



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

2

Pediatric Healthcare Epidemiology

Jane D. Siegel and Judith A. Guzman-Cottrill

The reduction of healthcare-associated infections (HAIs) is an important component of patient safety programs. Five of the 16 Hospital National Patient Safety Goals for 2016 of The Joint Commission (formerly the Joint Commission on Accreditation of Healthcare Organizations) target prevention of HAIs.¹ Hospitals have learned from high-reliability organizations (e.g., the aviation industry) the importance of adopting changes that include the leadership's commitment to achieving zero patient harm, a fully functional culture of safety throughout the organization, and the widespread deployment of highly effective process improvement tools.² Involvement of new stakeholders for improving patient safety and outcomes related to HAIs (e.g., Children's Hospitals' Solutions for Patient Safety, Children's Hospital Association, individual states' mandatory HAI public reporting programs, the Centers for Medicare and Medicaid Services, The Joint Commission) has broadened the arena for HAI prevention efforts. Knowledge of the complexities of prevention and control of HAIs in children is critical to many different leaders of children's healthcare facilities. One framework for patient safety in children's hospitals that includes infection prevention and control (IPC) was developed by the Ohio Children's Hospital Solutions collaborative and demonstrates the effectiveness of hospitalwide collaboration.³ As more disciplines in healthcare become engaged in prevention of HAIs as well as antimicrobial stewardship, it is the responsibility of the healthcare epidemiologist and the IPC staff (infection preventionists, healthcare epidemiologists) to educate the facility leadership on the discipline of IPC.

IPC for the pediatric population is a unique discipline that requires understanding of various host factors, sources of infection, routes of transmission, behaviors required for care of infants and children, pathogens and their virulence factors, treatments, preventive therapies, and behavioral theory. Although the term *nosocomial* still applies to infections that are acquired in acute care hospitals, the more general term, *healthcare-associated infections* (HAIs), is preferred because much care of high-risk patients, including patients with medical devices (e.g., central venous catheters, ventilators, ventricular shunts, peritoneal dialysis catheters), has shifted to ambulatory settings, rehabilitation or chronic care facilities, and the home; thus, the geographic location of acquisition of the infection often cannot be determined.

The principles of transmission of infectious agents in healthcare settings and recommendations for prevention are reviewed in the Healthcare and Infection Control Practices Advisory Committee (HICPAC) Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Healthcare Settings, 2007⁴ and in the Management of Multidrug Resistant Organisms in Healthcare Settings, 2006 document.⁵ As new pathogens emerge, epidemiologists will continue to learn more about preventing transmission; therefore, for such pathogens, the most up-to-date guidance posted on the Centers for Disease Control and Prevention (CDC) or the World Health Organization (WHO) website should be consulted. The experience treating Ebola virus disease (EVD) in the United States in 2014 is the most recent example of changes in the usual infection prevention paradigm that were required, with emphasis on the hierarchy of controls⁶ and donning and doffing of personal protective equipment (PPE) with trained observers.⁷ A detailed discussion of HAIs can be found in Chapters 99 and 100. This chapter focuses on the components of an effective pediatric hospital epidemiology program.

RISK FACTORS FOR HEALTHCARE-ASSOCIATED INFECTIONS IN CHILDREN

Unique aspects of HAIs in children are summarized in the following sections. Specific risks and pathogens are addressed in several other chapters in this textbook.

Host or *Intrinsic* Factors

Intensive care units (ICUs), oncology services, and gastroenterology services caring for patients with short gut syndrome who are dependent on total parenteral nutrition (and lipids) have the highest rates of bacterial and fungal infection associated with central venous catheters. A newer definition of mucosal barrier injury laboratory-confirmed bloodstream infection (MBI-LCBI) currently is used by the National Healthcare Safety Network (NHSN) of the CDC to distinguish bacteremia that represents translocation of gut microorganisms related to mucosal barrier injury in patients with oncologic conditions, hematopoietic stem cell transplantation (HSCT), and intestinal failure from bacteremia associated with central venous catheters.⁸ HAIs can result in substantial morbidity and mortality, as well as lifetime physical, neurologic, and developmental disabilities. Host (i.e., intrinsic) factors that make children particularly vulnerable to infection include immaturity of the immune system, congenital abnormalities, and congenital or acquired immunodeficiencies. Children with congenital anomalies have a high risk of HAI if their unusual anatomic features predispose them to contamination of normally sterile sites. Moreover, these children require prolonged and repeated hospitalizations, undergo many complex surgical procedures, and have extended exposure to invasive supportive and monitoring equipment.

Innate deficiencies of the immune system in prematurely born infants, who may be hospitalized for prolonged periods and exposed to intensive monitoring, supportive therapies, and invasive procedures, contribute to the relatively high rates of infection in the neonatal ICU (NICU). All components of the immune system are compromised in neonates, and the degree of deficiency is proportional inversely to gestational age (see Chapter 9). The underdeveloped skin of the very low birth weight (<1000 g) infant provides another mode of pathogen entry.

Populations of immunosuppressed children have expanded with the advent of more intense immunosuppressive therapeutic regimens used for oncologic conditions, HSCT, solid-organ transplantation, and rheumatologic conditions and inflammatory bowel disease for which immunosuppressive agents and tumor necrosis factor- α -inhibiting agents (infliximab [Remicade]) and other immune modulators are used. Genetic mutations in the genes for the transmembrane conductance regulator (CFTR) in children with cystic fibrosis result in thick secretions, chronic endobronchial infections, and gastrointestinal malabsorption. Knowledge of the epidemiology of infection of patients with cystic fibrosis and effective methods to prevent patient-to-patient transmission have expanded with the use of newer molecular diagnostic methods, resulting in a 2013 update in the Infection Prevention and Control Guideline for Cystic Fibrosis.⁹ Fortunately, the population of children with perinatally acquired human immunodeficiency virus (HIV) infection and acquired immunodeficiency syndrome (AIDS) has decreased dramatically since 1994, but new cases of sexually transmitted HIV infection continue to be diagnosed in teens who receive care in children's hospitals. Finally, young infants who have not yet been immunized, or immunosuppressed children who do not respond to vaccines or who lose antibody during disease or treatment (e.g., patients with nephrotic syndrome), have increased susceptibility to vaccine-preventable diseases.

Sources or *Extrinsic* Factors

The source of many HAIs is the endogenous flora of the patient. An asymptotically colonizing pathogen can invade a patient's bloodstream or be transmitted to other patients on the hands of healthcare personnel (HCP) or on shared equipment. Other important sources of HAIs in infants and children include the mother in the case of neonates,

invasive monitoring and supportive equipment, blood products, total parenteral nutrition fluids, lipids, infant formula and human milk, HCP, and other contacts, including adult and sibling visitors. Maternal infection with *Neisseria gonorrhoeae*, *Treponema pallidum*, HIV, hepatitis B virus, parvovirus B19, *Mycobacterium tuberculosis*, herpes simplex virus, or group B *Streptococcus*, or colonization with multidrug-resistant organisms (MDROs), pose substantial threats to the neonate. During perinatal care, procedures such as fetal monitoring using scalp electrodes, fetal transfusion and surgical procedures, umbilical cannulation, and circumcision are potential risk factors for infection. Intrinsically contaminated powdered formulas and infant formulas prepared in contaminated blenders or improperly stored or handled have resulted in sporadic and epidemic infections in the nursery (e.g., *Cronobacter* [formerly *Enterobacter*] *sakazakii*), but such infections have become less frequent since the pathogenesis was defined and contamination reduced.¹⁰ Human milk that has been contaminated by maternal flora or by organisms transmitted through breast pumps has caused isolated serious infections and epidemics. The risks of neonatal hepatitis, cytomegalovirus (CMV) infection, and HIV infection from human milk warrant further caution for handling and use of banked breast milk. With increasing numbers of procedures being performed by pediatric interventional radiologists,¹¹ an understanding of appropriate aseptic technique, as well as prevention and management of infectious complications, by interventional radiologists is important.¹²

Construction, renovation, demolition, and excavation in and near healthcare facilities are important sources of environmental fungi, (e.g., *Aspergillus* spp., agents of mucormycoses, *Fusarium* spp., *Scedosporium* spp., *Bipolaris* spp.).¹³ Immunocompromised patients and patients in the pediatric ICU (PICU) and NICU are at greatest risk for fungal infection, and case fatality rates can be $\geq 50\%$, especially if diagnosis and treatment are delayed.

Practices Related to Care of Infants and Young Children. Several practices must be evaluated with respect to the potentially associated risk of infection. A significant association between reduced levels of nurse staffing and appropriately trained nurses has been demonstrated to increase risk of infection in many studies in both children and adults.^{4,14,15} Theoretical concerns exist that infection risk also will increase in association with the innovative practices of co-bedding of twins and kangaroo care in the NICU because of increased opportunity for skin-to-skin exposure of multiple-gestation infants to each other and to their mothers, respectively. Neither the benefits nor the safety of co-bedding multiple-birth infants in the hospital setting has been demonstrated.¹⁶ Overall, the infection risk is reduced with kangaroo care, but transmission of tuberculosis and respiratory syncytial virus (RSV) has occurred in kangaroo mother care units in South Africa.¹⁷ Parents providing kangaroo care should be monitored for the presence of skin infections.

Antimicrobial Selective Pressure. Exposure to vancomycin and to third-generation cephalosporins contributes substantially to the increase in infections caused by vancomycin-resistant *Enterococcus* (VRE)¹⁸ and multidrug-resistant gram-negative bacilli, including extended spectrum β -lactamase (ESBL)-producing organisms¹⁹ and carbapenem-resistant Enterobacteriaceae²⁰ (CRE) in children. Additionally, exposure to third-generation cephalosporins also is a risk factor for the development of invasive candidiasis in low birth weight infants in the NICU.²¹ Studies of the human microbiome using culture-independent methods have demonstrated the bacterial community diversity on mucosal surfaces and the profound suppressive effect of antimicrobial agents on the population of protective bacteria, *Firmicutes*, thus increasing the risk of colonization and subsequent invasive disease caused by pathogenic bacteria.²²

TRANSMISSION

Routes

The principal modes of transmission of infectious agents are direct and indirect contact, droplet, and airborne.⁴

Contact. Most infectious agents are transmitted by the contact route on the hands of HCP or through shared items; many pathogens can be transmitted by more than 1 route. Viruses, bacteria, and *Candida* spp. can be transmitted horizontally. Toddlers often share waiting rooms, playrooms, toys, books, and other items and therefore have the potential of transmitting pathogens directly and indirectly to one another. Contaminated bath

toys were implicated in an outbreak of multidrug-resistant *Pseudomonas aeruginosa* in a pediatric oncology unit.²³ Although the source of most *Candida* HAIs is the patient's endogenous flora, horizontal transmission, most likely through HCP hands, has been demonstrated in studies using typing by pulsed gel electrophoresis in the NICU and in a pediatric oncology unit.^{24,25} Newer molecular diagnostic methods (e.g., whole genome sequencing) are more sensitive and specific than pulsed gel electrophoresis and have proven to be valuable in identifying outbreaks of a variety of pathogens in both pediatric and adult settings.^{26,27}

Droplet. Infectious respiratory droplets $>5 \mu\text{m}$ in diameter are generated from the respiratory tract by coughing, sneezing, or talking or during such procedures as suctioning, intubation, chest physiotherapy, or pulmonary function testing. Transmission of infectious agents by the droplet route requires exposure of mucous membranes to large respiratory droplets within 3 to 6 feet (1 to 2 m) of the infected person. Large respiratory droplets do not remain suspended in the air for prolonged periods, and they settle on environmental surfaces. The dynamics of infectious aerosols can be affected by a variety of factors including characteristics of specific strains of bacteria, temperature, humidity, and number of air exchanges in a room. Adenovirus, influenza virus, and rhinovirus are transmitted primarily by the droplet route, whereas RSV is transmitted primarily by the contact route.²⁸ Although influenza virus can be transmitted by the airborne route under unusual conditions of reduced air circulation or low absolute humidity, ample evidence indicates that transmission of influenza is prevented by droplet precautions and, in the care of infants, the addition of contact precautions.²⁹

Airborne. Droplet nuclei that arise from desiccation of respiratory droplets and are $<5 \mu\text{m}$ in diameter and contain infectious agents remain suspended in the air for prolonged periods and travel long distances on air currents.⁴ Susceptible persons who have not had face-to-face contact or been in the same room as the source person can inhale such infectious particles. *M. tuberculosis*, varicella-zoster virus (VZV), and rubeola virus are the agents most frequently transmitted by the airborne route. Although transmission of *M. tuberculosis* by the airborne route can occur rarely from an infant or young child with active tuberculosis, the more frequent source is the adult visitor with active pulmonary tuberculosis that has not yet been diagnosed; thus screening of visiting family members is an important component for control of tuberculosis in pediatric healthcare facilities.³⁰

Some agents (e.g., severe acute respiratory syndrome–coronavirus [SARS-CoV]) can be transmitted as small-particle aerosols under special circumstances of aerosol-generating procedures (e.g., endotracheal intubation, bronchoscopy); therefore, an N95 or higher respirator is indicated for persons in the same airspace when these procedures are performed, but an airborne infection isolation room (AIIR) may not always be required. Roy and Milton³¹ proposed the following classification for aerosol transmission when evaluating routes of SARS-CoV transmission:

1. **Obligate:** Under natural conditions, disease occurs following transmission of the agent only through small-particle aerosols (e.g., tuberculosis).
2. **Preferential:** Natural infection results from transmission through multiple routes, but small-particle aerosols are the predominant route (e.g., measles, varicella).
3. **Opportunistic:** Agents naturally cause disease through other routes, but under certain environmental conditions they can be transmitted by fine-particle aerosols.

