

## ORIGINAL PAPER

doi: 10.5455/medarch.2020.74.332-336

MED ARCH. 2020 OCT; 74(5): 332-336

RECEIVED: SEP 12, 2020 | ACCEPTED: OCT 20, 2020

<sup>1</sup>Department of Surgery, Sultan Agung Islamic University, Semarang, Indonesia

<sup>2</sup>Stem Cell and Cancer Research, Sultan Agung Islamic University, Semarang, Indonesia

<sup>3</sup>Department of Pathological Anatomy, Sultan Agung Islamic University, Semarang, Indonesia

<sup>4</sup>Department of Postgraduate Biomedical Science, Sultan Agung Islamic University, Semarang, Indonesia

<sup>5</sup>Department of Surgery, Faculty of Medicine, Universitas Sumatera Utara, Medan, Indonesia

<sup>6</sup>Sultan Agung, Islamic University, Semarang, Indonesia

**Corresponding author:** Agung Putra, Stem Cell And Cancer Research (SCCR), Medical Faculty, Sultan Agung Islamic University (UNISSULA), Semarang 50112, Central Java, Indonesia.  
E-mail: dr.agungptr@gmail.com phone: +628164251646. ORCID ID: <http://www.orcid.org/0000-0001-6519-0384>.

# Mesenchymal Stem Cell-injected Omental Patch More Effective Promoting Wound Healing in Bowel Perforation Animal Model

Vito Mahendra Ekasaputra<sup>1</sup>, Agung Putra<sup>2,3,4</sup>, Adi Muradi Muhar<sup>5</sup>, Jherin Varessa<sup>6</sup>, Rafeni Bunga Cikita<sup>6</sup>, Satya A.P. Wijaya<sup>6</sup>, Mohammad Ariq Nazar<sup>2</sup>

## ABSTRACT

**Introduction:** Bowel perforation (BP) occurs as the complication of many gastrointestinal problems. Omental patch (OP) is one of the methods to place omentum flaps in the perforated area. Mesenchymal stem cells (MSCs) may increase regeneration process in all tissues.

**Aim:** to demonstrate the role of MSC in accelerating of wound healing process by analyzing fibroblast and collagen appearance in perforated bowel conditions. **Methods:** Using a BP rabbit model, 18 rabbit were randomly assigned into three groups: combination of umbilical cord (UC)-MSCs injection and OP (T1), OP only (T2) and vehicle control (Veh). Hematoxylin-eosin staining and Masson's trichrome staining were performed to analyze the level of fibroblast and collagen. Wound length were measured using standardized caliper. **Results:** The study showed a significant ( $P < 0.05$ ) increase of fibroblast and collagen amount on T1 and T2, in which T1 was higher than T2. This result was also followed by the decrease of wound length.

**Conclusion:** The combination of MSCs and OP-sutured in perforated bowel are better to accelerate wound healing than OP only in BP cases.

**Keywords:** Mesenchymal stem cells, fibroblast, collagen, wound length, Bowel perforation.

## 1. INTRODUCTION

Bowel perforation (BP) is the most disastrous complication of gastroduodenal ulcers, that potentially lead to peritonitis and eventually sepsis (1, 2). Although the incidence of BP is 7 to 10 in 100.000 population, the BP is a life-threatening catastrophe that requires immediate surgical repair (3). The one minimum invasive technique for BP treatment is using a laparoscopic closure (4). However, It takes more operative time and necessarily needs trained personnel that not available everywhere yet. This procedure is not the best choice to control the BP in the majority of hospitals. On the other hand, the simple closure of the BP using the omentum patch (OP) has been suggested method (5). Most of them were healed completely, nevertheless under a certain condition such as elderly patients, they have a high risk of death, owing to abnormal healing and gastrointestinal leakage (6). While majority studies highlight the benefits of mesenchymal stem cells (MSCs) in repairing tissue damage including in accelerating wound healing (7, 8). Therefore, combining the MSCs to OP in regard to control BP is a rational option to achieve the optimum healing.

