

miR-644a is a tumor cell-intrinsic mediator of sex bias in glioblastoma

Ellen S. Hong, Sabrina Z. Wang, András K. Ponti, Nicole Hajdari, Juyeun Lee[®], Erin E. Mulkearns-Hubert, Josephine Volovetz, Kristen E. Kay, Justin D. Lathia[®], Andrew Dhawan

All author affiliations are listed at the end of the article

*Corresponding Authors: Justin D. Lathia, Department of Cardiovascular and Metabolic Sciences, Lerner Research Institute, Cleveland Clinic, 9500 Euclid Avenue (NC10), Cleveland, Ohio 44195, USA (lathiaj@ccf.org); Andrew Dhawan, Department of Cardiovascular and Metabolic Sciences, Lerner Research Institute, Cleveland Clinic, 9500 Euclid Avenue (NC10), Cleveland, Ohio 44195, USA (dhawana@ccf.org).

Abstract

Background. Biological sex is an important risk factor for glioblastoma (GBM), with males having a higher incidence and poorer prognosis. The mechanisms for this sex bias are thought to be both tumor intrinsic and tumor extrinsic. MicroRNAs (miRNAs), key posttranscriptional regulators of gene expression, have been previously linked to sex differences in various cell types and diseases, but their role in the sex bias of GBM remains unknown.

Methods. We leveraged previously published paired miRNA and mRNA sequencing of 39 GBM patients (22 male, 17 female) to identify sex-biased miRNAs. We further interrogated a separate single-cell RNA-sequencing dataset of 110 GBM patients to examine whether differences in miRNA target gene expression were tumor cell-intrinsic or tumor cell extrinsic. Results were validated in a panel of patient-derived cell models.

Results. We identified 10 sex-biased miRNAs ($p_{\text{adjusted}} < .1$), of which 3 were more highly expressed in males and 7 more highly expressed in females. Of these, miR-644a was higher in females, and increased expression of miR-644a target genes was significantly associated with decreased overall survival (HR 1.3, $P = .02$). Furthermore, analysis of an independent single-cell RNA-sequencing dataset confirmed sex-specific expression of miR-644a target genes in tumor cells ($P < 10^{-15}$). Among patient-derived models, miR-644a was expressed a median of 4.8-fold higher in females compared to males.

Conclusions. Our findings implicate miR-644a as a candidate tumor cell-intrinsic regulator of sex-biased gene expression in GBM.

Key Points

- miR-644a is more highly expressed in female GBM patients.
- Lower miR-644a target gene expression is associated with improved overall survival.
- miR-644a target genes are higher in male GBM cells but not in other cell types.

Glioblastoma (GBM) is the most common primary malignant brain tumor in adults. Despite an increasing scientific understanding of disease biology,^{1,2} the median overall survival of patients with GBM is 17 months and has not improved since 2005.³ The varying mechanisms of a tumor cell's intrinsic resistance to therapy, such as transcriptional programs,⁴ are the subject of significant investigation.

Male biological sex has emerged as a key risk factor for both increased incidence and decreased survival in GBM.^{5,6} This sex bias is believed to arise from both tumor-intrinsic and tumor-extrinsic factors. Tumor-intrinsic sex differences include differences in enhancer landscapes, glutamine metabolism, and oncogenic transformation.^{7,8} Tumor-extrinsic factors contributing to sex bias include microglial activation and anti-tumor

Importance of the Study

MicroRNAs (miRNAs) are non-coding RNAs that regulate gene expression at the posttranscriptional level and were previously linked to glioblastoma (GBM) growth and therapeutic resistance. miRNAs play a role in the sex bias of various cell types and diseases, but how miRNAs contribute to sex differences in GBM is not well elucidated. We show that 10 miRNAs are differentially expressed between males and females and identify miR-644a as more

highly expressed in female GBM patients. Using single-cell RNA-seq data, we demonstrate that sex differences in miR-644a target gene expression are tumor cell-intrinsic. Likewise, decreased miR-644a target gene expression is associated with improved overall patient survival. Our findings reveal miR-644a as a novel sex-biased miRNA in GBM and a possible target for sex-specific precision therapies with limited collateral damage.

immunity.^{9,10} However, the mechanisms underlying these sex-specific differences remain unknown.

We investigated microRNAs (miRNAs) as possible mediators of tumor-intrinsic sex differences in GBM. miRNAs are 19- to 25-nucleotide single-stranded RNAs that negatively regulate the expression of target genes through base pairing to the 3' untranslated region of the mRNA transcript.¹¹ miRNAs are important for tumorigenesis, globally dysregulating the transcriptome in numerous cancer types.¹² In GBM, miRNAs, such as miR-7, have been shown to play key roles in tumor growth by repressing transcripts such as *EGFR*.¹³ Furthermore, miRNA profiles of tumors have been shown to reliably subtype GBM and predict prognosis^{14,15} highlighting their clinical utility.

The role of miRNAs is context- and cell-type-dependent, highlighting the need for miRNAs to be evaluated in specific cell types and diseases.¹⁶ Notably, over 100 miRNAs are encoded on the X chromosome, while only 2 miRNAs are encoded on the Y chromosome,¹⁷ which has led to studies of the association between miRNAs and sex differences. Additionally, both male and female sex hormones are known to influence miRNA expression.^{18,19} Sex differences in miRNA expression have been reported in multiple tissues, including lung and brain.^{20,21} Various diseases, including lupus and metabolic syndrome, also display a sex bias in miRNA expression.^{22,23}

Sex bias in miRNA expression has also been reported in multiple cancers. These cancer types include those that are sex-specific (prostate, uterine) or have a substantial sex bias in terms of incidence, such as breast.²⁴ However, cancers with a less pronounced sex bias, such as colorectal cancer, lung adenocarcinoma, and hepatocellular carcinoma,^{25,26} also exhibit a sex difference in miRNA expression. How miRNA expression differs between males and females with GBM and contributes to sex differences is unknown. Here, we leverage paired miRNA and mRNA sequencing of GBM samples to investigate sex-biased miRNA expression and report in a tumor-intrinsic manner the increased expression of miR-644a in biological females with associated repression of miR-644a target genes.

Materials and Methods

miRNA and mRNA Analysis Using Patient Data from Kashani et al. and Gliovis

We retrieved data from the Kashani et al.²⁷ and Gliovis,²⁸ which utilize data from the Cancer Genome Atlas (TCGA).²⁹ Briefly, Kashani and colleagues performed RNA

sequencing on 100 ng total RNA using the nCounter_Human_miRNA_Expression_Panel_Assay_Kit_H_miRNA_V3 or the nCounter_Human_PanCancer_pathway_panel (NanoString, Seattle, USA) spiked with 30 additional genes implicated in autophagy, epithelial mesenchymal transition, and DNA repair processes.

For the TCGA dataset, miR-644a target gene expression was obtained for the "TCGA GBM" cohort through the Gliovis web browser.²⁸ Additionally, matching clinical data for patients in the "TCGA_GBM" cohort was downloaded.

Differential Expression Analysis

Differential expression analysis of the miRNA sequencing was performed using DESeq2 (v1.30.1).³⁰ miRNAs with an adjusted *P*-value <.1 were selected as differentially expressed miRNAs. R v4.0.5 was used.

miRNA Gene Target Prediction

miRNA gene targets were predicted using miRNAatop (v1.24.0).³¹ The default settings of including all 5 possible target databases were used: DIANA version 5.0, Miranda 2010 release, PicTar 2005 release, TargetScan 7.1, and miRDB 5.0. The default minimum source number of 2 was used, and the union of all targets found was taken as the initial set of targets for a given miRNA. miR-644a target genes were further filtered by removing target genes whose expression had positive Spearman's correlations with miR-644a expression.

