



Review

miRNA-Mediated Epigenetic Regulation of Treatment Response in RA Patients—A Systematic Review

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Abstract: This study aimed to evaluate the role of microRNAs (miRNA) as biomarkers of treatment response in rheumatoid arthritis (RA) patients through a systematic review of the literature. The MEDLINE and Embase databases were searched for studies including RA-diagnosed patients treated with disease-modifying antirheumatic drugs (DMARDs) that identify miRNAs as response predictors. Review inclusion criteria were met by 10 studies. The main outcome of the study was the response to treatment, defined according to EULAR criteria. A total of 839 RA patients and 67 healthy donors were included in the selected studies. RA patients presented seropositivity for the rheumatoid factor of 74.7% and anti-citrullinated C-peptide antibodies of 63.6%. After revision, 15 miRNAs were described as treatment response biomarkers for methotrexate, anti-tumour necrosis factor (TNF), and rituximab. Among treatments, methotrexate presented the highest number of predictor miRNAs: miR-16, miR-22, miR-132, miR-146a and miR-155. The most polyvalent miRNAs were miR-146a, predicting response to methotrexate and anti-TNF, and miR-125b, which predicts response to infliximab and rituximab. Our data support the role of miRNAs as biomarkers of treatment response in RA and point to DMARDs modifying the miRNAs expression. Nevertheless, further studies are needed since a meta-analysis that allows definitive conclusions is not possible due to the lack of studies in this field.

Keywords: microRNA; rheumatoid arthritis; treatment; biomarker; disease-modifying antirheumatic drug; systemic review



Citation: Mucientes, A.; Lisbona, J.M.; Mena-Vázquez, N.; Ruiz-Limón, P.; Manrique-Arija, S.; Fernández-Nebro, A. miRNA-Mediated Epigenetic Regulation of Treatment Response in RA Patients—A Systematic Review. *Int. J. Mol. Sci.* **2022**, *23*, 12989. <https://doi.org/10.3390/ijms232112989>

Academic Editor: George N. Goulielmos

Received: 13 September 2022

Accepted: 24 October 2022

Published: 27 October 2022

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1. Introduction

Rheumatoid arthritis (RA) is an immune-mediated inflammatory disease characterised by chronic synovial inflammation, progressive and irreversible joint destruction and functional disability. It has been estimated that RA affects 0.5–1% of the adult population worldwide [1]. Nevertheless, the detailed mechanisms underlying the pathogenesis of RA, disease activity and its severity remain not fully elucidated [2]. Consequently, there is no reliable biomarker for RA diagnosis. A holistic understanding of RA including both genetic and epigenetic (DNA methylation, microRNA and histone modifications) perspectives is needed for early diagnosis and personalised treatment [3–5]. Currently, there is no cure for RA. Disease-modifying antirheumatic drugs (DMARDs) are the conventional therapeutic approach to treat RA. In recent years, the therapeutic options were expanded by developing tumour necrosis factor (TNF) inhibitors and other biological agents that have improved both the management and prognosis of RA [6]. Despite these benefits, clinical practice

has demonstrated that these biological agents are not effective for all RA patients: 20% to 40% of RA patients discontinue the treatment due to several reasons, such as lack of efficacy in treatment, adverse events or inability to afford its high cost [7,8]. Therefore, identifying reliable markers of response to treatment will benefit the patient's quality of life and optimise healthcare resources.

Small RNAs (sRNA) are RNA molecules, usually non-coding, and 18–30 nucleotides in length. MicroRNAs (miRNA), the most studied sRNAs, are endogenous, non-coding, single-stranded, highly conserved, and 20–22-nucleotide-long RNAs [9]. To date, miRNA genes are estimated to constitute 1–2% of the complete genome, and more than 2000 miRNAs have been identified [10]. After being transcribed, mature miRNAs interact with their messenger RNA (mRNA) targets by hybridising into complementary sequences in the mRNA 3'-UTR regions [1]. This binding results in translational repression or mRNA degradation, regulating the gene expression [11]. Around a third of the total protein-coding genes are controlled by miRNAs, whereas 60% of the genes present miRNA-binding domains [12]. Thus, miRNAs act as an epigenetic control agent determining gene expression, acquiring a pivotal role in biological processes. miRNAs present good stability, and they can be detected in tissue samples and different biological fluids: blood, serum, saliva, plasma, and urine [9,13]. Furthermore, miRNAs can be determined by relative economic, simple and reproducible assays.

The described characteristics give miRNAs a potential role as biomarkers of diseases in which they have been described as being involved. Thus, miRNAs are used as biomarkers of cancers, cardiovascular and autoimmunity diseases, including RA [14–16]. In RA patients, it has been communicated altered levels of miRNA in blood, plasma, synovial fluid and cells lining the joint capsule [17–19]. This makes sense, as miRNAs are related to the proliferation and differentiation of inflammatory cytokines, synovial cells and osteoclast [10]. Moreover, miRNAs contribute to inflammation formation and immune response, which are altered in autoimmune diseases [20,21]. Described altered miRNA in RA may be potential biomarkers of the disease state, gravity, prognosis or treatment response: Bhanji et al. described the role of miRNA in RA progression in 2007 [22], and Filkova et al. proposed that both disease severity and duration, or the effect of treatment, could modulate the levels of circulating miRNAs in established RA patients [23].

All these data indicate a major role of the miRNAs in the RA epigenome. As stated above, markers of treatment response are needed. Therefore, it is plausible to consider miRNA levels as biomarkers of treatment response in the RA context. In recent years, several studies have tested miRNAs as biomarkers of both conventional synthetic DMARDs (csDMARDs) and biological DMARDs (bDMARDs). All the studies referenced present differences in methodology and results [1,13,24–30]. Hence, we aim to systematically review and resume the recent literature that analyses the role of miRNAs as biomarkers of treatment response in RA patients and provide an overview of the current literature. To our knowledge, this is the first work addressing this objective.

2. Methods

2.1. Search Strategy and Studies Selection

A systematic search was conducted for papers studying the expression of miRNAs in RA in relation to the therapeutic response in relation with the therapeutic response at both Medline and Embase databases using the following MeSH terms, Entry Terms and text-free: "Rheumatoid arthritis" and "miRNA" or "microRNA" or "microRNAs" and "variant" or "variants" or "mutation" or "polymorphism" or "polymorphisms" (Supplementary Table S1). Moreover, a secondary manual search of the related articles was also performed. It was restricted to the English language and human studies. The review protocol followed the declaration of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement (Figure 1). The research question was "Can miRNAs be useful as biomarkers of therapeutic response in RA?" and it was structured following the PICOS methodology (population, intervention, comparison, outcomes,

and study design). The search was performed by two researchers (A.M. and J.M.L.) who independently reviewed article titles and abstracts. Disagreement between reviewers on the inclusion/exclusion of studies was solved by consensus or with the assistance of a third reviewer (N.M-V.).