MSCs as multipotent stromal progenitor cells express several surface markers such as CD105, CD90, and CD73, and less express CD11b, CD14, CD19 or CD79a, CD45, CD34, or Human Leucocyte Antigen (HLA) class II. Most studies reported that MSCs shown the capacity to differentiate into multiple tissue-forming cell lineages and promote tissue regeneration by releasing various growth factor (9). On the other hand, omentum tissues also contain mesothelial cells that have properties of MSCs known as adipose-derived MSCs (AD-MSCs). Specifically, AD-MSCs have the ability to promote tissue repair and differentiate into cartilage, bone, tendon, and fat under specific conditions (10). Previous studies reported that MSCs have been used to repair a variety of organ injuries including lung, peritoneum, kidney, and liver, with encouraging results (11-15).

Under certain condition, MSCs can migrate into the damaged tissues and show immunomodulatory capabilities to inhibit an inflammatory response

© 2020 Vito Mahendra Ekasaputra, Agung Putra, Adi Muradi Muhar, Jherin Varessa, Rafeni Bunga Cikita, Satya A.P. Wijaya, Mohammad Ariq Nazar

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

(9). Furthermore, the active MSCs may activate fibroblast cells to synthesize higher amounts of collagen including extracellular matrix to accelerate wounds healing (16). Nevertheless, the little bit amount of MSCs in the normal omentum tissues and mostly inactive state is a point crucial in healing acceleration. Therefore to activate and optimize the MSCs are needed. Thus, injecting MSCs to OP sutured in BP site could give a better outcome in wound healing process rather than MSCs or OP procedure only. However, there is no yet study reported the combination of MSCs and OP in BP. In this study, we investigated the effects of the combination of MSCs and OP-sutured in perforated bowel to accelerate wound healing of BP cases by analyzing the increase of fibroblast quantity as one of the healing process markers.

**2. AIM**

The aim of this study was to demonstrate the role of MSCs in accelerating of wound healing process by analyzing fibroblast and collagen appearance in perforated bowel conditions.

**3. MATERIAL AND METHODS**

**Isolation and culture of MSCs**

MSCs were isolated from female New Zealand white rabbits' umbilical cords. Phosphate Buffer Solution (PBS) (Gibco TM Invitrogen, NY, USA) with 5% Pen-strep antibiotic were used as a transport medium. The Wharton's jelly was separated from umbilical cord and minced evenly then placed into the 75 cm2 flask containing Dulbecco's Modified Eagle's Medium (DMEM) (Sigma-Aldrich, Louis St, MO) mixed with 10% Fetal Bovine Serum (FBS) (Gibco) TM Invitrogen, NY, USA), and 100 IU/mL penicillin/streptomycin (Sigma-Aldrich). The cultured Wharton's Jelly was incubated in 5% CO2 and 37°C incubator. The medium was changed in three days intervals and the MSCs will emerge in seven to 10 days. After reaching 80% confluence, the MSCs were passaged by trypsin. The 4th passage was used for the experiment. This study was approved by the Institutional Review Board of the Ethics Committee of the Medical Department, Sultan Agung Islamic University, Semarang, Indonesia.

**BP animal models**

Eighteen New Zealand white male rabbits weighing between 2000-3000 grams were veterinary at 24°C with 12h light-dark cycle and fed with water and food. Briefly, sutured gastric perforation in rabbits was performed under sterile conditions, as described previously the rabbits were anesthetized by intraperitoneal administration of ketamine (80 mg per kg body weight) after fasting for 12h. A midline abdominal incision was made to expose the stomach was exposed along the midline abdominal incision. A 2-cm vertical perforation was made at the gastric body using a scalpel, allowing the gastric lumen to open into the

abdominal cavity. Subsequently, the gastric perforation was disinfected with iodophor, and closed employing 5/0 non-absorbable suture (Ethicon, Johnson & Johnson, Somerville, New Jersey, USA) at 4-mm intervals.