This conceptual framework can explain rare occurrences of airborne transmission of agents that are transmitted most frequently by other routes (e.g., smallpox, SARS, influenza, noroviruses). Concern about airborne transmission of influenza arose during the 2009 influenza A (H1N1) pandemic. However, the conclusion from all published experiences during the pandemic was that droplet transmission is the usual route of transmission, and surgical masks were noninferior to N95 respirators in preventing laboratory-confirmed influenza in HCP.^{32,33} Concerns about unknown or possible routes of transmission of agents that can cause severe disease and have no known treatment often result in more extreme prevention strategies. Therefore, recommended precautions could change as the epidemiology of emerging agents is defined and these controversial issues are resolved. Although no evidence supports airborne transmission of the Ebola virus under usual circumstances in the field, the aerosolization of body fluids that contain high titers of Ebola virus requires additional protection.³⁴

Healthcare Personnel

Transmission of microbes between children and HCP is a risk because of the very close contact that occurs during care of infants and young children and is facilitated by overcrowding, understaffing, and too few appropriately trained nurses in pediatric facilities.^{4,14} Staffing levels and composition are important components of an effective IPC program. HCP rarely are the source of outbreaks of HAIs caused by bacteria and fungi, but when they are, certain factors are usually present that increase the risk of transmission (e.g., sinusitis, draining otitis externa, respiratory tract infections, dermatitis, onychomycosis, wearing of artificial nails).^{35–37} Persons with direct patient contact who were wearing artificial nails have been implicated in outbreaks of *P. aeruginosa* and ESBL-producing *Klebsiella pneumoniae* in NICUs; therefore, the use of artificial nails or extenders is prohibited in persons who have direct contact with high-risk patients.⁴ Several published studies have shown that infected pediatric HCP, including resident physicians, transmitted *Bordetella pertussis* to other patients and can be the source of other vaccine-preventable infections in healthcare.^{38,39}

PATHOGENS

Although no agreed-on definition for what constitutes an “epidemiologically important organism” has yet been established, the following characteristics apply and are presented for guidance to infection control staff in the 2007 HICPAC Guideline for Isolation Precautions in Healthcare Settings⁴:

1. A propensity for transmission within healthcare facilities based on published reports and the occurrence of temporal or geographic clusters of infection in >2 patients (e.g., VRE, methicillin-resistant *Staphylococcus aureus* [MRSA], and methicillin-susceptible *S. aureus* [MSSA], *Clostridium difficile*, norovirus, RSV, influenza, rotavirus, *Enterobacter* spp., *Serratia* spp., group A *Streptococcus*). A single case of healthcare-associated invasive disease caused by certain pathogens (e.g., group A *Streptococcus* postoperatively or in burn units; *Legionella* sp.; *Aspergillus* sp.) should trigger an investigation.
2. Association with antimicrobial resistance (e.g., MRSA, VRE, ESBL-producing gram-negative bacilli (GNB), CRE, *Burkholderia cepacia*, *Ralstonia* spp., *Stenotrophomonas maltophilia*, and *Acinetobacter* spp.). Infections caused by intrinsically resistant GNB also suggest possible contamination of water or medication.
3. Association with serious clinical disease and increased morbidity and mortality (e.g., MRSA and MSSA, group A *Streptococcus*).
4. A newly discovered or reemerging pathogen (e.g., vancomycin-intermediate or vancomycin-resistant *S. aureus* [VISA, VRSA], *C. difficile*).

Pathogens associated with HAIs in children differ from those in adults in that respiratory viruses are more frequently associated with transmission in pediatric healthcare facilities. Respiratory viruses (e.g., RSV, parainfluenza, adenovirus, human metapneumovirus) have been implicated in outbreaks in high-risk units. As more respiratory viruses and gastrointestinal pathogens are identified by using highly sensitive molecular methods, epidemiologic studies will be required to define further the risk of transmission in healthcare facilities and the clinical significance of positive antigen detection test results.^{40,41} Healthcare-associated outbreaks of varicella, measles, and rotavirus infection now are rare events because of the consistent use of vaccines by children and HCP.

The emergence of community-associated MRSA isolates characterized by the unique *scc mec* type IV element was first observed among infants and children. As rates of colonization with community-associated MRSA at the time of hospital admission increased, so did transmission of community strains, most often USA 300, within the hospital and especially within the NICU, thus making prevention especially challenging. Other MDROs (e.g., VRE, ESBLs, and CRE, especially *K. pneumoniae*) have emerged as the most challenging healthcare-associated pathogens in both pediatric and adult settings, and otherwise healthy children in the community can be colonized asymptotically with these MDROs.⁴² GNB, including ESBL and other multidrug-resistant isolates, are more frequent than MRSA and VRE in many PICUs and NICUs. Patients who are transferred from chronic care facilities may be colonized with MDR GNB at the time of admission to the PICU. Trends in targeted MDROs are tracked in the National Nosocomial Infections Surveillance system

(NNIS), now NHSN ICUs. HAIs caused by MDROs are associated with increased length of stay, increased morbidity and mortality, and increased cost, in part because of the delay in initiating effective antimicrobial therapy.^{43,44} Although the prevalence of specific MDROs is lower in pediatric institutions, the same principles of target identification and interventions to control MDROs apply in all settings.

C. difficile is an important pathogen in children, as it is in adults, especially in children receiving chemotherapy. Testing for *C. difficile* in the first year of life is not advised because of the high asymptomatic colonization rate with toxigenic strains in this age group.

Candida spp. are the third most frequent pathogens associated with bloodstream infections in US NICUs. There is considerable center-to-center variability in both the incidence of invasive candidiasis and the proportion of *Candida* infections caused by *Candida non-albicans* spp., most of which are resistant to fluconazole. Risk factors for *Candida* infections include prolonged length of stay in an ICU, use of central venous catheters, intralipids, histamine (H₂)-blocking agents, and exposure to third-generation cephalosporins. GNB and *Candida* spp. are especially important pathogens for HAIs in patients with intestinal failure who are receiving total parenteral nutrition, and these organisms can cause repeated episodes of sepsis. The incidence of *Candida* infections had increased in incidence in most PICUs and NICUs during the 1990s, but the rate of *C. albicans* and non-*albicans* central line-associated bloodstream infections decreased by 75% in all birth weight categories from 1999 to 2009,⁴⁵ likely a result of improved infection control practices, antimicrobial stewardship, and use of fluconazole prophylaxis in the very low birth weight preterm infants. The most recently published clinical practice guidelines of the Infectious Diseases Society of America (IDSA) recommend the use of oral or intravenous fluconazole prophylaxis in infants weighing <1000 g at birth in NICUs with high rates (>10%) of invasive candidiasis, based on high quality of evidence to support efficacy and safety.⁴⁶ Additionally, empiric antifungal therapy in preterm infants of ≤1000 g birth weight is associated with improved survival rates without adverse outcomes.⁴⁷ The staff members of each NICU first must optimize infection control practices and then assess the remaining risk of *Candida* infections. Finally, environmental fungi (e.g., *Aspergillus*, *Fusarium*, *Scedosporium*, *Bipolaris*, agents of mucormycosis) are important sources of infection for severely immunocompromised patients; meticulous attention to the conditions of the internal environment of any facility that provides care for severely immunocompromised patients is required, as well as prevention of possible exposure to construction dust in and around healthcare facilities.¹³ With the advent of more effective and less toxic antifungal agents and improved outcomes, it is important to identify promptly the infecting agent by obtaining tissue samples and to determine susceptibility to candidate antifungal agents.

PREVENTION PROGRAMS

Prevention remains the mainstay of infection control and requires special considerations in children. The goals of IPC are to prevent the transmission of infectious agents among individual patients or groups of patients, visitors, and HCP who care for them. As new pathogens emerge, new strategies for prevention emerge. The experience treating EVD in the US in 2014 and 2015 is the most recent example of changes in the usual infection prevention paradigm that were required, with a renewed emphasis on the 3 tiers of the hierarchy of controls (e.g., engineering, administration, and PPE), donning and doffing of PPE, and use of trained observers.^{6,7} If prevention cannot always be achieved, the strategy of early diagnosis, treatment, and containment is critical.

A series of IPC guidelines have been developed and updated at varying intervals by the HICPAC/CDC, IDSA, Society for Healthcare Epidemiology of America (SHEA), American Academy of Pediatrics, Association for Professionals in Infection Control and Epidemiology, and others to provide evidence-based and rated recommendations for practices that are associated with reduced rates of HAIs, especially those infections associated with the use of medical devices and surgical procedures. Recommended isolation precautions by infectious agent also can be found in the most recent edition of the *Red Book Report of the Committee on Infectious Diseases* of the American Academy of Pediatrics.

Prevention *bundles* are groups of 3 to 5 evidence-based “best practices” with respect to a process that individually improve care, but when applied together result in substantially greater reduction in infection

rates. Adherence to the individual measures within a bundle is readily measured. Bundled practices are used most frequently for prevention of device- or procedure-related HAIs, but they can be applied to prevention of any type of HAI.

Administrative Factors

The importance of certain administrative measures for a successful IPC program has been demonstrated. A white paper published by SHEA summarizes the necessary infrastructure for an effective IPC program in modern times. The paper addresses the expansion of IPC responsibilities from a relatively narrow focus on acute infectious disease events in the acute care hospital, surveillance, and implementation of recommended isolation precautions to a broader set of activities across the continuum of care requiring team work within and beyond individual facilities, usually including large networks.⁴⁸ Because IPC comprises one component of the institutional culture of safety, it is critical to obtain support from the senior leadership of healthcare organizations to provide necessary fiscal and human resources for a proactive, successful IPC program. Critical elements requiring administrative support include access to the following: (1) appropriately trained healthcare epidemiologists and IPC personnel; (2) clinical microbiology laboratory services needed to support infection control outbreak investigations, including ability to perform molecular diagnostic testing; (3) data-mining programs and information technology specialists; (4) multidisciplinary programs to ensure judicious use of antimicrobial agents and control of resistance; (5) development of effective educational information for delivery to HCP, patients, families, and visitors; and (6) local and state health department resources for preparedness. Provision of adequate numbers of well-trained infection preventionists and bedside nursing staff is critical for success.

Infection Prevention and Control Team

An effective IPC program improves safety of patients and HCP and decreases short-term and long-term morbidity, mortality, and healthcare costs.⁴⁹ The IPC committee of a facility establishes policies and procedures to prevent or reduce the incidence and costs associated with HAIs. This committee should be one of the strongest and most accessible committees in the facility; committee composition should be considered carefully and limited to active, authoritative participants who have well-defined committee responsibilities and who represent major groups within the hospital. The chairperson should be a good communicator with expertise in IPC issues, healthcare epidemiology, and clinical pediatric infectious diseases. Important functions of the IPC committee are regular review of IPC policies and development of new policies as needed. Annual review of all policies is required by The Joint Commission and can be accomplished optimally by careful review of a few policies each month. With the advent of unannounced inspections, a constant state of readiness is required.

The hospital epidemiologist or medical director of the pediatric IPC department usually is a physician with training in pediatric infectious diseases and dedicated expertise in healthcare epidemiology. In multi-specialty medical centers where infants and children comprise a small proportion of patients, pediatric infectious disease experts should be consulted for management of pediatric IPC issues and report to the broader IPC leadership. The skillsets, training, and competencies needed for success as a healthcare epidemiologist were summarized in another white paper published by the SHEA.⁵⁰ Certification for healthcare epidemiologists has not yet been developed.

Infection preventionists are specialized professionals with advanced training, and preferably certification, in IPC. Although most infection preventionists are registered nurses, other professionals, including microbiologists, medical technologists, pharmacists, and epidemiologists, are successful in this position. Pediatric patients should have infection preventionist services provided by professionals with expertise and training in the care of children. In a large, general hospital, at least 1 infection preventionist should be dedicated to IPC services for children. The responsibilities of infection preventionists have expanded greatly and include the following:

1. Surveillance and IPC in facilities affiliated with primary acute care hospitals (e.g., ambulatory clinics, day-surgery centers, long-term care facilities, rehabilitation centers, home care) in addition to the primary hospital
2. Oversight of occupational health services related to IPC (e.g., assessment of risk and administration of recommended prophylaxis following exposure to infectious agents, tuberculosis screening, influenza and pertussis vaccination, respiratory protection fit testing, administration of other vaccines as indicated during infectious disease crises such as preexposure smallpox vaccine in 2003 and pandemic influenza A [H1N1] vaccine in 2009)
3. Preparedness planning for annual influenza outbreaks, pandemic influenza, SARS, Middle East respiratory syndrome (MERS), bio-weapons attacks, and EVD
4. Adherence monitoring for selected IPC practices
5. Oversight of risk assessment and implementation of preventive measures associated with construction, renovation, and other environmental conditions associated with increased infection risk
6. Participation in antimicrobial stewardship programs, focusing on prevention of transmission of MDROs
7. Evaluation of new products and medical devices that could be associated with increased infection risk (e.g., endoscopes,⁵¹ contaminated injectable medications⁵²) and introduction and assessment of performance after implementation of modified products
8. Mandatory public reporting of HAI rates in states according to enacted legislation
9. Increased communication with the public and with local public health departments concerning infection control-related issues
10. Participation in local and multicenter reporting and research projects

IPC programs must be adequately staffed to perform all the foregoing activities. Thus the ratio of 1 infection preventionist to 250 beds that was associated with a 30% reduction in the rates of nosocomial infection in the Study on Efficacy of Nosocomial Infection Control (SENIC) performed in the 1970s no longer is sufficient because the complexity of patient populations and responsibilities have increased. Many experts recommend that a ratio of 1 infection preventionist to 100 beds is more appropriate for the current workload, but no study has been performed to confirm the effectiveness of that ratio. No information is available on the number of IPC personnel required outside acute care, but it is clear that persons well trained in IPC must be available for all sites where healthcare is delivered. Data collected from a member workforce survey conducted in 2015 by the Association for Professionals in Infection Control and Epidemiology are expected to help determine the optimal number of infection preventionists for different healthcare settings based on the current responsibilities and demographics of infection preventionists.

