PREVENTION AND EARLY DETECTION (N ARBER, SECTION EDITOR)

Lifestyle Modifications and Colorectal Cancer

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Abstract Many studies suggest that Western lifestyle and dietary factors may be responsible for the high incidence of colorectal cancer in industrialized countries. Consumption of high amounts of red and processed meat and low intake of fiber and multiple protective phytochemicals found in fruits, vegetables, and whole grains might be responsible for the high incidence of this neoplasm in the Western world. Additionally, obesity, lack of physical activity, tobacco and alcohol use, sleep deprivation, and other factors have been proven to further increase the risk of colorectal cancer. Identifying and understanding the mechanisms through which they impact colon carcinogenesis is needed for the introduction of protective lifestyle recommendations.

Keywords Colorectal cancer · Prevention · Lifestyle modification · Phytochemicals · Obesity · Physical activity

Introduction

Colorectal cancer (CRC) is the second commonest neoplasm in women and the third commonest neoplasm in men. According to the World Health Organization GLOBOCAN database, in 2008 approximately 1.2 million new cases of CRC were diagnosed and 608,000 people died of CRC [1]. Despite screening programs designed for early detection of CRC having been introduced in many countries, the incidence of this neoplasm has not decreased, partly because of low compliance [2]. The occurrence of this disease varies approximately 25-fold in different world regions. The highest CRC incidence is noted in highly industrialized countries, whereas in developing

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countries CRC is not diagnosed so commonly [3]. Additionally, the number of new CRC cases increases in countries that undergo rapid economic transformations and adopt a Western lifestyle. This phenomenon occurred in eastern European countries, Asia, and some South American countries in the last three decades [4]. This observation strongly suggests that one of the key mechanisms of carcinogenesis of this neoplasm is associated with environmental factors [5, 6]. Therefore, identifying those mechanisms seems to be the most important goal in avoiding risk factors and developing prophylactic strategies leading to decreased incidence of CRC.

Many environmental factors have been studied, with imbalanced diet, alcohol consumption, tobacco use, obesity, lack of physical activity, and sleep deprivation being identified as the main CRC etiological factors [7]. Alimentary factors seem to have a major role, and the highest CRC incidence is noted in countries where the diet consists mainly of red and processed meat with low amounts of fiber sources, i.e., fruits, vegetables, and grains. On the other hand, some dietary factors have been proven to have chemopreventive properties [7]. Many molecular mechanisms of environmental factors promoting carcinogenesis have been discovered recently, which gives hope that it will be possible to reduce the incidence of CRC.

Dietary Factors

Fiber-Rich Foods

One of the earliest hypotheses that indicated diet as a risk factor for CRC was based on the observation that in the areas of low incidence of this neoplasm (Africa, Asia) the consumption of fiber is greater than in the Western world [8]. A high-fiber diet consists mainly of vegetables, fruits, and grains. Their presence in meals contributes to decreased transit time through the gastrointestinal tract, dilution of colonic contents, and enhancement of bacterial fermentation, which leads to increased production of short-chain fatty acids (acetate,



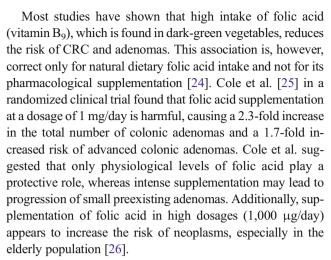
propionate, and butyrate) [9]. Moreover, these substances were found to induce apoptosis in CRC cells in rats [10]. Dietary fiber has also been proved to have an antiinflammatory function, decreasing the production of IL-6 and TNF α and also cyclooxygenase 2 (COX-2) and inducible nitric oxide synthase (iNOS) gene expression [11, 12]. Additionally short-chain fatty acids interfere with numerous regulators of the cell cycle, proliferation, and apoptosis, such as βcatenin, p53, p21, Bax, and caspase 3 genes in an animal model of CRC [13]. A meta-analysis based on 13 retrospective cohort studies presented by Howe et al. [14] in 1992 indicated that the risk of CRC is reduced by approximately 50 % in individuals who consume a high-fiber diet in comparison with the general population. However, prospective cohort studies performed in the USA did not confirm that fruit, vegetable, and grain intake decreases the risk of CRC [15, 16].

Another theory for the protective role of high-fiber diet indicates that such meals contain anticancer phytochemicals such as polyphenols, carotenoids, terpenes, and thioethers that interfere with cancer intracellular signaling cascades [17]. The presence of these substances is mainly observed in cruciferous and green leafy vegetables, onion, garlic, and citrus fruits. The above-mentioned studies presenting no chemopreventive role of high-fiber diet in CRC were conducted in the USA, where the most consumed vegetables are potatoes, lettuce, and tomatoes, which do not consist of many protective phytochemicals [18, 19]. The European Prospective Investigation into Cancer and Nutrition (EPIC) analyzed fiber intake and incidence of CRC in more than 500,000 individuals, and showed that doubling total fiber consumption decreases the incidence of this neoplasm by 40 % [20]. Moreover, in the observed population the incidence of CRC in individuals consuming high amounts of fruits and vegetables was 25 % lower than that in the remaining group [21].

The data presented clearly indicate that the risk of CRC can be decreased by consumption of plant-derived meals rich not only in fiber but also in protective phytochemicals. Many authors suggest that consumption of fruits and legumes, especially cruciferous and green leafy vegetables, might impact the prevalence of this disease.

B Vitamins

Foods rich in fiber, beside containing protective phytochemicals, are often rich in numerous vitamins. The B vitamins have received special attention mostly because of their role in DNA synthesis, repair, and methylation [22]. Recent molecular in vitro studies also showed that low folate levels enhance invasiveness of colon cancer cells mediated by activation of the Hedgehog Shh signaling pathway through promoter hypomethylation and stimulation of the nuclear factor κB (NF- κB) pathway [23].



The chemopreventive function of folic acid is partially explained by its abilities to suppress the loss of heterozygosity of the tumor suppressor gene *DCC* (deleted in CRC) and to attenuate the *EGFR* gene by the folic acid metabolite 5-methyltetrahydrofolate [27].

Vitamin B_6 , or pyridoxal phosphate, which is also found in numerous fruits, vegetables, and grains, is another important protective anticancer substance. Larson et al. [28] presented a meta-analysis of prospective studies, and showed that the risk of CRC decreases by 49 % for every 100 pmol/mL increase of pyridoxal phosphate concentration in the serum.

