

Impact of Target-Mediated Elimination on the Dose and Regimen of Evolocumab, a Human Monoclonal Antibody Against Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9)

The Journal of Clinical Pharmacology
2017, 57(5) 616–626
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of Clinical Pharmacology published by
Wiley Periodicals, Inc. on behalf of
American College of Clinical Pharma-
cology
DOI: 10.1002/jcph.840

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Abstract

Understanding the pharmacokinetic (PK) and pharmacodynamic (PD) relationship of a therapeutic monoclonal antibody against proprotein convertase subtilisin/kexin type 9 (PCSK9) exhibiting target-mediated drug disposition (TMDD) is critical for selecting optimal dosing regimens. We describe the PK/PD relationship of evolocumab using a mathematical model that captures evolocumab binding and removal of unbound PCSK9 as well as reduction in circulating low-density lipoprotein cholesterol (LDL-C). Data were pooled from 2 clinical studies: a single-dose escalation study in healthy subjects (7–420 mg SC; n = 44) and a multiple-dose escalation study in statin-treated hypercholesterolemic patients (14 mg weekly to 420 mg monthly [QM] SC; n = 57). A TMDD model described the time course of unbound evolocumab concentrations and removal of unbound PCSK9. The estimated linear clearance and volume of evolocumab were 0.256 L/day and 2.66 L, respectively, consistent with other monoclonal antibodies. The time course of LDL-C reduction was described by an indirect response model with the elimination rate of LDL-C being modulated by unbound PCSK9. The concentration of unbound PCSK9 associated with half-maximal inhibition (IC₅₀) of LDL-C elimination was 1.46 nM. Based on simulations, 140 mg every 2 weeks (Q2W) and 420 mg QM were predicted to achieve a similar time-averaged effect of 69% reduction in LDL-C in patients on statin therapy, suggesting that an approximate 3-fold dose increase is required for a 2-fold extension in the dosing interval. Evolocumab dosing regimens of 140 mg Q2W or 420 mg QM were predicted to result in comparable reductions in LDL-C over a monthly period, consistent with results from recently completed phase 3 studies.

Keywords

monoclonal antibody, pharmacokinetics, pharmacokinetic/pharmacodynamic, target-mediated drug disposition, PCSK9, LDL-C

Low-density lipoprotein cholesterol (LDL-C) lowering is a crucial component of the management of atherosclerotic cardiovascular disease (ASCVD) and has broadly proven benefits for primary and secondary prevention of ASCVD complications.¹ Low-density lipoprotein receptors (LDLR) on hepatocytes bind apolipoproteins B and E on circulating lipoprotein particles followed by internalization of the LDLR-lipoprotein complex. In the acidic environment of the endosome, LDL dissociates from the LDLR and is subsequently catabolized in lysosomes while the LDLR recycles back to the cell surface.² LDLR recycling is critical for the maintenance of cellular and whole-body cholesterol balance. Proprotein convertase subtilisin/kexin type 9 (PCSK9) is an enzymatically inactive serine protease predominantly secreted by hepatocytes. PCSK9 associates with LDLR and targets the internalized receptor for degradation, preventing recycling and thereby regulating serum LDL-C levels.³

Compelling human genetic data generated from studies of several thousand patients provide strong validation for the role of PCSK9 in modulating LDL-C levels and coronary heart disease (CHD) risk in humans.

PCSK9 “gain-of-function” mutations are associated with elevated LDL-C levels (eg, >300 mg/dL) and CHD risk, whereas “loss-of-function” (LOF) mutations are associated with lower LDL-C levels (eg, ≤100 mg/dL) and reduced CHD risk.^{4–10} Two adults with LOF mutations in both alleles have been reported.^{10,11} Despite having no detectable circulating PCSK9¹⁰

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Submitted for publication 18 May 2016; accepted 11 October 2016.

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and extremely low LDL-C levels (< 20 mg/dL),^{10,11} these individuals are otherwise healthy. Patients with heterozygous LOF mutations exhibit modestly lower serum levels of PCSK9 and as much as an 88% reduction in the incidence of CHD over a 15-year period relative to noncarriers of the mutations.⁶

Hydroxymethyl glutaryl coenzyme A reductase inhibitors, or statins, are currently the LDL-C-lowering treatment of choice for patients with increased cardiovascular risk. However, many patients either do not achieve their LDL-C targets with statins or are unable to tolerate statins at the dose recommended for their level of risk. Failure to achieve the LDL-C target may be due to high baseline LDL-C concentrations, diminished responsiveness, or an inability to tolerate adequate statin dosages.¹² Patients with inadequately controlled LDL-C values are at increased risk of future cardiovascular events.

Several studies indicate that statin therapy increases the levels of PCSK9.³ It is hypothesized that the cholesterol-lowering effect of statins may be attenuated by this increase in PCSK9 levels. Therefore, the use of a PCSK9 antagonizing therapy, alone or as an add-on to statin therapy, may be a particularly effective strategy to lower LDL-C, especially in high-risk patients and in patients who cannot tolerate high statin doses.

Evolocumab is a fully human monoclonal immunoglobulin IgG2 approved for use as an adjunct treatment for hyperlipidemia. Evolocumab binds to human PCSK9 with high affinity in vitro (dissociation constant [K_D] = 8.0 pM) and prevents its interaction with LDLR. Clinical studies have demonstrated robust reductions in LDL-C following administration of evolocumab in patients with primary hyperlipidemia and mixed dyslipidemia as monotherapy,¹³ in combination with a statin,¹⁴ in statin-intolerant patients,¹⁵ in patients with heterozygous familial hypercholesterolemia (HeFH),¹⁶ and in patients with homozygous familial hypercholesterolemia.¹⁷

This analysis described the pharmacokinetic (PK) and pharmacodynamic (PD) relationship of evolocumab using a mathematical model that captured evolocumab binding and removal of unbound PCSK9, and subsequent reduction in circulating LDL-C. This modeling exercise helps to understand the implications of target-mediated elimination on evolocumab dose and regimen selection, particularly given the higher PCSK9 concentrations observed in hypercholesterolemic patients treated with statins.

