells and their microenvironment, playing a pivotal role in the intricate process of tumor metastasis. The IncRNA refers to an RNA molecule that exceeds 200 nt in length and lacks the ability to encode proteins itself.<sup>57</sup> In recent years, numerous studies have elucidated the functions and underlying mechanisms of exosomal lncRNA in facilitating metastasis of CRC. Exosomal IncRNA PCAT1 derived from CRC cells regulates the activity of miR-329-3p/Netrin-1-CD146 complex to facilitate EMT and liver metastasis in CRC through the mediation of circulating tumor cells.<sup>58</sup> A previous study demonstrated that the expression level of UCA1 in serum exosomes from CRC patients was upregulated, thereby augmenting the proliferation and migration abilities of CRC of the miR-143/MYO6 axis. Furthermore, UCA1 was found to be overexpressed in CRC tissues and acted as a competitive endogenous RNA for miR-143, consequently regulating CRC cell proliferation and metastasis.<sup>59</sup> Xu et al. discovered that exosomal MALAT1 from CRC engulfs miR-26a/26b, thereby regulating fucosyltransferase 4 (FUT4) and activating phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR) pathway, ultimately promoting malignant behavior in CRC cells.<sup>60</sup>

The precise role of exosomal proteins, particularly membrane-bound proteins, remains incompletely understood. Previous studies have elucidated the pivotal role and underlying mechanisms of exosome-associated proteins derived from CRC cells in facilitating tumor invasion, migration, and metastasis.<sup>30</sup> Among the ADAM protein family, A disintegrin and metalloproteinase 17 (ADAM 17) is a membrane protein. Exosome-derived ADAM17 levels are elevated in both mCRC cells and serum from mCRC patients. Additionally, it has been discovered that exosomal ADAM the migration ability of CRC cells and contributes-metastatic niche formation by disrupting E-cadherin connections. Furthermore, the potential of circulating exosomal ADAM17 as a biomarker for predicting tumor metastasis in CRC patients has been convincingly demonstrated.<sup>61</sup> Moreover, exosomes derived from CRC cells with upregulated Ca2+-dependent secretion activator protein 1 (CAPS1) exhibited the ability to augment the migratory potential of normal colonic epithelial FHC cells, while inhibition of exosome release using GW4869 can impede FHC cell migration. Overexpression of CAPS1 induces alterations in the expression profile of exosomal proteins implicated in cellular migration, bone morphogenetic protein 4 potentially playing a crucial role in CAPS1-mediated cell migration.<sup>62</sup>

Exosomes derived from diverse malignant tumor cells induce the reprogramming of MSCs within the TME, resulting in their differentiation into CAFs.<sup>63-65</sup> The reciprocal communication between tumor cells and CAFs, as well as the biomechanical remodeling of the ECM by CAFs, significantly contributed to the migratory and invasive properties of tumor cells.<sup>66</sup> In a study investigating the regulation of fibroblast phenotype in the TME by exosomes derived from early-stage (SW480) and late-stage (SW620) CRC cells, both CRC cell lines induced quiescent fibroblasts with normal characteristics ( $\alpha$ -SMA-, CAV+, FAP+, VIM+) to differentiate into CAFs-like cells (CAFs-like;  $\alpha$ -SMA+, CAV-, FAP+, VIM+). However, the activated CAFs exhibited diverse biological functions. Early CRC cells induced fibroblast activation with potent



proliferation-promoting and-promoting capabilities, accompanied by elevated expression of angiogenesis-promoting proteins (IL8, RAB10, NDRG1) and proliferation-promoting proteins (SA1008, FFPS). In contrast, late-stage CRC cells triggered fibroblast activation that displayed remarkable invasive potential within the ECM through upregulation of pre-invasive regulatory factors involved in membrane protrusions (PDLIM1, MYO1B) and matrix remodeling proteins (MMP11, EMMPRIN, ADAM10). The conserved features of fibroblast activation mediated by exosomes encompass augmented ECM secretion, including collagen type I α-1 and Tenascin-C/X, as well as oncogenic transformation and metabolic reprogramming involving the downregulation of caveolin-1 and upregulation of glycogen metabolism (GAA), amino acid biosynthesis (SHMT2, IDH2), and membrane transporters for glucose (GLUT1), lactate (MCT4), and amino acids (SLC1A5/3A5).<sup>67</sup> Zhou et al. discovered that LINC01915 facilitates the conversion of normal fibroblasts (NFs) into CAFs via the miR-92a-3p/KLF4/CH25H axis. Additionally, LINC01915 impeded the uptake of EVs from CRC cells by NFs through the miR-92a-3p/KLF4/CH25H axis, thereby inhibiting angiogenesis and preventing the transformation of NFs into CAFs to suppress tumor growth.<sup>68</sup> Bhome et al. investigated the impact of EMT status on fibroblast phenotypes and the regulation of EVs as mediators of intercellular communication in CRC. Epithelial CRC-derived EVs suppressed TGF-β-induced myoblast differentiation, while mesenchymal CRC-derived EVs do not exhibit this effect, with miR-200 (miR-200 a/b/c, -141) playing a crucial role in driving these responses.<sup>69</sup> In another study, exosomal miR-146a-5p and miR-155-5p facilitated CXCL12/CXCR7-induced metastasis in CRC through intricate interplay with CAFs. Their study unraveled that exosomes derived from CRC cells overexpressing CXCR7 can be internalized by CAFs, thereby promoting their activation via JAK2-STAT3/NF-κB signaling pathway by targeting cytokine signal transduction inhibitor 1 (SOCS1) and zinc finger and BTB structure domain 2 (ZBTB2). The activated CAFs exhibited a substantial elevation in the levels of pro-inflammatory cytokines interleukin-6, tumor necrosis factor-alpha, transforming growth factor  $\beta$ , as well as CXCL12, which collectively instigate EMT in CRC.<sup>70</sup>

#### Angiogenesis and changes in vascular permeability

Tumor angiogenesis, the process of neovascularization from pre-existing vessels, facilitates the provision of oxygen and nutrients to tumor cells, thereby promoting their active proliferation. The distinctive characteristics of primary tumor vasculature and the TME contribute to the enhanced permeability and retention (EPR) effect. Augmented vascular permeability is a hallmark feature of tumor blood vessels and plays a pivotal role in driving the EPR effect.<sup>10</sup> Emerging evidence suggests that tumor cell-derived exosomes facilitate the transfer of bioactive substances to endothelial cells (ECs) or modulate angiogenesis alterations through intermediary cells such as fibroblasts and immune cells, thereby promoting tumor metastasis (Figure 2).<sup>71,72</sup>

Angiogenesis plays a pivotal role in all stages of cancer development, exerting profound influence on tumor progression. Exosomes derived from CRC encompass a diverse array of bioactive molecules that intricately modulate angiogenic processes and permeability alterations, thereby potentiating the invasive and metastatic potential of cancer cells.<sup>73</sup> Hu et al. discovered a significant elevation in miR-1229 levels within serum exosomes derived from CRC patients, which exhibited a strong correlation with tumor size, lymph node metastasis, TNM stage, and unfavorable prognosis. Furthermore, their investigation revealed that CRC cell-derived exosomal miR-1229 facilitated angiogenesis by targeting HIPK2 and activating the VEGF pathway. Inhibition of miR-1229 using anti-miR-1229 effectively suppressed tumor growth and angiogenesis in a xenograft mouse model.<sup>74</sup> Hong et al. discovered that EVs derived from SW480 CRC cells exhibited an enrichment of cell cycle-related mRNAs known to stimulate endothelial cell proliferation, thereby implicating cancer cell-derived EVs in the facilitation of tumor growth and metastasis of angiogenesis.<sup>75</sup> In another study, Exosomal circTUBGCP4 facilitated the formation of tip cells and promoted metastasis in colon cancer by suppressing miR-146b-3p and activating the AKT signaling pathway in endothelial cells.<sup>6</sup> Growth/differentiation factor 15 (GDF15), a member of the TGF- $\beta$ /bone morphogenetic protein (BMP) superfamily, exhibits expression in diverse tissue types. Exosomes derived from colon cancer cells resistant to 5-fluorouracil are enriched with GDF15 and possess the ability to promote angiogenesis through activation of the Smad/POSTN axis.<sup>77</sup> Moreover, EVs derived from CRC cells were found to induce Egr-1 activation and facilitate migration of endothelial cells as angiogenesis via the ERK1/2 and JNK signaling pathways.<sup>78</sup> In a separate investigation, exosomal B7-H3 derived from CRC facilitated tumor angiogenesis and metastasis by activating the AKT1/mTOR/VEGFA signaling pathway.<sup>79</sup> Moreover, fibroblasts activated by exosomes derived from SW480, exhibited augmented proliferation and angiogenesis-promoting functions, characterized by elevated expression levels of angiogenic factors (IL8, RAB10, NDRG1) and proteins associated with cellular proliferation (SA1008, FFPS).<sup>67</sup>

Tight junctions and adherens junctions are present endothelial cells, and disruption of tight junctions results in increased vascular permeability.<sup>80</sup> The tight junctions consist of two proteins, Zonula occludens-1 (ZO-1) and Claudin-5. Exosome-mediated delivery of miR-29a from tumor cells regulates the expression of ZO-1, Claudin-5, and occludin by targeting Kruppel-like factor 4 (KLF4). *In vivo* experiments confirmed that exosomal miR-29a promoted liver metastasis in CRC mice.<sup>72</sup> In a separate study, exosomal miR-25-3p derived from CRC effectively modulated the expression of VEGFR2, ZO-1, occludin, and Claudin 5 in endothelial cells, thereby facilitating vascular angiogenesis and permeability. Meanwhile, exosomal miR-25-3p originating from CRC cells significantly induced hepatic and pulmonary vascular leakage while promoting metastasis in CRC.<sup>71</sup>

Hypoxia, a crucial driver of tumor progression, regulates angiogenesis, invasion, metabolism, and genetic instability.<sup>81</sup> Huang et al. discovered that CRC cells under hypoxic conditions promote the proliferation and migration of endothelial cells through activation of the Wnt/β-catenin signaling pathway mediated by EVs.<sup>82</sup> Furthermore, they observed that these exosomes rich in Wnt4 are dependent on HIF-1α and can be inhibited by the ICG-001 inhibitor. Moreover, knockdown of RAB27a impeded CRC cell-induced proliferation and migration of endothelial cells mediated by EVs.<sup>82</sup> Cellular prion protein (PrPC) is a cell surface glycoprotein that has been found to play an important role in tumor metastasis in recent years. Yun et al. demonstrated that the hypoxic TME induces an up-regulation of exosomal PrPC expression in CRC, thereby modulating the biological behavior of CRC cells and promoting tumor metastasis, including enhanced endothelial permeability,





#### Figure 2. Exosomes derived from CRC promote angiogenesis and alter vascular permeability

For instance, by targeting KLF4, the exosomal miR-29a from CRC cells modulated the expression of ZO-1, Claudin-5, and occludin to disrupt the integrity of the vascular endothelial barrier and facilitate metastasis in CRC.

migration, invasion, and secretion of angiogenic factors. Notably, the progression of CRC in xenografted mouse models was effectively inhibited by anti-PrPC antibody.<sup>83</sup>

#### Immune escape

The involvement of exosomes in the dynamic interactions between tumor cells and the immune microenvironment enables them to regulate multiple processes associated with tumor metastasis. In most solid tumors, macrophages constitute the predominant component of the tumor stroma, with their diversity often characterized two opposing polarization states: pro-inflammatory M1 and anti-inflammatory M2 macrophages.<sup>84,85</sup> TAMs, primarily derived from peripheral blood monocytes recruited to tumor masses, constitute distinct subpopulations of macrophages that support tumor growth and are classified as M2 macrophages.<sup>86,87</sup> In addition to macrophages, the metastasis of tumor cells is also regulated by various immune cells including neutrophils, NKs, and T cells. Cancer cells infiltrate the circulatory system with the assistance of immune cells and employ multiple mechanisms to evade recognition by cytotoxic lymphocytes and phagocytes.<sup>88,89</sup> Upon arrest distant capillaries, cancer cells extravasate into the parenchyma of target organs and initiate colonization. Subsequently, established cancer cells must acquire resistance against immune responses and other host tissue defenses in order to ensure survival (Figure 3).<sup>9</sup>

Exosomes derived from tumors can induce macrophage polarization, leading to the activation of either anti-inflammatory M2 immune cells that promote tumor growth or pro-inflammatory M1 immune cells tumor growth, thereby modulating the M1/M2 ratio in the TME and regulating CRC progression.<sup>42,86,90</sup> Exosomal miRNAs derived from CRC cells have been extensively investigated in previous studies on immune evasion during metastasis. Zhao et al. revealed that exosomal miR-934 derived from CRC cells induces polarization of M2 macrophages through PTEN downregulation and activation of the PI3K/AKT signaling pathway.<sup>91</sup> In addition, reprogramming of macrophages into tumor-supportive M2 macrophages was facilitated by mutant p53 exosome-mediated miR-1246.<sup>87</sup> In another study, exosome derived fragments of cytoskeleton proteins from CRC cells were found to have the potential to induce phenotypic transitions between M1 and M2 states.<sup>92,93</sup> Takano et al. reported that exosomal miR-203 derived from CRC patients can modulate TAMs implicated in CRC metastasis.<sup>94</sup> Exosomal miR-203a-3p from CRC cells induced M2 macrophage polarization by controlling PTEN and activating the PI3K/AKT signaling pathway.<sup>95</sup> Sun et al. demonstrated that exosomes derived from CRC stem cells delivered miR-17-5p to HCT116 cells, thereby suppressing





#### Figure 3. Exosomes derived from CRC are involved in tumor immune escape

For example, the exosomal miR-203a-3p derived from CRC induced macrophage M2 polarization by modulating PTEN and activating the PI3K/AKT signaling pathway, thereby exerting its role in facilitating tumor metastasis.

anti-tumor immunity and promoting their malignant behavior. Conversely, the overexpression of speckle-type POZ protein (SPOP), an E3 ubiquitin ligase adaptor with tumor suppressor properties, exhibited an opposing effect. Upregulated miR-17-5p enhances tumor cell growth by upregulating programmed death ligand 1 (PD-L1) to inhibit SPOP expression and impairs anti-tumor immunity in CRC.<sup>96</sup> Yang et al. discovered that EMT-associated CRC cells facilitated M2-like polarization of macrophages through direct exosomal transfer, leading to a significant elevation in miR-106b-5p levels within macrophages. Mechanistically, the upregulated miR-106b-5p activated the PI3KY/AKT/mTOR signaling cascade by directly suppressing programmed cell death 4 (PDCD4) at the post-transcriptional level, thereby promoting M2 polarization of macrophages.<sup>97</sup> Hypoxia-induced exosomal miR-135a-5p is associated with the development, clinical severity, and prognosis of CRLM. Mechanistically, Kupffer cells phagocytize exosomes containing highly expressed miR-135a-5p from the bloodstream and transport them to the liver. Exosomal miR-135a-5p activates the large tumor suppressor kinase 2 (LATS2)-yes-associated protein (YAP)-matrix metal-loproteinase 7 (MMP7) axis, thereby promoting CRLM. Moreover, the intricate process is associated with immunosuppression signaling mediated by cluster of differentiation 30-TNF receptor-associated factor 2 (TRAF2)-p65.<sup>98</sup>

In addition to exosomal miRNAs, other genetic or metabolic molecules present in tumor-derived exosomes, such as IncRNAs, circRNAs, and others, exert regulatory effects on the metastasis of CRC through various mechanisms including immune evasion.<sup>18,99</sup> The plasma level of exosomal IncRPPH1 was elevated in untreated patients with CRC and significantly decreased following tumor resection. Exosomes derived from CRC cells facilitated the transfer of IncRPPH1 into macrophages, leading to M2 polarization and consequently promoting CRC cells metastasis and proliferation. Moreover, both *in vitro* and *in vivo* experiments demonstrated that IncRPPH1 played a crucial role in driving CRC metastasis. Mechanistically, IncRPPH1 interacted with tubulinβ-III (TUBB3) to prevent its ubiquitination, thereby inducing metastasis of CRC cells.<sup>100</sup> Additionally, IncKCNQ1OT1, originating from CRC cells, modulated M2 macrophage polarization by regulating the miR-30a-5p/ubiquitin-specific protease 22 (USP22) signaling axis, thereby ubiquitinating and promoting immune evasion of CRC. Furthermore, cancer cell-derived exosomes promote remodeling of the lymphatic system and trigger sentinel lymph node metastasis.<sup>57</sup>Exosomes harboring Inc-HOXB8-1:2 derived from neuroendocrine-differentiated CRC cells competitively sequestered hsa-miR-6825-5p, resulting in the upregulation of CXCR3 expression and subsequent recruitment of TAMs with M2 polarization, thereby facilitating the progression of neuroendocrine-differentiated CRC.<sup>101</sup> In a separate investigation, exosomal IncBANCR derived from SW620 and HCT-15 cells facilitated EV-mediated polarization of M2 macrophages and enhanced CRC metastasis through modulation of the RhoA/ROCK signaling pathway.<sup>102</sup>





#### Figure 4. Exosomes derived from CRC are involved in the formation of pre-metastatic niches

The exosomal HSPC 111 from CRC induced changes in the lipid metabolism of CAFs through phosphorylation of ACLY, resulting in an up-regulation of A-CoA levels. This process promoted the formation of a pre-metastatic niche and facilitates CRLM.