Red Meat and Processed Meat Intake

The consumption of red meat (beef, pork, lamb, veal, and mutton) is high in developed countries. A recent study by Daniel et al. [29] showed that its average daily intake in the USA is 128 g. Most consumed meat undergoes thermal processing or earlier preservation [29]. A recent meta-analysis of 21 prospective large-cohort studies indicates that the risk of CRC increases approximately linearly with higher intake of red and processed meat: the risk increased by 14 % for every daily 100-g increase of red meat intake [30••].

There are a few possible mechanisms that might explain that fact. One of the important factor leading to carcinogenesis might be increased intake of heme, which is present in red meat in high concentrations. Poultry and fish, considered as white meat, have tenfold lower amounts of this porphyrin pigment. Dietary heme is degraded in the small intestine by the enzyme heme oxygenase 1, releasing free ferrous iron [31]. Iron promotes the production of reactive oxygen species (ROS), especially H_2O_2 , which induces genetic mutations and expression of numerous cytokines (IL-6, IL-8, TNF α , NF- κ B), leading to increased cytotoxicity and stimulation of an inflammatory response [32].

Another mechanism that may be responsible for CRC development is connected to malondialdehyde, which is a product of lipid peroxidation. Malondialdehyde is a carcinogen, the



action of which leads to DNA degeneration, causing single-strand breaks and double-strand breaks [33]. Additionally, during digestion of red meats, N-nitroso compounds are created, resulting in the formation of DNA complexes by binding them through telomere stabilizing proteins (TRF2) [33]. Consumption of red meat leads also to excess production of oxysterols and aldehydes, which induces transforming growth factor β expression and promotes cell proliferation [34].

Processing of red meat, especially by frying or grilling at high temperatures, causes degradation of muscle creatinine and amino acids, resulting in the formation of numerous carcinogenic heterocyclic amines [35, 36].

The data presented indicate that consuming poultry, fish, or legumes instead of red meat as the main source of proteins would be beneficial in decreasing the risk of CRC. Additionally, avoiding meat processed at high temperatures might also have a protective effect.

Grape Seed Extract

Grape seed extract (GSE) is a mixture of polyphenols, mainly proanthocyanidins. Molecular in vivo studies have shown that it inhibits aberrant β -catenin, cyclin D1, and c-myc expression, preventing cycle cell disruption. Additionally, GSE reduces expression of iNOS and COX-2, decreasing oxidative cellular stress [37]. Derry et al. [38] performed in vitro studies on CRC cell lines, and showed that GSE induces their apoptosis, mainly due to activation of casapses 3, 8, and 9 and also generation of ROS. Moreover the proapoptotic function of GSE was limited only to cancer cells and there was no effect in normal colonocytes.

However, the available data are based only on animal models and in vitro studies on human CRC cell lines, but GSE supplementation seems to hold promise for chemoprevention.

Silibinin

Silibinin is a falvonolignan extracted from milk thistle (Sylibum marianum). It is the main component of silymarin—a supportive medication used in the treatment of many liver diseases. Several in vitro and in vivo studies have shown its chemopreventive role in skin cancer, lung cancer, prostate cancer, bladder cancer, and CRC by targeting DNA mutation mechanisms, proliferation, metastatic signaling, and inflammation processes [39]. Silibinin was found to inhibit the Wnt/β-catenin pathway, causing further decrease of cyclin D1 and c-myc expression [39, 40]. Moreover, silibinin deactivates antiapoptotic proteins such as BCL-2, MCL-1, X-linked inhibitor of apoptosis protein, and survivin [37]. On the other hand, it upregulates transcription of the death receptors DR4 and DR5, promoting apoptosis [41]. A recent in vitro study on human CRC cell lines showed that silibinin inhibits TNF α activation of NF- κ B. Furthermore, it decreases expression of COX-2 and iNOS.

Additionally, silibinin causes endoplasmic reticulum stress and inhibits mitochondrial glucose uptake in CRC cells [42]. Importantly, silibinin is not toxic to normal colonic epithelium. Hoh et al. [43] conducted a study evaluating silibinin pharmacodynamics. Its levels were monitored in plasma, liver, and colonic tissue, and high concentrations of this substance were found in colonocytes in patients receiving this herbal supplement orally. This promising finding supports the need for further investigations on the chemopreventive properties of silibinin.

Curcumin

Curcumin is a phenolic extract of the spice turmeric (Curcuma longa) with antioxidative potential used in ancient medicine. Numerous early-phase clinical trials have proved the safety and low systemic concentration of curcumin after oral admission in CRC [44, 45•]. Curcumin is effective in promoting apoptosis and inhibiting DNA mutations, cancer cell proliferation, metastasis, and inflammation. Curcumin upregulates glutathione S-transferases and induces an ROS concentration that leads to p21 protein upregulation, inhibiting cancer cell growth [46, 47]. Curcumin interferes with the mitogenactivated protein kinase (MAPK) pathway, leading to decreased production of TNF α and COX-2 and downregulation of NF-kB and IL-6 expression, preventing development of inflammation [48, 49]. Lower NF-kB levels also result in downregulation of c-myc, cyclin D1 and BCL-2 genes, modulating the cell cycle [50].

Interesting results were obtained in in vitro studies on the colon cancer cell line HT-29 incubated with the combination of curcumin and the COX-2 inhibitor sulindac sulfone. The effects of such treatment resulted in a synergistic inhibitory effect on cell growth, cell cycle arrest, and induction of apoptosis of cancer cells [51]. Similar results were found in rats treated with 1,2-dimethylhydrazine. The curcumin and sulindac sulfone treatment reduced the number of aberrant crypt foci by 75 % comparing with the control group [51]. This finding may be clinically important since addition of curcumin allows lower and less toxic sulindac sulfone doses to be used, resulting in safer treatment of CRC.

Another study evaluated the effect of Coltect (a dietary supplement containing curcumin) used together with 5-aminosalicylic acid in vivo on rats treated with 1,2-dimethylhydrazine. Coltect reduced the number of aberrant crypt foci similarly as 5-aminosalicylic acid by 40 %. The use of both substances resulted in a synergistic effect, reducing the number of aberrant crypt foci by 70 % [52].