Methods

Study Participants

The model included results from a phase 1a study in healthy subjects (Study 20080397) and a phase 1b study in hypercholesterolemic patients stably treated

with statins (Study 20080398). Details of the study designs have been reported previously.¹⁸ The protocols and study procedures were approved by an institutional review board for each site.

Study Design

In the ascending single-dose phase 1a study, healthy subjects were randomized to a single dose of placebo or evolocumab ranging from 7 mg to 420 mg administered via subcutaneous (SC) injection. In the ascending multiple-dose phase 1b study, hypercholesterolemic patients receiving low- to moderate-dose statin therapy were randomized in 5 cohorts to SC placebo or evolocumab at doses of 14 mg once weekly (QW) \times 6 doses, 35 mg QW \times 6 doses, 140 mg once every 2 weeks (Q2W) \times 3 doses, 280 mg Q2W \times 3 doses, or 420 mg once monthly (QM; every 4 weeks) \times 2 doses. Two additional patient cohorts that received either high-dose statin therapy or had been diagnosed with HeFH received either SC placebo or evolocumab 140 mg Q2W \times 3 doses.

PK/PD Sampling and Assays

Venous blood samples were collected for PK and PD measurements after administration of placebo or evolocumab. All PD blood samples were obtained after overnight fasting (> 10 hours). The blood samples were collected into tubes containing no anticoagulant and allowed to clot at room temperature for 30 to 60 minutes. The samples were then centrifuged, and serum aliquots were prepared and stored at -80°C for unbound evolocumab and unbound PCSK9 measurements.

Unbound evolocumab was measured in serum using a validated enzyme-linked immunosorbent assay (ELISA). The ELISA was designed to measure unbound evolocumab in test samples by using highly specific anti-idiotypic antibodies for capture and detection of evolocumab. By virtue of binding to the antigen-combining site of evolocumab, the anti-idiotypic antibodies bound to evolocumab that was not bound to PCSK9. The assay limits of quantification ranged from 0.8 $\mu\text{g/mL}$ to 10 $\mu\text{g/mL}$. The validated procedure required accuracy (percentage difference from nominal concentration) and precision (percentage coefficient of variation) of $\pm 15\%$ and $\leq 15\%$, respectively, for the standard, and $\pm 20\%$ and $\leq 15\%$, respectively, for the QC samples.

Unbound PCSK9 was measured in serum using a qualified ELISA method.¹⁹ Measurements of PCSK9 quantified the unbound serum concentration by using evolocumab for capture and a second biotin-conjugated anti-PCSK9 antibody for detection. Thus PCSK9 that was bound to evolocumab was not measured. Standard and QC samples were prepared in 100% fetal

bovine serum because of the high and variable levels of endogenous PCSK9 in human serum. The QC samples in each run were 45.0, 140, and 500 ng/mL PCSK9 in fetal bovine serum. The assay lower and upper limits of quantification were 15 ng/mL and 1200 ng/mL, respectively. The validated procedure required accuracy (percentage difference from nominal concentration) and precision (percentage coefficient of variation) of $\pm 15\%$ and $\leq 20\%$, respectively, for the standard and $\pm 25\%$ and $\leq 20\%$, respectively, for the QC samples. LDL-C was measured using a homogeneous direct assay after overnight fasting.

Data Analysis

Maximum observed concentration (C_{max}) and area under the serum concentration curve from time zero to the time of the last quantifiable concentration (AUC_{last}) were determined using noncompartmental methods. Population PK/PD data were analyzed using the nonlinear mixed-effects modeling software program NONMEM (version 7.2)²⁰ on the NONMEM High Performance Cluster. A simultaneous approach to the PK/PD analysis was undertaken using the stochastic approximation expectation-maximization and importance sampling estimation methods sequentially. The M3 method was used for unbound evolocumab and un-

bound PCSK9 to simultaneously model observations as either continuous or categorical data when observations were above or below the limit of quantification, respectively.^{21–23} All statistical analyses were performed using either TIBCO Spotfire S+ for Windows version 8.2 (TIBCO Software Inc, Palo Alto, California) or R software 2.10.1 or higher (The R Foundation for Statistical Computing, Vienna, Austria).

The target-mediated drug disposition (TMDD) model was used to analyze concentration-time data using a previously described model framework.^{24,25} Unbound evolocumab and unbound PCSK9 were analyzed using the steady-state approximation of the TMDD model that incorporated binding between evolocumab and PCSK9, turnover of PCSK9, and elimination of drug-target complex, as shown in Figure 1.²⁶ Equations used for the model are shown in equations 1 to 3:

$$\frac{dA_{\text{depot}}}{dt} = -k_a \cdot A_{\text{depot}} \quad (1)$$

$$\frac{dTDA}{dt} = k_a \cdot A_{\text{depot}} - k \cdot \text{FDC} \cdot V - \frac{k_{\text{int}} \cdot \text{TLC} \cdot \text{FDC} \cdot V}{k_{\text{ss}} + \text{FDC}} \quad (2)$$