GO Analysis

Gene ontology (GO) analysis was performed using www.geneontology.org. The default settings of searching for "biological process" with species "*Homo sapiens*" were used.

Kaplan-Meier Curve

The Kaplan-Meier survival plot was generated using R package survminer (v0.4.9). The fit was performed on survival, in days, and biological sex of patient. The log rank test was used for significance calculations.

Survival Analysis

The Cox proportional hazard ratio was calculated using the R package survival (v3.2.10). The miR-644a target gene

expression score was defined as the mean of the log-normalized expression of negatively correlated miR-644a target genes. The variables included in the Cox model for overall survival include: miR-644a target gene expression score, biological sex, and age. miR-644a target gene expression score was calculated by taking the mean expression of the genes *MAKP9* and *PTPRR*.

Single-Cell RNA-Sequencing Data and Analyses

The “Core GBmap” was downloaded from <https://cellxgene.cziscience.com/collections/999f2a15-3d7e-440b-96ae-2c806799c08c>. Seurat (v4.3.0) was used to process the data. Default Seurat preprocessing steps were used, including `NormalizeData`, `FindVariableFeatures`, `ScaleData`, `RunPCA`, `FindNeighbors`, and `FindClusters`. Only cells that originated from a patient with known biological sex were kept. Only cell types that had at least 500 cells per sex were kept.

As above, miRNAatp was used to predict miR-644a targets. miR-644a target gene expression for each cell was retrieved, all genes with zero counts were removed from the analyses, and the mean expression of males and females was calculated across each cell type. The Wilcoxon rank-sum test was used to test for statistically significant differences between target expression in males and females on a cell type-specific basis.

To calculate z-scores for male to female miRNA target gene expression ratios, only genes that were expressed in at least 50% of all cells were retained. Next, the average log-normalized expression across all male and female cells for each miRNA and corresponding target gene was calculated. The target genes for each miRNA predicted using miRNAatp for Figure 5 are included as Supplementary Table S3. All above analyses were done using R v4.2.2.

Real-Time Quantitative PCR

RNA was extracted from cells following the standard RNeasy kit protocol (Qiagen; 74134), and concentrations were measured using a NanoDrop (ThermoFisher) spectrophotometer. cDNA was synthesized using qScript cDNA SuperMix (Quanta Biosciences; 101414-102). qPCR was performed using Fast SYBR Green Mastermix (Applied Biosystems; 01120793) and an Applied Biosystems QuantStudio 3. Primer sequences are available in Supplementary Table S5. During qPCR analysis, threshold cycle values were normalized to *LMNA*.

RNA-Sequencing Analysis

Expression of miR-644a targets in patient-derived GBM models was investigated using publicly available RNA-sequencing datasets. The L0, L1, and L2 data were retrieved from E-MTAB-13161.³² The HW1, PB1, RN1, SB2b, and WK1 data were retrieved from Supplemental Dataset 4.³³ The 23M and DI318 data were retrieved from GSE119834.³⁴ Each individual model was rank-normalized, and the top 10% of miR-644a gene targets were evaluated. The mean of the ranks for miR-644a gene targets was calculated per model, and a one-sided Wilcoxon test was performed.

Patient-Derived Cell Models

L0, L1, and L2 were provided by Dr. Brent Reynolds (University of Florida) and have been described previously.³⁵ DI318 cells were obtained from the Rose Ella Burkhardt Brain Tumor Center biorepository (Cleveland Clinic) and have been described previously.³⁶ 3691 and 3832 were received from Dr. Jeremy Rich (University of Pittsburgh) and have been described previously.³⁷ 23M was received from Dr. Erik Sulman (New York University) and has been described previously.³⁸ SB2b, WK1, PB1, RN1, MMK1, and HW1 were received from Dr. Andrew Boyd (University of Queensland) and have been described previously.³³

Cell Culture

All cells were grown at 37 °C with 5% CO₂. The L0, L1, L2, DI318, G3691, G3832, and GSC23 lines were grown in Neurobasal medium minus phenol red (Gibco) with 1X B-27 supplement (Gibco), 1 mmol/L sodium pyruvate, 2 mmol/L L-glutamine, 50 U/mL penicillin/streptomycin, 20 ng/mL hEGF, and 20 ng/mL hFGF2 (R&D Systems). The remaining cell lines were grown in DMEM-F12 (Cleveland Clinic Media Preparation Core), 50 U/mL penicillin/streptomycin, 1% N2 supplement (Thermo Fisher Scientific), 20 ng/mL EGF, and 20 ng/mL FGF-2 (R&D Systems). Cells were passaged regularly using Accutase (Stem Cell Technologies) and phosphate-buffered saline.

Data and Code Availability

All raw genomic data that were the source of analyses performed in this study have already been published. The matched miRNA and mRNA sequencing data were obtained from the authors of Kashani et al. 2023. The TCGA data obtained from GlioVis are available at <http://gliovis.bioinfo.cnio.es/>. The single-cell RNA-seq Seurat object is available at <https://cellxgene.cziscience.com/collections/999f2a15-3d7e-440b-96ae-2c806799c08c>. The RNA-sequencing data were previously published at E-MTAB-13161, GSE119834.

All code necessary to reproduce the analyses in the figures of this manuscript is available on GitHub at https://github.com/esh81/miR644a_sexbias_GBM.