Surveillance

Facilitywide or Systemwide Surveillance

Surveillance for HAIs consists of a systematic method of determining the incidence and distribution of infections acquired by hospitalized patients. The CDC recommends the following: (1) prospective surveillance on a regular basis by trained infection preventionists, using standardized definitions; (2) analysis of infection rates using established epidemiologic and statistical methods (e.g., calculation of rates using appropriate denominators that reflect duration of exposure; use of statistical process control charts for trending rates); (3) regular use of data in decision making; and (4) employment of an effective and trained healthcare epidemiologist who develops IPC strategies and policies and serves as a liaison with the medical community and administration.⁵³⁻⁵⁵ The CDC has established a set of standard definitions of HAIs that have been validated and accepted widely with updates posted on the CDC NHSN website. Standardization of surveillance methodology has become especially important with the advent of state legislation for mandatory reporting of HAI rates to the public. The NHSN now receives, analyzes, and reports data from >17,000 healthcare facilities in the US. A standardized infection ratio (SIR) that takes into account differences in risk among healthcare settings, unit types, procedures, and patient populations has been included in summary reports of HAI rates since 2009.⁵⁶ The Centers for Medicare and Medicaid Services and most states use the NHSN data for public reporting of HAI rates on their websites. Although much effort has been directed toward making these data understandable and useful to consumers, interpretation of

BOX 2.1 Sources of Data for Surveillance

Clinical rounds with physicians or nurses, or both

Review of:

- Patients' orders
- Radiology reports and databases
- Pharmacy reports and databases
- Operating room diagnoses and procedures
- Microbiology: bacteriology, virology, mycology, mycobacteriology, serology reports, autopsy reports, data-mining reports

Postdischarge surveillance, especially for surgical site infections

Public health surveillance

Review of:

- Employee health reports
- Admission diagnoses
- Outpatient diagnoses
- Administrative databases, but these should not be used as a sole source because of inaccurate coding of healthcare-associated infections

these data by the public remains difficult, and more research is needed to optimize methods of data display to the public.⁵⁷ New York State is the first state to have published an improvement in process and outcomes of central line-associated bloodstream infection rates in NICUs following implementation of a public reporting program.⁵⁸

Although various surveillance methods are used, the basic goals and elements are similar and include using standardized definitions of infection, finding and collecting cases of HAIs, tabulating data, using appropriate denominators that reflect duration of risk, analyzing and interpreting the data, reporting important deviations from endemic rates (epidemic, outbreaks) to the bedside care providers and to the facility administrators, implementing appropriate control measures, auditing adherence rates for recommended processes, and assessing efficacy of the control measures. Medical centers can use different methods of surveillance, as outlined in Box 2.1. Most experts agree that a combination of methods enhances surveillance and reliability of data, and some combination of clinical chart review and database retrieval is important. Whatever data collection systems are used, validation is required. Administrative databases created for the purposes of billing should not be used as the sole source to identify HAIs because of overestimates and underestimates that result from inaccurate coding of HAIs.⁵⁹ Use of software designed specifically for IPC data entry and analysis facilitates real-time tracking of trends and timely intervention when clusters are identified. The IPC team should participate in the development and update of electronic medical record systems for a healthcare organization, to ensure that surveillance needs will be met.

Targeted Pathogen-Specific Active Surveillance Cultures

Controversy has surrounded the role of obtaining active surveillance cultures from all patients admitted to an acute care hospital, especially to an ICU, to detect asymptomatic colonization with MRSA or VRE and then placing those persons on Contact Precautions in an endemic setting, a practice referred to as a *vertical* approach.^{60,61} More recently published experiences demonstrate the benefits of a *horizontal* approach to reduce the risk of transmission of a broader variety of pathogens,⁶¹ and a framework for a less restrictive approach has been published.⁶² Contributing factors to the benefits of the horizontal approach include the following: (1) widespread implementation of bundled prevention practices, including limiting use of unnecessary medical devices; (2) improved understanding and more consistent implementation of Standard Precautions, especially hand hygiene; (3) establishment of the safety and efficacy of universal decolonization using chlorhexidine bathing in ICUs^{63,64} and NICUs for infants weighing >1000 g at birth⁶⁵; (4) improving environmental cleaning; and (5) promoting antimicrobial stewardship. A program of active surveillance cultures and Contact Precautions is best reserved for implementation in a targeted fashion (i.e., in units with an indication of ongoing transmission of MRSA, VRE, or other MDROs)

according to 2006 guidelines, if transmission continues after standardized horizontal interventions have been completely implemented.³ At this time, no formal recommendation has been made to discontinue routine use of Contact Precautions for patients with asymptomatic colonization with MRSA or VRE in an endemic setting; thus each IPC program must determine practice based on local conditions and follow with close auditing and surveillance for potential adverse outcomes.

The microbiology laboratory can provide online culture information about individual patients, outbreaks of infection, antibiograms (antibiotic susceptibility patterns of pathogens summarized periodically), and employee infection data. The laboratory also can assist with surveillance cultures and facilitation of molecular typing of isolates during outbreak investigations. Rapid diagnostic testing of clinical specimens for identification of respiratory and gastrointestinal tract viruses and *B. pertussis* is especially important for pediatric facilities. The IPC division and the microbiology laboratory must communicate daily because even requests for cultures or other diagnostic testing from physicians (e.g., *M. tuberculosis*, *Neisseria meningitidis*, *C. difficile*) can identify patients early who are infected, are at high risk of infection, or require isolation. If microbiology laboratory work is outsourced, it is important to ensure that the services needed to support effective ICP be available, as delineated in a 2013 guideline developed by the IDSA and the American Society for Microbiology.⁶⁶

Control of unusual infections or outbreaks in the community generally is the responsibility of the local or state public health department; however, the individual facility must be responsible for preventing transmission within that facility. Public health agencies can be helpful, particularly in alerting hospitals of community outbreaks so that outpatient and inpatient diagnosis, treatment, necessary isolation, and other preventive measures are implemented promptly to avoid further spread. Conversely, designated persons in the hospital must notify public health department personnel of reportable infections to facilitate early diagnosis, treatment, and infection control in the community. Benefits of community or regional collaboratives of individual healthcare facilities and local public health departments for prevention of HAIs, especially those caused by MDROs, have been demonstrated, and this collaboration should be encouraged.^{4,67}

Antimicrobial Stewardship

The rapid increase of MDROs is a public health threat. Between 20% and 50% of antibiotics prescribed in US hospitals are either inappropriate or unnecessary.⁶⁸ In 2014, the President's Council of Advisors on Science and Technology submitted a 78-page *Report to the President on Combating Antibiotic Resistance* that raised awareness of antimicrobial resistance to a national level.⁶⁹ A National Action Plan based on this report was released in March 2015, and funding was made available for its implementation.

Antimicrobial stewardship was defined in a consensus statement by the IDSA, SHEA, and Pediatric Infectious Diseases Society in 2012 as "coordinated interventions designed to improve and measure the appropriate use of antibiotic agents by promoting the selection of the optimal antibiotic regimen, including dosing, duration and route of administration."⁷⁰ Antimicrobial stewardship programs are collaborative partnerships among infection preventionists, healthcare epidemiologists, clinical pharmacists, and microbiologists. Hospital administrative support for the infrastructure required for ongoing measurement and reporting of antimicrobial use and other related outcome measures, including feedback to prescribers, is a critical component of a successful antimicrobial stewardship program. An antimicrobial stewardship program can optimize clinical outcomes while decreasing unintended consequences of antimicrobial use, including the emergence of resistant organisms. Additionally, use of specific antimicrobial agents can alert the IPC program to the presence of potentially infectious patients (e.g., with pulmonary tuberculosis, MDROs). National guidelines exist for developing and implementing an institutional antimicrobial stewardship program, including core components for acute care hospitals and for long-term care facilities.^{68,70} The National Quality Forum and its partners have also developed a Playbook that provides additional guidance for implementation of antimicrobial stewardship programs in acute care. The knowledge and skills required for antimicrobial stewardship leaders also have been defined.^{50,71}

The effectiveness of antimicrobial stewardship programs in achieving improved patient outcomes is evident in pediatric acute care hospitals,^{72,73} including the NICU,^{74,75} in ambulatory settings, and in long-term care facilities. The area of antimicrobial stewardship, however, requires additional research to establish optimal methods in various pediatric specialty populations. One practice from the CDC GET SMART program that can be implemented by each prescriber in most settings is the antibiotic “time out” that consists of reviewing patient data at 48 to 72 hours of treatment to determine which of the following is indicated: (1) continue antibiotic treatment; (2) change to a narrower-spectrum agent; (3) change from a parenteral to an oral agent; or (4) shorten or conclude therapy.⁶⁸

Isolation Precautions

Isolation of patients with potentially transmissible infectious diseases is a strategy proven to prevent transmission of infectious agents in healthcare settings. Many published studies, performed in both adult and pediatric settings, provide a strong evidence base for most recommendations for isolation precautions and for limiting outbreaks. However, controversies exist concerning the most clinically and cost-effective measures for preventing certain HAIs, especially those associated with MDROs. As discussed earlier in the section on surveillance, a call has gone out to reconsider recommendations for isolation of patients who are asymptotically colonized with MRSA or VRE, but no definite recommendation has been made by the HICPAC/CDC, SHEA, or Association for Professionals in Infection Control and Epidemiology.

Since 1970, the guidelines for isolation developed by CDC have responded to the needs of the evolving US healthcare systems. For example, universal precautions became a required standard in response to the HIV epidemic that emerged in the 1980s and the need to prevent acquisition of bloodborne pathogens (e.g., HIV, hepatitis B and C viruses) by HCP through skin, eye, mucous membrane, or parenteral contact with blood or other potentially infectious materials from persons not known to be or suspected of being infected. Universal precautions were modified and have been known as Standard Precautions since publication of the 1996 Guideline for Isolation. The federal Needlestick Safety and Prevention Act, signed into law in November 2000, authorized the Occupational Safety and Health Administration’s revision of its Bloodborne Pathogens Standard more explicitly to require the use of safety-engineered sharp devices.⁷⁶

Evidence and recommendations are provided for the prevention of transmission of MDROs such as MRSA, VRE, VISA, VRSA, and GNB.^{4,5} The components of a protective environment for prevention of environmental fungal infections in HSCT recipients are summarized.⁴ Finally, evidence-based, rated recommendations for administrative measures that are necessary for effective prevention of infection in healthcare settings are provided.

Standard Precautions

The most recent Guideline for Isolation Precautions published in 2007⁴ reaffirms *Standard Precautions*, a combination of universal precautions and body substance isolation, as the foundation of transmission prevention measures. Critical thinking is required for HCP to recognize the importance of body fluids, excretions, and secretions in the transmission of infectious pathogens and take appropriate protective precautions by using PPE (e.g., masks, gowns, gloves, face shields, or goggles) and safety devices when exposure is likely even if an infection is not suspected or known. In addition, these updated guidelines provide recommendations for Standard Precautions in all settings in which healthcare is delivered (acute care hospitals, ambulatory surgical and medical centers, long-term care facilities, and home health agencies). The components of Standard Precautions are summarized in [Table 2.1](#). In the most recent HICPAC/CDC isolation guideline,⁴ safe injection practices are included as a component of Standard Precautions. Despite the emphasis on safe injection practices, transmission of hepatitis B and C viruses continues to be reported in ambulatory care settings as a result of failure to follow recommended practices, thus indicating a need to reiterate the established effective practices.⁷⁷ During 2008 to 2014, 45 outbreaks (≥2 cases) of viral hepatitis related to healthcare were reported to the CDC; of these, 42 (95%) occurred in nonhospital settings. A review of clusters of transmission of hepatitis C virus in dialysis centers from 2005 to 2015

identified the following potential infection control breaches: (1) use of multidose vials for heparin or saline administration; (2) poor compliance with hand hygiene before and after patient contacts or after touching a possibly contaminated surface; (3) failure to change gloves between patient contacts or after contact with a potentially contaminated surface; (4) failure to disinfect environmental surfaces adequately; (5) unsafe injection practices; (6) failure to disinfect shared equipment between patient uses; (7) lack of a separate area for medication preparation; and (8) failure to have clean and dirty utility rooms clearly separated.⁷⁸

Two additions were made to Standard Precautions in 2007: (1) *respiratory hygiene or cough etiquette* for source containment by people with signs and symptoms of respiratory tract infection and (2) *use of a mask* by personnel inserting an epidural anesthesia needle or performing a myelogram when prolonged exposure of the puncture site is likely. Both components have a strong evidence base.

Implementation of Standard Precautions requires the availability of PPE in proximity to all patient care areas. HCP with exudative lesions or weeping dermatitis must avoid direct patient care and handling of patient care equipment. Persons having direct patient contact should be able to anticipate exposure incurring risks and steps to take if a high-risk exposure occurs. Exposures of concern are as follows: exposures to blood or other potentially infectious material defined as an injury with a contaminated sharp object (e.g., needlestick, scalpel cut); a spill or splash of blood or other potentially infectious material onto nonintact skin (e.g., cuts, hangnails, dermatitis, abrasions, chapped skin) or onto a mucous membrane (e.g., mouth, nose, eye); or blood exposure covering a large area of normal skin. Patient-related duties that do *not* constitute high-risk exposures include handling food trays or furniture, pushing wheelchairs or stretchers, using restrooms or telephones, having personal contact with patients (e.g., giving information, touching intact skin, bathing, giving a back rub, shaking hands), or performing clerical or administrative functions for a patient.

If hands or other skin surfaces are exposed to blood or other potentially infectious material, the area should be washed immediately with soap and water for at least 10 seconds and rinsed with running water for at least 10 seconds. For an eye, nose, or mouth splash with blood or body fluids, the area should be irrigated immediately with a large volume of water. If a skin cut, puncture, or lesion is exposed to blood or other potentially infectious material, the area should be washed immediately with soap and water for at least 10 seconds and rinsed with 70% isopropyl alcohol. Any exposure incident should be reported immediately to the occupational health department to determine whether blood samples are required from the source patient and the exposed person and if immediate prophylaxis is indicated.

All HCP should know where to find the exposure control plan specific to each place of employment, whom to contact, where to go, and what to do if inadvertently exposed to blood or body fluids. Important resources include the occupational health department, the emergency department, and the infection control or hospital epidemiology division. The most important recommendation in any accidental exposure is to seek advice and intervention immediately because the efficacy of recommended prophylactic regimens is improved with shorter intervals after exposure, such as for hepatitis B immune globulin administration after exposure to hepatitis B virus or for antiretroviral therapy after percutaneous exposure to HIV. Chemoprophylaxis following exposure to HIV-infected material is most effective if it is initiated as soon as possible, but within hours of exposure.⁷⁹ The current guidelines recommend using ≥3 drugs for post-exposure prophylaxis of HIV independent of the severity of exposure. Updates are posted on the CDC website as they are developed. Reporting a work-related exposure is required for subsequent medical care and workers’ compensation.

