

Original article

Contents lists available at ScienceDirect

European Journal of Radiology Open



journal homepage: www.elsevier.com/locate/ejro

Radiogenomics study to predict the nuclear grade of renal clear cell carcinoma $\stackrel{\star}{\sim}$

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HIGHLIGHTS

• A radiomics model and genomics model was developed for predicting the histopathologic nuclear grade with localized clear cell renal cell carcinoma (ccRCC).

• The CT radiomics models exhibited higher predictive performance than mRNA models.

• The association between radiomics features and mRNA related to nuclear grade is not universal.

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Keywords: Kidney Radiomics Neoplasms-Primary Computer Applications Radiogenomics Pathological nuclear grade

ABSTRACT

Purpose: To develop models based on radiomics and genomics for predicting the histopathologic nuclear grade with localized clear cell renal cell carcinoma (ccRCC) and to assess whether macro-radiomics models can predict the microscopic pathological changes. *Method:* In this multi-institutional retrospective study, a computerized tomography (CT) radiomic model for nuclear grade prediction was developed. Utilizing a genomics analysis cohort, nuclear grade-associated gene modules were identified, and a gene model was constructed based on top 30 hub mRNA to predict the nuclear grade. Using a radiogenomic development cohort, biological pathways were enriched by hub genes and a radiogenomic map was created. *Results:* The four-features-based SVM model predicted nuclear grade with an area under the curve (AUC) score of 0.94 in validation sets, while a five-gene-based model predicted nuclear grade with an AUC of 0.73 in the ge-

0.94 in validation sets, while a five-gene-based model predicted nuclear grade with an AUC of 0.73 in the genomics analysis cohort. A total of five gene modules were identified to be associated with the nuclear grade. Radiomic features were only associated with 271 out of 603 genes in five gene modules and eight top 30 hub genes. Differences existed in the enrichment pathway between associated and un-associated with radiomic features, which were associated with two genes of five-gene signatures in the mRNA model.

Conclusion: The CT radiomics models exhibited higher predictive performance than mRNA models. The association between radiomic features and mRNA related to nuclear grade is not universal.

1. Introduction

Radiogenomics focus on discovering correlations between radiomic features and genomics [1]. Based on the principle that medical images

are the outcome of complex interactions at the genetic and molecular levels [2], radiogenomic studies have revealed associations between imaging features and genetic information of tumours in glioblastoma, gastric, ovarian and other cancers [3–5]. Nuclear grade of ccRCC is

https://doi.org/10.1016/j.ejro.2023.100476

Received 7 December 2022; Received in revised form 11 January 2023; Accepted 16 January 2023

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Abbreviations: ccRCC, Clear cell renal cell carcinoma; WHO/ISUP, World Health Organization and International Society of Urological Pathology; FDR, False discovery rate; GLRLM, Gray level run length matrix; GLSZM, Gray level size matrix; NGTDM, Neighborhood gray tone difference matrix; KEGG, KOBAS-Kyoto Encyclopedia of Genes and Genomes; TCGA, The cancer genome atlas; TCIA, The cancer imaging archive; WGCNA, Weighted gene co-expression network; PPI, Protein-Protein Interaction Networks.

^{*} Grant number: GSWSKY2020-15 and grant number. LCCX2021006.

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Fig. 1. Overview of the study cohort and design.

 Table 1

 Demographic and clinical information of the study cohorts.

	train	test	p value
	N = 103	N = 57	
Age	61.000	60.000	0.056
	[51.000;69.000]	[50.000;75.500]	
Sex			0.052
female	32 (31.068%)	14 (24.561%)	
male	71 (68.932%)	43 (75.439%)	
Pathological-nuclear- grading			0.001
low	59 (57.282%)	31 (54.386%)	
high	44 (42.718%)	26 (45.614%)	

