

Review Article

Encapsulation of Gold Nanorods with Porphyrins for the Potential Treatment of Cancer and Bacterial Diseases: A Critical Review

Nthabeleng Hlapisi ¹, Tshwafo E. Motaung ¹, Linda Z. Linganiso,¹
Oluwatobi S. Oluwafemi,^{2,3} and Sandile P. Songca⁴

¹Department of Chemistry, University of Zululand, X1001, KwaDlangezwa, KwaZulu-Natal, South Africa

²Department of Applied Chemistry, University of Johannesburg, P.O. Box 17011, Doornfontein, Johannesburg 2028, South Africa

³Centre for Nanomaterials Science Research, University of Johannesburg, Johannesburg, South Africa

⁴Department of Chemistry, University of Kwazulu Natal, Kwazulu Natal, South Africa

Correspondence should be addressed to Nthabeleng Hlapisi; nsisitha@gmail.com and Tshwafo E. Motaung; motaungt@unizulu.ac.za

Received 1 August 2018; Accepted 4 February 2019; Published 30 April 2019

Guest Editor: Rais A. Khan

Copyright © 2019 Nthabeleng Hlapisi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Cancer and bacterial diseases have been the most incidental diseases to date. According to the World Health Report 2018, at least every family is affected by cancer around the world. In 2012, 14.1 million people were affected by cancer, and that figure is bound to increase to 21.6 million in 2030. Medicine therefore sorts out ways of treatment using conventional methods which have been proven to have many side effects. Researchers developed photothermal and photodynamic methods to treat both cancer and bacterial diseases. These methods pose fewer effects on the biological systems but still no perfect method has been synthesized. The review serves to explore porphyrin and gold nanorods to be used in the treatment of cancer and bacterial diseases: porphyrins as photosensitizers and gold nanorods as delivery agents. In addition, the review delves into ways of incorporating photothermal and photodynamic therapy aimed at producing a less toxic, more efficacious, and specific compound for the treatment.

1. Introduction

Drugs based on metals have since been used during ancient times; however, modern metal-based medicine dates to about 50 years ago through the use of cisplatin; since then, the antitumor activity of these metals has been studied [1–5]. Gold salts and arsenic compounds have been used for the treatment of arthritis and syphilis, respectively. Heavy metals, rhodium, indium, palladium, and noble metals have the ability to be used as therapeutic agents. The noble metal nanoparticles work better when ligands are attached to them as opposed to when they operate singularly as free ligands; in addition, antibiotic compounds can also be bound to the metals, thereby increasing their efficacy or to avoid the resistance by drugs [6, 7]. In their elemental and coordinated states, metals act as various agents: antibiotic, antimalarial, antibacterial, antiviral, antitubercular, antimycotic, antiarthritis, and anti-inflammatory. In their elemental states,

nanoparticle formulations are used as carrier agents for drugs and to toxify bacteria but this toxicity of bacteria in humans is still a subject of research [8]. The specificity and selectivity of target drugs that only hit the affected cells and tissue have been a challenge in the therapeutic field, but the use of receptor nanoparticles has been implemented to overcome this challenge. Other challenges as highlighted by Akhtar et al. [9] are primarily on the basis of the design of a nanocarrier, the drug loading efficiency, how stable the ligands are on the nanoconjugates, maximum receptor-ligand interactions, and the time the targeted receptor would be expressed, the toxicity of the nanoparticle, and immunity to the blood system [10]. Gold nanorods (AuNRs) are used in photodynamic and photothermal therapy for clinical purposes on the basis of their optical and chemical properties. Porphyrin is one of the common photosensitizers used for the treatment of tumors and antibacterial infections. Nanoparticles have been introduced to the clinical field as to

prevent or assist in the traditional methods of treating cancer.

Nobel metal nanoparticles have strong electric fields at the surface; hence, the absorption and scattering of electromagnetic radiation by those nanoparticles are enhanced, making them ideal choices in the photothermal cancer therapy. The best option is to use agents that are active at the near-infrared region for minimal light extinction by the intrinsic chromophores in the tumor tissues. AuNRs with suitable aspect ratios have the ability to absorb and scatter light in the NIR (650–900 nm), hence used in molecular imaging and photothermal cancer therapy. Solid tumors have leaky blood vessels with cell junction gaps from 100 nm to 780 nm, and this permeable vasculature allows for the extravasation of gold nanoparticles into the tumor matrix. The optimal aspect ratio as experimentally obtained by Singh et al. for *ex vivo* experiments was determined as 4.0 ± 0.06 as per TEM results. The aspect ratio resulted in the SPR peak of the AuNR to be around 808 nm which is in the NIR and allows for deep optical tissue penetration [11].

Some scientists however argue for the use of nanoparticles as tumor-targeted delivery systems in the clinical environment. An example is seen where Van de Meel et al. [12] state that the “unappreciation” of nanomedicine is due to clinical trials practised on mice as opposed to humans and that only 0.7% of the researched nanomedication even reaches clinical trials. He however concluded that researchers should focus not only on the understanding of the biology and chemistry behind nanocarriers but also on the idealistic “disease-driven” approach on their potential.

The use of gold nanoparticles has been of interest lately due to their impeccable properties as carrier agents of drugs; the properties include the fact that a wide range of organic molecules can adhere and be bonded to the nanoparticles, they have low levels of toxicity, their absorption is very tunable, and they have small sizes and high surface area to the volume ratio. These properties however ideally work when the surface of the gold particles is modified and tuned to suit the use [13]. Gold nanoparticles are biologically inert, have high dispersity, are noncytotoxic, are biocompatible, and are optically tunable [14]. The treatment of cancer has traditionally made use of the systematic delivery of an anticancer agent which unfortunately leads to the accumulation of the agent to the tumor cell; a new method has however been developed which ensures that the anticancer agent is delivered through the lymphatic network. Oladipo et al. used the polymeric AuNRs (PAuNRs) to deliver drugs from an accessory lymph node to a proper auxiliary lymph node containing a tumor in order to treat lymph node metastases [15]. Coating of nanoparticles with alkane thiols monolayer has become of great attraction as resulting nanoconjugates have unique optical and electronic properties that depend and can be controlled by the particle size and the nature of the protecting molecules [16].

Gold nanoparticles possess unique optical properties. AuNRs particularly have two distinctive absorption bands: one is caused by light absorbed on the short axis (transverse) and another due to light absorbed on the long axis (longitudinal). The ability of the AuNRs to be optically

controlled and the fact that they are very sensitive to changes in the local environment makes them excellent materials for sensing, photothermal therapy, and imaging [17]. AuNRs applications are vast; they can be used in biomedical technologies, plasmon-enhanced spectroscopies, and optical and optoelectronic devices. AuNRs exhibit special optical properties in cancer diagnostics and treatment. Their advantages however still bring a few questions: their toxicity in the blood system, their biodistribution, and their fate *in vivo*. Still to be explored is a further optical control of gene delivery and use of nanorods for *in vivo* spectroscopic tumor detection combined with organic dye molecules [18].

Photodynamic therapy is one of the most effective regimens in the treatment of cancer, precancerous inductions, actinic keratosis, infections, dermatology, cardiovascular illness, and wound healing. For photodynamic therapy to successfully proceed, three elements have to be present: oxygen, a photosensitizer, and light [19]. Porphyrins are photosensitizers which selectively accumulate in tumor cells, releasing cytotoxic substances and hence destroy the cancerous cells with minimal harm to the surrounding healthy cells. Exposure of a photosensitizer to photoactivating light enhances the destruction of malignant tissues. Clinically, common photosensitizers used are the haematoporphyrin derivative (HPD) and the Photofrin II, which is a purified form of HPD [20]. Porphyrin is a naturally occurring heterocyclic compound shown in Figure 1 [21].

Heterocyclic compounds occur widely in nature in compounds such as hemes and chlorophylls. These compounds are used in medicines, pharmaceutical, agrochemical, and energy materials. Polyheteroatomic heterocycles are used in clinical therapy as they allow for direct interactions with the biological targets which are often not possible with single heterocyclic compounds [22]. Porphyrins are heterocyclic compounds with semiconducting properties; these compounds can be used for a number of applications: artificial photosynthesis, catalysis, molecular electronics, sensors, nonlinear optics, and solar cells. The functionality of porphyrins depends on their crystallography, which depends on the plane of the macrocycle and the conjunction of the pi-bonding to the adjacent macrocycle or how the macrocycles are interconnected with the ligands by covalent bonds. The modification of the basic porphyrin structure enhances its semiconductivity property and hence its ability to be used as a photosensitizer. Porphyrins also have the tumor-localizing ability [23].

Encapsulation of gold nanorods with porphyrin has been achieved before to obtain novel multifunctional nanoparticles. The porphyrin structures were, for example, doped with silica shells to protect them from the external bioenvironment [24]. The resulting multifunctional nanoparticles are good candidates for both photosensitization and two-photon imaging and also image-guided therapy.

The use of imaging therapy has many advantages over conventional therapy. The merits of two imaging therapy include the bioimaging that provides for deeper penetration (light) and has a 3D imaging capability. Metal nanoparticles especially gold nanorods have shown an excitingly good

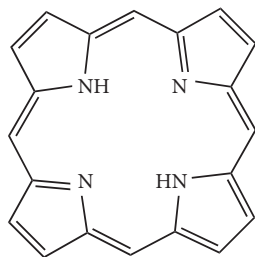


FIGURE 1: Structure of porphyrin.

promise in two-photon imaging. They have high two-photon luminescence which is approximately 58 times greater than that of a single rhodamine molecule [25]. The biological transparency window is around 6000–10000 nm which coincides with the longitudinal plasmon band of the gold nanorods. The nanorods alone however lack the targeting ability, hence with combination with porphyrins will create a more specific selective multifunctional compound.

The purpose of this study is to revise the coating performed on nanorods with porphyrin for further modification for the treatment of cancer and bacteria using photodynamic and photothermal therapy [26–29].

2. Literature Review

2.1. Gold Nanoparticles. Generally, metal nanoparticles have been used for cancer diagnosis and treatment for the past decades [30–34]. Multifunctional nanoparticles have specifically been used for a number of aspects such as targeting, imaging, and therapy as a way to overcome challenges imposed by conventional treatment (radiotherapy, chemotherapy, and surgery). Amongst all, the use of iron oxide nanoparticles received overwhelming attention primarily due to super magnetic properties for magnetic resonance imaging (MRI). In fact, magnetic nanoparticles other than gold and carbon nanoparticles could be degraded to the respective metal cations in the body especially in acidic media, increasing the toxicity of the long-term nanoparticles residing in the body [35]. Certain metal nanoparticles interact with light, and that proved to be an opportunity for biophonic nanomedicine [36, 37]. One of the main fields that use nanoparticles (NPs) is the image-guided therapy; it uses multifunctional nanoparticles based on surface plasmon resonance absorption property which is tunable in the near-infrared region [38, 39]. Typical examples are gold nanorods, for which optical properties could be explained better by the Mie–Gans theory [40]. The sensitivity of the plasmon resonance frequency of the nanorods towards the refractive index of its surroundings makes them suitable for use in biological sensing. They are also proven to be very sensitive to thermal environments and possess nonlinear optical response properties [41, 42]. Synthesis of the nanorods is one of the main parameters that are controlled for optimization of properties [43]. An important technology investigated in this review as a promising factor for the image-guided therapy and diagnosis is the passivation of gold nanorods [44, 45].

Gold nanoparticles have different shapes ranging from spherical, suboctahedral, octahedral, decahedral, icosahedral multiple twinned, irregular shaped, tetrahedral, nanotriangles, nanoprisms, hexagonal platelets, and nanorods [46–48]. Fluorescent nanoparticles have good biocompatibility for molecular imaging and metabolites used for cellular functions in cancer. On the other hand, nanorods show special optical and chemical properties for biological applications. They also have unique anisotropic geometry for tuneable absorption in the visible and near-infrared region. These phenomena make them useful for biosensing, photothermal therapy, and gene delivery [49–52]. Generally, nanoparticles are used for their light scattering and absorbing abilities, and scattering is more important for microscopy and optical coherence tomography (OCT) [53].

The effects of the nanoparticles can be detected using various imaging apparatus: magnetic resonance imaging, nuclear imaging, and photoacoustic imaging. After the administration of the nanoparticle to the tumor cell, they land on a solid target tissue, and their activity can be changed by an external stimulus. An example is photothermal therapy, in which the light energy is converted to heat energy for damaging the cancer cells [54, 55]. The dependence of the excitation of the noble metal nanorod energy lies in the dimensions of the nanorods and its environment. This enhances tunable optical properties which can be demonstrated by the Mie–Gans theory. It states that there is a direct dependence of the geometry end cap of the nanoparticles and the particles' size, which affects the peak position in the absorption spectra. For AuNRs, this gives rise to the two absorption bands by the longitudinal band and the transverse band [56–58].

2.1.1. Properties of Gold Nanorods. The optical properties of AuNRs are of great importance and make them excellent to use in therapy. In the past, the use of gold nanoparticles was limited to additives for aesthetic purposes, but emerging studies indicated that the peculiar optical properties at nanoscale of the particles brought other dimensions. For instance, the increase in rod length of the gold nanorod increases the longitudinal band red shift and the extinction coefficient [59, 60], as shown in Figure 2 [60, 62–66]. As a result, the nanostructures are used in sensing, imaging, and photothermal therapy. One property which qualifies the nanorods to be used in as biological sensors is that their absorption band changes with the refractive index of the local material. This phenomenon allows for very accurate sensing. They are often identified by their aspect ratio and are cylindrical rods with a width less than 10 nm to over 40 nm and length to several nanometers; an example is seen in Figure 3. For one to fully comprehend the unique features of nanorods, the robust extinction coefficient for predicting the concentration at an absorbance has to be known. The AuNRs are effective for the detection of sequences of infectious agents for diseases like HIV-1 [68].

AuNRs absorb at the near-infrared region where the maximum radiation through the tissue occurs, hence used for *in vivo* imaging and photothermal therapy. Nanorods

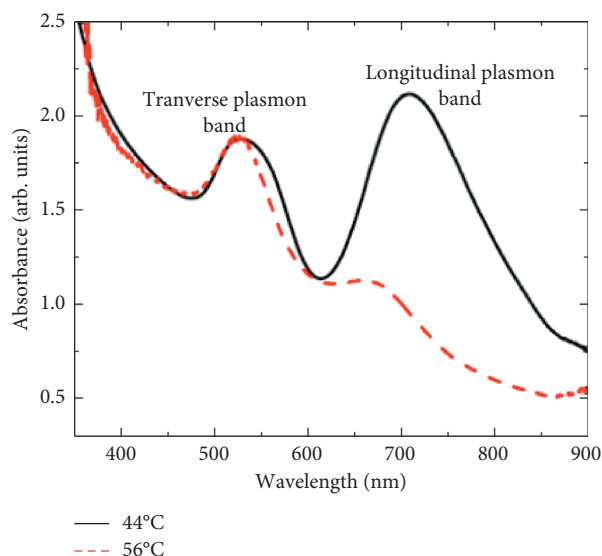


FIGURE 2: UV-Vis spectrum of AuNRs at different preparation temperatures [61].

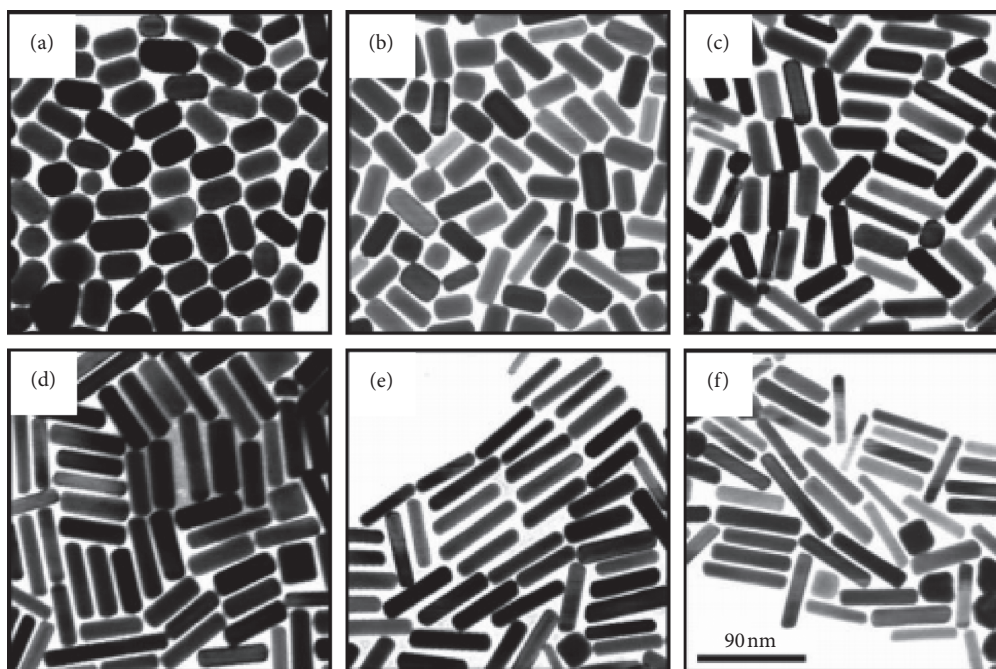


FIGURE 3: Illustration of gold nanorods of different aspect ratios in AgNO_3 growth solution with concentration (a) 0.03, (b) 0.05, (c) 0.075, (d) 0.1, (e) 0.125, and (f) 0.15 mmol^{-1} [67].

exhibit two bands of surface plasmon resonance (SPR), which are a product of the conduction band along their long and short axes. On the visible region, the transverse band and the longitudinal band occur on the near-infrared region (NIR) [69–72]. Tailoring of the nanorods during synthesis enables the absorption bands to be of desired wavelength in the near-infrared region, and this can be attributed to the fact that the longitudinal length of the nanorods can be tuned via its aspect ratio. Their luminous energy can be converted into heat energy which is influenced by the

maximum penetration of light in tissues favorable for photothermal therapy [73–76]. AuNRs have a higher distinct local field enhancement resulting in a significant surface-enhanced Raman spectroscopy (SERS) activity. Their modifiable surface area enables the incorporation of drugs for use as drug delivery agents [77–79].

The distinctive optical and electrical properties of AuNRs depend on size and aspect ratio. For the nanorods to efficiently work, several aspects need to be addressed: improvement of the synthesis of AuNRs in terms of reproducibility and

efficiency, full understanding of the direct characterization of nanorod geometry functionalization, and lastly, to find a cost-effective and sensitive method that would be used for nanorod sensing. Another important aspect is coating which also depends on synthesis that may lead to covalent or noncovalent bonds at the interface [80–82].

X-ray diffraction (XRD) shows a face-centred cubic close packing arrangement of gold nanorods in specific areas. The XRD peaks are very consistent with ones of metallic gold. Sharp peaks resembling ones of gold are observed in Figure 4, hence confirming the crystalline nature of the gold nanorods. Figure 4 shows an XRD pattern of gold nanorods [83].

Various diffraction patterns are presented at 2θ with values 38.4° , 44.4° , and 64.6° which correspond to the following diffraction planes: (111), (200), and (220), respectively. The peaks perfectly index to a cubic face-centred structure of a gold metal. The XRD showed a lattice constant of $a = 4.082 \text{ \AA}$ which is in agreement with 4.079 \AA , which is the standard diffraction pattern of a cubic gold metal (CAS: 7440-57-5) by Pallares [83].

2.1.2. Synthesis of Gold Nanorods. The nanorods are mainly synthesized using the wet chemical method and the hard template directed (an example in Figure 5) method [84, 85]. The wet chemical techniques involve the reduction of metal ions at gold surfaces in the presence of various surfactants; the technique however produces hybrids of nanospheres and other shapes clearly dominated by rods and that affects the ideal optical response of the gold nanorods. For high purity nanorods, the hard template directed methods are utilized by making use of the polycarbonate membrane and the anionic aluminum oxide (AAO) as templates [86, 87]. A typical example of the wet chemical method is the synthesis of AuNRs by seeding. In this method, a spherical seed of nanoparticles is added to a gold salt solution containing ascorbic acid, silver nitrate, and cetyltrimethylammonium bromide (CTAB) which then produces nanoparticles with a rod-like shape, as shown in Figures 5 and 6 [88]. CTAB enhances the rod-like shape of nanoparticles by preferably binding to the sides of the nanoparticles, and the concentration of gold nanoparticles is reduced slowly by the growth of the nanorods particles. Variation of silver nitrate concentration is used for the alteration of the rod length. The synthesis is followed by the centrifugation with distilled water to purify the nanoparticles by removing excess CTAB which is cytotoxic, unreacted metal ions, and the ascorbic acid.

The hard template directed method includes photochemical and electrochemistry methods, from which nanorods are grown in an electrolyte solution under regulated current between two electrolytes [89–92]. A gold metal plate acts as an anode and a platinum plate as a cathode. Both electrodes are immersed into an electrolytic solution containing a cationic surfactant.