Furthermore, curcumin promotes cancer cell apoptosis by inducing expression of proapoptotic proteins (Bax, Bim, Bak, Noxa) and inhibiting expression of antiapoptotic proteins (BCL-2, BCL-xl) [53]. Curcumin also plays a role in decreasing vascular endothelial growth factor (VEGF) and matrix



metalloproteinase 9 expression, which prevents the formation of metastases [50]. Other recently presented data show that curcumin inhibits CRC growth in a murine model and in vitro by downregulating microRNA-21, which restores the function of the phosphatase and tensin homolog tumor suppressor gene [54]. Additionally, Patel et al. [55] have shown in in vitro studies that curcumin inhibits HER2 and insulin-like growth factor 1 (IGF-1) receptor expression in CRC cells that are resistant to 5-fluorouracil and oxaliplatin used as chemotherapy for this neoplasm.

Curcumin seems to be one of the most promising widely available chemopreventive agents which reduces CRC risk. However, studies focusing on the influence of dietary intake of meals rich in curcumin on the incidence of this CRC are required.

Epigallocatechin 3-Gallate

Epigallocatechin 3-gallate (EGCG) is a polyphenol found in tea leaves of *Camellia sinensis*. Animal model studies suggest its chemopreventive function on various levels [53]. EGCG is a strong antioxidant, preventing ROS formation. Additionally, it decreases the expression of growth factors (epidermal growth factor, IGF-1, VEGF), blocking cancer cell proliferation and metastasis formation [56]. EGCG also blocks the cell cycle through downregulation of the MAPK/extracellular-signal-regulated kinase 1/2 and p21 signaling pathways [57]. Furthermore, it induces apoptotic abilities by upregulation of p53 and p21 [58]. However, a clinical phase II study examining the effectiveness of EGCG in the treatment of advanced ovarian cancer did not show any benefits of this substance, and further investigations in CRC and other tumors are needed [59].

Probiotics

Intestinal microbial flora regulate multiple physiological functions related to cell proliferation, differentiation, and stimulation of immunity. The DNA mutations in CRC reduce the integrity of colonic epithelium, allowing bacterial translocation and further activation of the cytokines IL-1, IL-2, and IL-23 and stimulation of T_h17 lymphocytes, which further promotes cell proliferation [60•]. Interestingly, extensive colonization with pathogenic *Fusobacterium* species, which was earlier described as a possible etiological factor for inflammatory bowel diseases, was discovered in CRC tumors [61]. Additionally, genotoxic strains of *Escherichia coli* leading to numerous DNA mutations are often cultured from CRC tissue [62].

Probiotics similarly impact the molecular mechanisms of cell growth and differentiation. Many in vitro and animal model studies have shown multiple possible pathways through which probiotics might prevent CRC. Their presence might competitively reduce the number of pathological colonic flora, reduce carcinogenic secondary bile acid production, and increase

production of short-chain fatty acids, modulating the inflammatory response [63]. The type of mechanism depends on the microbial species. *Saccharomyces boulardi*, a yeast species, has antioxidant and anti-inflammatory effects. Polyamines produced by this species reduce the oxidative stress. Additionally, in vitro and animal model studies have shown that *S. boulardi* downregulates the MAPK/extracellular-signal-regulated kinase 1/2 signaling pathway, which might explain its potential anti-proliferative effect [64]. Other probiotics have different potential anticancer mechanisms: *Bifidobacterium animalis* modulates IGF-1 expression, and *Bacillus polyfermenticus* suppresses epidermal growth factor receptor pathway signaling [65, 66]. However, the lack of convincing data from clinical studies does not allow probiotics to be recommended as chemopreventive agents in CRC.

Alcohol

The risk of CRC and colonic adenomas is associated with alcohol consumption. Several prospective cohort studies have shown that consumption of more than 30 g of ethanol per day results in a multivariative risk of CRC of 1.16, whereas consumption of more than 45 g of ethanol per day increases that risk to 1.41 [67]. Metabolism of alcohol is based mainly on catalysis and oxidation by alcohol dehydrogenase, catalase, and cytochrome P450, resulting in the formation of acetaldehyde, which is a class 1 carcinogen and is responsible for chromosome damage [68•]. An in vitro study on ethanol and acetaldehyde genotoxicity showed a dose-dependent increase in the number of DNA strand breaks particularly in colonic mucosa cells and lymphocytes [69].

In addition to promoting carcinogen activity, long-term alcohol consumption decreases the absorption of group B vitamins (B₁, B₂, B₁₂, folic acid), which causes increased cell vulnerability to oxidative stress [68•]. Increased ROS levels induce the NADPH oxidase cascade, leading to activation of the phosphoinositide 3-kinase (PI3K)/AKT and VEGF pathway, which promotes proliferation and metastasis [70].

Additionally, alcohol blocks expression of the enzyme cytochrome P450 2E1, which is involved in vitamin A synthesis. The low vitamin A levels decrease the expression of activator protein 1—a transcription factor which controls cell differentiation and proliferation [68•].

Therefore, it is reasonable to recommend the reduction of alcohol consumption, which reduces the risk of CRC development.

Tobacco

Many studies have shown that tobacco use increases the risk of CRC and colonic adenomas [70]. The multicenter,



prospective study EPIC, conducted on 465,879 individuals with a mean follow-up of 8.7 years, shows that such risk is observed in both past and present smokers (relative risk 1.21) and most CRC tumors in smokers are located on the right side of the colon [71]. The main carcinogens found in tobacco smoke are aromatic amines, nitrosamines, heterocyclic amines, and polycyclic aromatic hydrocarbons. These substances undergo metabolism through cytochromes P450, leading to the formation of aberrant DNA and further gene mutation (KRAS, BRAF, MYC) [71]. Nitrosamines have the ability to activate and bind to nicotinic acetylcholine receptors, which results in an increase of intracellular ROS concentrations. The oxidative stress leads to activation of the NF-kB and COX-2 inflammatory pathways and also promotes the MAPK proliferative signaling cascade [72]. Nicotine activates βadrenoreceptors, which triggers inflammatory and metastatic signaling through the COX-2, metalloproteinase 2, and VEGF pathways [73].

There is no doubt that avoiding tobacco use decreases the risk of CRC and other neoplasms and also prevents the development of cardiovascular disease, pulmonary disease, and other diseases.