Moreover, the upregulation of exosomal circVCP can potentially enhance M1/M2 macrophage polarization through modulation of the miR-9-5p/NRP1 axis, thereby contributing to the progression of CRC.<sup>103</sup> CRC cells induced T cell apoptosis *in vitro* and *in vivo* by releasing EVs carrying Fas ligand and tumor necrosis factor-related apoptosis-inducing ligand.<sup>104</sup>

#### Formation of pre-metastatic niches

Tumors instigate the establishment of microenvironments in distant organs through diverse communication mediators prior to metastasis, thereby facilitating the survival and proliferation of tumor cells upon their arrival at these sites. These predetermined microenvironments are commonly referred to as pre-metastatic niches.<sup>11,105</sup> Pre-metastatic niches are established through systemic signals emanating from the primary tumor prior to the arrival of cancer cells, which recruit supportive stromal cells and other cellular constituents. The perivascular microenvironment facilitates the extravasation of cancer cells and their dissemination along the basement membrane of capillaries. Specialized niches are orchestrated by secretory products released by cancer cells, serving as an autocrine or paracrine source for recruiting supportive signals from stromal components. Cancer cells infiltrate host tissue's endogenous stem cell niches, thereby directly occupying favorable microenvironments (Figure 4).<sup>9</sup>

Exosomes derived from CRC cells have the ability to disseminate through body fluids over considerable distances, thereby facilitating the establishment of pre-metastatic niches and subsequent tumor colonization.<sup>106</sup> Exosomes can modulate the ECM to facilitate adhesion of circulating tumor cells, such as by augmenting fibronectin deposition in hepatic tissue. In a previous study, exosomes derived from CRC cells carrying miR-21 have the ability to induce an inflammatory pre-metastatic niche in macrophages through activation of the miR-21/Toll-like receptor 7/IL-6 axis.<sup>107</sup> In a murine model of mCRC, exosomal miR-25-3p selectively targeted transcription factors Krüppel-like factors 2 and 4, leading to downregulation of VEGFR2 expression as well as tight junction components occludin and claudin-5 in the liver and lungs, thereby facilitating extravasation and enhancing vascular permeability.<sup>71</sup> In another study, it was reported that circulating Kupffer cells (KCs) in CRC patients could phagocytose exosomes containing highly expressed miR-135a-5p from the bloodstream and promote the formation of pre-metastatic niches, thereby contributing to the development and clinical severity of CRLM.<sup>98</sup> The expression of miR-221/222 consistently exhibited upregulation in serum exosomal samples obtained from patients with CRLM. Exosomes derived from CRC cells, which carry miR-221/222, play a pivotal role in the establishment of the pre-metastatic niche by suppressing SPINT1 expression and



activating hepatocyte growth factor (HGF), thereby facilitating CRC colonization.<sup>108</sup> Exosomal miR-10a derived from CRC cells suppressed the proliferation and migration activity of normal human lung fibroblasts (NHLFs), as well as downregulates the expression levels of IL-6, IL-8, and IL-1β in NHLFs. Their study provided valuable insights into the role of exosomal miR-10a derived from CRC cells in inducing phenotypic alterations in NHLFs and enhanced our understanding of potential mechanisms underlying CRC lung metastasis.<sup>109</sup> The CXCL10, belonging to the CXC chemokine family, binds to the CXCR3 receptor and exerts its biological effects. Exosomal CXCL10 RNA has the potential to serve as a novel biomarker for liver metastasis from mCRC and could be targeted for prevention and treatment of these patients.<sup>110</sup> In another study, compared with paired normal tissues, the level of ANGPTL1 protein in exosomes derived from CRC is significantly downregulated. Exosomal ANGPTL1, which is mainly taken up by Kupffer cells (KCs) and downregulates intracellular MMP9 levels by inhibiting the JAK2-STAT3 signaling pathway, regulated CRLM and prevented vascular leakage in liver PMNs.<sup>111</sup> Zhang et al. demonstrated that exosomal HSPC111 can modulate the lipid metabolism of CAFs by phosphorylating ATP-citrate lyase, promoting the formation of pre-metastatic niches and distant metastases. Consistently, patients with CRC who have developed liver metastasis exhibited higher levels of HSPC111 in serum exosomes, primary tumors, and CAFs within the liver metastases compared to those without liver metastasis.<sup>112</sup> A study identified that CRC-derived exosomal ADAM17 is involved in the formation of pre-metastatic niches and promotes the migration ability of CRC cells by cleaving E-cadherin. Moreover, overexpression of exosomal ADAM17 in CRC highlighted its function as a tumor-promoting factor, both *in vitro* and *in vivo*.<sup>61</sup>

#### FIBROBLAST-DERIVED EXOSOMES

CAFs, present in the TME adjacent to cancer cells, possess the ability to remodel the ECM, secrete soluble factor and EV, and coordinate their biological functions for the regulation of tumor development. Fibroblasts typically remain quiescent in normal tissue but can undergo activation upon tissue injury.<sup>17,113</sup> Although the exact definition of CAFs is still controversial, cells that are negative for epithelial, endothelial, and leukocyte markers with an elongated morphology and lacking the mutations found within cancer cells, are generally considered as CAFs.<sup>114</sup> CAFs exhibit significant heterogeneity and can be categorized into distinct subtypes based on the differential expression of specific biomarkers. These diverse subtypes possess varying functions, with certain subtypes demonstrating anti-cancer properties while others exhibit ing pro-cancer characteristics.<sup>113</sup> In most instances, CAFs exhibit pro-carcinogenic functions. CAFs play a pivotal role in ECM remodeling through the secretion of ECM components and enzymes. Moreover, CAFs not only serve as physical barriers but also orchestrate the regulation of various cell types within the TME by releasing soluble, EVs, and ECM constituents, thereby promoting cancer cell proliferation, invasion, migration, angiogenesis, and therapy resistance. Additionally, CAFs possess the ability to modulate cancer cell metabolism and facilitate immune evasion. Exosomes derived from CAFs serve as crucial mediators of intercellular communication, involving intercellular communication and exerting regulatory effects on various aspects of the TME in CRC. Notably, their predominant influence lies in promoting cancer cell invasion and migration, and thus facilitate tumor metastasis.<sup>17</sup>

Fibroblasts within the TME undergo activation and transformation into CAFs in response to regulatory exosomes released by tumor cells. Subsequently, CAFs release exosomes that promote cell proliferation, migration, and invasion, while exosomes derived from NFs do not significantly impact the tumor biology of CRC cells.<sup>115</sup> Cyclin-dependent kinase inhibitor 1A (CDKN1A) serves as a potent suppressor of cyclin-dependent kinases. Shi et al. discovered that miR-345-5p exhibited significant upregulation in exosomes derived from CAFs, compared to exosomes from normal fibroblasts (NFs). This elevated expression facilitated the progression and metastasis of CRC by interacting with CDKN1A.<sup>115</sup> Zhang et al. isolated and characterized exosomes derived from CAFs and NFs, revealing that CAFs-derived exosomes exhibited elevated expression levels of miR-17-5p compared to NFs-derived exosomes. In addition, these CAFs-derived exosomes delivered miR-17-5 to CRC cells by directly targeting of runt-related transcription factor 3 (RUNX3) to activate transforming growth factor  $\beta$  (TGF- $\beta$ ) signaling pathway. The interaction between RUNX3 and the oncogene MYC, along with their binding to the promoter region of TGF- $\beta$ 1, ultimately led to the activation of the TGF- $\beta$  pathway and facilitated tumor progression. Moreover, the RUNX3/MYC/ TGF- $\beta$ 1 signaling cascade sustains autocrine TGF- $\beta$ 1 to activate CAFs, inducing increased release of exosomal miR-17-5p into CRC cells, thus establishing a positive feedback loop that promotes CRC progression.<sup>116</sup> Exosomal transfer of miR-92a-3p derived from CAFs regulated CRC cells, resulting in a significant miR-92a-3p levels within CRC cells. Mechanistically, enhanced expression of miR-92a-3p activated the Wnt/β-catenin pathway in CRC cells, thereby promoting cell stemness, EMT, metastasis, and resistance to 5-fluil/ oxaliplatin (5-FU/L-OHP) through direct inhibition of FBXW7 and MOAP1-mediated mitochondrial apoptosis. Additionally, the expression of miR-92a-3p in CRC tissues is significantly upregulated and exhibits a negative correlation with the expression levels of FBXW7 and MOAP1. Moreover, elevated serum levels of miR-92a-3p are strongly associated with metastasis and resistance to chemotherapy in patients with CRC.<sup>117</sup> Jiang et al. demonstrated that exosomes derived from CAFs co-cultured with CRC cells induce the upregulation of miR-181b-3p, which in turn promotes the malignant progression of CRC by targeting SNX2.<sup>118</sup> Zhang et al. revealed that CAFs-Exo facilitated the delivery of miR-625-3p to CRC cells, thereby, invasion, EMT, and chemoresistance through inhibition of the CELF2/WWOX pathway.<sup>119</sup> Additionally, miR-17-5p facilitated the metastasis of CRC through modulation of the RUNX/TGF-β1 signaling axis. Prolonged autocrine TGF-B1 stimulation activated CAFs and enhanced the release of exosomal miR-17-5p into CRC cells, thus establishing a positive feedback loop that drives CRC progression.<sup>116</sup> Hu et al. demonstrated that exosomes derived from CAFs mediated EMT in CRC cells by transferring miR-92a-3p. Mechanistically, upregulated expression of exosomal miR-92a-3p activated the Wnt/β-catenin pathway in CRC cells, thereby promoting cancer progression and chemoresistance by directly inhibiting FBXW7 and MOAP1-mediated mitochondrial apoptosis. Clinically, there is a negative correlation between the expression levels of FBXW7 and MOAP1, while high levels of exosomal miR-92a-3p in serum are closely associated with metastasis and chemotherapy resistance in CRC patients.<sup>117</sup> Furthermore, miR-625-3p



derived from CAFs-Exo may enhance the migratory potential, invasiveness, EMT, and chemoresistance of CRC cells through the inhibition of the CELF2/WWOX signaling pathway.<sup>119</sup> Upregulation of exosomal miR-200b-3p counteracted the stimulatory effect exert hypoxic CAFs on CRC cell proliferation in vitro and in vivo. Moreover, administration of miR-200b-3p agomir enhanced the sensitivity of SW480 cells to 5-FU through downregulation of ZEB1 and E2F3 expression, thus suppressing colon cancer cell migration, invasion, and stemness.<sup>120</sup> In a previous investigation, CAFs were identified as mediators of circN4BP2L2 delivery to CRC cells via exosomes, modulating cell proliferation and migration through the miR-664b-3p/HMGB3 signaling pathway.<sup>121</sup> Hypoxia induced the secretion of cyclin EIF3K from CAFs, and knockdown of circEIF3K in exosomes derived from CAFs significantly attenuated the proliferation, invasion, and angiogenesis of HCT116 and SW620 cells. Furthermore, their study elucidated that the circEIF3K/miR-214/PD-L1 axis mediated hypoxia-induced CRC progression through exosomes derived from CAFs.<sup>122</sup> LINC00659, which is significantly upregulated in exosomes derived from CAFs, facilitated the transfer of exosomal LINC00659 to CRC cells. It interacted with miR-342-3p and enhanced the expression of annexin A2 (ANXA2), promoting CRC cell proliferation, invasion, and migration.<sup>31</sup> Additionally, IncCCAL is transferred from CAFs to cancer cells via exosomes and directly interacts with the mRNA stabilizing protein HuR, thus activating β-catenin pathway. It confers chemoresistance and inhibits apoptosis in CRC cells.<sup>123</sup> LncRNAH19, a maternally imprinted gene, plays a crucial role in embryonic development and growth control. Exosomes derived from CAFs were found to be enriched with IncRNAH19. By transferring exosomal IncRNAH19, CAFs can activate the  $\beta$ -catenin pathway in CRC cells and act as competitive endogenous RNA sponges for miR-141 in CRC, regulating tumor development and chemotherapy resistance.<sup>124</sup> Furthermore, CAFs activated by exosomes derived from SW480 had highly pro-proliferative and pro-angiogenic functions, and showed high expression of proteins promoting angiogenesis (IL8, RAB10, NDRG1) and proliferation (SA1008, FFPS).<sup>67</sup>

#### **IMMUNE CELL-DERIVED EXOSOMES**

The immune system of cancer patients is frequently found to be in a state of suppression. Mounting evidence suggested that exosomes play a pivotal role in the immune microenvironment, facilitating tumor development by coordinating interactions between tumor cells and immune cells.<sup>125</sup> Exosomes derived from immune cells modulate crucial processes within the immune system, including antigen presentation, immune signal transduction, and intercellular communication among immune cells. These exosomes exert their effects by modifying the functionality of recipient cells, thereby establishing a conducive microenvironment for tumor progression and metastasis.<sup>126</sup> The TME harbors a diverse array of immune cells, including DCs, macrophages, neutrophils, NKs, and myeloid-derived suppressor cells (MDSCs), which collectively exert regulatory control over the growth, invasion, and metastasis of tumor cells through the secretion of exosomes.<sup>85</sup> In CRC, particular attention has been directed toward investigating the immunomodulatory role played by exosomes derived from TAMs.<sup>97</sup> Further investigation is warranted to elucidate the functions and mechanisms of immune cell-derived exosomes in mCRC, with the aim of identifying more effective, rational, and safe treatment strategies. It includes exploring the correlation between exosomes derived from immune-suppressed tumors and responses to immune therapy, which may enhance the efficacy of immune checkpoint inhibitors.

