



Review

The Effects of Physical Exercise on Saliva Composition: A Comprehensive Review

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Abstract: Saliva consists of organic and inorganic constituents. During exercise, analysis of the saliva can provide valuable information regarding training stress, adaptation and exercise performance. The objective of the present article was to review the effect of physical exercise on saliva composition. The shift in the composition of the saliva, during and after a workout, reflects the benefits of exercise, its potential risks and the capability of the saliva to serve as a health indicator. The type and the frequency of training, the physical condition and the athletes' general health influence the hormones, immunoglobulins and saliva enzymes. The correlation between saliva and physical exercise has to be further investigated and the available knowledge to be applied for the benefit of the athletes during sports activities.

Keywords: oral health; saliva; sports dentistry; physical exercise; sports medicine



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1. Introduction

Salivary glands are exocrine organs that produce a large amount of fluid. Through the saliva, electrolytes and other substances are transferred from the inner of the glands to the oral cavity. The mean volume of the fluid is estimated to be 750 mL/day, which almost represents 20% of the overall plasma volume [1,2]. Apart from the major salivary glands (2–5 mL/min), saliva is also secreted by several minor glands at a rate of 0.5 mL/min [3].

Salivary glands are stimulated by parasympathetic cholinergic nerves and sympathetic adrenergic nerves [4]. Parasympathetic stimulation increases regional blood flow and saliva, consisting of low organic and inorganic components [5]. Sympathetic stimulation results in the saliva of low volume, containing high levels of K⁺ and proteins [6]. The autonomous nervous system regulates salivary secretion. Catecholamines might also play a role in the secretion of electrolytes and proteins [7]. During exercise, hormone response analysis can provide valuable information regarding training stress, adaptation, dehydration and exercise performance [8].

Saliva comprises 99% water and 1% organic and inorganic constituents [9,10]. Although saliva's organic and inorganic components are usually present in low concentration, compared with the serum, some proteins such as α-amylase are synthesized in the glands and presented in higher levels [11–13]. Other organic components, which can be detected in the saliva are vitamin C, maltase, urea, uric acid, albumin, mucin, creatinine, amino acids, lactase and hormones such as testosterone, cortisol, etc. Moreover, amounts of CO₂ are presented and so are immunoglobulins such as IgA, IgG, IgM [14].

The objective of the present article was to review the effect of physical exercise on saliva composition based on clinical studies.

2. Materials and Methods

The review focuses on clinical trials, directly investigating the effects of physical exercise or various sports on the composition of the saliva. A search was conducted in the PubMed and Scopus databases, implementing a Boolean strategy, to identify eligible articles. A literature search until September 2021 was performed, for articles written in English, without any restriction upon publication date. The search strategy included the following terms: (“physical exercise” OR “sports” OR “athletes”) AND (“saliva” or “salivary”). The references of the selected articles were also evaluated to identify themes that may fail to be detected by the search mentioned above strategy.

3. Results

The search strategy revealed 4487 articles. Following the removal of duplicates, 2498 articles were remained. Based on the title and the abstract of the studies, 2,289 articles were removed. After the evaluation of the remaining articles against the eligibility criteria, a total of 125 articles were concluded in the present review. Physical exercise has a significant impact on the composition of saliva. Therefore, the data which were extracted from the selected studies are presented individually for each substance of the saliva, in the following categories: salivary secretion, lysozyme, lactoferrin, lactate, oral peroxides, nitric oxide, salivary A-amylase (sAA), salivary cortisol (S-Cortisol), steroids-testosterone, salivary immunoglobulin A (IgA), immunoglobulin G (IgG), immunoglobulin M (IgM), insulin-like growth factor 1 (s-IGF-1), salivary MicroRNAs and melatonin. In Table 1, the available studies for each of the components of the saliva, along with their primary outcomes are presented.

Table 1. Primary outcomes of the included studies, separately for each investigated parameter of the saliva.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|-------------------------------|-------------------------------|------------------|--|---|
| A-amylase (sAA) | Yasuda N [15] | 2021 | 11 Males cycling | Increase in A-amylase activity after moderately long-lasting exercise, regardless of exogenous carbohydrate availability |
| | Wunsch K [16] | 2019 | 42 Males Acute exercise (24.1 ± 3) 42 Males placebo exercise (23.8 ± 2.3) | Increase in A-amylase concentration after moderate-to-high ergometer cycling. No difference in A-amylase peak level between habitual and acute exercise. |
| | Allgrove J [17] | 2008 | 10 Males (23) | Increase A-amylase concentration after exercise, followed by a return to pre-existing values 1 h post-exercise. (Cycling) An increase in s-IgA was independent of exercise intensity. |
| | Li TL [18] | 2004 | 8 Males (28.9 ± 1.8) | Increase in A-amylase activity after exercise (cycling, 60% VO max, 2 h) |
| | Walsh NP [19] | 1999 | 8 Well trained males (25 ± 1) | Decrease in A-amylase concentration immediately after exercise. (cycling) |
| | Chatterton R [20] | 1996 | 47 Medical students | Increase in A-amylase concentration. After exercise |
| Cortisol (S-cortisol) | Hough J [21] | 2021 | 23 Active males, cycle ergometer (21 ± 3) | Increase in the s-cortisol immediately after the exercise. Decrease in the s-cortisol 30 min post-exercise |
| | Ushiki K [22] | 2020 | 54 Participants (22) | Different rates of change in s-cortisol, depending on the intensity of the exercise. Decrease of s-cortisol levels after morning exercise. Increase of s-cortisol levels after afternoon exercise. |
| | Pearlmutter P [23] | 2020 | 22 Athletes 26 Non-athletes (22 ± 3) | Decrease in s-cortisol concentration after exercise. Workout intensity affected changes among athletes and non-athletes. In non-athletes, the s-cortisol concentration decreased more compared to athletes. |

Table 1. Cont.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|-------------------------------|-------------------------------|------------------|--|---|
| | Rahman MS [24] | 2019 | 945 Participants | Reduced s-cortisol levels after 12 weeks of physical exercise |
| | Wunsch K [16] | 2019 | 42 Males Acute exercise (24.1 ± 3) 42 Males placebo exercise (23.8 ± 2.3) | Increase in s-cortisol concentration after moderate-to-high ergometer cycling. Habitual exercise can reduce s-cortisol peak concentrations. |
| | Wood CJ [25] | 2018 | 164 Females (20 ± 3.4) | Increase in s-cortisol levels until 10 min of walking. The decline is s-cortisol levels after 10 min of walking. Participants with a more excellent fitness presented lower s-cortisol levels during walking. |
| | Crewther BT [26] | 2013 | 14 Rugby players (23.3 ± 3.5) | Decrease in salivary cortisol before games. |
| | Gillum T [27] | 2013 | 14 Marathon runners (43.7 ± 9.9) | Increase in s-cortisol concentration after exercise. |
| | Ida I [28] | 2013 | 18 Outpatients (45.9 ± 13.3) | Decrease in s-cortisol before, immediately after and ten minutes after the exercise session. |
| | He CS [29] | 2010 | 8 Basketball players (20.5 ± 0.3) | Increase in s-cortisol concentration during the intensive training and competition period |
| | Budde S [30] | 2010 | 40 High school students (14.35) | Increase in s-cortisol post-exercise. No effect of physical activity level on the s-cortisol before and after exercise. (Sprinting) |
| | Allgrove J [31] | 2009 | 16 Active adults (22 ± 4) | Increase in s-cortisol levels post-exercise. |
| | Thomas NE [32] | 2009 | 17 School children (15.5 ± 0.4) | Increase in salivary cortisol after exercise. (Cycling) |
| | Allgrove J [17] | 2008 | 10 Males (23) | No change in s-cortisol levels immediately after exercise. (Cycling) Increase in s-cortisol levels, 1 h after the training. Cortisol levels were higher in high-intensity training. |
| | Gozansky WS [33] | 2005 | 12 Participants (23–65) | Significant correlation of salivary cortisol with serum cortisol. |
| | Neary JP [34] | 2002 | 8 Physical education students Females (21 ± 2) | Significant correlation among the levels of s-cortisol, serum cortisol and urinary cortisol. |
| | Sugano A [35] | 2000 | 7 Participants (61.9 ± 11.8) | Decrease in s-cortisol levels post-exercise. (Water exercise) Decrease in s-cortisol levels post-exercise. (Land stretching) No difference in s-cortisol levels among water and land physical exercise. |
| | Filaire E [36] | 1996 | 10 Swimmers (18.5 ± 1.2) 14 Handball players 7 Sedentary participants | Increase in s-cortisol post-exercise only in handball players. Higher s-cortisol concentrations the following to exercise morning for handball players compared to swimmers and sedentary participants. |
| | Port K [37] | 1991 | 6 Males | Increase in s-cortisol levels, especially in intensive training (Ergometer cycling) |
| | Cook NJ [38] | 1986 | 8 Marathon runners (35.1 ± 8.1) | Increase in salivary cortisol during the marathon. Return to baseline levels, some hours after the marathon. |
| Cystatins | Sant'Anna M [39] | 2019 | 20 Pentathletes (28 ± 6) | Increase in the secretion of S-type cystatins and cystatin C after aerobic and anaerobic exercise |
| Ferritin | Franco-Martinez L [40] | 2019 | 18 Males (21.2 ± 4.2) | No difference in ferritin concentration in saliva after exercise. No correlation between blood lactate and salivary ferritin |