Obesity

Many studies have indicated that obesity, defined as body mass index (BMI) exceeding 25 kg/m², is a risk factor for multiple neoplasms. This has been confirmed by a recent meta-analysis by Renehan et al. [74]. Another systematic review concentrating only on CRC and adenoma incidence indicated their higher prevalence in obese individuals, particularly in men. Compared with BMI below 23.0 kg/m², for BMI of 23.0–24.9, 25.0–27.4, and 27.5–29.9 kg/m² and BMI greater than 30 kg/m², the risk of CRC was 14, 19, 24, and 41 %, respectively [75••, 76]. Moreover, abdominal visceral adipose tissue volume was proposed to be a more adequate risk factor for CRC than BMI or waist circumference, explaining the higher prevalence of this neoplasm in obese men than in obese women [77••].

The mechanisms for increased cancer incidence in obese individuals have not been fully elucidated. Adipose tissue is considered as a metabolically active organ releasing numerous hormones and cytokines and stimulating T cells that promote low-intensity chronic inflammation and insulin resistance [78]. Additionally, increased serum triglyceride concentration in obese individuals further enhances this process [79]. Hyperinsulinemia, which is the result of insulin resistance, contributes, together with IGF-1, to increased cell proliferation and potential carcinogenesis [80]. Moreover, adipose tissue secretes adipohormones such as leptin, adiponectin, and resistin which contribute to promotion of inflammation and carcinogenesis [81]. Booth et al. [82] have shown that

serum leptin and adiponectin levels were elevated in CRC patients and patients with colonic adenomas compared with healthy individuals.

The data presented suggest that body mass reduction, which decreases chronic inflammation, glucose intolerance, and dyslipidemia, might impact CRC incidence and prevent the development of multiple obesity-related diseases.

Physical Activity

Many chronic diseases, including cardiovascular, pulmonary, and musculoskeletal diseases, type 2 diabetes, and many types of cancer, are associated with insufficient physical activity [81]. A meta-analysis by Wolin et al. [83] indicates that most physically active individuals have 24 % lower risk of CRC development than those who have a sedentary lifestyle. The same authors conducted another systematic review, showing 16 % reduction in the incidence of colonic adenomas and 35 % reduction in the incidence of large colonic polyps in the physically active group [84••].

The key mechanisms that explain the protective role of physical activity focus on increased insulin sensitivity, lower insulin levels, decreased body mass, and decreased adipose tissue volume, leading to reduction of chronic inflammation [85].

Current guidelines of the American Institute of Cancer Research recommend a minimum of 60 min of moderate or 30 min of intense physical activity daily to promote health [86]. However, excessive exercise might be harmful, leading to increased oxidative stress and consequent DNA damage. Such observations were not found during moderate exercise [86].

Ashgar et al. [87] found in rat models that physical activity induces superoxide dismutase activity, which through the transcription factor Nrf2 initiates the expression of detoxifying enzymes such as glutathione S-transferases.

Another protective mechanism induced by physical activity focuses on the role of the adipohormone leptin, which induces proliferative signaling pathways through activation of the MAPK and PI3K/AKT cascades. Moderate exercise was found to decrease serum leptin levels [88]. Additionally, high adipose tissue volume induces low-grade inflammation, mainly through TNFα overexpression. Such chronic inflammation is believed to promote carcinogenesis [89]. Physical activity promotes the production of IL-6 and decreases the expression of iNOS and TNF α both in plasma and in colonic mucosa, leading to enhanced immunity and also increased lipolysis in adipose tissue [90, 91]. Moreover, Ju at el. [92] showed in an animal model that physical activity inhibits the IGF-1/insulin-like growth factor binding protein 3 expression and abnormal β -catenin signaling pathway. The aberrant β catenin pathway induces further modulation of oncogenic



genes such as c-myc and VEGF expression; therefore, maintaining the equilibrium of this cascade might be an important anticancer mechanism.

The clinical and molecular studies presented clearly indicate that physical activity leads to metabolic transformations resulting in a decrease of CRC risk. Additionally, weight reduction induced by physical exercise in overweight and obese individuals might also be protective.

Circadian Rhythm

Some authors have investigated carcinogenesis related to chronic sleep deprivation or disruption of circadian rhythm. Hrushesky et al. [93] have shown that nightshift workers, both men and women, have 50 % risk of developing CRC.

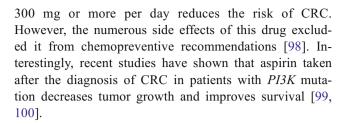
It has been proven that cell proliferation, differentiation, apoptosis, and DNA repair mechanisms have different day and night activities [93]. Some of the enzymes expressed in rhythmic manner are cytochromes P450 involved in detoxication mechanisms. Another such gene is the p21/Weel gene, which regulates the cell cycle, DNA replication, and mitosis [94]. Additionally, so-called circadian clock genes have been described and studied recently: *PER1*, *PER2*, *PER3*, and *CLOCK* [95]. Mice with a silenced *Per2* gene showed no p53 protein activity, which resulted in dysfunction of the DNA repair and apoptosis pathways [95]. Moreover *PER2* expression was shown to be high in cancer cells and to promote the c-myc, cyclin D, β-catenin, and VEGF signaling pathways [96].

The data presented suggest that maintaining a regular and adequate daily amount of sleep contributes to CRC prevention.

Other Factors

Numerous studies have indicated that oxidative stress is one of the key promoters of carcinogenesis, as presented earlier. Various protective substances have been examined. Vitamin E (α -tocopherol) has been shown to decrease colonic epithelial DNA damage evoked by bleomycin in in vitro studies [97]. Another study indicated that gene mutations in colonic mucosa induced by nickel could be attenuated by quercetin, a flavonoid obtained from many fruits and vegetables [98].

There are a variety of medications with a chemopreventive role in CRC development, but describing their function is beyond the scope of this article. However, recent data on the effects of aspirin intake in prevention of CRC are noteworthy. Aspirin inhibits COX-2, reducing the oxidative stress and limiting gene mutations. Previous studies have shown that aspirin at dosages of



Conclusion

The studies presented indicate that diet and lifestyle habits could impact the incidence of CRC. Therefore, it is reasonable to suggest general recommendations for lifestyle modifications that would decrease the risk of CRC. Replacing red and processed meat, highly saturated fats, refined starches, and sugars with fish, poultry, vegetables, fruits, and grains as sources of polyunsaturated fats, carbohydrates, and proteins might lower the risk of this neoplasm. Additionally avoiding smoking and alcohol consumption, reducing obesity, especially by physical activity, and maintaining adequate sleep patterns also appear to be very beneficial. Moreover, introducing a chemopreventive agent such as curcumin, GSE, EGCG, or silibinin might further lower the incidence of CRC.