Exosomes derived from CRC cells reprograms macrophages, establishing a tumor-promoting microenvironment that facilitates the progression of tumor cells through a series of biological events and ultimately promotes metastasis.<sup>97,127</sup> In a recent study, analysis of exosomes derived from the plasma of CRC patients revealed that the integrin alpha M (ITGAM) subpopulation of macrophage-derived exosomes exhibited an inhibitory effect on development of CRC. Furthermore, this study also identified the potential value of integrin subunit beta 3 (ITGB3) and ITGAM subpopulation of exosomes as biomarkers and therapeutic targets for early diagnosis of CRC.<sup>37</sup> The Switch/Sucrose Non-Fermentable (SWI/SNF) complex plays a crucial role in early embryonic development, inflammation, and immune response. Numerous studies have consistently reported a significant decrease in BRG1 expression, which serves as a core motor of the SWI/SNF complex, within cancerous tissues. In the TME, exosomes derived from M2 macrophages actively downregulated BRG1 through miR-21-5p and miR-155-5p mechanisms to facilitate migration and invasion of cells. Furthermore, M2 macrophages secrete exosomes containing WNT that induce CRC stem cell activity involved in metastasis development.<sup>32</sup> In addition to macrophages, exosomes derived from N2-like neutrophil-derived exosomes promoted EMT and angiogenesis. The aberrant expression of miR-4780 in exosomes derived from N2-like neutrophil-derived exosomes promoted EMT and angiogenesis. The aberrant expression of miR-4780 in exosomes derived from N2-like neutrophils has been demonstrated to play a pivotal role in the metastasis and progression of tumors in CRC models featuring liver metastasis. Moreover, miR-4780 exhibited regulatory effects on its target gene SOX11, thereby EMT and angiogenesis in CRC.<sup>128</sup>

#### **OTHER CELLS-DERIVED EXOSOMES**

The establishment of TME and the progression and metastasis of tumors rely not only on the interplay among tumor cells, but also on the cross-talk between tumor cells and diverse cell.<sup>30</sup> In the context of CRC, exosomes derived from diverse cellular sources within the TME, including tumor cells, fibroblasts, immune cells, as well as MSCs, hepatocytes, and Schwann cells, collectively orchestrate various processes involved in tumor cell metastasis.<sup>66</sup> These processes encompass augmented migratory capacity of primary tumors, facilitation of EMT, suppression of immune response, establishment of pre-metastatic niches, and sustenance of growth at secondary metastatic sites.

MSCs exist in many tissues and have the ability to trigger immune response, tissue healing, cell proliferation, and tumor progression control. As regulators of the TME, MSCs release exosomes and their involvement in tumor occurrence and metastasis is controlled by many growth factor receptors, including EGFR and PDGFR.<sup>129</sup> Elevated ITGA6 expression is correlated with unfavorable overall survival in patients with CRC. Exosomes derived from MSCs carrying miR-3940-5p target ITGA6 and subsequently deactivate TGF-β1, thereby inhibiting invasion and EMT of CRC cells, as well as suppressing tumor growth and metastasis.<sup>130</sup> Jahangiri et al. discovered that exosomes derived from MSCs



possess the capability to impede the proliferation, migration, invasion, and metastasis of CRC cells through modulation of the miR-100/ mTOR/miR-143 axis, while concurrently inducing apoptosis. Furthermore, it was proposed that exosomal miR-100 derived from MSCs could be regarded as a promising therapeutic strategy for CRC.<sup>131</sup> Exosomes derived from human umbilical cord MSCs (hUC-MSCs) carrying miR-1827 suppressed M2 macrophage polarization by downregulating SUCNR1 expression, thus inhibiting the proliferation, migration, and invasion of CRC cells. Moreover, in vivo experiments demonstrated that exosomes derived from hUC-MSCs carrying miR-1827 effectively block CRLM. These findings provided a theoretical foundation for comprehending the mechanism underlying exosome-based targeted therapy for CRC. <sup>132</sup> Previous studies have demonstrated that exosomes derived from hUC-MSCs and Human bone marrow MSCs (hBM-MSCs) containing tumor-suppressive miR-3940-5p can inhibit the proliferation, migration, and invasion of CRC cells by regulating the ITGA6/TGF-β1 signaling pathway.<sup>133</sup> In a separate investigation, miR-203a-3p derived from EVs originating from hepatocytes played a pivotal role in the hepatic colonization of CRC cells. Mechanistically, miR-203a-3p derived from hepatocyte-derived EVs upregulated E-cadherin expression in CRC cells, downregulated Src expression and facilitated MET in CRC cells.<sup>134</sup> Wang et al. demonstrated that EVs derived from hepatocytes in fatty liver enhanced the progression of CRLM by promoting oncogenic YAP signaling and creating an immune-suppressive microenvironment.<sup>135</sup> The involvement of the peripheral nervous system (PNS) in TME components has been documented in relation to the progression and metastasis of diverse cancer types.<sup>136</sup> Schwann cells, the principal glial cells in the PNS, have been demonstrated to actively contribute to the dissemination and metastasis of lung and pancreatic cancer through direct cellular interaction or paracrine signaling.<sup>137,138</sup> In a recent study, it was discovered that the intercellular communication between Schwann cells and colon cancer cells played a crucial role in promoting the proliferation and metastasis of colon cancer. Furthermore, nerve growth factor (NGF) and miR-21-5p from EVs had been identified as potential therapeutic targets for combating colon cancer.<sup>139</sup>

#### APPLICATION OF EXOSOMES IN THE DIAGNOSIS AND TREATMENT

Exosomes, emerging liquid biopsy options and being stable information carriers, hold great promise as biomarkers for the diagnosis or treatment of metastasis of CRC.<sup>33</sup> With many advantages, including good stability and the ability to carry different bioactive molecules, exosomes participate in intercellular communication and regulate the proliferation, invasion and metastasis of tumor cells.<sup>18</sup> In addition, exosomes are significant nanomaterials due to their excellent biocompatibility and long-term stability in the bloodstream, making them ideal targeted drug carriers for cancer treatment.<sup>23,140</sup> As a result, exosomes possess immense potential and value in tumor diagnosis, differentiation, and therapy.

Exosomes exhibit remarkable stability and are rich in bioactive molecules such as ncRNAs and proteins that play a key role in multiple steps of tumor metastasis.<sup>18</sup> Additionally, exosomes can be readily detected in various bodily fluids including blood, urine, cerebrospinal fluid, and saliva, thereby exhibiting immense potential as diagnostic markers for metastasis of CRC.<sup>23</sup> The diagnostic significance of exosome-related biomarkers in the metastasis of CRC had been reported by several previous studies. In a recent study, liquid biopsy based on a panel of exosomal miRNAs (miR-181b, miR-193b, miR-195, and miR-411) robustly identified T1 CRC patients at risk for lymph node metastasis in the preoperative setting (AUC, 0.84; 95%CI 0.70–0.98).<sup>141</sup> In another study, the accuracy of epithelial growth factor (EGFR) and ITGB3 in plasma exosomes for distinguishing CRC from mCRC was (AUC, 0.91; 95%CI 0.76–1.00) and (AUC, 0.87; 95%CI 0.72–1.00).<sup>37</sup> Zeng et al. found that the expression level of miR-25-3p from circulating exosomes was significantly higher in CRC patients with metastasis than those without metastasis and could be used as a blood biomarker for CRC metastasis.<sup>71</sup> In a study, it was discovered that tumor-secreted exosomal miR-208b promoted the expansion of regulatory T cells by targeting programmed cell death factor 4 (PDCD4), potentially leading to reduced sensitivity to oxaliplatin therapy, offering a novel target for immunotherapy.<sup>142</sup> Moreover, Shao et al. evaluated the level of immune cell infiltration in the microenvironment by constructing a risk model of exosome related genes, resulting in a high degree of accuracy. This approach offered a novel perspective for evaluating immunotherapy efficacy in patients with CRC.<sup>143</sup>

The treatment of metastatic malignant tumors using exosome-based therapies has witnessed a gradual increase in recent years, primarily encompassing drug delivery vectors and targeted therapy specifically designed for exosomes.<sup>144–146</sup> Considering the intricate TME surrounding CRC, achieving safe and efficacious drug delivery to CRC cells currently presents one of the most significant challenges. In recent decades, significant advancements have been achieved in the field of targeted therapies for CRC through the utilization of drug delivery systems based on synthetic nanomaterials.<sup>12,36,147–149</sup> Compared to traditional nanoparticles, EVs or exosomes offered significant advantages in terms of biocompatibility, phagocytic clearance evasion, and intrinsic homing ability. As such, they have the potential to become a new generation of drug delivery systems.<sup>12,150,151</sup> hBM-MSCs exhibit remarkable proficiency in large-scale exosome production, thereby offering a promising avenue for the delivery of miRNAs in cancer therapy. In a previous study, exosomal miR-99b-5p secreted by hBM-MSCs exerted inhibitory effects on the proliferation, invasion, and migration of CRC cells through targeted regulation of FGFR3.<sup>152</sup> Additionally, Liang et al. used engineered exosomes to simultaneously deliver anticancer drug 5-FU and miR-21 inhibitor oligonucleotide (Mir-21i) to CRC cells expressing Her2, and showed good therapeutic potential.<sup>153</sup> Additionally, Huang et al. developed a novel engineered exosome delivery system based on primary cells, enabling simultaneous delivery of siRNAs targeting Coiled-coil domain-containing protein 80 (CCDC80) and enhanced sensitivity to chemotherapy. Their findings demonstrated the remarkable anti-tumor metastasis efficacy of this exosome in a mice model of CRLM.<sup>140</sup> The findings of a recent study demonstrated that exosome circTUBGCP4, produced by CRC cells, stimulated vascular endothelial cell tilt to enhance angiogenesis and promote tumor metastasis through the activation of the Akt signaling pathway. These results offered novel insights for future applications of engineered exosomes in the treatment of mCRC.<sup>76</sup> Van et al.'s isolated EVs from CT26 colon cancer cells and 4T1 mouse breast cancer cells, followed by electroporation-mediated loading



with doxorubicin (DOX). The findings demonstrated that the vector exhibited excellent biocompatibility, high drug-loading capacity, controllable drug release kinetics, and remarkable selectivity toward CRC cells. Notably, it significantly impeded tumor growth in BALB/c mice bearing colorectal tumor.<sup>150</sup> Reprogramming TAMs to adopt a pro-inflammatory M1 phenotype represents an innovative strategy for tumor immunotherapy, while the M2 phenotype is regulated by crucial transcription factors such as signal transducers and transcriptional activator 6 (STAT 6).<sup>154</sup> The natural killer group 2 member D (NKG2D) receptor facilitates the interaction between anti-tumor EVs derived from NK cells and spheroids from CRC cells.<sup>155</sup> CRC cells exhibit elevated expression levels of the NKG2D ligand MICA/B, and the susceptibility of tumor tissue to NK cell-derived EVs is associated with differential expression of the NKG2D ligand MICA/B, which can be inhibited by anti-NKG2D antibodies. Their study demonstrated the potential of engineered exosomes targeting NK cells in cancer immunotherapy.<sup>155</sup> Programmed cell death ligand-1 (PD-L1) antibodies bind to and are consumed by the exosome PD-L1 in peripheral blood.<sup>156</sup> One study found that sulindac down-regulated PD-L1 by blocking the NF-κB signaling pathway, thereby reducing the amount of exosome PD-L1 secreted by tumor cells and enhancing the efficacy of PD-L1 immunotherapy in patients with pMMR phenotype CRC.<sup>157</sup> Exosomes present an appealing therapeutic target for malignancies with metastatic disease. Various drugs have been identified to inhibit tumor-derived exosomes by targeting their formation, suggesting the possibility of repurposing them for metastatic anticancer therapy. For instance, apatinib used in the treatment of mCRC may exhibit a potential anti-tumor mechanism in CRC through multiple pathways, including inhibition of tumor-promoting exosome secretion.<sup>145</sup> These novel approaches for anti-tumor activity offered a fresh perspective on the treatment of exosomes in CRC.

#### **CURRENT CHALLENGES AND FUTURE PROSPECTS**

Metastasis is the primary determinant of unfavorable prognosis in patients with CRC.<sup>5</sup> Exosomes play a pivotal role in facilitating bidirectional communication between CRC cells and recipient cells, as well as orchestrating the establishment of a dynamic network within the TME.<sup>30</sup> This regulation is crucial for controlling pivotal processes such as tumor cell proliferation, invasion and migration, EMT, remodeling of ECM leading to physical alterations and metastasis angiogenesis, modulation of vascular permeability, along with immunosuppressive effects.<sup>17,30</sup> (Table 1). In recent years, there has been an increasing use of exosome-based liquid biopsy and drug delivery systems in clinical diagnosis and treatment, indicating the broad application potential of exosomes in the medical field.<sup>23,140</sup> It provides a novel perspective for future diagnosis, treatment, and prognostic assessment of mCRC. However, there remain numerous challenges associated with utilizing exosomes for the diagnosis and treatment of CRC.

Exosomes, as crucial mediators of communication within the circulatory system, exert distinct regulatory roles in various events impacting tumor cell metastasis.<sup>23</sup> Exosomes derived from diverse cellular sources exhibit unique molecular compositions and respond to intricate genetic information, encompassing the promotion of tumor cell invasion and migration, facilitation of angiogenesis, modulation of tumor immunity, and facilitation of pre-metastatic niche formation.<sup>17,18</sup> Exosomes play a pivotal role in either promoting or inhibiting tumor metastasis, thereby indicating their potential utility as valuable diagnostic and prognostic markers for alterations in the TME and mCRC.<sup>18</sup> Enhanced understanding of the underlying molecular mechanisms governing tumor metastasis and intercellular communication after various treatments will facilitate the refinement of therapeutic strategies aimed at eradicating residual tumors.<sup>18,33</sup> Despite notable progress made in understanding the functions and mechanisms of exosomes in metastasis of CRC, several pivotal questions persist unanswered for further investigation. Specifically, a comprehensive comprehension of exosome communication within the TME throughout metastatic progression is lacking, encompassing aspects such as their recognition, binding dynamics, and functional mechanisms across various recipient cells.<sup>30</sup> Although the current evidence substantiates the pivotal role of exosomes in facilitating tumor cell dissemination through EMT induction, their involvement in metastatic growth, such as reactivation of metastasis or MET, remains largely unexplored.<sup>30</sup> Moreover, future investigations on exosomes in cancer should prioritize exploring the heterogeneity of exosomes derived from tumor patients and elucidating their specificity in carrying biological information.<sup>37,158,160</sup> Additionally, the underlying mechanisms behind the selective packaging of cellular components into exosomes remain largely unknown. Similarly, there is limited understanding regarding the impact of exosomes derived from metastatic cells on primary tumors, as well as the precise quantity and timing required for functional effects on recipient cells during the metastatic process. Furthermore, the mechanisms underlying pre-metastatic niche formation are still poorly understood. The 'seed/soil' hypothesis provided the framework for explaining the phenomenon of tumor metastasis.<sup>161</sup> Addressing these inquiries will enhance our comprehension of exosome involvement in metastasis of CRC and offer novel insights for developing exosome-based anti-metastatic therapies.