Table 1. Cont.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|-------------------------------|-------------------------------|--|---|---|
| Immunoglobulin A (s-IgA) | Yasuda N [15] | 2021 | 11 Males | No change in s-IgA after moderately long-lasting exercise (2 h, cycling) |
| | Rapson A [41] | 2020 | 46 Participants (23.7 ± 3.5) | Salivary free light chains (FLCs) follow the same pattern with the fluctuation of s-IgA. |
| | Tiernan C [42] | 2020 | 19 Rugby players (19.7 ± 1.1) | A decrease in 65% or more of sIgA was associated with an increased risk within the following 2 weeks for contracting an upper respiratory tract infection. Not a significant association between s-IgA levels and training load. |
| | Engels HJ [43] | 2018 | 50 Female participants | Increase in s-IgA concentration immediately after exercise. Return to the s-IgA pre-exercise levels 2 h after exercise. |
| | Gillum T [44] | 2014 | 18 Participants | Increase in s-IgA, 1 h after exercise |
| | Gillum T [27] | 2013 | 14 Marathon runners (43.7 ± 9.9) | Decrease in s-IgA concentration after exercise. |
| | He CS [29] | 2010 | 8 Basketball players (20.5 ± 0.3) | Decrease in s-IgA concentration during the training and competition period |
| | Davison G [45] | 2009 | 12 Males cycling | Increase in s-IgA after exercise |
| | Allgrove J [31] | 2009 | 16 Active adults (22 ± 4) | Increase in s-IgA concentration post-exercise. |
| | Allgrove J [17] | 2008 | 10 Males (23) | Increase in s-IgA concentration after exercise, followed by a return to pre-existing values 1 h post-exercise. (Cycling) An increase in s-IgA was independent of exercise intensity |
| | Sari-Sarraf V [46] | 2006 | 8 Participants (24.1 ± 3.3) | No difference in s-IgA concentration and secretion rate during or after training. (standing, walking, jogging, cruising and sprinting) |
| | Costa RJS [47] | 2005 | 32 Male triathletes, (32.1 ± 9) | Increased s-IgA concentration post-exercise in high carbohydrate consuming group. |
| | Tzai-Li Li [48] | 2005 | 25 Participants (29) | Increase in s-IgA concentration after exercise, followed by a return to pre-existing values 2 h post-exercise. (Cycling) |
| | Tiollier E [49] | 2005 | 21 Military cadets | No difference in s-IgA levels after 3-weeks of training. Increased s-IgA levels after 5 days of training and return to pre-training levels after 1 week of recovery. |
| | Li TL [18] | 2004 | 8 Males cycling (28.9 ± 1.8) | Increase in s-IgA concentration after exercise (60% V _O max, 2 h) No difference in s-IgA secretion rate after training. |
| | Akimoto T [50] | 2003 | 45 Participants (64.9 ± 8.4) | Increase in s-IgA concentration 12 months of physical exercise training. Increase in s-IgA concentration after 4 months of physical exercise training. |
| | Nehlsen-Cannarella S [51] | 2000 | 20 Elite female rowers (22.6 ± 0.5) 19 Non-athletic females (24.6 ± 0.8) | 77% Higher s-IgA concentration in the rowers compared to non-athletes |
| | Walsh NP [19] | 1999 | 8 Well trained males (25 ± 1) | S-IgA secretion rate was not affected by the exercise. (cycling) |
| Shimizu K [52] | 2007 | 114 Men (71.6 ± 0.4) 170 Women (71 ± 0.3) Elderly volunteers | The S-IgA flow rate and secretion rate increased when physical activity was improved. | |
| Gleeson M [53] | 1995 | 26 Elite swimmers (16–24) 12 Athletic participants (19–41) | Increase in s-IgA levels in professional swimmers immediately after exercise. There is no difference in pre-exercise s-IgA levels in professional swimmers compared to the athletic participant. | |

Table 1. Cont.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|--|-------------------------------|----------------------------------|--|--|
| Immunoglobulin G (s-IgG) | Mackinnon LT [54] | 1994 | 10 Joggers (20–35) 7 Marathon runners (20–35) 14 Swimmers (16–20) | No change in s-IgA secretion rates in Joggers after exercise, irrespective of exercise intensity. Decrease in s-IgA secretion rates in marathon runners after 90 min of running after the first day of exercise. There is no difference in s-IgA concentration, in stale compared to well-trained swimmers, during a season of 6 months. |
| | Mackinnon LT [55] | 1993 | 12 Physical education students (17–25) | Increase in S-IgA concentration after training, A decline in S-IgA flow rate after training. (Cycling) |
| | McDowell SL [56] | 1992 | 24 Novice runners (22.1 ± 3) | Decrease in s-IgA levels immediately after the exercise. Increased compared to prior exercise, s-IgA concentration 1 h post-exercise. S-IgA levels are independent of salivary cortisol. |
| | Tomasi T [57] | 1982 | 8 Nationally ranked skiers (23.5) 8 Non-competitive skiers | Decrease in s-IgA levels after exercise. (skiing) Lower s-IgA levels of the skiers compared to non-competitive athletes. |
| | Nehlsen-Cannarella S [51] | 2000 | 20 Elite female rowers (22.6 ± 0.5) 19 Non-athletic females (24.6 ± 0.8) | No difference in s-IgG concentration among the rowers compared to non-athletes |
| Immunoglobulin M (s-IgM) | Gleeson M [53] | 1995 | 26 Elite swimmers (16–24) 12 Athletic participants (19–41) | Higher s-IgG levels in professional swimmers compared to the athletes post-exercise. There is no difference in s-IgG levels among the professional swimmer and athletic participants post-exercise. |
| | Mackinnon LT [55] | 1993 | 12 Physical education students (17–25) | Increase in S-IgG concentration after training, A decline in S-IgG flow rate after training. (Cycling) |
| | Tomasi T [57] | 1982 | 8 Nationally ranked skiers (23.5) 8 Non-competitive skiers (25.5) | Same s-IgG levels prior and post-exercise. No difference in s-IgG levels among the skiers and non-competitive athletes. |
| Insulin-like growth factor I (s-IGF-I) | Nehlsen-Cannarella S [51] | 2000 | 20 Elite female rowers (22.6 ± 0.5) 19 Non-athletic females (24.6 ± 0.8) | No difference in s-IgM concentration among the rowers compared to non-athletes |
| | Gleeson M [53] | 1995 | 26 Elite swimmers (16–24) 12 Athletic participants (19–41) | Higher s-IgM levels in professional swimmers compared to the athletes post-exercise. There is no difference in s-IgM levels among the professional swimmer and athletic participants post-exercise. |
| | Mackinnon LT [55] | 1993 | 12 Physical education students (17–25) | Increase in S-IgM concentration after training, A decline in S-IgM flow rate after training. (Cycling) |
| Lactate | Antonelli G [58] | 2009 | 18 Cyclists (19 ± 1) | Increase in s-IGF-I after exercise. |
| | Antonelli G [59] | 2007 | 15 Volleyball players 14 Sedentary females | Lower s-IGF-I in athletes compared to sedentary females before exercise. |
| | Almasi G [60] | 2021 | 31 Elite adolescent athletes | Increase in the concentration of salivary lactate after exercise. (200 m freestyle swimming) |
| | Hermann R [61] | 2019 | 32 Males (24.3 ± 3.3) | Increase in the concentration of salivary lactate after ergometer |
| | Franco-Martinez L [40] | 2019 | 18 Males (21.2 ± 4.2) | Increase in the concentration of lactate in the saliva after exercise. (Sprinting) Lactate in saliva was correlated with blood lactate only in untrained subjects |
| | Santos RV [62] | 2006 | 15 Expert marathon racers | Increase in the concentration of salivary lactate after 18 km of running. |
| Segura R [63] | 1996 | 9 Amateur sportsmen (22.2 ± 1.9) | Increase in the concentration of salivary lactate both for anaerobic and aerobic exercise. | |

Table 1. Cont.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|-------------------------------|-------------------------------|------------------|--|--|
| Lysozyme, lactoferrin | Ohkuwa T [64] | 1995 | 7 Long-distance runners (18.6 ± 0.8) 5 Sprinters (19.3 ± 1.1) | Increase the salivary lactate concentration both in 400-m and in the 3000-m run. Higher lactate concentration after 400-m run for sprinters compared to long-distance runners. |
| | Port K [37] | 1991 | 6 Males | Steadily increase of the lactate throughout the exercise |
| | Gillum T [44] | 2017 | 11 Participants (20.3 ± 0.8) | Increase in lysozyme secretion rate after exercise. (ran for 45 min at 75% of VO ₂ peak) |
| | De Feo P [65] | 1989 | 9 Male Participants (21.1 ± 1.1) 9 Female (22.4 ± 2.4) | Increase in lactoferrin secretion rate after exercise. Increase in lysozyme secretion rate after exercise. |
| | Gillum T [27] | 2013 | 14 Marathon runners (43.7 ± 9.9) | Increase in lactoferrin concentration after exercise. Lysozyme concentration does not change during exercise. |
| | He CS [29] | 2010 | 8 Basketball players (20.5 ± 0.3) | Decrease in lactoferrin concentration during the training and competition period |
| | West NP [66] | 2010 | 17 Elite rowers (24.3 ± 4) 18 Sedentary individuals (27.2 ± 7) | 60% decreased lactoferrin concentration before exercise in elite rowers compared to sedentary individuals 50% increase in the lysozyme concentration and 50% increase in lactoferrin after graded exercise. |
| | Allgrove J [17] | 2008 | 10 Males (23) | Increased lysozyme concentration after exercise for 1 h. (Cycling) |
| Melatonin | Carlson LA [67] | 2019 | 12 Regularly exercising men | Increased salivary melatonin after morning exercise compared to afternoon exercise. |
| MicroRNAs | Hicks S [68] | 2020 | Former football players (73 ± 8) Younger participants (20 ± 5) | Non-invasive measurement of saliva miRNAs, (miR-340-5p, miR-339-3p, miR-361-5p, miR-28-3p) may have utility to identify individuals at risk for chronic concussion symptoms. |
| Nitric Oxide | Di Pietro V [69] | 2018 | 52 Rugby Athletes (26) | Differentially expressed miRNAs could be particularly suitable for concussion assessment. |
| | Gonzalez D [70] | 2008 | 24 Participants (27.2 ± 9.6) | No change in nitric concentration after aerobic exercise. |
| | Panosian AG [71] | 1999 | 109 Athletes (32–44) | Increase in nitric oxide concentration after exercise in amateur athletes. |
| Peroxides | Damirchi A [72] | 2010 | 10 University students | Increase in peroxide secretion rate at the 75%VO ₂ (2 max) after exercise. Decrease 1 h after exercise. |
| | Gillum T [27] | 2013 | 14 Marathon runners (43.7 ± 9.9) | Salivary flow rate not changed by the exercise. |
| | Damirchi A [72] | 2010 | 10 University students | The salivary flow rate does not change by the exercise. (Treadmill runs) |
| | Allgrove J [31] | 2009 | 16 Active adults (22 ± 4) | Decrease in saliva flow rate during exercise, followed by a return to pre-existing values 1 h post-exercise. (cycling) |
| | Allgrove J [17] | 2008 | 10 Males (23) | Salivary flow rate not changed by the exercise. (Cycling) |
| | Shimizu K [52] | 2007 | 114 Men (71.6 ± 0.4) 170 Women (71 ± 0.3) Elderly volunteers | No difference in salivary flow rate when physical activity is improved. |
| | Sari-Sarraf V [46] | 2006 | 8 Participants (24.1 ± 3.3) | Decrease in saliva flow rate during exercise. (standing, walking, jogging, cruising and sprinting) Duration of exercise significantly influenced the reduction in saliva flow rate. |
| | Tzai-Li Li [48] | 2005 | 25 Participants (29) | Decrease in saliva flow rate during exercise, followed by a return to pre-existing values 1 h post-exercise. (cycling) |
| | Li TL [18] | 2004 | 8 Men cycling (28.9 ± 1.8) | Decrease in saliva flow rate after exercise (60% V _O max, 2 h) |
| | Akimoto T [50] | 2003 | 45 Participants (64.9 ± 8.4) | No difference in saliva flow rate after 12 months of physical exercise training. |
| | Walsh NP [73] | 2002 | 15 Cyclists | Decrease in saliva flow rate after exercise |

