



#### Received: 2021/10/29, Revised: 2021/12/03, Accepted: 2021/12/19, Published: 2021/12/31

©2021 Sara Santarossa et al.; Licence Physical Activity and Nutrition. This is an open access article distributed under the terms of the creative commons attribution license (https:// creativecommons.org/licenses/by-nc/2.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the orginal work is properly cited.

\*Corresponding author : Sara Santarossa, Ph.D.

Department of Public Health Sciences, Henry Ford Health System, 1 Ford Place, Detroit, Michigan 48202, United States of America

Tel: 313-874-7960

E-mail: ssantar1@hfhs.org

©2021 The Korean Society for Exercise Nutrition

[Purpose] To determine whether physical activity (PA), primarily the recommended 60 minutes of moderate-to-vigorous PA, is associated with gut bacterial microbiota in 10-year-old children.

[Methods] The Block Physical Activity Screener, which provides minutes/day PA variables, was used to determine whether the child met the PA recommendations 16S rRNA sequencing was performed on stool samples from the children to profile the composition of their gut bacterial microbiota. Differences in alpha diversity metrics (richness, Pielou's evenness, and Faith's phylogenetic diversity) by PA were determined using linear regression, whereas beta diversity (unweighted and weighted UniFrac) relationships were assessed using PERMANOVA. Taxon relative abundance differentials were determined using DESeq2.

[Results] The analytic sample included 321 children with both PA and 16S rRNA sequencing data (mean age [SD] =10.2 [0.8] years; 54.2% male; 62.9% African American), where 189 (58.9%) met the PA recommendations. After adjusting for covariates, meeting the PA recommendations as well as minutes/ day PA variables were not significantly associated with out richness, evenness, or diversity ( $p \ge 0.19$ ). However, meeting the PA recommendations (weighted UniFrac  $R^2 = 0.014$ , p = 0.001) was significantly associated with distinct gut bacterial composition. These compositional differences were partly characterized by increased abundance of Megamonas and Anaerovorax as well as specific Christensenellaceae\_R-7\_group taxa in children with higher PA

[Conclusion] Children who met the recommendations of PA had altered gut microbiota compositions. Whether this translates to a reduced risk of obesity or associated metabolic diseases is still unclear.

[Keywords] physical activity, gastrointestinal microbiome, 16S rRNA, gut microbiota, children

#### [Abbreviations]

P	A	Physical activity
V	VHEALS	Wayne County Health Environment Asthma and
		Allergy Longitudinal Study
Н	IFHS	Henry Ford Health System
С	TAB	cetyltrimethylammonium bromide
С	DTUs	operational taxonomic units
В	KPAS	Block Physical Activity Screener
Ν	1PA	estimated daily moderate physical activity minutes
V	'PA	estimated daily vigorous physical activity minutes
R	RPA	estimated daily recreational physical activity minutes
Ν	1ET	metabolic equivalent
Ν	(IVPA	estimated daily moderate-to-vigorous physical
		activity minutes
В	MI	body mass index

## Associations of physical activity with gut microbiota in pre-adolescent children

Sara Santarossa<sup>1\*</sup> / Alexandra R. Sitarik<sup>1</sup> / Christine Cole Johnson<sup>1</sup> / Jia Li<sup>1</sup> / Susan V. Lynch<sup>2</sup> / Dennis R. Ownby<sup>3</sup> / Alex Ramirez<sup>1,4</sup> / Germaine LM. Yong<sup>2</sup> / Andrea E. Cassidy-Bushrow<sup>1</sup>

1. Department of Public Health Sciences, Henry Ford Health System, Michigan, USA

2. Department of Medicine, University of California, California, USA

Department of Pediatrics, Georgia Regents University, Georgia, USA
Wayne State University School of Medicine Detroit, Michigan, USA

## INTRODUCTION

Children can achieve substantial health benefits by engaging in moderate-to-vigorous physical activity (PA) for periods of time that add up to 60 minutes or more each day (current recommendations for ages 6-17 years)<sup>1</sup>. It appears that, as in adults, the total amount of PA is more important, for achieving health benefits, than any component of the activity (i.e., frequency, intensity, or duration) or a specific mix of activities (i.e., aerobic, muscle strengthening, and bone strengthening)<sup>1</sup>. However, based on the 2018 United States Report Card on PA for Children and Youth, only about 24% of children meet the recommended 60 minutes of PA every day<sup>2</sup>. PA is not only vital to the physical, psychological, social, and cognitive health of children<sup>3,4</sup>, but it may also play an important role in shaping the human gut microbiota<sup>5-7</sup>.

The human gut microbiota has been associated with maintaining host physiological states, including immune responses, metabolism, and mental and physical development<sup>8</sup>. Thus, understanding the mechanisms that maintain microbial balance in the gut is important. Changes in the composition of the gut microbiota have been associated with short- and long-term health conditions, such as overweight, obesity<sup>9</sup>, atopy manifestations, metabolic syndromes, chronic inflammatory diseases<sup>10,11</sup>, increased risk of allergy and asthma<sup>12,13</sup>, and neurodevelopmental disorders<sup>14</sup>. Generally, a high microbial diversity is thought to be associated with a healthy gut microbiota, whereas loss of diversity appears to correlate with incidence of disease<sup>15</sup>. The human gut microbiome is a complex community that begins to develop at infancy<sup>16-18</sup>. While the gut microbiome composition is largely determined in early life<sup>19-27</sup>, a variety of factors may perturb its composition across the life course. However, little is known about these factors in pre-adolescents<sup>28,29</sup>.

There is a significant lack of research regarding the effects of lifestyle factors such as PA on the gut microbiome of children, as most of the studies have focused on adults<sup>30,31</sup>.

PA has recently emerged as a probable modulator of the human gut microbiome<sup>32-34</sup>. Animal studies have shown that PA training independently alters the composition and functional capacity of the gut microbiota<sup>35-38</sup> and increases the relative abundance of butyrate-producing taxa<sup>39</sup>. The influence of PA on the human gut microbiota has been reported in several studies<sup>40-42</sup>. For example, increased levels of Faecalibacterium prausnitzii, Roseburia hominis, and Akkermansia muciniphila were observed in women who performed at least 3 hours of PA per week<sup>40</sup>. Moreover, longitudinal intervention studies ranging from six to eight weeks in duration suggest that exercise has independent effects on the gut microbiota43,44. Recently, studies exploring the role of PA on the gut microbiome of children have been published<sup>5-7</sup>. Previous studies have reported that exercise-induced alterations of the gut microbiota in children may depend on BMI and exercise modality and intensity and an obesity-related deleterious microbiota profile may be positively modified by exercise<sup>6</sup>. Specifically, the gut microbiota of children who exercise daily displays enrichment of Firmicutes and the genera Clostridiales, Lachnospiraceae, and Erysipelotrichaceae<sup>6</sup>.

Despite the growing body of evidence showing that the gut microbiota influences health and disease in childhood, and the well-established benefits of PA on children's health<sup>4</sup>, more research is needed on the role of PA in the composition of the gut microbiota in children. While published studies support an association<sup>5-7</sup>, these studies were limited due to factors such as broad age range (7-18 years)<sup>6</sup>, lack of validated instruments to assess PA<sup>6,7</sup>, and limited race/ ethnic diversity. Thus, the primary aim of this study was to determine whether there is an association between PA and gut bacterial microbiome in a cohort of 10-year-old children from a racially and socioeconomically diverse Wayne County Health Environment Asthma and Allergy Longitudinal Study (WHEALS) birth cohort.

## METHODS

#### **Study population**

Data were analyzed from WHEALS, a birth cohort study of 1,258 maternal-child pairs living in the Detroit metropolitan area. Details of the cohort have been published<sup>45-47</sup>. In brief, 21- to 49-year-old pregnant women who were receiving prenatal care at Henry Ford Health System's (HFHS) obstetrics clinics were recruited between 2003 and 2007. They were interviewed at the initial recruitment, as well as during the follow-up visits at 1, 6, 12, and 24 months, and during a phone interview at 36 months postpartum. At 10 years of age, the children enrolled into WHEALS were invited for an additional follow-up clinic visit to perform various measurements of health (i.e., a general health check-up), as well as to complete a questionnaire. Children of 10 years were selected because, while approximately 1/3 of obese preschoolers become obese adults, approximately 1/2 of obese school-age children will be obese as adults<sup>48</sup>. Thus, by studying body size at the age of 10 years, we are studying a time point that reveals a much greater propensity for adult obesity. Written informed consent was obtained from all the participants, including children. This study was approved by the HFHS Institutional Review Board (#1881).



#### **Stool specimen collection**

The subjects who agreed to participate in the 10-year visit were each provided with a stool sample collection kit (Protocult Collection Device paper sample collector, Ability Building Center, Rochester, MN, USA; and a Sarstedt collection tube, Nümbrecht, Germany) to collect a single fecal sample for analysis. The collection kits were distributed among the participants in small insulated cooler bags containing ice packs to keep the samples cold during post-collection transit. The participants were provided with a sheet of instructions inside the cooler bag. They were instructed to freeze their ice packs for at least one day prior to sample collection. The ice packs were rated to remain frozen for 36-48 h in an insulated container. All stool samples were retrieved from the participants or delivered to the research facility within 24h of collection. The stool samples were then stored at -80 °C. 16S rRNA sequencing was used to profile the bacterial gut microbiota present in these stool specimens.