The solution normally is placed in an ultrasonic bath to avoid aggregation of the gold nanoparticles before the addition of acetone and cyclohexane to the electrolyte

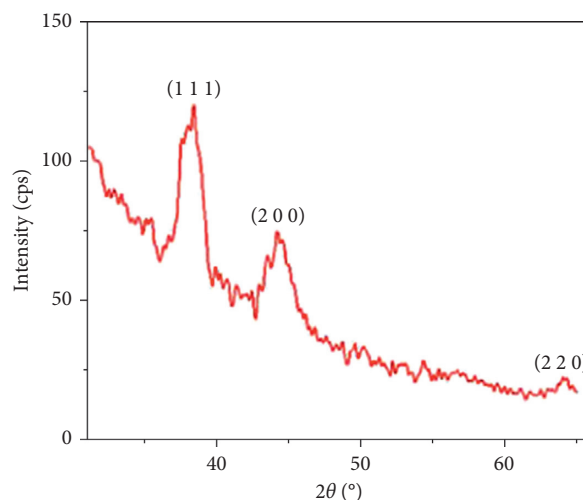


FIGURE 4: XRD pattern of gold nanorods as prepared by the sonoelectrochemical method [83].

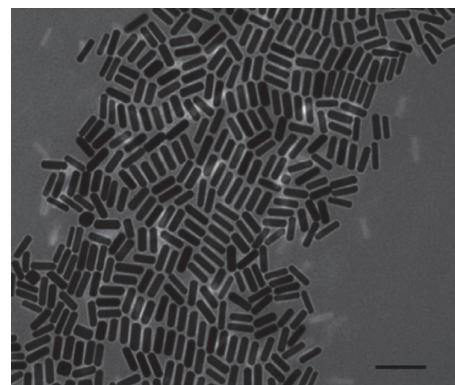


FIGURE 5: AuNRs prepared in the presence of a direction agent CTAB. Scale: 100 nm (TEM micrograph).

solution. Another electrochemical method can be used where the metal (gold) can be electrochemically deposited inside a nanoporous polycarbonate template of alumina membranes, as shown in Figure 7 [94]. In photochemical methods, gold salt is irradiated with UV light in the presence of CTAB and tetradodecylammonium bromide.

2.1.3. The Seed-Mediated Method/Green Method Synthesis. Although the synthesis of AuNRs using the template method has been perfected and produces the AuNRs, other methods are also used as greener methods to give comparable results as the template method. Gole and Murphy [95] used the seed-mediated method as the alternative route to the template method. In this method as summarized in Figure 8, gold seeds of size 3–4 nm are initially synthesized by chemical reduction using a strong reduction agent (sodium borohydride) in the presence of a capping agent (citrate). Following the step, the seeds are added to the solution of more metal salt, a weak reducing agent, and a surfactant directing agent (CTAB). The method generally results in monodisperse, stable gold nanorods with different aspect ratios.

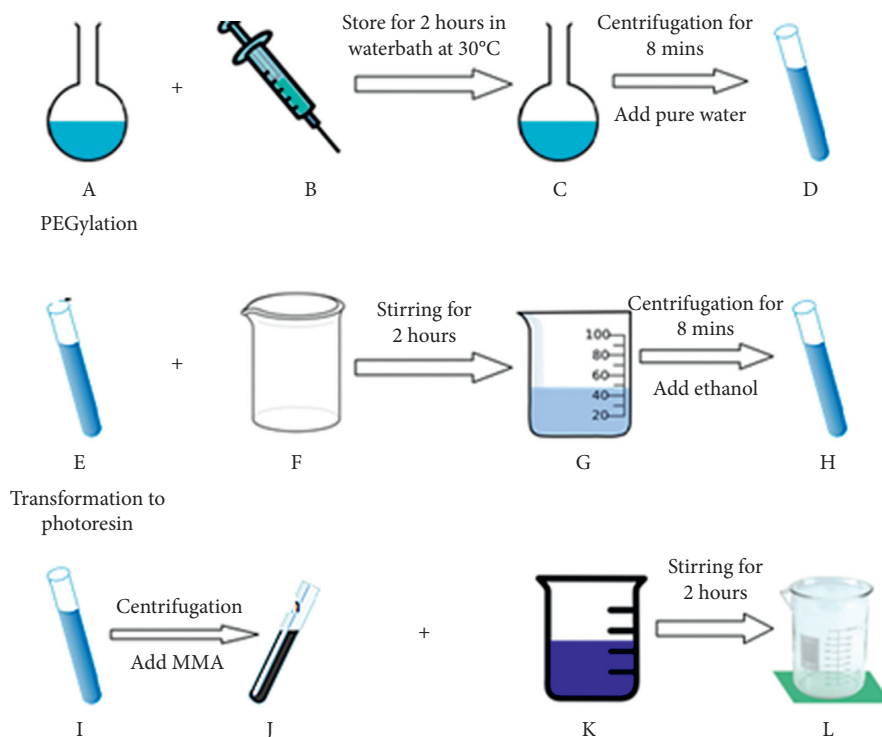


FIGURE 6: (A) Growth solution: CTAB, HAuCl_4 , AgNO_3 , H_2SO_4 , and AA. (B) Seed solution: CTAB, HAuCl_4 , and NaBH_4 . (C) Gold nanorods dispersed in pure water. (D) Redispensation of gold nanorods in pure water. (E) Gold nanorods dispersing in water. (F) mPEG-SH solution. (G) Gold nanorods wrapped in mPEG-SH, in pure water. (H) Gold nanorods wrapped in mPEG-SH, dispersing in ethanol. (I) Gold nanorods dispersing in ethanol. (J) Gold nanorods dispersing in MMA. (K) Photopolymerisation resin. (L) Gold nanorods dispersing in photoresin.

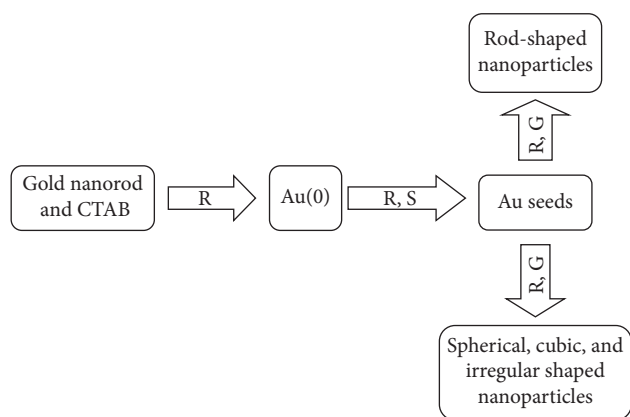


FIGURE 7: Illustration of the template method to synthesize gold nanorods and reduction of Au(I). S, forming Au seeds; G, growth of the Au seeds [93].

Moreover, another alternative green method is to synthesize gold nanorods without the use of CTAB but using gelatin as a capping agent. The method was reported to have resulted in higher media stability and enhance photostability [96].

2.1.4. Characterization of Gold Nanorods. Recent developments are done to develop nanotechnologies for specific drug delivery and multimodal activities [96–98]. Due to their distinctive chemical, physical, and photonic properties, AuNRs

have been used for cancer therapy, diagnostic, and therapeutic applications. Ali et al. stated that the low resolution from some equipment limits crucial information about the nanorods [99]. Many researchers characterised the nanoparticles by using electron microscopy (EM), atomic force microscopy (AFM), dynamic light scattering (DLS) and static light scattering (SLS), X-ray diffraction (XRD), and the Fourier-transform infrared spectroscopy (FTIR), as indicated in Table 1 [112–114].

Table 1 shows the properties, synthesis, characterization, and applications of AuNRs.

2.1.5. Application of the Gold Nanorods. A lot of studies indicated that the nanoparticles have the potential to be consumed as drug carriers, used in the detection and treatment of tumors, monitoring of treatment response, and to guide the therapeutic regimens [115, 116]. In addition, plasmon resonant gold nanorods are used as multifunctional agents for image-guided therapies. As the carriers, they promote the circulation time of drugs to take longer and improve and ensure that drugs do not degrade until they reach their targets. In the event of a tumor, the drug would land on the target still effective for an uptake through permeability, retention effect, and the receptor-mediated endocytosis. The large surface to volume ratio of nanocarriers helps them carry agents for chemotherapy, anti-angiogenic, or gene therapy delivered to tumor sites for enhanced treatment [117, 118].

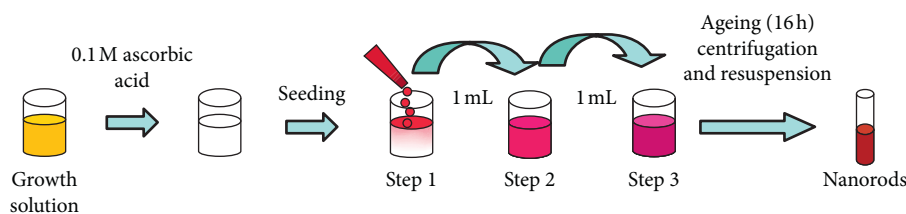


FIGURE 8: The general scheme used for the three step seed-mediated method [95].

TABLE 1: Summary of properties, synthesis, characterization, and applications of AuNRs

Synthesis	Characterization	Limitation	Application
Wet chemical synthesis [100–102], template method [103–105]	SEM, TEM	Limited yield due to the CTAB bilayer	Sensing, photothermal therapy, imaging
Electrochemical route [106–108]	TEM	N/A	Sensing, photothermal therapy, imaging
Seed-mediated method [109–111]	TEM, ETEM	N/A	Sensing, photothermal therapy, imaging

The versatility of nanoparticles enables them to be used in various ways. For example, in therapy, one way is for the treatment of ovarian cancer. This required a coat with thioglucose, sensitizers to produce ROS to damage the cancer cells [119–122]. The use of the nanorods over nanospheres or any other nanoshapes is also due to narrower line widths at more or less the same resonance frequencies because of reduced radiation-damping effects. In fact, nanorods have the ability to emit two-photon luminescence (TPL) signals that are used for single photon detection, which is suitable for biological imaging purposes [123–125].

The use of AuNRs however has a few challenges. For example, under intense illumination, the shape of the rod changes to a nanosphere and longitudinal NIR resonance loss. This normally happens in the photothermal therapy. A coating with thermally stable material was effective to overcome the shape change [100, 101, 126]. The modifications of gold nanoparticles for clinical value are of great importance, taking note of uptake and cell targeting. Theranostic systems have been founded from many building blocks, including hybrid, organic and or inorganic nanoparticles, superparamagnetic iron oxide, and plasmonic gold nanoparticles, which have been extensively studied due to their unique physical properties. The hybrid of these particles can be used for *in vitro* or *in vivo* imaging, magnetic targeting, and photothermal therapy. The coating/passivation or functionalization of the gold nanoparticles has currently received enormous attention due to more multifunctional competitive applications [102, 103].

2.1.6. Passivation of Gold Nanorods. The significance of magnetic nanoparticles has been growing due to use in more important and exciting applications like use in biomedical diagnosis, catalysis, and photoluminescent materials. Due to their unstable nature to the surrounding environment, their applications are inhibited as they readily oxidise on exposure to air due to their large surface areas. Methods have been developed to increase oxygen resistance on metal nanoparticles such as plasma spraying, phosphating, and electrolyte deposition [104–107]. Consideration has to be made

when encapsulating metal nanoparticles so as not to increase the thickness of the metal and hence interfering with the bulk metallic properties of the metal. One of the most ideal methods of passivation is the atomic layer deposition (ALD) method, which provides an ultrathin layer on the metal nanoparticle [108–111]. This method however does not guarantee the conformation of the geometry of the nanoparticles, hence only excellent for application in which precise morphology is not required [127, 128]. One of the methods used for the stabilization of metal nanoparticles is the attachment to dendrimers, ensuring and controlling the stability, size, and solubility of nanoparticles in a range of less than 1–5 nm diameter [129–131]. Examples of dendrimers include PAMAM and PPI as shown in Figure 9 [132]. Ligands and polymers have been used for the past years for the stabilization of metal nanoparticles and their application in catalysis and biocatalysis; in particular, their use with gold nanoparticles has been reported to yield excellent results.

The optical properties of AuNRs can be changed by slight changes in the shape or size of the nanorod; hence, it is important to ensure a stable environment for them. Sensing, imaging, and biomedical applications of AuNRs all come as a consequence of the ability of the AuNRs to be tunable, which is attributed to their optical properties contributed by their rod-like shape. Murphy et al. elaborates on the interfacial chemistry of nanorods [133]; three interfaces occur on the surface of nanorods, the gold-surfactant interface, hydrophobic surfactant bilayer, and a surfactant interface. These interfaces help in altering the nanorod properties in terms of stability, against aggregation and toxicity and how easily they can be assembled. Lastly, the solvent accessible interface provides a platform or directs how the nanorod can interact with other particles, macromoles, and living cells [134–136].

A common method of the synthesis of AuNRs uses CTAB, which makes the functionalization of AuNRs using ligands a challenge. During the synthesis using this method, a double layer of CTAB is formed for the passivation; this layer is a problem in terms of bioconjugation, adsorption of DNA nonspecifically, cytotoxicity, and the stability of the

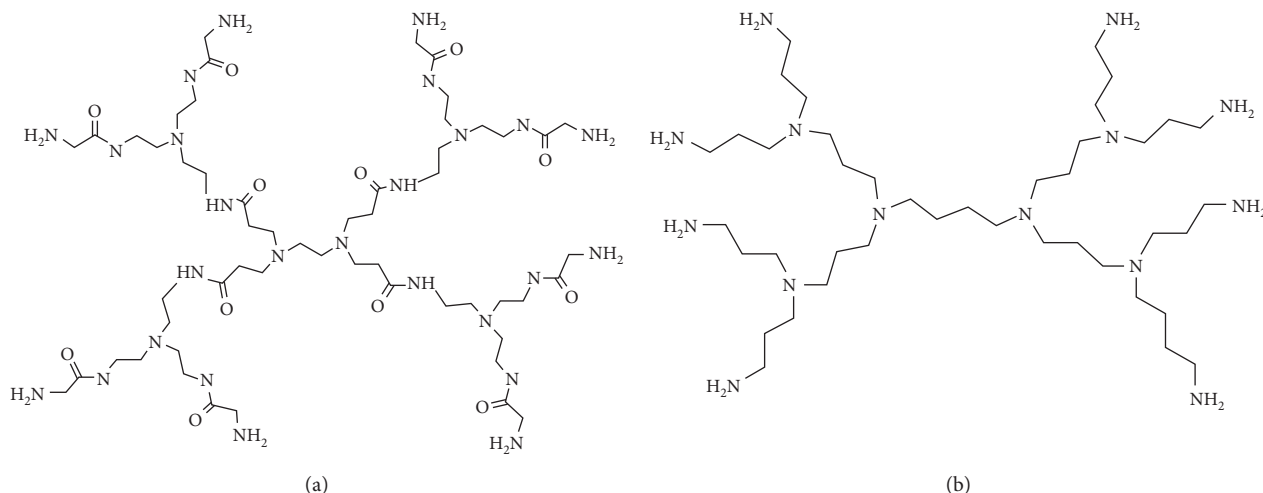


FIGURE 9: (a) A G1 PAMAM dendrimer and (b) G1 PPI dendrimer.

nanorod and hence limits the use of these AuNRs for biological applications [137–140]. Absorption associated with AuNRs enables them to be used for dark-field light scattering observation or biological media. Even though CTAB is very toxic to the biological environment, complete removal from the AuNRs would yield unstable AuNRs in colloidal dispersion forming aggregates. To obtain functional AuNRs, CTAB has to be replaced by amphiphilic molecules. Cationic cerasone-forming lipids and cationic nonsilylated lipids are used to passivate AuNRs [141]. Moreover, the amphiphilic molecules were added as to overcome the shortcomings brought by CTAB especially the fact that it has high cytotoxicity preparation of phosphatidylcholine passivated AuNRs (PC-AuNRs) which have low cytotoxicity and efficiently used as photosensitizers using pulsed light [142]. The pulsed light however reshapes the nanorods into nanospheres, hence no absorption spectra at the near-IR region. This helps in the damage of only tumor cells not healthy cells even with successive irradiation. The combination of photosensitizers and light induces the selectivity for tumors from healthy cells and also use of light produces heat just around the photosensitizer, hence the destruction of cells. Figure 10 [119] shows the absorption spectra of a PC-AuNRs in solution. It shows a normal NR spectrum with two SP peaks: ~ 900 nm for the LSPR and ~ 520 nm for the TSPR.

Ferric acid at room temperature as an etching agent for AuNRs is also used to passivate gold nanorods. The method used decreased the length of the AuNRs but not their diameter due to the oxidation of AuNRs by ferric ions, the shortening of only the length helps in the provision of the nanorod's desired aspect ratios and selective optical and also selectively removes other nanostructures. The shortening in length of the nanorods was monitored by TEM and the UV-vis absorption spectroscopy, and the results showed the reduction of electron potential of the gold species by halide ions and acceleration in oxidation of the AuNRs by ferric ions [121].

Other ways to encapsulate the particles include polyelectrolyte coatings such as poly(diallyldimethylammonium

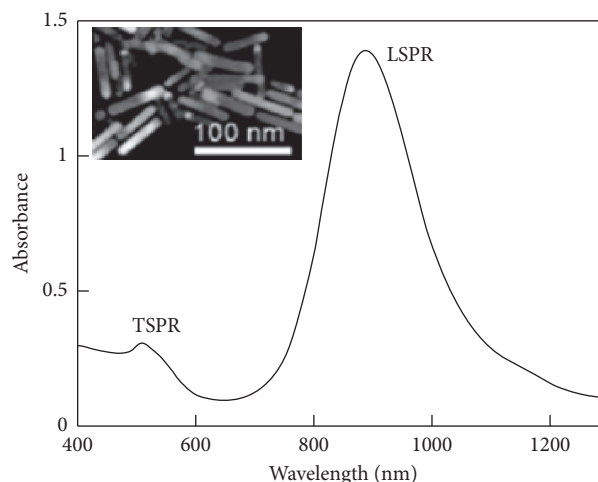


FIGURE 10: PC-AuNRs absorption spectrum, 0.5 nM Au nanorods (TEM image of PC-NRs).

chloride) (PDADMAC), poly(4-styrenesulfonic acid) (PSS), poly(acrylic acid) (PAA), or poly(allylamine) hydrochloride (PAH); these molecules reduce the interaction of CTAB with cells. PEGyated AuNRs are also used, which to some extent replace the CTAB in the molecule hence reducing toxicity [122]. Figure 11 illustrates the encapsulation with polymer-based molecules.

The treatment of particles with PEG-SH replaces the CTAB in the molecules which is toxic, replaces it with PEG (polyethylene glycol), and it also enhances the particles to be more stable under different conditions. Takahashi et al. stated that the PEG as a linker to attach antibodies is still a field to be explored further [91]. Lastly, a hydrophobic polymeric precursor, polyvinyl acetate (PVA) which changes into polyvinyl alcohol which is hydrophilic, is used to replace the nonbiocompatible and toxic CTAB [143].

