

ORIGINAL ARTICLE

Population Pharmacokinetics and Antimalarial Pharmacodynamics of Piperaquine in Patients With *Plasmodium vivax* Malaria in Thailand

J Tarning^{1,2}, P Thanan¹, AP Phyo³, KM Lwin³, W Hanpithakpong¹, EA Ashley^{1,2,3}, NPJ Day^{1,2}, F Nosten^{1,2,3} and NJ White^{1,2}

Dihydroartemisinin-piperaquine is an effective drug in the treatment of *Plasmodium falciparum* and *P. vivax* malaria. The objective of this study was to evaluate the population pharmacokinetics and pharmacodynamics of piperaquine in patients with *P. vivax* malaria in Thailand after a standard regimen of dihydroartemisinin-piperaquine to determine whether residual piperaquine prevents or delays the emergence of *P. vivax* relapse. Sparse blood samples were collected from 116 patients. Piperaquine pharmacokinetics were described well by a three-compartment distribution model. Relapsing *P. vivax* malaria was accommodated by a constant baseline hazard (8.94 relapses/year) with the addition of a surge function in a fixed 3-week interval and a protective piperaquine effect. The results suggest that a large proportion of the first relapses were suppressed completely by residual piperaquine concentrations and that recurrences resulted mainly from emergence of the second or third relapse or from reinfection. This suggests a significant reduction in *P. vivax* morbidity when using dihydroartemisinin-piperaquine compared with other antimalarial drugs with shorter terminal postprophylactic effects.

CPT Pharmacometrics Syst. Pharmacol. (2014) 3, e132; doi:10.1038/psp.2014.29; published online 27 August 2014

Plasmodium vivax is the most difficult of the five species of human malaria parasites to eliminate. It is the most geographically widespread human malaria parasite, causing an estimated 130–390 million cases every year.¹ *P. vivax* is transmitted within 95 countries in America, Africa, and Asia.^{2,3} In total, 2.85 billion people were estimated to be at risk of *P. vivax* transmission in 2009, especially in Central and Southeast Asia, where more than 90% of the world's *P. vivax* transmission occurs.³ Acute *P. vivax* malaria is rarely life threatening, but it causes substantial morbidity in higher transmission areas as a result of multiple relapses from latent liver-stage parasites.⁴ These repeated infections may result in life-threateningly severe anemia due to destruction of both infected and non-infected red blood cells. *P. vivax* is an important cause of low birth weight.^{4–7}

Chloroquine, a blood schizonticide, is used in first-line treatment and prophylaxis of *P. vivax* malaria in most endemic areas. However, *P. vivax* resistance to chloroquine, which was first observed in Papua New Guinea in 1989,⁸ is increasing in many parts of the world.⁹ A combination of two drugs with different mechanisms of action is preferred for the treatment of malaria to reduce the risk of the development of resistant parasite strains. The fixed combination of dihydroartemisinin-piperaquine (DHA-PQ) is recommended by the World Health Organization as the first-line treatment of uncomplicated *P. falciparum* malaria¹⁰ and has replaced chloroquine in the first-line treatment of resistant *P. vivax* malaria in Indonesia. DHA-PQ is currently available as a fixed-dose combination, administered once daily over 3 days. A meta-analysis pooled from 12 different studies in 6 countries between 2003 and 2006 showed high efficacy against *P. falciparum* malaria (polymerase chain reaction-corrected cure rates of 98.7% at

day 28) and good tolerance (4.8% total incidence of early vomiting) of DHA-PQ.¹¹ DHA-PQ also has excellent efficacy against *P. vivax* malaria.^{12–14}

Piperaquine is distributed extensively into tissues, resulting in multiphasic distribution and a slow elimination from the systemic circulation (elimination half-life ($t_{1/2}$) of ~18–28 days). Similar pharmacokinetic properties have been reported in children but these pediatric studies suggest that small children need a higher weight-based dose to achieve exposure comparable to that in older children.¹⁵ Most piperaquine pharmacokinetic studies have sampled venous plasma. However, capillary sampling is a desirable alternative for field pharmacokinetic studies, especially in children. The population pharmacokinetics and pharmacodynamics of piperaquine have not been reported previously in patients with *P. vivax* malaria.

In South East Asia and Oceania, *P. vivax* relapses at 3-week intervals. Treatment with efficacious slowly eliminated antimalarials results in substantially delayed emergence of the relapse, but it has been unclear whether the first relapse is prevented (and so the second relapse is observed) or delayed. The aim of this study was to characterize the population pharmacokinetics and antimalarial pharmacodynamics of piperaquine in the treatment of *P. vivax* malaria and to use this information to determine whether piperaquine prevents or delays the first *P. vivax* relapse.

RESULTS

Safety and efficacy

This study was conducted at Shoklo Malaria Research Unit clinics, Mae Sot, Thailand, an area of low and seasonal malaria transmission located at the Thailand–Myanmar

¹Mahidol-Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand; ²Centre for Tropical Medicine, Nuffield Department of Medicine, University of Oxford, Oxford, UK; ³Shoklo Malaria Research Unit, Mahidol-Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, Tak, Thailand. Correspondence: J Tarning (joel@tropmedres.ac)

Received 8 April 2014; accepted 19 May 2014; published online 27 August 2014. doi:10.1038/psp.2014.29