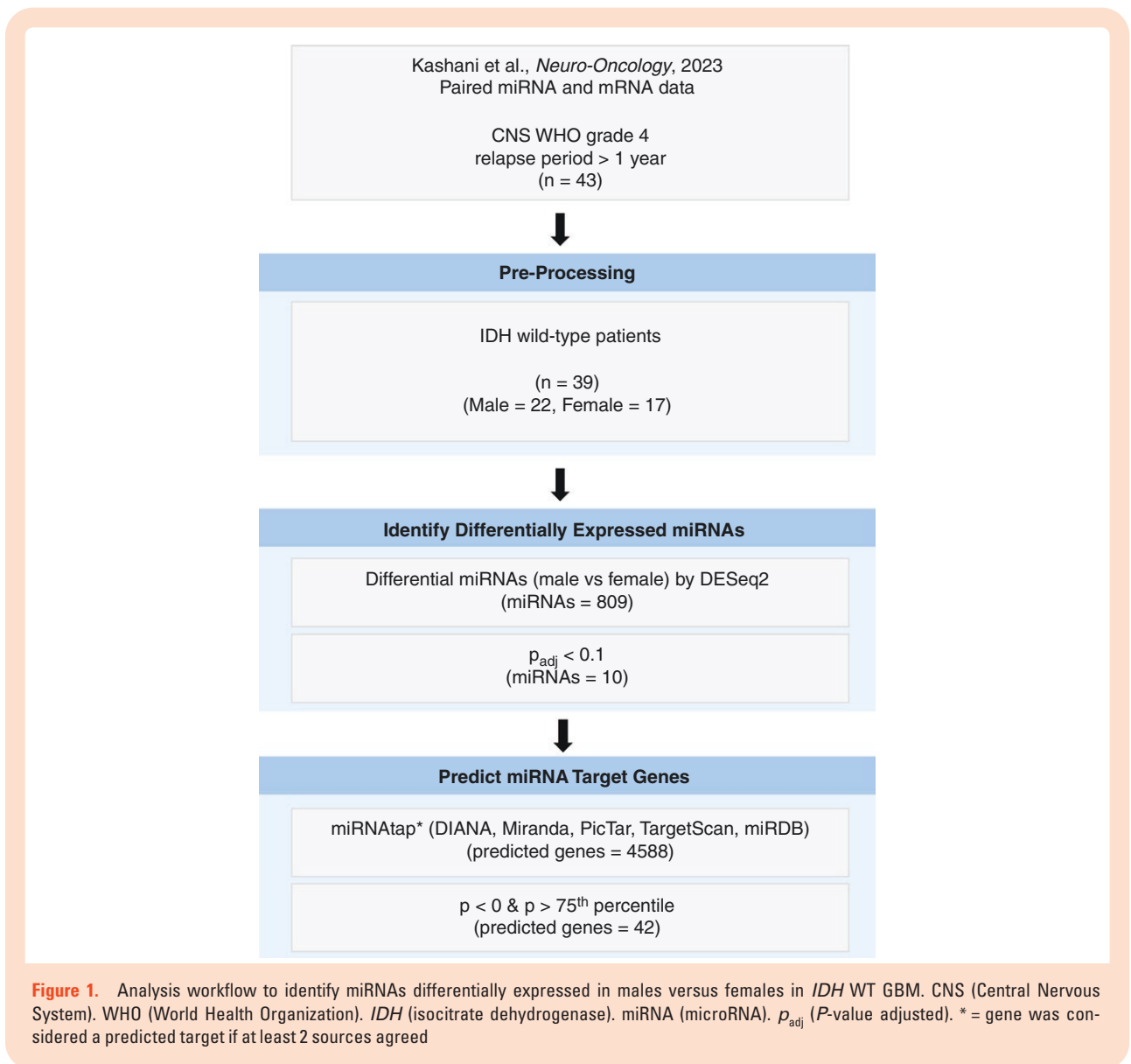
Ethics Statement

Studies were conducted in accordance with the Institutional Biosafety Committee of the Cleveland Clinic Foundation.

Results

Combined Sample Characteristics

For our comparative analysis, we leveraged tumor specimens from a cohort of 43 glioblastoma patients from the University of Bern during the years 1999–2016.²⁷ Importantly, this dataset contained matched mRNA and miRNA NanoString



sequencing on tissue specimens from this cohort. The cohort consisted of 39 *IDH* wild-type patients, of whom 22 were biological males (herein referred to males) and 17 were biological females (herein referred to females) (Supplementary Table S1). The median age at diagnosis was 57 years (range 41–79 years) (Supplementary Table S1).

To validate the findings from the Kashani et al. cohort in a larger and independent cohort, TCGA²⁹ database was utilized. Of the 538 patients, a total of 142 were *IDH* wild-type patients, with 93 male patients and 49 female patients (Supplementary Table S1). The median age at diagnosis was 62 years (range 24–89 years) (Supplementary Table S1).

miR-644a Shows Significantly Higher Expression in GBM Samples from Females

Differential expression analysis (Figure 1) of the 22 females and 17 males in the Kashani et al. dataset for miRNA

expression revealed statistically significant differences in 10 of 809 miRNAs (Figure 2A). Of these 10 microRNAs, 3 microRNAs were more highly expressed in males and 7 were more highly expressed in females. While the fold changes are relatively modest between samples, this effect was not driven by outliers. We chose to examine miR-644a further based on its previously reported differences in males versus females in several other cancers such as non-small cell lung cancer and hepatocellular carcinoma.^{39,40}

miR-644a Negatively Regulates Genes Associated With Cell Adhesion and Differentiation in GBM

We performed GO analysis on predicted miR-644a targets, highlighting its regulation of genes known to act in cell adhesion and development (Figure 2B). Among the predicted mRNA targets negatively correlated with miR-644a expression (Methods, Supplementary Table S2), *MAPK9*

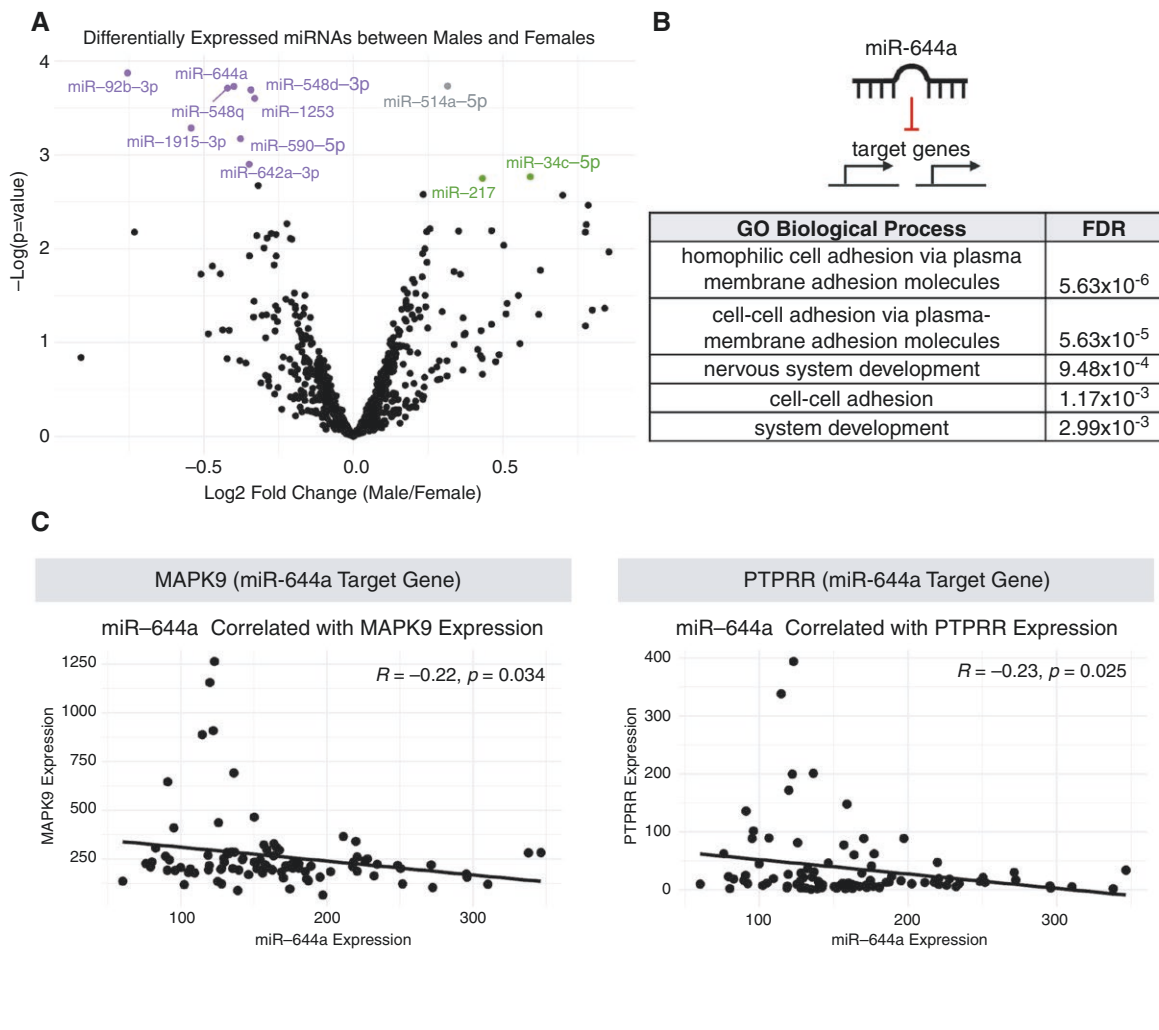


Figure 2. Differential expression analysis reveals sex-biased microRNAs. **(A)** Volcano plot showing results of DESeq2 analysis. miR-514a-5p was excluded from further analyses due to its genomic location on the X chromosome. **(B)** Top GO Biological Processes for target genes predicted to be regulated by miR-644a. FDR (False Discovery Rate). **(C)** Scatter plot of miR-644a expression in Kashani et al. dataset correlated with miR-644a target gene expression. (Left) MAPK9 and (right) PTPRR. R = Pearson's correlation.