AuNRs perform adequately in their retained shape; hence, a suitable coat has to be used for such. CTAB is the main inhibitor in AuNRs application due to instability; hence, thiol monolayers are used which provide for better

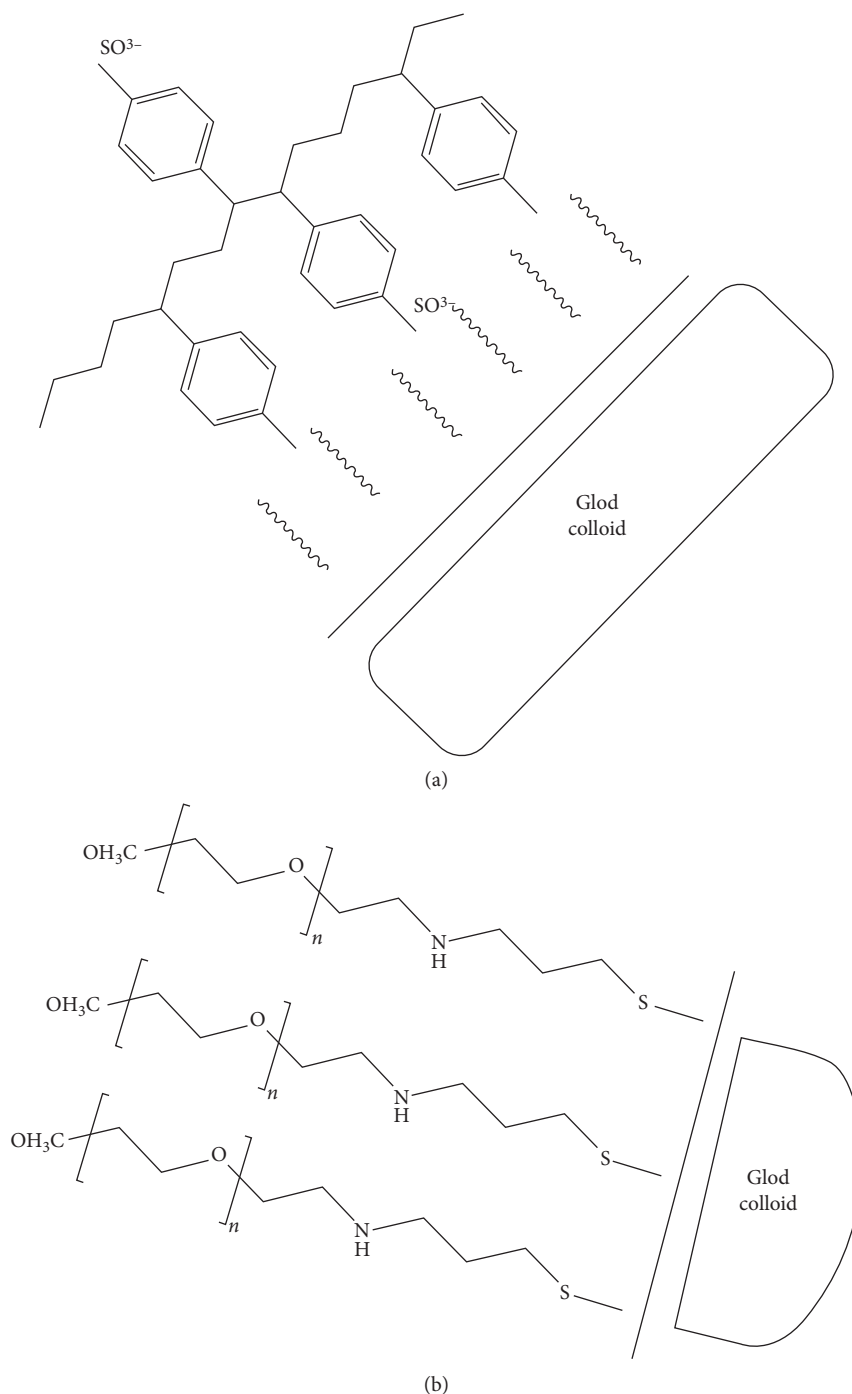


FIGURE 11: Surface modification of AuNRs. (a) PSS, encapsulation occurs on the CTAB bilayer through electrostatic adsorption. (b) mPEG-SH, replacement of the CTAB by PEG by the gold sulphur binding [123].

stability and organic media for compatibility [144]. Polymer (including porphyrin) coated nanorods are used as either ionic or cationic molecules with different charge densities, and these also give a promising future as biological delivery agents [145, 146].

2.1.7. The Replacement of Quantum Dots over Gold Nanorods. Quantum dots (QDs) are nanocrystals with semi-conducting properties. These materials comprise elements

from Group II to VI or III to V, and their sizes range from 2 nm to 19 nm. QDs such as AuNRs have unique optical and chemical properties [147]. These materials have an excellent future in biomedical imaging and detection; however, due to the heavy metals and colloidal instability, these pose limitation in terms of their use in diagnosis and therapy for both cancer and other diseases. Moreover, QDs still have questions pertaining to their cytotoxicity and their size increase after coating. The materials also show impressive results in imaging tumor neovasculature which

is done late after clinical diagnosis of the development of the cancer. Gold nanorods have very high absorption spectra in the NIR; this is where light penetration into the tissue is very high, around 10 cm deep. Gold nanorods relative to quantum dots are easy to synthesize, they have tunable optical properties, and they can also be multifunctional [11].

2.2. Porphyrins. Porphyrins and metalloporphyrins are tetraazamacrocyclic compounds found in nature, and the compounds participate in important biological processes such as photosynthesis. Importantly, the ability of a porphyrin photosensitizer to be located in a biological medium depends strongly on their peripheral substituent groups as well as their axial ligand; the two factors determine solubility, chemical affinity, redox potential, and other properties [148–150].

2.2.1. Synthesis of Porphyrin and Characterization. Porphyrins exist as two types, β -substituted and mesosubstituted porphyrins, as illustrated in Figure 12 [151], where the β -substituted porphyrins mirror naturally occurring porphyrins while mesosubstituted are synthesized. Meso-substituted porphyrins are widely used in biomedical and material chemistry [152].

The synthesis of the mesosubstituted porphyrins from nonporphyrins follows basic steps which are seen in the methods described here. The first synthesis was performed by Rothermund in 1936, and the method starts with the reaction of an aldehyde and pyrrole with both reactants at high temperature and concentration in a bomb reactor with no added oxidant. The addition of zinc acetate to the reaction only increases the yield of tetraphenylporphyrins twofold, but this is not observed with any other porphyrins [153–157].

Another method was developed by Adler and Longo so as to modify the Rothermund method by increasing the yields. The method was developed around the 60s and uses high temperature and concentration, but unlike the Rothermund method, it takes place in the presence of air under reflux in propionic acid. The method allows for synthesis of various porphyrins at relatively higher yields but polymerized pyrrole yield also increases, hence reduced porphyrin yields because it contaminates the product. The porphyrin can be isolated in two ways: treating with DDQ (2,3-dichloro-5,6-dicyano-1-4-benzoquinone) and refluxing with toluene or by using column chromatography. Scheme 1 illustrates the synthesis by the Adler–Longo method [158]. Figures 13 and 14 show a reported method scheme as performed by Lindsey in 1987.

The Adler–Longo reaction is used to react substituted benzaldehydes with pyrroles to produce the corresponding porphyrins in yields of about 20%. The method provides crystalline and pure product relative to the Rothermund reaction product, and it can also be used with different aldehydes to produce porphyrins but it poses certain problems; the high temperatures and concentrations hinder the synthesis of porphyrins with sensitive functional

groups [144, 160, 161]. The reaction method as established by Alder, Longo, and Shergalis shows that before the porphyrin is cyclised, it goes through a carbinol step as shown in Figure 15 [162].

Due to the drawbacks brought by the Adler–Longo method, the Lindsey method was developed. The method synthesizes porphyrins under milder conditions and is a two-step one-flask reaction. The method enhances the Adler–Longo method, in that porphyrins that cannot be synthesized by the latter can be synthesized by the Lindsey method; this method can yield porphyrins from some sensitive aldehydes. In this method, pyrrole, benzaldehyde, triethylorthoacetate, and a water quencher are stirred at room temperature at equimolar concentrations with boron triflate in dichloromethane (DCM). After an interval of 30–40 minutes, formation of porphyrinogen occurs. DDQ is added to oxidise the porphyrinogen to produce the porphyrin in a yield of 30–40%. Figure 16 [163] shows the reaction path from an aldehyde to a porphyrin. Figure 14 illustrates a reaction scheme for the formation of porphyrinogen [164–171].

2.2.2. Modifications in the Structure

(1) First-, Second-, and Third-Generation Photosensitizers. Photofrin has intrigued synthesis of a lot of other photosensitizers such as photoheme which is used to treat lung, skin, and breast cancer, and these are the first-generation photosensitizers [172–184]. Second-generation photosensitizers were developed so as to improve on first-generation photosensitizers. These photosensitizers are chemically pure and absorb light at around 650 nm, and they have relatively less skin photosensitivity. A lot of research has been conducted and shows that the sensitising efficiency of a compound increases with decreasing polarity [185]. The use of Photofrin has triggered the synthesis of more porphyrin-based photosensitizers mainly to produce the ideal photosensitizer. Numerous new porphyrin-based photosensitizers have been produced: first- and second-generation photosensitizers including hematoporphyrin monomethyl ether (HMME), photocarcinogen (PsD-007), second-generation hematoporphyrin derivatives (HiPorphyrin), and 5-ALA (aminolevulinic acid hydrochloride) [186]. The activation of a porphyrin and its derivatives by light triggers its relaxation to the ground state in three ways: nonradiative decay, emitting a photon, or by the transfer of energy [187–189]. Photofrin, which is the purified form of hematoporphyrin, was the first accepted photosensitizer for PDT, for the treatment of various cancers. Photofrin has certain properties required for an ideal photosensitizer but it has a few challenges; it has a weak long wavelength at 630 nm which is below the wavelength for maximum tissue penetration of deep tumors; secondly, it has a long photosensitivity to the skin. The surface of porphyrins can also be modified by different methods including surfactant-resistant, ionic, mixed porphyrin, sonic cation-assisted, and metal coordination self-assembly [190–192].

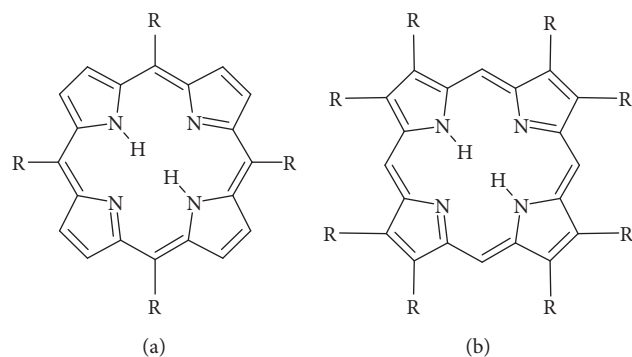


FIGURE 12: Structures of the (a) mesosubstituted and (b) β -substituted porphyrins.

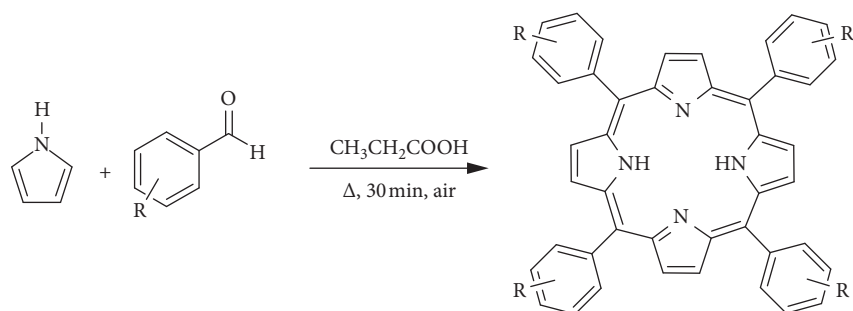


FIGURE 13: Synthesis of mesosubstituted tetraphenylporphyrin in open air [158].

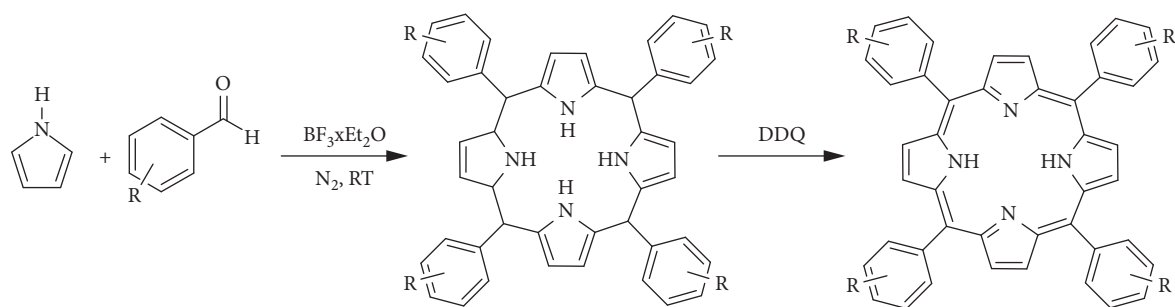


FIGURE 14: Schematic method as by Linsley in 1987 [159].

2.2.3. Modification of Porphyrins. Porphyrins can be modified by expanding their pi-electron conjugation. This is achieved by increasing the heterocyclic rings or bridging carbons around the porphyrin framework. The resulting chromophores absorb strongly in the red region, 650–800 nm as opposed to the 18 pi-electrons in normal porphyrins [193–195]. Modification to the porphyrin structure enhances the absorbing ability of the porphyrin; for example, the reduction of the ring produces a ring of chlorine which absorbs around 660 nm in the red spectra. This allows for the modulation of the light-activated analysis [196].

2.2.4. Porphyrin in Photosensitization. The use of two-photon imaging over conventional imaging has many advantages including the deeper light penetration, 3D image capability, low background fluorescence, and reduced damage to the surrounding tissue. The use of gold nanorods

as contrast agents of two-photon imaging is very essential due to the fact that they exhibit high two-photon luminescence which is 58x more than that of a rhodamic molecule [197, 198]. In the two-photon imaging, mesoporous silica nanoparticles have been used, so as to evade the shortcomings of photosensitizer delivery systems; they are used as carriers in chemical catalysis, drug delivery, and cell labelling. Mesoporous silica nanoparticles have high pore volume and surface area, hence ease for the production of singlet oxygen which is easily released from the matrix. The structures also show high biocompatibility and tolerance to most organic solvents; these compounds are also easily functionalized for targeting tumor cells *in vivo* [199–206].

2.2.5. Application of Porphyrins. The application of porphyrins is based on their peculiar characteristics including that they have rigid and planar geometries, photothermal,

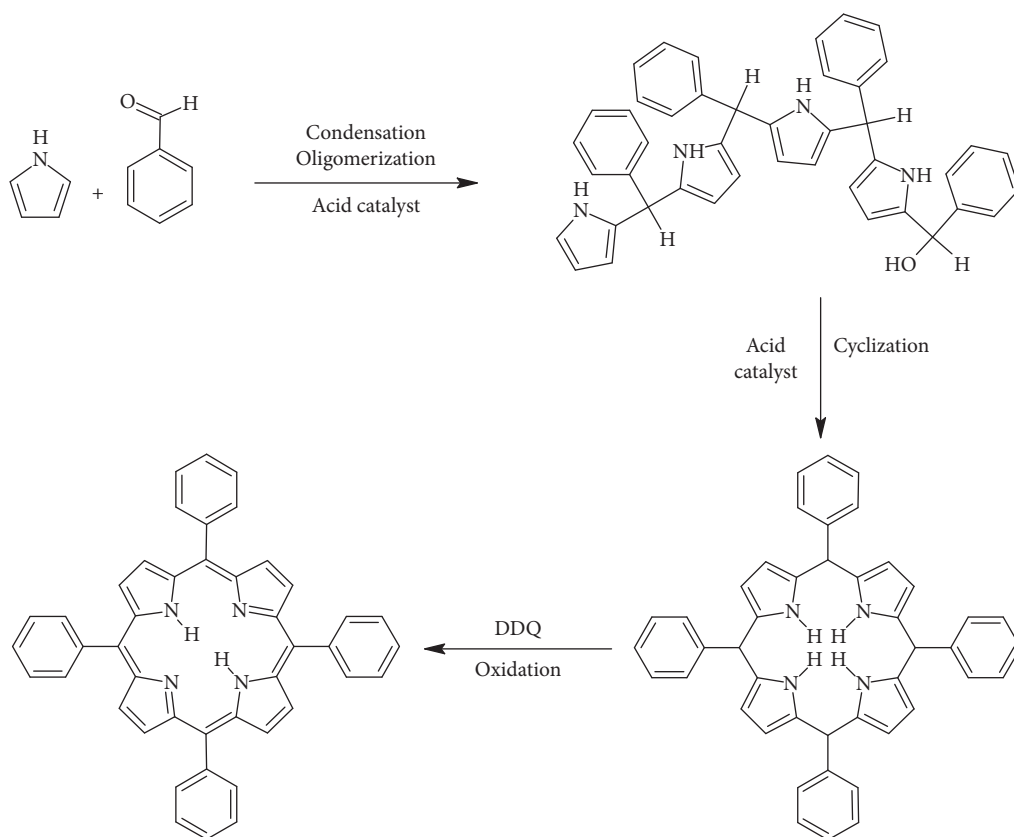


FIGURE 15: Reaction transformation from an aldehyde to a porphyrin.

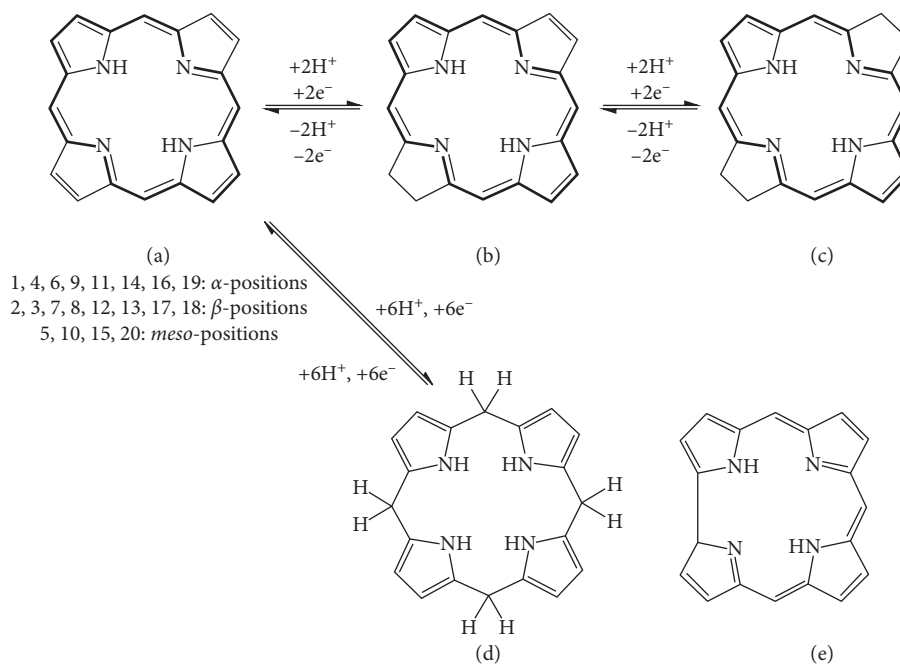


FIGURE 16: (d) The tetrapyrrole derivative porphynogen, a synthetic precursor of the porphyrin ligand. (a) Porphyrin. (b) Chlorin. (c) Bacteriochlorin. (d) Porphyrinogen. (e) Corrin.

and spectroscopic properties that can be tailored readily; they are multifunctional, biocompatible, and serve as electron donors [207, 208].

Photodynamic therapy is used for the treatment of tumor and malignant diseases. Its basis is on the administration of the photosensitizer which is followed by

irradiation of light at a specific wavelength, and this results in reactive species like radicals or singlet oxygen which ultimately destroy the tumor cells; this phenomenon is illustrated in Figure 17 [209]. The selectivity of PDT depends on the concentration of the photosensitizer, on the normal and tumor cells, and on the exposure to light of the side being treated [210].

New antibacterial approaches include the use of photosensitizers activated with visible light; as in PDT, the photosensitizers accumulate in microbial cells to induce phototoxic reactions. The excited PS in the triplet state in the presence of oxygen induces the production of reactive oxygen species (ROS), which then continually induces further reactions in the bacterial cell wall and the lipid membranes. Maich [211] reports that a specific photosensitizer with the ability to only target the bacteria without causing harm to the surrounding environment is still a challenge [212].

The low-dark toxicities of porphyrin macrocycles and the fact that porphyrins selectively localize on a wide range of tumors have led to them being one of the best photosensitizers relative to other macrocycles. Porphyrins and compounds based on porphyrins strongly absorb in the visible part of the optical spectrum; they are noncytotoxic in the dark, have high chemical stability, have high affinity for serum proteins, can be modified to have favorable pharmacokinetic properties, and are stable when complexed with different metals but still maintain their *in vivo* tumor localization properties [213, 214].

Most photosensitizers used in PDT and fluorescence diagnosis (FD) are based on the porphyrin structure because generally porphyrin accumulates on cancer cells as opposed to the surrounding nearby healthy cells and because their fluorescing properties are used for the detection of the cancer cells. Therapy resulting from the photosensitizers is based on the production of singlet oxygen when the photosensitizer is exposed to light. A major setback however for PDT is that it does not work best on distal metastasis [215].

Porphyrins and their analogs are adopted in many nanotechnologies due to their supramolecular design, flexibility, robustness, and unique photophysical and chemical properties and are used in technologies such as catalysts, sensors, molecular electronics and solar energy upconversion [216]. Porphyrins also have a visible light absorption and synthetic versatility, thus can be applied to the optoelectronics and act as photosensitizers [217]. Engelmann et al. [218] demonstrates the use of two cationic porphyrins to understand factors influencing the binding of the porphyrins to liposomes and mitochondria and how efficient their photodynamic reaction is in entocytes. The results showed that binding and photodynamic efficiency were inversely proportional to the number of positively charged groups but directly proportional to *n*-octanol/water partition coefficients [219, 220].

2.2.6. Encapsulation of Gold Nanorods with Porphyrins.

The use of nanoparticles has been very impressive, but when used by themselves, they pose certain shortcomings; for example, when entering the blood system, macrophages

and phagocytes readily passivate them leading to accumulation in the liver, spleen, and lymphatic system; this leads to toxicity, causes oxidative stress, and draws out an immune response [220]. Theranostic agents to be developed have to be the ones that contain both therapeutic and imaging properties. A lot of chemotherapeutic agents cannot be used as theranostic agents because they comprise less drug than the imaging doses; moreover, they are not tumor specific; antibodies on the other hand have the ability to be used as theranostic agents but they are relatively expensive. Porphyrins and derivatives however have the ability to fluoresce when excited by light, hence allowing for imaging before or after therapy; these molecules also allow for the attachment of tumor-targeting moieties at peripheral positions for the development of tumor-specific agents [221].