Table 1 Clinical outcomes and patient demographics of patients with *Plasmodium vivax* malaria enrolled in this study

| | Efficacy study | PK cohort ^a | PK-PD cohort ^b |
|---|-------------------|------------------------|---------------------------|
| Total number of patients | 241 | 116 | 62 |
| Total number of venous/capillary samples | — | 435/356 | 386/312 |
| Total dose of piperaquine base (mg/kg) | 30.8 (23.1–46.2) | 30.1 (27.2–36.1) | 30.1 (27.2–36.1) |
| Continuous and categorical admission covariates | | | |
| Age (years) | 18 (1–57) | 23.5 (3–57) | 29 (11–56) |
| Height (cm) | 154 (65–179) | 160 (88–176) | 160 (115–176) |
| Body weight (kg) | 45 (7–74) | 50 (9–67) | 50 (23–67) |
| Body temperature (°C) | 37 (36–41) | 37 (36–40.4) | 37 (36–40.1) |
| Days of fever before admission | 2 (0–8) | 2 (1–8) | 2 (1–7) |
| Hematocrit (%) | 37 (22–50) | 39 (22–50) | 39 (27–49) |
| Initial parasitemia (μl^{-1}) | 6,106 (80–70,255) | 7,185 (80–58,714) | 6,802 (80–45,166) |
| Gametocytomia (yes/no) | 165/76 | 65/51 | 34/28 |
| Sex (male/female) | 169/72 | 90/26 | 25/37 |
| Treatment outcomes ^c | | | |
| Number of patients with no recurrent malaria | 120 | 31 | 31 |
| Number of patients with recurrent <i>P. vivax</i> malaria | 109 | 75 | 31 |
| Time to recurrent <i>P. vivax</i> malaria (days) | 56 (14–65) | 56 (21–65) | 56 (35–65) |
| Parasitemia at recurrent <i>P. vivax</i> malaria (μl^{-1}) | 352 (16–39,144) | 735 (16–39,144) | 735 (16–31,617) |
| Number of patients with recurrent <i>P. falciparum</i> malaria | 10 | 8 | — |
| Time to recurrent <i>P. falciparum</i> malaria (days) | 37 (28–49) | 40 (35–49) | — |
| Number of patients with recurrent mixed malaria | 2 | 2 | — |
| Time to recurrent mixed malaria (days) | 42 (35–49) | 42 (35–49) | — |

All values are given as median (range) unless otherwise specified.

PK-PD, pharmacokinetics/pharmacodynamics.

^aBased on 62 patients enrolled for pharmacokinetic modeling and an additional 54 patients who provided a single plasma sample only at the time of recurrent malaria.

^bBased on 62 patients for whom both pharmacokinetic and pharmacodynamic data were available. ^cAs confirmed by polymerase chain reaction genotyping.

border. This pharmacokinetic study was nested into a larger efficacy trial in 500 patients with *P. vivax* malaria infections. Full clinical details have been published elsewhere and DHA-PQ was found to be an effective alternative treatment with a lower cumulative risk of recurrent *P. vivax* malaria at 9 weeks compared with chloroquine (54.9 vs. 79.1%).¹⁶ Two-hundred and fifty Karen and Burmese patients with *P. vivax* malaria infections were enrolled in the DHA-PQ-treated arm of the efficacy study (Table 1). All patients received DHA-PQ (Duo-cotecxin, Beijing Holley-Cotec Pharmaceuticals, China) tablets containing 40mg of dihydroartemisinin and 320mg of piperaquine phosphate. A standard weight-based regimen (2.2mg/kg/day of dihydroartemisinin and 17.8mg/kg/day of piperaquine phosphate) was administered with milk once daily for 3 days. The fixed combination of DHA-PQ was well tolerated with no serious adverse events reported during follow-up. In this population modeling, nine patients were excluded: two patients did not meet the inclusion criteria, five patients had an incomplete course of medication, one patient self-medicated with other drugs, and one patient withdrew consent. Overall, 45% (109/241) of patients presented with recurrent *P. vivax* malaria during the follow-up period at a median (range) of 56 (14–65) days (Table 1).

Pharmacokinetics of piperaquine

A total of 791 plasma concentrations (435 venous and 356 capillary samples) from 116 patients were quantified and used in the pharmacokinetic modeling. Venous and capillary plasma concentrations were transformed into their natural logarithms and modeled simultaneously using

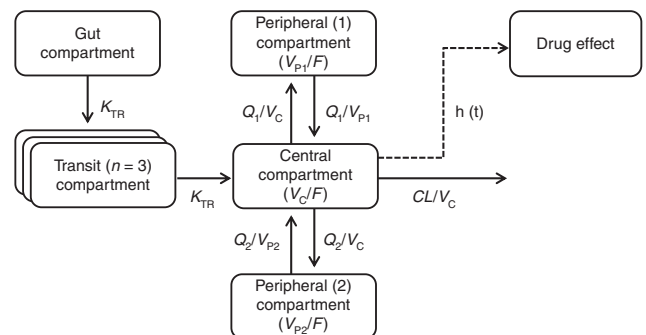


Figure 1 Final piperaquine population pharmacokinetic-pharmacodynamic model in patients with *P. vivax* malaria. K_{TR} is the transit absorption rate constant ($K_{TR} = (n + 1)/\text{mean absorption transit time}$), CL is the elimination clearance, V_C is the apparent central volume of distribution, V_p is the apparent peripheral volume of distribution, Q is the intercompartment clearance, and F is the relative oral bioavailability.

nonlinear mixed-effects modeling. Substantial differences in matched venous and capillary blood concentrations have been reported in a previous clinical study.¹⁷ A proportional venous-capillary transformation factor was, therefore, used to compensate for the difference between sampling matrices, resulting in 41% higher capillary concentrations compared with venous concentrations.

The initial structural base models were parameterized as elimination clearance (CL/F), intercompartment clearance(s) (Q/F), apparent volume of distribution of the central compartment (V_C/F), apparent volume of distribution of the