Following those regulations would not only decrease the risk of CRC but would also improve overall health status and decrease the incidence of other neoplasms and cardiovascular disease, pulmonary disease, musculoskeletal disease, and multiple other diseases.

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Compliance with Ethics Guidelines

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Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.



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References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance
 - Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer. 2010;127(12):2893–917.
 - Bretthauer M. Evidence for colorectal cancer screening. Best Pract Res Clin Gastroenterol. 2010;24:417–25.
 - Huxley RR, Ansary-Moghaddam A, Clifton P, Czernichow S, Parr CL, Woodward M. The impact of dietary and lifestyle risk factors on risk of colorectal cancer: a quantitative overview of the epidemiological evidence. Int J Cancer. 2009;125:171–80.
 - Center MM, Jemal A, Ward E. International trends in colorectal cancer incidence rates. Cancer Epidemiol Biomarkers Prev. 2009;18:1688–94.
 - Gingras D, Beliveau R. Colorectal cancer prevention through dietary and lifestyle modifications. Cancer Microenviron. 2011;4(2):133–9.
 - Slomka M, Kaczka A, Malecka-Panas E. Environmental factors and the risk of colorectal adenomas and cancer. Clin Exp Med Lett. 2008;49(2):77–81.
 - Derry MM, Raina K, Agarwal C, Agarwal R. Identifying molecular targets of lifestyle modifications in colon cancer prevention. Front Oncol. 2013;3:119.
 - Burkitt DP. Related disease—related cause? Lancet. 1969;2: 1229–31.
 - Scharlau D, Borowicki A, Habermann N, et al. Mechanisms of primary cancer prevention by butyrate and other products formed during gut flora-mediated fermentation of dietary fibre. Mutat Res. 2009;682:39–53.
- Vergara-Castaneda HA, Guevara-Gonzalez RG, Ramos-Gomez M, Reynoso-Camacho R, Guzman-Maldonado H, Feregrino-Perez AA, et al. Non-digestible fraction of cooked bean (Phaseolus vulgaris L.) cultivar Bayo Madero suppresses colonic aberrant crypt foci in azoxymethane-induced rats. Food Funct. 2010;1:294–300.
- Reddy BS, Hirose Y, Cohen LA, Simi B, Cooma I, Rao CV. Preventive potential of wheat bran fractions against experimental colon carcinogenesis: implications for human colon cancer prevention. Cancer Res. 2000;60:4792

 –7.
- Kaczmarczyk MM, Miller MJ, Freund GG. The health benefits of dietary fiber: beyond the usual suspects of type 2 diabetes mellitus, cardiovascular disease and colon cancer. Metab Clin Exp. 2012;61:1058–66.
- Feregrino-Perez AA, Berumen LC, Garcia-Alcocer G, Guevara-Gonzalez RG, Ramos-Gomez M, Reynoso-Camacho R, et al. Composition and chemopreventive effect of polysaccharides from common beans (Phaseolus vulgaris L.) on azoxymethane-induced colon cancer. J Agric Food Chem. 2008;56:8737–44.
- Howe GR, Benito E, Castelleto R, et al. Dietary intake of fiber and decreased risk of cancers of the colon and rectum: evidence from the combined analysis of 13 case-control studies. J Natl Cancer Inst. 1992;84:1887–96.

- Willett WC, Stampfer MJ, Colditz GA, et al. Relation of meat, fat, and fiber intake to the risk of colon cancer in a prospective study among women. N Engl J Med. 1990;323:1664–72.
- Fuchs CS, Giovannucci EL, Colditz GA, et al. Dietary fiber and the risk of colorectal cancer and adenoma in women. N Engl J Med. 1999;340:169–76.
- Surh YJ. Cancer chemoprevention with dietary phytochemicals. Nat Rev Cancer. 2003;3:768–80.
- Krebs-Smith SM, Kantor LS. Choose a variety of fruits and vegetables daily: understanding the complexities. J Nutr. 2001;131:487S-501S.
- Johnston CS, Taylor CA, Hampl JS. More Americans are eating "5 a day" but intakes of dark green and cruciferous vegetables remain low. J Nutr. 2000;130:3063

 –7.
- Bingham SA, Day NE, Luben R, et al. Dietary fibre in food and protection against colorectal cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC): an observational study. Lancet. 2003;361:1496–501.
- van Duijnhoven FJ, Bueno-De-Mesquita HB, Ferrari P, et al. Fruit, vegetables, and colorectal cancer risk: the European Prospective Investigation into Cancer and Nutrition. Am J Clin Nutr. 2009;89: 1441–52.
- Powers HJ. Interaction among folate, riboflavin, genotype, and cancer, with reference to colorectal and cervical cancer. J Nutr. 2005;135:2960S–6S.
- 23. Wang TP, Hsu SH, Feng HC, Huang RF. Folate deprivation enhances invasiveness of human colon cancer cells mediated by activation of sonic hedgehog signaling through promoter hypomethylation and cross action with transcription nuclear factor-kappa B pathway. Carcinogenesis. 2012;33 (6):1158-68.
- Giovannucci E. Epidemiologic studies of folate and colorectal neoplasia: a review. J Nutr. 2002;132:23508–5S.
- Cole BF, Baron JA, Sandler RS, et al. Folic acid for the prevention of colorectal adenomas: a randomized clinical trial. JAMA. 2007;297:2351–9.
- 26. Mason JB, Kim SJ. Am J Gastroenterol. 2010;105:1914-6.
- Majumdar AP, Kodali U, Jaszewski R. Chemopreventive role of folic acid in colorectal cancer. Front Biosci. 2004;9:2725–32.
- Larsson SC, Orsini N, Wolk A. Vitamin B₆ and risk of colorectal cancer. A meta-analysis of prospective studies. JAMA. 2010;303: 1077–83.
- Daniel CR, Cross AJ, Koebnick C, Sinha R. Trends in meat consumption in the USA. Public Health Nutr. 2011;14:575–83.
- 30. Chan DS, Lau R, Aune D, Vieira R, Greenwood DC, Kampman E, et al. Red and processed meat and colorectal cancer incidence: meta-analysis of prospective studies. PLoS ONE. 2011;6:e20456. The article is a meta-analysis of 24 prospective cohort studies conducted in the USA, in ten European countries as part of EPIC, and in Finland, Sweden, the Netherlands, Japan, Australia, China, and Singapore. The data presented provide very strong evidence that "the risk increase in colorectal cancer estimated in linear dose-response models was 14% for every 100 g/day increase of total red and processed meats, 25% in colon cancer; and 31% in rectal cancer." The authors suggest that limiting red meat consumption could be a primary preventive measure in CRC.
- Ishikawa S, Tamaki S, Ohata M, Arihara K, Itoh M. Heme induces DNA damage and hyperproliferation of colonic epithelial cells via hydrogen peroxide produced by heme oxygenase: a possible mechanism of heme-induced colon cancer. Mol Nutr Food Res. 2010;54:1182–91.
- 32. Knobel Y, Weise A, Glei M, Sendt W, Claussen U, Pool-Zobel BL. Ferric iron is genotoxic in non-transformed and pre-neoplastic human colon cells. Food Chem Toxicol. 2007;45:804–11.