Table 1. Cont.

| Evaluated Parameter of Saliva | First Author/Reference Number | Publication Date | Populations (Mean Age) | Primary Outcomes |
|-------------------------------|-------------------------------|------------------------|--|---|
| Testosterone | Nehlsen-Cannarella S [51] | 2000 | 20 Elite female rowers (22.6 ± 0.5) 19 Non-athletic females (24.6 ± 0.8) | No difference in saliva secretion rate among the professionals and non-athletic participants. |
| | Walsh NP [19] | 1999 | 8 Well trained males (25 ± 1) | The saliva flow rate was not affected by the exercise. (cycling) |
| | Blannin A [74] | 1998 | 18 Male with mixed physical fitness (23 ± 1) | Saliva flow rate reduced by moderate or high-intensity exercise |
| | Steenberg P [75] | 1997 | 42 Triathletes (34.1 ± 7.3) | Reduced saliva flow rate after the race |
| | Pilardeau P [76] | 1990 | 12 Male | In normoxia or hypoxia, there is no difference in saliva flow rate after exercise. However, in the situation of acute hypoxia, reduced saliva flow rate after exercise. |
| | Hough J [21] | 2021 | 23 Active males, cycle ergometer (21 ± 3) | Increase in the salivary testosterone immediately after the exercise. Decrease in the salivary testosterone 30 min post-exercise |
| | Cook CJ [77] | 2014 | 20 Rugby players (21.5 ± 1.4) | Increase in salivary testosterone after functional improvement. (Training) |
| | Crewther BT [26] | 2013 | 14 Rugby players (23.3 ± 3.5) | Increase in salivary testosterone before winning games. |
| | Budde S [30] | 2010 | 40 High school students (14.35) | Increase in the salivary testosterone after exercise. No effect of activity level on the salivary testosterone prior and after exercise. (Sprinting) |
| | Crewther BT [78] | 2010 | 4 Male (20.8 ± 3.5) 4 Female (20.8 ± 4.6) Olympic weightlifters | Significant correlation of pre-workout salivary testosterone, with the Olympic total lift, only for male weightlifters. |
| | Thomas NE [32] | 2009 | 17 School children (15.5 ± 0.4) | Increase in salivary testosterone after exercise. (Cycling) |
| | Filaire E [79] | 2000 | 14 National handball players (24.1 ± 2.6) 10 Sedentary women (23.5 ± 3.4) | Higher salivary testosterone for sedentary women, compared to professional players, at resting. Correlation among testosterone and dehydroepiandrosterone (DHEA). |
| | Cook NJ [38] | 1986 | 8 Marathon runners (35.1 ± 8.1) | Increase in salivary testosterone during the marathon. Increased salivary testosterone concentration, the days after the marathon. |
| | Uric acid | Franco-Martinez L [40] | 2019 | 18 Males (21.2 ± 4.2) |
| Gonzalez D [70] | | 2008 | 24 Participants (27.2 ± 9.6) | Increase in uric acid by aerobic exercise. |

3.1. Salivary Secretion

Parasympathetic and sympathetic neural systems regulate saliva secretion. Each one has a different effect on its secretion. When the sympathetic neural system stimulates saliva secretion, it consists more of proteins such as α -amylase and cystatin. When the parasympathetic system stimulates saliva secretion, its volume is mainly increased [80]. Physical exercise seems to increase the salivary flow rate and protein (e.g., amylase, lysozyme and MUC5B) secretion [81]. The decreased saliva flow may influence the concentration of the saliva substances, such as the metabolites [55]. S-IgA's protective role is dependent on its secretion rate [54].

It has been reported that in healthy individuals, unstimulated saliva is secreted at rest at the rate of 0.30–0.65 mL/min, whereas stimulated saliva flows at a rate of 1.5–6.0 mL/min [82]. Saliva flow rate increases during exercise to a secretion rate of 0.78–0.94 and decreases after recovery [73–75]. Under the physical exercise, as the flow rate of the saliva increases, the concentration of Na^+ and HCO_3^- is raised. Na^+ , Ca^{2+} , Cl^- , HCO_3^- and proteins increase, whereas K^+ shows little change [82]. Following the physical activity, the increase in salivary proteins may be associated with adrenergic activity [83]. Increased plasma catecholamines may also cause an α -amylase increase during exercise [84]. Salivary

and serum cortisol increase linearly with the intensity of exercise [85]. Secretion of S-type cystatins and cystatin C is also increased by physical exercise [39].

Exercise, performed in normoxia and hypoxia, did not affect saliva flow rate or α -amylase concentrations. On the other hand, acute hypoxia increases mean saliva flow rate, both at rest and after exercise and a decrease in mean saliva K^+ concentration, at rest and after exercise [39]. In addition, food consumption during the exercise increases saliva's flow rate and the secretion of specific proteins, as lysozyme and α -amylase, but not s-IgA secretion [76].

3.2. Lysozyme and Lactoferrin

Lysozyme and lactoferrin are the main saliva's antimicrobial proteins. Lysozyme and lactoferrin act synergistically to augment immunity [45]. Lysozyme breaks down the polysaccharide wall of their cell, thus facilitating the destruction of mainly gram-positive bacteria [86]. Furthermore, exercise activates neutrophils, potentially causing the release of lysozyme and lactoferrin into the saliva [87,88]. The lysozyme concentration in the saliva and its secretion rate is negatively influenced by psychological stress [89]. Lactoferrin has anti-inflammatory and anti-microbial roles, preventing bacterial growth by sequestering ferric iron from the bacteria and directly interacting and damaging bacterial membranes [90,91].

Lower salivary lactoferrin concentrations were found in elite rowers than in the non-exercising control group over a training season [66]. Moreover, lactoferrin concentration in saliva decreased over a competitive training season in basketball players [29]. On the other hand, acute running increases lactoferrin and lysozyme expression in both men and women [27,92].

3.3. Lactate

Lactate is an essential source of energy for the metabolism of skeletal muscle. Measuring blood lactate concentration provides information regarding changes in glycolysis and the capacity of the anaerobic work [92]. Saliva lactate is possibly formed by passive diffusion from salivary glands and blood [64]. Blood lactate and saliva lactate are highly correlated, with most kinds of exercise [63]. It has been suggested that salivary lactate increases due to an increase in the lactate concentration of the blood, which leads to an increase in the permeability of the blood–saliva barrier during exercise [93]. Salivary concentrations of lactate during training have been estimated as lower than those of the lactate of the blood [62]. Lactate levels can also be used to assess the possibility of overtraining, as they decrease during intense exercise [60]. Lactate levels seem to be independent of individuals' fitness and alteration in anxiety [61].

3.4. Oral Peroxides–Nitric Oxide

One of the highest value components of the saliva antioxidant system is the enzyme nitric oxide [94]. The paramount importance of the salivary antioxidant system is to decompose hydrogen peroxide produced by bacteria. Then, the enzymes inactivate bacterial glycolytic enzymes, thus destroying the oral bacteria [95]. Exercise with moderate intensity increases the activity of salivary peroxidase. The lower the training power, the longer the peroxidase remains at the high activity level [72,96]. Exercise-induced stress also induces the production of salivary nitric oxide [97].

Oral peroxide increased only in beginner athletes and not in well-trained athletes and professionals [71,98]. So, it can be a measure of the adaptation of the subject to intense and heavy exercise. Furthermore, exercise increases saliva uric acid and total antioxidant activity, while the saliva lipid hydroperoxides decrease. Thus, it seems increment in uric acid and total antioxidant activity prevent the lipid hydroperoxides from being generated, making oral peroxide a marker of oxidative stress in saliva [70].