Table 2 Population pharmacokinetic-pharmacodynamic parameters of the final model describing piperazine in patients with *Plasmodium vivax* malaria

| Parameter estimates | Venous estimates | | | Capillary estimates |
|------------------------------------|--|-----------------------------------|---|-----------------------------|
| | Population estimates ^a (%RSE ^b) | 95% CI ^b for estimates | %CV ^a for IIV (%RSE ^b) | 95% CI ^b for IIV |
| Pharmacokinetics parameters | | | | |
| CL/F (l/h) | 54.1 (5.56) | 48.0–59.7 | — | — |
| V_c/F (l) | 2,770 (10.4) | 2,209–3,298 | — | — |
| Q_1/F (l/h) | 663 (19.7) | 475–1,023 | 138 (32.2) | 75.0–222 |
| V_{p1}/F (l) | 10,600 (10.7) | 8,549–12,962 | 36.5 (40.2) | 18.8–51.7 |
| Q_2/F (l/h) | 72.7 (12.1) | 56.3–93.5 | — | — |
| V_{p2}/F (l) | 25,900 (12.8) | 21,336–33,547 | 45.2 (71.0) | 0.431–82.5 |
| MTT (h) | 1.93 (9.16) | 1.60–2.30 | 54.5 (38.4) | 33.6–80.0 |
| F (%) | 1 fixed | — | 46.5 (18.9) | 36.0–55.1 |
| TF | 1.41 (2.11) | 1.35–1.46 | — | — |
| σ_1 | 0.270 (5.59) | 0.239–0.299 | — | — |
| σ_2 | 0.321 (5.79) | 0.283–0.354 | — | — |
| Pharmacodynamics parameters | | | | |
| θ_{BHZ} (relapses/year) | 8.94 (22.1) | 4.80–11.7 | — | — |
| θ_{SA} (%) | 123 (23.7) | 61.8–165 | — | — |
| θ_{SW} (h) | 49.5 (54.3) | 35.7–80.4 | — | — |
| θ_{SHP} | 4 fixed | — | — | — |
| PC_{50} (ng/ml) | 6.92 (15.5) | 6.15–11.3 | — | — |
| Γ | 9.28 (25.6) | 5.09–14.0 | — | — |

CL/F , elimination clearance; F , relative oral bioavailability; MTT, mean transit absorption time; V_c/F , apparent central volume of distribution; Q_1/F , intercompartmental clearance(s); TF, venous-to-capillary transformation factor; V_{p1}/F , apparent peripheral volume(s) of distribution; σ_1 , residual error of piperazine capillary concentrations; σ_2 , residual error of piperazine venous concentrations; θ_{BHZ} , baseline hazard; θ_{SA} , amplitude of surge function; θ_{SW} , width of surge function; θ_{SHP} , shape factor of surge function; PC_{50} , 50% protective venous concentration for 50% reduction in baseline hazard; Γ , shape factor of E_{max} function.

^aComputed population mean parameter estimates from NONMEM. Coefficient of variation (%CV) for interindividual variability is calculated as $100 \times (\text{EXP}(\text{mean estimate}) - 1)^{1/2}$. ^bRelative standard errors (RSE) are calculated as $100 \times \text{standard deviation}/\text{mean value}$ from the nonparametric bootstrap method of the final pharmacokinetic model ($n = 1,000$) and pharmacodynamic model ($n = 500$). The 95% confidence intervals (95% CI) are displayed as the 2.5–97.5 percentiles of the bootstrap estimates.

Table 3 Secondary parameters of the final piperazine model in patients with *Plasmodium vivax* malaria

| | Venous plasma | Capillary plasma |
|-------------------------------------|-----------------------|-------------------------|
| C_{max} (ng/ml) | 162 (50.6–457) | 228 (71.3–645) |
| T_{max} (h) | 3.0 (1.6–9.3) | 3.0 (1.6–9.3) |
| CL/F (l/h/kg) | 1.08 (1.01–1.66) | 0.770 (0.716–1.18) |
| V_d/F (l/kg) | 802 (455–1,451) | 572 (324–1,033) |
| $t_{1/2}$ (days) | 28.8 (15.5–58.3) | 28.9 (15.5–58.4) |
| $AUC_{0-\infty}$ (h \times ng/ml) | 27,160 (9,443–78,470) | 38,250 (13,289–110,510) |
| Day 7 (ng/ml) | 33.0 (11.4–95.2) | 46.5 (16.0–134) |

All values are given as median (range) unless otherwise specified. Secondary parameter estimates are calculated from the empirical Bayesian *post hoc* estimates.

$AUC_{0-\infty}$, area under the concentration–time curve from time 0 to infinity; C_{max} , maximum concentration; CL/F , elimination clearance; day 7, predicted concentration at day 7; T_{max} , time to maximum concentration; $t_{1/2}$, terminal elimination half-life; V_d/F , total volume of distribution.

peripheral compartment(s) (V_p/F), and absorption rate constant. Several absorption, distribution, and covariate models were fitted to the data to construct the best performing model. Piperazine pharmacokinetics in patients with *P. vivax* malaria were best described by a three-compartment distribution model (Figure 1). A three-compartment disposition model resulted in a significant improvement in model fit compared with a two-compartment model (difference in objective function value (ΔOFV) of 108) with no additional benefit of an additional peripheral compartment ($\Delta\text{OFV} = 0.57$). A transit-compartment model with a fixed number of transit

compartments ($n = 3$, transit rate constant was set to be identical to absorption rate constant) described the absorption phase better than all other absorption models ($\Delta\text{OFV} > 54.2$). A relative bioavailability parameter (i.e., fixed to 100% for the population) was implemented to allow quantification of the interindividual variability in the absorption of piperazine and resulted in a significant improvement in the model fit ($\Delta\text{OFV} = 81.8$).

Body weight, incorporated as a fixed allometric function, improved the model fit significantly ($\Delta\text{OFV} = 31.0$) and resulted in better precision and decreased interindividual variability in clearance and volume parameters. Body temperature on admission on V_{p2}/F ($\Delta\text{OFV} = 11.5$) and age on Q_1/F ($\Delta\text{OFV} = 5.46$) were selected in the forward covariate selection but could not be retained in the backward deletion. The estimated interindividual variability was small for the transformation factor, CL/F , Q_2/F , and V_c/F , and could, therefore, be removed with no significant impact in the final model ($\Delta\text{OFV} = 2.40$). Final pharmacokinetic parameter estimates and secondary parameter estimates are reported in Tables 2 and 3.