- O'Callaghan NJ, Toden S, Bird AR, Topping DL, Fenech M, Conlon MA. Colonocyte telomere shortening is greater with dietary red meat than white meat and is attenuated by resistant starch. Clin Nutr. 2012;31:60–4.
- Biasi F, Mascia C, Poli G. The contribution of animal fat oxidation products to colon carcinogenesis, through modulation of TGF-β1 signaling. Carcinogenesis. 2008;29:890–4.
- Sugimura T, Wakabayashi K, Nakagama H, Nagao M. Heterocyclic amines: mutagens/carcinogens produced during cooking of meat and fish. Cancer Sci. 2004;95:290–9.
- Martinez ME, Jacobs ET, Ashbeck EL, et al. Meat intake, preparation methods, mutagens and colorectal adenoma recurrence. Carcinogenesis. 2007;28:2019–27.
- Velmurugan B, Gangar SC, Kaur M, Tyagi A, Deep G, Agarwal R. Silibinin exerts sustained growth suppressive effect against human colon carcinoma SW480 xenograft by targeting multiple signaling molecules. Pharm Res. 2010;27:2085–97.
- Derry M, Raina K, Agarwal R, Agarwal C. Differential effects of grape seed extract against human colorectal cancer cell lines: the intricate role of death receptors and mitochondria. Cancer Lett. 2013;334:69–78.
- Rajamanickam S, Agarwal R. Natural products and colon cancer: current status and future prospects. Drug Dev Res. 2008;69:460–71.
- Kaur M, Velmurugan B, Tyagi A, Agarwal C, Singh RP, Agarwal R. Silibinin suppresses growth of human colorectal carcinoma SW480 cells in culture and xenograft through down-regulation of beta-catenin-dependent signaling. Neoplasia. 2010;12:415–24.
- Kauntz H, Bousserouel S, Gosse F, Raul F. The flavonolignin silibinin potentiates TRAIL-induced apoptosis in human colon adenocarcinoma and in derived TRAIL-resistant metastatic cells. Apoptosis. 2012;17:797–809.
- Raina K, Agarwal C, Agarwal R. Effect of silibinin in human colorectal cancer cells: targeting the activation of NF-κB signaling. Mol Carcinog. 2013;52:195–206.
- 43. Hoh C, Boocock D, Marczylo T, Singh R, Berry DP, Dennison AR, et al. Pilot study of oral silibinin, a putative chemopreventive agent, in colorectal cancer patients: silibinin levels in plasma, colorectum, and liver and their pharmacodynamics consequences. Clin Cancer Res. 2006;12:2944–50.
- Hatcher H, Planalp R, Cho J, Torti FM, Torti SV. Curcumin: from ancient medicine to current clinical trials. Cell Mol Life Sci. 2008:65:1631–52.
- 45.• Shehzad A, Wahid F, Lee YS. Curcumin in cancer chemoprevention: molecular targets, pharmacokinetics, bioavailability, and clinical trials. Arch Pharm. 2010;343(9):489–99. Curcumin is a phenolic extract of the spice turmeric (Curcuma longa) with chemopreventive potential. The article presents recent data regarding the molecular background on the interaction curcumin with numerous signaling pathways. Additionally, the authors describe promising results of early-phase clinical trials of this substance in the treatment of neoplasms (multiple myeloma, pancreatic cancer, and colon cancer)..
- Ye SF, Hou ZQ, Zhong LM, Zhang QQ. Effect of curcumin on the induction of glutathione S-transferases and NADP(H):quinone oxidoreductase and its possible mechanism of action. Yao Xue Xue Bao. 2007;42:376–80.
- Yogosawa S, Yamada Y, Yasuda S, Sun Q, Takizawa K, Sakai T. Dehydrozingerone, a structural analogue of curcumin, induces cell-cycle arrest at the G2/M phase and accumulates intracellular ROS in HT-29 human colon cancer cells. J Nat Prod. 2012;75: 2088–93.
- 48. Camacho-Barquero L, Villegas I, Sanchez-Calvo JM, Talero E, Sanchez-Fidalgo S, Motilva V, et al. Curcumin, a Curcuma longa constituent, acts on MAPK p38 pathway modulating COX-2 and iNOS expression in chronic experimental colitis. Int Immunopharmacol. 2007;7:333–42.