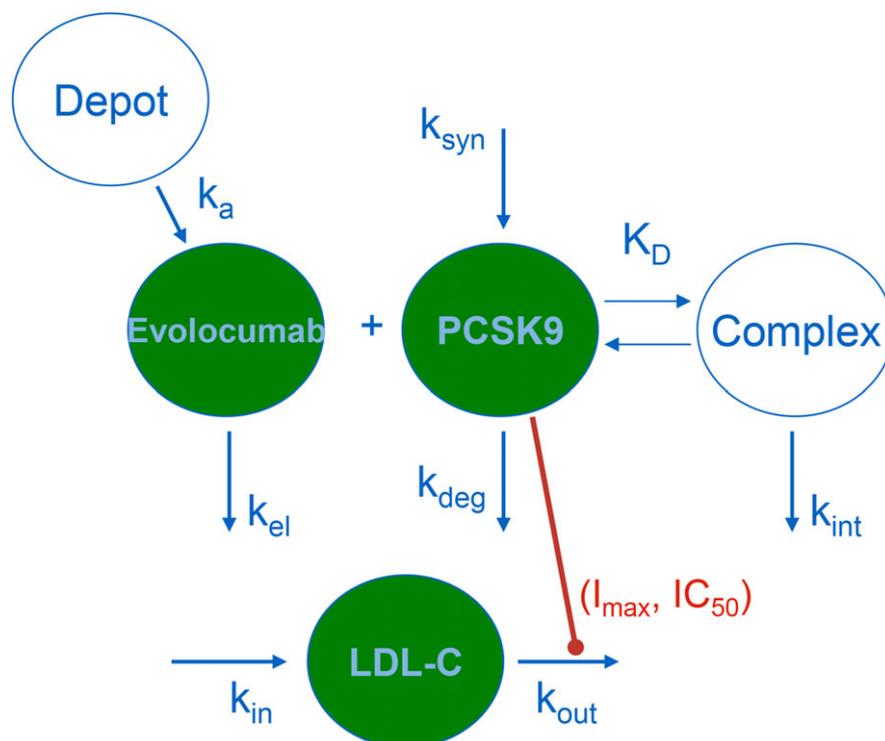


Figure 1. Schematic of the PK/PD model structure. Abbreviations: I_{max} , maximal inhibition; IC_{50} , PCSK9 concentration associated with half-maximal inhibition; k_a , absorption rate constant; k_{el} , elimination rate constant; k_{syn} , PCSK9 production rate constant; k_{deg} , PCSK9 elimination rate constant; K_D , dissociation constant; k_{int} , evolocumab-PCSK9 complex elimination rate constant; k_{in} , LDL-C production rate constant; k_{out} , LDL-C elimination rate constant; PCSK9, proprotein convertase subtilisin/kexin type 9.

$$\frac{dTLC}{dt} = k_{syn} - k_{deg} \cdot TLC - \frac{(k_{int} - k_{deg}) \cdot FDC \cdot TLC}{(k_{ss} + FDC)} \quad (3)$$

These equations described the PK of evolocumab in the depot (A_{depot}) after SC administration, the total drug amount in the central compartment (TDA), and the total ligand concentration (TLC), respectively. A 1-compartment open model with linear elimination from the central compartment was parameterized by volume of distribution in the central compartment (V) and drug clearance (CL). Absorption after SC administration was described by a first-order process (k_a) from the depot compartment to the central compartment. Bioavailability (F) was fixed to 72% (Amgen internal data). The total drug concentration (TDC; nM) was calculated as shown in equation 4:

$$TDC = \frac{TDA}{V} \quad (4)$$

A zero-order production rate constant (k_{syn}) and a first-order degradation rate constant (k_{deg}) described the production and degradation of PCSK9, related to the baseline PCSK9 ($BASE_{PCSK9}$) level as k_{syn}/k_{deg} . The quasi-steady-state approximation of the full TMDD model was selected.^{25,27} In this model approximation, binding of evolocumab and PCSK9 to form a complex was described by the steady-state constant (k_{ss}), which represented the ratio of the sum of the complex internalization rate (k_{int} , here representing the elimination of complex) and dissociation rate constant (k_{off}) to the drug-target association rate constant (k_{on}), where the K_D was defined as the ratio of k_{off}/k_{on} .

The free drug concentration (FDC; nM) was determined from TLC in equation 5:

$$FDC = \frac{1}{2} \left[(TDC - TLC - k_{ss}) + \sqrt{(TDC - TLC - k_{ss})^2 + 4k_{ss} TDC} \right] \quad (5)$$

The free ligand concentration (FLC; nM) was calculated as shown in equation 6:

$$FLC = TLC - (TDC - FDC) \quad (6)$$

Total drug or ligand referred to the sum of bound and free species under the assumption of one-to-one stoichiometry of binding between drug and target.

An indirect response model was used to describe the effect of PCSK9 on LDL-C removal,^{28,29} consistent

with PCSK9's role in the recycling of LDLR as shown in equation 7:

$$\frac{dLDL}{dt} = k_{in} - k_{out} \cdot \left(1 - \frac{I_{max} \cdot FLC}{IC_{50} + FLC} \right) \cdot LDL \quad (7)$$

The model included parameters for the zero-order production rate constant (k_{in}) and the first-order degradation rate constant (k_{out}) of LDL-C. The maximal k_{out} was the rate of LDL-C degradation in the absence of PCSK9. The ability of PCSK9 to modulate k_{out} was estimated with the potency parameters IC_{50} (the serum unbound PCSK9 concentration associated with 50% of the maximal k_{out}) and I_{max} (the theoretical maximal proportional change in k_{out} due to PCSK9).

Between-subject variability (BSV) was modeled as log-normally distributed. BSV for k_{deg} and k_{int} were fixed at 0%. Proportional error models described residual error for unbound evolocumab and unbound PCSK9. A proportional and additive error model described residual error for LDL-C.