3.5. Salivary A-Amylase (sAA)

Some non-immunological salivary proteins can inhibit bacterial adherence to the oral cavity. One protein is a-amylase, which can bind to several oral bacteria [99–101]. Salivary a-amylase is the predominant enzyme in saliva. It is responsible for the degradation of starch and glycogen to maltose and has been used as a sympathetic nervous system activation biomarker [102]. Both a-amylase and cortisol of the saliva serve as markers to stress response of exertion [18]. However, salivary a-amylase activity is a more sensitive, exercise-induced stress marker than cortisol, as it is produced locally in the salivary glands, controlled by the autonomous nervous system. The cortisol is transported from blood to saliva [103]. Beta-adrenergic agonists are capable of stimulating salivary a-amylase release without increasing salivary flow [104].

Salivary a-amylase increased in acute exercise and the magnitude depended on exercise intensity [16,105]. Two hours of moderate exercise seems to lead to enhanced a-amylase activity [15]. Salivary a-amylase concentrations predict plasma catecholamine levels, particularly norepinephrine, under various stressful conditions and maybe a more direct endpoint of catecholamine activity [20]. Salivary a-amylase responses are quick within one to a few minutes, even faster from blood cortisol levels and it declines rapidly after removing the stress factor [20,65].

3.6. Salivary Cortisol (S-Cortisol)

Cortisol is the primary glucocorticoid produced by the adrenal cortex that regulates blood glucose homeostasis [65,106]. It is released in stressful situations and leads to an increase in blood-glucose [107]. High salivary cortisol concentrations are related to impaired insulin sensitivity [108]. Free cortisol is more increased than salivary cortisol [109]. After intensive training, periods with elevated cortisol associated with the mild hypoglycemic state seem to produce an immunosuppressive state and decrease plasma glutamine concentration [110]. Cortisol is responsible for 95% of the glucocorticoid activity in the human body [111]. The secretion of cortisol due to exercise is not immediate [37].

Salivary cortisol is expected to be decreased during non-exercising [112]. Low-intensity exercise also reduces the levels of salivary cortisol [112,113]. During moderate-intensity exercise, its levels remain almost stable [114,115]. Exercise of high intensity influences the secretory process of the adrenal cortex and starts cortisol releasing in adults and adolescents [30,116]. Heavy training significantly increases the amount of salivary cortisol immediately after exercise. Endurance exercise produces higher plasma cortisol than acute high-intensity exercise [79,117]. A study suggests that salivary cortisol is lower during water than land exercises [36]. This contrasts with another research work that indicates that the salivary cortisol concentrations similarly significantly decreased with water exercise and land stretching [35]. Physical activity seems to increase the diminished due to poor sleep quality awakening cortisol levels [110].

Physical fitness is associated with cortisol secretion during psychological stress [25,34,118–120]. Psychological stress factors can contribute to higher values of cortisol [34,118]. A relation between salivary cortisol and anxiety has been suggested [119,120]. Depressive patients before and 10 min after the exercise sessions appear to have significantly decreased levels of salivary-free cortisol [23,24,28]. In addition, physical exercise has been found to decline the rate of cortisol [12,22]. Salivary cortisol measurements can detect the circadian rhythm of the athletes, assisting in the prevention of overtraining syndrome [121].

Carbohydrates during exercise decrease glutamine depletion, cortisol and so the immune activity [122,123]. On the other hand, a diet with low carbohydrates suppresses immune activity and increases cortisol in plasma [123]. In addition, carbohydrate intake during prolonged exercise decreases stress hormones responses [123]. According to other studies, carbohydrates did not affect saliva flow rate and s-IgA concentration during a single bout of exercise [48,51]. However, post-exercise consumption of chocolate milk,

which contains carbohydrates, proteins, fluid and electrolytes, is associated with lower saliva-cortisol response and higher saliva flow rate than water [124,125].

3.7. Steroids–Testosterone

Steroid hormones detectable in saliva include cortisol, androgens including testosterone and dehydroepiandrosterone (DHEA), estrogens and progesterone and aldosterone. Some serum components can transfer freely through the lipid-rich cell membrane into the salivary gland acinar cells and diffuse into the saliva. However, this mechanism is applied only to some lipid-soluble components such as steroid hormones. Salivary steroids are suggested to provide a more sensitive marker of changes than plasma ones [33]. Salivary testosterone (sal-T), in unison with cortisol (sal-C), has been used as a marker of anabolic status [26,38]. Adrenal glands secrete DHEA. A strong relationship between salivary and plasma DHEA has been reported [126]. It has also been suggested as an analog in salivary testosterone measurements to assess exercise response in females [79].

Salivary measures of testosterone are a reliable indicator of its plasma concentrations [127]. Both testosterone and cortisol can be increased at a significant rate with hypertrophic exercising [128]. Salivary testosterone is increased linearly during exercise and reaches its peak after the end of the training [32]. Salivary testosterone seems to be a valuable tool to assess the performance of the athletes and their readiness to train at a certain intensity level and assist with the designing of workouts for optimal gains [78]. This can be explained as testosterone contributes to neuromuscular performance and the muscles' long-term development [129]. The measurement of steroids of saliva samples throughout a competitive event can provide meaningful data regarding exercise's psychological and physical stress and highlight overtraining [21,38].

3.8. Salivary Immunoglobulin A (s-IgA)

Immunoglobulin A (IgA) is the pre-dominant immunoglobulin in the mucosal immune system [130]. IgA is produced by long-lived plasma B cells, which are influenced by T cell-generated cytokines [131]. It is found in the saliva, intestinal secretions, bronchoalveolar lavage fluid, urine and other mucosal fluids and it is also associated with resistance to specific infections [132].

Salivary immunoglobulin A (s-IgA) plays an essential role in immunity as the first line of defense against potential pathogens [133]. Older people who follow a daily moderate-intensity exercise program appear to have higher levels of S-IgA, than others of the same age who do not exercise. In addition, moderate to intense exercise can increase the secretion of salivary S-IgA in older adults to improve their immune function [52]. S-IgA also presented an increased post-exercise when combined with a high carbonated diet, suggesting that carbohydrates enhance the immune activity during exercise [47]. On the contrary, others indicated no effect of carbohydrate ingestion on saliva immunoglobulin concentrations or secretion rates [51]. Finally, a study demonstrated that a fed or fasted state 2 h before exercise does not influence resting s-IgA [31].

As far as the relationship between exercise and s-IgA, the majority of the studies conclude that s-IgA decreases after exercise [27,134], others report no change [19,46] and others show increased levels of s-IgA post-exercise [17,43,50]. S-IgA measurement seems to be a good way which shows the over-training [135,136]. The s-IgA may decrease over prolonged periods of intensive training in elite athletes. This reduction is attributed to neurohormonal factors related to physical and psychological stress during intensive daily exercise [49]. In addition, no significant association between changes in s-IgA levels and those in cortisol levels during exercise [49,56]. Low temperatures, such as in ski races, might depress the activity of secretion of s-IgA [57].

High training loads can decrease s-IgA and suppress immune function, as lower concentrations of salivary IgA or chronic salivary IgA deficiencies are associated with an increased frequency of upper respiratory tract infections (URTI) [36,137]. However, more studies are required to clarify the relationship between the components of the saliva and

the incidence of URTI. The coaches can use this information to predict athletes' immune function to help reduce the risk of upper respiratory tract infection [42]. S-IgA fluctuation is seemed to be mirrored by the secretion of salivary free light chains [41].

3.9. Immunoglobulin G (IgG) & Immunoglobulin M (IgM)

All salivary immunoglobulins contribute to mucosal immunity and defense against upper respiratory tract infections. However, only a few studies have evaluated s-IgM and s-IgG, under physical exercise. It seems that s-IgG levels remain unchanged during exercise, while s-IgM levels decrease and are restored within 24 h [105].

3.10. Insulin-like Growth Factor 1 (s-IGF-1)

According to a study in young female volleyball players, free IGF-1 in saliva levels was decreased in well-trained athletes, compared with sedentary groups [59]. In contrast, in another study, salivary IGF-1 was increased after exercise, while plasma IGF-1 was not [58]. Salivary IGF-1 can be more sensitive. Training is suggested to increase human growth hormone hGh secretion, which is regulated by the hypothalamus. The increase seems to be attributed to insulin-like growth factor I (IGF-I) [138].

3.11. Salivary MicroRNAs

Salivary microRNAs can reflect critical biological processes related to a trauma, such as hypoxia, neurogenesis, axon repair and cell death. MicroRNAs, expressed from the saliva, seem to be an accurate non-invasive alternative to diagnose a traumatic brain injury due to a concussion [68,69].

3.12. Melatonin

Melatonin is a hormone found naturally in the human body, regulating sleep-wake cycles. Physical exercise during the afternoon can decrease melatonin secretion compared to the morning exercise. A non-invasive evaluation of melatonin can be performed [67].

3.13. Uric Acid

The effect of exercise in the concentration of uric acid in saliva has to be further investigated, as the available studies come to opposite conclusions. Aerobic exercise, such as long-distance running, has a significant impact, increasing the concentration of uric acid [70]. On the other hand, it seems that explosive physical exercise, such as short sprints, does not significantly influence the concentration of uric acid in saliva [40,139].

4. Conclusions

A significant part of the scientific literature has investigated the relation of physical exercise with the physical and biological properties of saliva. The shift in the composition of the saliva, during and after a workout, reflects the benefits of exercise, its potential risks and the capability of the saliva to serve as a health indicator. Saliva analysis can be used as a non-invasive method to measure exercise-induced changes, adaptation in hormones, lactate accumulation and shift in immunological markers. The type and the frequency of training, the physical condition and the athletes' general health can influence hormones, immunoglobulins and saliva enzymes. The correlation between saliva and physical exercise has to be further investigated, especially for the organic components of the saliva. The available knowledge has to be applied to benefit sports activities. Athletes and coaches must consider monitoring salivary hormones during training or competitions for consistency or assessing overall workouts. Intelligent, easy to access affordable, non-invasive devices have to be further developed to take advantage of saliva's information.