## **DNA extraction**

DNA from the stool samples was extracted using the modified cetyltrimethylammonium bromide (CTAB) method as previously described for bacterial profiling49,50. Briefly, 500 µl of modified CTAB extraction buffer was added to 25 mg of stool in a 2 ml Lysing Matrix E (LME) tube (MP Biomedicals, Irvine, CA, USA) and the mixture incubated at 65 °C for 15 min. Samples were bead-beaten (5.5 m/s, 30 s) in a Fastprep-24 (MP Biomedicals, Irvine, CA, USA) and then centrifuged (16,000  $\times$  g, 5 min). Subsequently, the top aqueous phase was transferred to a 2 ml polypropylene 96-well plate (USA Scientific, Ocala, FL, USA). Modified CTAB extraction buffer (500 µl) was added to each LME tube, bead-beaten, and centrifuged to collect a total of 1 ml of aqueous phase per sample. After adding 1 ml of phenol:chloroform:isoamyl alcohol (25:24:1) to the collected aqueous supernatant, the samples were centrifuged (3,200  $\times$ g, 20 min, 4 °C), and the resulting upper aqueous phase was transferred to a new 2 ml polypropylene 96-well plate (USA Scientific). Polyethylene glycol/NaCl (2% v/v) was added to the collected aqueous supernatant and the mixture incubated at room temperature for 2 h. The samples were then centrifuged  $(3,200 \times g, 60 \text{ min}, 4 \text{ °C})$ , washed with ice-cold 70% ethanol, and resuspended in 30 µl of TE buffer (Invitrogen; Waltham, MA, USA).

## PCR conditions and library preparation for bacterial biomarker sequencing

The V4 region of the 16S rRNA bacterial gene was amplified using primers designed by Caporaso *et al.*<sup>51</sup>. PCR was performed in 25  $\mu$ l reactions using 0.025 U Takara Hot Start ExTaq (Takara Mirus Bio Inc.; San Jose, CA, USA), 1× Takara buffer with MgCl<sub>2</sub>, 0.4 pmol/ $\mu$ l of F515 and R806 primers, 0.56 mg/ml of bovine serum albumin (Roche Applied Science; Penzberg, Germany), 200  $\mu$ M of dNTPs, and 10 ng of gDNA. The reactions were performed in triplicate with the following conditions: initial denaturation at 98 °C for 2 min, 30 cycles at 98 °C for 20 s, annealing at 50 °C



for 30 s, extension at 72 °C for 45 s, and final extension at 72 °C for 10 min. Amplicons from the technical triplicates were pooled and verified using a 2% TBE agarose e-gel (Life Technologies; Carlsbad, CA, USA), cleaned up and normalized using SequalPrep Normalization Plates (Applied Biosystems), and quantified using the Qubit dsDNA HS Assay Kit (Invitrogen, Waltham, MA, USA). The samples were pooled in equal amounts (5 ng), purified using AMPure SPRI beads (Beckman Coulter; Brea, CA, USA), quantified using KAPA SYBR (KAPA Biosystems; Wilmington, MA, USA), denatured and diluted to 2 nM, and 5 pmol was loaded onto the Illumina Nextseq cartridge with 40% (v/v) of denatured 12.5 pM PhiX spike-in control.

## **Biomarker sequence data processing**

Paired-end sequences were assembled using FLASH v1.2.7<sup>52</sup>, requiring a minimum base pair overlap of 25 bp, and demultiplexed by barcode using QIIME v1.9.1<sup>53</sup>. Quality filtering was performed using USEARCH v8.0.1623<sup>54</sup> to remove reads with > 2 expected errors. Quality reads were dereplicated at 100% sequence identity, clustered at 97% sequence identity into operational taxonomic units (OTUs), filtered of chimeric sequences by UCHIME<sup>55</sup>, and mapped back to the resulting OTUs using UPARSE<sup>56</sup>. The sequence reads that failed to cluster with a reference sequence were clustered *de novo*. Taxonomy was assigned to the OTUs using the Greengenes v13\_5 database<sup>57</sup>. Sequences were aligned using PyNAST<sup>58</sup>, and a phylogenetic tree was constructed using FastTree 2.1.3<sup>59</sup>.

### PA assessment at 10 years of age

The block PA screen (BKPAS), which has been validated for use in 8- to 17-year-old children<sup>60</sup>, was supplied to the children during their 10-year study visit. The BKPAS is an 18-item PA screener developed by NutritionQuest that uses a self-report of activity over the previous 7 days to provide information about the frequency and duration of PA.

Overall, five total exposure variables were used to quantify the level of PA. Three of these variables were directly obtained from NutritionQuest: estimated daily moderate PA (MPA) minutes, estimated daily vigorous PA (VPA) minutes, and estimated daily recreational PA (RPA) minutes. The analysis program assigns each item to a metabolic equivalent (MET) value and classifies each item as either moderate or vigorous MET activity. It combines these MET values with the screener responses to calculate the number of MET minutes per day for each screener line item. It then calculates the number of MET minutes spent in the moderate and vigorous categories. The estimated daily moderate-to-vigorous PA (MVPA) minutes was calculated by summing the number of MPA and VPA minutes. Total MVPA was then used to determine if the child met the recommendation of 60 or more minutes of daily MVPA<sup>1</sup>. For this study, the primary exposure was meeting the recommendations of PA guidelines, as this most closely aligns with clinical recommendations, while the minutes per day variables were considered secondary exposures.

## **Potential confounders**

Self-reported education of mothers was collected during prenatal interviews. Household income of the parent/guardian was recorded during the 10-year visit. The season at the 10-year visit was noted. Detroit residency criteria (as a measure of urban vs. suburban residence) were assessed using the location of residence (or last known location) provided during the 10-year-old visit. Prenatal and delivery records of mothers were abstracted to determine the child's sex and delivery type. The child's race was maternally reported at the 24-month postpartum visit. Breastfeeding status was maternally reported at the 1-month and 6-month postpartum visits.

At the 10-year-old follow-up visit, clinical research assistants measured the height and weight of children using protocols adapted from the PhenX Toolkit<sup>61</sup>. Body mass index (BMI) Z-score was calculated to define the BMI category (i.e., obese/overweight vs. normal/underweight), and overweight or obesity was defined as  $\geq 85^{\text{th}}$  percentile according to the Centers for Disease Control and Prevention growth charts (2000)<sup>62</sup>. The Block Kids Food Screener<sup>63</sup> was also included in the 10-year visit. Briefly, the children were asked about the frequency and quantity of consumption of various foods in the previous week to determine the food group and nutrient estimates. Latent class analysis was then used to determine the dietary patterns at age 10 as processed/energy-dense foods, variety plus high intake, and healthy<sup>45</sup>. The Pubertal Development Scale<sup>64</sup> was also included in the 10-year questionnaire to quantify pubertal development. Antibiotic use within 4 weeks prior to the 10year stool sample collection was recorded using both chart abstraction and maternal reports.

#### **Statistical analysis**

The statistical significance level was set at 0.05. Meeting the PA recommendation was associated with potential confounders using chi-square tests and ANOVA, while Kruskal-Wallis tests and Spearman correlations were used for the minutes/day variables. The rarefied OTU table was used to calculate the alpha (richness, Pielou's evenness, and Faith's phylogenetic diversity) and beta (unweighted and weighted UniFrac)65 diversity metrics. PA was associated with alpha diversity metrics using linear regression models. Associations with beta diversity were assessed using PERMANO-VA with 10,000 permutations, using the adonis function of the R vegan package<sup>66</sup>. The multivariate homogeneity of group dispersions assumption of PERMANOVA was evaluated using the betadisper function of the R vegan package<sup>66</sup>. See Additional File 1; Table S1 for alpha and beta diversity definitions. The unrarefied OTU table was used to examine OTU and other taxa-level differences. Taxonomic testing was performed using DESeq2 in the R packages phyloseg67 and DESeq268, with a false discovery rate (FDR)69 adjusted p-value < 0.05 considered significant.

The set of confounders to adjust for throughout analyses was determined using the "disjunctive cause criterion" of the variables associated with the exposure or outcome, or both<sup>70</sup>. Among the set of hypothesized confounders, those



that exhibited associations with any of the PA variables, alpha diversity, or beta diversity were retained for the adjustment. This included prenatal maternal education, household income at age 10, location of residence at age 10, season during the 10-year visit, sex of the child, race of the child, breastfeeding status at 1-month, BMI category at age 10, and dietary pattern at age 10. Child sex, race, and BMI category at age 10 were prespecified as potential effect modifiers and were tested using interaction terms.