Porphyrins have a very rigid highly stable macrocyclic structure; hence, they are used as ligands in chemistry and biology; moreover when coordinated to metals, they give new characteristics. Metal-porphyrin chemistry is very important in biomimetic and chemical applications [222, 223]. The removal of the internal protons from the “free base” porphyrin gives a tetradentate chelating dianion molecule with an ability to coordinate a metal at the central cavity, as demonstrated in Figure 18. Research reports show that porphyrins can coordinate with lanthanides, some actinides, and a few main group metals [224].

Metals bounded to porphyrins can be in a range of oxidation states; $-2, -6, (d^0 - d^{10})$ spin; $S = 0$ to $S = 5/2$; and a coordination ranging from 4 to 8. Restriction on the metal coordinated with the porphyrin confines them to only have two mutually *trans*-coordination sites as shown in Figure 18 [225]. Peripheral substitution or enhancement of porphyrin is an approach for modifying the steric and electronic factors of metals but rearranging the meridional or facial types is not possible.

The versatility of the porphyrin ligand has led to a very large number of designed porphyrin complexes. Paths followed for the insertion or $M-L_n$ fragment have been explored and depend on the nature of the metal source. Reactions of porphyrins and metals can be of different paths which basically depend on the previous oxidation state of the metal and coordination, reduction, or oxidation of the metal.

The noncovalent interaction of porphyrins and their derivatives are very important in biological systems. Derivatives that possess a positive charge on the mesoposition interact well with DNA, nucleotides, and a lot of aromatic substrates [226]. Neutral porphyrins are reactive but the dications are inert. Metalloporphyrin reactions depend on a number of factors which influence the nature of the reaction, the rate at which the metal incorporates is decreased by the electron withdrawing group in the β -substituent and the solvent in use also influences the reactions [227].

Two photon-imaging systems have attracted a lot of research due to a number of advantages including the fact that there is enhanced penetration of light in the tissue or cells. The systems also have low fluorescence, and the photodamage on the living cells is reduced with their use. Using photodynamic therapy to treat cutaneous malignant

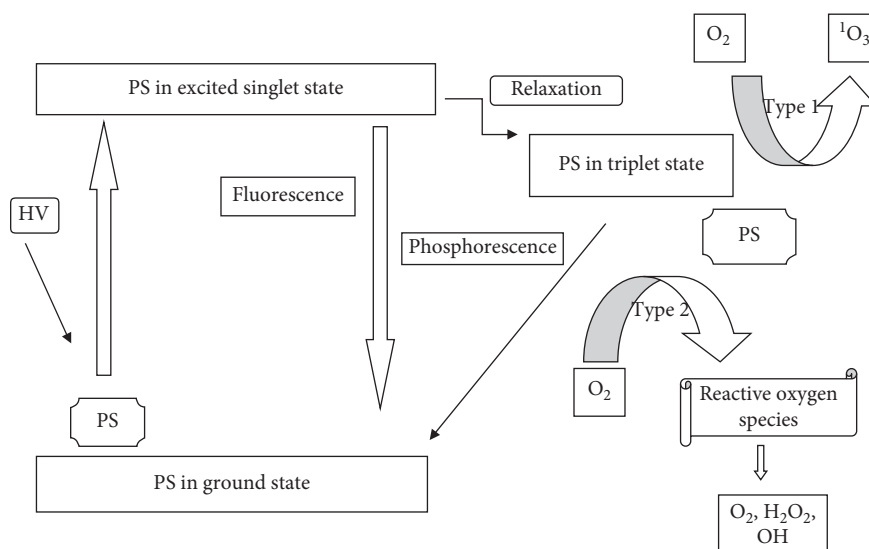


FIGURE 17: Schematic representation of a photosensitizer before and after excitation.

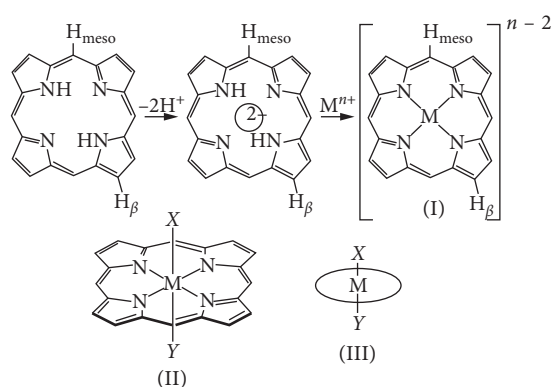


FIGURE 18: Schematic mechanism of a porphyrin complex presenting a tetra-coordinated metal in its tetrapyrrolic core. (i) Metal *trans*-coordinated to two ligands. (ii) Simplified presentation.

and intraperitoneal tumors has been researched to be one of the excellent pathways to use. The method however uses photosensitizers which currently have a lot of pitfalls, problems that can be solved by development of a new improved photosensitizer or enhancing the already existing ones. The aim will be to increase the cytotoxicity, selectivity, and protection against degradation.

The encapsulation of porphyrin with silica or an additional entity protects it from biological degradation. Multifunctional nanoparticles have been shown to produce more singlet oxygen than the porphyrin on its own. An example is illustrated by Zhao et al. [228] by using a two-photon imaging system with high photosensitisation and varying different image contrast agents, photosensitizers, and carriers. Here, a multifunctional nanocomposite (AuNRs/mSiO₂/HP) was designed and used. The nanoparticles comprise a gold nanorod core with a porphyrin-doped mesoporous silica shell. Results included the fact that the triplet oxygen production was enhanced with the addition of the silica coating as opposed to pure porphyrins. These were

monitored by the incorporation of ADDBA (anthracenediyl-bis(methylene)dimalonic acid), which readily reacts with newly produced singlet oxygen to produce endoperoxide. Endoperoxide on the other hand decreases the ADDBA absorption which is around 350 nm. The absorption spectra of the composite material embedded with ADDBA can be seen in Figure 19 [228].

Research has shown that enhancing the surface of gold nanorods enables them to be versatile, hence can tune their properties to desired use. Using alkyl thiol to encapsulate the gold not only anchors the nanorod but also enhances the stability of the nanorod and fills the space potentially between the gold nanorod and bulky porphyrin molecule so as to provide space to insert C₆₀ fullness molecules which increase stability to the porphyrin-gold nanorod hybrid structure. Work done by Xue and colleagues [229] showed that protection of nanorods with a monolayer of thiols and porphyrins has very interesting result.

Modification of gold nanorods can also be done by incorporation of a porphyrin with an antitumor drug, for example, doxorubicin (DOX) used usually for the treatment of a number of cancers. The goal is to increase and improve on the multidrug resistance, nanotargeted delivery, and the toxicity of the drugs. Multifunctional nanocomposites of meso-tetrakis-(4-sulfonatophenyl) porphyrin (TPPS), gold nanorods, and DOX (DOX@TPPS-AuNRs) were synthesized by Bera and his colleagues [230]. The resulting composite material showed improved cellular uptake by the cells and showed no cell toxicity. Figure 20 [231] shows UV-Vis spectrum of the composite which significantly shows the SPR (surface plasmon resonance) of TPPS-AuNRs at 523 nm, and one of the pure AuNRs is at 525 nm in aqueous solution. The noticeable peak broadening was observed due to the presence of porphyrin which meant that there was a strong association of the gold surface to the porphyrin.

The excellent imaging and sensing properties shown by AuNRs qualify them as excellent for photothermal therapy; on the other hand, porphyrin and derivatives show excellent

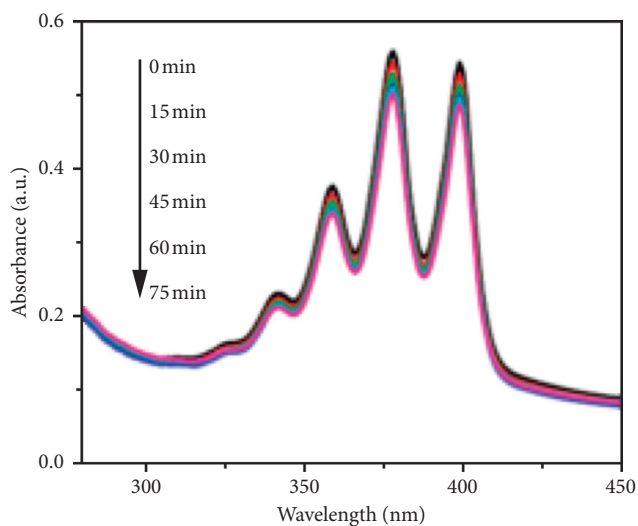


FIGURE 19: Graph showing a decrease in absorption of a composite AuNRs/mSiO₂/HP with ADDBA with increased illumination time.

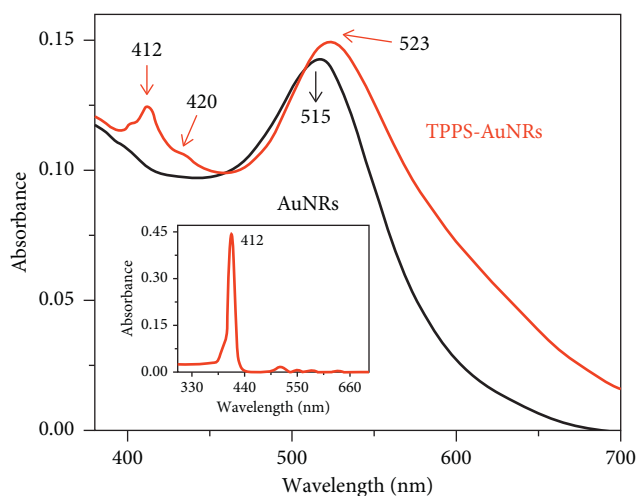


FIGURE 20: UV-Vis spectrum of TPPS-AuNRs and AuNRs.

properties as photosensitizers due to the fact that they have more than one absorption band in the near-infrared region; hence, they can be manipulated to be used to penetrate deeper into the tissue for photodynamic therapy. Photodynamic therapy shows minimal invasion to tissues while photothermal therapy is specific to tumor cells or affected area. Photodynamic therapy limitations are still major issues up to date especially based on singlet oxygen production, selectivity to the target tissue, and the concentration so as not to damage the normal cells [231].

The agglomeration of AuNRs with porphyrins is to increase the specificity and targeting. The system incorporates two modalities: the photothermal and photodynamic stability which will hence increase the chance of tumor and bacterial diseases to be treated. Incorporation of PTT and PDT as a dual technology for cancer and bacterial infections is likely to ensure selectivity and improved efficacy to the system. This study researches and further experiments

on both areas with a view to finding a more specific and less invasive combination technology.

2.3. Phototherapy

2.3.1. Applications of Photodynamic Therapy. Photodynamic therapy (PDT) is the use of a photosensitive agent on tissues to treat cancer or bacterial infections followed by photo-irradiation. This is a clinical treatment used in different diseases [12, 232–234]. It takes place by the administration of a photosensitizer in the body of a patient to accumulate on the tumor. The tumor is irradiated with biothermal light (500–650 nm), leading to an excited photosensitizer [149, 235]. Combination of the excited triplet photosensitizer and molecular oxygen results in a singlet oxygen [¹O₂] which is the main mediator of the destruction of the cell induced by PDT. The generation of the [¹O₂] has a very short life span and a limited diffusion rate, resulting in photo-oxidation of the tumor [236, 237]. A lot of photosensitizing agents have been formed but they have not been tested clinically due to factors such as (a) poor selectivity in terms of the target tissue and the healthy tissue, (b) the absorption spectra at short wavelengths, and (c) high accumulation rates in skin [236].

The use of photodynamic therapy extends to research of quantum dots, porphyrin, and micelles as photosensitizers. The discovery of PDT in the 1900s erupted as a cancer, HIV coronary heart, and psoriasis treatment agent [237–239]. PDT is a modal system which provides binary selectivity, which is accomplished by the increase in accumulation in the target tissue and limiting irradiation. The photosensitizer used in PDT has to be cytotoxic to damage only the affected tissue. An enhancement can be made by attachment to the part which has high affinity to the target tissue [240–243].

The tendency of a photosensitizer used in PDT to accumulate only in cancerous cells or tissue and not on the normal tissue is apparently explained by two reasons. It is taken up mostly by hyperproliferative cells than by normal resting cells, and their uptake by neovascular endothelial cells are accelerated, both of which characteristics can be seen in solid tumors [244, 245].

Photodynamic therapy is used to deactivate microorganisms by not leading to the selection of mutant-resistant chains compared to traditional antibiotics [246, 247]. The use of PDT has been improved by the incorporation of nanoparticles which have been used as delivery agents of the photosensitizer or to improve the inactivation kinetics.

Inorganic nanoparticles such as TiO₂ can also be used and have the capability of inactivating microorganisms [248]. Various developments of tumor target photodynamic therapy have been made; but until now, there is no universal approach due to spontaneous properties of tumors. For the ideal PDT development, the PDT has to be tailored, have an appropriate target strategy, and carefully select the tumor type and the stage of the disease [249].

Photodynamic therapy fundamentally relies on the accumulation of a photosensitizer of a tumor cell or tissue after administration [250, 251]. This phenomenon involves the

production of reactive oxygen and free radicals which are cytotoxic, and the main source of the photobiological activity is however the [$^1\text{O}_2$] which causes damage of cells either by apoptosis or necrosis [252, 253].

The use of supraparamagnetic iron oxide nanoparticles as MRI contrast agents for brain tissue has been proven very effective. Nanomaterials are used as delivery agents across the blood-brain barrier and to specifically transport drugs to cellular compartments such as the nucleus [254, 255].

Antimicrobial photodynamic therapy (aPDT) has been an effective treatment to damage bacteria. Enhancing antimicrobial photodynamic therapy (aPDT) with nanoparticles is a growing field to minimize the use of antibiotics to treat infectious diseases. The technology has many advantages including the increment of [$^1\text{O}_2$] yield of the photosensitizer [256]. A model was made by Hashimoto et al. [257] based on a burned wound and blood stream infection for the verification of aPDT. Two wavelengths were tested *in vitro*, blue and LEDs on a pathogen with resistance to antibiotics and using HB: La^{3+} as a photosensitizer. Experiments were also done *in vivo* on mice; both experiments proved that aPDT can be used in the treatment of bacteria in burned patients using PDT [258].

2.3.2. Mechanism of PDT Cytotoxicity. Singlet oxygen has a relatively short lifetime (see Figure 15), i.e., 810–320 nanoseconds, thereby making its diffusion to be limited to 10–55 nm in cells, the implication being that cells only near the photosensitizer will be damaged [259, 260].

2.3.3. Photodynamic Therapy Using Porphyrin. Photodynamic therapy is the recommended method to treat cancer in the developing countries due to its relative inexpensiveness and user-friendliness [260]. In addition, Chen and Zhang [261] proposed an innovation in which luminescent nanoparticles *in vivo* were used instead of supporting the PDT with external light. The use of these nanoparticles apparently reduces potential damage to the surrounding healthy cells with reasonable costs. The method normally combines both radiotherapy and photodynamic therapy through the attachment of luminescent nanoparticles with photosensitizers such as porphyrin [262]. No external light source is required for this treatment but rather exposure of the photosensitizer to ionizing radiation. The combination enhances the efficiency to damage cancer cells and reduce radiation. Nonetheless, the radiation has to be sufficient enough to produce enough light for PDT, and the prediction of level radiation is usually difficult [263, 264].

2.3.4. Applications of Photothermal Therapy. Photothermal therapy (PTT) is a treatment minimally invasive which occurs by conversion of photon energy to heat energy [265]. The selectivity of the therapy is obtained by the control of incident radiation used and accompanied by the conduct of some proactive molecules or nanoparticles. The photoexcitation relaxation of the particles induces heat transfer to the surrounding affected environment [266, 267].

Nanomaterials with photothermal effects are of great interest to researchers especially in the biological imaging and therapeutics. Nanorods and nanocages possess photothermal properties, hence their use in MRI imaging, infrared thermal imaging, and photothermal ablation of cancer tissues [19, 268–274].

2.3.5. Photothermal Therapy Using Gold Nanorods. Gold nanorods (AuNRs) attracted researchers due to their easy and quick synthesis and simplified bioconjugation, high and strong absorption through their cross-section, and the fact that their optical extinction is tuneable [275–279]. Varying the aspect ratios of the AuNRs influences longitudinal plasmon absorbance shift throughout the visible region [95, 280–285]. AuNRs scatter light through paths of extensive elastic scattering and intensive light scattering of molecular vibrations near the metal surface. The effect is called surface-enhanced Raman scattering (SERS) which is a result of oscillations around the nanoparticle upon radiation [286–289]. These phenomena qualified gold nanorods as chemical sensors as indicated in Figure 21.

In addition, many researchers used gold nanorods as photothermal agents to damage *Pseudomonas aeruginosa*, a Gram-negative pathogen [291–295]. Their work represented gold nanorods which absorbed light at 785 nm and were conjugated to antibodies specific to bacteria. The results positively indicated 75% destruction of infected cells.

2.3.6. Photothermal and Photodynamic Therapy. Recent research has been on nanocomposites based on plasmonic nanoparticles and fluorescent or photodynamic dyes. These are then used for simultaneous therapy and diagnosis [95, 282, 283]. Metal nanoparticles used in the methods are combined with photodynamic drugs using cross-linking procedures or by the use of electrostatic interactions [296–300].

The combination for simultaneous therapeutic purposes was fairly researched. Teretyvk et al. [301] incorporated PDT and PTT by the use of fabricated AuNRs/ SiO_2 -HP composite nanoparticles which successfully decreased large tumor volumes and to damaged solid tumor cells, using the steps (see Figure 22) to fabricate the gold nanorod. On the other hand, Liu et al. [303] synthesized MoS_2 nanosheets which are water-soluble and functionalized them with lipoic acid terminated polyethylene glycol (LA-PEG) to obtain MoS_2 -PEG. These are apparently stable in physiological solutions and also have the ability to be loaded with photosensitizers. Results of the nanocomposites formed using Ce_6 as a photosensitizer in PTT and PDT *in vivo* showed an enhanced tumor necrosis. Jang et al. [304, 305] proposed that for PTT and PDT to work effectively, the distance between the gold nanorod and the photosensitizer must be manipulated. They used an AuNRs- AlPcS_4 composite, and his results proved that the nanocomposite was effective for NIR fluorescence imaging for affected cancer sites and to improve *in vivo* therapeutic efficacy [298, 306–310].

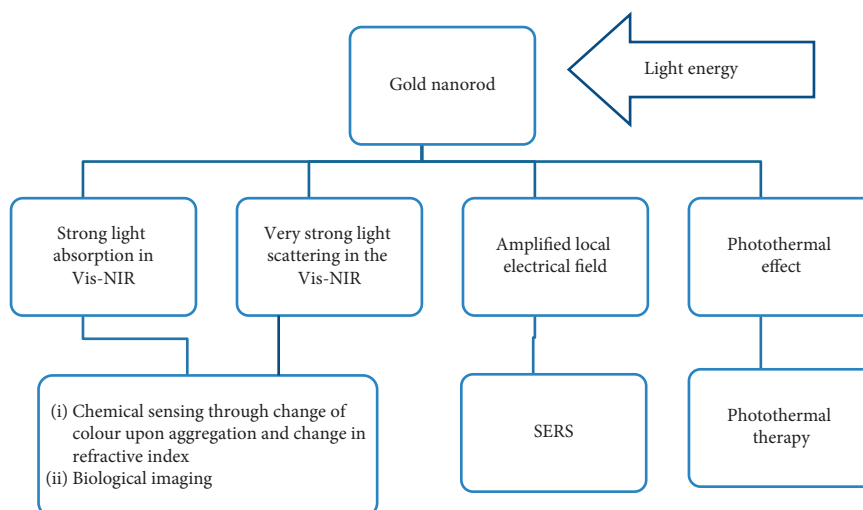


FIGURE 21: AuNRs effects and applications [290].

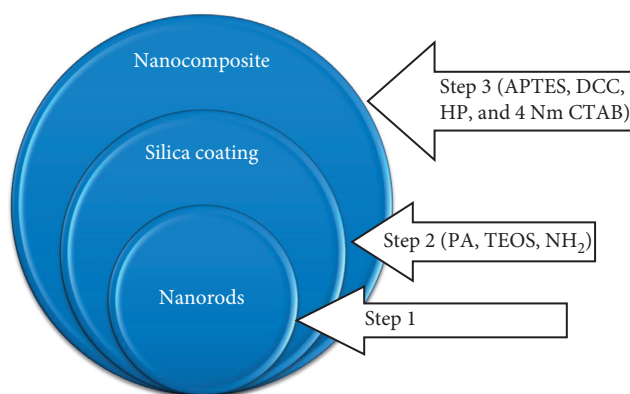


FIGURE 22: Scheme of the fabrication of AuNRs/SiO₂-HP. IPA, isopropyl alcohol; TEOS, tetraethyl orthosilicate; APTES, 3-aminopropyltriethoxysilane; HP, hematoporphyrin; DCC, dicyclo-carbodiimide; CTAB, cetyltrimethylammonium bromide [302].