The assessment of model adequacy and decisions about increasing model complexity were driven by the data and guided by goodness-of-fit criteria including (1) visual inspection of diagnostic scatter plots (predicted vs observed concentration), histograms of individual random effects, and visual predictive checks³⁰; (2) precision of parameter estimates; and (3) objective function value (OFV) and the stability of the objective function. A decrease in OFV of 10.8 points, corresponding to a χ^2 P -value of 0.001 with 1 degree of freedom, was considered significant. Model evaluation also included prediction-corrected visual predictive checks (VPC).³¹ PK, PCSK9, and LDL-C observations were placed in time bins corresponding to the nominal collection times listed in the study protocol: 4, 8, 12, or 24 hours postdose on day 1 and on days 3, 4, 5, 6, 7, 8, 11, 15, 22, 29, 36, 43, 57, 71, 85. Observations were assigned to bins by determining the closest nominal time postdose corresponding to the actual time postdose.

Simulations

Simulations were performed with the final PK/PD model using Berkeley Madonna Version 8.3.14.³² The time course of LDL-C response was simulated following repeated SC administration of evolocumab at 140 mg Q2W, 280 mg QM, and 420 mg QM for 12 weeks in a patient population stably treated with statins with the predicted baseline PCSK9 in the population PK/PD model (5.27 nM [379 ng/mL]).

Results

Baseline characteristics are shown in Table S1. A total of 73 patients were treated with evolocumab, and 28

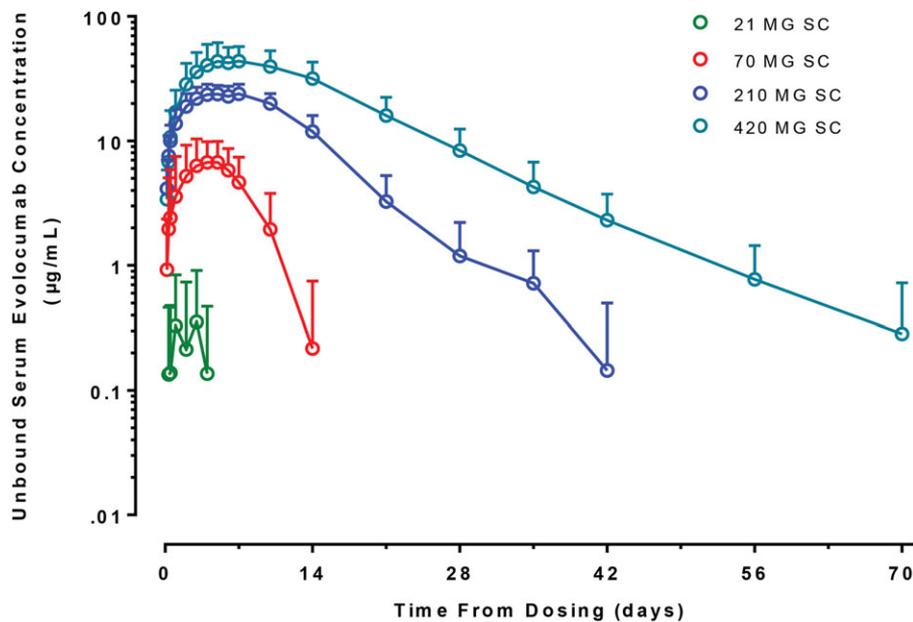


Figure 2. Observed unbound evolocumab concentration-time profiles after single administration to non-statin-treated healthy subjects. The open circles represent the arithmetic mean of the observed data, and the error bars represent the standard deviation.

Table 1. Summary of Unbound Evolocumab Pharmacokinetic Parameters After 140 mg SC Evolocumab Every 2 Weeks in Hypercholesterolemic Patients Treated With Low- to Moderate-Intensity or High-Intensity Statins

Parameter (Mean \pm SD)	Low-Moderate (n = 6)	High (n = 9)	Ratio (High/Low-Moderate)
Baseline PCSK9 (ng/mL)	385 \pm 88	487 \pm 214	1.26
C_{max} (μ g/mL)	20 \pm 13	16 \pm 11	0.80
AUC_{last} (day· μ g/mL)	226 \pm 249	181 \pm 157	0.80

Low-moderate: atorvastatin <40 mg, rosuvastatin <20 mg, simvastatin <40 mg, or simvastatin 80 mg. High: atorvastatin 80 mg or rosuvastatin 40 mg. AUC_{last} , area under the concentration-time curve from time zero to the last quantifiable concentration; C_{max} , maximum observed concentration; PCSK9, proprotein convertase subtilisin/kexin type 9.

patients received placebo, in the phase 1a and 1b studies. Most were male (73.3%) and white (83.2%), with a mean age and body weight of 45.5 years and 81.4 kg, respectively. The baseline PCSK9 was 1.58-fold higher in the phase 1b study relative to the phase 1a study due to the inclusion of hypercholesterolemic patients on stable statin therapy (> 97%) in the phase 1b study. Baseline LDL-C was similar between the evolocumab and placebo groups in both studies.

Figure 2 shows the observed mean \pm SD serum unbound evolocumab concentration-time profiles after a single SC administration of 21, 70, 210, or 420 mg evolocumab in healthy subjects. Unbound evolocumab concentrations were below the detection limit for all healthy subjects after a single 7 mg SC dose of evolocumab and were not included in the figure. Treatment with a single SC administration of evolocumab resulted in nonlinear elimination of unbound evolocumab over the dose range studied in healthy subjects. The terminal half-life of unbound evolocumab was dose dependent with the longest half-life observed after 420 mg SC evolocumab.

Unbound evolocumab noncompartmental PK parameters following 140 mg Q2W in hypercholesterolemic patients treated with low- to moderate-intensity statins or high-intensity statins are shown in Table 1. A 26% increase in baseline PCSK9 and a 20% decrease in unbound evolocumab C_{max} and AUC_{last} were observed in patients on high-intensity statins compared with patients on low- to moderate-intensity statins, consistent with target-mediated elimination of evolocumab.