Transmission-Based Precautions

Transmission-Based Precautions are designed for patients with documented or suspected infection with pathogens for which additional precautions beyond Standard Precautions are needed to prevent transmission. The 3 categories of Transmission-Based Precautions are *Contact Precautions*, *Droplet Precautions*, and *Airborne Precautions*, and they are based on the likely routes of transmission of specific infectious agents. Transmission-based precautions are combined for infectious agents that have more than 1 route of transmission. When used singly or in

TABLE 2.1 Recommendations for Application of Standard Precautions for the Care of All Patients in All Healthcare Settings

Component	Recommendations for Performance
Hand hygiene	Perform before touching patients and before donning gloves; after touching blood, body fluids, secretions, excretions, contaminated items; immediately after removing gloves; between patient contacts. Alcohol-containing antiseptic hand rubs preferred except when hands are visibly soiled with blood or other proteinaceous materials or if exposure to spores (e.g., <i>Clostridium difficile</i> , <i>Bacillus anthracis</i>) is likely to have occurred
Gloves	Use for touching blood, body fluids, secretions, excretions, contaminated items; for touching mucous membranes and nonintact skin
Gown	Wear during procedures and patient care activities when contact of clothing or exposed skin with blood or body fluids, secretions, and excretions is anticipated
Mask, ^a eye protection (goggles), face shield	Wear during procedures and patient care activities likely to generate splashes or sprays of blood, body fluids, secretions (especially suctioning, endotracheal intubation) to protect healthcare personnel. For patient protection, the person inserting an epidural anesthesia needle or performing myelograms should use a mask when prolonged exposure of the puncture site is likely to occur
Soiled patient-care equipment	Handle in a manner that prevents transfer of microorganisms to others and to the environment; wear gloves if equipment is visibly contaminated; perform hand hygiene
Environmental control	Develop procedures for routine care, cleaning, and disinfection of environmental surfaces, especially frequently touched surfaces in patient care areas
Textiles and laundry	Handle in a manner that prevents transfer of microorganisms to others and to the environment
Safe injection practices (use of needles and other sharps)	Do not recap, bend, break, or hand-manipulate used needles; if recapping is required, use a one-handed scoop technique only; use needle-free safety devices when available; place used sharps in a puncture-resistant container. Use a sterile, single-use, disposable needle and syringe for each injection given. Single-dose medication vials are preferred when medications are administered to >1 patient
Patient resuscitation	Use a mouthpiece, resuscitation bag, or other ventilation device to prevent contact with mouth and oral secretions
Patient placement	Prioritize for a single-patient room if the patient is at increased risk of transmission, is likely to contaminate the environment, does not maintain appropriate hygiene, or is at increased risk of acquiring infection or developing adverse outcome following infection
Respiratory hygiene and cough etiquette ^b	Instruct symptomatic persons to cover the mouth or nose when sneezing or coughing; use tissues and dispose in no-touch receptacle; observe hand hygiene after soiling of hands with respiratory secretions; wear a surgical mask if tolerated or maintain spatial separation, >1–2 m (3–6 feet) if possible

^aDuring aerosol-generating procedures on patients with suspected or proven infections transmitted by aerosols (e.g., severe acute respiratory syndrome), wear a fit-tested N95 or higher respirator in addition to gloves, gown, and face and eye protection.
^bSource containment of infectious respiratory secretions in symptomatic patients, beginning at the initial point of encounter (e.g., triage and reception areas in emergency departments and physician offices).
 Modified with permission from Siegel JD, Rhinehart E, Jackson M, et al. Guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings, 2007. *Am J Infect Control* 2007;35(Suppl 2):S65–S164.

combination, such precautions always are used in addition to Standard Precautions. Transmission-based precautions applied at the time of initial contact, based on the clinical presentation and the most likely pathogens are referred to as *Empiric Precautions* or *Syndromic Precautions*. This approach is useful especially for emerging agents (e.g., SARS-CoV, avian influenza, pandemic influenza), for which information concerning routes of transmission is evolving. The categories of clinical presentation are as follows: diarrhea, central nervous system, generalized rash or exanthem, respiratory, skin or wound infection. Single-patient rooms always are preferred for children needing Transmission-Based Precautions. If single-patient rooms are unavailable, cohorting of patients, and preferably of staff, according to clinical diagnosis is recommended. The experience of treating EVD in the US in 2014 led to the development of special precautions after viral transmission to 2 nurses occurred as a result of patients' extraordinarily high viral loads and large volumes of emitted body fluids.⁶ PPE for all transmission-based precautions is donned *upon entry* into the room to protect against acquisition of pathogens from contaminated surfaces, even if direct contact with the patient is not intended.

Although targeted Contact Precautions and universal gowning and gloving are effective for preventing transmission of infectious agents, potential adverse effects in patients placed on Contact Precautions have been described (e.g., depression, fewer visits from the healthcare team, increased rates of hypoglycemia or hyperglycemia, increased falls).⁸⁰ Additionally, adherence to Contact Precautions decreases as the number of patients on Contact Precautions increases.⁸¹ Finally, a simulation study demonstrated contamination of HCP skin and clothing during doffing of gowns and gloves⁸²; this study effectively demonstrated the PPE lessons learned during the SARS and EVD experiences. Evidence

supports the importance of applying Contact Precautions only when indicated, obtaining training on the use of PPE, having effective PPE readily available, and practicing consistent and precise use of PPE.⁸³

Table 2.2 lists the 3 categories of isolation based on routes of transmission and their necessary components. Table 2.3 lists precautions by syndromes, to be used when a patient has an infectious disease and the agent is not yet identified. For infectious agents that are more likely to be transmitted by the droplet route (e.g., pandemic influenza), droplet precautions (with use of surgical mask) are appropriate; however, during an aerosol-generating procedure, N95 or higher respirators are indicated.⁸⁴

Environmental Measures

Contaminated environmental surfaces and noncritical medical items have been implicated in transmission of several infectious agents, including VRE, *C. difficile*, *Acinetobacter* spp., MRSA, and RSV in healthcare settings.^{4,85,86} Pathogens on surfaces are transferred to the hands of HCP and are then transferred to patients or items to be shared. Occupying a room previously occupied by a patient with a key pathogen is a risk factor for acquiring that pathogen during a hospital stay. Most often, the failure to follow recommended procedures for cleaning and disinfection contributes more than does the specific pathogen to the environmental reservoir during outbreaks. Education of environmental services personnel combined with direct observation and feedback was associated with a persistent decrease in VRE acquisition in a medical ICU. Use of a standardized cleaning checklist and implementation of monitoring for adherence to recommended environmental cleaning practices are important determinants of success. Visual markers (e.g., invisible fluorescein powder) and adenosine triphosphate bioluminescence technologies are

TABLE 2.2 Transmission-Based Precautions^a

Component	Contact	Droplet	Airborne
Hand hygiene	Per Standard Precautions Perform 5 moments of hand hygiene, and upon entry into room Soap and water preferred over alcohol hand rub for <i>Clostridium difficile</i> , <i>Bacillus anthracis</i> spores	Per Standard Precautions Perform 5 moments of hand hygiene, and upon entry into room	Per Standard Precautions Perform 5 moments of hand hygiene, and upon entry into room
Gown	Yes; don before or upon entry into room	Per Standard Precautions Add to droplet precautions for infants, young children, or presence of diarrhea	Per Standard Precautions and, if infectious, draining skin lesions present
Gloves	Yes; don before or upon entry into room	Per Standard Precautions. Add for infants, young children and/or presence of diarrhea	Per Standard Precautions Add for infants, young children or presence of diarrhea
Mask	Per Standard Precautions	Yes; don before or upon entry into room	Don N95 particulate respirator or higher before entry into room
Goggles or face shield	Per Standard Precautions	Per Standard Precautions Always for SARS, avian influenza	Per Standard Precautions Always for SARS, avian influenza
N95 or higher respirator (Always don before entry into room)	When aerosol-generating procedures performed for influenza, SARS, VHF ^b	When aerosol-generating procedures performed for influenza, SARS, VHF	Yes; don before entry into room
Room placement	Single-patient room preferred Cohort similar infections if single-patient rooms unavailable	Single-patient room preferred Cohort similar infections if single-patient rooms unavailable	Single-patient room Negative air pressure; 12 air changes/hr for new construction, 6 air changes/hr for existing rooms
Environmental measures	Increased frequency, especially in the presence of diarrhea, transmission of <i>C. difficile</i> , norovirus Bleach for VRE, <i>C. difficile</i> , norovirus	Routine	Routine
Transport	Mask patient if coughing Cover infectious skin lesions PPE not routinely required for transporter	Mask patient	Mask patient Cover infectious skin lesions

^aIn addition to Standard precautions, use Transmission-Based Precautions for patients with highly transmissible or epidemiologically important pathogens for which additional precautions are needed.

^bIncludes Ebola virus. Consult most recent Centers for Disease Control and Prevention and World Health Organization guidelines for recommended infection control precautions for Ebola virus disease.

PPE, personal protective equipment; SARS, severe acute respiratory syndrome; VHF, viral hemorrhagic fever; VRE, vancomycin-resistant *Enterococcus*.

Modified with permission from Siegel JD, Rhinehart E, Jackson M, et al. Guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings, 2007. Am J Infect Control 2007;35(Suppl 2):S65–S164.

also useful for monitoring effective environmental cleaning and providing immediate feedback to workers.⁸⁷ A program of environmental cleaning should be developed collaboratively by the IPC and environmental services departments. Certain infectious agents (e.g., rotavirus, noroviruses, *C. difficile*) can be resistant to some routinely used hospital disinfectants; thus when ongoing transmission occurs despite appropriate cleaning procedures, a 1:10 dilution of 5.25% sodium hypochlorite (household bleach) or other special disinfectants are indicated.

“No-touch” automated room decontamination technologies have been developed and added to room turnover procedures in some facilities. Ultraviolet light irradiation and hydrogen peroxide vapor systems have been shown to reduce surface contamination with common pathogens and decrease the risk of acquiring HAIs caused by those pathogens when these systems are added to a terminal cleaning regimen.^{85–87} At specific wavelengths, ultraviolet light breaks the molecular bonds in DNA, thus destroying the organisms. Ultraviolet technology also has been considered as a method of disinfecting PPE, as a risk mitigation strategy for HCP caring for patients with EVD.⁸⁸ These technologies supplement, but do not replace, standard cleaning and disinfection because surfaces must be physically cleaned of particulate matter and debris. Other disadvantages of these systems are that they cannot be used when people are in the rooms, room turnover is delayed, and the systems are expensive to purchase. No recommendations have been made for routine use or specific indications because research on antimicrobial effectiveness, cost effectiveness, and feasibility of these systems is ongoing.

Self-disinfecting surfaces can be created by altering the structure of the surface material or by incorporating a material that has antimicrobial activity.^{85–87} Copper has antimicrobial activity against a wide range of organisms including bacteria and fungi. Thus, incorporating copper into high-touch surfaces such as toilet seats, bed rails, door handles, or countertops is a novel infection prevention strategy that has been shown to reduce bacterial colony counts compared with control surfaces in healthcare settings.⁸⁹ However, no recommendation for routine use has yet been made.

Disinfection, Sterilization, and Removal of Infectious Waste

Disinfection and sterilization as they relate to IPC have been reviewed,⁹⁰ and the HICPAC/CDC developed comprehensive guidelines in 2008.⁹¹ *Cleaning* is the removal of all foreign material from surfaces and objects. This process is accomplished using soap and enzymatic products. Failure to remove all organic material from items before disinfection and sterilization reduces the effectiveness of these processes. *Disinfection* is a process that eliminates all forms of microbial life except the endospore. Disinfection usually requires liquid chemicals. Disinfection of an inanimate surface or object is affected adversely by the following: the presence of organic matter; a high level of microbial contamination; use of too dilute germicide; inadequate disinfection time; an object that

TABLE 2.3 Clinical Syndromes or Conditions Warranting Empiric Transmission-Based Precautions in Addition to Standard Precautions Pending Confirmation of Diagnosis^a

Clinical Syndrome or Condition ^b	Potential Pathogens ^c	Empiric Precautions (Always Includes Standard Precautions)
DIARRHEA		
Acute diarrhea with a likely infectious cause in an incontinent or diapered patient	Enteric pathogens ^d	Contact Precautions (pediatrics and adult)
MENINGITIS		
	<i>Neisseria meningitidis</i>	Droplet Precautions for first 24 hr of antimicrobial therapy; mask, face, and eye protection for intubation
	Enteroviruses	Contact Precautions for infants and children
	<i>Mycobacterium tuberculosis</i>	Airborne Precautions if pulmonary infiltrate Airborne Precautions plus Contact Precautions if potentially infectious draining body fluid present
RASH OR EXANTHEMS, GENERALIZED, ORIGIN UNKNOWN		
Petechial or ecchymotic exanthem with fever (general)	<i>N. meningitidis</i>	Droplet Precautions for first 24 hr of antimicrobial therapy
If traveled in an area with an ongoing outbreak of viral hemorrhagic fever in the 10 days before onset of fever	Ebola, Lassa, Marburg viruses	Airborne Precautions plus Contact Precautions, with face and eye protection, emphasizing safety sharps and barrier precautions when blood exposure likely In the United States, asymptomatic persons can be managed in Ebola assessment centers. Transfer symptomatic persons with infection to biocontainment units Use a single-use fluid-resistant or impermeable gown that extends to at least midcalf or a coverall without an integrated hood. Two pairs of gloves should be worn. Use a single-use fluid-resistant or impermeable boot cover. A single-use fluid resistant or impermeable apron should be worn to cover the torso if the patient has vomiting or diarrhea Consult CDC, WHO websites for current recommendations
Vesicular	Varicella-zoster, herpes simplex, variola (smallpox), vaccinia viruses	Airborne plus Contact Precautions Contact Precautions only if herpes simplex, localized zoster in an immunocompetent host, or vaccinia viruses most likely
Maculopapular with cough, coryza, and fever	Rubeola (measles) virus	Airborne Precautions
RESPIRATORY INFECTIONS		
Cough, fever, or upper-lobe pulmonary infiltrate in an HIV-negative patient or a patient at low risk for HIV infection	<i>M. tuberculosis</i> , respiratory viruses, <i>Streptococcus pneumoniae</i> , <i>Staphylococcus aureus</i> (MSSA or MRSA)	Airborne Precautions plus Contact Precautions until <i>M. tuberculosis</i> ruled out Droplet Precautions if respiratory viruses most likely
Cough, fever, or pulmonary infiltrate in any lung location in an HIV-infected patient or a patient at high risk for HIV infection	<i>M. tuberculosis</i> , respiratory viruses, <i>Streptococcus pneumoniae</i> , <i>Staphylococcus aureus</i> (MSSA or MRSA)	Airborne Precautions plus Contact Precautions Use eye and face protection if aerosol-generating procedure performed or contact with respiratory secretions anticipated If tuberculosis is unlikely and no AIIRs or respirators are available, use Droplet Precautions instead of Airborne Precautions Tuberculosis more likely in HIV-infected than in HIV-uninfected persons
Cough, fever, or pulmonary infiltrate in any lung location in a patient with a history of recent travel (10–21 days) to a country with an outbreak of SARS or avian influenza	<i>M. tuberculosis</i> , severe acute respiratory syndrome virus–coronavirus (SARS-CoV), avian influenza	Airborne Precautions plus Contact Precautions in addition to eye protection If SARS and tuberculosis unlikely, use Droplet Precautions instead of Airborne Precautions
Respiratory infections, particularly bronchiolitis and pneumonia, in infants and young children	Respiratory syncytial virus, parainfluenza virus, adenovirus, influenza virus, human metapneumovirus	Contact Precautions plus Droplet Precautions Droplet Precautions can be discontinued when adenovirus and influenza have been ruled out
SKIN OR WOUND INFECTION		
Abscess or draining wound that cannot be covered	<i>Staphylococcus aureus</i> (MSSA or MRSA), group A streptococcus	Contact Precautions Add droplet precautions for the first 24 hr of appropriate antimicrobial therapy if invasive group A streptococcal disease is suspected

^aInfection control professionals should modify or adapt this table according to local conditions. To ensure that appropriate empiric precautions are always implemented, hospitals must have systems in place to evaluate patients routinely according to these criteria as part of preadmission and admission care.