## RESULTS

A total of 383 10-year-old children studied under WHEALS provided stool samples for 16S rRNA sequencing, of which 321 (84%) also completed the BKPAS. The children who met PA recommendations spent an average of 139 minutes per day on MVPA (SD = 86, min = 61, max = 494) compared with the average of 33 minutes per day among children who did not meet the recommendations (SD=16, min=0, max=59). Children with and without BK-PAS data were compared to examine potential sources of bias. This analysis revealed only one significant variable, season at the 10-year visit (p = 0.033), with children who completed the BKPAS being more likely to have the 10-year visit in the spring or summer. This likely reflects the fact that children completing the visit during the spring or summer had more time to complete all study activities.

## Factors associated with PA

We first examined factors associated with PA at 10 years of age. None of the examined factors were significantly associated with meeting the recommendation of 60 MVPA minutes per day (Table 1). For the minutes/day variables,

Table 1. Association between maternal and child characteristics and meeting the recommendations of daily physical activity minutes.

Variable	ا مرما	MET Recommenda Minutes of Daily Mo Physical	ation of 60 or More oderate-to-Vigorous Activity	n-value <sup>a</sup>
Variable	Level	No. N=132	Yes. N=189	p-value
		N (Column %) o	or N, Mean (SD)	
0	Male	66 (50%)	108 (57.1%)	0.000
Sex	Female	66 (50%)	81 (42.9%)	0.206
DNI Cotomery at Are 10	Normal/Under	81 (61.4%)	132 (69.8%)	0.444
Bini Calegory at Age 10	Obese/Overweight	51 (38.6%)	57 (30.2%)	0.114
	DP1: Processed/EDF	53 (40.2%)	57 (30.2%)	
Dietary Pattern (DP) at Age 10	DP2: Variety + High Intake	42 (31.8%)	84 (44.4%)	0.060
	DP3: Healthy	37 (28%)	48 (25.4%)	
	Never Breastfed	23 (17.6%)	40 (21.7%)	
Breastfeeding Status at 1-Month	Mixed Feeding	85 (64.9%)	118 (64.1%)	0.534
-	Breastfeeding Only	23 (17.6%)	26 (14.1%)	
	Never Breastfed	23 (17.6%)	40 (21.7%)	
Breastfeeding Status at 6-Months	Mixed Feeding	98 (74.8%)	134 (72.8%)	0.524
	Breastfeeding Only	10 (7.6%)	10 (5.4%)	
Mada of Dalitana	Vaginal	85 (64.9%)	122 (64.6%)	0.054
Mode of Delivery	C-section	46 (35.1%)	67 (35.4%)	0.951
	White	30 (22.7%)	44 (23.3%)	
Child Race	African American	86 (65.2%)	116 (61.4%)	0.683
	Other/Mixed	16 (12.1%)	29 (15.3%)	
Mother's Education	HS Diploma or Less	19 (14.4%)	30 (15.9%)	0 747
Mother's Education	Some College or More	113 (85.6%)	159 (84.1%)	0.717
	<\$40K	36 (27.3%)	59 (31.2%)	
	\$40K-<\$80K	43 (32.6%)	52 (27.5%)	
Household Income at 10-Year Visit	\$80K-<\$120K	23 (17.4%)	38 (20.1%)	0.794
	\$120 or more	21 (15.9%)	30 (15.9%)	
	Missing/Refused to Answer	9 (6.8%)	10 (5.3%)	
Location of Residence (or Last Known	Suburban	69 (54.3%)	103 (57.5%)	0 577
Location) at 10-Year Visit	Urban	58 (45.7%)	76 (42.5%)	0.577
Antibiotic Use Within 4 Weeks Prior to 10-Year	No	130 (98.5%)	188 (99.5%)	0.266
Stool Sample Collection	Yes	2 (1.5%)	1 (0.5%)	0.300
	Winter	40 (30.3%)	45 (23.8%)	0.558
Season at the 10 Year Visit	Spring	28 (21.2%)	49 (25.9%)	
Season at the 10-real visit	Summer	39 (29.5%)	56 (29.6%)	
	Fall	25 (18.9%)	39 (20.6%)	
10-Year Pubertal Development Score		126, 1.95 (0.53)	184, 1.92 (0.48)	0.551

<sup>a</sup>Chi-square test for categorical variables and ANOVA for numerical variables



Table 2. /	Association between mater	nal and child characteri	stics and estimated da	ily physical activity minute	s.

Variable	Level	Level N		Estimat ate ac	ed daily tivity mi	moder- nutes	Estimated daily vigorous activity minutes			Estimated daily moderate-to-vigorous activity minutes			Estimated daily recreational activity minutes		
			Median	IQR	<i>p</i> - valueª	Median	IQR	<i>p</i> - valueª	Median	IQR	<i>p-</i> valueª	Median	IQR	<i>p-</i> value <sup>ª</sup>	
Sex	Male Female	174 147	53.4 53.1	68.1 73.5	0.974	15 8.4	42.6 30	0.021	74.1 70.2	88.8 78.6	0.38	55.2 46.8	75 66	0.354	
BMI Category at	Normal/Under	213	52.5	68.4	0 905	15	35.7	0 347	73.5	77.1	0.455	55.5	66.6	0 128	
Age 10	Overweight	108	54.45	73.95	0.305	8.4	32.55	0.547	69.75	91.35	0.400	42.6	72.75	0.120	
Dietary Pat-	DP1: Pro- cessed/EDF	110	47.25	56.7		8.4	27.9	0 757	62.1	80.1	0.005	39.3	70.2		
tern (DP) at Age 10	DP2: Variety + High	126	61.35	72	0.028	15	36.3	0.757	78.9	88.2	0.085	60.6	72.6	0.03	
	DP3: Healthy	85	53.1	79.2		15	31.5		72.3	85.8		49.2	62.4		
Breastfeed-	Never breastfed	63	71.7	94.8		16.8	55.8		93	105.9		60	95.4		
ing Status at	Mixed Feeding	203	51.9	63	0.171	12.6	31.5	0.03	72.3	79.2	0.104	48.9	70.5	0.263	
1-Month	Breastfeeding Only	49	46.8	61.8		7.5	30		61.8	62.4		48.6	60		
Breastfeed-	Never breastfed	63	71.7	94.8		16.8	55.8		93	105.9		60	95.4		
ing Status at	Mixed Feeding	232	51.3	61.35	0.14	10.5	31.8	0.055	71.85	77.55	0.075	48.9	71.7	0.174	
6-Months	Breastfeeding Only	20	45.6	64.65		8.4	16.8		59.7	55.8		43.5	42		
Mode of	Vaginal	207	53.1	69.9	0.881	12.6	33.6	0.275	72.6	81	0.834	49.2	73.5	0.689	
Delivery	C-section White	113 74	53.4 45.15	70.8 58.5		15 16.8	29.4 38.4		74.4 69	94.5 73.5		55.5 51.9	62.4 60		
Child Race	African	202	56.1	69	0.221	12.15	33.6	0.708	73.05	88.8	0.604	46.8	76.5	0.498	
	Other/Mixed	45	61.8	78		8.4	34.2		84.9	86.4		60	58.2		
Mother's Edu-	HS diploma or less	49	61.8	64.5	0.393	12.6	34.2	0.972	72.3	84.9	0.524	53.4	62.4	0.669	
cation	Some college or more	272	51.9	69.9		12.6	31.5		72.6	83.4		51	72.15		
	<\$40K	95	62.1	77.4		8.4	33.6		75	87.9		46.8	67.5		
Household	\$40K-<\$80K	95 61	49.2	85.8		10.5	30 22 6		72.3	95.7 75.0		46.8	82.5		
Income at 10-	\$120 or more	51	51.5 47.1	60.6 64.2	0.376	16.8	33.0 46.8	0.483	76.9 67.2	75.9 83.7	0.901	53.4	92.4	0.818	
Year Visit	Missing/	10	1 A A	117		16.0	10.9		66	46.2		51	546		
	Answer	19	44.4	44.7		10.0	40.0		00	40.2		51	54.0		
Location of Residence (or	Suburban	172	55.35	68.85	0 602	11.1	27.9	0.562	73.5	75.45	0 824	52.2	64.05	0 553	
Location) at	Urban	134	51.6	76.5	0.002	12.6	36.3	0.502	72.45	98.7	0.024	47.7	80.1	0.000	
Antibiotic															
Use Within 4 Weeks Prior	No	318	53.4	70.8	0.425	13.8	31.5	0.074	72.6	84.6	0.214	51	69.37	0.27	
Stool Sample Collection	Yes	3	41.7	87		0	6.3		41.7	80.7		30	57.3		
	Winter	85	48.9	75		8.4	30		64.8	93		46.2	70.2		
Season at the 10-Year Visit	Spring Summer	77 95	66.3 51.9	73.8 69.9	0.385	11.7 15	35.7 27 9	0.674	91.8 72 3	88.8 68 1	0.678	60 48 6	74.4 60	0.561	
	Fall	64	49.5	60.6		15.9	34.2		69.9	81.15		52.2	70.05		
		N	Spear	man r	<i>p</i> - value⁵	Spearm	nan r	<i>p</i> - value <sup>b</sup>	Spearr	nan r	<i>p</i> - value <sup>b</sup>	Spearr	man r	 value <sup>b</sup>	
10-Year Pu- bertal Devel- opment Score		310	0.0	69	0.228	-0.04	17	0.413	0.03	33	0.565	-0.0	24	0.671	

<sup>a</sup>Kruskal-Wallis test p-value. <sup>b</sup>Spearman correlation p-value.