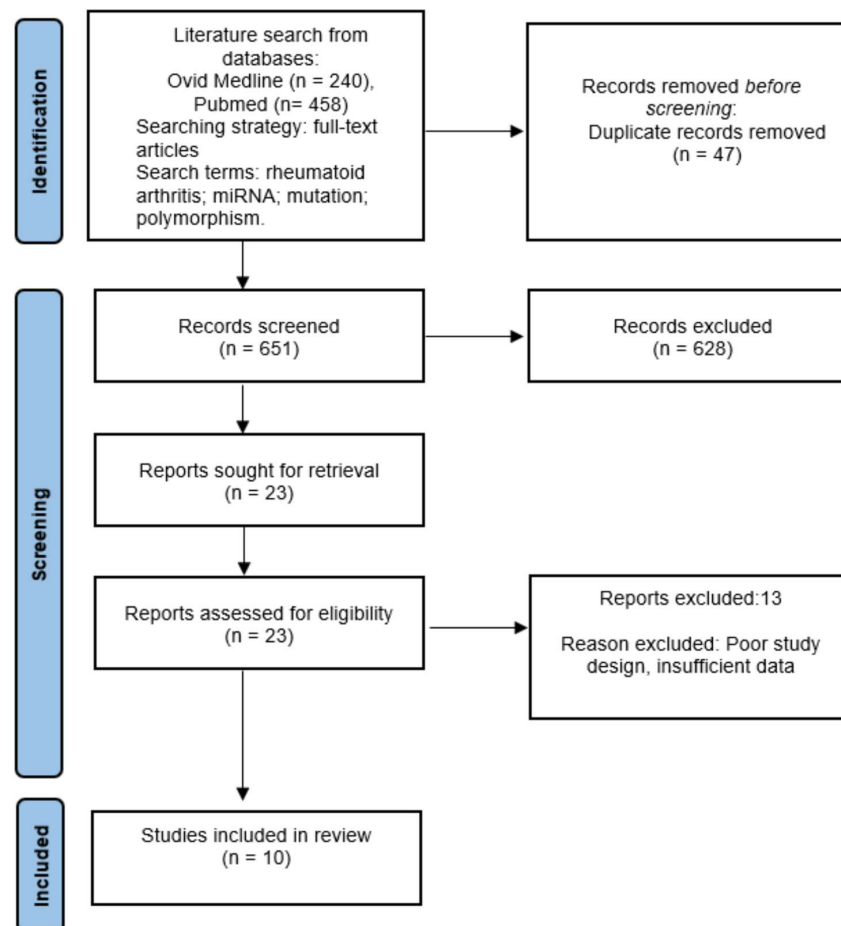


Figure 1. Flow diagram of study selection according to PRISMA statement.

2.2. Inclusion and Exclusion Criteria

These were the inclusion criteria for this review: (1) English language; (2) clinical trials, transversal studies, and case-control studies including adult RA patients according to ACR/EULAR classification criteria [17] treated with csDMARDs or bDMARDs; and (3) studies identifying miRNAs. Exclusion criteria were (1) editorials, narrative reviews, congress abstracts, case reports or case series with less than 30 cases; (2) inadequate description of methodology; (3) lack of data to evaluate the response to treatment; and (4) duplicated publications.

Regarding inclusion criterion 3, only studies that determine the specific association between miRNAs and treatment response were included. Thus, studies that communicated only the association between specific miRNAs with other RA characteristics were excluded.

2.3. Outcome Measures

The main outcome was a good therapeutic response measured by the disease activity score using 28 joint counts (DAS28) [31]. Response to treatment was defined according to EULAR criteria, based on the changes in the DAS28 score: an improvement in DAS28 over ≥ 1.2 and a DAS28 value ≤ 3.2 after treatment was considered a good response; a DAS28 value between 3.2 and 5.1 and a reduction between 0.6 and 1.2 was considered a moderate response; and patients with a DAS28 score > 5.1 or a reduction in DAS28 under 0.6 were considered non-responders [32].

2.4. Data Extraction and Measures of Study Quality

The whole text was read in the case of articles whose titles or abstracts met the inclusion criteria. Not fulfilling an eligibility criterion led the study to being excluded. In addition to the main and secondary outcome measures, we extracted information on the authors, year of publication, sex, age, autoantibodies, treatment groups, treatment response, miRNA associated and sample types of the patients. In order to evaluate the quality of the studies, the level of the evidence was assessed using the Scottish Intercollegiate Guidelines Network (SIGN) grading system [33].

3. Results and Discussion

3.1. Search

A systemic search was carried out, and 698 articles published between 1998 and 2022 were identified. Of them, 44 were duplicated and therefore eliminated from the study. After checking both the title and abstract, 628 articles were excluded. Finally, 13 articles were excluded after being fully read due to different causes (Table S2). Thus, 10 articles fulfilled the established inclusion criteria that constitute the focus of this systematic review (Figure 1).

3.2. Characteristics of the Included Studies

Eight observational studies and two clinical trials were included. The SIGN grading system results confirmed that all included studies present a low or very low bias risk (Table S3). Table 1 resumes the general characteristics of the included studies. Among the studies, 1/10 analysed the whole microRNome [24]; 5/10 assayed arrays of a different number of miRNAs—377 [26], 91 [28], 84 [27], 758 [13], and undetermined [1]; finally, 4/10 opted for a fine-mapping strategy by analysing a low number of specific miRNAs [25,29,30,34]. Regarding the type of sample, in all studies, peripheral blood was collected and, if needed, processed to obtain plasma, serum or peripheral blood mononuclear cells before miRNA isolation: 2/10 studies used plasma samples, 4/10 studies used serum samples, 2/10 studies used whole blood samples, 1/10 both whole blood and plasma samples, and 1/10 used peripheral blood mononuclear cells. Therefore, all studies analysed levels of circulating miRNAs.

The main characteristics of the patients are shown in Table 1. A total of 839 RA patients were included in the 10 studies after taking into account that 2/10 studies presented the same 108 RA patients, belonging to the OPERA cohort [35], but analysed different miRNAs [26,27]. Among the studies, 2/11 presented discovery and validation cohorts [13,28]; these cohorts were composed of 90 and 125 RA patients, respectively. Only in 3/10 studies, a healthy donor group was included [30,34], adding a total of 67 healthy subjects. Moreover, 2/10 studies included patients with different conditions: 13 ankylosing spondylitis patients, and 13 B lymphoma patients in addition to RA patients [30,34]. Overall, the female sex was predominant (74.5%), and the mean \pm SD age was 53.5 ± 5.3 years. All studies give information about RA patients' autoantibodies, and only Ciechomska et al. do not indicate if RA patients had previously had an inadequate response to at least one DMARD and/or were naïve to treatment (Table S3). Most RA patients presented seropositivity for rheumatoid factor (RF) of 74.7% and anti-citrullinated C-peptide antibodies (ACPA) of 63.6%. Moreover, patients included in the study of Luque-Tevar et al. [24] showed elevated titres for RF (112.9 ± 205.8) and ACPA (343.3 ± 762.6). Finally, information about glucocorticoid administration was lacking in 3/10 studies (Table S3). Only Cuppen et al. verified whether glucocorticoids have an effect on treatment response [13]. Their results showed a significant difference between responders and non-responders regarding the use of glucocorticoids in adalimumab (ADA) treatment (responders: 13%, non-responders: 50%, $p = 0.01$), but not in etanercept (ETN) treatment (responders: 20%, non-responders: 33%, $p = 0.25$).