and *PTPRR* emerged as the most significant candidate target genes negatively regulated by miR-644a (Figure 2C). *MAPK9* is a kinase involved in the JNK signaling pathway and is associated with prognosis in GBM,⁴¹ and *PTPRR* is a protein tyrosine phosphatase shown to inhibit *ERK1/2* in prostate cancer.⁴² For all downstream analyses, *MAPK9* and *PTPRR* were used as miR-644a target genes in GBM.

Increased miR-644a Target Gene Expression is Associated With Decreased Overall Survival

To determine if miR-644a is associated with outcome in GBM patients, we performed survival analyses. Firstly, in the Kashani et al. cohort, we showed that males survive shorter than females (male median survival = 30 months, female median survival = 36 months, HR 1.5 log rank, $P < .0001$) (Figure 3A). Additionally, we asked if miR-644a

expression independently of sex had potential to be useful for revealing univariate prognostic ability in GBM, but likely due to the small cohort size of the Kashani dataset, this did not reveal significance (Supplementary Figure S1). We then examined the distribution of miR-644a expression in males versus females and found that, as expected from the differential miRNA expression analysis (Figure 2A), females express 1.3-fold higher miR-644a compared to males (Figure 3B). Altogether, males in the Kashani et al. cohort have decreased miR-644a expression and reduced overall survival compared to females.

To validate the analyses from the Kashani et al. cohort, we assessed the larger TCGA cohort, which has 142 patients. We performed a Cox proportional hazard ratio analysis on *IDH* wild-type tumors, evaluating survival as it related to miR-644a target gene expression, summarized into the miR-644a score, fully detailed in the Methods (Figure 3C). Multivariate Cox proportional hazards analysis

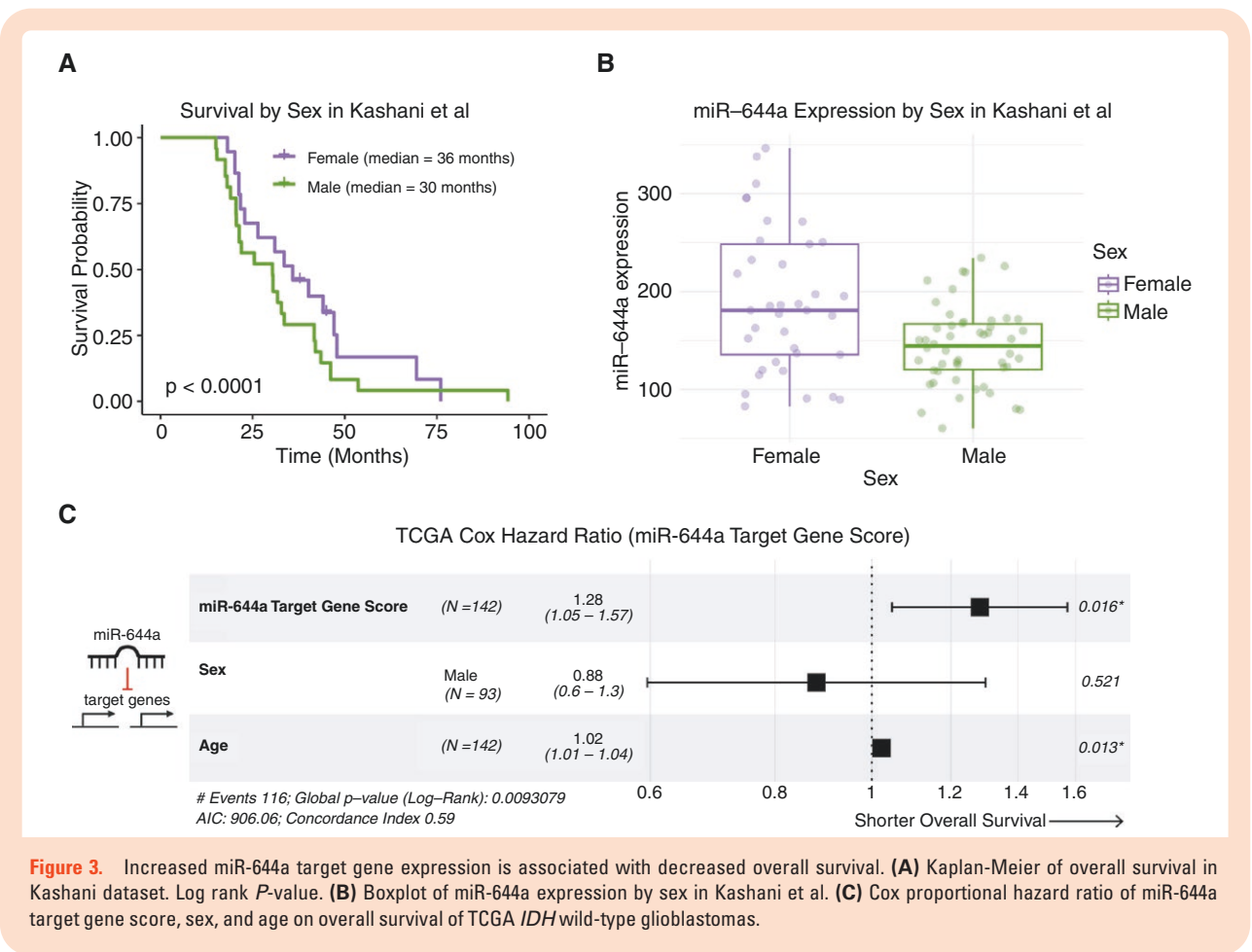


Figure 3. Increased miR-644a target gene expression is associated with decreased overall survival. **(A)** Kaplan-Meier of overall survival in Kashani dataset. Log rank P -value. **(B)** Boxplot of miR-644a expression by sex in Kashani et al. **(C)** Cox proportional hazard ratio of miR-644a target gene score, sex, and age on overall survival of TCGA *IDH* wild-type glioblastomas.

on TCGA samples revealed an association between decreased overall survival and increased miR-644a target gene expression (HR 1.3 (95% CI 1.1–1.6), $P = .02$) (Figure 3C). In other words, higher expression of miR-644a target genes, which implies lower miR-644a activity, is associated with worse survival. As males have reduced expression of miR-644a and greater expression of miR-644a target genes, this finding suggests that reduced miR-644a expression in males may contribute to their poorer overall survival.

miR-644a Target Genes are More Highly Expressed in Male Tumor Cells Compared to Nontumor Cells

We then surveyed miR-644a target gene expression in an independent single-cell RNA-seq dataset to ascertain how miR-644a target gene expression varied among the cellular subtypes within GBM. We utilized the publicly available single-cell RNA-seq resource GBmap, which curated over 300,000 single cells from 110 glioblastoma samples (27 females and 53 males). This resource annotated cells as one of 10 cell types, of which we retained 8 with sufficient representation across both males and females (Figure 4A).