considered to affect prognosis and surgical benefits, and its preoperative evaluation can provide effective guidelines for clinical treatments [6]. Therefore, predicting ccRCC nuclear grade is important in clinical practice. Studies evaluating the ccRCC nuclear grade prediction revealed that performance of a CT-radiomics-based model was higher than that of a The Cancer Genomics Atlas (TCGA) transcriptomics-based model, as indicated by the AUCs in the validation set which were 0.90 and 0.82, respectively [7,8]. Radiogenomic studies for prognosis and prediction of ccRCC suggested the existence of correlations between radiomic features and prognosis-related pathways such as cell cycle and microvascular infiltration [9–11]. The answer to predicting nuclear grade of ccRCC may lie in deciphering the association between nuclear grade-related radiomic features and genomics. However, the underlying mechanisms governing this association have yet to be elucidated. Under this light, the present study aimed to a) predict the nuclear grade of ccRCC by investigating the correlation between CT radiomic features and mRNA data, b) decipher the underlying genomic mechanisms of the tested radiomic models, and c) provide insights in the interpretation of relevant radiomic features.

2. Materials and methods

2.1. Study design

The design of the study included three main stages: First, we extracted the suitable CT radiomic features for building a model. Second, we identified gene modules related to nuclear grade and hub genes for building the genomics model. Finally, we mapped the radiomic features with gene modules, hub genes and gene signatures (Fig. 1).

2.2. Patient information sources

In this multi-institutional retrospective study, a total of 645 images of patients were retrieved from the kits21 public imaging dataset, The Cancer Imaging Archive (TCIA), The Cancer Genomics Atlas (TCGA), and the radiology department of local hospital from January 2015 to December 2021. The acquisition of images and genomic data from public database sources such as TCIA and TCGA and, the approval of this study by the ethics committee of the local institution, waived the requirement to obtain informed patient consent. The study included all patients with pathologically confirmed ccRCC that underwent a preoperative enhanced CT scan, except those with multiple regional tumours. Patients were divided into four subgroups. Subgroup 1 comprised of 103 patients with preoperative CT images (corticomedullary) collected from the kits21 dataset and used to develop the radiomics model. Subgroup 2 included 23 patients with images from local hospitals, used to construct a validation set with subgroup 4 for radiomics analysis. Subgroup 3 included 519 patients with mRNA sequencing data obtained from TCGA, used as a genomics analysis cohort for gene module identification and development of gene models. Subgroup 4 has served as a subset of subgroup 3, consisting of 34 patients with both preoperative CT corticomedullary images and mRNA sequencing data. Subgroup 4 was used in combination with subgroup 2 as a radiomics validation set and for analysing the potential association between radiomic features and mRNA expression.



Fig. 2. Selection of radiomic features and the development of radiomics model. a: Values of four selected radiomic features of high and low nuclear grades, b: ROC curve of six classified algorithms in the training set, c: The ROC curve of six classified algorithms in the validation set.

2.3. Evaluation of pathological nuclear grade

Nuclear grade information from the local hospital originated from the pathology department of local hospital and evaluated by two independent pathologists according to the WHO/ISUP grade system, while all kidney biopsy specimens were re-examined independently by a pathologist with 7 years of genitourinary pathology experience. Grades between 1 and 2 of the WHO grade system were considered as low and grades between 3 and 4 were considered as high.

2.4. CT images

All CT examinations at the local hospital were conducted using spiral scanners to acquire images of corticomedullary phase on the CT system. All patients were given 400 mL of water orally 25 min before the examination and injected with a dose of 1.5 mL/kg of a non-ionic contrast agent. After 25–30 s, diaphragm to kidney images of the corticomedullary phase were obtained. The image parameters were as follows: (1) manufacturer: Siemens; (2) gantry rotation time 0.5 s; (3) tube voltage: 120 kV; (4) detector collimation: 64×0.6 mm. all images were reconstructed as a 5 mm thick reconstructed section.