^bPatients with the syndromes or conditions listed may have atypical signs or symptoms (e.g., neonates and adults with pertussis may not have paroxysmal or severe cough). The clinician's index of suspicion should be guided by the prevalence of specific conditions in the community, as well as clinical judgment.

^cThe organisms listed under the column "Potential Pathogens" are not intended to represent the complete, or even most likely, diagnoses, but rather possible etiologic agents that require additional precautions beyond standard precautions until they can be excluded.

^dThese pathogens include enterohemorrhagic *Escherichia coli* O157:H7, *Shigella* spp., hepatitis A virus, noroviruses, rotavirus, and *Clostridium difficile*.

AIIR, airborne infection isolation room; CDC, Centers for Disease Control and Prevention; CoV, coronavirus; HIV, human immunodeficiency virus; MRSA, methicillin-resistant *Staphylococcus aureus*; MSSA, methicillin-susceptible *Staphylococcus aureus*; SARS, severe acute respiratory syndrome; VHF, viral hemorrhagic fever; WHO, World Health Organization.

can harbor microbes in protected cracks, crevices, and hinges; and pH and temperature.

Sterilization is the eradication of all forms of microbial life, including fungal and bacterial spores. Sterilization is achieved by physical and chemical processes such as steam under pressure, dry heat, ethylene oxide, and liquid chemicals. The Spaulding classification of patient care equipment as *critical*, *semicritical*, and *noncritical* items with regard to sterilization and disinfection is used by the CDC. *Critical items* require sterilization because they enter sterile body tissues and carry a high risk of causing infection if they are contaminated; *semicritical items* require disinfection because they may contact mucous membranes and nonintact skin; and *noncritical items* require routine cleaning because they come in contact only with intact skin. If noncritical items used on patients requiring Transmission-Based Precautions, especially Contact Precautions, must be shared, these items should be disinfected between uses. Guidelines for specific objects and specific disinfectants are published and updated by the CDC. Multiple published reports and manufacturers similarly recommend the use and reuse of objects with appropriate sterilization, disinfection, or cleaning recommendations. Recommendations in guidelines for reprocessing endoscopes to avoid contamination focus on training of personnel, meticulous manual cleaning, high-level disinfection followed by rinsing and air-drying, and proper storage.⁹² However, outbreaks of MDR GNB infections associated with exposure to duodenoscopes used for retrograde cholangiopancreatography that have been reprocessed according to recommendations suggest a need for new endoscope reprocessing technologies.^{51,93} These endoscopes have a complex design with long, narrow channels, crevices that are difficult to access with a cleaning brush, right-angle turns, and heavy microbial contamination following procedures. Until new methods are developed, meticulous adherence to recommended processes with enhancements should be followed. Medical devices that are designed for single use (e.g., specialized catheters, electrodes, biopsy needles) must be reprocessed by third parties or hospitals according to the guidance issued by the Food and Drug Administration (FDA) in August, 2000 with amendments in September, 2006; such reprocessors are considered and regulated as “manufacturers.” Available data show that single-use devices reprocessed according to the FDA regulatory requirements are as safe and effective as new devices.

Deficiencies in disinfection and sterilization leading to infection have resulted either from failure to adhere to scientifically based guidelines or failures in the disinfection or sterilization processes. When such failures are discovered, an investigation must be completed, including notification of patients and, in some cases, testing for infectious agents. A guidance document for risk assessment and communication to patients in such situations is published.⁹⁴

Healthcare facility waste is all biologic or nonbiologic waste that is discarded and not intended for further use. *Medical waste* is material generated as a result of use with a patient, such as for diagnosis, immunization, or treatment, and it includes soiled dressings and intravenous tubing. *Infectious waste* is that portion of medical waste that potentially could transmit an infectious disease. Microbiologic waste, pathologic waste, contaminated animal carcasses, blood, and sharps are all examples of infectious waste. Methods of effective disposal of infectious waste include incineration, steam sterilization, drainage to a sanitary sewer, mechanical disinfection, chemical disinfection, and microwave treatment. State regulations guide the treatment and disposal of regulated medical waste. Recommendations are available for developing and maintaining a program within a facility for safe management of medical waste.⁹⁵

Visitation Policies

Special visitation policies are required in pediatric units, especially the high-risk units, because acquisition of a seemingly innocuous viral infection in neonates and in children with underlying diseases can result in unnecessary evaluations and empiric therapies for suspected septicemia as well as serious, life-threatening disease. All visitors with signs or symptoms of respiratory or gastrointestinal tract infection should be restricted from visiting patients in healthcare facilities. Increased restrictions may be required during a community outbreak (e.g., SARS, pandemic influenza, enterovirus D68). During respiratory virus season, the number of visitors can be limited and the age restriction increased. It is preferred for all visitors to be immunized against influenza during

influenza season. Several children’s hospitals provide influenza vaccine or tetanus, diphtheria, and acellular pertussis (Tdap) vaccine, or both, to household contacts at no charge, thereby supporting the cocooning strategy endorsed by the Advisory Committee on Immunization Practices and the American Academy of Pediatrics.⁹⁶

For patients requiring Contact Precautions, the use of PPE by visitors is determined by the nature of the interaction with the patient and the likelihood that the visitor will frequent common areas on the patient’s unit or interact with other patients and their families. It is important to distinguish parents or guardians from nonhousehold visitors when determining whether the visitor should wear PPE. The risk-benefit decision should weigh not only the specific pathogen in question, but also the effect of parental or guardian PPE on breastfeeding, bonding, and family participation in the child’s medical care. For family members who are rooming in with children who have prolonged hospitalizations, restriction of visitation to other patients is emphasized. A SHEA expert guidance document has been published to summarize the principles to follow to prevent transmission of infectious agents by visitors to patients because few data are available to inform evidence-based recommendations.⁹⁷

Although most pediatricians encourage visits by siblings in inpatient areas, the medical risk must not outweigh the psychosocial benefit. Families favorably regard sibling visitation, and no evidence indicates increased bacterial colonization or subsequent bacterial infection in the neonate or older child who has been visited by siblings. Strict guidelines for sibling visitation should be established and enforced in an effort to maximize visitation opportunities and minimize risks of transmission of infectious agents, most frequently viruses. The following recommendations regarding visitation can guide policy development:

1. Sibling visitation is encouraged in the well-child nursery and NICU, as well as in areas for care of older children.
2. Before visitation, parents should be interviewed by a trained staff nurse concerning the current health status of the sibling. Siblings should not be allowed to visit if they are delinquent in recommended vaccines, have fever or symptoms of an acute illness, or are within the incubation period following exposure to a known infectious disease. After the interview, the physician or nurse should place a written consent for sibling visitation in the patient’s permanent record and a name tag indicating that the sibling has been approved for visitation for that day.
3. Asymptomatic siblings who recently were exposed to varicella but who previously were immunized can be assumed to be immune.
4. The visiting sibling should visit only his or her sibling and not be allowed in playrooms with groups of patients.
5. Visitation should be limited to periods of time that ensure adequate screening, observation, and monitoring of visitors by medical and nursing staff members.
6. Children should perform hand hygiene before and after contact with the patient or upon entry and departure from the patient’s room.
7. During the entire visit, sibling activity should be supervised by parents or another responsible adult.

Animals in Healthcare Facilities

Animal-assisted therapy can be of substantial clinical benefit to the child hospitalized for prolonged periods; therefore it is important for healthcare facilities to provide guidance for safe visitation. Many zoonoses and infections are attributable to animal exposure (see Chapter 89). Most infections result from inoculation of animal flora through a bite or scratch or self-inoculation after contact with the animal, the animal’s secretions or excretions, or contaminated environment. Although few data support a true evidence-based guideline for animal visitation (including personal pets) in healthcare facilities, updated expert guidance is provided in the SHEA Expert Guidance on Animals in Healthcare Facilities: Recommendations to Minimize Potential Risk, which includes a review of the literature related to animal-assisted activities.⁹⁸

Prudent visitation policies should limit visitation to animals that: (1) are domesticated; (2) do not require a water environment; (3) do not bite or scratch; (4) can be brought to the hospital in a carrier or easily walked on a leash; (5) are trained to defecate and urinate outside or in appropriate litter boxes; (6) can be bathed before visitation; and (7) are known to be free of respiratory, dermatologic, and gastrointestinal tract disease. Despite the established risk of salmonellosis

associated with reptiles (e.g., turtles, iguanas), many reports of outbreaks of invasive disease associated with reptiles continue to be published⁹⁹; reptiles should be excluded from pet visitation programs, and families should be advised not to have pet reptiles in the home with young infants or immunocompromised persons. Exotic animals that are imported should be excluded because of unpredictable behavior and the potential for transmission of unusual pathogens (e.g., monkeypox in the US in 2003).^{100,101} Visitation should be limited to short periods and confined to designated areas. Visiting pets must have a certificate of immunization from a licensed veterinarian. Children should observe hand hygiene after contact with animals. Most pediatric facilities restrict pet interaction with severely immunosuppressed patients and patients in ICUs.

Occupational Health

Occupational health and student health collaboration with the IPC department of a healthcare facility is required by the Occupational Safety and Health Administration. HCP are at increased risk of infection in hospitals caring for children because (1) children have a high incidence of infectious diseases, (2) personnel can be susceptible to many pediatric pathogens, (3) pediatric care requires close contact, (4) children lack good personal hygiene, (5) infected children can be asymptomatic, and (6) HCP are exposed to multiple family members who also may be infected.

The occupational health department is an educational resource for information on infectious pathogens in the healthcare workplace. In concert with the IPC service, occupational health provides preemployment education and respirator fit testing and annual retraining for all employees regarding routine health maintenance, available recommended and required vaccines, Standard and Transmission-Based Precautions, and exposure control plans. Screening for tuberculosis at regular intervals, as determined by the facility's risk assessment, can use either tuberculin skin testing or interferon- γ release assays.¹⁰² With new pathogens being isolated, new diseases and their transmission described, and new prophylactic regimens and treatment available, it is mandatory that personnel have an up-to-date working knowledge of IPC and know where and what services, equipment, and therapies are available for HCP.

All HCP should be screened by history or serologic testing, or both, to document their immune status to specific agents, and immunization should be provided for the following for all employees who are nonimmune and who do not have contraindications to receiving the vaccine: diphtheria toxoid, hepatitis B virus, influenza (yearly), mumps, poliomyelitis, rubella, rubeola, varicella, and Tdap. The 2006 Advisory Committee on Immunization Practices recommendation to administer a single dose of Tdap to certain HCP was amended in 2011 to have no restriction based on age or time interval since the last Td dose. Providing vaccines at no cost to HCP increases acceptance.

Influenza vaccine coverage among HCP has increased over time to 77% overall for the 2014 to 2015 influenza season, with the highest coverage rate of 90% in HCP working in hospitals and the lowest rate of 64% in long-term care settings.¹⁰³ Although mandatory influenza vaccination programs for all employees in healthcare facilities are endorsed by many professional societies,^{104,105} some facilities have had success using novel strategies that include incentives, without a mandate.¹⁰⁶ Publications from several large institutions, including children's hospitals, indicate that mandatory programs with only medical and religious exemptions are well received, and only rare employees are terminated for failure to be vaccinated.^{107,108}

Special Concerns of Healthcare Personnel

HCP who have common underlying medical conditions should be able to obtain general information on wellness and screening when needed from the occupational health service. HCP with direct patient contact who have infants <1 year of age at home often are concerned about acquiring infectious agents from patients and transmitting them to their susceptible children. An immune healthcare worker who is exposed to VZV does not become a silent "carrier" of VZV. However, pathogens to which the healthcare worker is partially immune or nonimmune can cause a severe, mild, or asymptomatic infection in the employee that can be transmitted to family members. Examples include influenza, pertussis, RSV and other

respiratory viruses, norovirus, and tuberculosis. Important preventive procedures for HCP with infants at home or who are pregnant are as follows: (1) consistent training and observance of Standard Precautions, Transmission-Based Precautions, and especially hand hygiene according to published recommendations; (2) annual influenza and 1-time Tdap immunization (unless pregnant, when a Tdap immunization during each pregnancy is recommended); (3) routine tuberculosis screening; (4) assurance of immunity or immunization against poliomyelitis, measles, mumps, hepatitis B, and rubella; (5) early medical evaluation for acute infectious illnesses; (6) routine, on-time immunization of infants; and (7) prompt initiation of prescribed prophylaxis or therapy following exposure or development of certain infections.

