

Genome Sequence of *Hafnia alvei* bta3_1, a Bacterium with Antimicrobial Properties Isolated from Honey Bee Gut

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***Hafnia alvei* bta3_1, a strain with antibacterial properties, was isolated from honey bee gut and cultured under aerobic and anaerobic conditions. To explore the potential genetic bases of its antibacterial and possible pathogenic properties, the complete genome of this organism was sequenced and analyzed.**

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Honey bees (*Apis mellifera*) are important agricultural pollinators (1) that have suffered declines in recent years (2). Microbial pathogens contribute to these declines, emphasizing the need to better understand microbial associates of honey bees. *Hafnia alvei* is one of several *Enterobacteriaceae* species that occur sporadically in bee gut and that may represent opportunistic pathogens. We sequenced and analyzed the genome of *H. alvei* bta3_1, a Gram-negative, facultatively anaerobic, rod-shaped bacterium isolated from honey bee gut. *H. alvei* bta3_1 showed antagonistic activity toward a tested strain of *Bacillus* sp., but no activity was detected toward *E. coli* EPI300, *Paenibacillus larvae*, or fungal strains.

DNA from *H. alvei* bta3_1 (3), was used to make a mate-pair library with average insert size of 2,933 bp (500–8,000 bp) that was end sequenced with Illumina HiSeq2000, resulting in 67,735,948 reads with average length of 76 bp. Ten percent of trimmed, filtered reads were randomly extracted and *de novo* assembled using CLC Genomic Workbench 4.6.1 with an optimal k-mer value of 43, yielding 291 contigs (>200 bp) of average 27.5-fold coverage. SSPACE (4) and Bowtie 2 2.0.2 (5) were used for scaffolding the pre-assembled contigs, and scaffolds were visualized using samtools 0.1.18 (6) and Tablet 1.12.09.03 (7), resulting in 8 super-scaffolds disrupted by rRNA operons. These were organized into linear draft genome sequences using mate-pair information. The gap-filled genomic sequences were manually revised using Tablet to finalize the consensus linear genome. Annotation was carried out using RAST (8), followed with manual curation by searching the automatically annotated coding sequences against GenBank nr and Pfam databases. Phylogenomic analyses were carried out using the core genomic data from 22 *Enterobacteriales* genomes produced in MicroScope (<https://www.genoscope.cns.fr>). A phylogenomic tree was constructed using the JC model calculated from ProtTest 3.0 (9) and bootstrap replicate branch supports were obtained using PhyML 3.0 in Seaview 4 platform (10).

The final assembly yielded a circular chromosome of 4,763,672 bp and 48.3% G+C. The genome contains 4,363 genes, including 4,270 protein-coding sequences, 72 tRNA genes, and 7 complete rRNA loci. Putative gene clusters were identified for the produc-

tion of colicin and tolerance to colicin, and siderophores. The presence of genes for production of bacteriocin and siderophore suggests that *H. alvei* can suppress or compete with other microbes inhabiting bee gut. Also present were genes for quorum sensing, biofilm formation, and motility (11–13). Phylogenomic analysis of *Hafnia* and 22 other *Enterobacteriaceae* based on 178 core genes clustered *H. alvei* bta3-1, *H. alvei* FB1, *H. alvei* ATCC 51873, and *Enterobacteriaceae* bacterium 9_2_54FAA together, with *Serratia* as the sister clade. Strain BIDMC_31, which was previously categorized as *H. alvei*, clustered with *Klebsiella*.

H. alvei bta3_1 isolated from honey bee gut with antimicrobial activity toward *Bacillus* sp. may represent an opportunistic gut pathogen in honey bees, and potentially interacts with other bacteria in the honey bee gut microbiota.

Nucleotide sequence accession numbers. The *H. alvei* bta3_1 genome project has been submitted to GenBank under project accession number PRJNA182589. The assembled and annotated *H. alvei* bta3_1 chromosome has been deposited in GenBank under accession number CP004083.

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REFERENCES

- Gallai N, Salles JM, Settele J, Vaissière BE. 2009. Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. *Ecol Econ* 68:810–821. <http://dx.doi.org/10.1016/j.ecolecon.2008.06.014>.
- United States Department of Agriculture. 2010. Colony Collapse Disorder Progress Report. United States Department of Agriculture, Washington, DC.
- Tian BY, Fadhil NH, Powell JE, Kwong WK, Moran NA. 2012. Long-term exposure to antibiotics has caused accumulation of resistance determinants in the gut microbiota of honeybees. *mBio* 3:e00377-12. <http://dx.doi.org/10.1128/mBio.00377-12>.
- Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics* 27:578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with bowtie 2. *Nat Methods* 9:357–359. <http://dx.doi.org/10.1038/nmeth.1923>.

6. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R; 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment of map format and SAMtools. *Bioinformatics* 25:2078–2079.
7. Milne I, Bayer M, Cardle L, Shaw P, Stephen G, Wright F, Marshall D. 2010. Tablet-next generation sequence assembly visualization. *Bioinformatics* 26:401–402. <http://dx.doi.org/10.1093/bioinformatics/btp666>.
8. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
9. Abascal F, Zardoya R, Posada D. 2005. ProtTest: selection of best-fit models of protein evolution. *Bioinformatics* 21:2104–2105. <http://dx.doi.org/10.1093/bioinformatics/bti263>.
10. Gouy M, Guindon S, Gascuel O. 2010. SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. *Mol Biol Evol* 27:221–224. <http://dx.doi.org/10.1093/molbev/msp259>.
11. Janda JM, Abbott SL. 2006. The genus *Hafnia*: from soup to nuts. *Clin Microbiol Rev* 19:12–18. <http://dx.doi.org/10.1128/CMR.19.1.12-28.2006>.
12. Padilla D, Remuzgo-Martínez S, Acosta F, Ramos-Vivas J. 2013. *Hafnia alvei* and *Hafnia paralvei*, taxonomy defined but still far from virulence and pathogenicity. *Vet Microbiol* 163:200–201. <http://dx.doi.org/10.1016/j.vetmic.2012.11.041>.
13. Vivas J, Padilla D, Real F, Bravo J, Grasso V, Acosta F. 2008. Influence of environmental conditions on biofilm formation by *Hafnia alvei* strains. *Vet Microbiol* 129:150–155. <http://dx.doi.org/10.1016/j.vetmic.2007.11.007>.