2.5. Radiomic analysis

2.5.1. Tumour segmentation

Image voxels were resampled to $1 \times 1 \text{ mm}^2$, and 25 Gray levels were used to discretize their intensities. Pre-testing of 60 randomly selected patients showed no significant difference in predictive performance between ROI (region of interest) and VOI (volume of interest) (Supplement Fig. 1). Hence, segmentation was performed in the largest

diameter cross-section along the tumour border by a radiologist with 8 years of experience using the ITK-snap software. Additionally, 30 patients were randomly selected for repeat segmentation by another senior radiologist to select for robust radiomic features.

2.5.2. Feature selection and radiomic model construction

Features of the segmented 2D tumour were extracted from the original images of cases using python package named Pyradiomics, following an intraclass correlation coefficient (ICC) test, the radiomic features with ICC less than 0.80 were excluded. The univariate Wilcoxon rank sum test was used to screen for radiomic features significantly associated with nuclear grade. Only radiomic features with ICC equal or more than 0.8 and univariate Wilcox-rank sum test p values less than 0.01 qualified for further analysis. Consequently, the maximum Relevance and Minimum Redundancy (mRMR) and Boruta's algorithms were used to select the features with a mean absolute correlation greater than 0.9. The selected radiomic features were used to construct radiomic models by using six classified algorithmic methods, which included Logistic regression (LR), Naive Bayes (NB), Decision Tree (DT), Random Forest (RF), Support vector machine (SVM), and XGBoost (XGB). Prediction performance was estimated by the AUC of the receiver operator characteristic curve (ROC). The optimal cut-off value was obtained from the Jorden index of the ROC curve in the training set. The accuracy (ACC), sensitivity (Se), specificity (Sp), negative prediction rate (NPV), and positive prediction rate (PPV) of the model were also calculated. The DeLong test was used for comparison between different AUCs.

	I.													
method	feature_numbers	train_auc	train_acc	train_sen	train_spe	train_ppv	train_npv	test_auc	test_acc	test_sen	test_spe	test_ppv	test_npv	cutoff
LR	4	0.759	0.733	0.714	0.746	0.667	0.786	0.711	0.707	0.704	0.710	0.679	0.733	0.36104249901878
		(0.661 - 0.858)	(0.635 - 0.816)	(0.499 - 0.833)	(0.424 - 0.865)	(0.583 - 0.700)	(0.676 - 0.810)	(0.572 - 0.850)	(0.573 - 0.819)	(0.370 - 0.889)	(0.290 - 0.903)	(0.526 - 0.727)	(0.529 - 0.778)	
NB	4	0.748	0.752	0.714	0.780	0.698	0.793	0.710	0.672	0.519	0.806	0.700	0.658	0.10187504141657
		(0.645 - 0.851)	(0.657 - 0.833)	(0.404 - 0.857)	(0.423 - 0.864)	(0.566 - 0.735)	(0.675 - 0.810)	(0.573 - 0.846)	(0.537 - 0.790)	(0.222 - 0.816)	(0.612 - 0.936)	(0.500 - 0.786)	(0.593 - 0.691)	
DT	4	0.723	0.733	0.667	0.780	0.683	0.767	0.700	0.707	0.593	0.806	0.727	0.694	0.47941468253968
		(0.633 - 0.813)	(0.635 - 0.816)	(0.371 - 0.799)	(0.508 - 0.872)	(0.545 - 0.721)	(0.682 - 0.786)	(0.582 - 0.817)	(0.573 - 0.819)	(0.333 - 0.751)	(0.506 - 0.923)	(0.600 - 0.772)	(0.588 - 0.722)	
RF	4	0.814	0.762	0.714	0.797	0.714	0.797	0.815	0.759	0.667	0.839	0.783	0.743	0.43
		(0.728 - 0.899)	(0.667 - 0.841)	(0.452 - 0.858)	(0.542 - 0.898)	(0.613 - 0.750)	(0.727 - 0.815)	(0.706 - 0.925)	(0.628 - 0.861)	(0.283 - 0.889)	(0.548 - 0.968)	(0.605 - 0.828)	(0.654 - 0.769)	
SVM	4	0.954	0.941	0.952	0.932	0.909	0.965	0.938	0.845	0.963	0.742	0.765	0.958	0.37022294632049
		(0.902 - 1.000)	(0.875 - 0.978)	(0.714 - 1.000)	(0.153 - 0.983)	(0.882 - 0.913)	(0.818 - 0.967)	(0.871 - 1.000)	(0.726 - 0.927)	(0.852 - 1.000)	(0.097 - 0.935)	(0.742 - 0.771)	(0.750 - 0.967)	
XGB	4	0.783	0.772	0.595	0.898	0.806	0.757	0.769	0.707	0.444	0.935	0.857	0.659	0.60353703796863
		(0.693 - 0.874)	(0.678 - 0.850)	(0.333 - 0.715)	(0.729 - 0.966)	(0.700 - 0.833)	(0.717 - 0.770)	(0.647 - 0.890)	(0.573 - 0.819)	(0.222 - 0.741)	(0.710 - 1.000)	(0.750-0.909)	(0.595 - 0.674)	