HCP who are, could be, or anticipate becoming pregnant should feel comfortable working in the healthcare workplace. In fact, with Standard Precautions and appropriate adherence to environmental cleaning and isolation precautions, vigilant HCP can be at less risk than a preschool teacher, childcare provider, or mother of children with many playmates in the home. Pathogens of potential concern to pregnant HCP include cytomegalovirus, hepatitis B virus, influenza, measles, mumps, parvovirus B19, rubella, VZV, *M. tuberculosis*, and, since 2015, Zika virus. The causal association between Zika virus and microcephaly and other neurodevelopmental abnormalities¹⁰⁹ has led to recommended precautions. Although Zika virus is more frequently acquired outside of healthcare, pregnant HCP are advised to follow safe injection practices for prevention of exposure to infectious blood.⁴ Pregnancy is an *indication* for influenza vaccine to prevent the increased risk of serious disease and hospitalization that occurs in women who develop influenza in the second or third trimester of pregnancy. In 2011, the CDC recommended universal immunization with Tdap (if previously not immunized with Tdap) for pregnant women after 20 weeks of gestation, and since 2012, the CDC recommends a dose of Tdap with each pregnancy.¹¹⁰ Pregnant workers should assume that all patients potentially are infected with cytomegalovirus and other "silent" pathogens and should use hand hygiene and gloves when handling body fluids, secretions, and excretions. [Table 2.4](#) summarizes information about infectious agents that are relevant to the pregnant woman working in healthcare. Chapters on each agent may be consulted for more specific information.

Infection Prevention and Control in the Nonacute Care Setting

The risk of HAIs in pediatric ambulatory settings is substantial, and it usually is associated with lack of adherence to routine IPC practices and procedures, especially disinfection, sterilization, and hand hygiene. Respiratory viral agents and *M. tuberculosis* are noteworthy pathogens transmitted in ambulatory settings. Transmission of RSV in an HSCT outpatient clinic has been demonstrated using molecular techniques.¹¹¹ Crowded waiting rooms, toys, furniture, lack of isolation of children, unwell children, contaminated hands, contaminated secretions, and susceptible HCP are only some of the factors that result in sporadic and epidemic illness in outpatient settings. The association of community-associated MRSA in HCP working in an outpatient HIV clinic with environmental community-associated MRSA contamination of that clinic indicates the potential for transmission in this setting.¹¹² Patient-to-patient transmission of *Burkholderia* species and *P. aeruginosa* in outpatient clinics for patients with cystic fibrosis has been confirmed and prevented by implementing recommended IPC methods.⁹ IPC guidelines and policies for pediatric outpatient settings, including office practices, were published by the American Academy of Pediatrics in 2007,¹¹³ reaffirmed in 2015, and are updated currently. Prevention strategies include definition of policies, education, and strict adherence to guidelines. In pediatrics, among the most important interventions are separation of children with respiratory tract illnesses from well children and consistent implementation of respiratory etiquette or cough hygiene. A guideline for IPC for outpatient settings with a checklist and a guideline for outpatient oncology settings can be found on the CDC website.¹¹⁴ Principles and recommendations for Safe Living after HSCT¹¹⁵ and for patients with cystic fibrosis⁹ are valuable contributions to management of infectious risks for specific populations in the ambulatory setting. A guideline based on data and expert consensus opinion for IPC in residential facilities for

Text continued on p. 25

TABLE 2.4 Pregnant Healthcare Personnel: Guide to Management of Occupational Exposure to Selected Infectious Agents

Agent or Disease	In-Hospital Source	Potential Effect on the Fetus	Perinatal Transmission	Maternal Screening	Prevention
<i>Bordetella pertussis</i> Pertussis (“whooping cough”)	Respiratory droplets from a coughing patient, HCP, visitor	No congenital syndrome	Maternal respiratory secretions	Documentation of date of vaccination with Tdap recommended during each pregnancy. Past history of pertussis disease is <i>not</i> protective and does not replace vaccine	Tdap in third trimester of each pregnancy (to prevent young infant from acquiring pertussis) and for every adult coming in contact with infant <12 months of age Breastfeeding <i>not</i> contraindicated <i>Standard Precautions</i> plus <i>Droplet Precautions</i>
Cytomegalovirus (CMV)/ congenital infection syndrome^a, hearing loss	Patient populations: neonates, toddlers in childcare, hemodialysis patients, immunocompromised hosts, transplant recipients HCP acquisition during pregnancy unlikely to be occupational Urine, saliva, blood, semen, vaginal secretions, breast milk, respiratory secretions if pneumonia present	Symptomatic congenital infection syndrome ^a 5%–10%; hearing loss 10%–15%; asymptomatic congenital infection; hepatitis, anemia, thrombocytopenia Hearing loss can have later onset	Primary infection 25%–50% Recurrent infection or second infection with new strain: 69% reduction in risk of transmission Symptomatic <5%–15%	Routine screening not recommended Preexisting maternal antibody incompletely protective for fetus	Efficacy of CMV immune globulin or ganciclovir for pregnant woman with primary infection not established No vaccine available Breastfeeding <i>not</i> contraindicated Pregnant HCP does not need to be restricted from care of known CMV-infected patient <i>Standard Precautions, especially Safe Injection Practices</i>
Ebola virus	Blood and body fluids from infected, clinically ill patients, especially large-volume diarrhea	Spontaneous abortion, stillbirth Neonatal survival rare.	Data on perinatal transmission limited and anecdotal; likely very high because Ebola virus is present in products of conception when tested	Routine screening not recommended, but a low threshold for diagnosing EVD in the pregnant woman is recommended PCR on blood is the recommended diagnostic test Case fatality rate of EVD in pregnant women is 90%	Restriction of pregnant women from care of persons with EVD Prolonged shedding of Ebola virus in breast milk Breastfeeding contraindicated, but duration unknown When caring for people not suspected of EVD, but during an EVD epidemic: <i>Standard Precautions, especially Safe Injection Practices plus Enhanced Precautions</i> in a biocontainment unit as defined on CDC and WHO websites
Hepatitis A virus (HAV)	Feces (most common), highest titer just before onset of jaundice; blood very rare Transmission in healthcare settings rare	None; transmission can occur at the time of delivery if mother still in the infectious phase and can cause hepatitis in infant	None	Routine screening not recommended	Vaccine is a killed virus vaccine and can be used safely in pregnancy Breastfeeding <i>not</i> contraindicated <i>Standard Precautions</i> Add <i>Contact Precautions</i> in acute phase
Hepatitis B virus (HBV)	Blood, body fluids, vaginal secretions, semen	Hepatitis; if acquired perinatally or at young age, increased risk for hepatocellular carcinoma	HBeAg and HBsAg ⁺ (10%) HBeAg ⁻ and HbsAg ⁺ (90%)	Routine HBsAg testing advised Documentation of vaccination	HBV vaccine during pregnancy if immunity not previously documented Neonate: HBIG in addition to routine vaccine at birth if mother HBsAg ⁺ or status unknown Breast feeding <i>not</i> contraindicated <i>Standard Precautions, especially Safe Injection Practices</i>

Continued

TABLE 2.4 Pregnant Healthcare Personnel: Guide to Management of Occupational Exposure to Selected Infectious Agents—cont'd

Agent or Disease	In-Hospital Source	Potential Effect on the Fetus	Perinatal Transmission	Maternal Screening	Prevention
Hepatitis C virus (HCV)	Blood, body fluids	Hepatitis	Transmission 5% (range 0%–25%)	Screening only if risk factors present: illicit IV drug user (mother or partner who is illicit IV drug user); history of organ transplant, transfusion of blood or blood products before 1992; hemodialysis; HBV or HIV infection; unexplained elevation of serum hepatic enzymes; history of tattooing; HCP with history of percutaneous exposure to blood test for HCV antibody (if positive, HCV RNA)	No vaccine or immune globulin available; postexposure treatment with antiviral agents investigational; Consult current guidelines for updates Breast feeding <i>not</i> contraindicated <i>Standard Precautions, especially Safe Injection Practices</i>
Herpes simplex virus (HSV) Fever blisters, cold sores, genital ulcers, encephalitis	Vesicular fluid, oropharyngeal and vaginal secretions, amniotic fluid	Sepsis, encephalitis, meningitis, mucocutaneous lesions, congenital malformation (rare)	Primary genital infection 33%–50% Recurrent genital infection 1%–2%	Routine antibody testing minimally useful Maternal type-specific serology for HSV-1 and HSV-2 antibodies should be considered when evaluating an asymptomatic neonate following vaginal or cesarean delivery to a woman with genital lesions that are characteristic of HSV	Since genital infection with HSV is the risk for the fetus, occupational acquisition unlikely to occur Oral suppressive antiviral therapy at or beyond 36 wk of gestation decreases shedding in women with a history of genital HSV lesions Breastfeeding contraindicated only if hepetic lesions are located on the breast <i>Standard Precautions Add Contact Precautions</i> for patients with skin lesions
Human immunodeficiency virus (HIV) Acquired immunodeficiency syndrome (AIDS)	Blood, body fluids, vaginal secretions, semen	No congenital syndrome Transmission primarily during delivery If infected, onset of symptoms usually at 12–18 months of age	Risk of transmission determined by maternal HIV viral load, duration of exposure to maternal blood, body fluids (including breast milk), and use of ART during pregnancy, labor, and postnatally in the infant If no ART: maternal viral load <10,000 copies/mL virus, rate of 2%; if load ≥10,000, rate up to 25% Rate reduced to <3% with perinatal and neonatal ART	Routine prenatal screening advised with repeat at end of pregnancy if high-risk behaviors HIV-exposed infants who receive ART from birth: repeated screens during first year HIV antibody screen with fourth-generation test; if positive, quantitative PCR	Antiretroviral chemoprophylaxis for occupational and nonoccupational exposures as recommended in most recent guidelines Treatment of infected woman with ART during pregnancy according to guidelines ART during labor and for infant of infected mother Repeated screening of treated HIV-exposed infant during first 4–6 months of life; check most recent guidelines Cesarean delivery reduces risk of HIV infection in infant if maternal viral load >1000 copies/mL or unknown near time of delivery Breast feeding <i>not</i> recommended if alternative source of nutrition available <i>Standard Precautions, especially Safe Injection Practices</i>

<p>Influenza virus Influenza (flu)</p>	<p>Respiratory droplets or sneezing; coughing patient, HCP, visitor</p>	<p>No congenital syndrome Influenza in mother can cause fetal hypoxia, premature labor, and fetal death Increased morbidity during third trimester of pregnancy Increased morbidity and mortality in pregnant women with 2009 influenza A (H1N1)</p>	<p>Maternal respiratory secretions</p>	<p>Documentation of vaccine received during current season</p> <p>Inactivated influenza vaccine (IIV) for all pregnant women during each influenza season to decrease risk of hospitalizations for cardiopulmonary complications in mother No risk if exposed to persons who received live attenuated influenza vaccine (LAIV) Breast feeding <i>not</i> contraindicated <i>Standard Precautions plus Droplet Precautions</i> Add <i>Contact Precautions</i> for infants and others who cannot contain their secretions Consider fitted N95 filtering facepiece respirator (or equivalent N95 respirator) for aerosol-generating procedures (e.g., bronchoscopy, nebulizer treatment), but AIIR not indicated</p>
<p>Measles (rubeola)</p>	<p>Respiratory secretions, coughing patient</p>	<p>Prematurity, spontaneous abortion; no congenital syndrome</p>	<p>Rare</p>	<p>Serology (IgG) if question after an exposure, but not routinely checked if adequate documentation provided Provider-documented disease or 2 doses measles-containing vaccine ≥12 months of age</p> <p>Vaccine^b Immune globulin IM or IGIV within 6 days of exposure if nonimmune <i>Standard Precautions plus Airborne Precautions</i></p>
<p>Mycobacterium tuberculosis, active disease (tuberculosis [TB])</p>	<p>Sputum, skin lesions, CSF if meningitis present</p>	<p>Neonatal tuberculosis; liver most frequently infected</p>	<p>Rare</p>	<p>Varies with PPD reaction size and chest radiograph result Anti-TB agents for active TB during pregnancy recommended PPD reliable and safe during pregnancy <i>Standard Precautions plus Airborne Precautions</i> for pulmonary or laryngeal TB. Add <i>Contact Precautions</i> if draining skin lesions</p> <p>PPD, IGRA (interferon-γ release assay, e.g., Quantiferon-Gold, T-Spot) Chest radiograph if indicated clinically or a past known positive PPD or IGRA result</p>
<p>Neisseria meningitidis, meningococcal disease, meningococcal meningitis, sepsis</p>	<p>Blood, respiratory secretions</p>	<p>No congenital syndrome Fetus at risk if mother develops severe disease during pregnancy</p>	<p>Rare</p>	<p>No routine screening</p> <p>Prompt chemoprophylaxis if close (within 3–6 feet) contact with respiratory secretions of patient with meningococcal disease with IM ceftriaxone or oral azithromycin Routine vaccine only if microbiologist and routinely exposed to isolates of <i>N. meningitidis</i> <i>Standard Precautions plus Droplet Precautions</i> until 24 hr after effective therapy initiated</p>

Continued

TABLE 2.4 Pregnant Healthcare Personnel: Guide to Management of Occupational Exposure to Selected Infectious Agents—cont'd

Agent or Disease	In-Hospital Source	Potential Effect on the Fetus	Perinatal Transmission	Maternal Screening	Prevention
Parvovirus B19 Fifth disease (slapped cheeks), fetal anemia, fetal hydrops	Respiratory secretions; blood of immunocompromised patients and patients with sickle cell disease during aplastic crisis Patient with Fifth disease no longer contagious once rash appears	Fetal hydrops, stillbirth No congenital syndrome Approximately 25% Fetal death <10%	Rare	No routine screening. Parvovirus B19 DNA can be detected in serum, leukocytes, respiratory secretions, urine, tissue specimens Specific IgM; IgG if exposure has occurred	No vaccine available Pregnant HCP can choose to defer care of immunocompromised patients with chronic infection or sickle cell disease during aplastic crisis <i>Standard Precautions</i> , especially <i>Safe Injection Practices</i> ; for infected immunocompromised patient, add <i>Droplet Precautions</i>
Rubella	Respiratory secretions	Congenital infection syndrome ^a 90% affected in first trimester 40%–50% affected overall	Rare	Documentation of 2 doses rubella-containing vaccine before pregnancy Routine rubella IgG testing during pregnancy Preconception screening recommended	Vaccine ^b Immune globulin after exposure of a susceptible woman to rubella does not provide protection against congenital rubella No congenital rubella syndrome described in association with vaccine given inadvertently during pregnancy Rubella-containing vaccine post partum if woman is nonimmune <i>Standard Precautions</i> <i>Droplet Precautions plus Congenital Rubella</i> until ≥1 year of age
Syphilis Rash, skin and genital lesions, central nervous system disease	Blood, lesion (especially large bullae of congenital syphilis), amniotic fluid	Congenital infection syndrome ^a Variable 10%–90%; ^c depends stage of maternal disease and trimester of the infection	Transmission possible, related to contagiousness of maternal lesions	Serum VDRL, RPR Serum FTA-ABS	Postexposure prophylaxis with penicillin or ceftriaxone if percutaneous exposure <i>Standard Precautions</i> , especially <i>Safe Injection Practices</i> (wear gloves when caring for patients with congenital, primary, and secondary syphilis until 24 hr of treatment completed)
Varicella zoster virus (VZV) Chickenpox, shingles	Respiratory secretions, vesicle fluid	Congenital infection syndrome Malformations, skin, limb, CNS, eye; chickenpox zoster Congenital syndrome 2%	High risk of infection and severe disease if maternal varicella within 5 days before or 2 days after delivery	Varicella IgG serology Past history of chickenpox unreliable	Vaccine (2 doses) ^b ; VarIZIG or (GIV, ideally within 96 hr of exposure if nonimmune Use the following precautions for patients with chickenpox and disseminated zoster: <i>Standard Precautions plus Airborne Precautions plus Contact Precautions</i>
Zika virus	Blood, body fluids of infected patients; mosquitoes in endemic area. Acquisition in the community more likely than in healthcare settings	Microcephaly, other neurodevelopmental abnormalities	Undefined at time of publication	rt-PCR on blood Serum IgG, IgM	Avoid outdoor work assignments in endemic areas Check CDC website for updated recommendations <i>Standard Precautions</i> , especially <i>Safe Injection Practices</i>

^aCongenital syndrome: varying combinations of jaundice, hepatosplenomegaly, microcephaly, thrombocytopenia, anemia, retinopathy, skin and bone lesions, and “blueberry muffin spots” (extramedullary hematopoiesis).