Following a single SC administration of evolocumab, rapid decreases in unbound PCSK9 were observed within 4 hours and resulted in > 80% suppression of unbound PCSK9 after doses \geq 21 mg SC evolocumab. Increasing the dose of evolocumab increased the duration of full suppression of unbound PCSK9 up to a maximum of 14 days with the highest dose evaluated of 420 mg SC evolocumab. On elimination of unbound evolocumab from serum, unbound PCSK9 concentrations gradually returned toward the baseline without evidence of rebound above the initial baseline values.

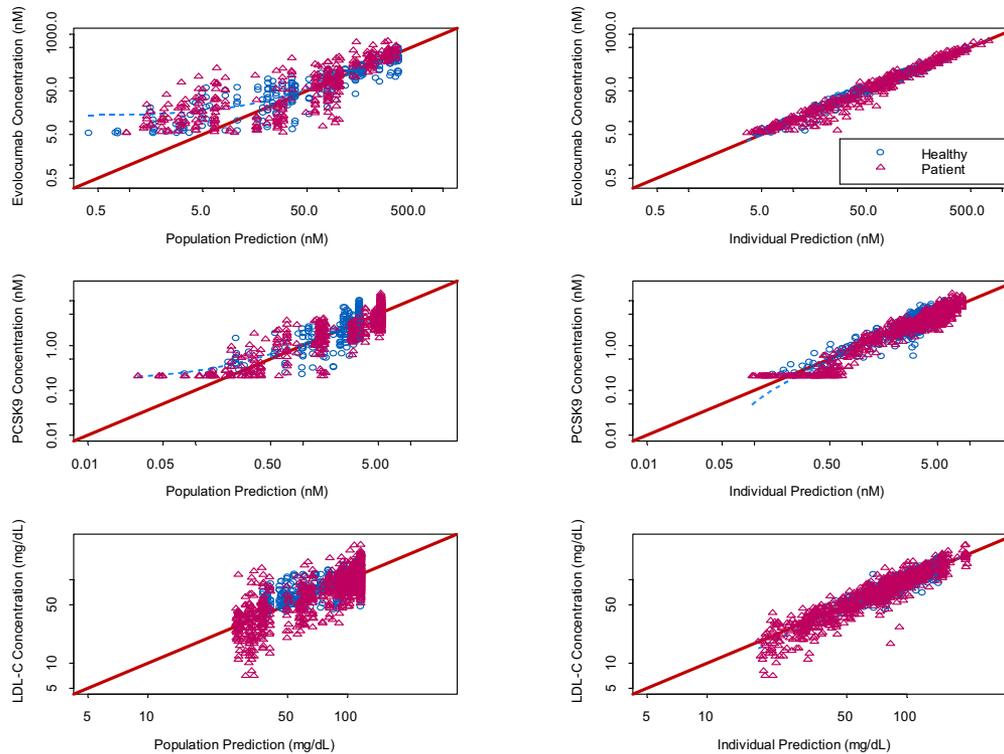


Figure 3. Diagnostic plots of model-predicted and observed unbound evolocumab concentrations, unbound PCSK9 concentrations, and LDL-C.

Table 2. Population Pharmacokinetic Parameter Estimates for Evolocumab Using a Target-Mediated Drug Disposition Model

Parameter	Units	Fixed Effects: Population Mean Parameter		Random Effects: Intersubject/Residual Variance	
		Estimate	SE (%RSE)	Estimate (~CV%)	SE (%RSE)
k_a	l/day	0.245	0.0290 (12)	0.807 (90)	0.157 (19)
V	L	2.66	0.156 (5.9)	0.158 (40)	0.0362 (23)
CL	L/day	0.256	0.0439 (17)	0.924 (96)	0.260 (28)
k_{deg}	l/day	2.12	0.156 (7.3)	0.00 ^a	—
BASE _{PCSK9}	nM	5.27	0.141 (2.7)	0.0348 (19)	0.00583 (17)
k_{ss}	nM	0.253	0.0579 (23)	1.17 (108)	0.233 (20)
k_{int}	l/day	0.0529	0.00184 (3.5)	0.00 ^a	—
θ^1	—	0.637	0.0259 (4.1)	0.00 ^a	—
Covariance ($k_a - V$)	—	—	—	0.253 (70)	0.0650 (26)
Covariance ($k_a - CL$)	—	—	—	-0.671 (-78)	0.182 (27)
Covariance ($V - CL$)	—	—	—	-0.183 (-48)	0.0775 (42)
σ^2 (evolocumab)	—	—	—	0.0576 (24)	0.00994 (4.1)
σ^2 (PCSK9)	—	—	—	0.0942 (31)	0.00625 (2.0)

CV%, coefficient of variation calculated as $\sqrt{\text{var}} \cdot 100$; %RSE, relative standard error of the estimate calculated as $(\text{SE}/\text{Estimate}) \cdot 100$; SE, standard error of the estimate; k_a , absorption rate constant; V, volume of distribution; CL, clearance; k_{deg} , PCSK9 degradation rate constant; BASE_{PCSK9}, baseline PCSK9; k_{ss} , steady-state constant; k_{int} , complex elimination rate constant; θ^1 , fold change in baseline PCSK9 for healthy subjects vs statin-treated patients.

^aIntersubject random variance was fixed at 0 in the pharmacokinetic model.

Based on the nonlinear elimination profile of evolocumab and consequent reductions in unbound PCSK9, a TMDD model was selected to describe evolocumab PK and PD (Figure 1). The data set used in the analysis included 101 individuals and 4910 observations including unbound evolocumab, unbound PCSK9, and LDL-C (Figure 3). Overall, there was agreement between observed and predicted values for

the 3 PK and PD endpoints of unbound evolocumab, unbound PCSK9, and LDL-C for both population and individual predictions. The diagnostic plots indicated that the TMDD model provided an adequate description of the data.