Table

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Table 3a

DeLong test among radiomics models of training set

name	p value
LR-NB	0.519020801
LR-DT	0.228144417
LR-RF	0.111431884
LR-SVM	0.0000743
LR-XGB	0.272050649
NB-DT	0.393096654
NB-RF	0.061762627
NB-SVM	0.0000552
NB-XGB	0.116238452
DT-RF	0.00091159
DT-SVM	0.00000251
DT-XGB	0.013907988
RF-SVM	0.000426971
RF-XGB	0.214224916
SVM-XGB	0.0000602

Table 3b

DeLong test among radiomics models of validation set.

name	p value
LR-NB	0.97683832
LR-DT	0.810259242
LR-RF	0.044645072
LR-SVM	0.000698819
LR-XGB	0.13932625
NB-DT	0.862149834
NB-RF	0.046600886
NB-SVM	0.00037982
NB-XGB	0.194359811
DT-RF	0.003248959
DT-SVM	0.0000378
DT-XGB	0.148733046
RF-SVM	0.016589642
RF-XGB	0.299101041
SVM-XGB	0.001676355

2.6. Nuclear grade-related gene module identification

2.6.1. Differential gene expression analysis for nuclear grade

The mRNA data of the genomics analysis cohort were processed using the "limma" R package to identify differentially expressed genes, with a differential expression threshold calculated according to the following formula: $mean(abs(logFc) + 2 \times sd(ABS(logFc)))$. Significance of differential genes was evaluated using a t-test, and differential expressed genes were considered the ones having false discovery rates (FDRs) less than 0.05 and fold changes (FCs) greater than the threshold, the differentially expressed genes (DEGs) was subsequently used in the WGCNA algorithm for further analysis.

2.6.2. Construction of weighted gene co-expression networks

Based on the genomics analysis cohort (subgroup 3), a scale-free network of DEGs was constructed using the WGCNA algorithm to identify gene modules related to nuclear grades. A soft threshold was calculated based on the corresponding scale independence (\mathbb{R}^2) and average connectivity. An adjacency matrix was constructed based on the soft threshold and the Pearson correlation coefficient between pairs of genes, which was then transformed into a topological overlap matrix (TOM) and the dissimilarity (1-TOM) was calculated. According to the similarity matrix, similar genes were grouped into one gene coexpression modules by hierarchical clustering, setting the number of gene within a module to be no less than 30 and the height threshold for gene module merging to 0.25. The association between gene modules and nuclear grade was visualized as a heat map, and the respective significance was expressed as gene significance (GS). Associations with a



Fig. 3. Two examples of radiomic features nuclear grade of ccRCC. a: corticomedullary images of high nuclear grade of ccRCC, b: corticomedullary images of low nuclear grade of ccRCC, c: histopathological images of high nuclear grade of ccRCC, d: histopathological images of low nuclear grade of ccRCC.



Fig. 4. Results of the WGCNA analysis of genomics analysis cohort. a: Genes with the same characteristics were grouped into a gene module, and each module was assigned a colour, b: The five-gene module that is shown to be related with the nuclear grade.

false discovery rate (FDR) corrected p value less than 0.05 were considered significant.