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References

- Schneyer, L.H.; Young, J.A.; Schneyer, C.A. Salivary secretion of electrolytes. *Physiol. Rev.* **1972**, *52*, 720–777. [[CrossRef](#)]
- Guyton, A.C. Secretory functions of the alimentary tract. In *Textbook of Medical Physiology*, 9th ed.; Guyton, A.C., Ed.; WB Saunders Company: Philadelphia, PA, USA, 1996; pp. 709–725.
- Dawes, C. Rhythms in salivary flow rate and composition. *Int. J. Chronobiol.* **1974**, *2*, 253–279. [[PubMed](#)]
- Garret, J.R. Structure and innervation of salivary glands. In *Scientific Foundations of Dentistry*; Cohen, B., Kramer, J.R.H., Eds.; Heinemann: London, UK, 1976; pp. 499–506.
- Suddick, R.P.; Dowd, F.J. Mechanisms of salivary secretion. In *The Biologic Basis of Dental Caries: An Oral Biology Textbook of Lewis Menaker*; Menaker, L., Ed.; Harper & Row Publishers: Hagerstown, MD, USA, 1986; pp. 67–118.
- Baum, B.J. Neurotransmitter control of secretion. *J. Dent. Res.* **1987**, *66*, 628–632. [[CrossRef](#)] [[PubMed](#)]
- Anderson, L.C.; Garret, J.R.; Johnson, D.A.; Kauffman, D.L.; Keller, P.J.; Thulin, A. Influence of circulating catecholamines on protein secretion into rat parotid saliva during parasympathetic stimulation. *J. Physiol.* **1984**, *352*, 163–171. [[CrossRef](#)]
- Beaven, C.M.; Gill, N.D.; Cook, C.J. Salivary testosterone and cortisol responses in professional rugby players after 4 resistance exercise protocols. *J. Strength Cond. Res.* **2008**, *22*, 426–432. [[CrossRef](#)]
- Young, J.A.; Van Lennep, E.W. Secretion of salivary and saltglands. In *Membrane Transport in Biology*; Giebisch, G., Tosteson, D.C., Ussing, H.H., Eds.; Springer: Berlin, Germany, 1979; pp. 563–574.
- Garret, J.R. Physiology of salivary glands. In *Scientific Foundations of Dentistry*; Cohen, B., Kramer, J.R.H., Eds.; Heinemann: London, UK, 1976; pp. 507–512.
- Schneyer, C.A. Salivary gland changes after isoproterenol induced enlargement. *Am. J. Physiol.* **1962**, *203*, 232–236. [[CrossRef](#)]
- Schneyer, L.H. Amylase content of separate salivary secretions of man. *J. Appl. Physiol.* **1956**, *9*, 453–455. [[CrossRef](#)]
- Schneyer, L.H.; Schneyer, C.A. Apparent synthesis of submaxillary gland amylase during pilocarpine administration. *Am. J. Physiol.* **1956**, *187*, 403–406. [[CrossRef](#)]
- Bartual, J. Fisiología y fisiopatología parotídea. In *Patología de la Parótida*; Clemente, M., Ed.; Fundación García Muñoz: Valencia, Spain, 1980; pp. 27–40.
- Yasuda, N.; Yamamoto, K.; Iwashita, N. Concurrent evaluation of salivary and urinary α -amylase activity following prolonged exercise with or without carbohydrate solution in aerobically active men. *Neuroendocrinol. Lett.* **2021**, *42*, 265–276.
- Wunsch, K.; Wurst, R.; Dawans, B.; Strahler, J.; Kasten, N.; Fuchs, R. Habitual and acute exercise effects on salivary biomarkers in response to psychosocial stress. *Psychoneuroendocrinology* **2019**, *106*, 216–225. [[CrossRef](#)] [[PubMed](#)]
- Allgrove, J.E.; Gomes, E.; Hough, J.; Gleeson, M. Effects of exercise intensity on salivary antimicrobial proteins and markers of stress in active men. *J. Sports Sci.* **2008**, *26*, 653–661. [[CrossRef](#)]
- Li, T.L.; Gleeson, M. The effect of single and repeated bouts of prolonged cycling and circadian variation on saliva flow rate, immunoglobulin A and alpha-amylase responses. *J. Sports Sci.* **2004**, *22*, 15–24. [[CrossRef](#)]
- Walsh, N.P.; Blannin, A.K.; Clark, A.M.; Cook, L.; Robson, P.J.; Gleeson, M. The effects of high-intensity intermittent exercise on saliva IgA, total protein and alpha-amylase. *J. Sports Sci.* **1999**, *17*, 129–134. [[CrossRef](#)] [[PubMed](#)]
- Chatterton, R.T.; Vogelsong, K.M.; Lu, Y.C.; Ellman, A.B.; Hudgens, G.A. Salivary alpha-amylase as a measure of endogenous adrenergic activity. *Clin. Physiol.* **1996**, *16*, 433–448. [[CrossRef](#)] [[PubMed](#)]
- Hough, J.; Leal, D.; Scott, G.; Taylor, L.; Townsend, D.; Gleeson, M. Reliability of salivary cortisol and testosterone to a high-intensity cycling protocol to highlight overtraining. *J. Sports Sci.* **2021**, *27*, 1–7. [[CrossRef](#)]
- Ushiki, K.; Tsunekawa, K.; Shoho, Y.; Martha, L.; Ishigaki, H.; Matsumoto, R.; Yanagawa, Y.; Nakazawa, A.; Yoshida, A.; Nakajima, K.; et al. Assessment of exercise-induced stress by automated measurement of salivary cortisol concentrations within the circadian rhythm in Japanese female long-distance runners. *Sports Med. Open* **2020**, *6*, 38. [[CrossRef](#)]
- Pearlmutter, P.; DeRose, G.; Samson, C.; Linehan, N.; Cen, Y.; Begdache, L.; Won, D.; Koh, A. Sweat and saliva cortisol response to stress and nutrition factors. *Sci. Rep.* **2020**, *10*, 19050. [[CrossRef](#)] [[PubMed](#)]
- Rahman, M.S.; Zhao, X.; Liu, J.J.; Torres, E.Q.; Tibert, B.; Kumar, P.; Kaldov, V.; Lindefors, N.; Forsell, Y.; Lavebratt, C. Exercise Reduces Salivary Morning Cortisol Levels in Patients with Depression. *Mol. Neuropsychiatry* **2019**, *4*, 196–203. [[CrossRef](#)]
- Wood, C.J.; Clow, A.; Hucklebridge, F.; Law, R.; Smyth, N. Physical fitness and prior physical activity are both associated with less cortisol secretion during psychosocial stress. *Anxiety Stress Coping* **2018**, *31*, 135–145. [[CrossRef](#)]
- Crewther, B.T.; Sanctuary, C.E.; Kilduff, L.P.; Carruthers, J.S.; Gaviglio, C.M.; Cook, C.J. The workout responses of salivary-free testosterone and cortisol concentrations and their association with the subsequent competition outcomes in professional rugby league. *J. Strength Cond. Res.* **2013**, *27*, 471–476. [[CrossRef](#)]
- Gillum, T.L.; Kuennen, M.R.; Gourley, C.; Schneider, S.; Dokladny, K.; Moseley, P. Salivary antimicrobial protein response to prolonged running. *Biol. Sport* **2013**, *30*, 3–8. [[CrossRef](#)] [[PubMed](#)]
- Ida, M.; Ida, I.; Wada, N.; Sohmiya, M.; Tazawa, M.; Shirakura, K. A clinical study of the efficacy of a single session of individual exercise for depressive patients, assessed by the change in saliva free cortisol level. *Biopsychosoc. Med.* **2013**, *7*, 18. [[CrossRef](#)] [[PubMed](#)]