Table 1. General characteristics of studies.

Author	Year	Country	microRNAs	Patients	Healthy Donors	Sex (% Women)	Age	Autoantibodies Information	Sample
Luque-Tevar et al. [24]	2012	Spain	Whole miRNome	104 RA patients	29 healthy donors	Patients: 65.5% Controls: 81.0%	Patients (mean \pm SD): 47 \pm 17.0 Controls (mean \pm SD): 51.2 \pm 10.5	ACPA, IU/mL (mean \pm SD): 343.3 \pm 762.6 RF, IU/mL (mean \pm SD): 112.9 \pm 205.8	Serum from whole blood
Krintel et al. [26]	2015	Denmark	377 miRNAs	180 RA patients (OPERA cohort)	None	Treatment group: 63% Placebo group: 69%	Treatment group: 56.2 (25.8–77.6) Placebo group: 54.2 (28.3–76.7)	Treatment group: ACPA positive: 60% RF positive: 70% Placebo group: ACPA positive: 70% RF positive: 74%	Whole blood
Sode et al. [27]	2017	Denmark	91 miRNAs	180 RA patients (OPERA cohort)	None	Treatment group: 63% Placebo group: 69%	Treatment group: 56.2 (25.8–77.6) Placebo group: 54.2 (28.3–76.7)	Treatment group: ACPA positive: 60% RF positive: 70% Placebo group: ACPA positive: 70% RF positive: 74%	Plasma from whole blood
Singh et al. [25]	2018	India	miR-132 miR-146a miR-155 let-7a	94 RA patients	None	86.2%	Patients (mean \pm SD): 40 \pm 17	RF positive: 85%	Whole blood
Ciechomska et al. [30]	2018	Poland	miRNA-5196	10 RA patients 13 AS patients	15 healthy controls	RA patients: 60% AS patients: 76.9% Controls: no data	RA patients: 59 (27–74) AS patients: 50 (32–59) Controls: no data	ACPA positive: 30% RF positive: 90%	Serum from whole blood
Castro-Villegas et al. [28]	2015	Spain	84 miRNAs.	Discovery cohort: 10 RA patients. Validation cohort: 85 RA patients.	None	Exploratory cohort: 90%. Validation cohort: 87.1%	Exploratory cohort: 54.6 (38–74) Validation cohort: 53.6 (24–72)	ACPA positive: 66.3% RF positive: 70.5%	Serum from whole blood
Duroux-Richard et al. [34]	2014	France	miR-125b	48 RA patients (32 treated with RTX)	13 healthy donors	84.7%	Patients (mean \pm SD): 58.8 \pm 7 Controls:	ACPA positive: 82.5%	Whole blood and serum
Cheng et al. [29]	2020	China	miR-125a miR-125b	96 active RA patients	None	80.2%	Patients (mean \pm SD): 58.6 \pm 10.0	ACPA positive: 62.5% RF positive: 71.9%	Plasma from whole blood
Cuppen et al. [13]	2016	Netherlands	758 miRNAs	RA patients were selected from the BiOCURA cohort. Discovery cohort: 80 RA patients. Validation cohort: 40 RA patients.	None	Discovery cohort: 76.3% Validation cohort: 67.5%	Discovery cohort (mean \pm SD): 55 \pm 11.0 Validation cohort (mean \pm SD): 56 \pm 10.0	Discovery cohort: ACPA positive: 71.3% RF positive: 73.8% Validation cohort: ACPA positive: 60% RF positive: 55%	Serum from whole blood
Liu et al. [1]	2019	China	microRNA array	92 active RA patients	None	80%	Patients (mean \pm SD): 55.6 \pm 8.8	ACPA positive: 77% RF positive: 82%	Peripheral blood mononuclear cells

Abbreviations. miRNAs: MicroRNAs; RA: rheumatoid arthritis. RF: rheumatoid factor; ACPA: anti-citrullinated C-peptide antibodies; AS, ankylosing spondylitis; SD: standard deviation.

3.3. miRNAs as Indicators for Treatment Response

Table 2 shows the microRNAs described as associated with the treatment response in the studies included in the present systemic review. These miRNAs were tested as biomarkers for both csDMARDs and bDMARDs.

Table 2. Baseline microRNAs associated with treatment response in RA.

Study	Treatment Groups	Treatment Response	Associated miRNA
Luque-Tevar et al. [24]	(1) IFX (2) ETN (3) ADA (4) GOL (5) CZP	At 3 months: Good: 35.4% Moderate: 31.7% No response: 32.9% At 6 months: Good: 49.4% Moderate: 20.2% No response: 30.4%	High levels of miR-106a were associated with good response.
Krintel et al. [26]	(1) ADA (2) Placebo	At 12 months: Good response (ADA): 72% Good response (Placebo): 63%	The combination of high expression of miR-886 with low expression of miR-22 was associated with a good response.
Sode et al. [27]	(1) MTX-ADA (2) MTX-Saline	At 3 months: Responders (MTX-ADA): 42.7% Responders (MTX-Saline): 24.2% At 12 months: Responders (MTX-ADA): 44.9% Responders (MTX-Saline): 28.6%	High levels of miR-27a were associated with a good response to MTX/ADA. High levels of miR-16 and miR-22 were associated with a good response to MTX
Singh et al. [25]	(1) MTX	At 4 months: Responders: 77.7%	Low levels of miR-132, miR-146a and miR-155 were associated with treatment response.
Ciechomska et al. [30]	(1) MTX-ETN (2) MTX-ADA	At 6 months: Responders: 100%	Expression of miR-5196 correlates with the RA state. (Low-size sample)
Castro-Villegas et al. [28]	(1) IFX (2) ETN (3) ADA	At 6 months: Responders: 89.5%	Expression of miR-23 and miR-223 as biomarkers and predictors of anti-TNF α /DMARDs combination therapy
Duroux-Richard et al. [34]	(1) RTX	At 3 months: Responders (Good/Moderate): 50%.	High expression of miR-125b was associated with good response in active RA patients.
Cheng et al. [29]	(1) IFX	At 24 weeks: Responders: 71.7%	miR-125b predicts treatment response at 24 weeks.
Cuppen et al. [13]	(1) ADA (2) ETN	At 12 months: Responders (ADA): 50% Responders (ETN): 50%	High levels of miR-99a and low levels of miR-143 were associated with ADA response. High levels of miR-23a and miR-197 were associated with ETN response.
Liu et al. [1]	(1) ETN	At 24 weeks: Responders: 65.2%	miR-146a predictive factor for good response. let-7a predictive factor for a worse response.

Abbreviations. RA: rheumatoid arthritis; IFX: infliximab, ETN: etanercept, ADA: adalimumab, GO: golimumab, CZP: certolizumab, MTX: methotrexate, RTX: rituximab.