2.7. Construction of radiogenomic maps

Radiogenomic association maps were created based on the radiogenomic development and genomic analysis cohorts including the gene modules related to nuclear grade, using the Spearman correlation with a significance p value of 0.05.

2.7.1. Screening of nuclear grade-related hub genes

Based on genomics analysis cohort, Protein-Protein Interaction Networks (PPI) were constructed by submitting the WGCNA results into the STRING database and setting the minimum interaction scores to 0.4. Then the results were imported into the Cytoscape software (v3.8.2), and the Degree Centrality of genes was calculated using the Degree algorithm. Top 30 genes were regarded as hub genes.

2.8. Identification of biological pathways for hub genes

The biological pathways of hub genes were identified using the R package "Cluster Profiler" for KOBAS Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis, based on the radiogenomic development cohort. A false positive rate (FDR) *p*-value less than 0.05 was considered as significant.

2.9. Multigene model for predicting nuclear grade

Based on the genomics cohort analysis, the genetic sample was randomly divided into training and validation sets in a 7:3 ratio. The



Fig. 5. Analysis of the radiogenomic development cohort. a: The radiogenomic map constructed by four radiomic features and related genes, b: Top 30 hub genes by the Degree algorithm, c: The radiogenomic map constructed by four radiomic features and 30 hub genes.

training and validation sets were balanced using the Synthetic Minority Over-Sampling Technique (SMOTE), and the dataset was normalized using the minmax method. The Pearson correlation coefficient (PCC) was calculated for each gene pair, and any one of the genes in pairs with PCC> 0.99 was randomly removed. Gene selection was performed using recursive feature elimination (RFE), before the construction of RF, SVM, DT, and LR models. All the above processes were performed by running the FAE software (V0.5.3) on python (3.7.6) (https://github.com/ salan668/FAE).

3. Statistical analysis of clinical data

Data was analysed using R 4.1.2. Nuclear grade, sex and age were analysed using Chi-square or Wilcoxon test. Statistical differences were significant at a *p* value less than 0.05.

4. Results

4.1. Clinical information of patients

The general clinical information of the study cohort is shown in Table 1. With the exception of the nuclear grade of ccRCC, no significant differences were found between age and gender.

4.2. Radiomic feature selection and model construction

A total of 107 radiomic features were extracted, including 14 morphological feature, 18 first-order features and 75 texture features.



Fig. 6. The KEGG results for hub genes distinguished by association. a: The pathway underlying radiomic feature-correlated genes, b: The pathway underlying radiomics feature non-correlated genes.

The result of ICC test revealed that 102 radiomic features was identified. After univariate Wilcox-rank sum test analysis, 11 radiomic features was screened. Finally, the four radiomic features were selected by mRMR and Boruta's algorithms (Fig. 2a). The values of neighbourhood Graytone difference matrix (NGTDM)-strength were reduced, while values of the other three radiomic features increased in the higher grades. The prediction performance of the six classifiers is shown in Fig. 2b and c. The SVM classification method achieved AUC and ACC values of 0.94 and 0.85 in the validation set, reaching the best performance among the six methods in both the training and validation sets with a cut-off value of 0.37 (Table 2). The results of the DeLong test between the SVM classifier and other classifiers also demonstrate that the prediction effectiveness of SVM was higher than the other five classes of classifiers (Table 3a and b). We showed corticomedullary images and histopathological images in a patient with high-grade ccRCC and a patient with low-grade ccRCC in the cohort (Fig. 3a-d).

4.3. Radiogenomic analysis and hub-related pathway identification

4.3.1. Differential genes associated with the nuclear grade

Based on genomics analysis cohort, mRNAs differential analysis was performed using a threshold value of 0.579. The differential expression analysis revealed 887 gene, including 451 down-regulated and 436 upregulated genes.