(Table 2), sex, dietary pattern at age 10, and breastfeeding status at 1-month were significantly associated with PA measures. Specifically, males had higher VPA minutes per day (p = 0.021) and children with diets characterized by processed and energy-dense foods had the least MPA (p = 0.028) and RPA (p = 0.030) minutes per day. Additionally, children who were never breastfed had the highest number of VPA minutes per day (p = 0.030).

## **Relationships between PA and microbiota diversity**

When the overall associations between PA and child gut alpha diversity metrics were assessed, no significant associations were found both before and after covariate adjustment (Table 3). However, in tests examining whether these associations differ by child sex, race, or BMI category at age 10, a total of 12 significant interactions were found, although only six of these had any within-group significance (Table 4). These six interactions modified the effects of overweight/ obesity at age 10, race, and sex. Specifically, there was a significant positive association between meeting PA recommendations and bacterial evenness among the normal/underweight children, but a non-significant negative association between the same among obese/overweight children. Additionally, race modified the association between VPA minutes and richness, where increased VPA minutes were associated with lower richness among mixed/other-race children only. Furthermore, sex modified the association between MPA minutes and diversity, where only among females, increased MPA minutes were associated with increased diversity.

## Relationships between PA and microbiota composition

Next, we evaluated the associations between PA and

Та	b	le :	3. /	Assoc	iat	ion	between	phy	sica	activi	ty and	l alp	bha	diversit	y metrics.
----	---	------	------	-------	-----	-----	---------	-----	------	--------	--------	-------	-----	----------	------------

microbiota composition (beta diversity; Table 5). Those who did or did not meet the recommendation of 60 MVPA minutes per day exhibited compositionally distinct gut microbiota both before and after covariate adjustment, based on weighted UniFrac (adjusted p = 0.003), which explained 1.3% of the variation in the composition. For a sensitivity analysis, we removed the three children who had used antibiotics in the 4 weeks prior to stool sample collection, and the results were still very similar (weighted UniFrac adjusted  $R^2 = 0.014$ , p = 0.002). Evaluation of the multivariate homogeneity of group dispersions assumption showed no significant difference in dispersions by group (p = 0.16), indicating that the significant PERMANOVA test result was due to true compositional distinction rather than unequal dispersions. In addition, estimated daily MPA minutes were initially associated with overall stool microbiota composition defined by unweighted UniFrac (p = 0.045,  $R^2 = 0.005$ ), but this was no longer significant after covariate adjustment  $(R^2 = 0.003, p = 0.334).$ 

Examination of the effect modifications by child sex, race, and BMI category at age 10 (Table 6) revealed three significant interactions for unweighted UniFrac/sex only. Specifically, the association between MVPA minutes as well as RPA minutes and composition was found to depend on the sex; however, none of the strata-specific *p*-values reached significance. Sex also modified the effect between MPA minutes and composition (unweighted UniFrac interaction p = 0.005), where an association was found among females (unweighted UniFrac R<sup>2</sup> = 0.012, p = 0.048), but not among males (unweighted UniFrac R<sup>2</sup> = 0.005, p =0.623).

Alpha Diversity Metric	Physical Activity Variable	Uı	nadjuste	d	Adjusted <sup>c</sup>			
Alpha Diversity metho	Thysical Activity variable	β	SE	<i>p</i> -value	β	SE	<i>p</i> -value	
Richness	Met recommendation of 60 or more minutes of daily moderate-to-vigorous physical activity	-1.21 <sup>b</sup>	7.78	0.88	-0.76 <sup>b</sup>	7.97	0.92	
	Estimated daily moderate activity minutes	5.13ª	4.04	0.21	3.50ª	4.14	0.40	
	Estimated daily vigorous activity minutes Estimated daily moderate-to-vigorous activity minutes Estimated daily recreational activity minutes	1.14ª 2.53ª 1.50ª	5.91 2.70 2.92	0.85 0.35 0.61	1.23ª 1.80ª 0.86ª	5.98 2.75 2.98	0.84 0.51 0.77	
Evenness	Met recommendation of 60 or more minutes of daily moderate-to-vigorous physical activity	0.005 <sup>b</sup>	0.005	0.33	0.004 <sup>b</sup>	0.006	0.46	
	Estimated daily moderate activity minutes	0.0007 <sup>a</sup>	0.003	0.79	0.0002 <sup>a</sup>	0.003	0.96	
	Estimated daily vigorous activity minutes	$-0.003^{a}$	0.004	0.40	$-0.003^{a}$	0.004	0.51	
	Estimated daily moderate-to-vigorous activity minutes	-0.0004ª	0.002	0.84	-0.001ª	0.002	0.79	
	Estimated daily recreational activity minutes	-0.001ª	0.002	0.63	-0.001ª	0.002	0.59	
Phylogenetic Diversity	Met recommendation of 60 or more minutes of daily moderate-to-vigorous physical activity	0.16 <sup>b</sup>	0.78	0.84	0.31 <sup>b</sup>	0.80	0.70	
	Estimated daily moderate activity minutes	0.65ª	0.40	0.11	0.55ª	0.42	0.19	
	Estimated daily vigorous activity minutes	0.17ª	0.59	0.77	0.14 <sup>a</sup>	0.60	0.81	
	Estimated daily moderate-to-vigorous activity minutes	0.33ª	0.27	0.23	0.27ª	0.28	0.33	
	Estimated daily recreational activity minutes	0.21ª	0.29	0.47	0.16 <sup>ª</sup>	0.30	0.61	

<sup>a</sup>Mean difference in alpha diversity metric for each 1-hour increase in a specific type of physical activity.

<sup>b</sup>Mean difference in alpha diversity metric comparing those who did and did not meet recommendations.

<sup>c</sup>Adjusted for prenatal maternal education, household income at age 10, location of residence at age 10, season at the 10-year visit, sex, child race, breastfeeding status at 1-month, BMI category at age 10, and dietary pattern at age 10. N = 300 due to some missing covariates.



Table 4. Association between physical activity and alpha diversity metrics by child sex, race, and BMI category at age 10.

Physical Activity	Effect	Interaction		Strata 0 <sup>b</sup>			Strata 1°			Strata 2	ł
Variable	Modifier	<i>p</i> -value <sup>a</sup>	β	SE	<i>p</i> -value	β	SE	<i>p</i> -value	β	SE	<i>p</i> -value
				Rich	ness						
Met recommendation of	Sex	0.208	-4.56	10.81	0.674	0.58	11.98	0.961			
60 or more minutes of	Race	0.802	-17.76	17.05	0.303	2.76	10.09	0.785	-27.39	27.34	0.325
physical activity	BMI	0.112	12.37	10.55	0.243	-18.86	13.63	0.170			
Estimated daily moderate	Sex	0.019	-1.32	5.29	0.803	12.33	6.74	0.070			
activity minutes	Race	0.112	4.80	8.51	0.576	5.85	5.41	0.281	-11.49	13.30	0.396
dourny minuted	BMI	0.568	7.41	5.30	0.164	0.73	7.53	0.923			
Estimated daily vigorous	Sex	0.338	-1.78	7.61	0.816	2.75	9.56	0.774	45.00	10.02	0.024
activity minutes	RACE	0.019	4.38	14.00 8.28	0.104	3.10 3.40	7.44 9.38	0.009	-45.00	19.03	0.031
Estimated daily moder-	Sex	0.045	-0.95	3.48	0.786	6.07	4.51	0.181			
ate-to-vigorous activity	Race	0.025	6.28	6.16	0.313	3.04	3.44	0.378	-13.99	8.98	0.131
minutes	BMI	0.552	4.42	3.67	0.230	1.08	4.58	0.814			
Estimated daily recreation-	Sex	0.064	-1.70	3.69	0.644	4.98	5.00	0.321			
al activity minutes	Race	0.041	5.33	6.76	0.434	2.02	3.68	0.583	-15.75	10.35	0.140
,	BMI	0.710	3.16	4.12	0.445	1.03	4.64	0.825			
Mot recommendation of				Even	iness						
60 or more minutes of	Sex	0.679	0.005	0.009	0.582	0.002	0.007	0.741			
daily moderate-to-vigorous	Race	0.785	0.016	0.013	0.225	0.002	0.007	0.802	0.004	0.015	0.802
physical activity	BMI	0.032	0.015	0.008	0.049	-0.014	0.009	0.128			
Estimate didations denote	Sex	0.783	0.000	0.004	0.954	-0.001	0.004	0.789			
activity minutes	Race	0.809	0.003	0.007	0.618	-0.001	0.004	0.837	0.008	0.007	0.271
activity minutes	BMI	0.004	0.007	0.004	0.063	-0.013	0.005	0.008			
Estimated deily vicerous	Sex	0.359	0.002	0.006	0.796	-0.012	0.006	0.041			
activity minutes	Race	0.308	0.008	0.011	0.446	0.000	0.005	0.957	-0.012	0.012	0.328
,	BMI	0.167	0.003	0.006	0.612	-0.010	0.006	0.109			
Estimated daily moder-	Sex	0.799	0.000	0.003	0.936	-0.003	0.003	0.252			
ate-to-vigorous activity	Race	0.854	0.003	0.005	0.488	0.000	0.002	0.916	0.002	0.005	0.736
minutes	BMI	0.011	0.004	0.003	0.130	-0.007	0.003	0.017			
Estimated daily recreation-	Sex	0.610	0.000	0.003	0.954	-0.005	0.003	0.091			
al activity minutes	Race	0.900	0.003	0.005	0.577	-0.001	0.003	0.711	0.002	0.006	0.714
	BMI	0.031	0.003	0.003	0.249	-0.007	0.003	0.030			
	-		F	hylogenet	tic Diversit	У					
Met recommendation of	Sex	0.087	-0.470	1.080	0.664	0.873	1.240	0.483			
daily moderate-to-vigorous	Race	0.854	-1.769	1.806	0.332	0.636	1.007	0.528	-2.53	2.71	0.358
physical activity	BMI	0.095	1.584	1.052	0.134	-1.975	1.392	0.160			
Estimated daily moderate	Sex	0.013	0.009	0.528	0.986	1.564	0.694	0.026			
activity minutes	Race	0.343	0.376	0.902	0.679	0.692	0.540	0.202	-0.65	1.33	0.628
activity minutes	BMI	0.389	0.990	0.527	0.062	-0.067	0.770	0.931			
Estimated daily vigorous	Sex	0.149	-0.401	0.760	0.599	0.747	0.990	0.452			
activity minutes	Race	0.057	1.802	1.498	0.235	0.308	0.744	0.680	-3.66	2.02	0.081
activity minutes	BMI	0.385	0.710	0.826	0.391	-0.037	0.959	0.969			
Estimated daily	Sex	0.021	-0.080	0.348	0.818	0.860	0.465	0.067			
moderate-to-vigorous	Race	0.124	0.538	0.654	0.415	0.345	0.344	0.317	-1.00	0.91	0.279
activity minutes	BIMI	0.292	0.616	0.300	0.094	-0.034	0.468	0.943			
Estimated daily	Daga	0.030	-0.194	0.300	0.000	0.700	0.010	0.141	1 00	1 0 /	0.210
minutes	RMI	0.100	0.410	0./1/	0.000	0.219	0.300	0.002	-1.00	1.04	0.310
	DIVII	0.555	0.012	0.411	0.210	-0.072	0.474	0.079			