Population PK parameter estimates for the TMDD model of evolocumab and unbound PCSK9 are presented in Table 2. The final TMDD model provided

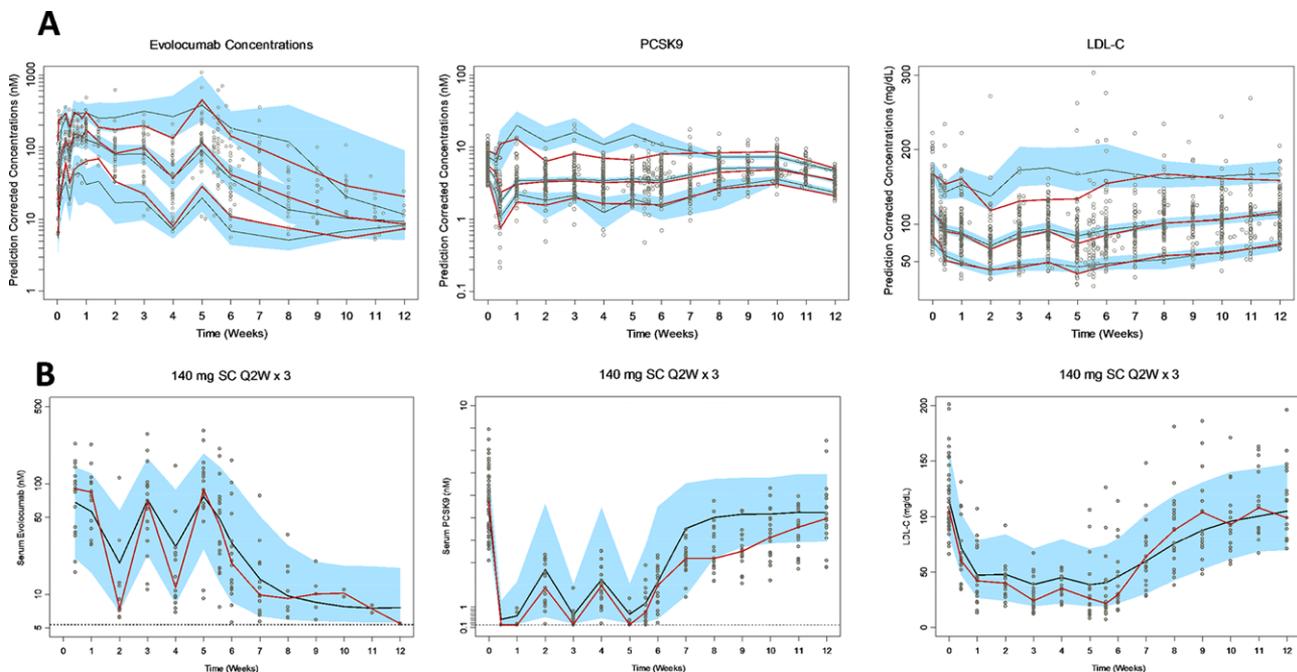


Figure 4. (A) Prediction-corrected visual predictive check. The red and black lines represent the median and the 5th and 95th percentiles of observed and predicted data, respectively. The blue-shaded region represents the 95% confidence interval of the respective median and 5th and 95th percentiles of predicted data. (B) Visual predictive check of 140 mg SC evolocumab Q2W \times 3, median \pm 90% prediction interval of simulated pharmacokinetics and pharmacodynamics (solid black line and blue-shaded region, respectively, for $n = 100$ simulations) and observed data (open circles). Red line represents the observed median.

Table 3. Population Pharmacokinetic/Pharmacodynamic Parameter Estimates for Evolocumab Using an Indirect Response Model for LDL-C

Parameter	Units	Fixed Effects: Population Mean Parameter		Random Effects: Intersubject/Residual Variance	
		Estimate	SE (%RSE)	Estimate (\sim CV%)	SE (%RSE)
k_{out}	1/day	0.305	0.0121 (4.0)	0.00 ^a	—
$BASE_{LDL-C}$	mg/dL	116	3.36 (2.9)	0.0465 (22)	0.00698 (15)
I_{max}	—	1.00 (fix)	—	0.00 ^a	—
IC_{50}	nM	1.46	0.152 (10)	0.481 (69)	0.106 (22)
σ^2 (proportional error)	—	—	—	0.0130 (11)	0.00505 (4.4)
σ^2 (additive error)	—	—	—	60.1 (7.8) ^b	0.519 (6.7)

CV%, coefficient of variation calculated as $\sqrt{\text{var}} \cdot 100$; %RSE, relative standard error of the estimate calculated as $(\text{SE}/\text{Estimate}) \cdot 100$; SE, standard error of the estimate; k_{out} , elimination rate constant for LDL-C; $BASE_{LDL-C}$, baseline LDL-C; I_{max} , maximal inhibition; IC_{50} , concentration associated with half-maximal inhibition.

^aIntersubject random variance was fixed at 0 in the pharmacokinetic/pharmacodynamic model.

^bRepresents the standard deviation and calculated as $\sqrt{\text{var}}$.

reasonably precise estimates of the structural parameters ($\leq 23\%$ relative standard error [RSE]) of all the fixed effect parameters. The residual variabilities for unbound evolocumab and unbound PCSK9 were 24% and 31%, respectively. The intersubject variabilities (%CV) for k_a , V , CL , $BASE_{PCSK9}$, and k_{ss} were 90%, 40%, 96%, 19%, and 108%, respectively. A covariate indicated that the baseline PCSK9 was 3.36 nM (or 242 ng/mL) in healthy subjects and 5.27 nM (or 379 ng/mL) in hypercholesterolemic patients stably treated with statins. The prediction-corrected VPC for all doses and the VPC for 140 mg Q2W indicated good agreement between the simulations and the observed data for unbound evolocumab and unbound PCSK9 (Figure 4).