29. He, C.S.; Tsai, M.L.; Ko, M.H.; Chang, C.K.; Fang, S.H. Relationships among salivary immunoglobulin A, lactoferrin and cortisol in basketball players during a basketball season. *Eur. J. Appl. Physiol.* **2010**, *110*, 989–995. [[CrossRef](#)] [[PubMed](#)]
30. Budde, H.; Pietrassyk-Kendziorra, S.; Bohm, S.; Voelcker-Rehage, C. Hormonal responses to physical and cognitive stress in a school setting. *Neurosci. Lett.* **2010**, *474*, 131–134. [[CrossRef](#)]
31. Allgrove, J.E.; Geneen, L.; Latif, S.; Gleeson, M. Influence of a fed or fasted state on the s-IgA response to prolonged cycling in active men and women. *Int. J. Sport Nutr. Exerc. Metab.* **2009**, *19*, 209–221. [[CrossRef](#)]
32. Thomas, N.E.; Leyshon, A.; Hughes, M.G.; Davies, B.; Gramham, M.; Bakerm, J.S. The effect of anaerobic exercise on salivary cortisol, testosterone and immunoglobulin (A) in boys aged 15–16 years. *Eur. J. Appl. Physiol.* **2009**, *107*, 455–461. [[CrossRef](#)]
33. Gozansky, W.S.; Lynn, J.S.; Laudenslager, M.L.; Kohrt, W.M. Salivary cortisol determined by enzyme immunoassay is preferable to serum total cortisol for assessment of dynamic hypothalamic-pituitary-adrenal axis activity. *Clin. Endocrinol.* **2005**, *63*, 336–341. [[CrossRef](#)]
34. Neary, J.P.; Malbon, L.; McKenzie, D.C. Relationship between Serum, Saliva and Urinary Cortisol and Its Implication During Recovery from Training. *J. Sci. Med. Sport* **2002**, *5*, 108–114. [[CrossRef](#)]
35. Sugano, A.; Nomura, T. Influence of water exercise and land stretching on salivary cortisol concentrations and anxiety in chronic low back pain patients. *J. Physiol. Anthropol. Appl. Human Sci.* **2000**, *19*, 175–180. [[CrossRef](#)]
36. Filaire, E.; Duche, P.; Lac, G.; Robert, A. Saliva cortisol, physical exercise and training: Influences of swimming and handball on cortisol concentrations in women. *Eur. J. Appl. Physiol.* **1996**, *74*, 274–278. [[CrossRef](#)] [[PubMed](#)]
37. Port, K. Serum and saliva cortisol responses and blood lactate accumulation during incremental exercise testing. *Int. J. Sports Med.* **1991**, *12*, 490–494. [[CrossRef](#)]
38. Cook, N.J.; Read, G.F.; Walker, R.F.; Harris, B.; Riad-Fahmy, D. Changes in adrenal and testicular activity monitored by salivary sampling in males throughout marathon runs. *Eur. J. Appl. Physiol.* **1986**, *55*, 634–638. [[CrossRef](#)]
39. Sant’Anna, M.L.; Oliveira, L.T.; Gomes, D.V.; Marques, S.T.F.; Provance, D.W.; Sorenson, M.M.; Salerno, V.P. Physical exercise stimulates salivary secretion of cystatins. *PLoS ONE* **2019**, *14*, e0224147. [[CrossRef](#)]
40. Franco-Martínez, L.; Tvarijonavičute, A.; Martínez-Subiela, S.; Márquez, G.; Martínez Díaz, N.; Cugat, R.; Cerón, J.J.; Jiménez-Reyes, P. Changes in lactate, ferritin, and uric acid in saliva after repeated explosive effort sequences. *J. Sports Med. Phys. Fit.* **2019**, *59*, 902–909. [[CrossRef](#)]
41. Rapson, A.; Collman, E.; Faustini, S.; Yonel, Z.; Chapple, I.L.; Drayson, M.T.; Richter, A.; Campbell, J.P.; Heaney, J.L.J. Free light chains as an emerging biomarker in saliva: Biological variability and comparisons with salivary IgA and steroid hormones. *Brain Behav. Immun.* **2020**, *83*, 78–86. [[CrossRef](#)]
42. Tiernan, C.; Lyons, M.; Comyns, T.; Nevill, A.M.; Warrington, G. Salivary IgA as a Predictor of Upper Respiratory Tract Infections and Relationship to Training Load in Elite Rugby Union Players. *J. Strength Cond. Res.* **2020**, *34*, 782–790. [[CrossRef](#)]
43. Engels, H.J.; Kendall, B.J.; Fahlman, M.M.; Gothe, N.P.; Bourbeau, K.C. Salivary immunoglobulin A in healthy adolescent females: Effects of maximal exercise, physical activity, body composition and diet. *J. Sports Med. Phys. Fit.* **2018**, *58*, 1096–1101. [[CrossRef](#)]
44. Gillum, T.; Kuennen, M.; Miller, T.; Riley, L. The effects of exercise, sex, and menstrual phase on salivary antimicrobial proteins. *Exerc. Immunol. Rev.* **2014**, *20*, 23–38. [[PubMed](#)]
45. Davison, G.; Allgrove, J.; Gleeson, M. Salivary antimicrobial peptides (LL-37 and alpha-defensins HNP1-3), antimicrobial and IgA responses to prolonged exercise. *Eur. J. Appl. Physiol.* **2009**, *106*, 277–284. [[CrossRef](#)] [[PubMed](#)]
46. Sari-Sarraf, V.; Reilly, T.; Doran, D.A. Salivary IgA response to intermittent and continuous exercise. *Int. J. Sports Med.* **2006**, *27*, 849–855. [[CrossRef](#)] [[PubMed](#)]
47. Costa, R.J.; Jones, G.E.; Lamb, K.L.; Coleman, R.; Williams, J.H.H. The Effects of a High Carbohydrate Diet on Cortisol and Salivary Immunoglobulin A (s-IgA) During a Period of Increase Exercise Workload amongst Olympic and Ironman Triathletes. *Int. J. Sports Med.* **2005**, *26*, 880–885. [[CrossRef](#)] [[PubMed](#)]
48. Li, T.; Gleeson, M. The effects of carbohydrate supplementation during repeated bouts of prolonged exercise on saliva flow rate and immunoglobulin A. *J. Sports Sci.* **2005**, *23*, 713–722. [[CrossRef](#)] [[PubMed](#)]
49. Tiollier, E.; Gomez-Merino, D.; Burnat, P.; Jouanin, J.C.; Bourrilhon, C.; Filaire, E.; Guezennec, C.Y.; Chennaoui, M. Intense training: Mucosal immunity and incidence of respiratory infections. *Eur. J. Appl. Physiol.* **2005**, *93*, 421–428. [[CrossRef](#)] [[PubMed](#)]
50. Akimoto, T.; Kumai, Y.; Akama, T.; Hayashi, E.; Murakami, H.; Soma, R.; Kuno, S.; Kono, I. Effects of 12 months of exercise training on salivary secretory IgA levels in elderly subjects. *Br. J. Sports Med.* **2003**, *37*, 76–79. [[CrossRef](#)]
51. Nehlsen-Cannarella, S.L.; Nieman, D.C.; Fagoaga, O.R.; Kelln, W.J.; Henson, D.A.; Shannon, M.; Davis, J.M. Saliva immunoglobulins in elite women rowers. *Eur. J. Appl. Physiol.* **2000**, *81*, 222–228. [[CrossRef](#)]
52. Shimizu, K.; Kimura, F.; Akimoto, T.; Akama, T.; Kuno, S.; Kono, I. Effect of free-living daily physical activity on salivary secretory IgA in elderly. *Med. Sci. Sports Exerc.* **2007**, *39*, 593–598. [[CrossRef](#)]
53. Gleeson, M.; Blannin, A.K.; Walsh, N.P.; Bishop, N.C.; Clark, A.M. Effects of low- and high-carbohydrate diets on the plasma glutamine and circulating leukocyte responses to exercise. *Int. J. Sport Nutr.* **1999**, *8*, 49–59. [[CrossRef](#)]
54. Mackinnon, L.T.; Hooper, S. Mucosal (secretory) immune system responses to exercise of varying intensity and during overtraining. *Int. J. Sports Med.* **1994**, *15*, 179–183. [[CrossRef](#)]
55. Mackinnon, L.T.; Ginn, E.; Seymour, G.J. Decreased salivary immunoglobulin A secretion rate after intense interval exercise in elite kayakers. *Eur. J. Appl. Physiol.* **1993**, *67*, 180–184. [[CrossRef](#)]