Currently, extensive research has elucidated the crucial role of exosomes as mediators of intercellular communication by investigating their structure, biological origin, secretion, and function.<sup>23,29</sup> Furthermore, their significant potential in the diagnosis and treatment of mCRC has been gradually unveiled. Previous preclinical and clinical evidence had demonstrated immense promise for utilizing exosomes as diagnostic biomarkers in liquid biopsies and as effective therapeutic agents for various diseases, including tumors and autoimmune disorders.<sup>48,140</sup> Particularly, the noteworthy value is their efficacy as efficient biological carriers for targeted drug delivery.<sup>36,161</sup> In contrast to conventional nanoparticles, exosomes of endogenous origin exhibit superior biocompatibility, evade phagocytosis, and have significantly reduced immunogenicity.<sup>36,162,163</sup> Specifically, exosomes can easily infiltrate the ECM of tumor tissues and are less affected by TME, thus overcoming the challenges of cell therapy.<sup>164</sup> The research and application of certain nanomaterials in tumor-related diseases concurrently offer novel insights for the advancement of exosomes as drug carriers.<sup>147,148,163</sup> However, in order to translate exosomes from the laboratory to the clinic, several challenges must be overcome. Firstly, liquid biopsies need to meet multiple criteria including minimal sample requirements, simplicity, efficiency, affordability, high sensitivity and specificity as well as reproducibility for blood, urine and other samples. The technology of microfluidic separation and detection holds promising potential for clinical application. It is crucial to establish standardized

| Table 1. Function and mechanism of exosomes derived from different cells as communication mediators in metastasis of CRC |                 |                |   |                        |                             |  |  |
|--|-----------------|----------------|---|------------------------|-----------------------------|--|--|
| Origin   | Exosomal cargos | Regulation     | Mechanism   | Biological function    | Reference                   |  |  |
| CRC  | miR-106b-3p     | Up-regulated   | Inhibit the expression of DLC-1   | Invasion and migration | Liu et al. <sup>43</sup>    |  |  |
| CRC  | LINC01915       | Down-regulated | Inhibit miR-92a-3p/KLF4/CH25H axis  | Invasion and migration | Zhou et al. <sup>68</sup>   |  |  |
| CRC  | circFMN2        | Up-regulated   | Activate miR-338- 3 p/MSI1 pathway  | Invasion and migration | Yu et al. <sup>52</sup>     |  |  |
| CRC  | miR-146a-5p     | Up-regulated   | Promote the expression of Cxcl12/Cxcr7 and regulate JAK2-STAT3/NF-κB pathway  | Invasion and migration | Wang et al. <sup>70</sup>   |  |  |
| CRC  | miR-155-5p      | Up-regulated   | Promote the experssion of Cxcl12/Cxcr7 and regulate JAK2-STAT3/NF-κB pathway  | Invasion and migration | Wang et al. <sup>70</sup>   |  |  |
| CRC  | circ-ABCC1      | Up-regulated   | Activate Wnt/β-catenin pathway  | Invasion and migration | Zhao et al. <sup>53</sup>   |  |  |
| CRC  | ADAM17          | Up-regulated   | Block the E-cadherin connections  | Invasion and migration | Sun et al. <sup>61</sup>    |  |  |
| CRC  | PCAT1           | Up-regulated   | Downregulate miR-329-3p and upregulate<br>Netrin-1 and CD146  | Invasion and migration | Fang et al. <sup>58</sup>   |  |  |
| CRC  | circCOG2        | Up-regulated   | Inhibit miR-1305/TGF-β2/SMAD3 pathway   | Invasion and migration | Gao et al. <sup>54</sup>    |  |  |
| CRC  | circEPB41L2     | Down-regulated | Regulate PTEN/AKT pathway and sponge miR-<br>21-5p and miR-942-5p   | Invasion and migration | Jiang et al. <sup>49</sup>  |  |  |
| CRC  | miR-548c-5p     | Down-regulated | Regulate HIF-1α/CDC42 pathway   | Invasion and migration | Yan et al. <sup>46</sup>    |  |  |
| CRC  | UCA1            | Up-regulated   | Inhibit miR-143/MYO6 axis   | Invasion and migration | Luan et al. <sup>59</sup>   |  |  |
| CRC  | MALAT1          | Up-regulated   | Activate the PI3K/AKT/mTOR pathway to sponge miR-26a/26b and regulate FUT4  | Invasion and migration | Xu et al. <sup>60</sup>     |  |  |
| CRC  | miR-1229        | Up-regulated   | Target HIPK2 and activate the VEGF pathway  | Angiogenesis           | Hu et al. <sup>74</sup>     |  |  |
| CRC  | circTUBGCP4     | Up-regulated   | Upregulate PDK2 to activate Akt signaling<br>pathway by sponging miR-146b-3p  | Angiogenesis           | Chen et al. <sup>76</sup>   |  |  |
| CRC  | B7-H3           | Up-regulated   | Activate the AKT1/mTOR/VEGFA pathway  | Angiogenesis           | Wu et al. <sup>79</sup>     |  |  |
| CRC  | GDF15           | Up-regulated   | Inhibit the Smad signaling pathway and<br>increase POSTN levels   | Angiogenesis           | Zheng et al. <sup>77</sup>  |  |  |
| CRC  | miR-25-3p       | Up-regulated   | Regulates the expression of VEGFR2, ZO-1,<br>Occludin and Claudin5 in endothelial cells by<br>targeting KLF2 and KLF4 | Angiogenesis           | Zeng et al. <sup>71</sup>   |  |  |
| CRC  | miR-17-5p       | Up-regulated   | Promote the expression of PD-L1 to inhibit<br>SPOP  | Immune escape          | Sun et al. <sup>96</sup>    |  |  |
| CRC  | circVCP         | Up-regulated   | Regulate the miR-9-5p/NRP1 axis   | Immune escape          | Tang et al. <sup>103</sup>  |  |  |
| CRC  | IncRNA BANCR    | Up-regulated   | Activate RhoA/Rock pathway by recruiting IGF2BP2  | Immune escape          | Ding et al. <sup>102</sup>  |  |  |
| CRC  | miR-203a-3p     | Up-regulated   | Regulate PTEN and activate the PI3K/Akt pathway.  | Immune escape          | Pei et al. <sup>95</sup>    |  |  |
| CRC  | IncRNA RPPH1    | Up-regulated   | Interacte with TUBB3 to prevent its ubiquitination  | Immune escape          | Liang et al. <sup>100</sup> |  |  |

CellPress
 OPEN ACCESS

(Continued on next page)

| Table 1. Continued |                 |                |   |                                    |                             |  |  |
|--------------------|-----------------|----------------|---|------------------------------------|-----------------------------|--|--|
| Origin             | Exosomal cargos | Regulation     | Mechanism   | Biological function                | Reference                   |  |  |
| CRC                | miR-106b-5p     | Up-regulated   | Activate the PI3K $\gamma$ /AKT/mTOR signaling cascade by directly suppressing PDCD4  | Immune escape                      | Yang et al. <sup>97</sup>   |  |  |
| CRC                | LncRNA KCNQ1OT1 | Up-regulated   | Modulate the miR-30a-5p/USP22 axis and regulate PD-1 ubiquitination   | Immune escape                      | Xian et al. <sup>57</sup>   |  |  |
| CRC                | miR-135a-5 p    | Up-regulated   | Activate the large tumor suppressor kinase<br>2-yes-associated protein-matrix<br>metalloproteinase 7 axis   | Formation of pre-metastatic niches | Sun et al. <sup>98</sup>    |  |  |
| CRC                | miR-21          | Up-regulated   | Activate miR-21/Toll-like receptor 7/IL-6 axis  | Formation of pre-metastatic niches | Liang et al. <sup>153</sup> |  |  |
| CRC                | circPABPC1      | Up-regulated   | Protect ADAM19 and BMP4 from miR-874-/<br>miR-1292-mediated degradation.  | Formation of pre-metastatic niches | Li et al. <sup>56</sup>     |  |  |
| CRC                | ANGPTL1         | Down-regulated | Downregulates intracellular MMP9 levels by inhibiting the JAK2-STAT3 signaling pathway  | Formation of pre-metastatic niches | Jiang et al. <sup>111</sup> |  |  |
| CRC                | miR-221/222     | Up-regulated   | Activate HGF by inhibiting SPINT1 expression  | Formation of pre-metastatic niches | Tian et al. <sup>108</sup>  |  |  |
| CRC                | miR-10a         | Up-regulated   | Reduces the proliferation and migration<br>activity of NHLFs and the expression levels of<br>IL-6, IL-8, and IL-1β in NHLFs   | Formation of pre-metastatic niches | Wang et al. <sup>109</sup>  |  |  |
| CAFs               | miR-200b-3p     | Up-regulated   | Inhibit colon cancer cell migration, invasion,<br>and stemness by downregulating the<br>expression of ZEB1 and E2F3   | Invasion and migration             | Gong et al. <sup>120</sup>  |  |  |
| CAFs               | miR-345-5p      | Up-regulated   | Promote CRC progression and metastasis by interacting with CDKN1A   | Invasion and migration             | Shi et al. <sup>115</sup>   |  |  |
| CAFs               | miR-92a-3p      | Up-regulated   | Activate the Wnt/β-catenin pathway in CRC<br>cells and promotes cell stemness, EMT,<br>metastasis, and resistance to 5-FU/L-OHP by<br>directly inhibiting FBXW7 and MOAP1-<br>mediated mitochondrial apoptosis. | Invasion and migration             | Zhou et al. <sup>68</sup>   |  |  |
| CAFs               | miR-181b-3p     | Up-regulated   | Promote malignant progression of CRC by targeting SNX2  | Invasion and migration             | Jiang et al. <sup>118</sup> |  |  |
| CAFs               | miR-625-3p      | Up-regulated   | Promote EMT and drug resistance in CRC by inhibiting the CELF2/WWOX pathway   | Invasion and migration             | Zhang et al. <sup>119</sup> |  |  |
| CAFs               | miR-17-5p       | Up-regulated   | Promotes colorectal cancer metastasis by regulating the RUNX3/MYC/TGF-β1 signaling axis   | Invasion and migration             | Zhang et al. <sup>116</sup> |  |  |
| CAFs               | circN4BP2L2     | Up-regulated   | Regulate cell proliferation and migration through the miR-664b-3p/HMGB3 pathway   | Invasion and migration             | Yang et al. <sup>121</sup>  |  |  |
| CAFs               | LINC00659       | Up-regulated   | Bind with miR-342-3p, and increase the expression of ANXA2  | Invasion and migration             | Zhou et al. <sup>31</sup>   |  |  |
|                    |                 |                |   |                                    | (Continued on next pag      |  |  |

14

iScience 27, 109350, April 19, 2024

iScience Review

CellPress OPEN ACCESS

| Table 1. Continued         |                 |                |  |  |                                 |  |  |  |
|----------------------------|-----------------|----------------|--|--|---------------------------------|--|--|--|
| Origin                     | Exosomal cargos | Regulation     | Mechanism  | Biological function                                    | Reference                       |  |  |  |
| CAFs                       | IncCCAL         | Up-regulated   | Interact with mRNA stabilizing protein HuR and activates the $\beta\mbox{-}catenin$ pathway  | Invasion and migration                                 | Deng et al. <sup>123</sup>      |  |  |  |
| CAFs                       | LncRNAH19       | Up-regulated   | Activate the β-catenin pathway and act as a competitive endogenous RNA sponge for miR-<br>141  | Invasion and migration                                 | Ren et al. <sup>124</sup>       |  |  |  |
| CAFs                       | circEIF3K       | Up-regulated   | Mediate hypoxia-induced CRC progression via<br>circEIF3K/miR-214/PD-L1   | Invasion and migration                                 | Yang et al. <sup>122</sup>      |  |  |  |
| CAFs                       | miR-200b-3p     | Up-regulated   | Inhibit colon cancer cell migration, invasion,<br>and stemness by downregulating the<br>expression of ZEB1 and E2F3                  | Invasion and migration                                 | Gong et al. <sup>120</sup>      |  |  |  |
| M2 macrophages             | BRG1            | Down-regulated | Downregulate BRG1 through miR-21-5p and<br>miR-155-5p to regulate the expression of SWI/<br>SNF complex                              | Invasion and migration                                 | Lan et al. <sup>32</sup>        |  |  |  |
| N2-like neutrophil         | miR-4780        | Up-regulated   | Target gene SOX11 to influence EMT and angiogenesis  | Invasion, migration and angiogenesis                   | Wang et al. <sup>128</sup>      |  |  |  |
| MSC                        | miR-3940-5p     | Up-regulated   | Target ITGA6 and inactivate TGF- $\beta$ 1 to inhibit the invasion and EMT   | Invasion and migration                                 | Li et al. <sup>130</sup>        |  |  |  |
| MSC                        | miR-100         | Up-regulated   | Regulate the miR-100/mTOR/miR-143 axis   | Invasion and migration                                 | Jahangiri et al. <sup>131</sup> |  |  |  |
| hUC-MSC                    | miR-1827        | Up-regulated   | Inhibit M2 macrophage polarization by<br>downregulating SUCNR1 expression  | Immune escape  | Chen et al. <sup>132</sup>      |  |  |  |
| hUC-MSC                    | miR-3940-5p     | Down-regulated | Active ITGA6/TGF-β1 pathway  | Invasion and migration                                 | Li et al. <sup>130</sup>        |  |  |  |
| hepatocytes                | miR-203a-3p     | Up-regulated   | Increase the expression of E-cadherin and inhibit the expression of Src to promote MET   | Formation of pre-metastatic niches                     | Xu et al. <sup>134</sup>        |  |  |  |
| hepatocytes in fatty liver | Rab27a          | Up-regulated   | Enhance the progression of CRC liver<br>metastasis by promoting oncogenicYAP<br>signaling and immune-suppressive<br>microenvironment | Formation of pre-metastatic niches and immun<br>escape | Huang and Feng <sup>82</sup>    |  |  |  |

CRC, Colorectal cancer; CRLM, Colorectal cancer liver metastasis; TME, Tumor microenvironment; ECM, Extracellular matrix; NKs, Natural killer cells; TAMs, Tumor-associated macrophages; EV, Extracellular vesicle; DCs, Dendritic cells; ECs, Endothelial cells; KCs, Kupffer cells; NHLFs, Normal human lung fibroblasts; MSCs, Mesenchymal stem cells; TDEs, Tumor-derived exosomes; BRG1, Brahma-related gene 1; MALAT1, Metastasis-associated lung adenocarcinoma transcript 1; EMT, Epithelial-mesenchymal transition; TUBB3, Tubulinβ-III; PDCD4: Programmed cell death 4; USP22, Ubiquitin-specific protease 22; IFN-2, Interferon-regulated factor 2; DLC-1, Dynein light chain 1; HIF-1α, Hypoxia-inducible factor 1 alpha; CDC42, Cell division control protein 42; ZEB1, zinc finger E-Box binding homeobox 1; SPOP, Speckle-type POZ protein; PD-L1, programmed death ligand 1; MSI1, Musashi1; PTEN, phosphatase and tensin homolog; ADAM 17, A disintegrin and metalloproteinase 17; BMP4, bone morphogenetic protein 4; GDF15, Growth/differentiation factor 15; ZO-1, Zonula occlusion 1; PAD4, Peptidyl arginine deiminase 4; CDKN1A, Cyclin-dependent kinase inhibitor 1A; NFs, Normal fibroblasts; SNF, Sucrose Non-Fermentable; RUNX3, runt-related transcription factor 3; ANXA2, annexin A2; YAP, Yes-associated protein; hUC-MSCs: human umbilical cord mesenchymal stem cells; MDSCs, Myeloid-derived suppressor cells; PNS, peripheral nervous system; NGF, nerve growth factor; PBA, Proximity-dependent barcoding assay; MET, mesenchymal-epithelial transition; EPR, enhanced permeability and retention; HGF, hepatocyte growth factor.