4.3.2. Identification of gene modules associated with nuclear grade

Based on genomics analysis cohort, we selected $\beta = 10$ as a suitable soft threshold to construct the scale-free network, and 887 differential genes were divided into five gene modules (Fig. 4a). Five gene modules (brown, yellow, turquoise, blue, and green modules) containing 603 genes were associated with nuclear grade (p < 0.05) (Fig. 4b), which were associated with radiomic features in the radiogenomic development cohort.

4.3.3. Construction of radiogenomic maps, hub gene selection and pathway identification

The radiogenomic development cohort revealed that 271 genes were associated with radiomic features while the remaining 332 genes were not associated with radiomic features, as shown in the respective radiogenomic map (Fig. 5a). The results of the hub genes are shown in Fig. 5b. The radiogenomic map revealed that 8 hub genes were associated with radiomic features, while 22 were not associated with the radiomic features (Fig. 5c). The radiomic features-related 8 hub genes were enriched in 10 KEGG pathways, including PI3K-Akt pathway, HIF-

1 signalling pathway, and MAPK signalling pathway (Fig. 6a). The remaining 22 hub genes were enriched in 19 KEGG pathways, including IL-17, Th17 cell differentiation and PD-L1 immune checkpoint signalling pathway, and other tumour-related signalling pathways (Fig. 6b).

4.4. Gene signature for nuclear grade prediction

Five hub genes were selected and used to develop a LR model with the highest AUC value of 0.72 in the validation set (Fig. 7a). The prediction performance and result of DeLong test among four algorithms is shown in Table 4a and b, Table 5a and b, respectively. The results of the analysis between radiomic features and gene signatures showed that F2, TEK was associated with the four radiomic features (Fig. 7b).

5. Discussion

In conclusion, a radiomics model used to predict the ccRCC nuclear grade was developed, and radiomic features were correlated with genes in order to classify the nuclear grade and explore the driving pathway of the radiomic features. In comparison with this study, most previous radiogenomic studies identified gene modules associated with radiomic features, but not gene modules that were not associated with radiomic features but which may also predict the clinical outcome. In order to avoid selection bias of gene modules, we identified gene modules based on clinical outcomes, including gene modules un-associated with the radiomic features. Furthermore, previous studies have not evaluated the predictive performance of gene modules, whereas in our study we constructed a hub gene model for predicting nuclear grade.

CT radiomic features able to identify the nuclear grade of ccRCC in the validation cohort with an AUC of 0.81 have been previously reported [12]. The respective CT based SVM model performed well in the validation cohort (AUC=0.82) [13]. These results are consistent with our study, where the constructed SVM model had a comparable performance in the validation set (AUC=0.94). In addition, we constructed a hub mRNA model for predicting the nuclear grade. Biological processes always involve interactions among genes expressed by interaction networks which are controlled by driving hub genes [14,15]. Exploring the predictive performance of hub genes can help understand the association between radiomic features and genes. Our hub mRNA genes achieved a comparable AUC (0.73) in the validation set, which is lower than that of similar previous studies (AUC=0.82) [8]. The different predictive performance between the two models suggests that single-dimension genes such as mRNA genes may not reflect the entire variation observed in nuclear grades. mRNA is also regulated by other factors such



Fig. 7. Prediction performance of the multigene model. a: The contribution of gene signatures in the final multigene model, b: ROC curve of the multigene model in training and validation sets, c: The radiogenomic map constructed by combining genes of the multigene model and radiomic features.

Table 4a

Prediction performance of the genomics models constructed by four classifiers of training set.

feature-selection of training set	Number	PosNum	NegNum	AUC	95% CIs	Std	Acc	Youden Index	Sen	Spe	PPV	NPV
MinMax_PCC_RFE_5_LR	364	197	167	0.7363	[0.6882-0.7895]	0.0262	0.7033	0.4541	0.7462	0.6527	0.7171	0.6855
MinMax_PCC_RFE_9_SVM MinMax_PCC_RFE_7_RF	364 364	197 197	167	0.7251	[1.0000–1.0000]	0.0265 0	0.0808	0.4691	0.7716	0.5868 1	0.0878 1	0.0855
MinMax_PCC_RFE_8_DT	364	197	167	1	[1.0000 - 1.0000]	0	1	1	1	1	1	1