Because miRNA was not quantified in these patients, we used the average expression of miR-644a target genes as a

surrogate for miRNA activity. At a single-cell level, we compared the average expression of miR-644a target genes in males versus females across each cell type (Figure 4B). Of all cell types, only malignant cells and resident brain cells, microglia, had significantly increased expression of miR-644a target genes in males (Figure 4B). In the remaining cell types, miR-644a target genes were more highly expressed in females compared to males. This suggests that miR-644a expression is driven in females by malignant cells, given the reduced expression of target genes in this cell subpopulation.

miR-644a Target Genes Are More Highly Expressed in Male Tumor Cells Relative to All Other miRNAs

To elucidate if male tumor cells specifically express miR-644a target genes more highly, we inspected the landscape of all miRNA target genes across cell types. For each cell type, we predicted miRNA target genes for every known miRNA in the genome and calculated the average expression in male and female cells (Figure 5A). We then normalized the distribution of male to female ratios for miRNA target genes per cell type (Figure 5B). Compared to target genes of all miRNAs, the z-score of miR-644a target

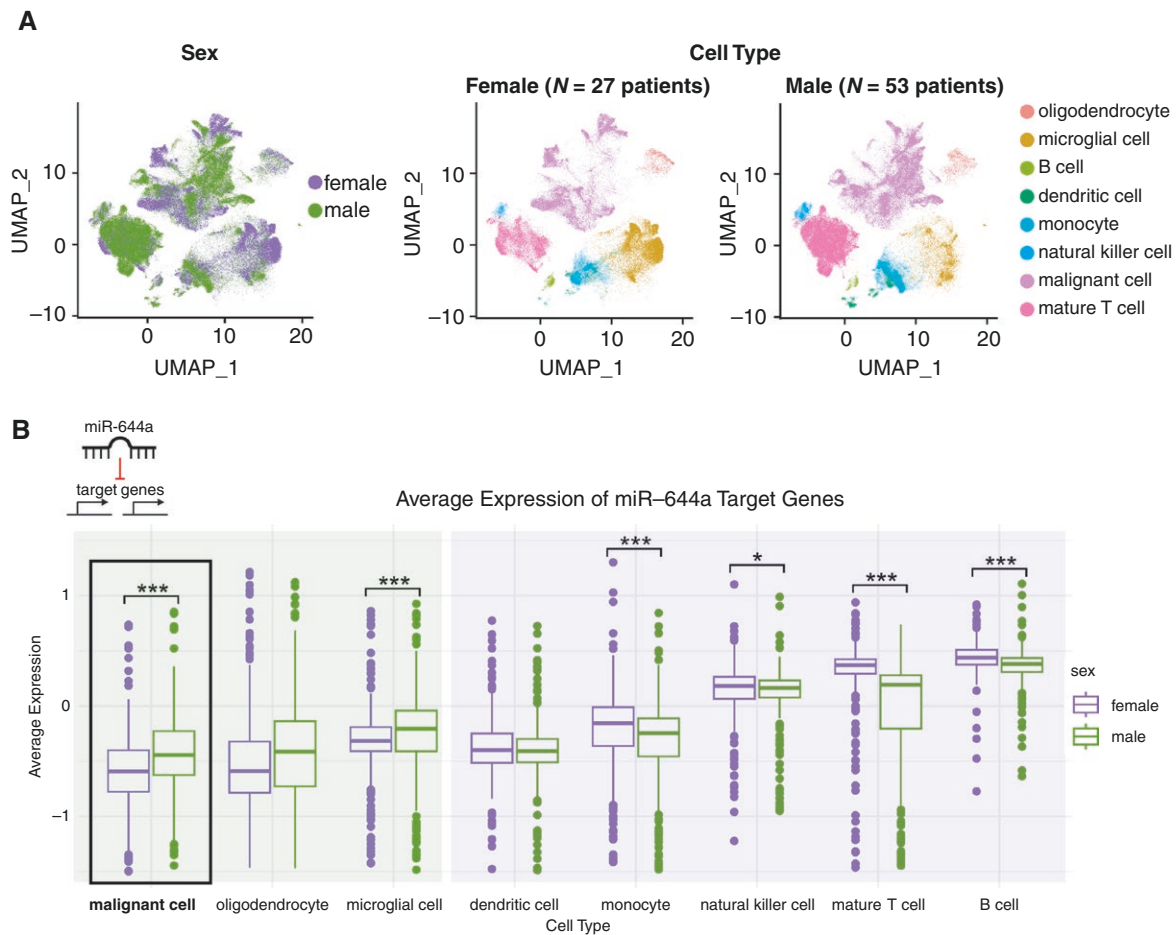


Figure 4. Male GBM cells more highly express miR-644a target genes. **(A)** UMAP plots of sex (left) and cell type (right) from GBmap. **(B)** Boxplot showing average expression (with zeros removed) of miR-644a target genes by cell type and sex in GBmap. Each point is one miR-644a target gene. *** P -value < .001 and * P -value < .05 using Wilcoxon test.

gene expression for males over females was positive only among malignant cells and oligodendrocytes (Figure 5C), again suggesting the sex bias in expression of miR-644a target genes is specific to primarily tumor cells.

miR-644a is More Highly Expressed in Female Patient-Derived Glioblastoma Models

Finally, we validated miR-644a expression in our cohort of patient-derived glioblastoma models. We extracted total RNA from 12 total patient-derived glioblastoma models and assayed miR-644a expression. While there is heterogeneity across the different models, overall, female models express on average 3.1-fold more miR-644a than male models (Figure 6A).

Moreover, we examined the expression of predicted miR-644a target genes in our patient-derived models and found, as expected, that females express miR-644a target genes at a lower level than males ($P < .05$) (Figure 6B). These results illustrate that miR-644a is more highly expressed in female patient-derived models, which have corresponding lower miR-644a target gene expression.

Discussion

miRNAs are key regulators of GBM growth, self-renewal, and therapeutic resistance. Here, we show that males and females with GBM differentially express a subset of miRNAs, and among these, miR-644a is more highly expressed in female patients in a tumor cell-intrinsic manner. Using the predicted target genes for miR-644a (*MAPK9* and *PTPRR*), we discover that higher expression of miR-644a target genes is associated with decreased overall survival. Using single-cell RNA-sequencing data, we demonstrate that miR-644a target genes are also differentially expressed in male and female malignant cells compared to other cell types. Finally, we validate the higher expression of miR-644a in a panel of female GBM patient-derived models relative to male patient-derived models. Taken together, our results across three independent datasets and 12 patient-derived models demonstrate that miR-644a is more highly expressed in females and may play a role in GBM sex bias.