**Administration of MSCs**

Eighteen New Zealand white male rabbits were selected and divided randomly into three groups (n = 6). Vehicle group (Veh) received no treatment and intervention, only sutured on the incision's mark. The treatment group has had an OP (T1) and MSCs-injected OP (T2) on the abraded ileum. Tissue was obtained from the rabbits after 14 days of observation and stored in phosphate-buffered saline (PBS, Sigma) with 1% (v/v) penicillin-streptomycin (Figure 1).

**In Vitro Differentiation**

The MSCs were grown in 10 % fetal bovine serum (FBS) in DMEM and osteogenic induction medium containing 10 mmol/L β-glycerophosphate, 10<sup>-7</sup> mol/L/0.1 μM dexamethasone, 50 μmol/L ascorbate-2-phosphate (Sigma-Aldrich, Louis St, MO), at 37°C and 5% CO2. The fixed cells were stained with 0.2% Alizarin Red solution (Sigma-Aldrich) to represent calcium deposition (cells used were from the 4th passage).

**Histopathological evaluation**

Wound tissues from Veh and treatment group were taken out from the healed wounds of the animals in excision and incision wound models for histopathological examinations. The thin sections were cut and stained with Hematoxylin and eosin (H&E) then observed under a microscope for the histopathological changes such as fibroblast proliferation, collagen formation, and angiogenesis.

**Wound length measurement**

Wound length measured before the rabbit was terminated. Wound length measured using calipers and calculated every rabbit in each group. All measurement was recorded in millimeters.

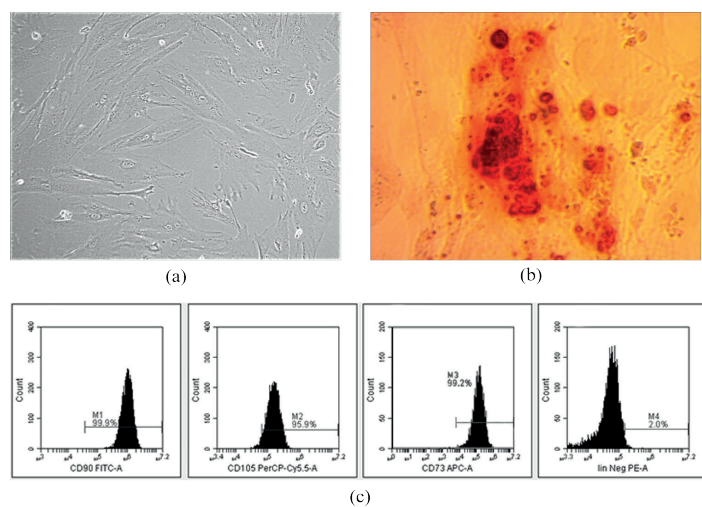


Figure 1. a) MSCs candidates from the in vitro culture showed fibroblast-like cell characteristics, b) Differentiation assays revealed that UC-MSCs could differentiate into osteocytes- via staining by Alizarin red which enabled osteocytes to appear red among the MSCs population, c) Flow cytometry analysis of UC-MSCs related immunophenotypes. Most UC-MSCs expressed positive markers (CD90, CD105, and CD73)