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Prediction performance of the genomics models constructed by four classifiers of validation set.

feature-selection of training set	Number	PosNum	NegNum	AUC	95% CIs	Std	Acc	Youden Index	Sen	Spe	PPV	NPV
MinMax_PCC_RFE_5_LR MinMax_PCC_RFE_9_SVM MinMax_PCC_RFE_7_RF	155 155 155	84 84 84	71 71 71	0.7341 0.7202 0.7076	[0.6496–0.8155] [0.6353–0.8042] [0.6278–0.7898]	0.0419 0.0417 0.0404	0.7161 0.6774 0.6581	0.4904 0.562 0.550	0.6786 0.5595 0.5595	0.7606 0.8169 0.7746	0.7703 0.7833 0.746	0.6667 0.6105 0.5978
MinMax_PCC_RFE_8_DT	155	84	71	0.5804	[0.5067–0.6600]	00.0394	0.5806	1	0.583	0.575	0.6203	0.5395

Table 5a

Results of the DeLong test between genomics models of training set.

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Classifier name	<i>p</i> value
LR-RF	< 0.0001
LR-SVM	0.402
LR-DT	< 0.0001
RF-SVM	< 0.0001
RF-DT	1
SVM-DT	< 0.0001

Table 5b

Results of the DeLong test between genomics models of validation set.

Classifier name	p value
LR-RF	0.399
LR-SVM	0.514
LR-DT	0.0003
RF-SVM	0.717
RF-DT	0.003
SVM-DT	0.002

as lncRNA, miRNA, and methylation modifications [16-18]. This is evident by the fact that combined models of mRNA and lncRNA achieved higher predictive efficacy compared with the two individual models [19]. Furthermore, a multi-omics model significantly outperformed the single-omics model [20]. These findings may explain the reason why the gene model we developed had a lower predictive performance. In a similar way, the differences in predictive performance between the radiomics model and the mRNA model utilized in our study suggest that radiomics may be influenced by and reflect multi-dimensional biological information. Previous studies have indicated that CT radiomic features can predict variation of different dimensions such as individual key genes and methylation phenotypes [21–24]. These results are also consistent with the theory that radiomic features can reflect changes in different dimensions, containing information not found in the genomic features [25]. By connecting different dimensions of biological information, the values of radiomic features may serve as indicators of the product of multi-omics molecule interaction through activation cascade.

One of the limitations in deploying radiomics analysis in clinical practice is the lack of interpretability. Previous studies have shown the ability of the four imaging features to reflect tumour progression. Firstorder_TotalEnergy has been reported to predict the aggressiveness of papillary thyroid cancer and the T-stage of colorectal cancer [26,27]. Both glrlm_RunLengthNonUniformity and glszm_SizeZoneNonUniformity can distinguish large nodular medullary thyroid carcinoma from papillary thyroid carcinoma. In addition, glszm_SizeZoneNonUniformity can predict the non-perfusion volume ratio (NPVR) after high-intensity focused ultrasound ablation of uterine fibroids, which could reflect the prognosis after treatment [28–30]. NGTDM_strength was found to be an independent preoperative predictor of T-stage in rectal cancer [31]. The rising popularity of radiogenomics suggests that radiomic features are increasingly relied on to elucidate biological mechanisms related to genes and pathways. A total of 13 MRI radiomic features were associated

with immune regulation, cell function, tumour proliferation and treatment response in glioblastoma [32]. CT radiomics have been shown to be predictive of postoperative metastasis of ccRCC, which was correlated with ECM-receptor interactions, focal adhesion and PI3-Akt pathways [33]. Our KEGG results are in agreement with prior research revealing the critical role of these pathways in relation to the nuclear grade. Other ccRCC-related pathways include the MAPK pathway, which has been associated with tumour aggressiveness [34] and the HIF-1 pathway demonstrated to be associated with nuclear grade [35]. Furthermore, the Ras signalling pathway, which is related to the HIF pathway, has been reported to be involved in the regulation of ccRCC progression and drug resistance [36]. CT radiomic features correlate with genes associated with clinical outcomes, such as postoperative distant metastasis, overall survival, and immune subtypes of ccRCC [10, 33,37]. Radiomic features-associated genes was enriched in critical pathways for ccRCC development, suggesting that four radiomic features was related to the MAPK, HIF-1 and Ras signalling pathways.