56. McDowell, S.L.; Hughes, R.A.; Hughes, R.J.; Housh, T.J.; Johnson, G.O. The effects of exercise training on salivary immunoglobulin A and cortisol responses to maximal exercise. *Int. J. Sports Med.* **1992**, *13*, 577–580. [[CrossRef](#)]
57. Tomasi, T.; Trudeau, F.; Czerinski, D.; Erredge, S. Immune parameters in athletes before and after strenuous exercise. *J. Clin. Immunol.* **1982**, *2*, 173–178. [[CrossRef](#)] [[PubMed](#)]
58. Antonelli, G.; Gatti, R.; Prearo, M.; De Palo, E.F. Salivary free insulin like growth factor-i levels: Effects of an acute physical exercise in athletes. *J. Endocrinol. Investig.* **2009**, *32*, 1–5. [[CrossRef](#)] [[PubMed](#)]
59. Antonelli, G.; Cappellin, E.; Gatti, R.; Chiappin, S.; Spinella, P.; De Palo, E.F. Measurement of free IGF-I saliva levels: Perspectives in the detection of GH/IGF axis in athletes. *Clin. Biochem.* **2007**, *13*, 545–550. [[CrossRef](#)] [[PubMed](#)]
60. Almási, G.; Bosnyák, E.; Móra, Á.; Zsákai, A.; Fehér, P.V.; Annár, D.; Nagy, N.; Sziráki, Z.; Kemper, H.C.G.; Szmodis, M. Physiological and Psychological Responses to a Maximal Swimming Exercise Test in Adolescent Elite Athletes. *Int. J. Environ. Res. Public Health* **2021**, *18*, 9270. [[CrossRef](#)] [[PubMed](#)]
61. Hermann, R.; Lay, D.; Wahl, P.; Roth, W.T.; Petrowski, K. Effects of psychosocial and physical stress on lactate and anxiety levels. *Stress* **2019**, *22*, 664–669. [[CrossRef](#)] [[PubMed](#)]
62. Santos, R.V.; Almeida, A.L.; Caperuto, E.C.; Martins, E.; Costa Rosa, L.F. Effects of a 30-km race upon salivary lactate correlation with blood lactate. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* **2006**, *145*, 114–117. [[CrossRef](#)]
63. Segura, R.; Javierre, C.; Ventura, J.L.L.; Lizarraga, M.A.; Campos, B.; Garrido, E. A new approach to the assessment of anaerobic metabolism: Measurement of lactate in saliva. *Br. J. Sports Med.* **1996**, *30*, 305–309. [[CrossRef](#)]
64. Ohkuwa, T.; Itoh, H.; Yamazaki, Y.; Sato, Y. Salivary and blood lactate after supramaximal exercise in sprinters and long-distance runners. *Scand. J. Med. Sci. Sports* **1995**, *5*, 285–290. [[CrossRef](#)]
65. De Feo, P.; Perriello, G.; Torlone, E.; Ventura, M.M.; Fanelli, C.; Santeusano, F.; Brunetti, P.; Gerich, J.E.; Bolli, G.B. Contribution of cortisol to glucose counterregulation in humans. *Am. J. Physiol.* **1989**, *257*, 35–42. [[CrossRef](#)] [[PubMed](#)]
66. West, N.P.; Pyne, D.B.; Kyd, J.M.; Renshaw, G.M.; Fricker, P.A.; Cripps, A.W. The effect of exercise on innate mucosal immunity. *Br. J. Sports Med.* **2010**, *44*, 227–231. [[CrossRef](#)]
67. Carlson, L.A.; Pobocik, K.M.; Lawrence, M.A.; Brazeau, D.A.; Koch, A.J. Influence of Exercise Time of Day on Salivary Melatonin Responses. *Int. J. Sports Physiol. Perform.* **2019**, *14*, 351–353. [[CrossRef](#)]
68. Hicks, S.D.; Olympia, R.P.; Onks, C.; Kim, R.Y.; Zhen, K.J.; Fedorchak, G.; DeVita, S.; Rangnekar, A.; Heller, M.; Zwibel, H.; et al. Saliva microRNA Biomarkers of Cumulative Concussion. *Int. J. Mol. Sci.* **2020**, *21*, 7758. [[CrossRef](#)]
69. Di Pietro, V.; Porto, E.; Ragusa, M.; Barbagallo, C.; Davies, D.; Forcione, M.; Logan, A.; Di Pietro, C.; Purrello, M.; Grey, M.; et al. Salivary MicroRNAs: Diagnostic Markers of Mild Traumatic Brain Injury in Contact-Sport. *Front. Mol. Neurosci.* **2018**, *11*, 290. [[CrossRef](#)] [[PubMed](#)]
70. González, D.; Marquina, R.; Rondón, N.; Rodríguez-Malaver, A.J.; Reyes, R. Effects of Aerobic Exercise on Uric Acid, Total Antioxidant Activity, Oxidative Stress, and Nitric Oxide in Human Saliva. *Res. Sports Med.* **2008**, *16*, 128–137. [[CrossRef](#)] [[PubMed](#)]
71. Panossian, A.G.; Oganessian, A.S.; Ambartsumian, M.; Gabrielian, E.S.; Wagner, H.; Wikman, G. Effects of heavy physical exercise and adaptogens on nitric oxide content in human saliva. *Phytomedicine* **1999**, *6*, 17–26. [[CrossRef](#)]
72. Damirchi, A.; Kiani, M.; Jafarian, V.; Sariri, R. Response of salivary peroxidase to exercise intensity. *Eur. J. Appl. Physiol.* **2010**, *108*, 1233–1237. [[CrossRef](#)]
73. Walsh, N.P.; Bishop, N.C.; Blackwell, J.; Wierzbicki, S.G.; Montague, J.C. Salivary IgA response to prolonged exercise in a cold environment in trained cyclists. *Med. Sci. Sports Exerc.* **2002**, *34*, 1632–1637. [[CrossRef](#)]
74. Blannin, A.K.; Robson, P.J.; Walsh, N.P.; Clark, A.M.; Glennon, L.; Gleeson, M. The effect of exercising to exhaustion at different intensities on saliva immunoglobulin A, protein and electrolyte secretion. *Int. J. Sports Med.* **1998**, *19*, 547–552. [[CrossRef](#)] [[PubMed](#)]
75. Steerenberg, P.A.; van Asperen, I.A.; van Nieuw Amerongen, A.; Biewenga, A.; Mol, D.; Medema, G.J. Salivary levels of immunoglobulin A in triathletes. *Eur. J. Oral Sci.* **1997**, *105*, 305–309. [[CrossRef](#)] [[PubMed](#)]
76. Pilardeau, P.; Richalet, J.; Bouissou, P.; Vaysse, J.; Larmignat, P.; Boom, A. Saliva flow and composition in humans exposed to acute altitude hypoxia. *Eur. J. Appl. Physiol. Occup. Physiol.* **1990**, *59*, 450–453. [[CrossRef](#)]
77. Cook, C.J.; Kilduff, L.P.; Beaven, C.M. Improving strength and power in trained athletes with 3 weeks of occlusion training. *Int. J. Sports Physiol. Perform.* **2014**, *9*, 166–172. [[CrossRef](#)]
78. Crewther, B.T.; Christian, C. Relationships between salivary testosterone and cortisol concentrations and training performance in Olympic weightlifters. *J. Sports Med. Phys. Fit.* **2010**, *50*, 371–375.
79. Filaire, E.; Lac, G. Dehydroepiandrosterone (DHEA) rather than testosterone shows saliva androgen responses to exercise in elite female handball players. *Int. J. Sports Med.* **2000**, *21*, 17–20. [[CrossRef](#)]
80. Chicharro, J.L.; Lucia, A.; Perez, M.; Vaquero, A.; Urena, R. Saliva composition and exercise. *Sports Med.* **1998**, *26*, 17–27. [[CrossRef](#)] [[PubMed](#)]
81. Ligtenberg, J.M.; Brand, S.; Petra, A.M.; Veerman, C.I. The effect of physical exercise on salivary secretion of MUC5B, amylase and lysozyme. *Arch. Oral Biol.* **2015**, *60*, 1639–1644. [[CrossRef](#)]
82. Watanabe, S.; Dawes, C. The effects of different foods and concentrations of citric acid on the flow rate of whole saliva in man. *Arch. Oral Biol.* **1988**, *33*, 1–5. [[CrossRef](#)]

83. Dawes, C. The effects of exercise on protein and electrolyte secretion in parotid saliva. *J. Physiol.* **1981**, *320*, 139–148. [[CrossRef](#)] [[PubMed](#)]
84. Thaysen, J.H.; Thorn, N.H.; Schwartz, I.L. Excretion of sodium, potassium, chloride and carbon dioxide in human parotid saliva. *Am. J. Physiol.* **1954**, *178*, 155–159. [[CrossRef](#)]
85. del Corral, P.; Mahon, A.D.; Duncan, G.E.; Howe, C.A.; Craig, B.W. The effect of exercise on serum and salivary cortisol in male children. *Med. Sci. Sports Exerc.* **1994**, *26*, 1297–1301. [[CrossRef](#)] [[PubMed](#)]
86. Leitch, E.C.; Willcox, M.D. Elucidation of the antistaphylococcal action of lactoferrin and lysozyme. *J. Med. Microbiol.* **1999**, *48*, 867–871. [[CrossRef](#)]
87. Pyne, D.B. Regulation of neutrophil function during exercise. *Sports Med.* **1994**, *17*, 245–258. [[CrossRef](#)] [[PubMed](#)]
88. Karatosun, H.; Cetin, C.; Baydar, M.L. Blood and saliva lactate levels during recovery from supramaximal exercise. *Saudi Med. J.* **2005**, *26*, 1831–1832.
89. Perera, S.; Uddin, M.; Hayes, J.A. Salivary lysozyme: A noninvasive marker for the study of the effects of stress on natural immunity. *Int. J. Behav. Med.* **1997**, *4*, 170–178. [[CrossRef](#)]
90. Jenssen, H.; Hancock, R.E. Antimicrobial properties of lactoferrin. *Biochimie* **2009**, *91*, 19–29. [[CrossRef](#)]
91. Ellison, R.T.; Giehl, T.J.; LaForce, F.M. Damage of the outer membrane of enteric gram-negative bacteria by lactoferrin and transferrin. *Infect. Immun.* **1988**, *56*, 2774–2781. [[CrossRef](#)]
92. Gillum, T.; Kuennen, M.; McKenna, Z.; Castillo, M.; Jordan-Patterson, A.; Bohnert, C. Exercise does not increase salivary lymphocytes, monocytes, or granulocytes, but does increase salivary lysozyme. *J. Sports Sci.* **2017**, *35*, 1294–1299. [[CrossRef](#)]
93. Shannon, I.L. Effect of exercise on parotid fluid corticosteroids and electrolytes. *J. Dent. Res.* **1967**, *46*, 608–610. [[CrossRef](#)]
94. Mandel, I.D. The role of saliva in maintaining oral homeostasis. *J. Am. Dent. Assoc.* **1989**, *119*, 298–304. [[CrossRef](#)] [[PubMed](#)]
95. Aune, T.M.; Thomas, E.L. Accumulation of hypothiocyanite ion during peroxidase-catalyzed oxidation of thiocyanate ion. *Eur. J. Biochem.* **1977**, *80*, 209–214. [[CrossRef](#)] [[PubMed](#)]
96. Damirchi, A.; Saati Zareei, A.; Sariri, R. Salivary antioxidants of male athletes after aerobic exercise and garlic supplementation on: A randomized, double blind, placebo-controlled study. *J. Oral Biol. Craniofac. Res.* **2015**, *5*, 146–152. [[CrossRef](#)] [[PubMed](#)]
97. Sone, R.; Eda, N.; Kosaki, K.; Endo, M.; Watanabe, K. Influence of acute high-intensity exercise on salivary nitric oxide levels. *J. Oral Sci.* **2019**, *61*, 307–312. [[CrossRef](#)]
98. Moncada, S.; Higgs, A. The L-arginine-nitric oxide pathway. *N. Engl. Med.* **1993**, *329*, 2002–2012.
99. Scannapieco, F.A.; Solomon, L.; Wadenya, R.O. Emergence in human dental plaque and host distribution of amylase-binding streptococci. *J. Dent. Res.* **1993**, *73*, 1627–1635. [[CrossRef](#)]
100. Bortner, C.A.; Miller, R.D.; Arnold, R.R. Effects of alpha-amylase on in vitro growth of Legionella pneumophila. *Infect. Immun.* **1983**, *41*, 44–49. [[CrossRef](#)] [[PubMed](#)]
101. Jespersgaard, C.; Hajishengallis, G.; Russell, M.W.; Michalek, S.M. Identification and characterization of a non immunoglobulin factor in human saliva that inhibits *Streptococcus mutans* glucosyltransferase. *Infect. Immun.* **2002**, *70*, 1136–1142. [[CrossRef](#)]
102. Papacosta, E.; Nassis, G.P. Saliva as a tool for monitoring steroid, peptide and immune markers in sport and exercise science. *Sci. Med. Sport* **2011**, *14*, 424–434. [[CrossRef](#)] [[PubMed](#)]
103. Rohleder, N.; Nater, U.M. Determinants of salivary a-amylase in humans and methodological considerations. *Psychoneuroendocrinology* **2009**, *34*, 469–485. [[CrossRef](#)]
104. Ehlert, U.; Erni, K.; Hebisch, G.; Nater, U. Salivary alpha-amylase levels after yohimbine challenge in healthy men. *J. Clin. Endocrinol. Metab.* **2006**, *91*, 5130–5133. [[CrossRef](#)] [[PubMed](#)]
105. Bishop, N.C.; Gleeson, M. Acute and chronic effects of exercise on markers of mucosal immunity. *Front. Biosci.* **2009**, *14*, 4444–4456. [[CrossRef](#)]
106. Tharp, G.D. The role of glucocorticoid in exercise. *Med. Sci. Sports* **1975**, *7*, 6–11. [[CrossRef](#)] [[PubMed](#)]
107. Levine, S. Influence of psychological variables on the activity of the hypothalamic-pituitary-adrenal axis. *Eur. J. Pharmacol.* **2000**, *405*, 149–160. [[CrossRef](#)]
108. Tam, C.S.; Frost, E.A.; Xie, W.; Rood, J.; Ravussin, E.; Redman, L.M. Pennington CALERIE Team. No effect of caloric restriction on salivary cortisol levels in overweight men and women. *Metabolism* **2014**, *63*, 194–198. [[CrossRef](#)]
109. Shimojo, M.; Ricketts, M.L.; Petrelli, M.D.; Moradi, P.; Johnson, G.D.; Bradwell, A.R.; Hewison, M.; Howie, A.J.; Stewart, P.M. Immunodetection of 11 beta-hydroxysteroid dehydrogenase type 2 in human mineralocorticoid target tissues: Evidence for nuclear location. *Endocrinology* **1997**, *138*, 1305–1311. [[CrossRef](#)]
110. Robson, P.J.; Blannin, A.K.; Walsh, N.P.; Castell, L.M.; Gleeson, M. Effects of exercise intensity, duration, and recovery on in vitro neutrophil function in male athletes. *Int. J. Sports Med.* **1999**, *20*, 1–8.
111. Wilmore, J.H.; Costill, D.L. *Fisiologia do Esporte e do Exercício*, 2nd ed.; Editora Manole: São Paulo, Brazil, 2001.
112. Ortega, E.; Collazos, M.E.; Maynar, M.; Barriga, C.; De la Fuente, M. Stimulation of the phagocytic function of neutrophils in sedentary men after acute moderate exercise. *Eur. J. Appl. Physiol. Occup. Physiol.* **1993**, *66*, 60–64. [[CrossRef](#)]
113. Davies, C.T.; Few, J.D. Effects of exercise on adrenocortical function. *J. Appl. Physiol.* **1973**, *35*, 887–891. [[CrossRef](#)] [[PubMed](#)]
114. Staessen, J.; Fiocchi, R.; Fagard, R.; Hespel, P.; Amery, A. Carotid baroreflex sensitivity at rest and during exercise is not influenced by opioid receptor antagonism. *Eur. J. Appl. Physiol. Occup. Physiol.* **1989**, *59*, 131–137. [[CrossRef](#)]
115. Stephenson, L.A.; Kolka, M.A.; Francesconi, R.; Gonzalez, R.R. Circadian variations in plasma renin activity, catecholamines and aldosterone during exercise in women. *Eur. J. Appl. Physiol. Occup. Physiol.* **1989**, *58*, 756–764. [[CrossRef](#)]