^bLive virus vaccine given before or after pregnancy.

^cART, antiretroviral therapy; CDC, Centers for Disease Control and Prevention; CNS, central nervous system; CSF, cerebrospinal fluid; EVD, Ebola virus disease; FTA-ABS, Fluorescent treponema antigen-antibody test; HBeAg, hepatitis B e antigen; HBIG, hepatitis B immune globulin; HBSAg, hepatitis B surface antigen; HCP, healthcare personnel; IgG, immunoglobulin G; IgM, immunoglobulin M; IGIV, immune globulin intravenous; IM, intramuscular; IV, intravenous; PCR, polymerase chain reaction; PPD, purified protein derivative; RPR, rapid plasma reagin test; Tdap, tetanus-diphtheria-acellular pertussis; VDRL, Venereal Disease Research Laboratory test; WHO, World Health Organization.

pediatric patients and their families provides practical guidance for settings where high-risk patients live with their families for varying periods of time.¹¹⁶ IPC challenges now are being addressed in long-term care facilities for children.¹¹⁷ More data are needed to determine the most effective and least restrictive practices.

All references are available online at www.expertconsult.com.

KEY REFERENCES

3. Lyren A, Brilli R, Bird M, et al. Ohio Children's Hospitals' solutions for patient safety: a framework for pediatric patient safety improvement. *J Healthc Qual* 2016;38:213–222.
8. See I, Iwamoto M, Allen-Bridson K, et al. Mucosal barrier injury laboratory-confirmed bloodstream infection: results from a field test of a new National Healthcare Safety Network definition. *Infect Control Hosp Epidemiol* 2013;34:769–776.
12. Huang SY, Philip A, Richter MD, et al. Prevention and management of infectious complications of percutaneous interventions. *Semin Intervent Radiol* 2015;32:78–88.
13. Kanamori H, Rutala WA, Sickbert-Bennett EE, Weber DJ. Review of fungal outbreaks and infection prevention in healthcare settings during construction and renovation. *Clin Infect Dis* 2015;61:433–444.
15. Rogowski JA, Staiger D, Patrick T, et al. Nurse staffing and NICU infection rates. *JAMA Pediatr* 2013;167:444–450.
20. Dirajlal-Fargo S, DeBiasi R, Campos J, Song X. Carbapenem-resistant Enterobacteriaceae in pediatric patients: epidemiology and risk factors. *Infect Control Hosp Epidemiol* 2014;35:447–449.
22. Langdon A, Crook N, Dantas G. The effects of antibiotics on the microbiome throughout development and alternative approaches for therapeutic modulation. *Genome Med* 2016;8:39.
48. Bryant KA, Harris AD, Could CV, et al. Necessary infrastructure of infection prevention and healthcare epidemiology programs: a review. *Infect Control Hosp Epidemiol* 2016;37:371–380.
50. Kaye KS, Anderson DJ, Cook E, et al. Guidance for infection prevention and healthcare epidemiology programs: healthcare epidemiologist skills and competencies. *Infect Control Hosp Epidemiol* 2015;36:369–380.
61. Septimus E, Weinstein RA, Perl TM, et al. Approaches for preventing healthcare-associated infections: go long or go wide? *Infect Control Hosp Epidemiol* 2014;35(suppl 2):S10–S14.
90. Rutala WA, Weber DJ. Disinfection and sterilization: an overview. *Am J Infect Control* 2013;41(suppl):S2–S5.

REFERENCES

1. The Joint Commission. National Patient Safety Goals. www.jointcommission.org/assets/1/6/2016_NPSG_HAP_ER.pdf.
2. Chassin MR, Loeb J. High-reliability health care: getting there from here. *Milbank Q* 2013;91:459–490.
3. Lyren A, Brilli R, Bird M, et al. Ohio Children's Hospitals' solutions for patient safety: a framework for pediatric patient safety improvement. *J Healthc Qual* 2016;38:213–222.
4. Siegel JD, Rhinehart E, Jackson M, et al. Guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings, 2007. *Am J Infect Control* 2007;35(suppl 2):S65–S164.
5. Siegel JD, Rhinehart E, Jackson M, et al. Management of multidrug-resistant organisms in healthcare settings, 2006. *Am J Infect Control* 2007;35(suppl 2):S165–S193.
6. Cummings KJ, Choi MJ, Esswein EJ, et al. Addressing infection prevention and control in the first U.S. community hospital to care for patients with Ebola virus disease: context for national recommendations and future strategies. *Ann Intern Med* 2016 May 10;[Epub ahead of print].
7. Hewlitt AL, Varkey JB, Smith PW, Ribner BS. Ebola virus disease: preparedness and lessons learned from two biocontainment units. *Curr Opin Infect Dis* 2015;28:343–348.
8. See I, Iwamoto M, Allen-Bridson K, et al. Mucosal barrier injury laboratory-confirmed bloodstream infection: results from a field test of a new National Healthcare Safety Network definition. *Infect Control Hosp Epidemiol* 2013;34:769–776.
9. Saiman L, Siegel JD, LiPuma JJ, and the Guideline Writing Committee. Infection prevention and control guideline for cystic fibrosis: 2013 update. *Infect Control Hosp Epidemiol* 2014;35(suppl 1):S1–S67.
10. Drudy D, Mullane NR, Quinn T, et al. *Enterobacter sakazakii*: an emerging pathogen in powdered infant formula. *Clin Infect Dis* 2006;43:322.
11. Hogan MJ. Infection in pediatric interventional radiology. *Pediatr Radiol* 2011;41(suppl 1):S99–S106.
12. Huang SY, Philip A, Richter MD, et al. Prevention and management of infectious complications of percutaneous interventions. *Semin Intervent Radiol* 2015;32:78–88.
13. Kanamori H, Rutala WA, Sickbert-Bennett EE, Weber DJ. Review of fungal outbreaks and infection prevention in healthcare settings during construction and renovation. *Clin Infect Dis* 2015;61:433–444.
14. Wilson S, Cert P, Bremner A, et al. The effect of nurse staffing on clinical outcomes of children in hospital: a systematic review. *Int J Evid Based Healthc* 2011;9:97–121.
15. Rogowski JA, Staiger D, Patrick T, et al. Nurse staffing and NICU infection rates. *JAMA Pediatr* 2013;167:444–450.
16. Lai NM, Foong SC, Foong WC, Tan K. Co-bedding in neonatal nursery for promoting growth and neurodevelopment in stable preterm twins. *Cochrane Database Syst Rev* 2016;(4):CD008313.
17. Baley J, Committee on Fetus and Newborn. Skin to skin care for term and pre-term infants in the neonatal ICU. *Pediatrics* 2015;136:596–599.
18. Haas EJ, Zaoutis TE, Parsad P, et al. Risk factors and outcomes for vancomycin-resistant enterococcus bloodstream infections in children. *Infect Control Hosp Epidemiol* 2010;31:1038.
19. Zaoutis TE, Goyal M, Chu JH, et al. Risk factors for and outcomes of bloodstream infection caused by extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella* species in children. *Pediatrics* 2005;115:942.
20. Dirajlal-Fargo S, DeBiasi R, Campos J, Song X. Carbapenem-resistant Enterobacteriaceae in pediatric patients: epidemiology and risk factors. *Infect Control Hosp Epidemiol* 2014;35:447–449.
21. Cotton CM, McDonald S, Stoll B, et al. The association of third-generation cephalosporin use and invasive candidiasis in extremely low birth-weight infants. *Pediatrics* 2006;118:717–722.
22. Langdon A, Crook N, Dantas G. The effects of antibiotics on the microbiome throughout development and alternative approaches for therapeutic modulation. *Genome Med* 2016;8:39.
23. Buttery JP, Alabaster SJ, Heine RG, et al. Multiresistant *Pseudomonas aeruginosa* outbreak in a pediatric oncology ward related to bath toys. *Pediatr Infect Dis J* 1998;17:509–513.
24. Reissa E, Lasker BA, Iqbal NJ, et al. Molecular epidemiology of *Candida parapsilosis* sepsis from outbreak investigations in neonatal intensive care units. *Infect Genet Evol* 2006;8:103.
25. Barchiesi F, Caggiano G, Falconi DiFrancesco L, et al. Outbreak of fungemia due *Candida parapsilosis* in a pediatric oncology unit. *Diagn Microbiol Infect Dis* 2004;49:269–271.
26. Conlan S, Thomas PJ, Deming C, et al. Single molecule sequencing to track plasmid diversity of hospital-associated carbapenemase-producing Enterobacteriaceae. *Sci Transl Med* 2014;6:254ra126.
27. Davis RJ, Jensen SO, Van Hal S, et al. Whole genome sequencing in real-time investigation and management of a *Pseudomonas aeruginosa* outbreak on a neonatal intensive care unit. *Infect Control Hosp Epidemiol* 2015;36:1058.
28. Hall CB, Douglas RG Jr. Modes of transmission of respiratory syncytial virus. *J Pediatr* 1981;99:100–103.
29. Brankston G, Gitterman L, Hirji Z, et al. Transmission of influenza A in human beings. *Lancet Infect Dis* 2007;7:257–265.
30. Munoz FM, Ong LT, Seavy D, et al. Tuberculosis among adult visitors of children with suspected tuberculosis and employees at a children's hospital. *Infect Control Hosp Epidemiol* 2002;23:568–572.
31. Roy CJ, Milton DK. Airborne transmission of communicable infection: the elusive pathway. *N Engl J Med* 2004;350:1710–1712.
32. Carlson AL, Budd AP, Perl TM. Control of influenza in healthcare settings: early lessons from the 2009 pandemic. *Curr Opin Infect Dis* 2010;23:293–299.
33. Ang B, Poh BF, Win MK, Chow A. Surgical masks for protection of healthcare personnel against pandemic novel swine-origin influenza A (H1N1)-2009: results from an observational study. *Clin Infect Dis* 2010;50:1011–1014.
34. Weber DJ, Fischer WA, Wohl DA, Rutala WA. Protecting healthcare personnel from acquiring Ebola virus disease. *Infect Control Hosp Epidemiol* 2015;36:1229–1232.
35. Vonberg R, Stamm-Balderjahn S, Hansen S, et al. How often do asymptomatic healthcare workers cause methicillin-resistant *Staphylococcus aureus* outbreaks? A systematic evaluation. *Infect Control Hosp Epidemiol* 2006;27:1123–1127.
36. Gupta A, Della-Latta P, Todd B, et al. Outbreak of extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* in a neonatal intensive care unit linked to artificial nails. *Infect Control Hosp Epidemiol* 2004;25:210–215.
37. Jefferies JMC, Cooper T, Yam T, Clarke SC. *Pseudomonas aeruginosa* outbreaks in the neonatal intensive care unit: a systematic review of risk factors and environmental sources. *J Med Microbiol* 2012;61:1051–1061.
38. Calderon TA, Coffin SE, Sammons J. Preventing the spread of pertussis in pediatric healthcare settings. *J Pediatr Infect Dis Soc* 2015;4:52–59.
39. Sydnor E, Perl TM. Healthcare providers as sources of vaccine-preventable diseases. *Vaccine* 2014;32:4814–4822.
40. Self WH, Williams DJ, Zhu Y, et al. Respiratory viral detection in children and adults: comparing asymptomatic controls and patients with community-acquired pneumonia. *J Infect Dis* 2016;213:584–591.
41. Buss SN, Leber A, Chapin K, et al. Multicenter evaluation of the BioFire FilmArray gastrointestinal panel for etiologic diagnosis of infectious gastroenteritis. *J Clin Microbiol* 2015;53:915–925.
42. Suwantarat N, Logan LK, Carroll KC, et al. The prevalence and molecular epidemiology of multidrug-resistant Enterobacteriaceae colonization in a pediatric intensive care unit. *Infect Control Hosp Epidemiol* 2016;37:535–543.
43. Maragakis LL. Recognition and prevention of multidrug-resistant gram-negative bacteria in the intensive care unit. *Crit Care Med* 2010;38(suppl):S345–S351.
44. Folgieri L, Livadiotti S, Carletti M, et al. Epidemiology and clinical outcomes of multidrug-resistant gram-negative bloodstream infections in a European tertiary pediatric hospital during a 12-month period. *Pediatr Infect Dis J* 2014;33:929–932.
45. Chitnis AS, Magill SS, Edwards JR, et al. Trends in *Candida* central line-associated bloodstream infections among NICUs, 1999–2009. *Pediatrics* 2012;130:e46–e52.
46. Pappas PG, Kauffman CA, Andes DR, et al. Clinical practice guideline for the management of candidiasis: 2016 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2016;62:e1–e50.
47. Greenberg RG, Benjamin DK, Gantz MG, et al. Empiric antifungal therapy and outcomes in extremely-low-birth-weight infants with invasive candidiasis. *J Pediatr* 2012;161:264–269.
48. Bryant KA, Harris AD, Could CV, et al. Necessary infrastructure of infection prevention and healthcare epidemiology programs: a review. *Infect Control Hosp Epidemiol* 2016;37:371–380.
49. Burke JP. Patient safety: infection control: a problem for patient safety. *N Engl J Med* 2003;348:651–656.
50. Kaye KS, Anderson DJ, Cook E, et al. Guidance for infection prevention and healthcare epidemiology programs: healthcare epidemiologist skills and competencies. *Infect Control Hosp Epidemiol* 2015;36:369–380.
51. Epstein L, Hunter JC, Arwady A, et al. New Delhi metallo- β -lactamase-producing carbapenem-resistant *Escherichia coli* associated with exposure to duodenoscopes. *JAMA* 2014;312:1447–1455.
52. Smith RM, Schaefer MK, Kainer MA, et al. Fungal infections associated with contaminated methylprednisolone injections. *N Engl J Med* 2013;369:1598–1609.
53. Haley RW. The scientific basis for using surveillance and risk factor data to reduce nosocomial infection rates. *J Hosp Infect* 1995;30(suppl):3–14.
54. Gaynes R, Richards C, Edwards J, et al. Feeding back surveillance data to prevent hospital-acquired infections. *Emerg Infect Dis* 2001;7:295–298.
55. Benneyan JC, Lloyd RC, Plsek PE. Statistical process control as a tool for research and healthcare improvement. *Qual Saf Health Care* 2003;12:458–464.
56. Dudeck MA, Edwards JR, Allen-Bridson K, et al. National Healthcare Safety Network report, data summary for 2013, device-associated module. *Am J Infect Control* 2015;43:206–221.
57. Masnick M, Morgan DJ, Sorkin JD, et al. Lack of patient understanding of hospital-acquired infection data published on the Centers for Medicare and Medicaid Services Hospital Compare website. *Infect Control Hosp Epidemiol* 2016;37:182–187.
58. Zachariah P, Furuya EY, Edwards J, et al. Compliance with prevention practices and their association with central line-associated blood stream infections in neonatal intensive care units. *Am J Infect Control* 2014;42:847–851.
59. McKibben L, Horan TC, Tokars JI, et al. Guidance on public reporting of healthcare-associated infections: recommendations of the Healthcare Infection Control Practices Advisory Committee. *Infect Control Hosp Epidemiol* 2005;26:580–587.
60. Morgan DJ, Murthy R, Munoz-Price LS, et al. Reconsidering contact precautions for endemic methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant *Enterococcus*. *Infect Control Hosp Epidemiol* 2015;36:1163–1172.
61. Septimus E, Weinstein RA, Perl TM, et al. Approaches for preventing healthcare-associated infections: go long or go wide? *Infect Control Hosp Epidemiol* 2014;35(suppl 2):S10–S14.
62. Bearman G, Stevens MP. Control of drug-resistant pathogens in endemic settings: contact precautions, controversies, and a proposal for a less restrictive alternative. *Curr Infect Dis Rep* 2012;14:620–626.