<sup>a</sup>Adjusted for prenatal maternal education, household income at age 10, location of residence at age 10, season at the 10-year visit, sex, child race, breastfeeding <sup>b</sup>If effect modifier is sex, strata 0 = male. If effect modifier is race, strata 0 = white. If effect modifier is BMI, strata 0 = normal/underweight.
<sup>c</sup>If effect modifier is sex, strata 1 = female. If effect modifier is race, strata 1 = black. If effect modifier is BMI, strata 1 = obese/overweight.
<sup>d</sup>If effect modifier is race, strata 2 = other/mixed. Otherwise, disregard (blank).



Table 5 Association	hotwoon	nhysical	activity	and hota divorsity	
Table 5. Association	Detween	physical	activity	and beta diversity.	

Data Diversity Matria	Dhusical Astivity Veriable	Unadj	usted	Adjusted <sup>a</sup>		
Beta Diversity Metric	Physical Activity variable	<i>p</i> -value	R <sup>2</sup>	<i>p</i> -value	R <sup>2</sup>	
Unweighted UniFrac	Met recommendation of 60 or more minutes of daily moderate-to- vigorous physical activity	0.451	0.003	0.469	0.003	
	Estimated daily moderate activity minutes	0.045	0.005	0.334	0.003	
	Estimated daily vigorous activity minutes	0.231	0.004	0.506	0.003	
	Estimated daily moderate-to-vigorous activity minutes	0.051	0.005	0.335	0.003	
	Estimated daily recreational activity minutes	0.194	0.004	0.491	0.003	
Weighted UniFrac	Met recommendation of 60 or more minutes of daily moderate-to- vigorous physical activity	0.001	0.014	0.003	0.013	
	Estimated daily moderate activity minutes	0.056	0.007	0.107	0.006	
	Estimated daily vigorous activity minutes	0.157	0.005	0.229	0.004	
	Estimated daily moderate-to-vigorous activity minutes	0.065	0.007	0.114	0.005	
	Estimated daily recreational activity minutes	0.086	0.006	0.117	0.005	

<sup>a</sup>Adjusted for prenatal maternal education, household income at age 10, location of residence at age 10, season at the 10-year visit, sex, child race, breastfeeding status at 1-month, BMI category at age 10, and dietary pattern at age 10. N = 300 due to some missing covariates.

Table 6.	Association	between pl	hysical activit	y and beta	diversity by	/ child sex,	race, and	BMI category	at age 10.

	Interaction		Strata 0 <sup>b</sup>					Strata 1 <sup>°</sup>				Strata 2 <sup>d</sup>			
Effect Modifier	<i>p</i> -valu	les <sup>a</sup>	Unw Un	eighted iFrac	Weig Un	ghted iiFrac	Unwe Uni	ighted Frac	Weig Unif	hted Frac	Unwei Unif	ghted Frac	Weig Uni	ghted Frac	
	Unweighted UniFrac	Weighted UniFrac	<b>R</b> <sup>2</sup>	<i>p</i> -value	<b>R</b> <sup>2</sup>	<i>p</i> -value	<b>R</b> <sup>2</sup>	<i>p</i> -value	<b>R</b> <sup>2</sup>	<i>p</i> -value	<b>R</b> <sup>2</sup>	<i>p</i> -value	<b>R</b> <sup>2</sup>	<i>p</i> -value	
Met recor	mmendation o	f 60 or more	e minut	tes of dail	y modei	rate-to-vig	orous p	hysical ac	tivity						
Sex	0.122	0.098	0.006	0.364	0.024	0.003	0.006	0.523	0.004	0.731					
Race	0.886	0.394	0.013	0.664	0.007	0.878	0.005	0.392	0.024	0.003	0.021	0.584	0.017	0.598	
BMI	0.242	0.793	0.005	0.276	0.012	0.027	0.010	0.347	0.026	0.029					
Estimated	d daily modera	te activity r	minute	s											
Sex	0.005	0.240	0.005	0.623	0.012	0.077	0.012	0.048	0.006	0.532					
Race	0.088	0.136	0.012	0.795	0.025	0.182	0.007	0.095	0.015	0.018	0.020	0.670	0.039	0.093	
BMI	0.471	0.161	0.006	0.258	0.003	0.675	0.011	0.253	0.032	0.006					
Estimated	d daily vigorou	is activity n	ninutes	;											
Sex	0.210	0.665	0.006	0.405	0.006	0.366	0.006	0.623	0.005	0.563					
Race	0.214	0.952	0.013	0.647	0.017	0.416	0.005	0.350	0.006	0.234	0.025	0.314	0.020	0.482	
BMI	0.420	0.163	0.004	0.675	0.005	0.336	0.010	0.316	0.017	0.107					
Estimated	d daily modera	te-to-vigor	ous act	tivity minu	utes										
Sex	0.026	0.314	0.005	0.560	0.011	0.103	0.010	0.091	0.006	0.516					
Race	0.083	0.520	0.012	0.811	0.013	0.602	0.007	0.115	0.012	0.035	0.022	0.505	0.029	0.233	
BMI	0.345	0.149	0.005	0.299	0.004	0.538	0.011	0.202	0.029	0.015					
Estimate	d daily recreat	ional activit	ty minu	ites											
Sex	0.035	0.615	0.006	0.487	0.009	0.165	0.009	0.195	0.005	0.607					
Race	0.181	0.679	0.011	0.854	0.012	0.669	0.006	0.211	0.011	0.054	0.021	0.586	0.022	0.418	
BMI	0.454	0.174	0.005	0.532	0.003	0.627	0.010	0.329	0.026	0.026					

<sup>a</sup>Adjusted for prenatal maternal education, household income at age 10, location of residence at age 10, season at the 10-year visit, sex, child race, breastfeeding status at 1-month, BMI category at age 10, and dietary pattern at age 10.

<sup>b</sup>If effect modifier is: sex, strata 0 = male; race, strata 0 = white and; BMI, strata 0 = normal/underweight.

If effect modifier is: sex, strata 1 = female; race, strata 1 = African American and; BMI, strata 1 = obese/overweight.

<sup>d</sup> If effect modifier is race, strata 2 = other/mixed. Otherwise, disregard (blank).

# Relationships between PA and gut bacterial relative abundance

We first evaluated taxonomic differences at the phylum level to determine large taxonomic shifts (Figure 1). Actinobacteria was significantly decreased in those who met the recommendations ( $p_{FDR} = 0.022$ ), while no other phyla reached statistical significance. Further analyses were then

performed to determine which bacterial taxa were differentially abundant between the children who did and did not meet the PA recommendations of 60 or more daily MVPA minutes. After covariate adjustment, two genera were significantly associated with meeting the PA recommendations (see Additional File 1; Table S2). Specifically, those who met the PA recommendations had significantly increased





Figure 1. Mean relative abundance at the phylum level among 10-year-old children who did and did not meet physical activity recommendations.