The mean parameter estimates from the simultaneous model with both venous and capillary data were also comparable with estimates when capillary and venous data were modeled separately. The final pharmacokinetic model displayed satisfactory goodness-of-fit diagnostics (Supplementary Figure S1 online) with a small epsilon-shrinkage of 15.0%. However, a relatively high η -shrinkage (up to 43.5%) could be seen for certain parameters in the

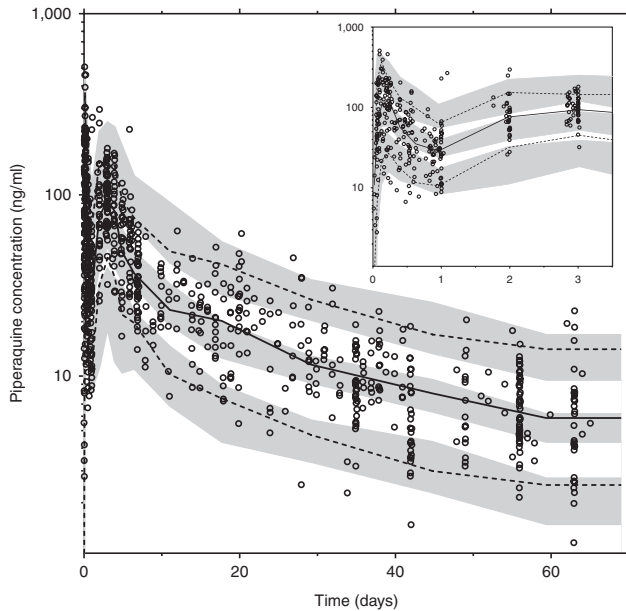


Figure 2 Prediction-corrected visual predictive check of the final model describing piperaquine pharmacokinetics in *P. vivax* malaria. Open circles represent the observed piperaquine concentrations. Solid black line represents the 50th percentile of the observations, and dash lines represent the 5th and 95th percentiles of the observations. Gray areas represent the 95% confidence intervals of the simulated 5th, 50th, and 95th percentiles from 1,000 simulations. The inset shows a prediction-corrected visual predictive check for the first 3 days of piperaquine treatment.

final model because of the sparse data. The numerical and prediction-corrected visual predictive checks resulted in 4.6% (95% CI: 2.7–8.1%) and 3.7% (95% CI: 2.5–8.2%) of piperaquine observations below and above the simulated 90% prediction interval, respectively (Figure 2).

Pharmacodynamics of piperaquine

Piperaquine inhibits asexual parasite multiplication but has no effect on liver-stage parasites. The influence of piperaquine exposure on the risk of recurrent *P. vivax* malaria was investigated in 62 patients for whom both pharmacokinetic and pharmacodynamic data were available. However, efficacy data were available from all the 241 patients included in the study. Pharmacodynamic parameters were evaluated using an interval-censoring time-to-event model,¹⁸ implemented with the Laplacian estimation method. The pharmacokinetic parameter and variability estimates were fixed to that of the final pharmacokinetic model and used in the pharmacokinetic-pharmacodynamic model. *P. vivax* malaria in tropical regions commonly displays frequent relapses which emerge at three-week intervals when the infections are treated with rapidly eliminated antimalarials.¹⁹ A multiple surge function was, therefore, implemented to characterize this periodicity in the risk of recurrent malaria. The constant hazard function with a periodically increased risk of relapses described the data well. Piperaquine had a significant ($\Delta\text{OFV} = 20.3$) protective effect on the risk of recurrent malaria infections with a required *in vivo* venous piperaquine concentration of 6.92 ng/ml for a 50% relative reduction in baseline hazard (PC_{50}). Sex was selected as a significant covariate

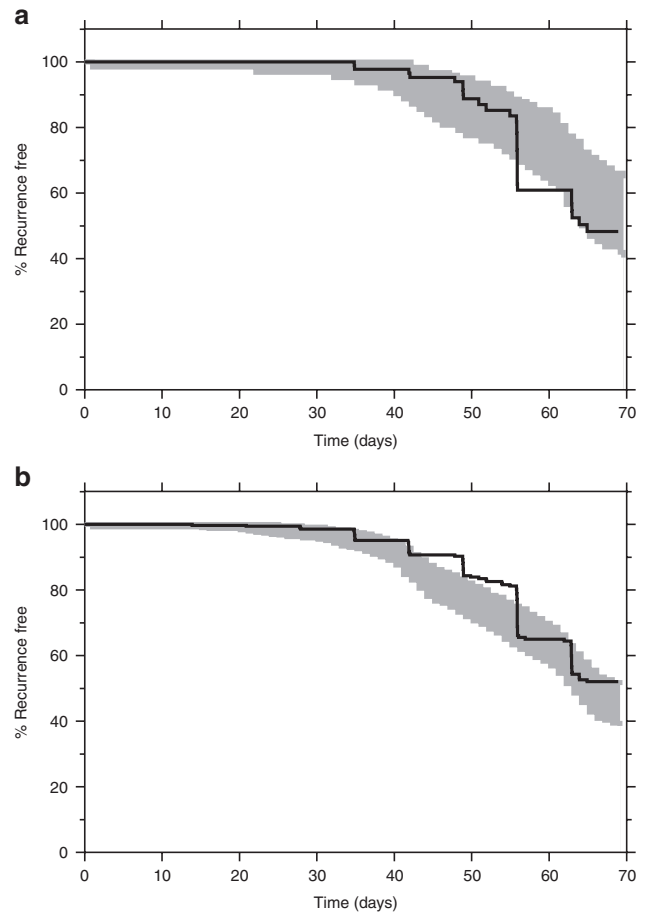


Figure 3 Visual predictive check of the final time-to-event model describing piperaquine pharmacodynamics in *P. vivax* malaria. Gray area represents the 95% prediction interval of the time to recurrent infections from 500 simulations. Solid line represents the observed Kaplan–Meier plot for (a) patients in the pharmacokinetic and pharmacodynamic cohort (62 patients, Table 1) and (b) all patients in the efficacy study (241 patients, Table 1).

($P < 0.05$) on baseline hazard in the forward selection step, but it was removed during the backward elimination resulting in a covariate-free pharmacodynamic model. Final pharmacodynamic parameters were well estimated with high precision (Table 2) and simulation-based diagnostics showed good agreement between simulated and observed recurrent malaria infections (Figure 3).