CellPress OPEN ACCESS





procedures for exosome isolation, purification and characterization while considering sample handling protocols. Meanwhile, developing high-throughput exosome analysis platforms that integrate various techniques such as nanoparticle tracking analysis (NTA), electron microscopy (EM) and flow cytometry can lead to improved exosome characterization.<sup>165</sup> Secondly, although exosomes hold great promise as novel biomarkers for cancer diagnosis and prognosis; enhancing their specificity and functional relevance remains a challenge. Proximity dependent barcode analysis (PBA), as a new high-throughput method for single exosome analysis, can distinguish different exosome subgroups by identifying specific combinations of exosome membrane proteins.<sup>159</sup> Third, the sources and composition of naturally occurring exosomes are diverse and intricate, rendering the sorting mechanism of exosomes not fully comprehended. Rigorous biosafety verification is imperative when employing natural or modified exosomes as drug delivery vehicles. Accurate identification of exosomal composition holds great potential for enhancing their therapeutic effectiveness while minimizing side effects. Lastly, addressing the challenge of mass production of engineered exosomes for therapeutic purposes emerges as a crucial task in future clinical translational processes.

#### Conclusions

Exosomes, serving as intercellular communication mediators, play a pivotal role in the progression of tumors. Gaining an in-depth understanding of the function and mechanism of exosomes derived from various cell types in mCRC contributes to a more profound comprehension of this disease. In recent years, research on tumor exosomes has explored their potential as biomarkers and drug delivery vehicles, thereby advancing precision cancer medicine. However, the complex intercellular communication mediated by exosomes and the engineered production of exosomes still pose numerous challenges. Overall, exosomes are anticipated to enhance our comprehension of the biology and oncology associated with CRC metastases, as well as facilitate the development of novel strategies for tumor diagnosis and treatment.

#### ACKNOWLEDGMENTS

The research was funded by the following grants : General Program of National Natural Science Foundation of China (No.82373293); Research Project of Jiangsu Commission of Health (No. ZD2022063); The fifth phase of the "333 Project" scientific research project of Jiangsu Province (No. BRA2020091); Primary Research and Development Plan of Jiangsu Province (No. BE2019759); the National Key Research and Development Program of China (No. 2018YFE0127300).

#### **AUTHOR CONTRIBUTIONS**

Conceptualization, Chunzhao Yu, Yimin E, and Chen Lu; writing—original draft preparation, Yimin E, Pengcheng Ji, Minjie Meng and Kuixuan Zhu; writing—review and editing, Chunzhao Yu, Jing Sun, and Zhengxia Liu; supervision, Chunzhao Yu and Wenyuan Li. The Figures in this manuscript were drawn by Figdraw, Yimin E. Yimin E, Chen Lu and Kuixuan Zhu contributed equally to this work and should be considered as co-first authors. All authors read and approved the final manuscript.

#### **DECLARATION OF INTERESTS**

The authors have declared that no competing interest exists.

#### REFERENCES

- Sung, H., Ferlay, J., Siegel, R.L., Laversanne, M., Soerjomataram, I., Jemal, A., and Bray, F. (2021). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA. Cancer J. Clin. 71, 209–249. https://doi.org/10.3322/caac.21660.
- Santos, D.A.R., Gaiteiro, C., Santos, M., Santos, L., Dinis-Ribeiro, M., and Lima, L. (2023). MicroRNA Biomarkers as Promising Tools for Early Colorectal Cancer Screening-A Comprehensive Review. Int. J. Mol. Sci. 24, 11023. https://doi.org/10.3390/ ijms241311023.
- Sarvizadeh, M., Ghasemi, F., Tavakoli, F., Sadat Khatami, S., Razi, E., Sharifi, H., Biouki, N.M., and Taghizadeh, M. (2019). Vaccines for colorectal cancer: an update. J. Cell. Biochem. 120, 8815–8828. https://doi.org/ 10.1002/jcb.28179.
- Zhu, X., and Li, S. (2023). Ferroptosis, Necroptosis, and pyroptosis in gastrointestinal cancers: the chief culprits of tumor progression and drug resistance. Adv. Sci. 10, e2300824. https://doi.org/10. 1002/advs.202300824.

- Dekker, E., Tanis, P.J., Vleugels, J.L.A., Kasi, P.M., and Wallace, M.B. (2019). Colorectal cancer. Lancet 394, 1467–1480. https://doi. org/10.1016/s0140-6736(19)32319-0.
- Zhou, H., Liu, Z., Wang, Y., Wen, X., Amador, E.H., Yuan, L., Ran, X., Xiong, L., Ran, Y., Chen, W., and Wen, Y. (2022). Colorectal liver metastasis: molecular mechanism and interventional therapy. Signal Transduct. Target. Ther. 7, 70. https://doi.org/10.1038/ s41392-022-00722-2.
- Abdul Pari, A.A., Singhal, M., and Augustin, H.G. (2021). Emerging paradigms in metastasis research. J. Exp. Med. 218, e20190218. https://doi.org/10.1084/jem. 20190218.
- Ni, Y., Liang, Y., Li, M., Lin, Y., Zou, X., Han, F., Cao, J., and Li, L. (2023). The updates on metastatic mechanism and treatment of colorectal cancer. Pathol. Res. Pract. 251, 154837. https://doi.org/10.1016/j.prp.2023. 154837.
- Massagué, J., and Obenauf, A.C. (2016). Metastatic colonization by circulating tumour cells. Nature 529, 298–306. https:// doi.org/10.1038/nature17038.

- Lahooti, B., Akwii, R.G., Zahra, F.T., Sajib, M.S., Lamprou, M., Alobaida, A., Lionakis, M.S., Mattheolabakis, G., and Mikelis, C.M. (2023). Targeting endothelial permeability in the EPR effect. J. Control. Release 361, 212–235. https://doi.org/10.1016/j.jconrel. 2023.07.039.
- Chen, E., and Yu, J. (2023). The role and metabolic adaptations of neutrophils in premetastatic niches. Biomark. Res. 11, 50. https://doi.org/10.1186/s40364-023-00493-6 (2023.
- Xiong, L., Wei, Y., Jia, Q., Chen, J., Chen, T., Yuan, J., Pi, C., Liu, H., Tang, J., Yin, S., et al. (2023). The application of extracellular vesicles in colorectal cancer metastasis and drug resistance: recent advances and trends. J. Nanobiotechnol. 21, 143. https:// doi.org/10.1186/s12951-023-01888-1 (2023.
- Franco, P.I.R., Rodrigues, A.P., de Menezes, L.B., and Miguel, M.P. (2020). Tumor microenvironment components: Allies of cancer progression. Pathol. Res. Pract. 216. https://doi.org/10.1016/j.prp.2019.152729.
- 14. Wu, T., and Dai, Y. (2017). Tumor microenvironment and therapeutic

response. Cancer Lett. 387, 61–68. https:// doi.org/10.1016/j.canlet.2016.01.043.

- Roma-Rodrigues, C., Mendes, R., Baptista, P.V., and Fernandes, A.R. (2019). Targeting Tumor Microenvironment for Cancer Therapy. Int. J. Mol. Sci. 20, 840. https://doi. org/10.3390/ijms20040840.
- Denton, A.E., Roberts, E.W., and Fearon, D.T. (2018). In Stromal Immunology, Vol 1060, B.M.J. Owens and M.A. Lakins, eds. Advances in Experimental Medicine and Biology, pp. 99–114.
- Li, C., Teixeira, A.F., Zhu, H.J., and ten Dijke, P. (2021). Cancer associated-fibroblastderived exosomes in cancer progression. Mol. Cancer 20, 154. https://doi.org/10. 1186/s12943-021-01463-y.
- Yao, J., Chen, Y., and Lin, Z. (2023). Exosomes: Mediators in microenvironment of colorectal cancer. Int. J. Cancer 153, 904–917. https://doi.org/10.1002/ijc.34471.
- Almohammai, A., Rahbarghazi, R., Keyhanmanesh, R., Rezaie, J., and Ahmadi, M. (2021). Asthmatic condition induced the activity of exosome secretory pathway in rat pulmonary tissues. J. Inflamm. 18, 14. https://doi.org/10.1186/s12950-021-00275-7.
- Shaban, S.A., Rezaie, J., and Nejati, V. (2022). Exosomes Derived from Senescent Endothelial Cells Contain Distinct Proangiogenic miRNAs and Proteins. Cardiovasc. Toxicol. 22, 592–601. https:// doi.org/10.1007/s12012-022-09740-y.
- Mahbubfam, S., Rezaie, J., and Nejati, V. (2022). Crosstalk between exosomes signaling pathway and autophagy flux in senescent human endothelial cells. Tissue Cell 76, 101803. https://doi.org/10.1016/j. tice.2022.101803.
- Théry, C., Witwer, K.W., Aikawa, E., Alcaraz, M.J., Anderson, J.D., Andriantsitohaina, R., Antoniou, A., Arab, T., Archer, F., Atkin-Smith, G.K., et al. (2018). Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. J. Extracell. Vesicles 7, 1535750. https://doi.org/10.1080/ 20013078.2018.1535750.
- Kalluri, R., and LeBleu, V.S. (2020). The biology, function, and biomedical applications of exosomes. Science 367. eaau6977-+. https://doi.org/10.1126/ science.aau6977.
- Li, S., Dong, R., Kang, Z., Li, H., Wu, X., and Li, T. (2023). Exosomes: Another intercellular lipometabolic communication mediators in digestive system neoplasms? Cytokine Growth Factor Rev. 73, 93–100. https://doi. org/10.1016/j.cytogfr.2023.06.005.
- Lozano-Ramos, I., Bancu, I., Oliveira-Tercero, A., Armengol, M.P., Menezes-Neto, A., Del Portillo, H.A., Lauzurica-Valdemoros, R., and Borràs, F.E. (2015). Size-exclusion chromatography-based enrichment of extracellular vesicles from urine samples. J. Extracell. Vesicles 4, 27369. https://doi.org/10.3402/jev.v4.27369.
- Welton, J.L., Loveless, S., Stone, T., von Ruhland, C., Robertson, N.P., and Clayton, A. (2017). Cerebrospinal fluid extracellular vesicle enrichment for protein biomarker discovery in neurological disease; multiple sclerosis. J. Extracell. Vesicles 6, 1369805. https://doi.org/10.1080/20013078.2017. 1369805.

- 27. Yang, E., Wang, X., Gong, Z., Yu, M., Wu, H., and Zhang, D. (2020). Exosome-mediated metabolic reprogramming: the emerging role in tumor microenvironment remodeling and its influence on cancer progression. Signal Transduct. Target. Ther. 5, 242–2139.
- Hoshino, A., Costa-Silva, B., Shen, T.L., Rodrigues, G., Hashimoto, A., Tesic Mark, M., Molina, H., Kohsaka, S., Di Giannatale, A., Ceder, S., et al. (2015). Tumour exosome integrins determine organotropic metastasis. Nature 527, 329–335. https:// doi.org/10.1038/nature15756.
- van Niel, G., D'Angelo, G., and Raposo, G. (2018). Shedding light on the cell biology of extracellular vesicles. Nat. Rev. Mol. Cell Biol. 19, 213–228. https://doi.org/10.1038/ nrm.2017.125.
- Lafitte, M., Lecointre, C., and Roche, S. (2019). Roles of exosomes in metastatic colorectal cancer. Am. J. Physiol. Cell Physiol. 317, C869–C880. https://doi.org/10. 1152/ajpcell.00218.2019.
- Zhou, L., Li, J., Tang, Y., and Yang, M. (2021). Exosomal LncRNA LINC00659 transferred from cancer-associated fibroblasts promotes colorectal cancer cell progression via miR-342-3p/ANXA2 axis. J. Transl. Med. 19, 8. https://doi.org/10.1186/s12967-020-02648-7.
- Lan, J., Sun, L., Xu, F., Liu, L., Hu, F., Song, D., Hou, Z., Wu, W., Luo, X., Wang, J., et al. (2019). M2 Macrophage-Derived Exosomes Promote Cell Migration and Invasion in Colon Cancer. Cancer Res. 79, 146–158. https://doi.org/10.1158/0008-5472.Can-18-0014.
- Titu, S., Gata, V.A., Decea, R.M., Mocan, T., Dina, C., Irimie, A., and Lisencu, C.I. (2023). Exosomes in Colorectal Cancer: From Physiology to Clinical Applications. Int. J. Mol. Sci. 24, 4382. https://doi.org/10.3390/ ijms24054382.
- Ahmadi, M., Jafari, R., Mahmoodi, M., and Rezaie, J. (2021). The tumorigenic and therapeutic functions of exosomes in colorectal cancer: Opportunity and challenges. Cell Biochem. Funct. 39, 468–477. https://doi.org/10.1002/cbf.3622.
- Rezaie, J., Nejati, V., Mahmoodi, M., and Ahmadi, M. (2022). Mesenchymal stem cells derived extracellular vesicles: A promising nanomedicine for drug delivery system. Biochem. Pharmacol. 203, 115167. https:// doi.org/10.1016/j.bcp.2022.115167.
- Xu, M., and Li, S. (2023). Nano-drug delivery system targeting tumor microenvironment: A prospective strategy for melanoma treatment. Cancer Lett. 574, 216397. https:// doi.org/10.1016/j.canlet.2023.216397.
- Guo, W., Cai, Y., Liu, X., Ji, Y., Zhang, C., Wang, L., Liao, W., Liu, Y., Cui, N., Xiang, J., et al. (2023). Single-Exosome Profiling Identifies ITGB3+and ITGAM plus Exosome Subpopulations as Promising Early Diagnostic Biomarkers and Therapeutic Targets for Colorectal Cancer. Research 6, 0041. https://doi.org/10.34133/ research.0041.
- Goldie, B.J., Dun, M.D., Lin, M., Smith, N.D., Verrills, N.M., Dayas, C.V., and Cairns, M.J. (2014). Activity-associated miRNA are packaged in Map1b-enriched exosomes released from depolarized neurons. Nucleic Acids Res. 42, 9195–9208. https://doi.org/ 10.1093/nar/gku594.
- Bai, S., Wang, Z., Wang, M., Li, J., Wei, Y., Xu, R., and Du, J. (2022). Tumor-Derived Exosomes Modulate Primary Site Tumor

Metastasis. Front. Cell Dev. Biol. 10, 752818. https://doi.org/10.3389/fcell.2022.752818.