An important characteristic in RA biologic treatment is whether the patient has shown an inadequate response to other(s) biologic treatment(s) or the patient is naïve to biologic drugs. Half of the included studies used a cohort composed of RA patients naïve to bDMARDs. Hence, Singh et al., after studying the expression in 94 RA patients, found that

miR-132, miR-146a and miR-155 are lowly expressed in MTX-responder RA patients [25]. Luque-Tevar et al., with a cohort composed of 104 RA patients and 29 healthy donors, observed that elevated levels of miR-106a were associated with a good response to several anti-TNF treatments [24]. In a previous study with a similar design, Castro-Villegas et al. informed that serum levels of miR-23 and miR-223 can predict response to anti-TNF treatment [28]. In their randomised clinical trial including treatment-naïve RA patients, Krintel et al. associated the low expression of miR-22 combined with miR-886 high expression with a good response to adalimumab (ADA), combined with methotrexate (MTX) [26], whereas Sode et al. concluded that higher expression of miR-27a prior treatment is associated with remission at 12 months in patients treated with the combination of ADA-MTX in their double-blinded placebo-controlled trial using the same cohort [27]. On the other hand, 4/10 of the studies included RA patients with inadequate response to at least one bDMARD, mostly anti-TNF. The initial results obtained by Cuppen et al. indicated that the expression of miR-99 and miR-143 predicts the ADA response, while the expression of miR-23a and miR-197 indicates the etanercept response, but no association kept the significance level in the validation cohort [13], whereas Liu et al. compared miRNAs levels between etanercept-responder and non-responder RA patients and validated two miRNAs as indicators of treatment response: miR-146a was overexpressed and let-7a was down-expressed in responders compared to non-responders [1]. Cheng et al. communicated that both miR-125a and miR-125b were elevated in RA patients compared to healthy controls, and the basal level of miR-125a predicted response after 24-week infliximab (IFX) treatment [29]. bDMARDs other than anti-TNF were also studied; in this sense, the results obtained by Duroux-Richard et al. indicated that elevated serum levels of miR-125b at disease flare are associated with good clinical response after 3 months of rituximab (RTX) treatment [34]. Furthermore, this predictive role was not limited to RA patients; it was also determined in B-cell lymphoma patients.

Finally, Ciechomska et al. did not inform about the prior use of bDMARDs, and after studying a small cohort of autoimmune patients, they established that the expression of miR-5196 positively correlates with the response to ADA and ETN combined with MTX in both RA and ankylosing spondylitis patients [30].

Disease-modifying antirheumatic drugs (DMARDs) are the conventional treatment for RA. Nevertheless, between 30 and 40% of the patients do not respond to the treatment, which involves maintained inflammatory activity, potential adverse effects, and continuous changes in treatment [36]. Hence, identifying reliable biomarkers for treatment response will have a positive impact on patient life quality and the optimisation of sanitary resources [37]. In this context, miRNAs are emerging as both potential targets for new therapeutic strategies and biomarkers of RA since the association of specific miRNAs with RA has been communicated in the last years [18,19,38]. Based on the dysregulated expression of miRNAs RA in patients, several studies analysing their potential role as biomarkers of treatment response have been published. Considering this, we aim to carry out a synthesis of the literature analysing the role of miRNAs as treatment response biomarkers in the RA context. Thus, the included literature analyses the miRNAs associated with response to csDMARDs [25], csDMARDs/bDMARDs combination [27,28,30] anti-TNF α [1,13,24,26,29], and rituximab [34]. It was not possible to include biomarkers for Jakus kinase inhibitor (JAKi), a promising new drug for RA treatment [39]. To date, no study identifying specific miRNAs as predictors of JAKi response has been published.

The most common DMARDs to treat RA is MTX. Therefore, identifying a reliable biomarker of MTX response is a high priority. In this sense, Singh et al. found that MTX-responder RA patients presented lower baseline levels of miR-132, miR-146a and miR-155, defined as potential biomarkers of responsiveness to MTX treatment [25]. Sode et al., in a placebo-controlled clinical trial, defined miR-27a as a potential biomarker of the response to ADA/MTX treatment, and both miR-16 and miR-22 as biomarkers of the response to MTX treatment [27]. Castro-Villegas et al. concluded that miR-23 and miR-223 are potential biomarkers of anti-TNF α /DMARD combination therapy [28]. Furthermore, in

responder patients, the expression of miR-23 and miR-223 correlates with CRP and DAS28, respectively. Finally, Cimoska et al. [30] observed that levels of miR-5196 decreased after both treatments and correlated with DAS28. These miRNAs are related to the immune and inflammatory response. Several works have communicated their important role in processes such as the regulation of the NF- κ B signalling pathway, activation of Th17 cells, mediation of intercellular crosstalk between immune cells, and regulation of IFN- γ , IL-1, IL-4, IL-5, IL-6, IL-8, and IL-12 signalling [38,40–44]. As expected, MTX was the most common DMARD used in the cohorts of all studies. Considering this together with the existing miRNAs dysregulation in RA patients, it is plausible that MTX, alone or combined with anti-TNF α drugs, could restore this dysregulation somehow. Further and specific studies are needed to confirm this hypothesis and describe the underlying mechanisms.

Luque-Tevar et al. evaluated various anti-TNF treatments and concluded that high levels of miR-106a were associated with good response [24]. Likewise, associations have been established with each anti-TNF individually. With ADA, a good therapeutic response has been shown with the overexpression of miR-886 and low expression of miR-22 by Krintel et al. [26], whereas in the study by Cuppen et al. [13], good response was associated with high levels of miR-99a and low levels of miR-143. Regarding ETN, in the study by Liu et al. [1], a good response to treatment was associated with miR-146a, while let-7a had a poor response. The study by Cuppen et al. [13] showed that high levels of miR-23a and miR-197 were associated with a good response. Finally, Cheng et al. [29] studied the response to IFX and determined that miR-125b predicts treatment response at 24 weeks. There is a similar occurrence with RTX; high expression of miR-125b was also observed in responders [32]. Many of the miRNAs evaluated in these studies were found to be increased in response to the treatments used, and it is described that their expression affects the course of RA pathology since among their roles are the release and production of inflammatory factors (such as IL-1 β , IL-6, and TNF- α) or the regulation of signalling pathways during inflammation, B cell differentiation, and apoptosis [45–48]. After treatment, most of the miRNAs evaluated in the analysed works showed a significant alteration in their expression, both in an increasing and decreasing manner, mainly in responder individuals. Since miRNAs are key regulators of gene expression and transcription, these changes are likely to restore the protein dysregulation that RA patients present prior to treatment [34,48]. In this way, previous studies indicated a decreased expression in RA patients in miR-16-3p, miR-132, miR-146a and miR-155 [25]; another study found that that decreased miRNA-146a and 155-3p constituted an abnormal T-regulatory phenotype in these patients [49]. Therefore, this points to an effect of treatment on miRNA expression patterns, but the mechanisms underlying the possible interaction between treatment and miRNAs expression are complex and remain unknown.