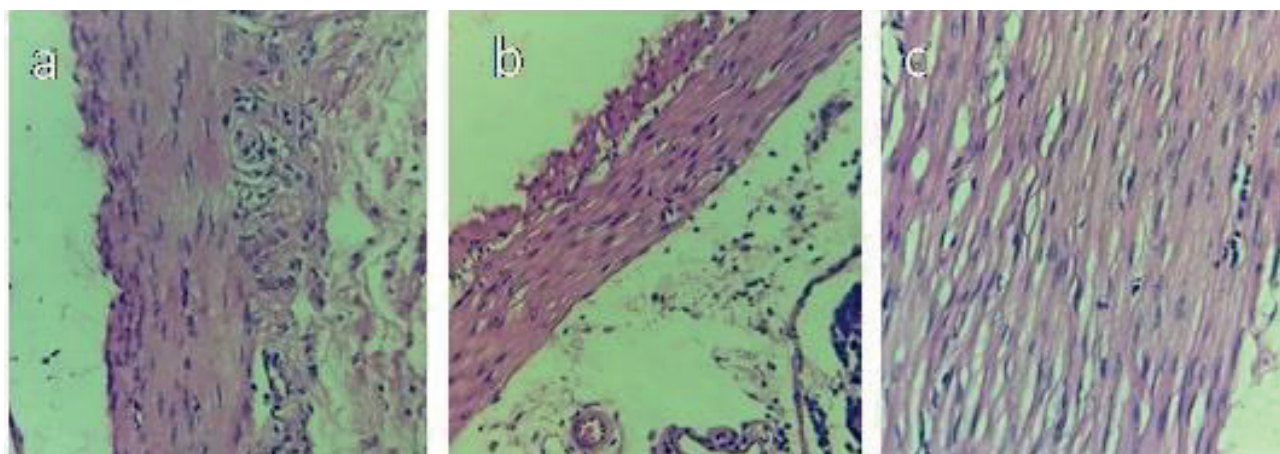


Figure 2. Histology analysis after H&E staining of paraffin-embedded bowel section a) vehicle group which lowest level of fibroblast and collagen, b) treatment group has had an OP only and c) MSCs-injected OP group

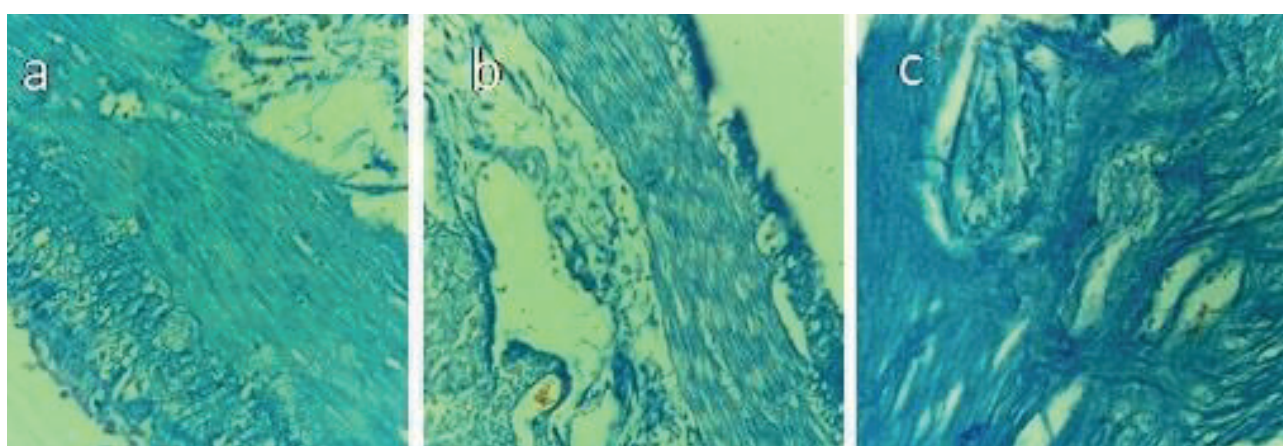


Figure 3. Histological appearance of the healing perforated wound a) vehicle group b) treatment group has had an omental patch only and c) MSCs-injected omental patch group

#### Statistical analysis

Statistical comparisons were performed using the One-Way ANOVA and Kruskal Wallis test (SPSS 23.0), and results were expressed as mean + SDs. Statistical correlations were performed using Pearson and Spearman tests.  $P < 0.05$  was considered statistically significant.

## 4. RESULTS

#### MSCs culture and characterization

Spindle-like shaped cells adhered to the culture flasks and differentiated into osteoblasts after being induced by the osteoblast differentiation medium. The characteristics of MSCs were expressing high levels of CD73 (99.2%), CD90 (96.7%), and CD105 (67.1%) in flow cytometric analysis.