- Huang, Y., Kanada, M., Ye, J., Deng, Y., He, Q., Lei, Z., Chen, Y., Li, Y., Qin, P., Zhang, J., and Wei, J. (2022). Exosome-mediated remodeling of the tumor microenvironment: From local to distant intercellular communication. Cancer Lett. 543, 215796. https://doi.org/10.1016/j.canlet.2022. 215796.
- Kimura, M., Kothari, S., Gohir, W., Camargo, J.F., and Husain, S. (2023). MicroRNAs in infectious diseases: potential diagnostic biomarkers and therapeutic targets. Clin. Microbiol. Rev. 36, e0001523. https://doi. org/10.1128/cmr.00015-23.
- Nail, H.M., Chiu, C.C., Leung, C.H., Ahmed, M.M.M., and Wang, H.M.D. (2023). Exosomal miRNA-mediated intercellular communications and immunomodulatory effects in tumor microenvironments. J. Biomed. Sci. 30, 69. https://doi.org/10. 1186/s12929-023-00964-w.
- Liu, H., Liu, Y., Sun, P., Leng, K., Xu, Y., Mei, L., Han, P., Zhang, B., Yao, K., Li, C., et al. (2020). Colorectal cancer-derived exosomal miR-106b-3p promotes metastasis by downregulating DLC-1 expression. Clin. Sci. 134, 419–434. https://doi.org/10.1042/ cs20191087.
- Bigagli, E., Luceri, C., Guasti, D., and Cinci, L. (2016). Exosomes secreted from human colon cancer cells influence the adhesion of neighboring metastatic cells: Role of microRNA-210. Cancer Biol. Ther. 17, 1062– 1069. https://doi.org/10.1080/15384047. 2016.1219815.
- Cho, W.C., Kim, M., Park, J.W., Jeong, S.Y., and Ku, J.L. (2021). Exosomal miR-193a and let-7g accelerate cancer progression on primary colorectal cancer and paired peritoneal metastatic cancer. Transl. Oncol. 14, 101000. https://doi.org/10.1016/j. tranon.2020.101000.
- 46. Yan, S., Ren, X., Yang, J., Wang, J., Zhang, Q., and Xu, D. (2020). Exosomal miR-548c-5p Regulates Colorectal Cancer Cell Growth and Invasion Through HIF1A/CDC42 Axis. OncoTargets Ther. 13, 9875–9885. https:// doi.org/10.2147/ott.S273008.
- Rezaei, R., Baghaei, K., Amani, D., Piccin, A., Hashemi, S.M., Asadzadeh Aghdaei, H., and Zali, M.R. (2021). Exosome-mediated delivery of functionally active miRNA-375-3p mimic regulate epithelial mesenchymal transition (EMT) of colon cancer cells. Life Sci. 269, 119035. https://doi.org/10.1016/j. lfs.2021.119035 (2021.
- Yi, Q., Yue, J., Liu, Y., Shi, H., Sun, W., Feng, J., and Sun, W. (2023). Recent advances of exosomal circRNAs in cancer and their potential clinical applications. J. Transl. Med. 21, 516. https://doi.org/10.1186/ s12967-023-04348-4 (2023.
- 49. Jiang, Z., Hou, Z., Li, L., Liu, W., Yu, Z., and Chen, S. (2021). Exosomal circEPB41L2 serves as a sponge for miR-21-5p and miR-942-5p to suppress colorectal cancer progression by regulating the PTEN/AKT signalling pathway. Eur. J. Clin. Invest. 51, e13581. https://doi.org/10.1111/eci.13581.
- Guo, X., Gao, C., Yang, D.H., and Li, S. (2023). Exosomal circular RNAs: A chief culprit in cancer chemotherapy resistance. Drug Resist. Updat. 67, 100937. https://doi. org/10.1016/i.drup.2023.100937.
- org/10.1016/j.drup.2023.100937. 51. Abdullah, S.T., Abdullah, S.R., Hussen, B.M., Younis, Y.M., Rasul, M.F., and Taheri, M. (2024). Role of circular RNAs and gut





microbiome in gastrointestinal cancers and therapeutic targets. Noncoding. RNA Res. 9, 236–252. https://doi.org/10.1016/j.ncrna. 2023.12.002.

- Yu, Q., Zhang, Y., Tian, Y., Peng, A., Cui, X., Ding, B., Yang, L., Liu, Y., Ju, Y., and Gao, C. (2023). Exosomal Circ\_FMN2 Derived from the Serum of Colorectal Cancer Patients Promotes Cancer Progression by miR-338-3p/MSI1 Axis. Appl. Biochem. Biotechnol. 195, 7322–7337. https://doi.org/10.1007/ s12010-023-04456-3.
- Zhao, H., Chen, S., and Fu, Q. (2020). Exosomes from CD133(+) cells carrying circ-ABCC1 mediate cell stemness and metastasis in colorectal cancer. J. Cell. Biochem. 121, 3286–3297. https://doi.org/ 10.1002/jcb.29600.
- Gao, L., Tang, X., He, Q., Sun, G., Wang, C., and Qu, H. (2021). Exosome-transmitted circCOG2 promotes colorectal cancer progression via miR-1305/TGF-beta 2/SMAD3 pathway. Cell Death Discov. 7, 281. https://doi.org/10.1038/s41420-021-00680-0.
- 55. Chen, C., Yu, H., Han, F., Lai, X., Ye, K., Lei, S., Mai, M., Lai, M., and Zhang, H. (2022). Tumor-suppressive circRHOBTB3 is excreted out of cells via exosome to sustain colorectal cancer cell fitness. Mol. Cancer 21, 46. https://doi.org/10.1186/s12943-022-01511-1.
- Li, Y., Hu, J., Wang, M., Yuan, Y., Zhou, F., Zhao, H., Qiu, T., and Liang, L. (2022). Exosomal circPABPC1 promotes colorectal cancer liver metastases by regulating HMGA2 in the nucleus and BMP4/ADAM19 in the cytoplasm. Cell Death Discov. 8, 335. https://doi.org/10.1038/s41420-022-01124-z.
- Xian, D., Niu, L., Zeng, J., and Wang, L. (2021). LncRNA KCNQ10T1 Secreted by Tumor Cell-Derived Exosomes Mediates Immune Escape in Colorectal Cancer by Regulating PD-L1 Ubiquitination via MiR-30a-5p/USP22. Front. Cell Dev. Biol. 9, 653808. https://doi.org/10.3389/fcell.2021. 653808.
- Fang, X., Xu, Y., Li, K., Liu, P., Zhang, H., Jiang, Y., Tang, J., and Li, Y. (2022). Exosomal IncRNA PCAT1 Promotes Tumor Circulating CellMediated Colorectal Cancer Liver Metastasis by Regulating the Activity of the miR-329-3p/Netrin-1-CD146 Complex. J. Immunol. Res. 2022, 9916228. https://doi.org/10.1155/2022/9916228.
- 59. Luan, Y., Li, X., Luan, Y., Zhao, R., Li, Y., Liu, L., Hao, Y., Oleg Vladimir, B., and Jia, L. (2020). Circulating IncRNA UCA1 Promotes Malignancy of Colorectal Cancer via the miR-143/MY06 Axis. Mol. Ther. Nucleic Acids 19, 790–803. https://doi.org/10.1016/ j.omtn.2019.12.009.
- 60. Xu, J., Xiao, Y., Liu, B., Pan, S., Liu, Q., Shan, Y., Li, S., Qi, Y., Huang, Y., and Jia, L. (2020). Exosomal MALAT1 sponges miR-26a/26b to promote the invasion and metastasis of colorectal cancer via FUT4 enhanced fucosylation and PI3K/Akt pathway. J. Exp. Clin. Cancer Res. 39, 54. https://doi.org/10. 1186/s13046-020-01562-6.
- Sun, J., Lu, Z., Fu, W., Lu, K., Gu, X., Xu, F., Dai, J., Yang, Y., and Jiang, J. (2021). Exosome-Derived ADAM17 Promotes Liver Metastasis in Colorectal Cancer. Front. Pharmacol. 12, 734351. https://doi.org/10. 3389/tphar.2021.734351.
- 62. Wu, B., Sun, D., Ma, L., Deng, Y., Zhang, S., Dong, L., and Chen, S. (2019). Exosomes

isolated from CAPS1-overexpressing colorectal cancer cells promote cell migration. Oncol. Rep. 42, 2528–2536. https://doi.org/10.3892/or.2019.7361.

- Yang, Y., Li, J., and Geng, Y. (2020). Exosomes derived from chronic lymphocytic leukaemia cells transfer miR-146a to induce the transition of mesenchymal stromal cells into cancer-associated fibroblasts. J. Biochem. 168, 491–498. https://doi.org/ 10.1093/jb/mva064.
- 64. Cho, J.A., Park, H., Lim, E.H., and Lee, K.W. (2012). Exosomes from breast cancer cells can convert adipose tissue-derived mesenchymal stem cells into myofibroblastlike cells. Int. J. Oncol. 40, 130–138. https:// doi.org/10.3892/ijo.2011.1193.
- 65. Cho, J.A., Park, H., Lim, E.H., Kim, K.H., Choi, J.S., Lee, J.H., Shin, J.W., and Lee, K.W. (2011). Exosomes from ovarian cancer cells induce adipose tissue-derived mesenchymal stem cells to acquire the physical and functional characteristics of tumor-supporting myofibroblasts. Gynecol. Oncol. 123, 379–386. https://doi.org/10. 1016/j.ygyno.2011.08.005.
- 66. Wang, M.Y., Su, Z.L., and Barnie, P.A. (2020). Crosstalk among colon cancer-derived exosomes, fibroblast-derived exosomes, and macrophage phenotypes in colon cancer metastasis. Int. Immunopharm. 81. https://doi.org/10.1016/j.intimp.2020. 106298.
- 67. Rai, A., Greening, D.W., Chen, M., Xu, R., Ji, H., and Simpson, R.J. (2019). Exosomes Derived from Human Primary and Metastatic Colorectal Cancer Cells Contribute to Functional Heterogeneity of Activated Fibroblasts by Reprogramming Their Proteome. Proteomics 19, e1800148. https://doi.org/10.1002/pmic.201800148.
- 68. Zhou, M., Wang, S., Liu, D., and Zhou, J. (2021). LINC01915 Facilitates the Conversion of Normal Fibroblasts into Cancer-Associated Fibroblasts Induced by Colorectal Cancer-Derived Extracellular Vesicles through the miR-92a-3p/KLF4/ CH25H Axis. ACS Biomater. Sci. Eng. 7, 5255–5268. https://doi.org/10.1021/ acsbiomaterials.1c00611.
- 69. Bhome, R., Emaduddin, M., James, V., House, L.M., Thirdborough, S.M., Mellone, M., Tulkens, J., Primrose, J.N., Thomas, G.J., De Wever, O., et al. (2022). Epithelial to mesenchymal transition influences fibroblast phenotype in colorectal cancer by altering miR-200 levels in extracellular vesicles. J. Extracell. Vesicles 11, e12226. https://doi.org/10.1002/jev2.12226.
- Wang, D., Wang, X., Song, Y., Si, M., Sun, Y., Liu, X., Cui, S., Qu, X., and Yu, X. (2022). Exosomal miR-146a-5p and miR-155-5p promote CXCL12/CXCR7-induced metastasis of colorectal cancer by crosstalk with cancer-associated fibroblasts. Cell Death Dis. 13, 380. https://doi.org/10.1038/ s41419-022-04825-6.
- Zeng, Z., Li, Y., Pan, Y., Lan, X., Song, F., Sun, J., Zhou, K., Liu, X., Ren, X., Wang, F., et al. (2018). Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. Nat. Commun. 9, 5395. https://doi.org/10.1038/s41467-018-07810-w.
- Liu, K., Dou, R., Yang, C., Di, Z., Shi, D., Zhang, C., Song, J., Fang, Y., Huang, S., Xiang, Z., et al. (2023). Exosome-transmitted miR-29a induces colorectal cancer

metastasis by destroying the vascular endothelial barrier. Carcinogenesis 44, 356–367. https://doi.org/10.1093/carcin/ bgad013.

- Duan, S.L., Fu, W.J., Jiang, Y.K., Peng, L.S., Ousmane, D., Zhang, Z.J., and Wang, J.P. (2023). Emerging role of exosome-derived non-coding RNAs in tumor-associated angiogenesis of tumor microenvironment. Front. Mol. Biosci. 10, 1220193. https://doi. org/10.3389/fmolb.2023.1220193.
- Hu, H.Y., Yu, C.H., Zhang, H.H., Zhang, S.Z., Yu, W.Y., Yang, Y., and Chen, Q. (2019). Exosomal miR-1229 derived from colorectal cancer cells promotes angiogenesis by targeting HIPK2. Int. J. Biol. Macromol. 132, 470–477. https://doi.org/10.1016/j. ijbiomac.2019.03.221.
- Hong, B.S., Cho, J.H., Kim, H., Choi, E.J., Rho, S., Kim, J., Kim, J.H., Choi, D.S., Kim, Y.K., Hwang, D., and Gho, Y.S. (2009). Colorectal cancer cell-derived microvesicles are enriched in cell cycle-related mRNAs that promote proliferation of endothelial cells. BMC Genom. 10, 556. https://doi.org/ 10.1186/1471-2164-10-556.
- Chen, C., Liu, Y., Liu, L., Si, C., Xu, Y., Wu, X., Wang, C., Sun, Z., and Kang, Q. (2023). Exosomal circTUBGCP4 promotes vascular endothelial cell tipping and colorectal cancer metastasis by activating Akt signaling pathway. J. Exp. Clin. Cancer Res. 42, 46. https://doi.org/10.1186/s13046-023-02619-y.
- Zheng, X., Ma, N., Wang, X., Hu, J., Ma, X., Wang, J., and Cao, B. (2020). Exosomes derived from 5-fluorouracil-resistant colon cancer cells are enriched in GDF15 and can promote angiogenesis. J. Cancer 11, 7116– 7126. https://doi.org/10.7150/jca.49224.
- Yoon, Y.J., Kim, D.K., Yoon, C.M., Park, J., Kim, Y.K., Roh, T.Y., and Gho, Y.S. (2014). Egr-1 Activation by Cancer-Derived Extracellular Vesicles Promotes Endothelial Cell Migration via ERK1/2 and JNK Signaling Pathways. PLoS One 9, e115170. https://doi.org/10.1371/journal.pone. 0115170.
- 79. Wu, R., Zhang, Y., Xu, X., You, O., Yu, C., Wang, W., and Mao, Y. (2023). Exosomal B7-H3 facilitates colorectal cancer angiogenesis and metastasis through AKT1/ mTOR/VEGFA pathway. Cell. Signal. 109, 110737. https://doi.org/10.1016/j.cellsig. 2023.110737.
- Ma, J., Wang, P., Liu, Y., Zhao, L., Li, Z., and Xue, Y. (2014). Krüppel-like factor 4 regulates blood-tumor barrier permeability via ZO-1, occludin and claudin-5. J. Cell. Physiol. 229, 916–926. https://doi.org/10. 1002/jcp.24523.
- Bakshi, H.A., Mkhael, M., Faruck, H.L., Khan, A.U., Aljabali, A.A.A., Mishra, V., El-Tanani, M., Charbe, N.B., and Tambuwala, M.M. (2024). Cellular signaling in the hypoxic cancer microenvironment: Implications for drug resistance and therapeutic targeting. Cell. Signal. *113*, 110911. https://doi.org/10. 1016/j.cellsig.2023.110911.
- Huang, Z., and Feng, Y. (2017). Exosomes Derived From Hypoxic Colorectal Cancer Cells Promote Angiogenesis Through Wnt4-Induced beta-Catenin Signaling in Endothelial Cells. Oncol. Res. 25, 651–661. https://doi.org/10.3727/ 096504016x14752792816791.
- Yun, C.W., Lee, J.H., Go, G., Jeon, J., Yoon, S., and Lee, S.H. (2021). Prion Protein of Extracellular Vesicle Regulates the



Progression of Colorectal Cancer. Cancers 13, 2144. https://doi.org/10.3390/ cancers13092144.