In the study by Chen et al. [29], a positive correlation was shown between the basal levels of miRNA-125 with C-reactive protein and of miRNA-125 b with DAS28-ESR in RA patients. In this sense, C-reactive protein and other parameters of disease activity are widely used in the clinical follow-up of patients with inflammatory arthritis and other inflammatory diseases. However, they are nonspecific, as their levels are increased in other inflammatory conditions or remain at normal levels in patients with active disease. Thus, other studies have shown a better correlation and specificity of miRNAs with disease activity than with C-reactive protein [50,51].

Some limitations in this review are related to miRNA research. Nomenclatures used by authors are not consistent since the terms 3p and 5p refer to mature miRNAs, and several studies do not specify whether the results refer to mature miRNAs [52]. Moreover, there is a variety of samples used in the studies: whole blood, serum and plasma. This variety is important, as it is known that there are significant differences in miRNA profiles depending on the tissue [53,54]. Furthermore, it has been established that miRNA expression depends on the stage of RA development [10]. Finally, the number of works aiming to identify miRNAs as biomarkers of treatment response in RA patients found in the literature is lacking, the cohorts included in these studies are small in size, and most of them are

not high-quality clinical trials. Furthermore, the treatment guideline significantly differs between studies (e.g., different treatment duration in studies analysing the response to the same treatment). As a consequence of these limitations, it was not possible to carry out a meta-analysis as desirable.

4. Conclusions

Overall, our data support the potential role of miRNA as biomarkers of response to different treatments in RA. Moreover, the results point to DMARDs modifying the miRNAs expression, which plays a pivotal role in the modulation of the inflammatory cascade. Three miRNAs stand out due to their polyvalence: miR-146a, which predicts response to MTX and ENT; miR-125b, which predicts response to IFX and RTX; and miR-22, which predicts response to MTX and ADA. Regarding treatments, MTX is the most studied, presenting 5 miRNAs (miR-16, miR-22, miR-132, miR-146a and miR-155) described as response biomarkers. To date, a meta-analysis is not possible to be carry out due mostly to the lacking number of studies. More studies are needed to confirm our results and establish validated predictive models of response to treatment in RA patients.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms232112989/s1>. References [55–67] are cited in the supplementary materials.

Author Contributions: Conceptualisation, N.M.-V. and A.F.-N.; methodology, A.M. and J.M.L.; investigation, A.M. and N.M.-V.; resources, A.M., J.M.L., P.R.-L., S.M.-A. and N.M.-V.; data curation, A.M., J.M.L. and N.M.-V.; writing—original draft preparation, A.M. and N.M.-V.; writing—review and editing, A.F.-N.; supervision, N.M.-V.; funding acquisition, A.F.-N. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by grant RICORS (REI) RD21/0002/0037 (Instituto de Salud Carlos III), PRL was supported by a “Sara Borrell” postdoctoral contract (CD19/00216) from the Instituto de Salud Carlos III -Madrid (Spain), co-funded by the Fondo Europeo de Desarrollo Regional-FEDER, FIS Grant PI18/00824 and FIS Grant PI22/01207 (Instituto de Salud Carlos III).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Liu, Y.; Han, Y.; Qu, H.; Fang, J.; Ye, M.; Yin, W. Correlation of microRNA expression profile with clinical response to tumor necrosis factor inhibitor in treating rheumatoid arthritis patients: A prospective cohort study. *J. Clin. Lab. Anal.* **2019**, *33*, e22953. [[CrossRef](#)]
2. Chang, C.; Xu, L.; Zhang, R.; Jin, Y.; Jiang, P.; Wei, K.; Xu, L.; Shi, Y.; Zhao, J.; Xiong, M.; et al. MicroRNA-Mediated Epigenetic Regulation of Rheumatoid Arthritis Susceptibility and Pathogenesis. *Front. Immunol.* **2022**, *13*, 838884. [[CrossRef](#)]
3. Guo, S.; Zhu, Q.; Jiang, T.; Wang, R.; Shen, Y.; Zhu, X.; Wang, Y.; Bai, F.; Ding, Q.; Zhou, X.; et al. Genome-wide DNA methylation patterns in CD4+ T cells from Chinese Han patients with rheumatoid arthritis. *Mod. Rheumatol.* **2017**, *27*, 441–447. [[CrossRef](#)] [[PubMed](#)]
4. Guo, S.; Xu, L.; Chang, C.; Zhang, R.; Jin, Y.; He, D. Epigenetic Regulation Mediated by Methylation in the Pathogenesis and Precision Medicine of Rheumatoid Arthritis. *Front. Genet.* **2020**, *11*, 811. [[CrossRef](#)] [[PubMed](#)]
5. Angiolilli, C.; Kabala, P.A.; Grabiec, A.M.; Van Baarsen, I.M.; Ferguson, B.S.; García, S.; Malvar Fernandez, B.; McKinsey, T.A.; Tak, P.P.; Fossati, G.; et al. Histone deacetylase 3 regulates the inflammatory gene expression programme of rheumatoid arthritis fibroblast-like synoviocytes. *Ann. Rheum. Dis.* **2017**, *76*, 277–285. [[CrossRef](#)] [[PubMed](#)]
6. Sparks, J.A. Rheumatoid Arthritis. *Ann. Intern. Med.* **2019**, *170*, ITC1–ITC16. [[CrossRef](#)]
7. Favalli, E.G.; Raimondo, M.G.; Becciolini, A.; Crotti, C.; Biggioggero, M.; Caporali, R. The management of first-line biologic therapy failures in rheumatoid arthritis: Current practice and future perspectives. *Autoimmun. Rev.* **2017**, *16*, 1185–1195. [[CrossRef](#)]