**Figure 2.** The number of significant OTUs (after covariate adjustment, FDR p < 0.05) within each genus by direction of association with meeting physical activity recommendations. Only the genera that had three or more significant OTUs are plotted.

abundance of *Megamonas* and *Anaerovorax* (both adjusted  $p_{\rm FDR} < 0.05$ ). Additionally, 186 OTUs reached statistical significance after covariate adjustment. Six OTUs of the genera *Christensenellaceae\_R-7\_group* were significant, five of which had higher abundance in those who met the recommendations. Most of the members of the significant genus *Subdoligranulum* (3/4) also had higher abundance in children who met the recommendations (Figure 2). On the contrary, most of the members of the significant genera *Blautia* (9/12), *Faecalibacterium* (4/5), *Roseburia* (3/4), and *Ruminococcaceae\_UCG-014* (3/4) were less abundant in children who met the recommendations.

## DISCUSSION

The aim of this study was to examine whether there is an association between PA and gut bacterial microbiome in 10-year-old children from a racially and socioeconomically diverse WHEALS birth cohort. In the current study, 59% of the children reported PA levels meeting the recommendations of 60 or more minutes of daily MVPA, compared with the nationally representative data of 42.5% of 6–11 year



old children from the 2018 United States Report Card on Physical Activity for Children and Youth<sup>2</sup>. After covariate adjustment, PA was not significantly associated with gut microbiota richness, evenness, or diversity at the age of 10. However, meeting recommendations of 60 or more minutes of daily MVPA was associated with distinct gut bacterial composition, which was partly characterized by an increased abundance of *Megamonas* and *Anaerovorax*, as well as specific *Christensenellaceae R-7 group* OTUs, among others.

In this study, OTUs of the genera Christensenellaceae R-7 group, and Subdoligranulum were enriched in children who met the PA recommendations. Christensenellaceae has been previously characterized as part of a healthy gut signature<sup>71</sup>. Various relationships between *Christensenellaceae* abundance and human health have been established, and its enrichment in individuals with normal BMI is considered the most validated association between the gut microbiome and BMI72,73. Moreover, Christensenellaceae has also been reported to be negatively correlated with multiple cardiometabolic risk factors including visceral fat mass, blood pressure, and metabolic syndrome<sup>72,74</sup>. A recent study found a higher relative abundance of *Christensenellaceae* R-7 group in adolescents with obesity who completed a six-week exercise program<sup>75</sup>. Furthermore, Subdoligranulum has been previously identified to be associated with a healthier metabolic status<sup>76</sup>. Therefore, our study suggests that meeting the PA recommendations may help to create a healthy gut signature in children.

This study reports a positive association between meeting the daily recommended duration of MVPA and the abundance of Megamonas and Anaerovorax in children. This is in contrast with a study on Italian adults with obesity in which Megamonas abundance was negatively correlated with the level of PA77. These conflicting results may be due to differences in species or strains of this genus or the dietary inputs of these populations, emphasizing the need for additional studies of these relationships across the life course. Research connecting Anaerovorax to PA is limited, with many of the current studies being conducted in animal models<sup>78-80</sup>. In mice subjected to a swimming test, those with high exercise capacity had significantly lower abundance of Anaerovorax than those with low exercise capacity<sup>78</sup>. Anaerovorax has been shown to produce butyrate<sup>79</sup>, which can increase insulin sensitivity and reduce adiposity, while increasing energy expenditure when supplemented in the diet of obese mice80. Further investigation into the relationship between Anaerovorax and PA in children is needed. Moreover, several other OTUs that we found to have altered abundance at different activity levels have also been explored in other studies. Although not as widely studied as Christensenellaceae in the context of cardiovascular health, increased exercise in adults has also been associated with a higher abundance of Faecalibacterium<sup>40</sup>. Conversely, we found a lower abundance of Faecalibacterium in children who met the PA recommendations. While this finding is contrary to the reported effects of exercise on Faecalibacterium in adults, further research is needed to determine whether Faecalibacterium may be more significantly influenced by lifestyle factors other than exercise. There are reports of higher levels of Faecalibacterium in the gut of US adolescents than in that of Egyptian adolescents, which could be due to the differing metabolic needs between a Western diet rich in processed foods, animal protein, and fats and a Mediterranean diet rich in plant foods<sup>81</sup>. Further research is needed to determine whether Faecalibacterium is more influenced by specific macro and micronutrients, as dietary patterns at age 10 were significantly associated with PA in the current study. Our results suggest that the abundance of Faecalibacterium may not be altered in children who meet PA minimums to the same extent as it might in adults who exercise at least 3 hours per week. In addition, our findings suggest lower abundance of Blautia in children who met PA recommendations, which appears to be similar to the results found in animal models<sup>82</sup> and adults<sup>83</sup>. We also found lower abundance of Roseburia in children who met PA recommendations, which is contrary to what has been found in studies focused on active adult women<sup>40</sup>. Moreover, previous studies on pediatric patients with obesity have reported that PA tended to increase the abundance of both Roseburia and Blautia genera5; however, the conditions of a controlled intervention study make it difficult to compare these results to our observational findings. Additional studies examining the complex interrelationships of PA and gut microbiota composition and health outcomes in humans are needed.

Several potential mechanisms have been proposed to explain the impact of PA on the gut microbiome. Exercise has many effects on metabolism, including decreased intestinal transit time<sup>84</sup>. Changes in intestinal transit time likely affects the gut microenvironment, altering intestinal pH, nutrient availability, and composition of mucus, which could alter the types and activities of the microbes that colonize the intestine<sup>33</sup>. Moreover, immune cells reside in close proximity to microbial communities in the tissues of the gut, and exercise might alter the gene expression of these immune cells, leading to the production of fewer pro-inflammatory and more anti-inflammatory cell-signaling proteins<sup>85</sup>. Differential effects on individual gut microbiome taxa require further exploration to understand how PA influences health in humans. Future studies that measure stool metabolites are required to further understand the potential mechanistic pathways.

The limitations of this study include the cross-sectional design and the collection of a single fecal sample, which is subject to day-to-day variability. Although we *a priori* hypothesized that PA would influence the gut microbiota composition, we cannot rule out reverse causality (i.e., gut microbiota composition influences a child's activity level), although the results from animal studies provide support for our proposed direction of relationship. Additionally, the BK-PAS evaluates PA based on the recall method with a short duration (7 days) that was only taken at a single time point. Future studies would benefit from a longitudinal measure of PA. Although the strength of our study was the use of validated self-report instruments for the assessment of PA and dietary intake, a future direction could include utilizing an objective measure of PA (e.g., activity tracking device or ac-



celerometer). Studying a well-characterized, racially diverse cohort of pre-adolescents is also a strength, as the current microbiome literature in this age group is limited and rarely includes individuals from different races and ethnicities. Additionally, a wealth of information has been collected on these children from infancy to 10-years of age, which allowed us to adjust for a large set of confounding covariates. Future studies should consider exploring the PA and microbiome across different pediatric age groups to aid in developing a better understanding of how the composition of the human gut microbiome varies across the life course.

Although PA has recently been established as a probable modulator of the gut microbiome, alterations in the abundance of specific taxa and the extent to which specific taxa affect the body at different life stages remain unclear. Moreover, this relationship is likely confounded by several environmental factors, including specific nutrient information, necessitating further studies that can establish more definitive relationships. In summary, this study provides observational support for the importance of regular PA in shaping the gut microbiome during a period of continued growth and development, preadolescence. Specific taxa associated with health were differentially abundant among pre-adolescents who met the PA recommendations. Future studies need to assess whether this translates to a reduced risk of obesity or associated metabolic diseases.

## ACKNOWLEDGEMENTS

We thank the participants of the WHEALS study.

## REFERENCES

- Piercy KL, Troiano RP, Ballard RM, Carlson SA, Fulton JE, Galuska DA, George SM, Olson RD. The physical activity guidelines for Americans. *Jama*. 2018;320:2020-8.
- Katzmarzyk PT, Denstel KD, Beals K, Carlson J, Crouter SE, McKenzie TL, Pate RR, Sisson SB, Staiano AE, Stanish H, Ward DS. Results from the United States 2018 report card on physical activity for children and youth. J Phys Act Health. 2018;15:S422-4.
- Janssen I, LeBlanc AG. Systematic review of the health benefits of physical activity and fitness in school-aged children and youth. *Int. J. Behav. Nutr.* 2010;7:1-6.
- Poitras VJ, Gray CE, Borghese MM, Carson V, Chaput JP, Janssen I, Katzmarzyk PT, Pate RR, Connor Gorber S, Kho ME, Sampson M. Systematic review of the relationships between objectively measured physical activity and health indicators in school-aged children and youth. *Appl Physiol Nutr Metab.* 2016;41:S197-239.
- Quiroga R, Nistal E, Estébanez B, Porras D, Juárez-Fernández M, Martínez-Flórez S, García-Mediavilla MV, de Paz JA, González-Gallego J, Sánchez-Campos S, Cuevas MJ. Exercise training modulates the gut microbiota profile and impairs inflammatory signaling pathways in obese children. *Exp. Mol. Med.* 2020; 52:1048-61.
- Bai J, Hu Y, Bruner D. Composition of gut microbiota and its association with body mass index and lifestyle factors in a cohort of 7–18

years old children from the American gut project. *Pediatric Obes.* 2019;14:e12480.