- Mills, C.D., Kincaid, K., Alt, J.M., Heilman, M.J., and Hill, A.M. (2000). M-1/M-2 macrophages and the Th1/Th2 paradigm. J. Immunol. 164, 6166–6173. https://doi. org/10.4049/jimmunol.164.12.6166.
- Rastin, F., Javid, H., Oryani, M.A., Rezagholinejad, N., Afshari, A.R., and Karimi-Shahri, M. (2024). Immunotherapy for colorectal cancer: Rational strategies and novel therapeutic progress. Int. Immunopharmacol. *126*, 111055. https:// doi.org/10.1016/j.intimp.2023.111055.
- Bied, M., Ho, W.W., Ginhoux, F., and Blériot, C. (2023). Roles of macrophages in tumor development: a spatiotemporal perspective. Cell. Mol. Immunol. 20, 983–992. https://doi.org/10.1038/s41423-023-01061-6.
- Cooks, T., Pateras, I.S., Jenkins, L.M., Patel, K.M., Robles, A.I., Morris, J., Forshew, T., Appella, E., Gorgoulis, V.G., and Harris, C.C. (2018). Mutant p53 cancers reprogram macrophages to tumor supporting macrophages to a exosomal miR-1246. Nat. Commun. 9, 771. https://doi.org/10.1038/ s41467-018-03224-w.
- Shang, Z., Ma, Z., Wu, E., Chen, X., Tuo, B., Li, T., and Liu, X. (2024). Effect of metabolic reprogramming on the immune microenvironment in gastric cancer. Biomed. Pharm. 170, 116030. https://doi. org/10.1016/j.biopha.2023.116030.
- Chuang, Y.M., Tzeng, S.F., Ho, P.C., and Tsai, C.H. (2024). Immunosurveillance encounters cancer metabolism. EMBO Rep. 25, 471–488. https://doi.org/10.1038/ s44319-023-00038-w.
- Shinohara, H., Kuranaga, Y., Kumazaki, M., Sugito, N., Yoshikawa, Y., Takai, T., Taniguchi, K., Ito, Y., and Akao, Y. (2017). Regulated Polarization of Tumor-Associated Macrophages by miR-145 via Colorectal Cancer-Derived Extracellular Vesicles. J. Immunol. 199, 1505–1515. https://doi.org/10.4049/jimmunol.1700167.
- Zhao, S., Mi, Y., Guan, B., Zheng, B., Wei, P., Gu, Y., Zhang, Z., Cai, S., Xu, Y., Li, X., et al. (2020). Tumor-derived exosomal miR-934 induces macrophage M2 polarization to promote liver metastasis of colorectal cancer. J. Hematol. Oncol. 13, 156. https:// doi.org/10.1186/s13045-020-00991-2.
- Baig, M.S., Roy, A., Rajpoot, S., Liu, D., Savai, R., Banerjee, S., Kawada, M., Faisal, S.M., Saluja, R., Saqib, U., et al. (2020). Tumorderived exosomes in the regulation of macrophage polarization. Inflamm. Res. 69, 435–451. https://doi.org/10.1007/s00011-020-01318-0.
- Chen, Z., Yang, L., Cui, Y., Zhou, Y., Yin, X., Guo, J., Zhang, G., Wang, T., and He, Q.Y. (2016). Cytoskeleton-centric protein transportation by exosomes transforms tumor-favorable macrophages. Oncotarget 7, 67387–67402. https://doi.org/10.18632/ oncotarget.11794.
- 94. Takano, Y., Masuda, T., Iinuma, H., Yamaguchi, R., Sato, K., Tobo, T., Hirata, H., Kuroda, Y., Nambara, S., Hayashi, N., et al. (2017). Circulating exosomal microRNA-203 is associated with metastasis possibly via inducing tumor-associated macrophages in colorectal cancer. Oncotarget *8*, 78598– 78613. https://doi.org/10.18632/ oncotarget.20009.

- Pei, W., Wei, K., Wu, Y., Qiu, Q., Zhu, H., Mao, L., Shi, X., Zhang, S., Shi, Y., Tao, S., et al. (2023). Colorectal cancer tumor cellderived exosomal miR-203a-3p promotes CRC metastasis by targeting PTEN-induced macrophage polarization. Gene 885, 147692. https://doi.org/10.1016/j.gene. 2023.147692.
- 96. Sun, W., Cui, J., Ge, Y., Wang, J., Yu, Y., Han, B., and Liu, B. (2022). Tumor stem cellderived exosomal microRNA-17-5p inhibits anti-tumor immunity in colorectal cancer via targeting SPOP and overexpressing PD-L1. Cell Death Discov. 8, 223. https://doi.org/ 10.1038/s41420-022-00919-4.
- 97. Yang, C., Dou, R., Wei, C., Liu, K., Shi, D., Zhang, C., Liu, Q., Wang, S., and Xiong, B. (2021). Tumor-derived exosomal microRNA-106b-5p activates EMT-cancer cell and M2subtype TAM interaction to facilitate CRC metastasis. Mol. Ther. 29, 2088–2107. https://doi.org/10.1016/j.ymthe.2021. 02.006.
- Sun, H., Meng, Q., Shi, C., Yang, H., Li, X., Wu, S., Familiari, G., Relucenti, M., Aschner, M., Wang, X., and Chen, R. (2021). Hypoxia-Inducible Exosomes Facilitate Liver-Tropic Premetastatic Niche in Colorectal Cancer. Hepatology 74, 2633–2651. https://doi.org/ 10.1002/hep.32009.
- Hánělová, K., Raudenská, M., Masařík, M., and Balvan, J. (2024). Protein cargo in extracellular vesicles as the key mediator in the progression of cancer. Cell Commun. Signal. 22, 25. https://doi.org/10.1186/ s12964-023-01408-6.
- 100. Liang, Z.X., Liu, H.S., Wang, F.W., Xiong, L., Zhou, C., Hu, T., He, X.W., Wu, X.J., Xie, D., Wu, X.R., and Lan, P. (2019). LncRNA RPPH1 promotes colorectal cancer metastasis by interacting with TUBB3 and by promoting exosomes-mediated macrophage M2 polarization. Cell Death Dis. *10*, 829. https:// doi.org/10.1038/s41419-019-2077-0.
- 101. Li, X., Lan, Q., Lai, W., Wu, H., Xu, H., Fang, K., Chu, Z., and Zeng, Y. (2022). Exosomederived Inc-HOXB8-1:2 induces tumorassociated macrophage infiltration to promote neuroendocrine differentiated colorectal cancer progression by sponging hsa-miR-6825-5p. BMC Cancer 22, 928. https://doi.org/10.1186/s12885-022-09926-1.
- 102. Ding, A.X., Wang, H., Zhang, J.M., Yang, W., and Kuang, Y.T. (2024). IncRNA BANCR promotes the colorectal cancer metastasis through accelerating exosomes-mediated M2 macrophage polarization via regulating RhoA/ROCK signaling. Mol. Cell. Biochem. 479, 13–27. https://doi.org/10.1007/s11010-023-04709-z.
- 103. Tang, Y., Hu, S., Li, T., and Qiu, X. (2023). Tumor cells-derived exosomal circVCP promoted the progression of colorectal cancer by regulating macrophage M1/M2 polarization. Gene 870, 147413. https://doi. org/10.1016/j.gene.2023.147413.
- 104. Huber, V., Fais, S., Iero, M., Lugini, L., Canese, P., Squarcina, P., Zaccheddu, A., Colone, M., Arancia, G., Gentile, M., et al. (2005). Human colorectal cancer cells induce T-cell death through release of proapoptotic microvesicles: Role in immune escape. Gastroenterology 128, 1796–1804. https://doi.org/10.1053/j.gastro.2005. 03.045.
- Kaplan, R.N., Riba, R.D., Zacharoulis, S., Bramley, A.H., Vincent, L., Costa, C., MacDonald, D.D., Jin, D.K., Shido, K., Kerns,

S.A., et al. (2005). VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 438, 820-827. https://doi.org/10.1038/ nature04186.

- Luo, X., Li, Y., Hua, Z., Xue, X., Wang, X., Pang, M., Xiao, C., Zhao, H., Lyu, A., and Liu, Y. (2023). Exosomes-mediated tumor metastasis through reshaping tumor microenvironment and distant niche. J. Control. Release 353, 327–336. https:// doi.org/10.1016/j.jconrel.2022.11.050.
- 107. Shao, Y., Chen, T., Zheng, X., Yang, S., Xu, K., Chen, X., Xu, F., Wang, L., Shen, Y., Wang, T., et al. (2018). Colorectal cancer-derived small extracellular vesicles establish an inflammatory premetastatic niche in liver metastasis. Carcinogenesis 39, 1368–1379. https://doi.org/10.1093/carcin/bgy115.
- 108. Tian, F., Wang, P., Lin, D., Dai, J., Liu, Q., Guan, Y., Zhan, Y., Yang, Y., Wang, W., Wang, J., et al. (2021). Exosome-delivered miR-221/222 exacerbates tumor liver metastasis by targeting SPINT1 in colorectal cancer. Cancer Sci. 112, 3744–3755. https:// doi.org/10.1111/cas.15028.
- 109. Wang, J., Liu, Y., Li, Y., Zheng, X., Gan, J., Wan, Z., Zhang, J., Liu, Y., Wang, Y., Hu, W., et al. (2021). Exosomal-miR-10a derived from colorectal cancer cells suppresses migration of human lung fibroblasts, and expression of IL-6, IL-8 and IL-1 beta. Mol. Med. Rep. 23, 84. https://doi.org/10.3892/ mmr.2020.11723.
- Lee, S., Park, Y.S., Kim, J.H., Lim, A.R., Hyun, M.H., Kim, B., Lee, J.W., Lee, S.B., and Kim, Y.H. (2022). Identification of Biomarkers Associated with Liver Metastasis Progression from Colorectal Cancer Using Exosomal RNA Profiling. Cancers 14, 4723. https://doi.org/10.3390/cancers14194723.
- 111. Jiang, K., Chen, H., Fang, Y., Chen, L., Zhong, C., Bu, T., Dai, S., Pan, X., Fu, D., Qian, Y., et al. (2021). Exosomal ANGPTL1 attenuates colorectal cancer liver metastasis by regulating Kupffer cell secretion pattern and impeding MMP9 induced vascular leakiness. J. Exp. Clin. Cancer Res. 40, 21. https://doi.org/10.1186/s13046-020-01816-3.
- 112. Zhang, C., Wang, X.Y., Zhang, P., He, T.C., Han, J.H., Zhang, R., Lin, J., Fan, J., Lu, L., Zhu, W.W., et al. (2022). Cancer-derived exosomal HSPC111 promotes colorectal cancer liver metastasis by reprogramming lipid metabolism in cancer-associated fibroblasts. Cell Death Dis. 13, 57. https:// doi.org/10.1038/s41419-022-04506-4.
- 113. Kanzaki, R., and Pietras, K. (2020). Heterogeneity of cancer-associated fibroblasts: Opportunities for precision medicine. Cancer Sci. 111, 2708–2717. https://doi.org/10.1111/cas.14537.
- 114. Sahai, E., Astsaturov, I., Cukierman, E., DeNardo, D.G., Egeblad, M., Evans, R.M., Fearon, D., Greten, F.R., Hingorani, S.R., Hunter, T., et al. (2020). A framework for advancing our understanding of cancerassociated fibroblasts. Nat. Rev. Cancer 20, 174–186. https://doi.org/10.1038/s41568-019-0238-1.
- 115. Shi, W., Liu, Y., Qiu, X., Yang, L., and Lin, G. (2023). Cancer-associated fibroblastsderived exosome-mediated transfer of miR-345-5p promotes the progression of colorectal cancer by targeting CDKN1A. Carcinogenesis 44, 317–327. https://doi. org/10.1093/carcin/bgad014.





- 116. Zhang, Y., Wang, S., Lai, Q., Fang, Y., Wu, C., Liu, Y., Li, Q., Wang, X., Gu, C., Chen, J., et al. (2020). Cancer-associated fibroblastsderived exosomal miR-17-5p promotes colorectal cancer aggressive phenotype by initiating a RUNX3/MYC/TGF-beta 1 positive feedback loop. Cancer Lett. 491, 22–35. https://doi.org/10.1016/j.canlet. 2020.07.023.
- 117. Hu, J.L., Wang, W., Lan, X.L., Zeng, Z.C., Liang, Y.S., Yan, Y.R., Song, F.Y., Wang, F.F., Zhu, X.H., Liao, W.J., et al. (2019). CAFs secreted exosomes promote metastasis and chemotherapy resistance by enhancing cell stemness and epithelial-mesenchymal transition in colorectal cancer. Mol. Cancer 18, 91. https://doi.org/10.1186/s12943-019-1019-x.
- 118. Jiang, Y., Qiu, Q., Jing, X., Song, Z., Zhang, Y., Wang, C., Liu, K., Ye, F., Ji, X., Luo, F., and Zhao, R. (2023). Cancer-associated fibroblast-derived exosome miR-181b-3p promotes the occurrence and development of colorectal cancer by regulating SNX2 expression. Biochem. Biophys. Res. Commun. 641, 177–185. https://doi.org/10. 1016/j.bbrc.2022.12.026.
- 1010/JOIC2022. (2020).
  119. Zhang, Y., Yin, C., Wei, C., Xia, S., Qiao, Z., Zhang, X.W., Yu, B., Zhou, J., and Wang, R. (2022). Exosomal miR-625-3p secreted by cancer-associated fibroblasts in colorectal cancer promotes EMT and chemotherapeutic resistance by blocking the CELF2/WWOX pathway. Pharmacol. Res. 186, 106534. https://doi.org/10.1016/j. phrs.2022.106534.
- 120. Gong, W., Guo, Y., Yuan, H., Chai, R., Wan, Z., Zheng, B., Hu, X., Chen, B., Gao, S., Dai, Q., et al. (2023). Loss of exosomal miR-200b-3p from hypoxia cancer-associated fibroblasts promotes tumorigenesis and reduces sensitivity to 5-Flourouracil in colorectal cancer via upregulation of ZEB1 and E2F3. Cancer Gene Ther. 30, 905–916. https://doi.org/10.1038/s41417-023-00591-5
- 121. Yang, K., Zhang, F., Luo, B., and Qu, Z. (2022). CAFs-derived small extracellular vesicles circN4BP2L2 promotes proliferation and metastasis of colorectal cancer via miR-664b-3p/HMGB3 pathway. Cancer Biol. Ther. 23, 404–416. https://doi.org/10.1080/ 15384047.2022.2072164.
- 122. Yang, K., Zhang, J., and Bao, C. (2021). Exosomal circEIF3K from cancer-associated fibroblast promotes colorectal cancer (CRC) progression via miR-214/PD-L1 axis. BMC Cancer 21, 933. https://doi.org/10.1186/ s12885-021-08669-9.
- 123. Deng, X., Ruan, H., Zhang, X., Xu, X., Zhu, Y., Peng, H., Zhang, X., Kong, F., and Guan, M. (2020). Long noncoding RNA CCAL transferred from fibroblasts by exosomes promotes chemoresistance of colorectal cancer cells. Int. J. Cancer 146, 1700–1716. https://doi.org/10.1002/ijc.32608.
- 124. Ren, J., Ding, L., Zhang, D., Shi, G., Xu, Q., Shen, S., Wang, Y., Wang, T., and Hou, Y. (2018). Carcinoma-associated fibroblasts promote the stemness and chemoresistance of colorectal cancer by transferring exosomal IncRNA H19. Theranostics *8*, 3932–3948. https://doi.org/ 10.7150/thno.25541.
- 125. Niu, L., Wang, Q., Feng, F., Yang, W., Xie, Z., Zheng, G., Zhou, W., Duan, L., Du, K., Li, Y., et al. (2024). Small extracellular vesiclesmediated cellular interactions between tumor cells and tumor-associated

macrophages: Implication for immunotherapy. Biochim. Biophys. Acta, Mol. Basis Dis. 1870, 166917. https://doi. org/10.1016/j.bbadis.2023.166917.