8. Nam, J.L.; Takase-Minegishi, K.; Ramiro, S.; Chatzidionysiou, K.; Smolen, J.S.; van der Heijde, D.; Bijlsma, J.W.; Burmester, G.R.; Dougados, M.; Scholte-Voshaar, M.; et al. Efficacy of biological disease-modifying antirheumatic drugs: A systematic literature review informing the 2016 update of the EULAR recommendations for the management of rheumatoid arthritis. *Ann. Rheum. Dis.* **2017**, *76*, 1113–1136. [[CrossRef](#)]
9. Wielinska, J.; Bogunia-Kubik, K. miRNAs as potential biomarkers of treatment outcome in rheumatoid arthritis and ankylosing spondylitis. *Pharmacogenomics* **2021**, *22*, 291–301. [[CrossRef](#)]
10. Kmiołek, T.; Paradowska-Gorycka, A. miRNAs as Biomarkers and Possible Therapeutic Strategies in Rheumatoid Arthritis. *Cells* **2022**, *11*, 452. [[CrossRef](#)]
11. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* **2004**, *116*, 281–297. [[CrossRef](#)]
12. Zhang, L.; Wu, H.; Zhao, M.; Chang, C.; Lu, Q. Clinical significance of miRNAs in autoimmunity. *J. Autoimmun.* **2020**, *109*, 102438. [[CrossRef](#)] [[PubMed](#)]
13. Cuppen, B.V.; Rossato, M.; Fritsch-Stork, R.D.; Concepcion, A.N.; Schenk, Y.; Bijlsma, J.W.; Radstake, T.R.; Lafeber, F.P.; all SRU investigators. Can baseline serum microRNAs predict response to TNF-alpha inhibitors in rheumatoid arthritis? *Arthritis Res. Ther.* **2016**, *18*, 189. [[CrossRef](#)]
14. Schwarzenbach, H.; Nishida, N.; Calin, G.A.; Pantel, K. Clinical relevance of circulating cell-free microRNAs in cancer. *Nat. Rev. Clin. Oncol.* **2014**, *11*, 145–156. [[CrossRef](#)] [[PubMed](#)]
15. Navickas, R.; Gal, D.; Laucevičius, A.; Taparuskaitė, A.; Zdanytė, M.; Holvoet, P. Identifying circulating microRNAs as biomarkers of cardiovascular disease: A systematic review. *Cardiovasc Res.* **2016**, *111*, 322–337. [[CrossRef](#)]
16. Ormseth, M.J.; Wu, Q.; Zhao, S.; Allen, R.M.; Solus, J.; Sheng, Q.; Guo, Y.; Ye, F.; Ramirez-Solano, M.; Bridges, S.L.; et al. Circulating microbial small RNAs are altered in patients with rheumatoid arthritis. *Ann. Rheum. Dis.* **2020**, *79*, 1557–1564. [[CrossRef](#)]
17. Aletaha, D.; Neogi, T.; Silman, A.J.; Funovits, J.; Felson, D.T.; Bingham, C.O., 3rd; Birnbaum, N.S.; Burmester, G.R.; Bykerk, V.P.; Cohen, M.D.; et al. 2010 Rheumatoid arthritis classification criteria: An American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum.* **2010**, *62*, 2569–2581. [[CrossRef](#)]
18. Stanczyk, J.; Pedrioli, D.M.L.; Brentano, F.; Sanchez-Pernaute, O.; Kolling, C.; Gay, R.E.; Detmar, M.; Gay, S.; Kyburz, D. Altered expression of MicroRNA in synovial fibroblasts and synovial tissue in rheumatoid arthritis. *Arthritis Rheum.* **2008**, *58*, 1001–1009. [[CrossRef](#)]
19. Pauley, K.M.; Satoh, M.; Chan, A.L.; Bubb, M.R.; Reeves, W.H.; Chan, E.K. Upregulated miR-146a expression in peripheral blood mononuclear cells from rheumatoid arthritis patients. *Arthritis Res. Ther.* **2008**, *10*, R101. [[CrossRef](#)]
20. Brennan, E.; Wang, B.; McClelland, A.; Mohan, M.; Marai, M.; Beuscart, O.; Derouiche, S.; Gray, S.; Pickering, R.; Tikellis, C.; et al. Protective Effect of let-7 miRNA Family in Regulating Inflammation in Diabetes-Associated Atherosclerosis. *Diabetes* **2017**, *66*, 2266–2277. [[CrossRef](#)]
21. Fan, Y.; Ding, S.; Sun, Y.; Zhao, B.; Pan, Y.; Wan, J. MiR-377 Regulates Inflammation and Angiogenesis in Rats After Cerebral Ischemic Injury. *J. Cell Biochem.* **2018**, *119*, 327–337. [[CrossRef](#)] [[PubMed](#)]
22. Bhanji, R.A.; Eystathiou, T.; Chan, E.K.L.; Bloch, D.B.; Fritzler, M.J. Clinical and serological features of patients with autoantibodies to GW/P bodies. *Clin. Immunol.* **2007**, *125*, 247–256. [[CrossRef](#)] [[PubMed](#)]
23. Filková, M.; Aradi, B.; Senolt, L.; Ospelt, C.; Vettori, S.; Mann, H.; Filer, A.; Raza, K.; Buckley, C.D.; Snow, M.; et al. Association of circulating miR-223 and miR-16 with disease activity in patients with early rheumatoid arthritis. *Ann. Rheum. Dis.* **2014**, *73*, 1898–1904. [[CrossRef](#)]
24. Luque-Tévar, M.; Perez-Sanchez, C.; Patiño-Trives, A.M.; Barbarroja, N.; Arias de la Rosa, I.; Abalos-Aguilera, M.C.; Marin-Sanz, J.A.; Ruiz-Vilchez, D.; Ortega-Castro, R.; Font, P.; et al. Integrative Clinical, Molecular, and Computational Analysis Identify Novel Biomarkers and Differential Profiles of Anti-TNF Response in Rheumatoid Arthritis. *Front. Immunol.* **2021**, *12*, 631662. [[CrossRef](#)] [[PubMed](#)]
25. Singh, A.; Patro, P.S.; Aggarwal, A. MicroRNA-132, miR-146a, and miR-155 as potential biomarkers of methotrexate response in patients with rheumatoid arthritis. *Clin. Rheumatol.* **2019**, *38*, 877–884. [[CrossRef](#)]
26. Krintel, S.B.; Dehlendorff, C.; Hetland, M.L.; Hørslev-Petersen, K.; Andersen, K.K.; Junker, P.; Pødenphant, J.; Ellingsen, T.; Ahlquist, P.; Lindegaard, H.M.; et al. Prediction of treatment response to adalimumab: A double-blind placebo-controlled study of circulating microRNA in patients with early rheumatoid arthritis. *Pharm. J.* **2016**, *16*, 141–146. [[CrossRef](#)] [[PubMed](#)]
27. Sode, J.; Krintel, S.B.; Carlsen, A.L.; Hetland, M.L.; Johansen, J.S.; Hørslev-Petersen, K.; Stengaard-Pedersen, K.; Ellingsen, T.; Burton, M.; Junker, P.; et al. Plasma MicroRNA Profiles in Patients with Early Rheumatoid Arthritis Responding to Adalimumab plus Methotrexate vs Methotrexate Alone: A Placebo-controlled Clinical Trial. *J. Rheumatol.* **2018**, *45*, 53–61. [[CrossRef](#)]
28. Castro-Villegas, C.; Pérez-Sánchez, C.; Escudero, A.; Filipescu, I.; Verdu, M.; Ruiz-Limón, P.; Aguirre, M.A.; Jiménez-Gomez, Y.; Font, P.; Rodríguez-Ariza, A.; et al. Circulating miRNAs as potential biomarkers of therapy effectiveness in rheumatoid arthritis patients treated with anti-TNF α . *Arthritis Res. Ther.* **2015**, *17*, 49. [[CrossRef](#)]
29. Cheng, P.; Wang, J. The potential of circulating microRNA-125a and microRNA-125b as markers for inflammation and clinical response to infliximab in rheumatoid arthritis patients. *J. Clin. Lab. Anal.* **2020**, *34*, e23329. [[CrossRef](#)]
30. Ciechomska, M.; Bonek, K.; Merdas, M.; Zarecki, P.; Swierkot, J.; Gluszko, P.; Bogunia-Kubik, K.; Maslinski, W. Changes in MiRNA-5196 Expression as a Potential Biomarker of Anti-TNF- α Therapy in Rheumatoid Arthritis and Ankylosing Spondylitis Patients. *Arch. Immunol. Ther. Exp.* **2018**, *66*, 389–397. [[CrossRef](#)]