3. Conclusions and Future Developments

The excellent imaging and sensing properties shown by gold nanorods qualify them as excellent for photothermal therapy. On the other hand, porphyrin and derivatives show excellent properties as photosensitizers due to the fact that they have more than one absorption band on the near-infrared region; the phenomena allow for the penetration deeper into the tissue, and hence, they have excellent use in photodynamic therapy. Photodynamic therapy shows minimal invasion to tissues while photothermal therapy is specific to tumor cells or affected area.

Photodynamic therapy limitation is still an issue up to date especially based on singlet oxygen production, selectivity to the target tissue, and the concentration so as not to damage the normal cells. Despite efforts of several researchers, an ideal photosensitizer is still not discovered, one that is safe and selective and the other that does not cause skin photosensitivity and pain. Many photosensitizers used recently have very low water solubility and tend to aggregate

under physiological conditions; moreover for clinical applications, the target tissue is still not recognised.

Incorporation of PTT and PDT as a dual technology for cancer and bacterial infections ensures selectivity and improved efficacy to the system. Previous attempts have been made using and varying different porphyrin or derivatives for photodynamic therapy or using different noble metals or carbon nanotubes for photothermal therapy or incorporating the two therapies but no ideal photosensitizer or an ideal specific drug which damages only the cancer cells has been discovered. This review is aimed at giving and overviewing of the studies made on the application of photothermal and photodynamic therapy by encapsulating gold nanorods with porphyrin for better cancer and bacterial therapy. A thorough research and further experimentation will be made on both so as to find a more specific and less invasive technology.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] P. a. Ma, H. Xiao, C. Li et al., "Inorganic nanocarriers for platinum drug delivery," *Materials Today*, vol. 18, no. 10, pp. 554–564, 2015.
- [2] F. Foroutan, N. H. de Leeuw, R. A. Martin et al., "Novel sol-gel preparation of (P₂O₅)_{0.4} - (CaO)_{0.25} - (Na₂O)_x - (TiO₂)_(0.35-x) bioresorbable glasses (X = 0.05, 0.1, and 0.15)," *Journal of Sol-Gel Science and Technology*, vol. 73, no. 2, pp. 434–442, 2015.
- [3] Y. Wang and Y. Sun, "Insulin-like growth factor receptor-1 as an anti-cancer target: blocking transformation and inducing apoptosis," *Current Cancer Drug Targets*, vol. 2, no. 3, pp. 191–207, 2002.
- [4] G. Barrera, "Oxidative stress and lipid peroxidation products in cancer progression and therapy," *ISRN Oncology*, vol. 2012, Article ID 137289, 21 pages, 2012.
- [5] J. Abtahi, F. Agholme, O. Sandberg, and P. Aspenberg, "Effect of local vs. systemic bisphosphonate delivery on

- dental implant fixation in a model of osteonecrosis of the jaw," *Journal of Dental Research*, vol. 92, no. 3, pp. 279–283, 2013.
- [6] A. Accardo, D. Tesaro, and G. Morelli, "Peptide-based targeting strategies for simultaneous imaging and therapy with nanovectors," *Polymer Journal*, vol. 45, no. 5, pp. 481–493, 2013.
- [7] I. Ramos-Álvarez, P. Moreno, S. A. Mantey et al., "Insights into bombesin receptors and ligands: highlighting recent advances," *Peptides*, vol. 72, pp. 128–144, 2015.
- [8] S. Medici, M. Peana, V. M. Nurchi, J. I. Lachowicz, G. Crisponi, and M. A. Zoroddu, "Noble metals in medicine," *Coordination Chemistry Reviews*, vol. 284, pp. 29–350, 2015.
- [9] M. J. Akhtar, M. Ahamed, H. A. Alhadlaq, S. A. Alrokayan, and S. Kumar, "Targeted anticancer therapy: overexpressed receptors and nanotechnology," *Clinica Chimica Acta*, vol. 436, pp. 78–92, 2014.
- [10] A. Accardo, L. Aloj, M. Aurilio, G. Morelli, and D. Tesaro, "Receptor binding peptides for target-selective delivery of nanoparticles encapsulate drugs," *International Journal of Nanomedicine*, vol. 9, pp. 1537–1557, 2014.
- [11] M. Singh, D. C. C. Harris-Birtill, Y. Zhou et al., "Application of gold nanorods for photothermal therapy in *ex vivo* human oesophagogastric adenocarcinoma," *Journal of Biomedical Nanotechnology*, vol. 12, no. 3, pp. 481–490, 2016.
- [12] R. Van De Meel, T. Lammers, and W. E. Hennink, "Cancer nanomaterials: oversold or unappreciated?," *Expert Opinion Drug Delivery*, vol. 14, no. 1, pp. 1–5, 2016.
- [13] D. Pissuwan, T. Niidome, and M. B. Cortie, "The forthcoming applications of gold nanoparticles in drug and gene delivery systems," *Journal of Controlled Release*, vol. 149, no. 1, pp. 65–71, 2008.
- [14] A. K. Khan, R. Rashid, G. Murtaza, and A. Zahra, "Gold nanoparticles: synthesis and application in drug delivery," *Journal of Pharmacy Research*, vol. 13, no. 7, pp. 1169–1174, 2014.
- [15] A. O. Oladipo, O. S. Oluwafemi, S. P. Songca et al., "A novel treatment for metastatic lymph nodes using lymphatic delivery and photothermal therapy," *Scientific Reports*, vol. 3, no. 7, p. 45459, 2017.
- [16] J. Ohyama, K. Hitomi, Y. Yasuhiro et al., "One-phase synthesis of small gold nanoparticles coated by a horizontal porphyrin monolayer," *Chemical Communications*, vol. 47, pp. 6300–6302, 2008.
- [17] D. Camestratic, *Synthesis of Polyheteroatomic Heterocycles; Relevance in Microwave Assisted Reactions*, Instituto Superior Técnico, Universidade Técnica de Lisboa, Lisbon, Portugal, 2014.
- [18] L. P. Cook, G. Brewer, and W. Wong-Ng, "Structural aspects of porphyrins for functional materials applications," *Crystals*, vol. 7, no. 7, pp. 1–22, 2017.
- [19] X. Huang, I. H. El-Sayed, W. Qian, and M. A. El-Sayed, "Cancer cell imaging and photothermal therapy in the near infra-red region by using gold nanorods," *Journal of the American Chemical Society*, vol. 128, no. 6, pp. 2115–2120, 2006.
- [20] A. Brioude, X. C. Jiang, and M. P. Pileni, "Structural aspects of porphyrins for functional materials applications," *Crystals*, vol. 109, no. 27, pp. 13138–13142, 2005.
- [21] X. Huang, S. Neretina, and M. A. El-Sayed, "Gold nanorods: from synthesis and properties to biological and biomedical application," *Advanced Materials*, vol. 21, no. 48, pp. 4880–4910, 2009.
- [22] H. Liang, H. Tian, M. Deng, and X. Chen, "Gold nanoparticles for cancer theranostics," *Chinese Journal of Chemistry*, vol. 33, no. 9, pp. 1001–1010, 2015.
- [23] G. Awah and Y. You, "Boron dipyrromethane (BODIPY)-based photosensitizers for photodynamic therapy," *RSC Advances*, vol. 2, pp. 11169–11183, 2012.
- [24] P. Wolf, E. Rieger, and H. Kerl, "Topical photodynamic therapy with endogenous porphyrins after applications of S-aminoleulinic acid," *Journal of the American Academy of Dermatology*, vol. 28, pp. 17–21, 1995.
- [25] Z. Song, A. O. Adeyemo, J. Baker et al., "Structure of porphyrin TPPS4 and its interaction with metal ions as elucidated by ¹H NMR and UV-visible spectra," *Georgia Journal of Science*, vol. 69, no. 2-3, pp. 89–101, 2011.
- [26] J. Winther, "Porphyrin photodynamic therapy in an experimental retinoblastoma model," *Ophthalmic Paediatric Genet*, vol. 8, no. 1, pp. 49–52, 1997.
- [27] V. Delic, J. W. D. Griffin, S. Zivkovic et al., "Individual amino acid supplementation can improve energy metabolism and decrease ROS production in neuronal cells overexpressing alpha-synuclein," *Neuromolecular Medicine*, vol. 19, no. 2-3, pp. 322–344, 2017.
- [28] R. W. Wagner, "Synthesis and excited-state photodynamics of a molecular square containing four mutually coplanar porphyrins," *Journal of Organic Chemistry*, vol. 63, no. 15, pp. 5042–5049, 1998.
- [29] D. Fenyo, B. T. Chait, T. E. Johnson, and J. S. Lindsey, "Laser desorption mass spectrometry of synthetic multiporphyrin arrays," *Journal of Porphyrins and Phthalocyanines*, vol. 1, no. 1, pp. 93–99, 1997.
- [30] E. C. A. Ojadi, H. Linschitz, M. Gouterman et al., "Sequential protonation of meso-(dimethylamino)phenyl porphyrins: charge-transfer excited states producing hyperporphyrins," *Journal of Physical Chemistry*, vol. 97, no. 50, pp. 13192–13197, 1993.
- [31] J. Li, A. Ambrose, S. I. Yang et al., "Template-directed synthesis, excited-state photodynamics, and electronic communication in a hexameric wheel of porphyrins," *Journal of the American Chemical Society*, vol. 121, no. 38, pp. 8927–8940, 1999.
- [32] S. Jain, D. G. Hirst, and J. M. O'Sullivan, "Gold nanoparticles as novel agents for cancer therapy," *British Journal of Radiology*, vol. 85, no. 1010, pp. 101–113, 2012.
- [33] W. Cai, T. Gao, H. Hong et al., "Applications of gold nanoparticles in cancer nanotechnology," *Nanotechnology*, vol. 1, pp. 17–32, 2008.
- [34] M. Marsh, E. Schelew, S. Wolf et al., "Gold nanoparticles for cancer treatment," *Physics*, pp. 1–9, 2009.
- [35] P. K. Jain, I. H. EL-Sayed, and L. A. EL-Sayed, "Au nanoparticles target cancer," *Nano Today*, vol. 2, no. 1, pp. 18–29, 2007.
- [36] S. Link and M. A. El-Sayed, "Shape and size dependence of radiative, non-radiative and photothermal properties of gold nanocrystals," *International Reviews in Physical Chemistry*, vol. 19, no. 3, pp. 409–453, 2000.
- [37] M. K. Yu, J. Park, and S. Jon, "Targeting strategies for multifunctional nanoparticles in cancer imaging and therapy," *Theranostics*, vol. 2, no. 1, pp. 1–42, 2012.
- [38] O. V. Salata, "Applications of nanoparticles in biology and medicine," *Journal of Nanobiotechnology*, vol. 2, p. 3, 2004.
- [39] W. H. De Jong and P. J. A. Borm, "Drug delivery and nanoparticles: applications and hazards," *International Journal of Nanoscience*, vol. 3, no. 2, pp. 133–149, 2008.

- [40] J. Conde, G. Doria, and P. Baptista, "Nobel metal nanoparticles applications in cancer," *Journal of Drug Delivery*, vol. 2012, Article ID 751075, 12 pages, 2012.
- [41] H. C. Huang, S. Barua, G. Sharma, S. K. Dey, and K. Rege, "Inorganic particles for cancer imaging and therapy," *Journal of Controlled Release*, vol. 155, no. 3, pp. 344–357, 2011.
- [42] M. P. Melancon, M. Zhou, and C. Li, "Cancer theranostics with near-infrared light-activatable multinodal nanoparticles," *Accounts of Chemical Research*, vol. 44, no. 10, pp. 947–956, 2011.
- [43] C. D. Chen, S. F. Cheng, L. K. Chau, and C. R. Chris Wang, "Sensing capability of localized surface plasmon resonance of gold nanorods," *Biosensors & Bioelectronics*, vol. 22, no. 6, pp. 926–932, 2007.
- [44] W. Lu, H. Wang, J. Zhang et al., "Gold nanorods: synthesis growth mechanism and purification," *Process Chemistry*, vol. 27, pp. 785–793, 2015.
- [45] H. Takahashi, T. Niidome, A. Nariai, Y. Niidome, and S. Yamada, "Gold nanorod: sensitized cell-death microscopic observation of single living cells irradiated by pulsed near-infrared laser light in the presence of gold nanorods," *Chemistry Letters*, vol. 35, no. 5, pp. 500–502, 2006.
- [46] J. W. Hotchkiss, A. B. Lowe, and S. G. Boyes, "Surface modification of gold nanorods with polymers synthesized by reversible addition-fragmentation chain transfer polymerization," *Chemistry of Materials*, vol. 19, no. 1, pp. 6–13, 2007.
- [47] R. Narayanan, "Nanoparticles of different shapes for biosensor applications. Functional nanoparticles for bioanalysis," *Nanomedicine, and Bioelectronic Devices*, vol. 1, pp. 281–292, 2012.
- [48] G. Doria, J. Conde, B. Veigas et al., "Noble metal nanoparticles for biosensing applications," *Sensors*, vol. 12, no. 2, pp. 1657–1687, 2012.
- [49] M. Holzinger, A. Goff, and S. Cosnier, "Nanomaterials for biosensing applications: a review," *Frontiers in Chemistry*, vol. 2, pp. 63–66, 2014.
- [50] A. Ahmed, M. Ahamad, L. B. Swami, and S. Ikram, "A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise," *Journal of Advanced Research*, vol. 7, no. 1, pp. 17–28, 2016.
- [51] W. Chen, S. Zhang, Y. Yu, H. Zhang, and Q. He, "Structural-engineering rationales of gold nanoparticles for cancer theranostic," *Advanced Materials*, vol. 28, no. 39, pp. 8567–8585, 2016.
- [52] Z. Zhang, J. Wang, and C. Chen, "Gold nanorods based platforms for light-mediated theranostic," *Theranostics*, vol. 3, no. 3, pp. 223–239, 2012.
- [53] D. Sharma, S. Kanchi, and K. Bisetty, "Biogenic synthesis of nanoparticles," *Arabian Journal of Chemistry*, vol. 11, pp. 1–25, 2015.
- [54] Q. Zhang, N. Iwakuma, P. Sharma et al., "Gold nanoparticles as a contrast agent for in-vivo tumor imaging and photoacoustic tomography," *Nanotechnology*, vol. 20, no. 39, pp. 1–9, 2009.
- [55] M. Sguizzato, E. Esposito, M. Drechsler et al., "Nanotechnology containing solid lipid nanoparticles as a tool for anticancer drug delivery: preliminary studies," *Journal of Chemistry*, vol. 2017, Article ID 3206298, 6 pages, 2017.
- [56] M. I. Tomey, J. Narula, and J. C. Kovacic, "Advances in the understanding of plaque composition and treatment options," *Journal of the American College of Cardiology*, vol. 63, no. 16, pp. 1604–1606, 2014.
- [57] S. W. Prescott and P. Mulvaney, "Gold nanorod extinction spectra," *Journal of Applied Physics*, vol. 99, no. 103, pp. 1–8, 2008.
- [58] E. K. Payne, K. L. Shuford, S. Park, G. C. Schatz, and C. A. Mirkin, "Multipole plasmon resonances in gold nanorods," *Journal of Physical Chemistry*, vol. 110, no. 5, pp. 2150–2154, 2006.
- [59] M. Liu, P. H. Yang, and J. Y. Cai, "Optical properties and biomedical application of gold nanorods," *Microbiology and Immunology*, vol. 36, no. 11, pp. 1402–1407, 2009.
- [60] K. Park, S. Biswas, S. Kanel, D. Nepal, and R. A. Vaia, "Engineering the optical properties of gold nanorods: independent tuning of surface plasmon energy, extinction coefficient, and scattering cross section," *Journal of Physical Chemistry*, vol. 118, no. 11, pp. 5918–5926, 2014.
- [61] A. M. Funston, C. Novo, T. J. Davis, and P. Mulvaney, "Plasmon coupling of gold nanorods at short distances in different geometries," *Nano Letters*, vol. 9, no. 4, pp. 1651–1658, 2009.
- [62] T. Ye, Z. Dai, F. Mei et al., "Synthesis and optical properties of gold nanorods with controllable morphology," *Journal of Physics: Condensed Matter*, vol. 28, no. 43, article 434002, 2016.
- [63] Y. Liu, E. N. Mills, and R. J. Composto, "Tuning optical properties of gold nanorods in polymer films through thermal reshaping," *Journal of Materials Chemistry*, vol. 19, no. 18, pp. 2704–2709, 2009.
- [64] Y. R. Toh, P. Yu, X. Wen, J. Tang, and T.-s. Hsieh, "Induced pH-dependent shift by local surface plasmon resonance in functionalized gold nanorods," *Nanoscale Research Letters*, vol. 8, no. 1, p. 103, 2013.
- [65] C. A. Peng and S. Pachpinde, "Longitudinal plasmonic detection of glucose using gold nanorods," *Nanomaterials and Nanotechnology*, vol. 4, no. 9, pp. 1–5, 2013.
- [66] B. Sadeghi, *Synthesis and Application of Nanorods: Nanotechnology and Nanomaterials*, Chapter 6, IntechOpen, London, UK, 2012.
- [67] P. Wang, M. Liu, G. Gao et al., "From gold nanorods to nanodumbbells: a different way to tailor surface plasmon resonances by a chemical route," *Journal of Materials Chemistry*, vol. 22, pp. 24006–24011, 2012.
- [68] S. K. Sharma, P. Mroz, T. Dai, Y.-Y. Huang, T. G. S. Denis, and M. R. Hamblin, "Photodynamic therapy for cancer and for infections: what is the difference?," *Israel Journal of Chemistry*, vol. 52, no. 8-9, pp. 691–705, 2012.
- [69] L. Feng, Z. Xuana, J. Mac et al., "Preparation of gold nanorods with different aspect ratio and the optical response to solution refractive index," *Journal of Experimental Nanoscience*, vol. 10, no. 4, pp. 258–26, 2015.
- [70] E. T. Castellana, R. C. Gamez, M. E. Gómez, and D. H. Russell, "Longitudinal surface plasmon resonance based gold nanorod biosensors for mass spectrometry," *Langmuir*, vol. 26, no. 8, pp. 6066–6070, 2010.
- [71] T. A. El-Brollosy, T. Abdallah, M. B. Mohamed et al., "Shape and size dependence of the surface plasmon resonance of gold nanoparticles studied by photoacoustic technique," *European Physical Journal Special Topics*, vol. 153, no. 1, pp. 361–364, 2008.
- [72] A. Sambou, P. D. Tall, K. Talla et al., "Control of the surface plasmon resonance of two configurations of nanoparticles: simple gold nanorod and gold/silica core/shell," *Nanoscience and Nanotechnology Research*, vol. 4, no. 1, pp. 1–6, 2017.

