



High-Quality Draft Genome Sequence of *Francisella tularensis* subsp. *holarctica* Strain OR96-0246

L. M. Atkins,^{a,b} M. E. Holder,^{a,b} N. J. Ajami,^{a,b} G. A. Metcalf,^c G. M. Weissenberger,^c M. Wang,^c V. Vee,^c Y. Han,^c D. M. Muzny,^c R. A. Gibbs,^c J. F. Petrosino^{a,b}

Department of Molecular Virology and Microbiology,^a Alkek Center for Metagenomics and Microbiome Research,^b and Human Genome Sequencing Center at Baylor College of Medicine,^c Houston, Texas, USA

The bacterial pathogen *Francisella tularensis* was recently renewed as a tier-one select agent. *F. tularensis* subsp. *tularensis* (type A) and *holarctica* (type B) are of clinical relevance. Here, we report the complete genome of a virulent *F. tularensis* type B strain and describe its usefulness in comparative genomics.

Received 1 July 2015 Accepted 6 July 2015 Published 13 August 2015

Citation Atkins LM, Holder ME, Ajami NJ, Metcalf GA, Weissenberger GM, Wang M, Vee V, Han Y, Muzny DM, Gibbs RA, Petrosino JF. 2015. High-quality draft genome sequence of *Francisella tularensis* subsp. *holarctica* strain OR96-0246. Genome Announc 3(4):e00898-15. doi:10.1128/genomeA.00898-15.

Copyright © 2015 Atkins et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to J. F. Petrosino, jpetrosi@bcm.edu.

Francisella tularensis is a Gram-negative intracellular coccobacillus and etiological agent of the zoonotic disease tularemia (1). Subspecies tularensis (type A) and holarctica (type B) are responsible for all tularemia-associated fatalities in the United States, with type A infections resulting in significantly higher mortality than type B infections (2). The Federal Select Agent Program classifies *F. tularensis* as a tier-one select agent due to its low infectious dose, high morbidity, and lack of a licensed vaccine (3, 4). An attenuated live vaccine strain (LVS) was empirically derived from repeated passage of a type B strain in the 1950s. While LVS offers less protection against inhalational type A, it is to date the only vaccine for which formal efficacy data in humans exists (5). LVS remains unlicensed, and its exact mechanism of attenuation is unknown.

Live attenuated type A strains are highly scrutinized as tularemia vaccines due to reversion concerns. Recent attempts to derive a live vaccine strain from type A were unsuccessful, as disruption of multiple loci abolished the protective ability of the attenuated strain (6–8). Therefore, virulent and potentially immunogenic type B strains are ideal sources of a live attenuated vaccine. To this end, a study reported success with an attenuated type B strain (FSC200 $\Delta clpB$), which afforded greater protection against respiratory SchuS4 (type A) challenge than LVS in mice (8). However, as a chaperone protein, ClpB is likely to interact with many proteins and therefore does not address specific mechanisms of *Francisella* attenuation or virulence.

Studies in type B strains offer the major advantage that new information will build off of current knowledge and positive attributes of LVS. *F. tularensis* subsp. *holarctica* strain OR96-0246 was isolated in 1996 from a primate facility outbreak in Oregon (9, 10). OR96-0246 NR-648 was obtained through BEI Resources, NIAID, NIH, and grown in cation-adjusted Mueller-Hinton (MHII) broth (Becton Dickinson) supplemented with sterile 0.1% glucose (Fisher), 0.025% ferric pyrophosphate (Sigma), and 2% reconstituted IsoVitaleX (Becton Dickinson). A total of 10 to 20 μ g of high-molecular-weight genomic DNA was extracted from overnight cultures and precipitated with ethanol (11). Extracts were plated on MHII agar (Becton, Dickinson) to confirm sterility. DNA was sequenced on the Pacific Biosciences RS II platform according to the manufacturer's protocols. Reads were fed into the Celera assembler and annotated using the NCBI tools Glimmer and GeneMark (12, 13).

We report a high-quality draft of the OR96-0246 genome as one circle, with a gap, comprising 1,879,883 bases with 32.2% GC content and an average reference coverage of 142. The postfilter read N_{50} is 16,096 bases, with a quality value of 0.833. Annotation features include 1,976 genes, 213 pseudogenes, 10 rRNAs, 38 tRNAs, and 1 noncoding RNA (ncRNA). Two 23-kb repetitive regions encode the *Francisella* pathogenicity island (FPI), of which one region is fully represented in the assembly. The assembly process stacked all of the internal reads of the repeat into the assembled copy, confirmed by double coverage depth (>350).