#### Histology Evaluation

To evaluate the effects treatment of UC-MSCs to BP animal model after 14 days, the healed wounds of the animals were stained by H&E. The Veh showed  $86.67 \pm 0.76$  of mean fibroblast. In T1 (OP only) demonstrated a better fibroblast visualization by  $106.17 \pm 0.477$ . The highest fibroblast visualized on T2 (OP and MSCs site injection) with  $12.2 \pm 0.73$ . There was a significant difference among all groups ( $p < 0.05$ ). The result showed significant increase of collagen formation ( $p < 0.05$ ) of in Veh

( $11.33 \pm 0.211$ ), T1 ( $12.83 \pm 0.167$ ), and T2 ( $13.67 \pm 0.211$ ) (Figure 2 and 3).

#### Wound length measurement

The wound healing was evaluated by wound length enclosure measurement after 14 days. The measurement showed significant difference ( $P < 0.05$ ) between the wound length in Veh, T1, and T2 (respectively  $0.92 \pm 0.155$ ;  $0.65 \pm 0.108$ ;  $0.02 \pm 0.007$  mm).

## 5. DISCUSSION

In BP cases, the OP method may accelerate wound healing by increasing some specific neovascularization and reducing complications afterward due to these omentum-contained MSCs (17-19). A little bit amount of these MSCs in the omentum tissue and mostly inactive state lead to inadequately repair in tissue damage including the BP cases. Injecting MSCs to OP-sutured in the BP site could accelerate wound healing process. We assumed this combination may help in sealing the perforation of the damaged area through neovascularization formation and cellular proliferation to accelerate the BP healing and potentially prevents recurrence. Therefore, to explore the effects of the combination of MSCs and OP-sutured in perforated bowel, we used the BP animal model as previous study and analyzed fibroblast quantity as one of the healing process markers.

This study showed a significant difference in fibroblast levels among treatment groups in which the highest level was in the combinations of OP and MSCs group. The fibroblast enhancement following MSCs administration had been considerably studied in some reports that associated with a collagen increase leading to the wound healing acceleration. This is in line with a previous study that reported MSCs become active in the injury area to promote wound closure whether using the release of paracrine molecules or transdifferentiation mechanisms (20-22). MSCs signaling has also been shown to positively regulate cell survival, proliferation, and migration, besides increasing fibroblast gene expression (23-25). This finding was supported by in vitro study that reported there were not fibroblast cells apoptosis when treated with MSCs. The rule of MSCs in wound healing mechanism was proposed by reepithelization improvement, angiogenesis formation, granulation tissue growth, extracellular matrix restoration, following the in-situ inflammation process is under control (26).

In this study, we supposed the controlled inflammation in BP areas triggered the proliferation and maturation of fibroblasts (27). There were double paracrine signaling loops between fibroblast cells and mesothelium containing endogenous MSCs, known as crosstalk or dynamic reciprocity to normalize tissue homeostasis after injury (28). Acute inflammatory cells at the wound sites initially promote fibroblast migration however the active exogenous MSCs concurrently with endogenous MSCs of omental release paracrine molecules to activate fibroblasts. The active fibroblasts known as myofibroblastic release platelet-derived growth factor (PDGF) and basic fibroblast growth factor (bFGF/FGF-2) to synthesize collagen and promote cross-linking for supporting wound closure (29, 30). This is in line with our finding in which there were a significant increase of collagen levels in the treatment group particularly in the combination between MSCs and OP groups and also positive correlation to fibroblast levels. The collagen accumulation dramatically affect wound healing.

This study has several limitations in which we did not analyze several growth factors secreted MSCs such as VEGF, FGF, and PDGF. Therefore, the understanding of the process underneath of the BP must be explored to get a comprehensive healing mechanism in BP cases.

## 6. CONCLUSION

In summary, this study demonstrates the combination of the MSCs and OP-sutured in perforated bowel are more active to accelerate wound healing in BP cases.