Males have a higher incidence and poorer prognosis of GBM. However, the mechanisms of this sex bias are still

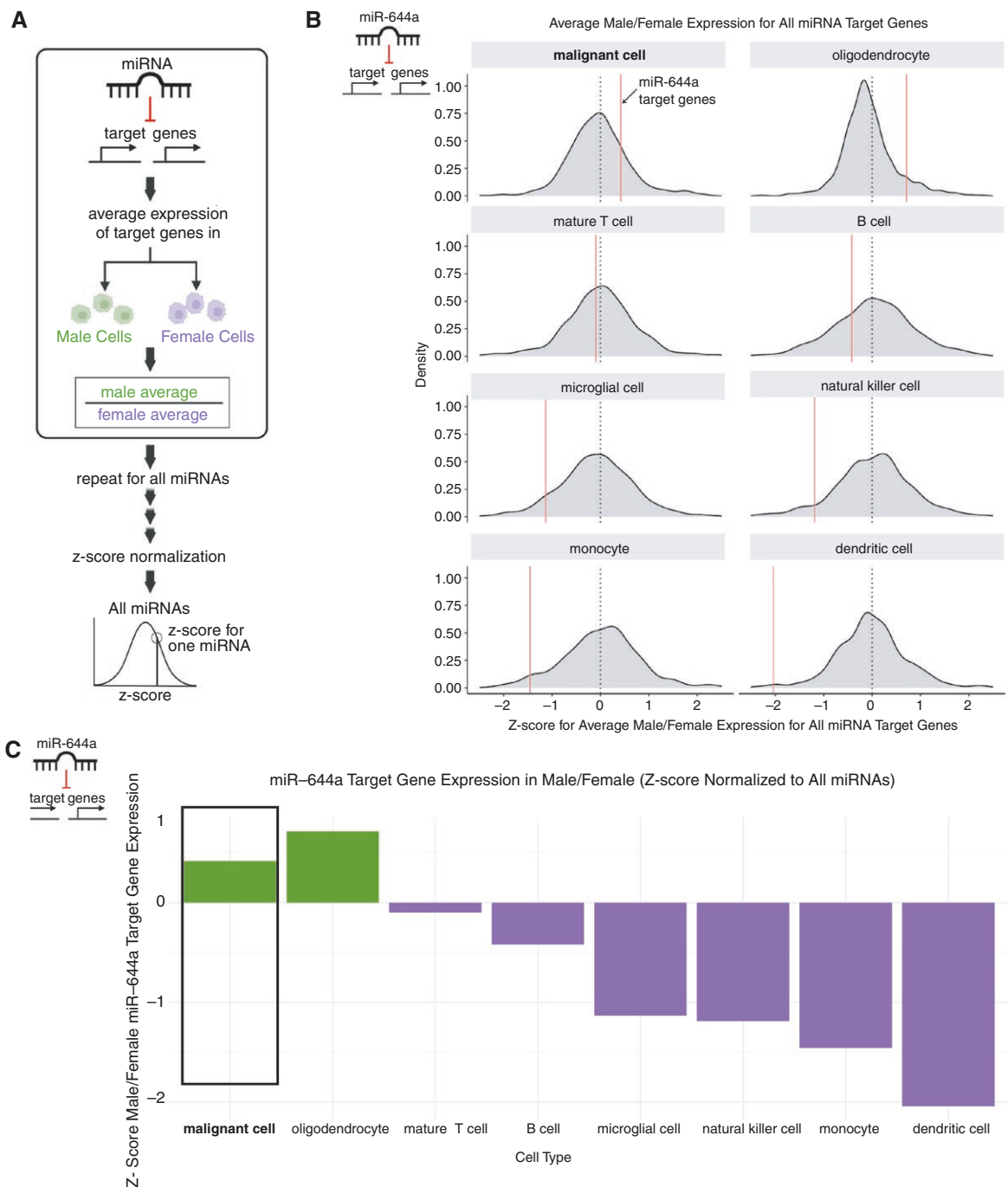


Figure 5. Male GBM cells more highly express miR-644a target genes relative to target genes of all other miRNAs. **(A)** Schematic of z-score normalization for all miRNA target genes in all cell types. **(B)** Z-score of distribution of male/female expression for all miRNAs. Red line shows average male/female expression for miR-644a target genes. **(C)** Bar plot showing miR-644a target gene expression between males and females as a z-score within each cell type.

an area of active investigation. miRNAs, as context-specific regulators of gene programs, warranted examination as a candidate mediator of sex-biased transcriptional differences. We demonstrated that increased expression of miR-644a target genes is associated with worse survival. The gene programs regulated by miR-644a are most

prominently cell adhesion and development, which have also been linked to sex differences in GBM⁴⁴⁴. Therefore, it is possible increased miR-644a expression in females leads to decreased GBM progression compared to males. Furthermore, miR-644a target genes were significantly overlapped with previously reported male-specific genes

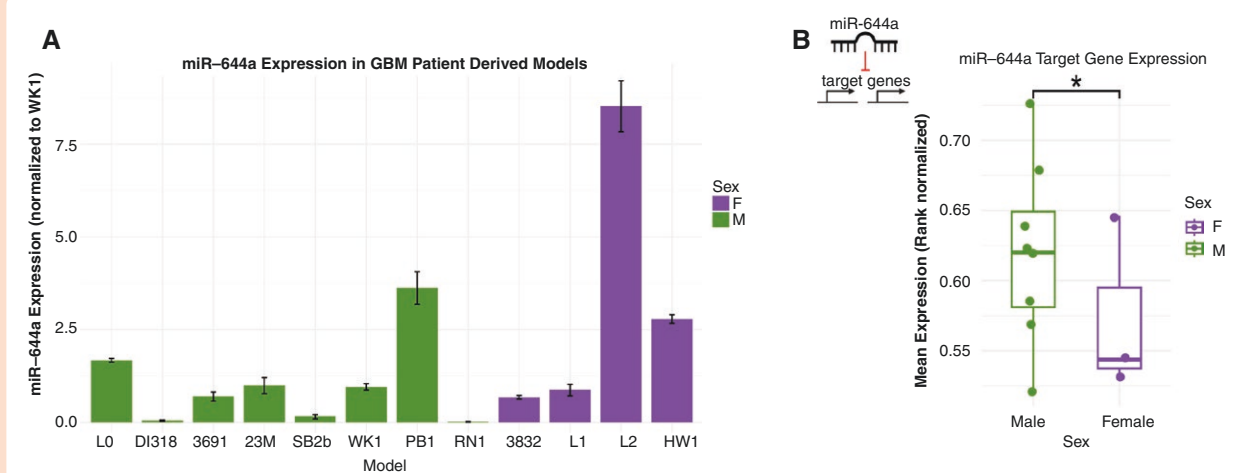


Figure 6. Female patient-derived GBM models more highly express miR-644a. qPCR of miR-644a in female and male patient-derived models. **(A)** Every $\Delta\Delta C^T$ is the average of 3 biological replicates (normalized to *LMNA*) and the fold change is normalized to WK1. **(B)** Boxplot representing the average expression per model of miR-644a target genes (rank-normalized per model). *One-sided Wilcoxon test $P < .05$.

but not female-specific genes,⁴³ supporting the function of miR-644a in repressing genes in females.