- [73] N. Thioune, N. Lidgi-Guigui, M. Cottat et al., "Study of gold nanorods-protein interaction by localized surface plasmon resonance spectroscopy," *Gold Bulletin*, vol. 46, no. 4, pp. 275–281, 2013.
- [74] J. Cao, T. Sun, and K. T. V. Grattan, "Gold nanorod-based localized surface plasmon resonance biosensors: a review," *Sensors and Actuators B: Chemical*, vol. 195, pp. 332–351, 2014.
- [75] S. Jayabal, A. Pandikumar, H. N. Lim, R. Ramaraj, T. Sun, and N. M. Huang, "A gold nanorod-based localized surface plasmon resonance platform for the detection of environmentally toxic metal ions," *Analyst*, vol. 140, no. 8, pp. 2540–2555, 2015.
- [76] J. Tao, L. Yong-Hua, Z. Rong-Sheng et al., "Effect of aspect ratio distribution on localized surface plasmon resonance extinction spectrum of gold nanorods," *Chinese Physics Letters*, vol. 25, no. 12, pp. 1–4, 2008.
- [77] V. S. Marangoni, J. Cancino-Bernardi, and V. Zucolotto, "Synthesis, physico-chemical properties and biomedical applications of gold nanorod—a review," *Journal of Biomedical Nanotechnology*, vol. 12, no. 6, pp. 1136–1158, 2016.
- [78] B. M. Geilich, A. L. van de Ven, G. L. Singleton, L. J. Sepúlveda, S. Sridhar, and T. J. Webster, "Silver nanoparticle-embedded polymersome nanocarriers for the treatment of antibiotic-resistant infections," *Nanoscale*, vol. 7, no. 8, pp. 3511–3519, 2015.
- [79] H. Takahashi, Y. Niidome, T. Niidome, Y. Niidome, and S. Yamada, "Photothermal reshaping of gold nanorods prevents further cell death," *Langmuir*, vol. 17, no. 17, pp. 4431–4435, 2006.
- [80] L. Vigdeman, B. P. Khanal, and E. R. Zubarev, "Functional gold nanorods: synthesis, self-assembly, and sensing applications," *Advanced Materials*, vol. 24, no. 36, p. 5016, 2012.
- [81] Y. Wang, K. C. L. Black, H. Luehmann et al., "Comparison study of gold nano hexapods, nanorods and nanocages for photothermal cancer treatment," *ACS Nano*, vol. 7, no. 3, pp. 2068–2077, 2013.
- [82] J. Li, B. Zhu, Z. Zhu et al., "Simple and rapid functionalization of gold nanorods with oligonucleotides using an mPEG-SH/tween 20-assisted approach," *Langmuir*, vol. 31, no. 28, pp. 7869–7876, 2015.
- [83] R. M. Pallares, X. Su, S. H. Lim et al., "Fine-tuning gold nanorod dimensions and plasmonic properties using the Hofmeister effects," *Journal of Materials Chemistry C*, vol. 00, no. 1, pp. 1–3, 2015.
- [84] K. Liu, Y. Zheng, X. Lu et al., "Biocompatible gold nanorods: one-step surface functionalization, highly colloidal stability, and low cytotoxicity," *Langmuir*, vol. 31, no. 17, pp. 4973–4980, 2015.
- [85] R. G. Rayavarapu, W. Petersen, L. Hartsuiker et al., "In vitro studies of polymer-coated gold nanorods," *Nanotechnology*, vol. 21, no. 14, article 145101, 2015.
- [86] Z.-C. Xu, C.-S. Shen, C.-W. Xiao et al., "Wet chemical synthesis of gold nanoparticles using silver seeds: a shape control from nanorods to hollow spherical nanoparticles," *Nanotechnology*, vol. 18, no. 11, pp. 115608–115613, 2007.
- [87] B. D. Busbee, S. O. Obare, and C. J. Murphy, "An improved synthesis of high-aspect-ratio gold nanorods," *Advanced Materials*, vol. 15, no. 5, pp. 414–416, 2003.
- [88] D. P. Sprünken, H. Omi, K. Furukawa et al., "Influence of the local environment on determining aspect distribution of gold nanorods," *Journal of Physical Chemistry*, vol. 111, no. 39, pp. 14299–14306, 2007.
- [89] L. Scarabelli, A. Sánchez-Iglesias, J. Pérez-Juste et al., "A 'Tips and Tricks' practical guide to the synthesis of gold nanorods," *Journal of Physical Chemistry Letters*, vol. 6, no. 21, pp. 4270–4279, 2015.
- [90] P. Wattanakull, MC Killingsworth, and D. Pissuwan, "Biological responses of T cells encapsulated with poly-electrolyte-coated gold nanorods and their cellular activities in a co-culture system," *Applied Nanoscience*, vol. 7, no. 8, pp. 667–679, 2017.
- [91] H. Takahashi, T. Niidome, A. Nariai, Y. Niidome, and S. Yamada, "Photothermal reshaping of gold nanorods prevents further cell death," *Nanotechnology*, vol. 17, no. 17, pp. 4431–4435, 2006.
- [92] C. Kinnear, D. Burnand, M. J. Clift, A. F. M. Kilbinger, B. Rothen-Rutishauser, and A. Petri-Fink, "Polyvinyl alcohol as a biocompatible alternative for the passivation of gold nanorods," *Journal of the American Chemical Society*, vol. 53, no. 46, pp. 12613–12617, 2014.
- [93] J. Stone, S. Jackson, and D. Wright, "Biological applications of gold nanorods," *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, vol. 3, no. 1, pp. 100–110, 2011.
- [94] T. A. Larson, B. James, A. Jesse, and K. Sokolov, "Hybrid plasmonic magnetic nanoparticles as molecular specific agent for MRI/optical imaging and photothermal therapy of cancer cells," *Nanotechnology*, vol. 18, no. 32, pp. 1–9, 2007.
- [95] A. Gole and C. J. Murphy, "Seed-mediated synthesis of gold nanorods: role of the size and nature of the seed," *Chemistry of Materials*, vol. 16, no. 19, pp. 3633–3640, 2004.
- [96] O. S. Oluwafemi, "Synthesis of gelatin stabilized high aspect ratio gold nanorods with enhanced biological stability as effective photothermal agents for cancer therapy," *Trends in Green Chemistry*, vol. 3, no. 3, p. 1, 2017.
- [97] H. Ke, J. Wang, Z. Dai et al., "Gold-nanoshelled microcapsules: theranostic agent for ultrasound contrast imaging and photothermal therapy," *Angewandte Chemie International Edition*, vol. 50, no. 13, pp. 3017–3021, 2011.
- [98] J. Becker, I. Zins, A. Jakab, Y. Khalavka, O. Schubert, and C. Sönnichsen, "Plasmonic focusing reduces ensemble linewidth of silver-coated gold nanorods," *Nano Letters*, vol. 8, no. 6, pp. 1719–1723, 2008.
- [99] H. S. Ali, S. Ahmed, and G. A. Khan, "Gold nanoparticles in cancer therapy and diagnosis," *Pharmaceutical Research*, vol. 28, no. 2, pp. 237–259, 2011.
- [100] A. Fernandez-Fernandez, R. Manchanda, and A. J. McGoron, "Theranostic applications of nanomaterials in cancer: drug delivery, image-guided therapy and multifunctional platforms," *Applied Biochemistry and Biotechnology*, vol. 165, no. 7–8, pp. 1628–1651, 2011.
- [101] F. Geng, K. Song, and J. Z. Xing, "Thio-glucose bound gold nanoparticles enhance radio-cytotoxic targeting of ovarian cancer," *Nanotechnology*, vol. 22, no. 28, article 285101, 2011.
- [102] T. Wolfe, D. Chatterjee, J. Lee et al., "Targeted gold nanoparticles enhance sensitization of prostate tumors to megavoltage radiation therapy in vivo," *Nanotechnology*, vol. 11, no. 5, pp. 1277–1283, 2015.
- [103] W. Xu, T. Luo, P. Li et al., "RGD-conjugated gold nanorods induce radiosensitization in melanoma cancer cells by downregulating $\alpha_v\beta_3$ expression," *International Journal of Nanomedicine*, vol. 7, pp. 915–924, 2012.
- [104] D. Kumar, I. Mutreja, K. Chitcholtan, and P. Sykes, "Cytotoxicity and cellular uptake of different sized gold nanoparticles in ovarian cancer cells," *Nanotechnology*, vol. 28, no. 47, article 475101, 2017.

- [105] T. B. Huff, L. Tong, Y. Zhao, M. N. Hansen, J.-X. Cheng, and A. Wei, "Hyperthermia effect of gold nanorods on tumor cells," *Nanomedicine*, vol. 2, no. 1, pp. 125–132, 2007.
- [106] N. Verellen, D. Denkova, B. D. Clercq et al., "Two-photon luminescence of gold nanorods mediated by higher order plasmon modes," *ACS Photonics*, vol. 2, no. 3, pp. 410–416, 2015.
- [107] T. Wang, D. Halaney, D. Ho et al., "Two-photon luminescence properties of gold nanorods," *Biomed Optical Express*, vol. 4, no. 4, pp. 584–595, 2013.
- [108] L. C. Kennedy, L. R. Bickford, N. A. Lewinski et al., "A new era for cancer treatment: gold nanoparticle-mediated thermal therapies," *Small*, vol. 7, no. 2, pp. 169–183, 2011.
- [109] R. Arvizo, R. Bhattacharya, and P. Mukherjee, "Gold nanoparticles: opportunities and challenges in nanomedicine," *Expert Opinion on Drug Delivery*, vol. 7, no. 6, pp. 753–763, 2010.
- [110] L. F. Hakim, C. L. Vaughn, H. J. Dunsheath et al., "Synthesis of oxidation resistance metal nanoparticles via atomic layer deposition," *Nanotechnology*, vol. 18, no. 34, pp. 1–7, 2007.
- [111] J. G. Thakare, C. Pande, R. S. Mulik, and M. M. Mahapatra, "Mechanical property evaluation of carbon nanotubes reinforced plasma sprayed YSZ-alumina composite coating," *Ceramics International*, vol. 44, no. 6, pp. 1–10, 2018.
- [112] S.-S. Chang, C.-W. Shih, C.-D. Chen et al., "The shape transition of gold nanorods," *Langmuir*, vol. 15, no. 3, pp. 701–709, 1999.
- [113] N. R. Jana, L. Gearheart, and C. J. Murphy, "Wet chemical synthesis of high aspect ratio cylindrical gold nanorods," *Journal of Physical Chemistry*, vol. 105, no. 19, pp. 4065–4067, 2001.
- [114] B. Nikoobakht and M. A. El-Sayed, "Preparation and growth mechanism of gold nanorods (NRs) using seed-mediated growth method," *Chemistry of Materials*, vol. 15, no. 10, pp. 1957–1962, 2003.
- [115] D. H. Kim, A. Wei, and Y. Y. Won, "Preparation of superstable gold nanorods via encapsulation into block copolymer micelles," *ACS Applied Materials & Interfaces*, vol. 4, no. 4, pp. 1872–1877, 2012.
- [116] X. Ye, L. Jin, H. Caglayan et al., "Improved size tunable synthesis of monodisperse gold nanorods through the use of aromatic additives," *ACS Nano*, vol. 6, no. 3, pp. 2804–2817, 2012.
- [117] J. S. Lee and J. Feijen, "Polymersomes for drug delivery: design, formation and characterization," *Journal of Controlled Release*, vol. 161, no. 2, pp. 473–483, 2012.
- [118] K. K. Upadhyay, A. K. Mishra, K. Chuttani et al., "The in vivo behavior and antitumor activity of doxorubicin-loaded poly (γ -benzyl L-glutamate)-block-hyaluronan polymersomes in Ehrlich ascites tumor-bearing BalB/c mice," *Nanomedicine: Nanotechnology, Biology and Medicine*, vol. 8, no. 1, pp. 71–80, 2012.
- [119] A. Gharatape, S. Davaran, R. Salehi, and H. Hamishehkar, "Engineered gold nanoparticles for photothermal cancer therapy and bacteria killing," *RSC Advances*, vol. 6, no. 112, pp. 111482–111516, 2016.
- [120] D. Jaque, L. Martínez Maestro, B. del Rosal et al., "Nanoparticles for photothermal therapies," *Nanoscale*, vol. 6, no. 16, pp. 9494–9530, 2014.
- [121] Z. Bao, X. Liu, Y. Liu, H. Liu, and K. Zhao, "Near-infrared light-responsive inorganic nanomaterials for photothermal therapy," *Asian Journal of Pharmaceutical Sciences*, vol. 11, no. 3, pp. 349–364, 2016.
- [122] Y. Niidome, K. Honda, K. Higashimoto et al., "Surface modification of gold nanorods with synthetic cationic lipids," *Chemical Communications*, vol. 36, pp. 3777–3779, 2007.
- [123] A. Pan, H. Zhang, Y. Li et al., "Disulfide-crosslinked nanomicelles confer cancer-specific drug delivery and improve efficacy of paclitaxel in bladder cancer," *Nanotechnology*, vol. 27, no. 42, article 425103, 2016.
- [124] T. Ci, L. Chen, L. Yu, and J. Ding, "Tumor regression achieved by encapsulating a moderately soluble drug into a polymeric thermogel," *Scientific Reports*, vol. 1, no. 4, p. 5473, 2014.
- [125] B. K. Lee, Y. H. Yun, and K. Park, "Smart nanoparticles for drug delivery: boundaries and opportunities," *Chemical Engineering Science*, vol. 125, pp. 158–164, 2015.
- [126] F. Gomez-Rivera, "Functionalization of gold nanorod-based immunosensors for label-free bio-detection with high sensitivity and specificity," *Society for Advancement of Hispanics/Chicanos and Native Americans in Science*, vol. 28, no. 1, 2014.
- [127] P. Bengtsson and T. Johannesson, "Characterization of microstructural defects in plasma-sprayed thermal barrier coatings," *Journal of Thermal Spray Technology*, vol. 4, no. 3, pp. 245–251, 1995.
- [128] S. Ohnishi, A. Nakasuga, and K. Nakagawa, "Synthesis of copper particles covered with cobalt-catalyzed carbon nanofibers and their application to air-curable conductive paste," *Japanese Journal of Applied Physics*, vol. 56, no. 7S2, pp. 1–7, 2017.
- [129] K. J. Park, J. M. Doub, T. Gougousi, and G. N. Parsons, "Microcontact patterning of ruthenium gate electrodes by selective area atomic layer deposition," *Applied Physics Letters*, vol. 86, no. 5, article 051903, 2005.
- [130] M. Leskela and M. Ritala, "Atomic layer deposition chemistry: recent developments and future challenges," *Angewandte Chemie International Edition*, vol. 42, no. 45, pp. 5548–5554, 2003.
- [131] H. Moshe and Y. Mastai, "Atomic layer deposition on self-assembled monolayers," in *Materials Science—Advanced Topics*, Springer, Berlin, Germany, 2009.
- [132] L. Niinistö, M. Nieminen, J. Päiväsaari, J. Niinistö, M. Putkonen, and M. Nieminen, "Advanced electronic and optoelectronic materials by Atomic Layer Deposition: an overview with special emphasis on recent progress in processing of high-k dielectrics and other oxide materials," *Physica Status Solidi*, vol. 201, no. 77, pp. 1375–1376, 2004.
- [133] C. J. Murphy, L. B. Thompson, A. M. Alkilany et al., "The many faces of gold nanorods," *Journal of Physical Chemistry Letters*, vol. 1, no. 19, pp. 2867–2875, 2010.
- [134] J. Schmidt, A. Merkle, R. Brendel et al., "Surface passivation of high-efficiency silicon solar cells by atomic-layer-deposited Al_2O_3 ," *Progress in Photovoltaics: Research and Applications*, vol. 16, no. 6, pp. 461–466, 2008.
- [135] R. M. Crooks, M. Zhao, L. Sun, V. Chechik, and L. K. Yeung, "Dendrimer-encapsulated metal nanoparticle synthesis, characterization and application to catalysis," *Accounts of Chemical Research*, vol. 34, no. 3, pp. 1–10, 2011.
- [136] Y. Niu and R. M. Crooks, "Dendrimer-encapsulated metal nanoparticles and their applications to catalysis," *Metallocentric Catalysis*, vol. 6, no. 8–10, pp. 1049–1059, 2003.
- [137] C. J. Murphy, L. B. Thompson, A. M. Alkilany et al., "The many faces of gold nanorods," *Journal of Physical Chemistry Letters*, vol. 1, no. 19, pp. 2867–2875, 2010.

- [138] S. Boulos, M. B. Prigozhin, Y. Liu et al., "The gold nanorod-biology interface: from proteins to cells to tissue," *Current Physical Chemistry*, vol. 3, no. 2, pp. 128–135, 2013.
- [139] W. Ahmed, C. Glass, E. S. Kooij, and J. M. van Ruitenbeek, "Tuning the oriented deposition of gold nanorods on patterned substrates nanotechnology," *Nanotechnology*, vol. 25, no. 3, article 035301, 2013.
- [140] M. Zhao and R. M. Crooks, "Intradendrimer exchange of metal nanoparticles," *Chemistry of Materials*, vol. 11, no. 11, pp. 3379–3385, 1999.
- [141] P. A. Oroskar, C. J. J. Jameson, and S. Murad, "Rotational behaviour of PEGylated gold nanorods in a lipid bilayer system," *Molecular Physics*, vol. 115, no. 9–12, pp. 1122–1143, 2016.
- [142] A. Wijaya and K. Humad-Shifferli, "Ligand customization and DNA functionalization of gold nanorods via round-trip phase transfer exchange," *Langmuir*, vol. 24, no. 18, pp. 9966–9969, 2008.
- [143] A. B. Golovin and O. D. Lavrentovich, "Electrically reconfigurable optical metamaterial based on colloidal dispersion of metal nano-rods in dielectric fluid," *Applied Physics Letters*, vol. 95, no. 25, article 254104, 2009.
- [144] A. D. Adler, F. R. Longo, F. Kampas, and J. Kim, "Preparation of metalloporphyrins," *Journal of Inorganic and Nuclear Chemistry*, vol. 32, no. 7, pp. 2443–2445, 1970.
- [145] L.-I. Chen, L. Jiang, Y.-I. Wang, J. Qian, and S. He, "Multilayered polyelectrolyte-coated gold nanorods as multifunctional optical contrast agents for cancer cell imaging," *Journal of Zhejiang University Science B*, vol. 11, no. 6, pp. 417–422, 2006.
- [146] D. Pan, M. Pramanik, A. Senpan, S. A. Wickline, L. V. Wang, and G. M. Lanza, "A facile synthesis of novel self-assembled gold nanorods designed for near-infrared imaging," *Journal of Nanoscience and Nanotechnology*, vol. 10, no. 12, pp. 8118–8123, 2010.
- [147] M. Fang, C.-W. Peng, D.-W. Pang, and Y. Li, "Quantum dots for cancer research: current status, remaining issues and future perspectives," *Cancer Biology & Medicine*, vol. 9, no. 3, pp. 151–163, 2013.
- [148] E. Locatelli, I. Monaco, and M. C. Franchini, "Surface modifications of gold nanorods for applications in nanomedicine," *RSC Advances*, vol. 5, no. 28, pp. 21681–21699, 2015.
- [149] C. Xue, Y. Xu, Y. Pang et al., "Organo-soluble porphyrin mixed monolayer protected gold nanorods with intercalated fullerenes," *Langmuir*, vol. 28, no. 14, pp. 5956–5963, 2012.
- [150] A. Gole and C. J. Murphy, "Polyelectrolyte-coated gold nanorods: synthesis, characterization and immobilization," *Chemistry of Materials*, vol. 17, no. 6, pp. 1325–1330, 2005.
- [151] L. M. Moreira, F. V. dos Santos, J. P. Lyon, M. Maftoum-Costa, C. Pacheco-Soares, and N. S. da Silva, "Photodynamic therapy: porphyrins and phthalocyanines as photosensitizers," *Journal of Chemistry*, vol. 61, no. 10, pp. 741–754, 2008.
- [152] J. Moan, "Porphyrin photosensitization and phototherapy," *Photochemistry and Photobiology*, vol. 43, no. 6, pp. 681–690, 1986.
- [153] R. Karami, A. Mohsenifar, N. S. M. Mesbah et al., "A novel nanobiosensor for the detection of paraoxon using chitosan-embedded organophosphorous hydrolase immobilized on Au nanoparticles," *Preparative Biochemistry and Biotechnology*, vol. 46, no. 6, pp. 559–566, 2015.
- [154] C. Thomas, "Exploring new routes towards photoactive assemblies," *Proquest*, vol. 115, pp. 1–229, 2016.
- [155] S. Kimura, E. Bill, E. Bothe et al., "Phenylthiyl radical complexes of gallium(III), iron(III), and cobalt(III) and comparison with their phenoxy analogues," *Journal of the American Chemical Society*, vol. 123, no. 25, pp. 6025–6039, 2001.
- [156] P. Bhyrappa, M. Sankar, and B. Varghese, "Mixed substituted porphyrins: structural and electrochemical redox properties," *Inorganic Chemistry*, vol. 45, no. 10, pp. 4136–4149, 2006.
- [157] G. T. Babcock, M. K. El-Deeb, P. O. Sandusky, M. M. Whittaker, and J. W. Whittaker, "Electron paramagnetic resonance and electron nuclear double resonance spectroscopies of the radical site in 3121 galactose oxidase and of thioether-substituted phenol model compounds," *Journal of the American Chemical Society*, vol. 114, no. 10, pp. 3121–3134, 1992.
- [158] D. Burdinski, E. Bothe, and K. Wieghardt, "Synthesis and characterization of tris(bipyridyl)ruthenium(II)-modified mono-, di-, and trinuclear manganese complexes as electron-transfer models for photosystem II," *Inorganic Chemistry*, vol. 39, no. 1, pp. 105–116, 2000.
- [159] P. Rothemund, "A new porphyrin synthesis. The synthesis of porphyrin," *American Chemical Society*, vol. 58, pp. 624–625, 1936.
- [160] P. Rothemund and A. R. Menott, "Porphyrin studies. IV. The synthesis of @,y,b-tetraphenylporphine," *Journal of the American Chemical Society*, vol. 63, no. 1, pp. 267–270, 1941.
- [161] P. Rothemund, "Porphyrin studies. III. The structure of the porphine² ring system," *Journal of the American Chemical Society*, vol. 61, no. 10, pp. 2912–2915, 1939.
- [162] N. I. Shvetsov-Shilovskii, R. G. Bobkova, N. P. Ignatova et al., "Compounds of two-coordinated phosphorus," *Journal of the American Chemical Society*, vol. 46, no. 6, pp. 514–529, 1977.
- [163] P. Angrish, *Syntheses of meso-Substituted Porphodimethenes and Porphyrins with Exocyclic Ring Systems*, University of Florida, Tampa, FL, USA, 2003.
- [164] A. D. Adler, F. R. Longo, and W. Shergalis, "Mechanistic investigations of porphyrin syntheses. I. preliminary studies on ms-tetraphenylporphine," *Journal of the American Chemical Society*, vol. 86, no. 15, pp. 3145–3314, 1964.
- [165] A. D. Adler, F. R. Longo, J. D. Finarelli, J. Goldmacher, J. Assour, and L. Korsakoff, "A simplified synthesis for meso-tetraphenylporphine," *Journal of Organic Chemistry*, vol. 32, no. 2, p. 476, 1967.
- [166] R. G. Littler, "Notes on the synthesis of meso-substituted porphyrins from pyrrol carbinols and the mechanisms of the rothemund reaction," *Journal of Heterocyclic Chemistry*, vol. 18, no. 4, p. 833, 1981.
- [167] S. Hiroto, Y. Miyake, and H. Shinokubo, "Synthesis and functionalization of porphyrins through organometallic methodologies," *Chemical Reviews*, vol. 117, no. 4, pp. 2910–3043, 2017.
- [168] J. S. Lindsey, I. C. Schreiman, H. C. Hsu, P. C. Kearney, and A. M. Marguerettaz, "Stepwise syntheses of unsymmetrical tetra-arylporphyrin," *Organic Chemistry*, vol. 52, no. 5, p. 827, 1987.
- [169] H. G. Graham, M. van der Auweraer, J.-C. Chambron et al., "Intramolecular energy transfer in bis-porphyrins containing diimine chelates of variable geometry as spacers," *Chemistry: A European Journal*, vol. 5, no. 7, pp. 2189–2100, 1999.
- [170] H. Du, R.-C. A. Fuh, J. Li, L. Andrew Corkan, and J. S. Lindsey, "PhotochemCAD: a computer aided design and