31. Anderson, J.K.; Zimmerman, L.; Caplan, L.; Michaud, K. Measures of rheumatoid arthritis disease activity: Patient (PtGA) and Provider (PrGA) Global Assessment of Disease Activity, Disease Activity Score (DAS) and Disease Activity Score with 28-Joint Counts (DAS28), Simplified Disease Activity Index (SDAI), *C. Arthritis Care Res.* **2011**, *63* (Suppl. S1), S14–S36. [[CrossRef](#)] [[PubMed](#)]
32. Wells, G.; Becker, J.-C.; Teng, J.; Dougados, M.; Schiff, M.; Smolen, J.; Aletaha, D.; van Riel, P.L. Validation of the 28-joint Disease Activity Score (DAS28) and European League Against Rheumatism response criteria based on C-reactive protein against disease progression in patients with rheumatoid arthritis, and comparison with the DAS28 based on eryth. *Ann. Rheum. Dis.* **2009**, *68*, 954–960. [[CrossRef](#)] [[PubMed](#)]
33. Miller, J. The Scottish Intercollegiate Guidelines Network (SIGN). *Br. J. Diabetes Vasc. Dis.* **2002**, *2*, 47–49. [[CrossRef](#)]
34. Duroux-Richard, I.; Pers, Y.-M.; Fabre, S.; Ammari, M.; Baeten, D.; Cartron, G.; Touitou, I.; Jorgensen, C.; Apparailly, F. Circulating miRNA-125b is a potential biomarker predicting response to rituximab in rheumatoid arthritis. *Mediat. Inflamm.* **2014**, *2014*, 342524. [[CrossRef](#)]
35. Hørslev-Petersen, K.; Hetland, M.L.; Junker, P.; Pødenphant, J.; Ellingsen, T.; Ahlquist, P.; Lindegaard, H.; Linauskas, A.; Schlemmer, A.; Dam, M.Y.; et al. Adalimumab added to a treat-to-target strategy with methotrexate and intra-articular triamcinolone in early rheumatoid arthritis increased remission rates, function and quality of life. The OPERA Study: An investigator-initiated, randomised, double-blind. *Ann. Rheum. Dis.* **2014**, *73*, 654–661. [[CrossRef](#)]
36. de Hair, M.J.H.; Jacobs, J.W.G.; Schoneveld, J.L.M.; van Laar, J.M. Difficult-to-treat rheumatoid arthritis: An area of unmet clinical need. *Rheumatology* **2018**, *57*, 1135–1144.
37. Mena-Vázquez, N.; Ruiz-Limón, P.; Moreno-Indias, I.; Manrique-Arija, S.; Tinahones, F.J.; Fernández-Nebro, A. Expansion of Rare and Harmful Lineages is Associated with Established Rheumatoid Arthritis. *J. Clin. Med.* **2020**, *9*, 1044. [[CrossRef](#)]
38. Niimoto, T.; Nakasa, T.; Ishikawa, M.; Okuhara, A.; Izumi, B.; Deie, M.; Suzuki, O.; Adachi, N.; Ochi, M. MicroRNA-146a expresses in interleukin-17 producing T cells in rheumatoid arthritis patients. *BMC Musculoskelet Disord.* **2010**, *11*, 209. [[CrossRef](#)]
39. Venetsanopoulou, A.I.; Voulgari, P.V.; Drosos, A.A. Janus kinase versus TNF inhibitors: Where we stand today in rheumatoid arthritis. *Expert Rev. Clin. Immunol.* **2022**, *18*, 485–493. [[CrossRef](#)]
40. Donate, P.B.; Alves de Lima, K.; Peres, R.S.; Almeida, F.; Fukada, S.Y.; Silva, T.A.; Nascimento, D.C.; Cecilio, N.T.; Talbot, J.; Oliveira, R.D.; et al. Cigarette smoke induces miR-132 in Th17 cells that enhance osteoclastogenesis in inflammatory arthritis. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2017120118. [[CrossRef](#)]
41. Calin, G.A.; Sevignani, C.; Dumitru, C.D.; Hyslop, T.; Noch, E.; Yendamuri, S.; Shimizu, M.; Rattan, S.; Bullrich, F.; Negrini, M.; et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 2999–3004. [[CrossRef](#)]
42. Chen, L.; Lu, Q.; Chen, J.; Feng, R.; Yang, C. Upregulating miR-27a-3p inhibits cell proliferation and inflammation of rheumatoid arthritis synovial fibroblasts through targeting toll-like receptor 5. *Exp. Ther. Med.* **2021**, *22*, 1227. [[CrossRef](#)] [[PubMed](#)]
43. Tian, T.; Zhou, Y.; Feng, X.; Ye, S.; Wang, H.; Wu, W.; Tan, W.; Yu, C.; Hu, J.; Zheng, R.; et al. MicroRNA-16 is putatively involved in the NF- κ B pathway regulation in ulcerative colitis through adenosine A2a receptor (A2aAR) mRNA targeting. *Sci. Rep.* **2016**, *6*, 30824. [[CrossRef](#)] [[PubMed](#)]
44. Salehi, E.; Eftekhari, R.; Oraei, M.; Gharib, A.; Bidad, K. MicroRNAs in rheumatoid arthritis. *Clin. Rheumatol.* **2015**, *34*, 615–628. [[CrossRef](#)] [[PubMed](#)]
45. Hong, B.K.; You, S.; Yoo, S.A.; Park, D.; Hwang, D.; Cho, C.S.; Kim, W.U. MicroRNA-143 and -145 modulate the phenotype of synovial fibroblasts in rheumatoid arthritis. *Exp. Mol. Med.* **2017**, *49*, e363. [[CrossRef](#)]
46. Lai, N.-S.; Yu, H.-C.; Yu, C.-L.; Koo, M.; Huang, H.-B.; Lu, M.-C. Anti-citrullinated protein antibodies suppress let-7a expression in monocytes from patients with rheumatoid arthritis and facilitate the inflammatory responses in rheumatoid arthritis. *Immunobiology* **2015**, *220*, 1351–1358. [[CrossRef](#)]
47. Bao, X.; Ma, L.; He, C. MicroRNA-23a-5p regulates cell proliferation, migration and inflammation of TNF- α -stimulated human fibroblast-like MH7A synoviocytes by targeting TLR4 in rheumatoid arthritis. *Exp. Ther. Med.* **2021**, *21*, 479. [[CrossRef](#)]
48. Moran-Moguel, M.C.; Petarra-Del Rio, S.; Mayorquin-Galvan, E.E.; Zavala-Cerna, M.G. Rheumatoid Arthritis and miRNAs: A Critical Review through a Functional View. *J. Immunol. Res.* **2018**, *2018*, 2474529. [[CrossRef](#)]
49. Zhou, Q.; Haupt, S.; Kreuzer, J.T.; Hammitzsch, A.; Proft, F.; Neumann, C.; Leipe, J.; Witt, M.; Schulze-Koops, H.; Skapenko, A. Decreased expression of miR-146a and miR-155 contributes to an abnormal Treg phenotype in patients with rheumatoid arthritis. *Ann. Rheum. Dis.* **2015**, *74*, 1265–1274. [[CrossRef](#)]
50. Chen, P.; Li, Y.; Li, L.; Yu, Q.; Chao, K.; Zhou, G.; Qiu, Y.; Feng, R.; Huang, S.; He, Y.; et al. Circulating microRNA146b-5p is superior to C-reactive protein as a novel biomarker for monitoring inflammatory bowel disease. *Aliment Pharmacol. Ther.* **2019**, *49*, 733–743. [[CrossRef](#)]
51. Schönaen, K.; Le, N.; von Arnim, U.; Schulz, C.; Malfertheiner, P.; Link, A. Circulating and Fecal microRNAs as Biomarkers for Inflammatory Bowel Diseases. *Inflamm. Bowel Dis.* **2018**, *24*, 1547–1557. [[CrossRef](#)] [[PubMed](#)]
52. Akhtar, M.M.; Micolucci, L.; Islam, M.S.; Olivieri, F.; Procopio, A.D. Bioinformatic tools for microRNA dissection. *Nucleic Acids Res.* **2016**, *44*, 24–44. [[CrossRef](#)] [[PubMed](#)]
53. Duroux-Richard, I.; Jorgensen, C.; Apparailly, F. What do microRNAs mean for rheumatoid arthritis? *Arthritis Rheum.* **2012**, *64*, 11–20. [[CrossRef](#)]