- **Author's contribution:** All authors were involved in all steps of preparation of this article. Final proof reading was made by the first author.
- **Conflict of interest:** None declared.
- **Financial support and sponsorship:** Nil.

## REFERENCES

1. Mouly C, Chati R, Scotté M, Regimbeau JM. Therapeutic management of perforated gastroduodenal ulcer: Literature review. *J Visc Surg.* 2013 Nov; 150: 333-340.
2. Aboobakar MR, Singh JP, Maharaj K, Mewa Kinoo S, Singh B. Gastric perforation following blunt abdominal trauma. *Trauma Case Rep.* 2017 Aug; 10: 12-15.
3. Macaluso C, McNamara. Evaluation and management of acute abdominal pain in the emergency department. *Int J Gen Med.* 2012 Sep; 789.
4. Shah FH, Mehta SG, Gandhi MD, Saraj. Laparoscopic Peptic Ulcer Perforation Closure: the Preferred Choice. *Indian J Surg.* 2015 Dec; 77(S2): 403-406.
5. Husain M, Khan R, Rehmani B, Haris H. Omental patch technique for the ileal perforation secondary to typhoid fever. *Saudi J Gastroenterol.* 2011; 17(3): 208.
6. Richter-Schrag HJ, Richter S, Ruthmann O, Olschewski M, Hopt UT, Fischer A. Risk Factors and Complications Following Percutaneous Endoscopic Gastrostomy: A Case Series of 1041 Patients. *Can J Gastroenterol.* 2011; 25(4): 201-206.
7. Hu MS, Borrelli MR, Lorenz HP, Longaker MT, Wan DC. Mesenchymal Stromal Cells and Cutaneous Wound Healing: A Comprehensive Review of the Background, Role, and Therapeutic Potential. *Stem Cells Int.* 2018; 2018: 6901983.
8. Hocking AM. Mesenchymal Stem Cell Therapy for Cutaneous Wounds. *Adv Wound Care (New Rochelle).* 2012; 1(4): 166-171.
9. Putra A, Ridwan FB, Putridewi AI, et al. The Role of TNF- $\alpha$  induced MSCs on Suppressive Inflammation by Increasing TGF- $\beta$  and IL-10. *Open Access Maced J Med Sci.* 2018; 6(10): 1779-1783.
10. Alatab S, Shekarchian S, Najafi I, et al. Systemic Infusion of Autologous Adipose Tissue-Derived Mesenchymal Stem Cells in Peritoneal Dialysis Patients: Feasibility and Safety. *Cell J.* 2019; 20(4): 483-495.
11. Wang R, Zhu C, Qiao P, Liu J, Zhao Q, Wang K, et al. Experimental treatment of radiation pneumonitis with human umbilical cord mesenchymal stem cells. *Asian Pac J Trop Med.* 2014 Apr; 7(4): 262-266.
12. Putra A, Rosdiana I, Darlan DM, et al. Intravenous Administration is the Best Route of Mesenchymal Stem Cells Migration in Improving Liver Function Enzyme of Acute Liver Failure. *Folia Med (Plovdiv).* 2020; 62(1): 52-58.
13. Muhar AM, Putra A, Warli SM, Munir D. Hypoxia-Mesenchymal Stem Cells Inhibit Intra-Peritoneal Adhesions Formation by Upregulation of the IL-10 Expression. *Open Access Maced J Med Sci.* 2019; 7(23):3937-3943.
14. Putra A, Pertiwi D, Milla MN, et al. Hypoxia-preconditioned MSCs Have Superior Effect in Ameliorating Renal Function on Acute Renal Failure Animal Model. *Open Access Maced J Med Sci.* 2019; 7(3): 305-310.
15. Sungkar T, Putra A, Lindarto D, Sembiring RJ. 2019. The Effect of Mesenchymal Stem Cells for The Reduction of Liver Fibrosis through Platelet Derived Growth Factor- $\alpha$  Regulation in Rats. *Biochem Cell Arch.* 2019. 19(Supp. 2): 4749-4753.
16. Rodriguez-Menocal L, Salgado M, Ford D, Van Badiavas E. Stimulation of Skin and Wound Fibroblast Migration by Mesenchymal Stem Cells Derived from Normal Donors and Chronic Wound Patients. *Stem Cells Transl Med.* 2012 Mar; 1(3): 221-229.
17. Kidwai R, Ansari MA. Graham Patch Versus Modified Graham Patch in the Management of Perforated Duodenal Ulcer. *J Nepalgunj Med Coll.* 2017 Jan 17; 13(1): 28.