- Tu SP, Jin H, Shi JD, Zhu LM, Suo Y, Lu G, et al. Curcumin induces the differentiation of myeloid-derived suppressor cells and inhibits their interaction with cancer cells and related tumor growth. Cancer Prev Res (Phila). 2012;5:205–15.
- Chen C, Liu Y, Chen Y, Xu J. C086, a novel analog of curcumin, induces growth inhibition and down-regulation of NFκB in colon cancer cells and xenograft tumors. Cancer Biol Ther. 2011;12: 797–807.
- Giladi N, Kazanov D, Shpitz B, Aroch I, Kraus S, Arber N. Curcumin potentiates the pro-apoptotic effects of sulindac sulfone in colorectal cancer. Expert Opin Investig Drugs. 2010;19 Suppl 1: S117–24.
- Aroch I, Kraus S, Naumov I, et al. Chemopreventive effects of Coltect, a novel dietary supplement, alone and in combination with 5-aminosalicylic acid in 1,2-dimethylhydrazine-induced colon cancer in rats. Ther Adv Gastroenterol. 2010;3(5):281–9.
- Yang CS, Wang X, Lu G, Picinich SC. Cancer prevention by tea: animal studies, molecular mechanisms and human relevance. Nat Rev Cancer. 2009;9:429–39.
- Roy S, Yu Y, Padhye SB, Sarkar FH, Majumdar AP, et al. Difluorinated-curcumin (CDF) restores PTEN expression in colon cancer cells by down-regulating miR-21. PLoS One. 2013. doi:10. 1371/journal.pone.0068543.
- Patel BB, Gupta D, Elliot AA, Sengupta V, Yu Y, Majumdar AP. Curcumin targets FOLFOX-surviving colon cancer cells via inhibition of EGFRs and IGF-1R. Anticancer Res. 2010;30(2):319–25.
- Khan N, Mukhtar H. Multitargeted therapy of cancer by green tea polyphenols. Cancer Lett. 2008;269:269–80.
- Larsen CA, Dashwood RH. (-)-Epigallocatechin-3-gallate inhibits Met-signaling, proliferation, and invasiveness in human colon cancer cells. Arch Biochem Biophys. 2010;501:52–7.
- Thakur VS, Ruhul Amin AR, Paul RK, Gupta K, Hastak K, Agarwal MK, et al. p53-dependent p21-mediated growth arrest pre-empts and protects HCT116 cells from PUMA-mediated apoptosis induced by EGCG. Cancer Lett. 2010;296:225–32.
- Trudel D, Labbe DP, Araya-Farias M, et al. A two stage singe arm, phase II study of EGCG-enriched green tea drink as a maintenance therapy in women with advanced stage ovarian cancer. Gynecol Oncol. 2013;13(2):357–61.
- 60.• Grivennikov SI, Wang K, Mucida D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. Nature. 2012;491(7423):254–8. The authors present a very detailed molecular analysis of the role of microbial colonic flora in the development of adenomas. The results indicate the importance of IL-17 and IL-23 signaling and mucosal barrier integrity in colon adenoma and cancer growth..
- Kostic AD, Gevers D, Pedamallu CS, et al. Genomic analysis identifies association of Fusobacterium with colorectal carcinoma. Genome Res. 2012;22(2):292–8.
- Arthur JC, Perez-Chanona E, Muhlbauer M, et al. Intestinal inflammation targets cancer-inducing activity of the microbiota. Science. 2012;338(6103):120–3.
- Chong ES. A potential role of probiotics in colorectal cancer prevention: review of possible mechanisms of action. World J Microbiol Biotechnol. 2013. doi:10.1007/s11274-013-1499-6.
- 64. Chen X, Kokkotou EG, Mustafa N, Bhaskar KR, Sougioultzis S, O'Brien M, et al. Saccharomyces boulardii inhibits ERK1/2 mitogen-activated protein kinase activation both in vitro and in vivo and protects against Clostridium difficile toxin A-induced enteritis. J Biol Chem. 2006;281:24449–54.
- Matsumoto M, Kurihara S, Kibe R, Ashida H, Benno Y. Longevity in mice is promoted by probiotic-induced suppression of colonic senescence dependent on up regulation of gut bacterial polyamine production. PLoS ONE. 2011;6: e23652.



- Park E, Jeon GI, Park JS, Paik HD. A probiotic strain of Bacillus polyfermenticus reduces DMH induced precancerous lesions in F344 male rat. Biol Pharm Bull. 2007;30:569–74.
- Cho E, Smith-Warner SA, Ritz J, et al. Alcohol intake and colorectal cancer: a pooled analysis of 8 cohort studies. Ann Intern Med. 2004;140:603–13.
- 68.• Haas SL, Ye W, Lohr JM. Alcohol consumption and digestive tract cancer. Curr Opin Clin Nutr Metab Care. 2012;15:457–67. This review summarizes recent evidence on the impact of alcohol consumption on the risk of gastrointestinal neoplasms. It presents current knowledge of the molecular bases of ethanol-induced carcinogenesis, mostly through oxidative stress mechanisms and DNA damage caused by the ethanol metabolite acetaldehyde. The authors also present the results of recent systematic reviews which indicate that alcohol intake is strongly related to increased risk of gastrointestinal cancers (including colon cancer)...
- Blasiak J, Trzeciak A, Malecka-Panas E, Drzewoski J, Wojewodzka M. In vitro genotoxicity of ethanol and acetaldehyde in human lymphocytes and the gastrointestinal tract mucosa cells. Toxicol In Vitro. 2000;14:287–95.
- Jacobson JS, Neugut AI, Murray T, Garbowski GC, Forde KA, Treat MR, et al. Cigarette smoking and other behavioral risk factors for recurrence of colorectal adenomatous polyps (New York City, NY, USA). Cancer Causes Control. 1994;5:215–20.
- Leufkens AM, Van Duijnhoven FJ, Siersema PD, et al. Cigarette smoking and colorectal cancer risk in the European Prospective Investigation into Cancer and Nutrition study. Clin Gastroenterol Hepatol. 2011;9(2):137–44.
- Ye YN, Liu ES, Shin VY, Wu WK, Cho CH. The modulating role of nuclear factor-κB in the action of α₇-nicotinicacetylcholine receptor and cross-talk between 5-lipoxygenase and cyclooxygenase-2 in colon cancer growth induced by4-(N-methyl-N-nitrosamino)-1-(3-pyridyl)-1-butanone. J Pharmacol Exp Ther. 2004;311:123–30.
- Wong HP, Yu L, Lam EK, Tai EK, Wu WK, Cho CH. Nicotine promotes colon tumor growth and angiogenesis through βadrenergic activation. Toxicol Sci. 2007;97:279–87.
- Renehan AG, Tyson M, Egger M, Heller RF, Zwahlen M. Bodymass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. Lancet. 2008;371: 569–78.
- 75.•• Ning Y, Wang L, Giovannucci EL. A quantitative analysis of body mass index and colorectal cancer: findings from 56 observational studies. Obes Rev. 2010;11:19–30. The publication is a meta-analysis of 56 studies, including 93,812 cases. The authors indicate that "BMI < 23.0 kg/m², BMI of 23.0-24.9, 25.0-27.4, 27.5-29.9 and > or = 30.0 kg/m² were associated with 14%, 19%, 24% and 41% increased risks of colorectal cancer, respectively." Additionally "the association was stronger for colon than rectal cancer, for men than women," These valuable data strongly emphasize the impact of obesity on CRC risk..
- Moghaddam AA, Woodward M, Huxley R. Obesity and risk of colorectal cancer: a meta-analysis of 31 studies with 70,000 events. Cancer Epidemiol Biomarkers Prev. 2007;16:2533