Nucleotide sequence accession number. The sequence of OR96-0246 has been deposited in NCBI GenBank under the accession no. CP011488.

ACKNOWLEDGMENTS

This work was supported by the NIH/NIAID funding source U54 AI057156 and generous funds gifted by the Alkek Foundation.

REFERENCES

- Francis E. 1925. Tularemia. JAMA 84:1243–1250. http://dx.doi.org/ 10.1001/jama.1925.02660430001001.
- Kugeler KJ, Mead PS, Janusz AM, Staples JE, Kubota KA, Chalcraft LG, Petersen JM. 2009. Molecular epidemiology of *Francisella tularensis* in the United States. Clin Infect Dis 48:863–870. http://dx.doi.org/10.1086/ 597261.
- 3. Centers for Disease Control and Prevention. HHS and USDA select agents and toxins: 7CFR, part 331, 9 CFR part 121, and 42 CFR part 73. Centers for Disease Control and Prevention, Atlanta, GA. http://www .selectagents.gov/SelectAgentsandToxinsList.html.
- Kaufmann AF, Meltzer MI, Schmid GP. 1997. The economic impact of a bioterrorist attack: are prevention and postattack intervention programs justifiable? Emerg Infect Dis 3:83–94. http://dx.doi.org/10.3201/ eid0302.970201.

- Burke DS. 1977. Immunization against tularemia: analysis of the effectiveness of live *Francisella tularensis* vaccine in prevention of laboratoryacquired tularemia. J Infect Dis 135:55–60. http://dx.doi.org/10.1093/ infdis/135.1.55.
- 6. Twine S, Shen H, Harris G, Chen W, Sjostedt A, Ryden P, Conlan W. 2012. BALB/c mice, but not C57BL/6 mice immunized with a $\Delta clpB$ mutant of *Francisella tularensis* subspecies *tularensis* are protected against respiratory challenge with wild-type bacteria: association of protection with postvaccination and postchallenge immune responses. Vaccine 30: 3634–3645. http://dx.doi.org/10.1016/j.vaccine.2012.03.036.
- Ryden P, Twine S, Shen H, Harris G, Chen W, Sjostedt A, Conlan W. 2013. Correlates of protection following vaccination of mice with gene deletion mutants of *Francisella tularensis* subspecies *tularensis* strain, SCHU S4 that elicit varying degrees of immunity to systemic and respiratory challenge with wild-type bacteria. Mol Immunol 54:58–67. http:// dx.doi.org/10.1016/j.molimm.2012.10.043.
- 8. Golovliov I, Twine SM, Shen H, Sjostedt A, Conlan W. 2013. A $\Delta clpB$ mutant of *Francisella tularensis* subspecies *holarctica* strain, FSC200, is a more effective live vaccine than *F. tularensis* LVS in a mouse respiratory

challenge model of tularemia. PLoS One 8:e78671. http://dx.doi.org/ 10.1371/journal.pone.0078671.

- Versage JL, Severin DD, Chu MC, Petersen JM. 2003. Development of a multitarget real-time TaqMan PCR assay for enhanced detection of *Francisella tularensis* in complex specimens. J Clin Microbiol 41:5492–5499. http://dx.doi.org/10.1128/JCM.41.12.5492-5499.2003.
- Farlow J, Wagner DM, Dukerich M, Stanley M, Chu M, Kubota K, Petersen J, Keim P. 2005. *Francisella tularensis* in the United States. Emerg Infect Dis 11:1835–1841. http://dx.doi.org/10.3201/ eid1112.050728.
- 11. Sambrook J, Russell DW. 2001. Appendix 8, p 9–10. Molecular cloning: a laboratory manual, 3rd ed, vol 3, Cold Spring Harbor, New York, Cold Spring Harbor Laboratory Press.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with Glimmer. Nucleic Acids Res 27: 4636-4641. http://dx.doi.org/10.1093/nar/27.23.4636.
- Lukashin AV, Borodovsky M. 1998. GeneMark.Hmm: new solutions for gene finding. Nucleic Acids Res 26:1107–1115. http://dx.doi.org/10.1093/ nar/26.4.1107.