- research tool in photochemistry,” *Photochemistry and Photobiology*, vol. 68, no. 2, pp. 141–142, 1998.
- [171] M. Dommaschk, C. Näther, and R. Herges, “Synthesis of functionalized perfluorinated porphyrins for improved spin switching,” *Journal of Organic Chemistry*, vol. 80, no. 17, pp. 8496–8500, 2015.
- [172] G. R. Geier III and J. S. Lindsey, “Investigation of porphyrin-forming reactions. Part 2. Examination of the reaction course in two-step, one-flask syntheses of meso-substituted porphyrins,” *Journal of the Chemical Society*, vol. 2, no. 5, pp. 687–700, 2001.
- [173] G. R. Geier III, B. J. Littler, and J. S. Lindsey, “Investigation of porphyrin-forming reactions. Part 3. The origin of scrambling in dipyrromethane aldehyde condensations yielding trans-A2B2-tetraarylporphyrins,” *Journal of the Chemical Society*, pp. 701–711, 2001.
- [174] G. R. Geier III, B. J. Littler, and J. S. Lindsey, “Investigation of porphyrin-forming reactions. Part 4.1 Examination of the reaction course in syntheses of porphyrins via dipyrromethanecarbinols,” *Royal Society of Chemistry*, pp. 712–718, 2001.
- [175] G. R. Geier III and J. S. Lindsey, “Effects of diverse acid catalysts on the reaction course in the two-step one-flask synthesis of meso-tetraphenylporphyrin,” *Royal Society of Chemistry*, vol. 6, no. 3, pp. 159–185, 2002.
- [176] G. Urata, “The chemistry of porphyrins and their precursors on the heme biosynthetic chain,” *Nihon Rinsho*, vol. 53, no. 6, pp. 1319–1328, 1995.
- [177] S. K. Pushpan, S. Venkatraman, V. G. Anand et al., “Porphyrin in photodynamic therapy—a search for ideal photosensitizers,” *Current Medicinal Chemistry-Anti-Cancer Agents*, vol. 2, no. 2, pp. 187–207, 2002.
- [178] Y. Wu, D. Xing, and W. R. Chen, “Single cell FRET imaging determination pathway of tumor cell apoptosis induced by photofin-PDT,” *Cell Cycle*, vol. 5, no. 7, pp. 729–734, 2006.
- [179] K. Moghissi, K. Dixon, M. Stringer, and J. A. C. Thorpe, “Photofin PDT for early stage oesophageal cancer: long term results in 40 patients and literature review,” *Thorax*, vol. 6, no. 3–4, pp. 159–166, 2009.
- [180] P. J. Muller and B. C. Wilson, “Photodynamic therapy of brain tumors—a work in progress,” *Lasers in Surgery and Medicine*, vol. 38, no. 5, pp. 384–389, 2006.
- [181] B.-H. Li, “Singlet oxygen quantum yields of porphyrin based photosensitizers for photodynamic therapy,” *Journal of Innovative Optical Health Sciences*, vol. 1, no. 1, pp. 141–149, 2008.
- [182] K. Stefflova, H. Li, J. Chen, and G. Zheng, “Peptide-based phamacodutation of a cancer-targeted optical imaging and photodynamic therapy agent,” *Bioconjugate Chemistry*, vol. 18, no. 2, pp. 379–388, 2007.
- [183] Y.-n. Wen, W.-s. Song, L.-m. An, Y.-q. Liu, Y.-h. Wang, and Y.-q. Yang, “Activation of porphyrin photosensitizers by semiconductor quantum dots via two-photon excitation,” *Applied Physics Letters*, vol. 95, article 143702, 2009.
- [184] T. Aida and S. Inoue, “Activation of carbon dioxide with aluminum porphyrin and reaction with epoxide. Studies on (tetraphenylporphinato) aluminum alkoxide having a long oxyalkylene chain as the alkoxide group,” *Journal of the American Chemical Society*, vol. 105, no. 5, pp. 1304–1309, 1983.
- [185] M. Adinehnia, U. Mazur, and K. W. Hipps, “Predicting the size distribution in crystallization of TSPP:TMPyP binary porphyrin nanostructures in a batch desupersaturation experiment,” *Crystal Growth & Design*, vol. 14, no. 12, pp. 6599–6606, 2014.
- [186] P. Guo, P. Chen, and M. Liu, “One-dimensional porphyrin nanoassemblies assisted via graphene oxide: sheet-like functional surfactant and enhanced photocatalytic behaviours,” *ACS Applied Materials & Interfaces*, vol. 5, no. 11, pp. 5336–5345, 2013.
- [187] C. J. Medforth, Z. Wang, K. E. Martin, Y. Song, J. L. Jacobsen, and J. A. Shelnutt, “Selfassembled porphyrin nanostructures,” *Chemical Communications*, vol. 21, no. 47, pp. 7261–7277, 2009.
- [188] Y. Arai and H. Segawa, “J-aggregation of protonated meso-tetrakis(sulfonatothienyl)porphyrin isomers: morphological selection of self-assembled nanostructures from structurally similar zwitterionic porphyrins,” *Chemical Communications*, vol. 46, no. 24, pp. 4279–4281, 2010.
- [189] Y. Arai, K. Tsuzuki, and H. Segawa, “Homogeneously mixed porphyrin J-aggregates with rod-shaped nanostructures via zwitterionic self-assembly,” *Physical Chemistry Chemical Physics*, vol. 14, no. 3, pp. 1270–1276, 2012.
- [190] J. Lee and S. J. Lee, “Microscopic crystalline rods from the self-assembly of mixed porphyrin building blocks,” *Inorganic Chemistry Communications*, vol. 14, no. 6, pp. 1014–1017, 2010.
- [191] J. Bozja, J. Sherrill, S. Michielsen, and I. Stojiljkovic, “Porphyrin-base, light-activated antimicrobial materials,” *Journal of Polymer Science Part A: Polymer Chemistry*, vol. 41, no. 15, pp. 2297–2303, 2005.
- [192] A. S. Derycke and P. A. de Witte, “Liposomes for photodynamic therapy,” *Advanced Drug Delivery Reviews*, vol. 56, no. 1, pp. 17–30, 2004.
- [193] J. C. Leroux, E. Roux, D. Garrec, K. Hong, and D. C. Drummond, “N-isopropylacrylamide copolymers for the preparation of pH-sensitive liposomes and polymeric micelles,” *Journal of Controlled Release*, vol. 72, no. 1–3, pp. 71–84, 2001.
- [194] S. H. Joo, J. Y. Park, C. K. Tsung, Y. Yamada, P. Yang, and G. A. Somorjai, “Thermally stable Pt/mesoporous silica core-shell nanocatalysts for high-temperature reactions,” *Nature Materials*, vol. 8, no. 2, pp. 126–131, 2009.
- [195] T. Maschmeyer, G. Sankar, F. Rey, and J. M. Thomas, “Heterogeneous catalysts obtained by grafting metallocene complexes onto mesoporous silica,” *Nature*, vol. 378, no. 6553, pp. 159–162, 1995.
- [196] N. K. Mal, M. Fujiwara, and Y. Tanaka, “Photocontrolled reversible release of guest molecules from coumarin-modified mesoporous silica,” *Nature*, vol. 421, no. 6921, pp. 350–353, 2003.
- [197] E. Aznar, M. D. Marcos, R. Martínez-Máñez et al., “pH- and photo-switched release of guest molecules from mesoporous silica supports,” *Journal of the American Chemical Society*, vol. 131, no. 19, pp. 6833–6843, 2009.
- [198] D. M. Huang, Y. Hung, B. S. Ko et al., “Highly efficient cellular labelling of mesoporous nanoparticles in human mesenchymal stem cells: implication for stem cell tracking,” *FASEB Journal*, vol. 19, no. 14, pp. 2014–2039, 2005.
- [199] L. Li, C.-K. Tsung, T. Ming et al., “Multifunctional meso-structured silica microspheres from an ultrasonic aerosol spray,” *Advanced Functional Materials*, vol. 18, no. 19, pp. 2956–2962, 2008.
- [200] L.-L. Li, H. Sun, C.-J. Fang, J. Xu, J.-Y. Jin, and C.-H. Yan, “Optical sensors based on functionalized mesoporous silica SBA-15 for the detection of multianalytes (H⁺ and Cu²⁺) in water,” *Journal of Materials Chemistry*, vol. 17, no. 42, pp. 4492–4498, 2007.

- [201] A. Vinu, N. Gokulakrishnan, V. V. Balasubramanian et al., "Three dimensional ultralarge-pore Ia3d mesoporous silica with various pore diameters and their application in biomolecule immobilization," *Chemistry*, vol. 14, no. 36, pp. 11529–11538, 2008.
- [202] P. Guo, G. Zhao, P. Chen et al., "Porphyrin nanoassemblies via surfactant-assisted assembly and single nanofiber nanoelectronic sensors for high-performance H₂O₂ vapor sensing," *Journal of the American Chemical Society*, vol. 8, no. 4, pp. 3402–3411, 2014.
- [203] T. Hasobe, "Photo- and electro-functional self-assembled architectures of porphyrins," *Physical Chemistry Chemical Physics*, vol. 14, no. 46, pp. 15975–15987, 2012.
- [204] C. Pavani, A. F. Uchoa, C. S. Oliveira, Y. Yamamoto, and M. S. Baptista, "Effect of zinc insertion and hydrophobicity on membrane interaction and PDT activity of porphyrin photosensitizers," *Photochemical and Photobiological Sciences*, vol. 8, no. 2, pp. 233–240, 2008.
- [205] T. Uno, K. Aoki, T. Shikimi, Y. Hiranuma, Y. Tomisugi, and Y. Ishikawa, "Copper insertion facilitates water-soluble porphyrin binding to rA, rU and rA, dT base pairs in duplex RNA and RNA, DNA hybrids," *Biochemistry*, vol. 41, no. 43, pp. 13059–13066, 2002.
- [206] A. B. Ormond and H. S. Freeman, "Dye sensitizers for photodynamic therapy," *Materials*, vol. 6, no. 3, pp. 817–840, 2013.
- [207] T. Maisch, C. Bosl, R. M. Szeimies, B. Love, and C. Abels, "Determination of the environmental efficacy of a new porphyrin-based photosensitizer against MRSA ex-vivo," *Photochemical & Photobiological Sciences*, vol. 6, no. 5, pp. 545–551, 2007.
- [208] T. Maisch, C. Bosl, R. M. Szeimies, N. Lehn, and C. Abels, "Photodynamic effects of novel XF porphyrin derivatives on prokaryotic and eukaryotic cells," *Antimicrob Agents Chemother*, vol. 49, no. 4, pp. 1542–1553, 2005.
- [209] H. Aghaie, M. Keshavarz, K. Zare et al., "Spectroscopic study in the interaction of three water-soluble porphyrins with calf thymus DNA," *Journal of Physical & Theoretical Chemistry*, vol. 1, no. 3, pp. 135–142, 2004.
- [210] E. B. Fleischer, "Structure of porphyrins and metalloporphyrins," *Accounts of Chemical Research*, vol. 3, no. 3, pp. 105–112, 1970.
- [211] T. Maisch, C. Bosl, R.-M. Szeimies, N. Lehn, and C. Abels, "Photodynamic effects of novel XF porphyrin derivatives on prokaryotic and eukaryotic cells," *Antimicrobial Agents and Chemotherapy*, vol. 49, no. 4, pp. 1542–1552, 2005.
- [212] Y. Tian, K. E. Martin, J. Shelnett et al., "Morphological families of self-assemble porphyrin structures and their photosensitization of hydrogen generation," *Chemical Communications*, vol. 47, pp. 6069–6071, 2011.
- [213] A. Lavi, H. Weitman, R. T. Holmes et al., "The depth of porphyrin in a membrane and the membranes physical properties affect the photosensitizing efficiency," *Biophysical Journal*, vol. 82, no. 4, pp. 2101–2110, 2002.
- [214] J. P. C. Tomé, M. G. P. M. S. Neves, A. C. Tomé et al., "Synthesis and antibacterial activity of new poly-S-lysine-porphyrin conjugates," *Journal of Medicinal Chemistry*, vol. 47, no. 26, pp. 6649–6652, 2004.
- [215] A. Srivatsan, J. R. Missert, S. K. Upadhyay, and R. K. Pandey, "Porphyrin based photosensitizers and corresponding multifunctional nanoplatforms for cancer –imaging and phototherapy," *Journal of Porphyrins and Phthalocyanines*, vol. 19, no. 1–3, pp. 109–134, 2015.
- [216] T. G. Smijs, R. N. van der Haas, J. Lugtenburg et al., "Photodynamic Treatment of the Dermatophyte Trichophyton Rubrum and its Mivronidia with Porphyrin Photosensitizers," *Photochemistry and Photobiology*, vol. 80, no. 2, pp. 197–202, 2004.
- [217] F. Würthner, C. C. You, and C. R. Saha-Möller, "Metallo-supramolecular squares: from structure to function," *Chemical Society Reviews*, vol. 33, no. 3, pp. 133–146, 2004.
- [218] F. M. Engelmann, I. Mayer, D. S. Gabrielli et al., "Interaction of cationic meso-porphyrins with liposomes, mitochondria and erythrocytes," *Journal of Bioenergetics and Biomembranes*, vol. 39, no. 2, pp. 175–185, 2007.
- [219] D. K. Cabbiness and D. W. Margerum, "Effect of macrocyclic structures on the rate of formation and dissociation of copper(II) complexes," *Journal of the American Chemical Society*, vol. 92, no. 7, pp. 2151–2153, 1970.
- [220] R. West and A. Hill, *Advances in Organometallic Chemistry*, vol. 58, pp. 289–342, 1st edition, 2001.
- [221] S. H. van Rijt and P. J. Sadler, "Current applications and future potential for bioinorganic chemistry in the development of anticancer drugs," *Drug Discovery Today*, vol. 14, no. 23–24, pp. 1089–1097, 2009.
- [222] P. Comba, H. Rudolf, and H. Wadepohl, "Synthesis and transition metal coordination chemistry of a novel hexadentate bispidine ligand," *Dalton Transactions*, vol. 44, no. 6, pp. 2724–2736, 2015.
- [223] M. Scirish and H. J. Scheureder, "Porphyrin derivatives as water soluble receptors for peptides," *Chemical Communications*, vol. 10, pp. 907–908, 1999.
- [224] F. R. Longo, E. M. Brown, D. J. Quimby, A. D. Adler, and M. Meot-Ner, "Kinetic studies on metal chelation by Porphyrins," *Annals of the New York Academy of Sciences*, vol. 206, no. 1, pp. 420–442, 1973.
- [225] X. Gong, T. Milic, C. Xu, J. D. Batteas, and C. M. Drain, "Preparation and characterization of porphyrin nanoparticles," *Journal of the American Chemical Society*, vol. 124, no. 48, pp. 14290–14291, 2002.
- [226] O. Penon, T. Patiño, L. Barrios et al., "A new pophyrin for the preparation of functionalized water-soluble gold nanoparticle with intrinsic toxicity," *ChemistryOpen*, vol. 4, no. 2, pp. 127–136, 2015.
- [227] K. Záruba, J. Králová, P. Řezanka, P. Poučková, L. Veverková, and V. Král, "Modified porphyrin–brucine conjugated to gold nanoparticles and their application in photodynamic therapy," *Organic and Biomolecular Chemistry*, vol. 8, no. 14, pp. 3202–3206, 2010.
- [228] T. Zhao, H. Wu, S. Q. Yao, Q.-H. Xu, G. Q. Xu, and G. Q. Xu, "Nanocomposites containing gold nanorods and porphyrin-doped mesoporous silica with dual capability of two-photon imaging and photosensitization," *Langmuir*, vol. 26, no. 18, pp. 14937–14942, 2010.
- [229] C. Xue, Y. Xu, Y. Pang et al., "Organo-soluble porphyrin mixed monolayer-protected gold nanorods with intercalated fullerenes," *Langmuir*, vol. 28, no. 14, pp. 5956–5963, 2012.
- [230] K. Bera, S. Maiti, M. Maity, C. Mandal, and N. C. Maiti, "Porphyrin-gold nanomaterial for efficient drug delivery to cancerous cells," *ACS Omega*, vol. 3, no. 4, pp. 4602–4619, 2018.
- [231] D. van Straten, V. Mashayekhi, H. S. de Bruijn, S. Oliveira, and D. Robinson, "Oncologic photodynamic therapy: basic principles, current clinical status and future directions," *Cancers*, vol. 9, no. 2, p. 19, 2017.
- [232] L. G. Levy, "Photosensitizers in photodynamic therapy," *Seminars in Oncology*, vol. 21, no. 150, pp. 4–10, 1994.