The individual patient parasite burden (Table 1) corresponded approximately to a total of 10^{11} parasites in a typical adult at admission. This assumes that *P. vivax* does not sequester substantially. DHA-PQ is administered for 3 consecutive days, covering two asexual cycles. Assuming a parasitocidal effect (parasite reduction ratio) of a 10,000-fold parasite reduction per cycle by dihydroartemisinin (without any additional effect by piperaquine) results in a 100,000,000-fold reduction in total parasite biomass during the first two cycles.²⁰ At an initial parasite burden of 10^{11} parasites, this would leave $\sim 1,000$ parasites to be eliminated by residual piperaquine concentrations to prevent recrudescence malaria. Piperaquine alone has not been adequately assessed in *P. vivax* malaria, but if it is similar to chloroquine, it should

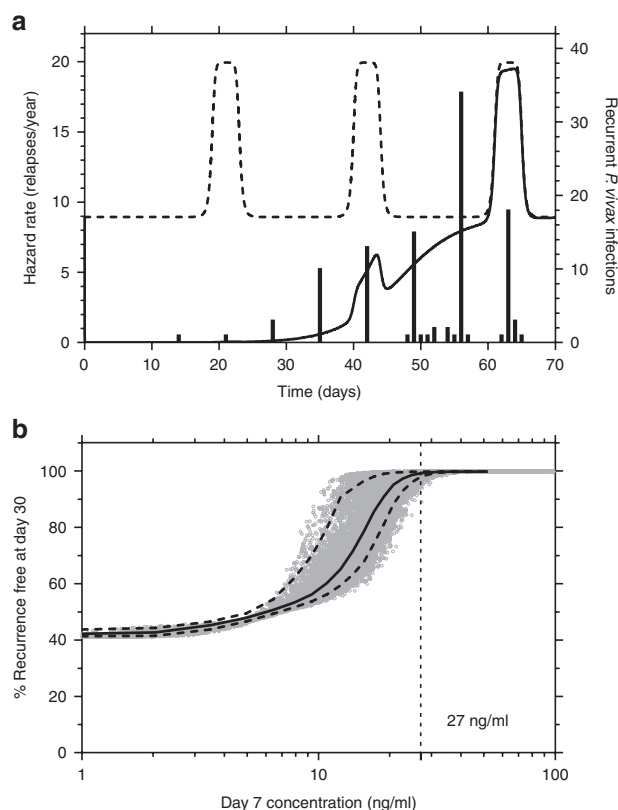


Figure 4 Relationship between piperazine treatment and risk of relapsing *P. vivax* malaria infections. **(a)** Mean hazard rate of relapsing *P. vivax* malaria versus time in the presence (solid line) and absence (dashed line) of a standard piperazine treatment (primary y-axis). The frequency of observed recurrent *P. vivax* infections for all patients in the efficacy study (241 patients, [Table 1](#)) is displayed on the secondary y-axis. The hazard of the first relapse (3 weeks) is completely suppressed and the hazard of the second relapse (6 weeks) is almost completely suppressed after piperazine treatment. The model-predicted hazard rate after piperazine treatment is in agreement with the observed frequency of observed recurrent *P. vivax* infections. **(b)** Simulated day-7 venous piperazine plasma concentrations and chance of remaining malaria free for 30 days. Open gray circles represent simulated patients. Solid black line represents the 50th percentile and dash lines represent the 5th and 95th percentiles of the prediction interval. The dashed vertical line indicates the predicted day-7 concentration that results in 99% chance of remaining malaria free for 30 days for a typical patient.

provide parasite reduction ratios of ~100 to 1,000 per cycle, so most patients should be parasite free 6–8 days post-treatment if piperazine levels are sufficient to sustain a maximum parasitocidal effect for this time. If the parasite burden is not eliminated completely, then these remaining parasites can multiply and cause a recrudescence infection. However, a high cure rate of the asexual infection is likely in this study as concentrations of piperazine after the second asexual cycle (i.e., ≥ 4 days) are highly likely to be able to eliminate ~1,000 parasites. In the first relapse of tropical *P. vivax* malaria (unaffected by drugs and assuming 1–10 schizonts and a subsequent parasite multiplication rate of ~10 per cycle), the hypnozoite-derived hepatic schizonts must liberate 13–17 days after the onset of the primary illness to produce a symptomatic infection at day 21. However, if residual piperazine concentrations

are sufficiently high, they could suppress and eliminate the asexual parasites derived from 10,000 to 100,000 released merozoites and thereby prevent the first relapse completely. If residual piperazine concentrations are not sufficiently high for complete elimination, they could cause an initial suppression, resulting in a delayed symptomatic first relapse. Only 14% (15 out of 109) of recurrent *P. vivax* infections occurred before day 39, which suggests that DHA-PQ treatment prevents a large proportion of the first relapses ([Figure 4](#)). The majority of infections (64%, 70 out of 109) occurred between day 42 and 56, suggesting that these are mainly delayed second relapses ([Figure 4](#)). There will be some contribution from reinfections, but as entomological inoculation rates are probably well below 0.5/person/year for *P. vivax* in this area, this contribution is likely to be small.

DISCUSSION

The fixed combination of DHA-PQ is a highly efficacious antimalarial drug with an excellent safety profile. Antimalarial treatment with DHA-PQ resulted in lower cumulative risk (54.9%) of recurrent *P. vivax* malaria compared with chloroquine (79.1%) in Thailand.¹⁶ This difference could be a result of the very potent artemisinin derivative which generally has a rapid onset of action against *P. vivax* malaria,²¹ but it is likely to result mainly from increasing chloroquine resistance and hence the superiority of piperazine. In fully sensitive *P. vivax* malaria, recrudescence rates are close to zero with chloroquine treatment. Previous studies in this region have shown high relapse rates of *P. vivax* within one month of treatment with a short acting artemisinin derivative.¹⁹ Thus, both slowly eliminated drugs prevent the first relapse. Whether the lower cumulative risk of recurrent *P. vivax* malaria, following DHA-PQ, results from a higher cure rate of the primary infection or a more effective suppression of the first or the second relapse has been unclear. As relapses can arise from parasites which are either genetically similar or genetically distinct to those causing the primary infection, it is not possible to distinguish with certainty between relapse, recrudescence, or reinfection in patients with *P. vivax* malaria who remain in the endemic area. Recrudescence can occur many weeks after treatment with slowly eliminated drugs, but the pharmacokinetic-pharmacodynamic data modeled in this study suggest that only a negligible proportion (<2%) of the recurrent *P. vivax* infections could be recrudescence.