An indirect response model described the turnover of LDL-C in which PCSK9 inhibited the elimination of LDL-C, such that decreasing PCSK9 levels due to evolocumab administration led to increased elimination of LDL-C, thereby decreasing serum LDL-C levels (Figure 1). The final PK/PD model described the time course of LDL-C. The final model provided reasonably precise estimates of the structural parameters ($\leq 10\%$ RSE) of all the fixed effect parameters, with proportional and additive residual variability of 11% and 7.8 mg/dL, respectively (Table 3). The intersubject variabilities for $BASE_{LDL-C}$ and IC_{50} were 22% and 69%, respectively. Initial attempts to estimate I_{max} suggested that it was near the boundary of 1.00, so its value was

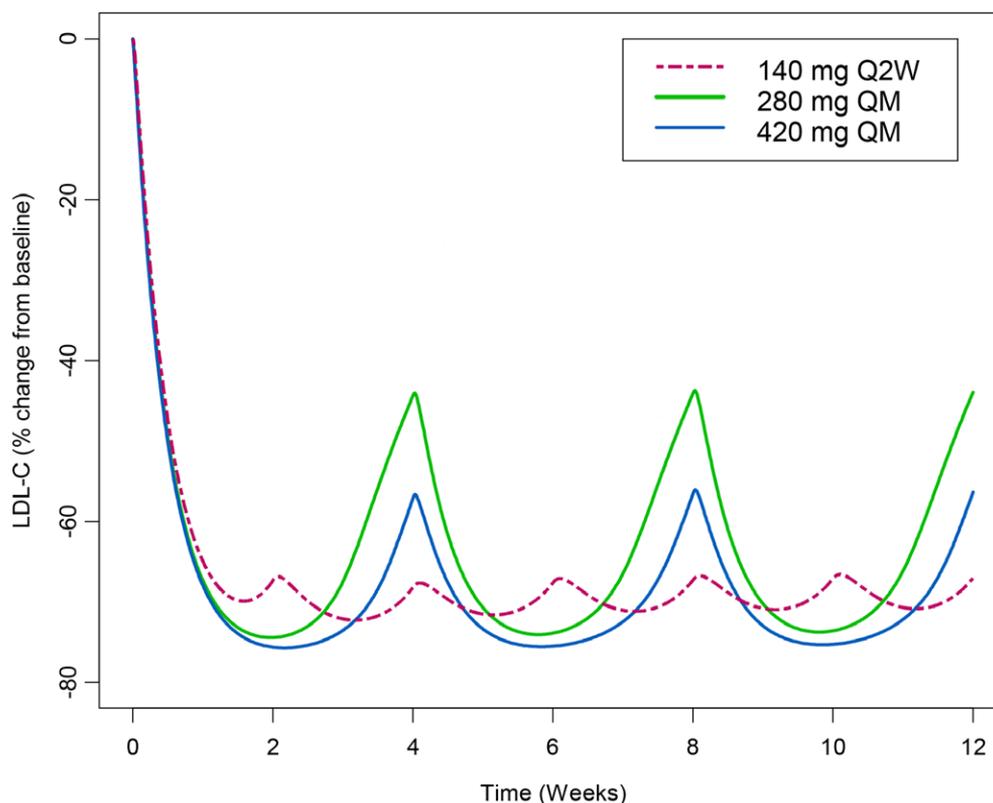


Figure 5. Model-predicted time course of LDL-C after multiple SC evolocumab doses.

fixed to 1.00. The mean (\pm standard error of the estimate) value of IC_{50} for inhibition of the elimination of LDL-C was 1.46 ± 0.152 nM. The prediction-corrected VPC for all doses and the VPC for 140 mg Q2W indicated good agreement between the simulations and the observed data for LDL-C (Figure 4).

Based on the final PK/PD model, simulations were performed to investigate the time course of LDL-C response after 140 mg SC Q2W, 280 mg SC QM, and 420 mg SC QM evolocumab in patients treated with stable statins (Figure 5). The simulations indicated that doubling the evolocumab dose from 140 mg SC Q2W to 280 mg SC QM to extend the dosing interval did not adequately maintain the reductions in LDL-C over the entire monthly dosing interval from weeks 8 to 12 after LDL-C reductions reached steady state. The time-averaged effects in the area under the LDL-C effect curve based on the simulations for evolocumab doses of 140 mg Q2W, 280 mg QM, and 420 mg QM were 68.9%, 63.5%, and 68.9%, respectively. Therefore, based on simulations from the PK/PD model, an approximate 3-fold increase in the dose to 420 mg SC QM evolocumab appeared to be required to maintain stable LDL-C reductions observed after 140 mg SC Q2W and to limit fluctuations in LDL-C over the dosing interval.

Discussion

Knowledge of the PK/PD relationship including the onset and offset of response is critical to defining optimal doses and regimens for novel therapeutics in different patient populations. Simulations based on the PK/PD relationship among unbound evolocumab, unbound PCSK9, and LDL-C following evolocumab administration were used to help support dose and regimen selection for clinical studies. The model was based on intensive, longitudinal data collected in 101 individuals (44 healthy subjects and 57 hypercholesterolemic patients treated with statins), including data from single administration or repeated dosing of evolocumab for 2-months. This PK/PD analysis leveraged the target-mediated interaction between evolocumab and PCSK9, and the impact on LDL-C, to evaluate the dose increment required to maintain maximal reduction in LDL-C while extending the dosing interval from Q2W to QM.