- [233] M. Panjehpour, R. C. DeNovo, M. G. Petersen et al., "Photodynamic therapy using Verteporfin (benzoporphyrin derivative monoacid ring A, BPD-MA) and 630 nm laser light in canine esophagus," *Lasers in Surgery and Medicine*, vol. 30, no. 1, pp. 26–30, 2002.
- [234] K. Bera, S. Maiti, M. Maity, C. Mandal, and N. C. Maiti, "Porphyrin-gold nanomaterial for efficient drug delivery to cancerous cells," *ACS Omega*, vol. 3, no. 4, pp. 4602–4619, 2018.
- [235] T. H. Foster, B. R. Giesselman, R. Hu, M. E. Kenney, and S. Mitra, "Intratumor administration of the photosensitizer Pc 4 affords photodynamic therapy efficacy and selectivity at short drug-light intervals," *Translational Oncology*, vol. 3, no. 2, pp. 135–141, 2010.
- [236] Z.-A. Chen, Y. Kuthati, R. K. Kankala et al., "Encapsulation of palladium porphyrin photosensitizer in layered metal oxide nanoparticles for photodynamic therapy against skin melanoma," *Science and Technology of Advanced Materials*, vol. 16, no. 5, pp. 1–14, 2015.
- [237] M. Scholz and M. Didie, "New trends in photodynamic therapy research," *Proceedings of Contributed Papers. Part III*, pp. 46–51, 2012.
- [238] D. Bechet, P. Couleaud, C. Frochot, M.-L. Viriot, F. Guillemain, and M. Barberi-Heyob, "Nanoparticles as vehicles for delivery of photodynamic therapy agents," *Trends in Biotechnology*, vol. 26, no. 11, pp. 612–621, 2008.
- [239] A. C. S. Samia, S. Dayal, and C. Burda, "Quarter dot-based energy transfer: perspectives and potential applications in photodynamic therapy," *Photochemistry and Photobiology*, vol. 82, no. 3, pp. 617–625, 2006.
- [240] T. Dai, B. B. Fuchs, J. J. Coleman et al., "Concepts and principles of photodynamic therapy as an alternative antifungal discovery platform," *Frontiers in Microbiology*, vol. 3, no. 120, pp. 1–13, 2012.
- [241] R. R. Allison, G. H. Downie, R. Cuenca, X.-H. Hu, C. J. H. Childs, and C. H. Sibata, "Photosensitizers in clinical PDT," *Photodiagnosis and Photodynamic Therapy*, vol. 1, no. 1, pp. 27–42, 2004.
- [242] J.-Y. Liu, C. Wang, C.-H. Zhu, Z.-H. Zhang, and J.-P. Xue, "Preparation and in vitro photodynamic activity of glucosylated zinc(II) phthalocyanines as underlying targeting photosensitizers," *Molecules*, vol. 22, no. 845, pp. 1–13, 2017.
- [243] Z. Kautzka, S. Clement, E. M. Goldys, and W. Deng, "Light-triggered liposomal cargo delivery platform incorporating photosensitizers and gold nanoparticles for enhanced singlet oxygen generation and increased cytotoxicity," *International Journal of Nanomedicine*, vol. 12, pp. 969–977, 2017.
- [244] T. Kiesslich, A. Gollmer, T. Maisch, M. Berneburg, and K. Plaetzer, "A comprehensive tutorial on vitro characterization of new photosensitizers for photodynamic antitumor therapy and photodynamic inactivation of microorganisms," *BioMed Research International*, vol. 2013, Article ID 840417, 17 pages, 2013.
- [245] C. J. Gomer, N. Rucker, A. Ferrario, and S. Wong, "New applications of photodynamic therapy," *Radiation Research*, vol. 120, no. 1, pp. 1–18, 1989.
- [246] J. Kou, D. Dou, and L. Yang, "Porphyrin photosensitizers in photodynamic therapy and its applications," *Oncotarget*, vol. 8, no. 46, pp. 81591–81603, 2017.
- [247] J. Zhang, C. Jiang, J. P. F. Longo et al., "An updated overview on the development of new photosensitizers for anticancer photodynamic therapy," *Acta Pharmaceutica Sinica B*, vol. 8, no. 2, pp. 137–146, 2017.
- [248] C. Staneloudi, K. S. Smith, R. Hudson et al., "Development and characterization of novel photosensitizer: scFv conjugates for use in photodynamic therapy of cancer," *Immunology*, vol. 120, no. 4, pp. 512–517, 2007.
- [249] A. Almeida, A. Cunha, N. C. M. Gomes, E. Alves, L. Costa, and M. A. Faustino, "Phage therapy and photodynamic therapy: low environmental impact approaches to inactivate microorganisms in fish farming plants," *Marine Drugs*, vol. 7, no. 3, pp. 268–313, 2009.
- [250] T. Huang, T. Dai, and M. R. Hamblin, "Antimicrobial photodynamic inactivation and photodynamic therapy for infections," *Methods in Molecular Biology*, vol. 635, pp. 155–173, 2010.
- [251] S. Perni, P. Prokopovich, J. Pratten, I. P. Parkin, and M. Wilson, "Nanoparticles: their potential use in antibacterial photodynamic therapy," *Photochemical and Photobiological Sciences*, vol. 10, no. 5, pp. 712–720, 2010.
- [252] N. Shirasu, S. O. Nam, and M. Kuroki, "Tumor targeted photodynamic therapy," *Anticancer Research*, vol. 33, no. 7, pp. 2823–2832, 2013.
- [253] C. M. Allen, W. M. Sharman, and J. E. van Lier, "Current status of phthalocyanines in the photodynamic therapy of cancer," *Phthalocyanines*, vol. 5, no. 2, pp. 161–169, 2009.
- [254] S. Wang, R. Gao, F. Zhou, and M. Selke, "Nanomaterials and singlet oxygen photosensitizers: potential application in photodynamic therapy," *Journal of Materials Chemistry*, vol. 14, no. 4, pp. 487–493, 2004.
- [255] Y. N. Konan, R. Gurny, and E. Allemann, "State of the art in the delivery of photosensitizers for photodynamic therapy," *Journal of Photochemistry and Photobiology*, vol. 66, no. 2, pp. 89–106, 2002.
- [256] R. Bonnett, "Photosensitizers of the porphyrin and phthalocyanine series for photodynamic therapy," *Chemical Society Reviews*, vol. 25, pp. 9–33, 1995.
- [257] M. C. Hashimoto, R. A. Prates, I. T. Kato, S. C. Núñez, L. C. Courrol, and M. S. Ribeiro, "Antimicrobial photodynamic therapy on drug resistant pseudomonas aeruginosa-induced infection: an in vivo study," *Photochemistry and Photobiology*, vol. 88, no. 3, pp. 590–595, 2012.
- [258] E. Paszko, C. Ehrhardt, M. O. Senge, D. P. Kelleher, and J. V. Reynolds, "Nanodrug applications in photodynamic therapy," *Photodiagnosis and Photodynamic Therapy*, vol. 8, no. 1, pp. 14–29, 2011.
- [259] J. Estelrich, M. J. Sánchez-Martín, and M. A. Busquets, "Nanoparticles in magnetic resonance imaging: from simple to dual contrast agents," *International Journal of Nanoscience*, vol. 10, pp. 1727–1741, 2015.
- [260] W. C. M. A. de Melo, A. N. Lee, J. R. Perussi, and M. R. Hamblin, "Electroporation enhances antimicrobial photodynamic therapy mediated by the hydrophobic photosensitizer, hypericin, electroporation enhances antimicrobial photodynamic inactivation," *Photodiagnosis and Photodynamic Therapy*, vol. 10, no. 4, pp. 647–650, 2013.
- [261] W. Chen and J. Zhang, "Using nanoparticles to enable simultaneous radiation and photodynamic therapies for cancer treatment," *Journal of Nanoscience and Nanotechnology*, vol. 6, no. 4, pp. 1159–1166, 2006.
- [262] P. Agostinis, K. Berg, K. A. Cengel et al., "Photodynamic therapy of cancer: an update," *CA: A Cancer Journal for Clinicians*, vol. 61, no. 4, pp. 250–281, 2011.
- [263] A. Casas, H. Fukuda, G. Di Venosa, and A. Batlle, "Photosensitization and mechanism of cytotoxicity induced by

- the use of ALA derivatives in photodynamic therapy," *British Journal of Cancer*, vol. 85, no. 2, pp. 279–284, 2001.
- [264] E. Otake, S. Sakuma, K. Torii et al., "Effect and mechanism of a new photodynamic therapy with glycoconjugated fullerene," *Photochemistry and Photobiology*, vol. 86, no. 6, pp. 1356–1363, 2010.
- [265] J. Usuda, H. Kato, T. Okunaka et al., "Photodynamic therapy (PDT) for lung cancers," *Journal of Thoracic Oncology*, vol. 1, no. 5, pp. 489–493, 2006.
- [266] S. P. Songca and O. S. Oluwafemi, "Photodynamic therapy: a new light for the developing world," *African Journal of Biotechnology*, vol. 12, no. 23, pp. 3590–3599, 2014.
- [267] L. Zou, H. Wang, B. He et al., "Current approaches of photothermal therapy in treating cancer metastasis with nanotherapeutics," *Theranostics*, vol. 6, no. 6, pp. 762–772, 2016.
- [268] W. Chen and J. Zhang, "Using nanoparticles to enable simultaneous radiation and photodynamic therapies for cancer treatment," *Journal of Nanoscience and Nanotechnology*, vol. 6, no. 4, pp. 1159–1166, 2006.
- [269] D. R. Cooper, D. Bekah, and J. L. Nadeau, "Gold nanoparticles and their alternatives for radiation therapy enhancement," *Frontiers in Chemistry*, vol. 2, no. 14, p. 86, 2014.
- [270] E. B. Dickerson, E. C. Dreaden, X. Huang et al., "Gold nanorod assisted near-infrared plasmonic photothermal therapy (PPTT) of squamous cell carcinoma in mice," *Cancer Letters*, vol. 269, no. 1, pp. 57–66, 2008.
- [271] J. Chen, D. Wang, J. Xi et al., "Immuno gold nanocages with tailored optical properties for targeted photothermal destruction of cancer cells," *Nano Letters*, vol. 7, no. 5, pp. 1318–1322, 2007.
- [272] C. M. Pitsillides, E. K. Joe, X. Wei, R. Rox Anderson, and C. P. Lin, "Selective cell targeting with light-absorbing microparticles and nanoparticles," *Biophysical Journal*, vol. 84, no. 6, pp. 4023–4032, 2003.
- [273] I. H. El-Sayed, X. Huang, and M. A. El-Sayed, "Selective laser photo-thermal therapy of epithelial carcinoma using anti-EGFR antibody conjugated gold nanoparticles," *Cancer Letters*, vol. 239, no. 1, pp. 129–135, 2006.
- [274] Q. Tian, J. Hu, Y. Zhu et al., "Sub-10 nm Fe₃O₄@Cu₂-XS core-shell nanoparticles for dual-modal imaging and photothermal therapy," *Journal of the American Chemical Society*, vol. 135, no. 23, pp. 8571–8577, 2013.
- [275] A. L. Mckenziei and J. A. S. Carruth, "Lasers in surgery and medicine," *Physics in Medicine and Biology*, vol. 29, no. 6, pp. 619–641, 1984.
- [276] L. O. Svaasand, C. J. Gomer, and E. Morinelli, "On the physical rationale of laser induced hyperthermia," *Lasers in Medical Science*, vol. 5, no. 2, pp. 121–128, 1990.
- [277] W. Lu, A. K. Singh, S. A. Khan, D. Senapati, H. Yu, and P. C. Ray, "Gold nano-popcon based targeted diagnosis nanotherapy treatment, and in situ monitoring of photothermal therapy response of prostate acancer cells using surface enhanced Raman spectroscopy," *Journal of the American Chemical Society*, vol. 132, no. 51, pp. 18103–18114, 2010.
- [278] S. Lal, S. E. Clare, and N. J. Halas, "Nanoshell-enabled photothermal cancer therapy: impending clinical impact," *Accounts of Chemical Research*, vol. 41, no. 12, pp. 1842–1851, 2012.
- [279] T. B. Huff, M. N. Hansen, Y. Zhao et al., "Controlling the cellular uptake of gold nanorods," *Langmuir*, vol. 23, no. 4, pp. 1596–1599, 2007.
- [280] J. M. Harris and R. B. Chess, "Effect of pegylation on pharmaceuticals," *Nature Reviews: Drug Discoveries*, vol. 2, no. 3, pp. 214–221, 2003.
- [281] J. Pérez-Juste, I. Pastoriza-Santos, L. M. Liz-Marzán, and P. Mulvaney, "Gold nanorods: synthesis, characterization and applications," *Coordination Chemistry Reviews*, vol. 249, no. 17–18, pp. 1870–1901, 2005.
- [282] R. Gans, "Form of ultramicroscopic particles of silver," *Annals of Physics*, vol. 14, pp. 270–284, 1915.
- [283] X. Huang, I. H. El-Sayed, W. Qian et al., "Cancer cells assemble and align gold nanorods conjugated to antibodies to produce highly enhanced, sharp and polarized surface Raman spectra: a potential cancer diagnostic marker," *Nano Letters*, vol. 7, no. 6, pp. 1591–1597, 2007.
- [284] M. Moskovits, "Surface-enhanced spectroscopy," *Reviews of Modern Physics*, vol. 57, no. 3, pp. 783–826, 1985.
- [285] C. Adura, S. Guerrero, E. Salas et al., "Stable conjugates of peptides with gold nanorods for biomedical applications with reduced effects on cell viability," *ACS Applied Materials & Interfaces*, vol. 5, no. 10, pp. 4076–4085, 2013.
- [286] A. M. Alkilany, L. B. Thompson, S. P. Boulos, P. N. Sisco, and C. J. Murphy, "Gold nanodots; their potential for photothermal therapeutics and drug delivery tempered by the complexity of their biological interactions," *Advanced Drug Delivery Reviews*, vol. 64, no. 2, pp. 190–199, 2012.
- [287] K. A. Willets and R. P. Van Duyne, "Localized surface plasmon resonance spectroscopy and sensing," *Annual Review of Physical Chemistry*, vol. 58, no. 1, pp. 267–297, 2007.
- [288] C. L. Haynes and A. M. R. P. Van Duyne, "Surface-enhanced Raman spectroscopy," *Analytical Chemistry*, vol. 77, no. 17, pp. 338A–346A, 2005.
- [289] J. N. Anker, W. P. Hall, O. Lyandres, N. C. Shah, J. Zhao, and R. P. Van Duyne, "Biosensing with plasmonic nanosensors," *Nature Materials*, vol. 7, no. 6, pp. 442–453, 2008.
- [290] E. Y. Lukianova-Hleb, E. Y. Hanna, J. H. Hafner, and D. Lapotko, "Tunable plasmonic nanobubbles for cell therapeutics," *Nanotechnology*, vol. 21, no. 8, article 85102, 2016.
- [291] T. Lammers, S. Aime, W. E. Hennink, G. Storm, and F. Kiessling, "Theranostic nanomedicine," *Accounts of Chemical Research*, vol. 44, no. 10, pp. 1029–1038, 2011.
- [292] N. Khlebtsov, V. Bogatyrev, L. Dykman et al., "Analytical and theranostic applications of gold nanoparticles and multifunctional nanocomposites," *Theranostics*, vol. 3, no. 3, pp. 167–180, 2013.
- [293] J. Wang, M. You, G. Zhu et al., "Photosensitizer-gold nanorod composite for targeted multimodal therapy," *Small*, vol. 9, no. 21, pp. 3678–3684, 2013.
- [294] B. Jang, J. Y. Park, C. H. Tung, I.-H. Kim, and Y. Choi, "Gold nanorod-photosensitizer complex for near-infrared fluorescence imaging and photodynamic/photothermal therapy in vivo," *ACS Nano*, vol. 5, no. 2, pp. 1086–1094, 2011.
- [295] G. Terentyuk, E. Panfilova, V. Khanadeev et al., "Gold nanorods with a hematoporphyrin-loaded silica shell for dual-modality photodynamic and photothermal treatment of tumors in vivo," *Nano Research*, vol. 7, no. 3, pp. 325–337, 2014.
- [296] T. Liu, C. Wang, W. Cui et al., "Combined photothermal and photodynamic therapy delivered by PEGylated MoS₂ nanosheets," *Nanoscale*, vol. 6, no. 19, pp. 11219–11225, 2014.
- [297] S. Hayden, L. A. Austin, R. D. Near et al., "Heterogeneous gold catalysts and catalysis," *Photochemistry and Photobiology*, vol. 269, p. 34, 2013.
- [298] A. M. A. Elhissi, W. Ahmed, I. U. Hassan, V. R. Dhanak, and A. D'Emmanuel, "Carbon nanotubes in cancer therapy and

- drug delivery," *Journal of Drug Delivery*, vol. 2012, Article ID 837327, 10 pages, 2012.
- [299] Z. Liu, J. T. Robinson, S. M. Tabakman, K. Yang, and H. Dai, "Carbon materials for drug delivery and cancer therapy," *Materials Today*, vol. 14, no. 7-8, pp. 1-8, 2012.
- [300] J.-W. Kim, E. I. Galanzha, E. V. Shashkov, H.-M. Moon, and V. P. Zharov, "Golden carbon nanotubes as multimodal photoacoustic and photothermal high-contrast molecular agents," *Nature Nanotechnology*, vol. 4, no. 10, pp. 688-694, 2009.
- [301] G. Terentyuk, E. Panfilova, V. Khanadeev et al., "Gold nanorods with a hematoporphyrin-loaded silica shell for dual-modality photodynamic and photothermal treatment of tumors in vivo," *Nano Research*, vol. 7, no. 3, pp. 325-337, 2014.
- [302] R. Saito, G. Dresselhaus, and M. S. Dresselhaus, *Physical Properties of Carbon Nanotubes*, Imperial College Press, London, UK, 1998.
- [303] R.-S. Liu, H. M. Chen, and S.-F. Hu, "Synthesis and characterization of long gold nanorods," *IEEE Transactions on Electrical and Electronic Engineering*, vol. 2, no. 4, pp. 468-472, 2007.
- [304] B. Jang, J.-Y. Park, C.-H. Tung, I.-H. Kim, and Y. Choi, "Gold nanorod-photosensitizer complex for near-infrared fluorescence imaging and photodynamic/photothermal therapy in vivo," *ACS Nano*, vol. 5, no. 2, pp. 1086-1094, 2011.
- [305] S. G. Bachhav and D. R. Patil, "Synthesis and characterization of polyaniline-multiwalled carbon nanotube nanocomposites and its electrical percolation behavior," *American Journal of Materials Science*, vol. 5, no. 4, pp. 90-95, 2015.
- [306] C. Xua, G. Wu, Z. Liu, D. Wu, T. T. Meek, and Q. Han, "Preparation of copper nanoparticles on carbon nanotubes by electroless plating method," *Materials Research Bulletin*, vol. 39, no. 10, pp. 1499-1505, 2004.
- [307] Z. M. Markovic, L. M. Harhaji-Trajkovic, B. M. Todorovic-Markovic et al., "In vivo comparison of the photothermal anticancer activity of graphene nanoparticles and carbon nanotubes," *Biomaterials*, vol. 32, no. 4, pp. 1121-1129, 2011.
- [308] V. Butkova, A. V. Fokina, V. N. Nevedomskaya et al., "A template method for carbon nanotube production from sugar water," *Nanotech*, vol. 26, pp. 1-5, 2013.
- [309] G. L. Hornyak, A. C. Dillon, P. A. Parilla et al., "Template synthesis of carbon nanotubes," *Pergamons*, vol. 12, no. 1-4, pp. 83-88, 1999.
- [310] F. Hekmat, B. Sohrabi, and M. S. Rahmanifar, *Effect of Template on the Structure of Carbon Nanotubes Grown by Catalytic Chemical Vapor Deposition Method*, pp. 1-5, Elsevier, New York, NY, USA, 2017.
- [311] H. S. Ali, S. Ahmed, and G. A. Khan, "Gold nanoparticles in cancer therapy and diagnosis," *Pharmaceutical Research*, vol. 28, no. 2, pp. 237-259, 2011.
- [312] M. Samim, C. K. Prashant, A. K. Dinda et al., "Synthesis and characterization of gold nanorods and their application for photothermal cell damage," *International Journal of Nanomedicine*, vol. 6, pp. 1825-1831, 2011.
- [313] R.-S. Liu, H. M. Chen, and S.-F. Hu, "Synthesis and characterization of long gold nanorods," *IEEE Transactions on Electrical and Electronic Engineering*, vol. 2, no. 4, pp. 468-472, 2007.
- [314] E. C. Dreaden, L. A. Austin, M. A. Mackey, and M. A. El-Sayed, "Size matters: gold nanoparticles in targeted cancer drug delivery," *Therapeutic Delivery*, vol. 3, no. 4, pp. 457-478, 2012.
- [315] J. Gautier, E. Allard-Vannier, E. Munnier, M. Soucé, and I. Chourpa, "Recent advances in theranostic nanocarriers of doxorubicin based on iron oxide and gold nanoparticles," *Journal of Controlled Release*, vol. 169, no. 1-2, pp. 48-61, 2013.
- [316] M. Ballauff and Y. Lu, "'Smart' nanoparticles: preparation, characterization and applications," *Polymer*, vol. 48, no. 7, pp. 1815-1823, 2007.
- [317] T. Ci, L. Chen, L. Yu et al., "Tumor regression achieved by encapsulating a moderately soluble drug into a polymeric thermogel," *Scientific Reports*, vol. 4, no. 1, p. 5473, 2014.
- [318] R. Zou, X. Guo, J. Yang et al., "Selective etching of gold nanorods by ferric chloride at room temperature," *Cry-EngComm*, vol. 11, pp. 2797-2803, 2009.
- [319] P. Rothemund, "The study of chlorophyll and photosynthesis," *Journal of the American Chemical Society*, vol. 63, pp. 267-270, 1935.
- [320] Z. C. Sun, Y. B. She, Y. Zhou, X. F. Song, and K. Li, "Synthesis, characterization and spectral properties of substituted tetraphenylporphyrin iron chloride complexes," *Molecules*, vol. 16, no. 4, pp. 2960-2970, 2011.
- [321] G. R. Geier III and G. S. Lindsey, "Investigation of porphyrin-forming reactions. Part 1. Pyrrole aldehyde oligomerization in two-step, one-flask syntheses of meso-substituted porphyrins," *Journal of the Chemical Society*, vol. 2, no. 5, pp. 677-686, 2001.
- [322] K. Berg, P. K. Selbo, A. Weyergang et al., "Porphyrin-related photosensitizers for cancer imaging and therapeutic applications," *Journal of Microscopy*, vol. 288, no. 2, pp. 133-147, 2005.
- [323] E. Fagadar-Cosma, A. Lascu, A. Palade et al., "Hybrid material based on 5-(4-pyridyl)-10,15,20-tris (4-phenoxyphenyl)- porphyrin and gold colloid for CO₂ detection," *Digest Journal of Nanomaterials and Biostructures*, vol. 11, no. 2, pp. 419-424, 2016.
- [324] J. T. Robinson, K. Welsher, S. M. Tabakman et al., "High performance in vivo near-IR (>1 μm) imaging and photothermal cancer therapy with carbon nanotubes," *Nano Research*, vol. 3, no. 11, pp. 779-793, 2010.
- [325] V. Kumar, J. Sinha, N. Verma, K. Nayan, C. S. Saimbi, and A. K. Tripathi, "Scope of photodynamic therapy in periodontics," *Indian Journal of Dental Research*, vol. 26, no. 4, pp. 439-442, 2015.
- [326] P. Li, Y.-W. Shi, B.-X. Li et al., "Photo-thermal effect enhances the efficiency of radiotherapy using Arg-Gly-Asp peptides-conjugated gold nanorods that target α_vβ₃ in melanoma cancer cells," *Journal of Nanobiotechnology*, vol. 13, no. 1, p. 52, 2015.