Pharmacokinetics of piperazine

The population pharmacokinetic properties of piperazine have been described previously in patients with uncomplicated *P. falciparum* malaria.^{15,22–24} This is the first study reporting the population pharmacokinetics of piperazine in patients with *P. vivax* malaria.

The structural three-compartment model described in this study was similar to that reported recently in young children and in pregnant and nonpregnant women with *P. falciparum* malaria.^{15,22,25} Simplified two-compartment structural models in children and adult patients have also been reported reflecting differences in follow-period and/or sampling strategies.^{23,24} A transit-compartment absorption model allowed more

flexibility in the absorption phase and provided a clear advantage over other absorption models, although there were not enough data in the absorption phase to estimate both K_{TR} and K_A separately. Large interindividual variability was seen in the absorption of piperazine as in previous studies.^{15,22,25} Data collected here were too sparse to allow an estimation of the variability between dose occasions, which is likely to have inflated the inter-individual variability. Indeed interindividual variability was high in this study with values up to 138%. Venous and capillary plasma concentrations were successfully modeled simultaneously with a transformation factor of 1.41 (relative standard error of 2.11%). This factor was similar to that from a linear regression between observed venous and capillary plasma concentrations (slope = 1.29), which further supports the appropriateness of the simultaneous model. However, the exact mechanism of this matrix-dependent difference cannot be elucidated from the data collected in this study. The advantage of a simultaneous modeling approach is that the model can be used to simulate drug exposures from any sampling technique and therefore enables literature comparisons.

Body weight was the only significant covariate in this model. This has also been reported in previous studies^{15,24} and was not unexpected, considering the strong biological prior of body weight as a covariate on pharmacokinetic disposition and elimination parameters.²⁶ All pharmacokinetic parameters were estimated with good precision (relative standard error < 20%) and the simulation-based diagnostics indicated excellent predictive performance of the final pharmacokinetic model.

Piperazine CL/F in this *P. vivax* study was 1.08 l/h/kg, total volume of distribution (V_D/F) was 802 l/kg, and $t_{1/2}$ was 28.8 days, which were comparable with previously published reports in patients with an uncomplicated *P. falciparum* malaria (mean (range) CL/F of 1.18 (0.90–1.4) l/h/kg, V_D/F of 789 (574–877) l/kg, and $t_{1/2}$ of 23.7 (18–28) days).^{22–24,27,28}

Pharmacodynamics of piperazine

The observed high relapse rate of *P. vivax* malaria was expected as DHA-PQ eliminates only blood-stage parasites and does not provide cure of the latent liver-stage parasites. It was expected that piperazine would delay the time to relapse, as does chloroquine, and this was supported by the pharmacodynamic results. Tropical frequent relapsing strains of *P. vivax* typically relapse at 3–4-week intervals, and the median time to relapse approximates three weeks (21 days) in Thailand after treatment with rapidly eliminated antimalarial drugs (such as artesunate, quinine, and halofantrine).¹⁹ In an area of much higher transmission in Papua, Indonesia, there were significantly more early recurrences of *P. vivax* malaria following artemether-lumefantrine treatment of *P. vivax* malaria compared with DHA-PQ (cumulative risk of 38 vs. 10% at day 42)¹³ reflecting the failure of the more rapidly eliminated lumefantrine to suppress the first relapse. However, with longer follow-up in a high transmission area, the cumulative reinfection rates converge as everyone is reinfected eventually. A critical and unanswered question has been whether the slowly eliminated drugs prevented, or simply delayed, the relapse. Explicitly, prevention would mean that the relapse emerged but was eliminated

by the residual drug levels, and thus, the first observed recurrence is in fact the second relapse. Delay would mean that the residual drug levels reduced multiplication of the relapse, so it reached patency after a substantial delay. In the former explanation, overall relapse numbers might be reduced as this would exhaust the liver burden of hypnozoites more rapidly. A pharmacometric time-to-event approach explaining relapsing *P. vivax* malaria, first presented for amodiaquine treatment in pregnant women,²⁹ described the data observed in this study well. An interval-censoring time-to-event model was successfully used since the exact times of relapses were unknown. The risk of relapsing *P. vivax* malaria in patients was explained by a constant baseline hazard (i.e., 8.94 relapses/year) with a 123% increased risk every third week. The pharmacodynamic model was significantly improved by adding a protective drug effect, indicating that piperazine cured the primary blood stage *P. vivax* infection and fully suppressed a large proportion of the first relapses that would have occurred three weeks after the initial infection. The pharmacodynamic model also indicated that the observed high recurrence rate is likely to be explained mainly by delayed second relapses, although it cannot be excluded that some recurrences are reinfections. This can only be mechanistically evaluated by an individual parasite-data-driven pharmacokinetic-pharmacodynamic model. Simulations using the final pharmacokinetic-pharmacodynamic model (Figure 4) demonstrated that minimum day-7 piperazine venous and capillary concentrations of 27 and 38 ng/ml, respectively, were needed to suppress the risk of relapse for 30 days (i.e., for 99% of simulated patients). This is in close agreement with a previously defined day-7 threshold venous concentration of 30 ng/ml in patients with *P. falciparum* malaria to minimize the risk of recrudescence infections.³⁰

In conclusion, the population pharmacokinetic properties of piperazine in patients with *P. vivax* malaria were described successfully by modeling venous and capillary plasma concentrations simultaneously. Body weight was the only significant covariate and resulted in increasing clearance and volumes with increasing body weight. Times to recurrent *P. vivax* infections were successfully modeled with a time-to-event approach and resulted in a significantly delayed time to recurrent infections during the post-treatment phase of piperazine treatment. This was explained by prevention of the first *P. vivax* malaria relapse by residual piperazine concentrations. Piperazine is a good candidate for the treatment of *P. vivax* malaria and the modeling conducted here demonstrated the added benefit of reduced morbidity compared with other more rapidly eliminated antimalarial drugs.