Empirical approaches to posology would assume that doubling the dose would be sufficient to extend the drug effect from 2 weeks to 4 weeks. However, given the nonlinear PK of evolocumab due to TMDD and the nonlinear PK/PD relationship between PCSK9 and LDL-C, this simplification was inappropriate for a monoclonal antibody directed against PCSK9. A

3-fold increase in the dose of evolocumab from 140 mg to 420 mg was required to obtain similar time-averaged reductions in LDL-C when the dosing interval was extended from Q2W to QM. Both doses were associated with more than 5% greater time-averaged reduction of LDL-C compared with the 280-mg QM dose of evolocumab. For statins, a similar difference (approximately 4% to 6%) in LDL-C reduction between lower-intensity and higher-intensity therapy has been used to support high-dose statin therapy in clinical practice.³³

The TMDD model and its approximations have been applied to describe nonlinear PK and reflect the influence of the target on drug disposition.^{24,25,27} Concentration-dependent binding between drug and target and subsequent elimination of the drug over the endogenous elimination pathways such as catabolism by the reticuloendothelial system. Here, availability of serum unbound PCSK9 concentrations facilitated model development and provided additional confirmation of the mechanism of action. The TMDD model parameters gave an indication of the extent and duration of PCSK9 suppression required to maintain consistent reduction in LDL-C. The linear PK parameters (ie, CL and V) for evolocumab were similar to those for other IgG antibodies.³⁴ Similarly, the k_{ss} value was within the reported range for TMDD models of other drugs and their targets. In comparing the *in vivo* k_{ss} to the *in vitro* K_D , k_{ss} is always greater than K_D , but it may be within the same range or up to 30 times greater.²⁶

PCSK9 may facilitate LDLR degradation via 2 routes: (1) intracellular endosomal association with LDLR and (2) internalization of secreted PCSK9 following cell surface association with LDLR.³⁵⁻³⁸ Intracellular cholesterol regulates the synthesis of PCSK9 and LDLR through the sterol regulatory element binding proteins known as SREBPs.³ In addition, clearance of PCSK9 is primarily mediated through its association with LDLR.³ The estimated k_{deg} of PCSK9 in this study was 2.12 day^{-1} , suggesting an elimination half-life of approximately 8 hours, consistent with the short half-life of injected recombinant PCSK9 observed after administration to mice.^{36,39} The higher baseline PCSK9 observed in patients treated with statins compared to healthy subjects with a similar baseline LDL-C was consistent with other reports.^{40,41} In addition, in hypercholesterolemic patients with a higher baseline PCSK9 due to high-intensity statin therapy, there was a 20% reduction in unbound evolocumab exposure. Thus, it is plausible that the high-turnover soluble target PCSK9 represents a sink for exogenous therapeutic proteins directed against PCSK9.

The indirect response model has been applied to capture the PD of drugs acting on LDL-C including treat-

ment with statins in hypercholesterolemic patients^{42,43} and with corticosteroids in rats.⁴⁴ Dietary cholesterol consumption and *de novo* hepatic cholesterol synthesis regulate production of very low density lipoprotein cholesterol and ultimately LDL-C, and internalization by hepatic LDLR is the predominant elimination mechanism for circulating LDL-C.⁴⁵ Here, the production and elimination mechanisms were represented as system parameters (k_{in} and k_{out} , respectively) in the indirect response model, with PCSK9 regulating k_{out} according to its known role in regulating LDLR expression. The maximal elimination rate (k_{out}) gave an estimate of the time to achieve a new equilibrium in the absence of PCSK9, taking into account the half-life of LDL-C. The estimated maximal k_{out} for LDL-C was 0.305 day^{-1} , suggesting an approximate half-life of approximately 2.3 days for the elimination of LDL-C in humans in the absence of PCSK9. Previously reported values of k_{out} in healthy subjects and patients treated with statins ranged from 0.105 to 0.350 day^{-1} .^{42,45,46} It is necessary to account for these delays in the onset and offset of LDL-C response to accurately estimate LDL-C reduction after administration of a monoclonal antibody such as evolocumab.

The model described in this report represented a simplified version of the biology of cholesterol metabolism as well as interactions between the drug and PCSK9 and between PCSK9 and rates of LDL-C elimination. This model did not include other known endogenous binding partners for PCSK9 such as LDL-C and LDLR.^{36,47} As mentioned, the TMDD model assumed there was no competition for the drug binding to its target. Exploration of a more complex physiologic model may help to better understand these interactions. In addition, we performed simulations using the typical values of PK/PD parameters using Berkley Madonna. Thus, the impact of dose regimen on interindividual variability in the response was not evaluated.

Conclusions

Based on simulations that included PCSK9-mediated nonlinear evolocumab elimination, 140 mg Q2W and 420 mg QM were predicted to achieve similar LDL-C responses, suggesting that an approximate 3-fold dose increase was required for a 2-fold extension in the dosing interval. Evolocumab 140 mg Q2W and 420 mg QM resulted in comparable reductions in LDL-C over a monthly period. Conclusions from modeling and simulation exercises using phase 1 study data were consistent with results of phase 3 studies,^{13,14} demonstrating the value of early characterization of PK/PD relationships to help inform dose and regimen selection.

Acknowledgments

We thank the volunteer participants and clinical teams that conducted the studies. Additionally, we would like to thank Alex Colbert for his bioanalytical support. Meera Kodukulla (Amgen Inc) and Jonathan Latham (on behalf of Amgen Inc) assisted the authors with the preparation and submission of this work.

Funding

This study was sponsored by Amgen, Inc. Meera Kodukulla of Amgen and Jonathan Latham of PharmaScribe, LLC (on behalf of Amgen) assisted the authors with preparation of the manuscript.

Declaration of Conflicting Interests

J.P.G., S.D., M.K., A.G., M.G.E., M.D., M.A.G., R.S., and S.M.W. were employees and stockholders of Amgen, Inc at the time this work was conducted. D.B. received honoraria from Amgen Inc, Sanofi-Aventis, Merck Sharp & Dohme, Pharma Dynamics, AstraZeneca, Pfizer, and Aegerion; and was a consultant to board: Merck Sharp & Dohme, Sanofi-Aventis, Gemphire, and Aegerion.

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Supporting Information

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