The mechanism behind the sex-biased expression of miR-644a remains unexplored. Unlike miRNAs encoded on the X chromosome that have been previously linked to sex differences, miR-644a is encoded on chromosome 20. Hence, the sex-biased expression of miR-644a is not entirely due to its chromosomal location. DNA methylation of miRNA promoters influences expression of the miRNA in various cancers such as colorectal cancer.⁴⁴ We therefore hypothesize that miR-644a may be differentially methylated in males versus females, contributing to its sex-specific expression, but this remains to be validated. Finally, aberrations in miRNA processing such as miRNA biogenesis and miRNA degradation can also affect miR-644a expression, but these have not yet been explored as contributors to sex differences in GBM.^{45,46}

There are limitations to our current study. In general, there is a scarcity of work investigating miRNAs due to the challenges in assaying their expression and function, which limits the feasibility of studies. There are no previous publications on differentially expressed miRNAs in males versus females with GBM. There are only a few findings in other cancer types such as non-small cell lung cancer, where males tended to have higher miR-644a expression,⁴⁰ and hepatocellular carcinoma, where males tended to have lower miR-644a expression.³⁹ The sparsity of available data underscores the importance of evaluating miRNA expression in specific contexts and disease and highlights the significance and novelty of our work.

Moreover, due to the lack of datasets with matched miRNA and mRNA sequencing, we were restricted to a small sample size. There is currently no published information on miR-644a targets in GBM, limiting us to using fewer predicted targets to not cloud our analyses with weaker statistical evidence. Although there have been some publications that have validated several miR-644a targets

in other cancers^{47,48}, true to the nature of the context-specificity of miRNAs, these previously validated targets are not statistically negatively correlated with miR-644a expression in GBM (Supplementary Table 4).

While Kashani and colleagues performed matched miRNA and mRNA sequencing, the sequencing was a targeted panel and did not assay the full transcriptome. Furthermore, as there is no miRNA information at the single-cell level, miRNA expression had to be extrapolated by examining target gene expression. Additionally, as protein expression may not correlate with miRNA or mRNA levels, the functional impact of these findings is yet to be determined.

Overall, these findings contribute molecular complexity to sex bias in GBM. miRNAs are attractive therapeutic targets because they utilize the cell's intrinsic machinery and act in context- and cell-type-specific ways. We showed that miR-644a target genes were increased in males compared to females in malignant cells, which highlights the potentially limited collateral damage to other cell types in the tumor. Thus, in summary, we demonstrate that miR-644a is a mediator of sex differences in GBM and a potential target for future tumor-intrinsic, sex-specific therapies.

Supplementary Material

Supplementary material is available online at *Neuro-Oncology Advances* (<https://academic.oup.com/noa>).

Keywords

miR-644a | miRNA | sex difference | tumor cell-intrinsic | glioblastoma

Lay Summary

Glioblastoma is a type of aggressive brain tumor that affects men more often than women, with women living longer than men. The researchers wanted to investigate whether certain genes might explain this difference. To do this, they looked at the amount of certain molecules that help control gene activity in samples from 39 patients with glioblastoma. Their results showed 10 of these molecules were different between men and women, with one called miR-644a being found at higher levels in women. They also found that higher amounts of this miR-644a molecule were linked to shorter survival.

Acknowledgments

The authors thank all members of the Lathia and Dhawan laboratories for thoughtful discussion and project conceptualization.

Conflict of interest statement

None declared.

Funding

National Institutes of Health grants R35 NS127083 (J.D.L.) and P01 CA245705 (J.D.L.). This work was also supported by the American Brain Tumor Association (J.L. and J.D.L.), American Academy of Neurology grant AAN2313JS (A.D.), Case Comprehensive Cancer Center (J.D.L.), and Cleveland Clinic/Lerner Research Institute (J.L., J.D.L., and A.D.).

Authorship Statement

E.S.H., J.D.L., and A.D. wrote the manuscript, with editing provided by J.L. and E.E.M.H. E.S.H., J.D.L., and A.D. designed the study, and E.S.H. performed primary data collection and analysis. S.Z.W., J.L., E.E.M.H., J.V., and K.E.K. assisted with experimental design. S.Z.W., A.K.P., N.H. assisted with the culture of patient-derived models and RNA extraction. S.Z.W., J.L., J.V., and K.E.K. assisted with interpretation of the data.

Affiliations

Department of Genetics and Genome Sciences, School of Medicine, Case Western Reserve University, Cleveland, OH, USA (E.S.H.); Medical Scientist Training Program (MSTP), School of Medicine, Case Western Reserve University, Cleveland, OH, USA (E.S.H., S.Z.W.); Department of Cardiovascular and Metabolic Sciences, Lerner Research Institute, Cleveland Clinic,

Cleveland, OH, USA (E.S.H., S.Z.W., A.K.P., N.H., J.L., E.E.M.-H., J.V., K.E.K., J.D.L., A.D.); Rose Ella Burkhardt Brain Tumor and Neuro-Oncology Center, Cleveland Clinic, Cleveland, OH, USA (J.D.L., A.D.); Department of Molecular Medicine, Cleveland Clinic Lerner College of Medicine of Case Western Reserve University, Cleveland, OH, USA (A.K.P., E.E.M.-H., J.V., K.E.K., J.D.L.); Case Comprehensive Cancer Center, Cleveland, OH, USA (J.D.L., A.D.); School of Medicine, Cleveland Clinic Lerner College of Medicine of Case Western Reserve University, Cleveland, OH, USA (A.D.)

References

1. Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med*. 2005;352(10):987–996.
2. Westphal M, Heese O, Steinbach JP, et al. A randomised, open label phase III trial with nimotuzumab, an anti-epidermal growth factor receptor monoclonal antibody in the treatment of newly diagnosed adult glioblastoma. *Eur J Cancer*. 2015;51(4):522–532.
3. Stupp R, Taillibert S, Kanner A, et al. Effect of tumor-treating fields plus maintenance temozolomide vs maintenance temozolomide alone on survival in patients with glioblastoma: a randomized clinical trial. *JAMA*. 2017;318(23):2306–2316.
4. Platten M, Wick W, Weller M. Malignant glioma biology: role for TGF-beta in growth, motility, angiogenesis, and immune escape. *Microsc Res Tech*. 2001;52(4):401–410.
5. Ostrom QT, Rubin JB, Lathia JD, Berens ME, Barnholtz-Sloan JS. Females have the survival advantage in glioblastoma. *Neuro Oncol*. 2018;20(4):576–577.
6. Ostrom QT, Gittleman H, Xu J, et al. CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2009–2013. *Neuro Oncol*. 2016;18(suppl_5):w1–v75.
7. Sponagel J, Jones JK, Frankfater C, et al. Sex differences in brain tumor glutamine metabolism reveal sex-specific vulnerabilities to treatment. *Med*. 2022;3(11):792–811.e12.
8. Kfoury N, Qi Z, Prager BC, et al. Brd4-bound enhancers drive cell-intrinsic sex differences in glioblastoma. *Proc Natl Acad Sci U S A*. 2021;118(16):e2017148118.
9. Lee J, Nicosia M, Hong ES, et al. Sex-biased t-cell exhaustion drives differential immune responses in glioblastoma. *Cancer Discov*. 2023;13(9):2090–2105.
10. Turaga SM, Silver DJ, Bayik D, et al. JAM-A functions as a female microglial tumor suppressor in glioblastoma. *Neuro Oncol*. 2020;22(11):1591–1601.
11. Mattick JS, Makunin IV. Non-coding RNA. *Hum Mol Genet*. 2006;15(Spec No. 1):R17–R29.
12. Dhawan A, Scott JG, Harris AL, Buffa FM. Pan-cancer characterisation of microRNA across cancer hallmarks reveals microRNA-mediated downregulation of tumour suppressors. *Nat Commun*. 2018;9(1):5228.
13. Kefas B, Godlewski J, Comeau L, et al. microRNA-7 inhibits the epidermal growth factor receptor and the Akt pathway and is down-regulated in glioblastoma. *Cancer Res*. 2008;68(10):3566–3572.
14. Srinivasan S, Patric IRP, Somasundaram K. A ten-microRNA expression signature predicts survival in glioblastoma. *PLoS One*. 2011;6(3):e17438.
15. Lages E, Guttin A, El Atifi M, et al. MicroRNA and target protein patterns reveal physiopathological features of glioma subtypes. *PLoS One*. 2011;6(5):e20600.