METHODS

Ethics approval.

Ethics approval was granted by the Faculty of Tropical Medicine Ethics Committee, Mahidol University, Bangkok, Thailand, and the Oxford Tropical Research Ethics Committee, Oxford University, UK. This study was registered in the ISRCTN Register (ISRCTN87827353). Details on the clinical study design and outcomes have been presented in full elsewhere.¹⁶

Drug regimen and blood collection.

All patients received a standard regimen of DHA-PQ. Patients who vomited the dose were excluded from the pharmacokinetic study. A full or half replacement dose was administered if vomiting occurred within 30 min of or between 30 min to 1 h after administration, respectively.

Venous blood samples (3 ml) and capillary blood samples (200 μ l) were drawn from each patient for piperazine plasma measurements. Samples were collected randomly over 69 days (4–7 venous samples and 1–11 capillary samples) from 62 patients and an additional venous sample (5 ml) was drawn from these patients at the time of recurrent *P. vivax* malaria. This random sampling allowed of an adequate coverage of the entire concentration–time profile. An additional 54 patients provided a single plasma sample only at the time of recurrent malaria. Patients were excluded from the study after recurrent malaria. Plasma samples were shipped on dry ice to the Department of Clinical Pharmacology, Mahidol-Oxford Tropical Medicine Research Unit, Thailand, for drug quantification.³¹

Population modeling.

Piperazine pharmacokinetics and pharmacodynamics were evaluated using nonlinear mixed-effects modeling (NONMEM version VII; Icon Development Solutions, Ellicott City, MD). Postprocessing and automation were performed using Pearl-Speaks-NONMEM version 3.5.3, Census version 1.2b2,³² and Xpose version 4.0³³ in the programming language R version 2.13.2 (The R Foundation for Statistical Computing). Measurements below the lower limit of quantification were less than 0.5% of the total data and therefore omitted. OFV, calculated by NONMEM as minus twice the log-likelihood up to constant, was used for model selection during the model-building process. A difference in the OFV (Δ OFV) of >3.84 was considered significant ($P < 0.05$) when comparing two nested models with 1 degree of freedom difference. Full details of the pharmacokinetic and pharmacodynamic modeling methodology can be found in supplementary material.

Acknowledgments. The authors gratefully thank all the patients who participated in this study. They also thank the staff at the Shoklo Malaria Research Unit, Mae Sot, Thailand, and all the staff involved in taking and processing the samples. This study was part of the Mahidol-Oxford Tropical Medicine Research Unit, funded by the Wellcome Trust of Great Britain.

Author Contributions. J.T., P.T., N.P.J.D., and N.J.W. wrote the manuscript. A.P.P., K.M.L., E.A.A., N.P.J.D., F.N., and N.J.W. designed the research. J.T., A.P.P., K.M.L., E.A.A., and F.N. performed the research. J.T., P.T., and W.H. analyzed the data.

Conflict of Interest. The Wellcome Trust is a UK-based charity that supports medical research and is independent of any drug companies. It has no financial links with the manufacturers of either the diagnostic tests or the drugs used in this study. The authors declared no conflict of interest.

Study Highlights**WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?**

- ✓ *Plasmodium vivax* malaria causes substantial morbidity in transmission areas as a result of multiple relapses. *P. vivax* resistance to chloroquine is increasing in many parts of the world. Dihydroartemisinin-piperazine is a promising candidate for the treatment of *P. vivax* malaria.

WHAT QUESTION DID THIS STUDY ADDRESS?

- ✓ The aim of this study was to evaluate the population pharmacokinetics and pharmacodynamics of piperazine in patients with *P. vivax* malaria in Thailand and to determine whether residual piperazine prevents or delays the emergence of *P. vivax* relapse.

WHAT THIS STUDY ADDS TO OUR KNOWLEDGE

- ✓ Piperazine pharmacokinetics and the biology of *P. vivax* malaria were successfully modeled. A large proportion of the first relapses were suppressed completely by residual piperazine concentrations, and recurrences resulted mainly from the second or the third relapse or from reinfection.

HOW THIS MIGHT CHANGE CLINICAL PHARMACOLOGY AND THERAPEUTICS

- ✓ Dihydroartemisinin-piperazine is a promising candidate for the treatment of *P. vivax* malaria and the modeling conducted here demonstrated the added benefit of reduced morbidity compared with other short-lived antimalarial drugs.

- Price, R.N., Tjitra, E., Guerra, C.A., Yeung, S., White, N.J. & Anstey, N.M. Vivax malaria: neglected and not benign. *Am. J. Trop. Med. Hyg.* **77**, 79–87 (2007).
- Guerra, C.A. *et al.* The limits and intensity of Plasmodium falciparum transmission: implications for malaria control and elimination worldwide. *PLoS Med.* **5**, e38 (2008).
- Guerra, C.A. *et al.* The international limits and population at risk of Plasmodium vivax transmission in 2009. *PLoS Negl. Trop. Dis.* **4**, e774 (2010).
- Anstey, N.M., Russell, B., Yeo, T.W. & Price, R.N. The pathophysiology of vivax malaria. *Trends Parasitol.* **25**, 220–227 (2009).
- Poesoprodjo, J.R. *et al.* Vivax malaria: a major cause of morbidity in early infancy. *Clin. Infect. Dis.* **48**, 1704–1712 (2009).
- Nosten, F. *et al.* Effects of Plasmodium vivax malaria in pregnancy. *Lancet* **354**, 546–549 (1999).
- Collins, W.E., Jeffery, G.M. & Roberts, J.M. A retrospective examination of anemia during infection of humans with Plasmodium vivax. *Am. J. Trop. Med. Hyg.* **68**, 410–412 (2003).
- Rieckmann, K.H., Davis, D.R. & Hutton, D.C. Plasmodium vivax resistance to chloroquine? *Lancet* **2**, 1183–1184 (1989).
- Baird, J.K. Chloroquine resistance in Plasmodium vivax. *Antimicrob. Agents Chemother.* **48**, 4075–4083 (2004).
- WHO. *World Health Organization Guidelines for the Treatment of Malaria*. (WHO, Geneva, Switzerland 2010).
- Zwang, J. *et al.* Safety and efficacy of dihydroartemisinin-piperazine in falciparum malaria: a prospective multi-centre individual patient data analysis. *PLoS ONE* **4**, e6358 (2009).
- Douglas, N.M., Anstey, N.M., Angus, B.J., Nosten, F. & Price, R.N. Artemisinin combination therapy for vivax malaria. *Lancet Infect. Dis.* **10**, 405–416 (2010).