18. Shah OJ, Bangri SA, Singh M, Lattoo RA, Bhat MY. Omental flaps reduces complications after pancreaticoduodenectomy. *Hepatobiliary Pancreat Dis Int*. 2015 Jun; 14(3): 313-319.
19. Moon KC, Lee JS, Han SK, Lee HW, Dhong ES. Effects of human umbilical cord blood-derived mesenchymal stromal cells and dermal fibroblasts on diabetic wound healing. *Cytotherapy*. 2017 Jul; 19(7): 821-828.
20. Hocking AM, Gibran NS. Mesenchymal stem cells: Paracrine signaling and differentiation during cutaneous wound repair. *Exp Cell Res*. 2010 Aug; 316(14): 2213-2219.
21. Hu MS, Borrelli MR, Lorenz HP, Longaker MT, Wan DC. Mesenchymal Stromal Cells and Cutaneous Wound Healing: A Comprehensive Review of the Background, Role, and Therapeutic Potential. *Stem Cells Int*. 2018; 2018: 1-13.
22. Otero-Viñas M, Falanga V. Mesenchymal Stem Cells in Chronic Wounds: The Spectrum from Basic to Advanced Therapy. *Adv Wound Care*. 2016 Apr; 5(4): 149-163.
23. Foote AG, Wang Z, Kendzierski C, Thibeault SL. Tissue specific human fibroblast differential expression based on RNA-sequencing analysis. *BMC Genomics*. 2019 Dec; 20(1).
24. Shabbir A, Cox A, Rodriguez-Menocal L, Salgado M, Badiavas EV. Mesenchymal Stem Cell Exosomes Induce Proliferation and Migration of Normal and Chronic Wound Fibroblasts, and Enhance Angiogenesis In Vitro. *Stem Cells Dev*. 2015 Jul 15; 24(14): 1635-1647.
25. Yin JQ, Zhu J, Ankrum JA. Manufacturing of primed mesenchymal stromal cells for therapy. *Nat Biomed Eng*. 2019 Feb; 3(2): 90-104.
26. Laverdet B, Micallef L, Lebreton C, Mollard J, Lataillade JJ, Coulomb B, et al. Use of mesenchymal stem cells for cutaneous repair and skin substitute elaboration. *Pathol Biol*. 2014 Apr; 62(2): 108-117.
27. Park SR, Kim JW, Jun HS, Roh JY, Lee HY, Hong I-S. Stem Cell Secretome and Its Effect on Cellular Mechanisms Relevant to Wound Healing. *Mol Ther*. 2018 Feb; 26(2): 606-617.
28. Lei Z, Singh G, Min Z, Shixuan C, Xu K, Pengcheng X, et al. Bone marrow-derived mesenchymal stem cells laden novel thermo-sensitive hydrogel for the management of severe skin wound healing. *Mater Sci Eng C*. 2018 Sep; 90: 159-167.
29. Etikala A, Bruce G, Hudkins K, Raghu G, Narayanan AS. LR8 expression in fibroblasts of healthy and fibrotic human tissues. *Biochem Biophys Rep*. 2017 Jul; 10: 165-171.
30. Pao SI, Chien KH, Lin HT, Tai MC, Chen JT, Liang CM. Effect of microgravity on the mesenchymal stem cell characteristics of limbal fibroblasts. *J Chin Med Assoc*. 2017 Sep; 80(9): 595-607.