Data in Brief 9 (2016) 155-158



Contents lists available at ScienceDirect

Data in Brief



journal homepage: www.elsevier.com/locate/dib

Data Article

Data on PRRSV infection promoted the subtype of porcine dendritic cells from mDCs to pDCs in vivo



Jinling Liu^a, Shu Wei^{b,1}, Lixia Liu^{a,1}, Lihui Yu^a, Chunyan Wang^c, Yujun Zhao^a, Fengping Shan^d, Guoshun Shen^{a,*}

^a Key Laboratory of Zoonosis of Liaoning Province, College of Animal Science & Veterinary Medicine, Shenyang Agricultural University, Shenyang 110866, P.R. China

^b The Preventive Center of Animal Disease of LiaoNing Province, No.95, Renhe Road, Shenbei District, Shenyang 110164, P.R. China

^c The 705 hospital Shen Xiang, Shenyang 110016, P.R. China

^d Department of Immunology, Basic School of Medicine, China Medical University, No. 92, North Second Road, Shenyang 110001, P.R. China

ARTICLE INFO

Article history: Received 21 July 2016 Received in revised form 23 August 2016 Accepted 26 August 2016 Available online 1 September 2016

Keywords: PRRSV Subtype Dendritic cells

ABSTRACT

The related study has confirmed that porcine reproductive and respiratory syndrome virus (PRRSV) infection may impair mature states of DCs can lead to suboptimal adaptive immune response ("The role of porcine reproductive and respiratory syndrome virus infection in immunephenotype and Th1/Th2 balance of dendritic cells" (Jinling Liu, We Shu, Liaxia Liu et al., 2016) [1]). In this data article, the porcine dendritic cells (DCs) isolated from porcine peripheral blood and spleen were collected after infection with PRRSV, then the characteristics of the subset differentiation of DCs were analyzed with FACS Calibur Cytometer and fluorescence microscope respectively. This data is an important foundation for further investigation into immune suppression by PRRSV infection, and that, it also provides new data for the development of potential antiviral therapies based on DCs and PRRS vaccines.

© 2016 Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

DOI of original article: http://dx.doi.org/10.1016/j.dci.2016.07.012

2352-3409/© 2016 Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

^{*} Corresponding author.

E-mail address: Ljlfree@sina.com (J. Liu).

¹ Contributed equally to this work.

http://dx.doi.org/10.1016/j.dib.2016.08.054

Subject area More specific subject area Type of data	Biology Microbiology and Immunity
How data was acquired	Microscope (Thermo) and FACS Calibur Cutometer (Recton-Dickinson
	Biosciences, San Jose, CA)
Data format	Analyzed
Experimental factors	Collected pigs infected with PRRSV
Experimental features	The distribution of DCs subsets was observed by flow cytometry and
-	fluorescence microscopy in the blood and spleen of infected PRRSV pigs.
Data source location	Liaoning Province
Data accessibility	Data is within this article

Specifications Table

Value of the data

- The data demonstrates that PRRSV infection could lead to adequate humoral immunity by predominantly polarizing pDCs and further supports our publication [1].
- This dataset provides new insights into the mechanism of immune suppression and persistent infection for PRRSV.
- The data is very important as a basis to further clarify the potential antiviral therapies based on DCs.

1. Data

We isolated the porcine peripheral blood and spleen of infected with PRRSV identified by RT-PCR and ELISA, subsets characteristics of DCs were assessed in vivo by flow cytometry (FCM) and fluorescence microscope respectively based on the key surface molecules for pDCs and mDCs. The analyzed data was presented in Fig. 1 and contained two types of data. DCs subtype analysis of porcine peripheral blood (Fig. 1a–b). DCs subtype analysis in porcine spleen (Fig. 1c–d).

2. Experimental design, materials and methods

2.1. Materials and methods

2.1.1. Screening experimental animals

According to the standard Ministry of Agriculture of China, the experiment pigs were obtained from healthy pigs free from all major diseases except for porcine reproductive and respiratory syndrome virus identified by RT-PCR and ELISA. After that, 4–10 week old piglets were conventionally reared, mixed breed [2]. They were housed in isolation rooms at the Animal Disease Center of LiaoNing Province under the approval of the Institutional Animal Care and Use Committee.

2.2. Analysis of DCs subtype in vivo under PRRSV infection

We aseptically collected the porcine peripheral blood from them with heparin anticoagulant (100 μ g/mL). Briefly, peripheral blood mononuclear cells (PBMC) were collected from blood by isolating the buffy coat after density centrifugation with lymphocyte-separating medium (Tianjin Hao Yang Biotechnology Company, China). CD14⁺ monocytes were isolated by positive selection of anti-CD14 immunomagnetic beads according to the manufacture's protocol (Miltenyi Biotec, Auburn, CA) [3–5]. Purified CD14⁺ monocytes were re-suspended in PBS at a concentration of 1 × 10⁶ cells/mL and



Fig. 1. The distribution of DCs subsets. (A) The distribution of mDC and pDC subsets identified with FCM for porcine blood. (B) Statistical graph of mDC and pDC subsets for porcine blood. (C-D). The distribution of mDC and pDC subsets identified with fluorescence microscopy for porcine spleen.

stained with 1 µL of each of anti-CD1–fluorescein isothiocyanate (FITC) and anti-CD172a allophycocyanin (APC). Afterwards, they were incubated in the dark at 4 °C for 30 min. The suspensions were then washed twice with PBS and the expressions of phenotype molecules mDCs and pDCs were analyzed with FACS Calibur Cytometer (Becton-Dickinson Biosciences, San Jose, CA) and fluorescence microscope [1]. The monocytes isolated from the healthy pigs free from all major diseases were uninfected PRRSV group (mock).

In addition, we prepared slices with the spleen tissue and the distribution of DCs subsets was examined by fluorescence microscopy after staining with specific molecular markers of mDCs and pDCs.

Acknowledgments

This study was funded by China Nature Science Project no.31502070 and Dr Start-up fund project of Liaoning no. 20141057. The authors thank all researchers who contributed to the work and we apologize to the researchers whose works could not be discussed here due to space limitations.

Transparency document. Supporting information

Transparency data associated with this article can be found in the online version at http://dx.doi. org/10.1016/j.dib.2016.08.054.

References

- Jinling Liu, We Shu, Liaxia Liu, et al., The role of porcine reproductive and respiratory syndrome virus infection in immunephenotype and Th1/Th2 balance of dendritic cells, Dev. Comp. Immunol. 65 (2016) 245–252.
- [2] Zhi-Jun Tian, Tong-Qing An, Yan-Jun Zhou, et al., An attenuated live vaccine based on highly pathogenic porcine reproductive and respiratory syndrome virus (HP-PRRSV)protects piglets against HP-PRRS, Veter. Microbiol. 138 (2009) 34–40.
- [3] R. Paillot, F. Laval, J.C. Audonnet, C. Andreoni, V. Juillard, Functional and phenotypic characterization of distinct porcine dendritic cells derived from peripheral blood monocytes, Immunology 102 (2001) 396–404.
- [4] Erika Silva-Campa, Lorena Cordoba, Lorenzo Fraile, Lilian Flores-Mendoza, María Montoya, Jesús Hernández, European genotype of porcine reproductive and respiratory syndrome (PRRSV) infects monocyte-derived dendritic cells but does not induce Treg cells, Virology 396 (2010) 264–271.
- [5] Crystal L. Loving, Susan L. Brockmeier, Randy E. Sacco, Differential type I interferon activation and susceptibility of dendritic cell populations to porcine arterivirus, Immunology 120 (2) (2007) 217–229.