- Moran-Ramos S, Lopez-Contreras BE, Villarruel-Vazquez R, Ocampo-Medina E, Macias-Kauffer L, Martinez-Medina JN, Villamil-Ramirez H, León-Mimila P, Del Rio-Navarro BE, Ibarra-Gonzalez I, Vela-Amieva M. Environmental and intrinsic factors shaping gut microbiota composition and diversity and its relation to metabolic health in children and early adolescents: a population-based study. *Gut Microbes.* 2020;11:900-17.
- Liang D, Leung RK-K, Guan W, Au WW. Involvement of gut microbiome in human health and disease: brief overview, knowledge gaps and research opportunities. *Gut Pathog.* 2018;10:1-9.
- Koleva PT, Bridgman SL, Kozyrskyj AL. The infant gut microbiome: evidence for obesity risk and dietary intervention. *Nutrients*. 2015;7:2237-60.
- Bernstein CN, Burchill C, Targownik LE, Singh H, Roos LL. Events within the first year of life, but not the neonatal period, affect risk for later development of inflammatory bowel diseases. *Gastroen*terology. 2019;156:2190-97.
- Milani C, Duranti S, Bottacini F, Casey E, Turroni F, Mahony J, Belzer C, Delgado Palacio S, Arboleya Montes S, Mancabelli L. The first microbial colonizers of the human gut: composition, activities, and health implications of the infant gut microbiota. *Microbiol Mol Biol Rev.* 2017;81:e00036-17.
- Johnson CC, Ownby DR. The infant gut bacterial microbiota and risk of pediatric asthma and allergic diseases. *Transl Res.* 2017;179:60-70.
- Roth R, Lynch K, Hyöty H, Lönnrot M, Driscoll KA, Bennett Johnson S, Group TS. The association between stressful life events and respiratory infections during the first 4 years of life: the environmental determinants of diabetes in the young study. *Stress and Health.* 2019;35:289-303.
- Sordillo JE, Korrick S, Laranjo N, Carey V, Weinstock GM, Gold DR, O'Connor G, Sandel M, Bacharier LB, Beigelman A. Association of the infant gut microbiome with early childhood neurodevelopmental outcomes: an ancillary study to the VDAART randomized clinical trial. *JAMA Netw Open.* 2019;2:e190905-e190905.
- Scott KP, Jean-Michel A, Midtvedt T, van Hemert S. Manipulating the gut microbiota to maintain health and treat disease. *Microb Ecol Health Dis.* 2015;26:25877.
- Bokulich NA, Chung J, Battaglia T, Henderson N, Jay M, Li H, Lieber AD, Wu F, Perez-Perez GI, Chen Y, Schweizer W. Antibiotics, birth mode, and diet shape microbiome maturation during early life. *Sci Transl Med.* 2016;8:343-82.
- Yassour M, Vatanen T, Siljander H, Hämäläinen AM, Härkönen T, Ryhänen SJ, Franzosa EA, Vlamakis H, Huttenhower C, Gevers D, Lander ES. Natural history of the infant gut microbiome and impact of antibiotic treatment on bacterial strain diversity and stability. *Sci Transl Med.* 2016;8:343-81.
- Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, Hidalgo G, Baldassano RN, Anokhin AP, Heath AC. Human gut microbiome viewed across age and geography. *Nature*. 2012;486:222-7.
- Aagaard K, Ma J, Antony KM, Ganu R, Petrosino J, Versalovic J. The placenta harbors a unique microbiome. *Sci Transl Med.* 2014; 6:237-65.
- Oh KJ, Lee SE, Jung H, Kim G, Romero R, Yoon BH. Detection of ureaplasmas by the polymerase chain reaction in the amniotic fluid

of patients with cervical insufficiency. *J Perinat Med.* 2010;38:261-68.

- Younes JA, Lievens E, Hummelen R, van der Westen R, Reid G, Petrova MI. Women and their microbes: the unexpected friendship. *Trends Microbiol.* 2018;26:16-32.
- Rackaityte E, Halkias J, Fukui EM, Mendoza VF, Hayzelden C, Crawford ED, Fujimura KE, Burt TD, Lynch SV. Viable bacterial colonization is highly limited in the human intestine in utero. *Nat Med.* 2020;26:599-607.
- Kim H, Sitarik AR, Woodcroft K, Johnson CC, Zoratti E. Birth mode, breastfeeding, pet exposure, and antibiotic use: associations with the gut microbiome and sensitization in children. *Curr Allergy Asthma Rep.* 2019;19:1-9.
- Lozupone CA, Stombaugh JI, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature*. 2012;489:220-30.
- Dicks L, Geldenhuys J, Mikkelsen L, Brandsborg E, Marcotte H. Our gut microbiota: a long walk to homeostasis. *Benef Microbes*. 2018;9:3-20.
- Jenmalm MC. The mother–offspring dyad: microbial transmission, immune interactions and allergy development. *J Intern Med.* 2017;282:484-95.
- Vuillermin PJ, Macia L, Nanan R, Tang ML, Collier F, Brix S. The maternal microbiome during pregnancy and allergic disease in the offspring. *Paper presented at: Seminars in immunopathology* 2017. 2017;39:669-75.
- Gschwendtner S, Kang H, Thiering E, Kublik S, Fösel B, Schulz H, Krauss-Etschmann S, Heinrich J, Schöler A, Schloter M, Standl M. Early life determinants induce sustainable changes in the gut microbiome of six-year-old children. *Sci Rep.* 2019;9:1-9.
- Nagpal R, Yamashiro Y. Gut microbiota composition in healthy Japanese infants and young adults born by C-section. *Ann Nutr Metab.* 2018;73:4-11.
- Martin R, Makino H, Cetinyurek Yavuz A, Ben-Amor K, Roelofs M, Ishikawa E, Kubota H, Swinkels S, Sakai T, Oishi K, Kushiro A. Early-life events, including mode of delivery and type of feeding, siblings and gender, shape the developing gut microbiota. *PLoS One.* 2016;11:e0158498.
- Zhao X, Zhang Z, Hu B, Huang W, Yuan C, Zou L. Response of gut microbiota to metabolite changes induced by endurance exercise. *Front microbiol.* 2018;9:765.
- Barton W. The exercise and diet-microbiome paradigm: influences of physical activity and dietary nutrition on the human gut microbiome. University College Cork; 2018.
- Mailing LJ, Allen JM, Buford TW, Fields CJ, Woods JA. Exercise and the gut microbiome: a review of the evidence, potential mechanisms, and implications for human health. *Exerc Sport Sci Rev.* 2019;47:75-85.
- Shahar RT, Koren O, Matarasso S, Shochat T, Magzal F, Agmon M. Attributes of physical activity and gut microbiome in adults: a systematic review. *Int J Sports Med.* 2020;41:801-14.
- Lambert JE, Myslicki JP, Bomhof MR, Belke DD, Shearer J, Reimer RA. Exercise training modifies gut microbiota in normal and diabetic mice. *Appl Physiol Nutr Metab.* 2015;40:749-52.
- Denou E, Marcinko K, Surette MG, Steinberg GR, Schertzer JD. High-intensity exercise training increases the diversity and metabolic capacity of the mouse distal gut microbiota during diet-induced obesity. *Am J Physiol - Endocrinol Metab.* 2016;310:E982-

93.