63. Huang SS, Septimus E, Kleinman K, et al. Targeted versus universal decolonization to prevent ICU infection. *N Engl J Med* 2013;368:2255–2265.
64. Derde LP, Cooper BS, Goossens H, et al. Interventions to reduce colonisation and transmission of antimicrobial-resistant bacteria in intensive care units: an interrupted time series study and cluster randomised trial. *Lancet Infect Dis* 2014;14:31–39.
65. Quach C, Milstone AM, Perpete C, et al. Chlorhexidine bathing in a tertiary care neonatal intensive care unit: impact on central line-associated bloodstream infections. *Infect Control Hosp Epidemiol* 2014;35:158–163.
66. Baron EJ, Miller M, Weinstein MP, et al. A Guide to utilization of the microbiology laboratory for diagnosis of infectious diseases: 2013 recommendations by the Infectious Diseases Society of America (IDSA) and the American Society for Microbiology (ASM). *Clin Infect Dis* 2013;57:e22–e121.
67. Haverkate MR, Bootsma MC, Weiner S, et al. Modeling spread of KPC-producing bacteria in long-term acute care hospitals in the Chicago region, USA. *Infect Control Hosp Epidemiol* 2015;36:1148–1154.
68. Centers for Disease Control and Prevention. Core Elements of Hospital Antibiotic Stewardship Programs. www.cdc.gov/getsmart/healthcare/implementation/core-elements.html.
69. President's Council of Advisors on Science and Technology. Report to the President on Combating Antibiotic Resistance. www.whitehouse.gov/sites/default/files/microsites/ostp/PCAST/pcast_carb_report_sept2014.pdf.
70. Barlam TF, Cosgrove SE, Abbo LM, et al. Implementing an antimicrobial stewardship program: guidelines by the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America. *Clin Infect Dis* 2016;62:e1–e27.
71. Cosgrove S, Hermsen E, Rybak M, et al. Guidance for the knowledge and skills required for antimicrobial stewardship leaders. *Infect Control Hosp Epidemiol* 2014;35:1444–1451.
72. Smith MJ, Gerber JS, Hersh AL. Inpatient antimicrobial stewardship on pediatrics: a systematic review. *J Pediatric Infect Dis Soc* 2015;4:e127–e135.
73. Magasarili HK, Giroto JE, Bennett NJ, Nicolau DP. Making a case for pediatric antimicrobial stewardship programs. *Pharmacotherapy* 2015;35:1026–1036.
74. Shipp KD, Chiang T, Karasick S, et al. Antimicrobial stewardship challenges in a referral neonatal intensive care unit. *Am J Perinatol* 2016;33:518–524.
75. Cantley JB, Patel SJ. Antimicrobial stewardship in the NICU. *Infect Dis Clin North Am* 2014;28:247–261.
76. Occupational Safety and Health Administration. Bloodborne Pathogens and Needlestick Prevention. www.osha.gov/SLTC/bloodborne/pathogens/index.html.
77. Williams IT, Perz JF, Bell BP. Viral hepatitis transmission in ambulatory health care settings. *Clin Infect Dis* 2004;38:1592–1598.
78. Weber DJ, Rutala WA, Fried MW. Hepatitis C virus outbreaks in hemodialysis centers: a continuing problem. *Infect Control Hosp Epidemiol* 2016;37:140–142.
79. Kuhar DT, Henderson DK, Struble KA, et al. Updated US Public Health Service guidelines for the management of occupational exposure to human immunodeficiency virus and recommendations for postexposure prophylaxis. *Infect Control Hosp Epidemiol* 2013;34:875–892.
80. Croft LD, Harris AD, Pineles L, et al. The effect of universal glove and gown use on adverse events in intensive care patients. *Clin Infect Dis* 2015;61:545–553.
81. Dhar S, Marchaim D, Tansek R, et al. Contact precautions: more is not necessarily better. *Infect Control Hosp Epidemiol* 2014;35:213–221.
82. Tomas ME, Kundrapu S, Thota P, et al. Contamination of health care personnel during removal of personal protective equipment. *JAMA Intern Med* 2015;175:1904–1910.
83. Doll M, Bearman GM. The Increasing visibility of the threat of health care worker self-contamination. *JAMA Intern Med* 2015;175:1911–1912.
84. Centers for Disease Control and Prevention. Prevention Strategies for Seasonal Influenza in Healthcare Settings. www.cdc.gov/flu/professionals/infectioncontrol/healthcaresettings.htm.
85. Weber DJ, Anderson D, Rutala WA. The role of the surface environment in healthcare-associated infections. *Curr Opin Infect Dis* 2013;26:338–344.
86. Otter JA, Yezli S, Salkeld JAG, French GL. Evidence that contaminated surfaces contribute to the transmission of hospital pathogens and an overview of strategies to address contaminated surfaces in hospital settings. *Am J Infect Control* 2013;41(suppl):S6–S11.
87. Carling PC, Huang SS. Improving healthcare environmental cleaning and disinfection: current and evolving issues. *Infect Control Hosp Epidemiol* 2013;34:507–513.
88. Jinadatha C, Simmons S, Dale C, et al. Disinfecting personal protective equipment with pulsed xenon ultraviolet as a risk mitigation strategy for health care workers. *Am J Infect Control* 2015;43:412–414.
89. Schmidt MG, Attaway HH, Fairey SE, et al. Copper continuously limits the concentration of bacteria resident on bed rails within the intensive care unit. *Infect Control Hosp Epidemiol* 2013;34:530–533.
90. Rutala WA, Weber DJ. Disinfection and sterilization: an overview. *Am J Infect Control* 2013;41:S2–S5.
91. Rutala WA, Weber DJ, the Healthcare Infection Control Practices Advisory Committee (HICPAC). Available at: www.cdc.gov/hicpac/pdf/guidelines/Disinfection_Nov_2008.pdf.
92. Petersen BJ, Chennat J, Cohen J, et al. Multisociety guideline on reprocessing flexible G.I. endoscopes: 2011. *Infect Control Hosp Epidemiol* 2011;32:527–537.
93. Rutala WA, Weber DJ. ERCP scopes: what can we do to prevent infections? *Infect Control Hosp Epidemiol* 2015;36:643–648.
94. Rutala WA, Weber DJ. How to assess risk of disease transmission to patients when there is a failure to follow recommended disinfection and sterilization guidelines. *Infect Control Hosp Epidemiol* 2007;28:146–155.
95. Centers for Disease Control and Prevention. Guidelines for Environmental Infection Control in Health-Care Facilities. Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee (HICPAC). *MMWR Recomm Rep* 2003;52(RR-10):1–42.
96. Guzman-Cottrill JA, Phillippi CA, Dolan SA, et al. Free vaccine programs to cocoon high-risk infants and children against influenza and pertussis. *Am J Infect Control* 2013;40:872–876.
97. Munoz-Price LS, Banach DB, Bearman G, et al. Isolation precautions for visitors. *Infect Control Hosp Epidemiol* 2015;36:747–758.
98. Murthy R, Bearman G, Brown S, et al. Animals in healthcare facilities: recommendations to minimize potential risks. *Infect Control Hosp Epidemiol* 2015;36:495–516.
99. Basler C, Bottichio L, Higa J, et al. Multistate outbreak of human *Salmonella poona* infections associated with pet turtle exposure—United States, 2014. *MMWR Morb Mortal Wkly Rep* 2015;64:804.
100. Reed KD, Melski JW, Graham MB, et al. The detection of monkeypox in humans in the Western Hemisphere. *N Engl J Med* 2004;350:342–350.
101. Pickering LK, Marano N, Bocchini JA, et al. Exposure to nontraditional pets at home and to animals in public settings: risk to children. *Pediatrics* 2008;122:876–886.
102. Starke JR. Interferon-gamma release assays for diagnosis of tuberculosis infection and disease in children. *Pediatrics* 2014;134:e1763–e1773.
103. Black CL, Yue X, Ball SW, et al. Influenza vaccination coverage among health care personnel—United States, 2014–15 influenza season. *MMWR Morb Mortal Wkly Rep* 2015;64:993–999.
104. Talbot T, Babcock H, Caplan A, et al. Revised SHEA position paper: influenza vaccination of healthcare personnel. *Infect Control Hosp Epidemiol* 2015;36:988–996.
105. Bernstein HH, Starke JR, American Academy of Pediatrics Committee on Infectious Diseases. Policy statement: recommendations for mandatory influenza immunization of all healthcare personnel. *Pediatrics* 2010;129:809–815.
106. Drees M, Wroten K, Smedley M, et al. Carrots and sticks: achieving high healthcare personnel influenza vaccination rates without a mandate. *Infect Control Hosp Epidemiol* 2015;36:717–724.
107. Babcock HM, Gemeinhart N, Jones M, et al. Mandatory influenza vaccination of health care workers: translating policy to practice. *Clin Infect Dis* 2010;50:459–464.
108. Feemster KA, Prasad P, Smith MJ, et al. Employee designation and health care worker support of an influenza vaccine mandate at a large pediatric tertiary care hospital. *Vaccine* 2011;29:1762–1769.
109. Petersen LR, Jamieson DJ, Powers AM, Honein MA. Zika virus. *N Engl J Med* 2016;374:1152–1163.
110. Centers for Disease Control and Prevention. Updated Recommendations for Use of Tetanus Toxoid, Reduced Diphtheria Toxoid, and Acellular Pertussis Vaccine (Tdap) in Pregnant Women—Advisory Committee on Immunization Practices (ACIP), 2012. *MMWR Morb Mortal Wkly Rep* 2013;62:131–134.
111. Machado AF, Sallum M, Boas L, et al. Molecular characterization of strains of respiratory syncytial virus identified in a hematopoietic stem cell transplant outpatient unit over 2 years: community or nosocomial infection? *Biol Blood Marrow Transplant* 2008;14:1348–1355.
112. Johnston CP, Cooper L, Ruby W, et al. Epidemiology of community-acquired methicillin-resistant *Staphylococcus aureus* skin infections among healthcare workers in an outpatient clinic. *Infect Control Hosp Epidemiol* 2006;27:1133–1136.
113. Committee on Infectious Diseases, American Academy of Pediatrics. Infection prevention and control in pediatric ambulatory settings. *Pediatrics* 2007;120:650–665.
114. Centers for Disease Control and Prevention. Guide to Infection Prevention for Outpatient Settings: Minimum Expectations for Safe Care. www.cdc.gov/HAI/settings/outpatient/outpatient-care-guidelines.html.
115. Yokoe D, Casper C, Dubberke E, et al. Safe living after hematopoietic cell transplantation. *Bone Marrow Transplant* 2009;44:509–519.
116. Guzman-Cottrill JA, Ravin KA, et al. SHEA guideline: infection prevention and control in residential facilities for pediatric patients and their families. *Infect Control Hosp Epidemiol* 2013;34:1003–1041.
117. Murray MT, Jackson O, Cohen B, et al. Impact of infection prevention and control initiatives on acute respiratory tract infections in a pediatric long-term care facility. *Infect Control Hosp Epidemiol* 2016;37:859–862.