16. Carroll AP, Tooney PA, Cairns MJ. Context-specific microRNA function in developmental complexity. *J Mol Cell Biol*. 2013;5(2):73–84.
17. Ghorai A, Ghosh U. miRNA gene counts in chromosomes vary widely in a species and biogenesis of miRNA largely depends on transcription or post-transcriptional processing of coding genes. *Front Genet*. 2014; 5:100.
18. Bhat-Nakshatri P, Wang G, Collins NR, et al. Estradiol-regulated microRNAs control estradiol response in breast cancer cells. *Nucleic Acids Res*. 2009;37(14):4850–4861.
19. Wang WLW, Chatterjee N, Chittur SV, Welsh J, Tenniswood MP. Effects of 1 α ,25 dihydroxyvitamin D3 and testosterone on miRNA and mRNA expression in LNCaP cells. *Mol Cancer*. 2011;10:58.
20. Murphy SJ, Lusardi TA, Phillips JL, Saugstad JA. Sex differences in microRNA expression during development in rat cortex. *Neurochem Int*. 2014;77:24–32.
21. Lin NW, Liu C, Yang IV, et al. Sex-specific differences in microRNA expression during human fetal lung development. *Front Genet*. 2022;13:762834.
22. Hewagama A, Gorelik G, Patel D, et al. Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun*. 2013;41:60–71.
23. Wang YT, Tsai PC, Liao YC, Hsu CY, Juo SHH. Circulating microRNAs have a sex-specific association with metabolic syndrome. *J Biomed Sci*. 2013;20(1):72.
24. Guo L, Zhang Q, Ma X, Wang J, Liang T. miRNA and mRNA expression analysis reveals potential sex-biased miRNA expression. *Sci Rep*. 2017;7:39812.
25. Bai PS, Hou P, Kong Y. Hepatitis B virus promotes proliferation and metastasis in male Chinese hepatocellular carcinoma patients through the LEF-1/miR-371a-5p/SRCIN1/pleiotrophin/Slug pathway. *Exp Cell Res*. 2018;370(1):174–188.
26. Skjefstad K, Johannessen C, Grindstad T, et al. A gender specific improved survival related to stromal miR-143 and miR-145 expression in non-small cell lung cancer. *Sci Rep*. 2018;8(1):8549.
27. Kashani E, Schnidrig D, Gheinani AH, et al. Integrated longitudinal analysis of adult grade 4 diffuse gliomas with long-term relapse interval revealed upregulation of TGF- β signaling in recurrent tumors. *Neuro Oncol*. 2023;25(4):662–673.
28. Bowman RL, Wang Q, Carro A, Verhaak RGW, Squatrito M. GlioVis data portal for visualization and analysis of brain tumor expression datasets. *Neuro Oncol*. 2017;19(1):139–141.
29. Weinstein JN, Collisson EA, Mills GB, et al. The Cancer Genome Atlas Pan-Cancer analysis project. *Nat Genet*. 2013;45(10):1113–1120.
30. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol*. 2014;15(12):550.
31. Pajak M, Simpson TI. miRNAtap.db: microRNA targets-aggregated predictions database use. Published online 2014.
32. Alshahrany N, Begum A, Siebzehrubl D, Jimenez-Pascual A, Siebzehrubl FA. Spatial distribution and functional relevance of FGFR1 and FGFR2 expression for glioblastoma tumor invasion. *Cancer Lett*. 2023;571:216349.
33. Stringer BW, Day BW, D'Souza RCJ, et al. A reference collection of patient-derived cell line and xenograft models of proneural, classical and mesenchymal glioblastoma. *Sci Rep*. 2019;9(1):4902.
34. Mack SC, Singh I, Wang X, et al. Chromatin landscapes reveal developmentally encoded transcriptional states that define human glioblastoma. *J Exp Med*. 2019;216(5):1071–1090.
35. Deleyrolle LP, Harding A, Cato K, et al. Evidence for label-retaining tumour-initiating cells in human glioblastoma. *Brain*. 2011;134(Pt 5):1331–1343.
36. Sundar SJ, Shakya S, Barnett A, et al. Three-dimensional organoid culture unveils resistance to clinical therapies in adult and pediatric glioblastoma. *Transl Oncol*. 2022;15(1):101251.
37. Guryanova OA, Wu Q, Cheng L, et al. Nonreceptor tyrosine kinase BMX maintains self-renewal and tumorigenic potential of glioblastoma stem cells by activating STAT3. *Cancer Cell*. 2011;19(4):498–511.
38. Farooqi A, Yang J, Sharin V, et al. Identification of patient-derived glioblastoma stem cell (GSC) lines with the alternative lengthening of telomeres phenotype. *Acta Neuropathol Commun*. 2019;7(1):76.
39. Liang W, Liao Y, Li Z, et al. MicroRNA-644a promotes apoptosis of hepatocellular carcinoma cells by downregulating the expression of heat shock factor 1. *Cell Commun Signal*. 2018;16(1):30.
40. Wu Z, Jiang H, Fu H, Zhang Y. A circGLIS3/miR-644a/PTBP1 positive feedback loop promotes the malignant biological progressions of non-small cell lung cancer. *Am J Cancer Res*. 2021;11(1):108–122.
41. Yang X, Jia Q, Liu X, et al. MAPK9 is Correlated with a Poor Prognosis and Tumor Progression in Glioma. *Front Biosci (Landmark Ed)*. 2023;28(3):63.
42. Munkley J, Lafferty NP, Kalna G, et al. Androgen-regulation of the protein tyrosine phosphatase PTPRR activates ERK1/2 signalling in prostate cancer cells. *BMC Cancer*. 2015;15(1):9.
43. Yang W, Warrington NM, Taylor SJ, et al. Sex differences in GBM revealed by analysis of patient imaging, transcriptome, and survival data. *Sci Transl Med*. 2019;11(473):eaao5253.
44. Qin J, Ke J, Xu J, et al. Downregulation of microRNA-132 by DNA hypermethylation is associated with cell invasion in colorectal cancer. *Onco Targets Ther*. 2015;8:3639–3648.
45. Mehraein Y, Schmid I, Eggert M, Kohlhasse J, Steinlein OK. DICER1 syndrome can mimic different genetic tumor predispositions. *Cancer Lett*. 2016;370(2):275–278.
46. Segalla S, Pivetti S, Todoerti K, et al. The ribonuclease DIS3 promotes let-7 miRNA maturation by degrading the pluripotency factor LIN28B mRNA. *Nucleic Acids Res*. 2015;43(10):5182–5193.
47. Sikand K, Singh J, Ebron JS, Shukla GC. Housekeeping gene selection advisory: glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and β -actin are targets of miR-644a. *PLoS One*. 2012;7(10):e47510.
48. Ebron JS, Shankar E, Singh J, et al. MiR-644a disrupts oncogenic trans-formation and Warburg effect by direct modulation of multiple genes of tumor-promoting pathways. *Cancer Res*. 2019;79(8):1844–1856.