116. Kirschbaum, C.; Hellhammer, D.H. Salivary cortisol in psychoneuroendocrine research: Recent developments and applications. *Psychoneuroendocrinology* **1994**, *19*, 313–333. [[CrossRef](#)]
117. Gleeson, M.; Nieman, D.C.; Pedersen, B.K. Exercise, nutrition and immune function. *J. Sport Sci.* **2004**, *22*, 115–125. [[CrossRef](#)]
118. Fekedulegn, D.; Innes, K.; Andrew, M.E.; Tinney-Zara, C.; Charles, L.E.; Allison, P.; Violanti, J.M.; Knox, S.S. Sleep quality and the cortisol awakening response (CAR) among law enforcement officers: The moderating role of leisure time physical activity. *Psychoneuroendocrinology* **2018**, *95*, 158–169. [[CrossRef](#)]
119. Lader, M. Anxiety and depression. In *Physiological Correlates of Human Behavior III: Individual Differences and Psychopathology*; Gale, A., Edwards, J.A., Eds.; Academic Press: London, UK, 1983; pp. 155–167.
120. Eck, M.V.; Berkhop, H.; Nicolson, N.; Sulon, J. The effects of perceived stress, traits, mood states, and stressful daily events on salivary cortisol. *Psychosom. Med.* **1996**, *58*, 447–458.
121. Bacurau, R.F.P.; Bassit, R.A.; Sawada, L.; Navarro, F.; Martins, E.; Rosa, L. Carbohydrate supplementation during intense exercise and the immune response of cyclists. *Clin. Nutr.* **2002**, *21*, 423–429. [[CrossRef](#)] [[PubMed](#)]
122. Bishop, N.C.; Blannin, A.K.; Rand, L.; Gleeson, M. Effects of carbohydrate and fluid intake on the blood leukocyte response to prolonged cycling. *J. Sport Sci.* **1999**, *17*, 26–27.
123. Gleeson, M.; Bishop, N.C. Special feature for the Olympics: Effects of exercise on the immune system: Modification of immune responses to exercise by carbohydrate, glutamine and anti-oxidant supplements. *Immunol. Cell Biol.* **2000**, *78*, 554–561. [[CrossRef](#)]
124. Slivka, D.; Hailes, W.; Cuddy, J.; Ruby, B. Caffeine and carbohydrate supplementation during exercise when in negative energy balance: Effects on performance, metabolism, and salivary cortisol. *Appl. Physiol. Nutr. Metab.* **2008**, *33*, 1079–1085. [[CrossRef](#)]
125. Papacosta, E.; Nassis, G.P.; Gleeson, M. Effects of acute postexercise chocolate milk consumption during intensive judo training on the recovery of salivary hormones, salivary SIgA, mood state, muscle soreness, and judo-related performance. *Appl. Physiol. Nutr. Metab.* **2015**, *40*, 1116–1122. [[CrossRef](#)]
126. Granger, D.A.; Schwartz, E.B.; Booth, A.; Curran, M.; Zakaria, D. Assessing dehydroepiandrosterone in saliva: A simple radioimmunoassay for use in studies of children, adolescents and adults. *Psychoneuroendocrinology* **1999**, *24*, 567–579. [[CrossRef](#)]
127. Morley, J.E.; Perry, H.M.; Patrick, P.; Dollbaum, C.M.; Kells, J.M. Validation of salivary testosterone as a screening test for male hypogonadism. *Aging Male* **2006**, *9*, 165–169. [[CrossRef](#)]
128. Kraemer, W.J.; Marchitelli, L.; Gordon, S.E.; Harman, E.; Dziados, J.E.; Mello, R.; Frykman, P.; Mccurry, D.; Fleck, S.J. Hormonal and growth factor responses to heavy resistance exercise protocols. *J. Appl. Physiol.* **1990**, *69*, 1442–1450. [[CrossRef](#)]
129. Viru, A.; Viru, M. Pre-conditioning of the performance in power events by endogenous testosterone: In memory of Professor Carmelo Bosco. *J. Strength Cond. Res.* **2005**, *19*, 6–8.
130. Tomasi, T.B. The discovery of secretory IgA and the mucosal immune system. *Immunol. Today* **1992**, *13*, 416–418. [[CrossRef](#)]
131. Salvi, S.; Holgate, S.T. Could the airway epithelium play an important role in mucosal immunoglobulin A production. *Clin. Exp. Allergy* **1999**, *29*, 1597–1605. [[CrossRef](#)] [[PubMed](#)]
132. Lamm, M.E. Interaction of antigens and antibodies at mucosal surfaces. *Annu. Rev. Microbiol.* **1997**, *51*, 311–340. [[CrossRef](#)]
133. Mackinnon, L.T.; Jenkins, D.G. Decreased salivary immunoglobulins after intense interval exercise before and after training. *Med. Sci. Sports Exerc.* **1993**, *25*, 678–683. [[CrossRef](#)]
134. Nieman, D.C. Immune function responses to ultramarathon race competition. *Med. Sport* **2009**, *13*, 189–196. [[CrossRef](#)]
135. Gleeson, M.; McDonald, W.A.; Cripps, W.A.; Pyne, D.B.; Clancy, R.L.; Fricker, P.A. The effect on immunity of long term intensive training in elite swimmers. *Clin. Exp. Immunol.* **1995**, *102*, 210–216. [[CrossRef](#)]
136. Mackinnon, L.T. Immunoglobulin, antibody, and exercise. *Exerc. Immunol. Rev.* **1996**, *2*, 1–35.
137. Asahi, Y.; Yoshikawa, T.; Watanabe, I.; Iwasaki, T.; Hasegawa, H.; Sato, Y.; Shimada, S.; Nanno, M.; Matsuoka, Y.; Ohwaki, M.; et al. Protection against influenza virus infection in polymeric Ig receptor knockout mice immunized intra nasally with adjuvant-combined vaccines. *J. Immunol.* **2002**, *168*, 2930–2938. [[CrossRef](#)] [[PubMed](#)]
138. Godfrey, R.J.; Madgwick, Z.; Whyte, G.P. The exercise-induced growth hormone response in athletes. *Sports Med.* **2003**, *33*, 599–613. [[CrossRef](#)]
139. Bardon, A.; Cedor, O.; Kollberg, H. Cystic fibrosis-like changes in saliva of healthy persons subjected to anaerobic exercise. *Clin. Chim. Acta* **1983**, *133*, 311–316. [[CrossRef](#)]