- Mahmoudi, F., Hanachi, P., and Montaseri, A. (2023). Extracellular vesicles of immune cells; immunomodulatory impacts and therapeutic potentials. Clin. Immunol. 248, 109237. https://doi.org/10.1016/j.clim.2023. 109237.
- 127. Szebeni, G.J., Vizler, C., Kitajka, K., and Puskas, L.G. (2017). Inflammation and Cancer: Extra- and Intracellular Determinants of Tumor-Associated Macrophages as Tumor Promoters. Mediators Inflamm. 2017, 9294018. https:// doi.org/10.1155/2017/9294018.
- 128. Wang, L., Shan, Y., Zheng, S., Li, J., and Cui, P. (2023). miR-4780 Derived from N2-Like Neutrophil Exosome Aggravates Epithelial-Mesenchymal Transition and Angiogenesis in Colorectal Cancer. Stem Cells Int. 2023, 2759679. https://doi.org/10.1155/2023/ 2759679.
- 129. Yuan, Y.G., Wang, J.L., Zhang, Y.X., Li, L., Reza, A.M.M.T., and Gurunathan, S. (2023). Biogenesis, Composition and Potential Therapeutic Applications of Mesenchymal Stem Cells Derived Exosomes in Various Diseases. Int. J. Nanomedicine 18, 3177– 3210. https://doi.org/10.2147/ijn.5407029.
- 3210. https://doi.org/10.2147/ijn.5407029.
  130. Li, T., Wan, Y., Su, Z., Li, J., Han, M., and Zhou, C. (2021). Mesenchymal Stem Cell-Derived Exosomal microRNA-3940-5p Inhibits Colorectal Cancer Metastasis by Targeting Integrin alpha 6. Dig. Dis. Sci. 66, 1916–1927. https://doi.org/10.1007/s10620-020-06458-1.
- 131. Jahangiri, B., Khalaj-Kondori, M., Asadollahi, E., Purrafee Dizaj, L., and Sadeghizadeh, M. (2022). MSC-Derived exosomes suppress colorectal cancer cell proliferation and metastasis via miR-100/ mTOR/miR-143 pathway. Int. J. Pharm. 627, 122214. https://doi.org/10.1016/j.ijpharm. 2022.122214.
- 132. Chen, J., Li, Z., Yue, C., Ma, J., Cao, L., Lin, J., Zhu, D., An, R., Lai, J., Guo, Y., and Gu, B. (2023). Human umbilical cord mesenchymal stem cell-derived exosomes carrying miR-1827 downregulate SUCNR1 to inhibit macrophage M2 polarization and prevent colorectal liver metastasis. Apoptosis 28, 549–565. https://doi.org/10.1007/s10495-022-01798-x (2023.
- 133. Guo, G., Tan, Z., Liu, Y., Shi, F., and She, J. (2022). The therapeutic potential of stem cell-derived exosomes in the ulcerative colitis and colorectal cancer. Stem Cell Res. Ther. 13, 138. https://doi.org/10.1186/ s13287-022-02811-5.
- 134. Xu, H., Lan, Q., Huang, Y., Zhang, Y., Zeng, Y., Su, P., Chu, Z., Lai, W., and Chu, Z. (2021). The mechanisms of colorectal cancer cell mesenchymal-epithelial transition induced by hepatocyte exosome-derived miR-203a-3p. BMC Cancer 21, 718. https://doi.org/10. 1186/s12885-021-08419-x.
- Wang, Z., Kim, S.Y., Tu, W., Kim, J., Xu, A., Yang, Y.M., Matsuda, M., Reolizo, L., Tsuchiya, T., Billet, S., et al. (2023).
   Extracellular vesicles in fatty liver promote a metastatic tumor microenvironment. Cell Metab. 35, 1209–1226.e13. https://doi.org/ 10.1016/j.cmet.2023.04.013.
- Silverman, D.A., Martinez, V.K., Dougherty, P.M., Myers, J.N., Calin, G.A., and Amit, M. (2021). Cancer-Associated Neurogenesis and Nerve-Cancer Cross-talk. Cancer Res.

81, 1431–1440. https://doi.org/10.1158/ 0008-5472.Can-20-2793.

- 137. Zhou, Y., Shurin, G.V., Zhong, H., Bunimovich, Y.L., Han, B., and Shurin, M.R. (2018). Schwann Cells Augment Cell Spreading and Metastasis of Lung Cancer. Cancer Res. 78, 5927–5939. https://doi.org/ 10.1158/0008-5472.Can-18-1702.
- Su, D., Guo, X., Huang, L., Ye, H., Li, Z., Lin, L., Chen, R., and Zhou, Q. (2020). Tumorneuroglia interaction promotes pancreatic cancer metastasis. Theranostics 10, 5029– 5047. https://doi.org/10.7150/thno.42440.
- Han, S., Wang, D., Huang, Y., Zeng, Z., Xu, P., Xiong, H., Ke, Z., Zhang, Y., Hu, Y., Wang, F., et al. (2022). A reciprocal feedback between colon cancer cells and Schwann cells promotes the proliferation and metastasis of colon cancer. J. Exp. Clin. Cancer Res. 41, 348. https://doi.org/10. 1186/s13046-022-02556-2.
- 140. Huang, C., Zhou, Y., Feng, X., Wang, J., Li, Y., and Yao, X. (2023). Delivery of Engineered Primary Tumor-Derived Exosomes Effectively Suppressed the Colorectal Cancer Chemoresistance and Liver Metastasis. ACS Nano 17, 10313– 10326. https://doi.org/10.1021/acsnano. 3c00668.
- 141. Miyazaki, K., Wada, Y., Okuno, K., Murano, T., Morine, Y., Ikemoto, T., Saito, Y., Ikematsu, H., Kinugasa, Y., Shimada, M., and Goel, A. (2023). An exosome-based liquid biopsy signature for pre-operative identification of lymph node metastasis in patients with pathological high-risk T1 colorectal cancer. Mol. Cancer 22, 2. https:// doi.org/10.1186/s12943-022-01685-8 (2023.
- Ning, T., Li, J., He, Y., Zhang, H., Wang, X., Deng, T., Liu, R., Li, H., Bai, M., Fan, Q., et al. (2021). Exosomal miR-208b related with oxaliplatin resistance promotes Treg expansion in colorectal cancer. Mol. Ther. 29, 2723–2736. https://doi.org/10.1016/j. ymthe.2021.04.028.
- 143. Shao, H., Yao, L., Tao, Y., and Huang, X. (2023). Identification and verification of an exosome-related gene risk model to predict prognosis and evaluate immune infiltration for colorectal cancer. Medicine (Baltim.) 102, e35365. https://doi.org/10.1097/md. 0000000000035365.
- Wang, L., Wang, D., Ye, Z., and Xu. (2023).
   J. engineering extracellular vesicles as delivery systems in therapeutic applications. Adv. Sci. 10, e2300552. https://doi.org/10. 1002/advs.202300552.
- 145. Zhao, L., Yu, Q., Gao, C., Xiang, J., Zheng, B., Feng, Y., Li, R., Zhang, W., Hong, X., Zhan, Y.Y., et al. (2022). Studies of the Efficacy of Low-Dose Apatinib Monotherapy as Third-Line Treatment in Patients with Metastatic Colorectal Cancer and Apatinib's Novel Anticancer Effect by Inhibiting Tumor-Derived Exosome Secretion. Cancers 14, 2492. https://doi.org/10.3390/ cancers14102492.
- 146. He, K., Wang, Z., Luo, M., Li, B., Ding, N., Li, L., He, B., Wang, H., Cao, J., Huang, C., et al. (2023). Metastasis organotropism in colorectal cancer: advancing toward innovative therapies. J. Transl. Med. 21, 612. https://doi.org/10.1186/s12967-023-04460-5 (2023.
- 147. Chen, Z., Yue, Z., Yang, K., and Li, S. (2022). Nanomaterials: small particles show huge possibilities for cancer immunotherapy. J. Nanobiotechnol. 20, 484. https://doi.org/ 10.1186/s12951-022-01692-3.

### iScience Review

# iScience

Review

- Kang, Y., and Li, S. (2023). Nanomaterials: Breaking through the bottleneck of tumor immunotherapy. Int. J. Biol. Macromol. 230, 123159. https://doi.org/10.1016/j.ijbiomac. 2023.123159.
- 149. Chen, Z., Yue, Z., Yang, K., Shen, C., Cheng, Z., Zhou, X., and Li, S. (2023). Four Ounces Can Move a Thousand Pounds: The Enormous Value of Nanomaterials in Tumor Immunotherapy. Adv. Healthc. Mater. 12, e2300882. https://doi.org/10.1002/adhm. 202300882.
- 150. Nguyen, V.D., Kim, H.Y., Choi, Y.H., Park, J.O., and Choi, E. (2022). Tumor-derived extracellular vesicles for the active targeting and effective treatment of colorectal tumors in vivo. Drug Deliv. 29, 2621–2631. https:// doi.org/10.1080/10717544.2022.2105444.
- Tran, P.H.L., Wang, T., Yin, W., Tran, T.T.D., Nguyen, T.N.G., Lee, B.J., and Duan, W. (2019). Aspirin-loaded nanoexosomes as cancer therapeutics. Int. J. Pharm. 572, 118786. https://doi.org/10.1016/j.ijpharm. 2019.118786.
- 152. Ning, S., Chen, Y., Li, S., Liu, M., Liu, H., Ye, M., Wang, C., Pan, J., Wei, W., Li, J., and Zhang, L. (2023). Exosomal miR-99b-5p Secreted from Mesenchymal Stem Cells Can Retard the Progression of Colorectal Cancer by Targeting FGFR3. Stem Cell Rev. Rep. 19, 2901–2917. https://doi.org/10.1007/s12015-023-10606-1 (2023.
- 153. Liang, G., Zhu, Y., Ali, D.J., Tian, T., Xu, H., Si, K., Sun, B., Chen, B., and Xiao, Z. (2020). Engineered exosomes for targeted codelivery of miR-21 inhibitor and chemotherapeutics to reverse drug resistance in colon cancer. J. Nanobiotechnology 18, 10. https://doi. org/10.1186/s12951-019-0563-2.

- 154. Kamerkar, S., Leng, C., Burenkova, O., Jang, S.C., McCoy, C., Zhang, K., Dooley, K., Kasera, S., Zi, T., Sisó, S., et al. (2022). Exosome-mediated genetic reprogramming of tumor-associated macrophages by exoASO-STAT6 leads to potent monotherapy antitumor activity. Sci. Adv. 8, eabj7002. https://doi.org/10.1126/ sciadv.abj7002.
- 155. Aarsund, M., Segers, F.M., Wu, Y., and Inngjerdingen, M. (2022). Comparison of characteristics and tumor targeting properties of extracellular vesicles derived from primary NK cells or NK-cell lines stimulated with IL-15 or IL-12/15/18. Cancer Immunol. Immunother. 71, 2227–2238. https://doi.org/10.1007/s00262-022-03161-0.
- Poggio, M., Hu, T., Pai, C.C., Chu, B., Belair, C.D., Chang, A., Montabana, E., Lang, U.E., Fu, Q., Fong, L., and Blelloch, R. (2019). Suppression of Exosomal PD-L1 Induces Systemic Anti-tumor Immunity and Memory. Cell 177, 414-427.e13. https://doi.org/10. 1016/j.cell.2019.02.016.
- 157. Yi, B., Cheng, H., Wyczechowska, D., Yu, Q., Li, L., Ochoa, A.C., Riker, A.I., and Xi, Y. (2021). Sulindac Modulates the Response of Proficient MMR Colorectal Cancer to Anti-PD-L1 Immunotherapy. Mol. Cancer Ther. 20, 1295–1304. https://doi.org/10.1158/ 1535-7163.Mct-20-0934.
- 158. Min, Y., Deng, W., Yuan, H., Zhu, D., Zhao, R., Zhang, P., Xue, J., Yuan, Z., Zhang, T., Jiang, Y., et al. (2024). Single extracellular vesicle surface protein-based blood assay identifies potential biomarkers for detection and screening of five cancers. Mol. Oncol. https://doi.org/10.1002/1878-0261.13586.

- 159. Wu, D., Yan, J., Shen, X., Sun, Y., Thulin, M., Cai, Y., Wik, L., Shen, Q., Oelrich, J., Qian, X., et al. (2019). Profiling surface proteins on individual exosomes using a proximity barcoding assay. Nat. Commun. 10, 3854. https://doi.org/10.1038/s41467-019-11486-1.
- Azubuike, U.F., and Tanner, K. (2023). Biophysical determinants of cancer organotropism. Trends Cancer 9, 188–197. https://doi.org/10.1016/j.trecan.2022.11. 002 (2023.
- 161. Rezaie, J., Etemadi, T., and Feghhi, M. (2022). The distinct roles of exosomes in innate immune responses and therapeutic applications in cancer. Eur. J. Pharmacol. 933, 175292. https://doi.org/10.1016/j. ejphar.2022.175292.
- 162. Samanta, S., Rajasingh, S., Drosos, N., Zhou, Z., Dawn, B., and Rajasingh, J. (2018). Exosomes: new molecular targets of diseases. Acta Pharmacol. Sin. 39, 501–513. https://doi.org/10.1038/aps. 2017.162.
- Zhu, X., and Li, S. (2023). Nanomaterials in tumor immunotherapy: new strategies and challenges. Mol. Cancer 22, 94. https://doi. org/10.1186/s12943-023-01797-9 (2023.
- 164. Choi, S.J., Cho, H., Yea, K., and Baek, M.C. (2022). Immune cell-derived small extracellular vesicles in cancer treatment. BMB Rep. 55, 48–56. https://doi.org/10. 5483/BMBRep.2022.55.1.133.
- 165. Zhou, X., Jia, Y., Mao, C., and Liu, S. (2024). Small extracellular vesicles: Non-negligible vesicles in tumor progression, diagnosis, and therapy. Cancer Lett. 580, 216481. https://doi.org/10.1016/j.canlet.2023. 216481.