 47.
- 77. Nam SY, Kim BC, Han KS, et al. Abdominal visceral adipose tissue predicts risk of colorectal adenoma in both sexes. Clin Gastroenterol Hepatol. 2010;8:443–50. The authors evaluated the associations between waist circumference, visceral adipose tissue volume, and colorectal adenoma type and prevalence in 3, 922 participants who underwent colonoscopy and computer tomography. The analysis provided strong evidence that "the risk of colorectal adenomas increased with increasing visceral adipose tissue volume in both sexes. Waist circumference was associated with colorectal adenomas in men, but not in women. High volume of visceral adipose tissue (>or=1000 cm³) had a positive association with larger adenomas (>or=10 mm) and multiple

- adenomas." The results presented indicate that abdominal visceral adipose tissue volume is a better predictor of colorectal adenomas than BMI or waist circumference in men and women.
- Gutierez DA, Puglisi MJ, Hasty AH. Impact of increased adipose tissue mass on inflammation, insulin resistance, and dyslipidemia. Curr Diabetes Rep. 2009;9(1):26–32.
- Chavez JA, Summers SA. Lipid oversupply, selective insulin resistance, and lipotoxicity: molecular mechanisms. Biochim Biophys Acta. 1801;2010:252–65.
- Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. Nat Rev Cancer. 2004:4:579–91.
- Kumor A, Daniel P, Pietruczuk M, Malecka-Panas E. Serum leptin, adiponectin, and resistin concentration in colorectal adenoma and carcinoma (CC) patients. Int J Colorectal Dis. 2009;24: 275–81.
- Booth FW, Chakravarthy MV, Gordon SE, Spangenburg EE.
 Waging war on physical inactivity: using modern molecular ammunition against an ancient enemy. J Appl Physiol. 2002:93:3–30
- Wolin KY, Yan Y, Colditz GA, et al. Physical activity and colon cancer prevention: a meta-analysis. Br J Cancer. 2009; 100:611–6.
- 84. •• Wolin KY, Yan Y, Colditz GA. Physical activity and risk of colon adenoma: a meta-analysis. Br J Cancer. 2011;104(5):882–5. This is a meta-analysis of 20 studies evaluating the association between physical activity and the prevalence of colon adenomas. The authors showed that "a significant inverse association between physical activity and colon adenomas was found with an overall RR of 0.84 (CI: 0.77-0.92). The association was similar in men (RR=0.81, CI: 0.67-0.98) and women (RR=0.87, CI: 0.74-1.02). The association appeared slightly stronger in large/advanced polyps (RR=0.70, CI: 0.56-0.88)." The study strongly emphasizes the important role of physical activity in CRC prevention.
- Handschin C, Spiegelman BM. The role of exercise and PGC1α in inflammation and chronic disease. Nature. 2008;454:463–9.
- 86. Wiseman M. The second World Cancer Research Fund/American Institute for Cancer Research expert report. Food, nutrition, physical activity, and the prevention of cancer: a global perspective. Proc Nutr Soc. 2008;67:253–6.
- Asghar M, George L, Lokhandwala MF. Exercise decreases oxidative stress and inflammation restores renal dopamine D1 receptor function in old rats. Am J Physiol Renal Physiol. 2007;293:F914–9.
- Cirillo D, Rachiglio AM, la Montagna R, Giordano A, Normanno N. Leptin signaling in breast cancer: an overview. J Cell Biochem. 2008;105:956–64.
- Drevon CA. Fatty acids and expression of adipokines. Biochim Biophys Acta. 2005;1740:287–92.
- Kim SY, Jun TW, Lee YS, Na HK, Surh YJ, Song W. Effects of exercise on cyclooxygenase-2 expression and nuclear factor-κB DNA binding in human peripheral blood mononuclear cells. Ann N Y Acad Sci. 2009;1171:464–71.
- Poulsen HE, Weimann A, Loft S. Methods to detect DNA damage by free radicals: relation to exercise. Proc Nutr Soc. 1999;58: 1007–14.
- Ju J, Nolan B, Cheh M, Bose M, Lin Y, Wagner GC, et al. Voluntary exercise inhibits intestinal tumorigenesis in Apc^{Min/+} mice and azoxymethane/dextransulfate sodiumtreated mice. BMC Cancer. 2008;8:316. doi:10.1186/1471-2407-8-316.
- Hrushesky WJ, Lannin D, Haus E. Evidence for an ontogenetic basis for circadian coordination of cancer cell proliferation. J Natl Cancer Inst. 1998;90:1480

 –4.
- Matsuo T, Yamaguchi S, Mitsui S, Emi A, Shimoda F, Okamura H. Control mechanism of the circadian clock for timing of cell division in vivo. Science. 2003;302:255–9.



- Ko CH, Takahashi JS. Molecular components of the mammalian circadian clock. Hum Mol Genet. 2006;15: R271-7.
- Wood PA, Yang X, Taber A, Oh EY, Ansell C, Ayers SE, et al. Period 2 mutation accelerates Apc^{Min/+} tumorigenesis. Mol Cancer Res. 2008;6:1786–93.
- Wozniak K, Arabski M, Malecka-Panas E, Drzewoski J, Blasiak J. DNA damage in human colonic mucosa cells induced by bleomycin and the protective action of vitamin E. Cell Mol Biol Lett. 2003;9:31–45.
- Blasiak J, Arabski M, Pertynski T, Malecka-Panas E, et al. DNA damage in human colonic mucosa cells evoked by nickel and protective action of quecetin – involvement of free radicals? Cell Biol Toxicol. 2002;18:279–88.
- Flossmann E, Rothwell PM. Effect of aspirin on long-term risk of colorectal cancer: consistent evidence from randomized and observational studies. Lancet. 2007;369:1603–13.
- Liao X, Lochhead P, Nishihara R. Aspirin use, tumor PIK3CA mutation, and colorectal-cancer survival. N Engl J Med. 2012;367(17):1596–606.