- Campbell SC, Wisniewski PJ, Noji M, McGuinness LR, Häggblom MM, Lightfoot SA, Joseph LB, Kerkhof LJ. The effect of diet and exercise on intestinal integrity and microbial diversity in mice. *PLoS One.* 2016;11:e0150502.
- Lamoureux EV, Grandy SA, Langille MG. Moderate exercise has limited but distinguishable effects on the mouse microbiome. *MSystems*. 2017;2:e00006-17.
- Evans CC, LePard KJ, Kwak JW, Stancukas MC, Laskowski S, Dougherty J, Moulton L, Glawe A, Wang Y, Leone V, Antonopoulos DA. Exercise prevents weight gain and alters the gut microbiota in a mouse model of high fat diet-induced obesity. *PLoS One*. 2014;9: e92193.
- Bressa C, Bailén-Andrino M, Pérez-Santiago J, González-Soltero R, Pérez M, Montalvo-Lominchar MG, Maté-Muñoz JL, Domínguez R, Moreno D, Larrosa M. Differences in gut microbiota profile between women with active lifestyle and sedentary women. *PLoS One.* 2017;12:e0171352.
- Dao MC, Everard A, Aron-Wisnewsky J, Sokolovska N, Prifti E, Verger EO, Kayser BD, Levenez F, Chilloux J, Hoyles L, Dumas ME. Akkermansia muciniphila and improved metabolic health during a dietary intervention in obesity: relationship with gut microbiome richness and ecology. *Gut.* 2016;65:426-36.
- Barton W, Penney NC, Cronin O, Garcia-Perez I, Molloy MG, Holmes E, Shanahan F, Cotter PD, O'Sullivan O. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut.* 2018;67:625-33.
- Allen JM, Mailing LJ, Niemiro GM, Moore R, Cook MD, White BA, Holscher HD, Woods JA. Exercise alters gut microbiota composition and function in lean and obese humans. *Med Sci Sports Exerc.* 2018;50:747-57.
- Cronin O, Barton W, Skuse P, Penney NC, Garcia-Perez I, Murphy EF, Woods T, Nugent H, Fanning A, Melgar S, Falvey EC. A prospective metagenomic and metabolomic analysis of the impact of exercise and/or whey protein supplementation on the gut microbiome of sedentary adults. *MSystems*. 2018;3:e00044-18.
- Sitarik AR, Kerver JM, Havstad SL, Zoratti EM, Ownby DR, Wegienka G, Johnson CC, Cassidy-Bushrow AE. Infant feeding practices and subsequent dietary patterns of school-aged children in a US birth cohort. *J Acad Nutr Diet.* 2020:121; 1064-79.
- Williams LK, McPhee RA, Ownby DR, Peterson EL, James M, Zoratti EM, Johnson CC. Gene-environment interactions with CD14 C-260T and their relationship to total serum IgE levels in adults. *J Allergy Clin Immunol.* 2006;118:851-7.
- Wegienka G, Havstad S, Joseph CL, Zoratti E, Ownby D, Woodcroft K, Johnson CC. Racial disparities in allergic outcomes in African Americans emerge as early as age 2 years. *Clin Exp Allergy.* 2012;42:909-17.
- Serdula MK, Ivery D, Coates RJ, Freedman DS, Williamson DF, Byers T. Do obese children become obese adults? A review of the literature. *Prev Med.* 1993;22:167-77.
- Fujimura KE, Sitarik AR, Havstad S, Lin DL, Levan S, Fadrosh D, Panzer AR, LaMere B, Rackaityte E, Lukacs NW. Neonatal gut microbiota associates with childhood multisensitized atopy and T cell differentiation. *Nat Med.* 2016;22:1187-91.
- 50. DeAngelis KM, Brodie EL, DeSantis TZ, Andersen GL, Lindow SE, Firestone MK. Selective progressive response of soil microbial





community to wild oat roots. ISME J. 2009;3:168-78.

- Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Huntley J, Fierer N, Owens SM, Betley J, Fraser L, Bauer M. Ultra-high-throughput microbial community analysis on the Illumina HiSeq and MiSeq platforms. *ISME J*. 2012;6:1621-4.
- Magoč T, Salzberg SL. FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics*. 2011; 27:2957-63.
- Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JI. QIIME allows analysis of high-throughput community sequencing data. *Nat Methods*. 2010;7:335-6.
- Edgar RC. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*. 2010;26:2460-1.
- Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics.* 2011;27:2194-200.
- Edgar RC. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat Methods*. 2013;10:996-8.
- DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, Huber T, Dalevi D, Hu P, Andersen GL. Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol.* 2006;72:5069-72.
- Caporaso JG, Bittinger K, Bushman FD, DeSantis TZ, Andersen GL, Knight R. PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinformatics*. 2010;26:266-7.
- Price MN, Dehal PS, Arkin AP. FastTree 2–approximately maximum-likelihood trees for large alignments. *PLoS One.* 2010;5: e9490.
- NutritionQuest. Block kids physical activity screener. https://nutritionquest.com/company/our-research-questionnaires/. Accessed December 23, 2020.
- Hamilton CM, Strader LC, Pratt JG, Maiese D, Hendershot T, Kwok RK, Hammond JA, Huggins W, Jackman D, Pan H, Nettles DS. The PhenX Toolkit: get the most from your measures. *Am J Epidemiol.* 2011;174:253-60.
- 62. Kuczmarski RJ. 2000 CDC growth charts for the United States: methods and development. *Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Health Statistics*; 2002.
- Hunsberger M, O'Malley J, Block T, Norris JC. Relative validation of block kids food screener for dietary assessment in children and adolescents. *Matern Child Nutr.* 2015;11:260-70.
- 64. Petersen AC, Crockett L, Richards M, Boxer A. A self-report measure of pubertal status: reliability, validity, and initial norms. *J Youth Adolesc.* 1988;17:117-33.
- Lozupone C, Lladser ME, Knights D, Stombaugh J, Knight R. UniFrac: an effective distance metric for microbial community comparison. *ISME J.* 2011;5:169-72.
- Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara RB, Simpson GL, Solymos P, Henry M, Stevens H, Szoecs E, Wagner H. Vegan: Community Ecology Package. *R package version 2.5-6.* 2019.
- McMurdie PJ, Holmes S. Phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. *PLoS One*. 2013;8:e61217.
- Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.*

2014;15:1-21.

- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol.* 1995:289-300.
- VanderWeele TJ. Principles of confounder selection. *Eur J Epidemiol.* 2019;34:211-9.
- Mancabelli L, Milani C, Lugli GA, Turroni F, Cocconi D, van Sinderen D, Ventura M. Identification of universal gut microbial biomarkers of common human intestinal diseases by meta-analysis. *FEMS Microbiol. Ecol.* 2017;93:fix153.
- Waters JL, Ley RE. The human gut bacteria Christensenellaceae are widespread, heritable, and associated with health. *BMC Biol.* 2019;17:83.
- Mazier W, Le Corf K, Martinez C, Tudela H, Kissi D, Kropp C, Coubard C, Soto M, Elustondo F, Rawadi G. A new strain of Christensenella minuta as a potential biotherapy for obesity and associated metabolic diseases. *Cells*. 2021;10:823.
- Li X, Li Z, He Y, Li P, Zhou H, Zeng N. Regional distribution of Christensenellaceae and its associations with metabolic syndrome based on a population-level analysis. *PeerJ*. 2020;8:e9591.
- Huang J, Liao J, Fang Y, Deng H, Yin H, Shen B, Hu M. Six-week exercise training with dietary restriction improves central hemodynamics associated with altered gut microbiota in adolescents with obesity. *Front Endocrinol.* 2020;11.
- Van Hul M, Le Roy T, Prifti E, Dao MC, Paquot A, Zucker J-D, Delzenne NM, Muccioli G, Clément K, Cani PD. From correlation to causality: the case of Subdoligranulum. *Gut Microbes*. 2020;12:1-13.
- Palmas V, Pisanu S, Madau V, Casula E, Deledda A, Cusano R, Uva P, Vascellari S, Loviselli A, Manzin A, Velluzzi F. Gut microbiota markers associated with obesity and overweight in Italian adults. *Sci Rep.* 2021;11:1-14.
- Tung Y-T, Hsu Y-J, Liao C-C, Ho S-T, Huang C-C, Huang W-C. Physiological and biochemical effects of intrinsically high and low exercise capacities through multiomics approaches. *Front Physiol.* 2019;10:1201.
- Matthies C, Evers S, Ludwig W, Schink B. Anaerovorax odorimutans gen. nov., sp. nov., a putrescine-fermenting, strictly anaerobic bacterium. *Int J Syst Evol Microbiol.* 2000;50:1591-4.
- Gao Z, Yin J, Zhang J, Ward RE, Martin RJ, Lefevre M, Cefalu WT, Ye J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes*. 2009;58:1509-17.
- Shankar V, Gouda M, Moncivaiz J, Gordon A, Reo NV, Hussein L, Paliy O. Differences in gut metabolites and microbial composition and functions between Egyptian and US children are consistent with their diets. *Msystems*. 2017;2.
- 82. Carbajo-Pescador S, Porras D, García-Mediavilla MV, Martínez-Flórez S, Juarez-Fernández M, Cuevas MJ, Mauriz JL, González-Gallego J, Nistal E, Sánchez-Campos S. Beneficial effects of exercise on gut microbiota functionality and barrier integrity, and gut-liver crosstalk in an in vivo model of early obesity and non-alcoholic fatty liver disease. *DMM*. 2019;12:dmm039206.
- Motiani KK, Collado MC, Eskelinen JJ, Virtanen KA, Löyttyniemi E, Salminen S, Nuutila P, Kalliokoski KK, Hannukainen JC. Exercise training modulates gut microbiota profile and improves endotoxemia. *Med Sci Sports Exerc.* 2020;52:94.
- 84. Gerard M, Fossa A, Folcarelli PH, Walker J, Bell SK. What patients value about reading visit notes: a qualitative inquiry of patient ex-



periences with their health information. *J Med Internet Res.* 2017; 19:e237.

 Yeager A. Exercise Changes Our Gut Microbes, But How Isn't Yet Clear. https://www.the-scientist.com/news-opinion/exercise-changes-our-gut-microbes--but-how-isnt-yet-clear-66281. Published 2019. Accessed June 28 2020.