54. Cheng, H.H.; Yi, H.S.; Kim, Y.; Kroh, E.M.; Chien, J.W.; Eaton, K.D.; Goodman, M.T.; Tait, J.F.; Tewari, M.; Pritchard, C.C.; et al. Plasma processing conditions substantially influence circulating microRNA biomarker levels. *PLoS ONE* **2013**, *8*, e64795. [[CrossRef](#)]
55. Ormseth, M.J.; Solus, J.F.; Sheng, Q.; Ye, F.; Wu, Q.; Guo, Y.; Oeser, A.M.; Allen, R.M.; Vickers, K.C.; Stein, C.M. Development and validation of a microRNA panel to differentiate between patients with rheumatoid arthritis, systemic lupus erythematosus, and control subjects. *Arthritis Rheumatol.* **2018**, *70*, 2070.
56. De La Cruz-Castillejos, J.C.; Barbosa-Cobos, R.E.; Becerril-Mendoza, L.T.; Lugo-Zamudio, G.E.; Ramírez-Bello, J.; Matias-Carmona, M.; Alemán-Avila, I. Evaluation of variants in miR-146a, miR-196a-2 and miR-499 and their association with susceptibility for rheumatoid arthritis and its extra-articular manifestations. *Ann. Rheum. Dis.* **2017**, *76*, 1133.
57. Heinicke, F.; Zhong, X.; Flåm, S.T.; Breidenbach, J.; Leithaug, M.; Mæhlen, M.T.; Lillegraven, S.; Aga, A.B.; Norli, E.S.; Mjaavatten, M.D. MicroRNA Expression Differences in Blood-Derived CD19+ B Cells of Methotrexate Treated Rheumatoid Arthritis Patients. *Front. Immunol.* **2021**, *12*, 663736. [[CrossRef](#)]
58. Pallio, G.; Mannino, F.; Irrera, N.; Eid, A.H.; Squadrito, F.; Bitto, A. Polymorphisms involved in response to biological agents used in rheumatoid arthritis. *Biomolecules* **2020**, *10*, 1203. [[CrossRef](#)]
59. Abdelaziz, M.M.; Gamal, R.M.; Khalifa, F.; Mosad, E.; Sadek, R.; Abd El Razik, D.I.; Kamal, D. MicroRNA146a gene polymorphism in patients with rheumatoid arthritis and the relevant value with disease activity and extra-articular manifestations. *Egypt Rheumatol.* **2022**, *44*, 97–101. [[CrossRef](#)]
60. Kádár, G.; Czibula, Á.; Szalay, B.; Nagy, K.; Pusztai, A.; Balog, A.; Monostori, E.; Vásárhelyi, B.; Szekanez, Z.; Kovács, L. Predictors of disease course after the discontinuation of biologic therapy in rheumatoid arthritis patients with long-term remission. *Ann. Rheum. Dis.* **2016**, *75*, 1007. [[CrossRef](#)]
61. Lim, M.-K.; Song, J.; Kim, S.A.; Yoo, J. MicroRNA-1915-3p in serum exosome is associated with disease activity of rheumatoid arthritis in Korea. *Ann. Rheum. Dis.* **2018**, *77*, 266.
62. Schordan, E.; Bilger, G.; Coq, M.; Danilin, S.; Mehdi, M.; Schumacher, M.; Firat, H. MiRNA profiling using HTG-EDGESEQ platform predicts response to anti-TNF alpha therapy in rheumatoid arthritis. *Ann. Rheum. Dis.* **2016**, *75*, 228. [[CrossRef](#)]
63. Jekic, B.; Vejnovic, D.; Milic, V.; Maksimovic, N.; Damjanovic, T.; Bunjevacki, V.; Novakovic, I.; Lukovic, L.; Damjanov, N.; Krajinovic, M. Association of 63/91 length polymorphism in the DHFR gene major promoter with toxicity of methotrexate in patients with rheumatoid arthritis. *Pharmacogenomics* **2016**, *17*, 1687–1691. [[CrossRef](#)] [[PubMed](#)]
64. Chen, Z.-Z.; Zhang, X.-D.; Chen, Y.; Wu, Y.-B. The role of circulating miR-146a in patients with rheumatoid arthritis treated by *Tripterygium wilfordii* Hook F. *Medicine* **2017**, *96*, e6775. [[CrossRef](#)]
65. Liu, Y.; Jeon, S.-M.; Caterina, M.J.; Qu, L. miR-544-3p mediates arthritis pain through regulation of FcγRI. *Pain* **2021**, *163*, 1497–1510. [[CrossRef](#)]
66. Lopez-Pedrerera, C.; Luque-Tevar, M.; Pérez-Sánchez, C.; Font, P.; Patiño-Trives, A.; Arias de la Rosa, I.; Abalos-Aguilera, M.; Torres-Granados, C.; Romero-Gomez, M.; Ruiz-Vilchez, D.; et al. Circulating Biomolecules as Potential Biomarkers of Early and Established response to TNFi Therapy in Rheumatoid Arthritis Patients. *Arthritis Rheumatol.* **2020**, *72* (Suppl. S10), 4015–4017.
67. Ciesla, M.; Kolarz, B.; Dryglewska, M.; Majdan, M. FCER1G gene methylation and mir-106/miR-17 as a new potential epigenetic markers in rheumatoid arthritis. *Ann. Rheum. Dis.* **2020**, *79* (Suppl. S1), 1347–1348. [[CrossRef](#)]