However, previous radiogenomic studies have not yet investigated the generality of this association. Interestingly, in the present study we found that hub genes which were not associated with radiomic features exhibited comparable ccRCC nuclear grade predictive performance. Our findings indicate that non-associated genes may also constitute an added value to the radiomic model. A glioma research study comparing a radiomic model separately with a combined genetic, clinical and radiomics model revealed that the C-index improved from 0.75 to 0.81 [38]. A deep learning survival prediction model combining MRI radiomic features with gene expression data, indicated that the added genetic markers contributed to improving the predictive performance of the radiomics model [39]. These studies indicate that genes have an added value for radiomics and deep learning models. Genomics could complement genomic-related information which cannot be reflected by radiomic features. Radiomics may contain biological information beyond a single dimension. Considering the possible complementary relationship between radiomics and genomics, combined models of radiogenomics may help comprehensively decipher tumour-related biological information.

However novel, our study has several limitations. First, the radiogenomic development cohort had a small sample size. Nevertheless, we have used multi-institutional data for radiomics analysis and utilized WGCNA results from genomics analysis for the radiogenomic development cohort ensuring the robustness of gene modules and radiomic features in distinguishing nuclear grades. Second, although we detected genes which were not associated with the radiomic features, we did not estimate the added value of those genes in the prediction performance of our model. The above shortcomings will be addressed in a follow-up study, where more samples of both imaging, gene expression as well as multi-omics data will be processed in order to enhance the validity and interpretability of radiomics. Finally, the corticomedullary phase used in this study may be difficult to obtain highly reproducible radiomic features, although we validated its stable predictive efficacy in the test set, and in the future study we will use the arterial phase to validate the efficacy of the classification of the four radiomic features.

In conclusion, the major findings of this study were the following: a) Both radiomic model constructed based on CT cortex medullary phase and the mRNA model from the TCGA database can predict nuclear grade, and b) The predictive radiomic features were associated with only a part of the hub genes related to nuclear grade. In conclusion, this study revealed differences in the performance between radiomic and genomic mRNA models for predicting nuclear grade of ccRCC. A non-universal association between the radiomic features and genetic data at the mRNA level was found, which could provide useful insights for future radiogenomic studies.

Fund information

This study was supported by the entrepreneurship project of Gansu University of Chinese Medicine (grant number. LCCX2021006).

CRediT authorship contribution statement

Xuan-ming He: Investigation, Formal analysis, Data curation, Writing – original draft, Visualization, Software. Jian-xin Zhao: Resources, Data curation. Di-liang He: Resources, Data curation. Jia-liang Ren: Formal analysis, Visualization, Software. Lian-ping Zhao: Project administration. Gang Huang: Conceptualization, Methodology, Validation, Writing – review & editing, Supervision, Funding acquisition.

Acknowledgements

Thanks to contribution of TCGA database for their platforms and datasets. And thanks to the researchers and study participants for their contributions and finance support of grant from innovation and entrepreneurship project of Gansu University of Chinese medicine. We would like to thank Editage (www.editage.cn) for English language editing.

Consent for publication

The authors affirm that human research participants provided informed consent for publication of all the images.

Author contributions

All authors contributed to the study conception and design, GH and XH designed the experimental flow. JZ and DH collected and downloaded the data, XH and RJ analyzed the data and wrote the manuscript. All authors read and approved the final manuscript.

Ethics approval

The studies involving human participants were reviewed and approved by The Ethics Committee of Gansu Provincial Hospital. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.ejro.2023.100476.

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