13. Ratcliff, A. *et al.* Two fixed-dose artemisinin combinations for drug-resistant falciparum and vivax malaria in Papua, Indonesia: an open-label randomised comparison. *Lancet* **369**, 757–765 (2007).
14. Karunajeewa, H.A. *et al.* A trial of combination antimalarial therapies in children from Papua New Guinea. *N. Engl. J. Med.* **359**, 2545–2557 (2008).
15. Tarning, J. *et al.* Population pharmacokinetics and pharmacodynamics of piperaque in children with uncomplicated falciparum malaria. *Clin. Pharmacol. Ther.* **91**, 497–505 (2012).
16. Phyo, A.P. *et al.* Dihydroartemisinin-piperaque versus chloroquine in the treatment of Plasmodium vivax malaria in Thailand: a randomized controlled trial. *Clin. Infect. Dis.* **53**, 977–984 (2011).
17. Ashley, E.A. *et al.* Comparison of plasma, venous and capillary blood levels of piperaque in patients with uncomplicated falciparum malaria. *Eur. J. Clin. Pharmacol.* **66**, 705–712 (2010).
18. Cox, E.H., Veyrat-Follet, C., Beal, S.L., Fuseau, E., Kenkare, S. & Sheiner, L.B. A population pharmacokinetic-pharmacodynamic analysis of repeated measures time-to-event pharmacodynamic responses: the antiemetic effect of ondansetron. *J. Pharmacokinet. Biopharm.* **27**, 625–644 (1999).
19. White, N.J. Determinants of relapse periodicity in Plasmodium vivax malaria. *Malar. J.* **10**, 297 (2011).
20. White, N.J. Pharmacokinetic and pharmacodynamic considerations in antimalarial dose optimization. *Antimicrob. Agents Chemother.* **57**, 5792–5807 (2013).
21. Pukrittayakamee, S. *et al.* Therapeutic responses to different antimalarial drugs in vivax malaria. *Antimicrob. Agents Chemother.* **44**, 1680–1685 (2000).
22. Tarning, J. *et al.* Population pharmacokinetics of dihydroartemisinin and piperaque in pregnant and non-pregnant women with uncomplicated malaria. *Antimicrob. Agents Chemother.* **56**, 1997–2007 (2012).
23. Hung, T.Y. *et al.* Population pharmacokinetics of piperaque in adults and children with uncomplicated falciparum or vivax malaria. *Br. J. Clin. Pharmacol.* **57**, 253–262 (2004).
24. Tarning, J. *et al.* Population pharmacokinetics of piperaque after two different treatment regimens with dihydroartemisinin-piperaque in patients with Plasmodium falciparum malaria in Thailand. *Antimicrob. Agents Chemother.* **52**, 1052–1061 (2008).
25. Hoglund, R.M. *et al.* A population pharmacokinetic model of piperaque in pregnant and non-pregnant women with uncomplicated Plasmodium falciparum malaria in Sudan. *Malar. J.* **11**, 398 (2012).
26. Holford, N.H. A size standard for pharmacokinetics. *Clin. Pharmacokinet.* **30**, 329–332 (1996).
27. Rijken, M.J. *et al.* Pharmacokinetics of dihydroartemisinin and piperaque in pregnant and nonpregnant women with uncomplicated falciparum malaria. *Antimicrob. Agents Chemother.* **55**, 5500–5506 (2011).
28. Nguyen, D.V. *et al.* Pharmacokinetics and ex vivo pharmacodynamic antimalarial activity of dihydroartemisinin-piperaque in patients with uncomplicated falciparum malaria in Vietnam. *Antimicrob. Agents Chemother.* **53**, 3534–3537 (2009).
29. Tarning, J. *et al.* Population pharmacokinetic and pharmacodynamic modeling of amodiaquine and desethylamodiaquine in women with Plasmodium vivax malaria during and after pregnancy. *Antimicrob. Agents Chemother.* **56**, 5764–5773 (2012).
30. Price, R.N. *et al.* Clinical and pharmacological determinants of the therapeutic response to dihydroartemisinin-piperaque for drug-resistant malaria. *Antimicrob. Agents Chemother.* **51**, 4090–4097 (2007).
31. Lindegardh, N., Annerberg, A., White, N.J. & Day, N.P. Development and validation of a liquid chromatographic-tandem mass spectrometric method for determination of piperaque in plasma stable isotope labeled internal standard does not always compensate for matrix effects. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **862**, 227–236 (2008).
32. Wilkins, J.J. NONMEMory: a run management tool for NONMEM. *Comput. Methods Programs Biomed.* **78**, 259–267 (2005).
33. Jonsson, E.N. & Karlsson, M.O. Xpose—an S-PLUS based population pharmacokinetic/pharmacodynamic model building aid for NONMEM. *Comput. Methods Programs Biomed.* **58**, 51–64 (1999).



This work is licensed under a Creative Commons Attribution 3.0 Unported License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/3.0/>

Supplementary information accompanies this paper on the *CPT: Pharmacometrics & Systems Pharmacology* website (<http://www.nature.com/psp>)