Epigenetic Inheritance From Normal Origin Cells Can Determine the Aggressive Biology of Tumor-Initiating Cells and Tumor Heterogeneity

Cancer Control Volume 29: 1–18 © The Author(s) 2022 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/10732748221078160 journals.sagepub.com/home/ccx

Jiliang Feng, PhD¹¹, Dawei Zhao, MD², Fudong Lv, MD¹, and Zhongyu Yuan, MS¹

Abstract

The acquisition of genetic- and epigenetic-abnormalities during transformation has been recognized as the two fundamental factors that lead to tumorigenesis and determine the aggressive biology of tumor cells. However, there is a regularity that tumors derived from lessdifferentiated normal origin cells (NOCs) usually have a higher risk of vascular involvement, lymphatic and distant metastasis, which can be observed in both lymphohematopoietic malignancies and somatic cancers. Obviously, the hypothesis of genetic- and epigeneticabnormalities is not sufficient to explain how the linear relationship between the cellular origin and the biological behavior of tumors is formed, because the cell origin of tumor is an independent factor related to tumor biology. In a given system, tumors can originate from multiple cell types, and tumor-initiating cells (TICs) can be mapped to different differentiation hierarchies of normal stem cells, suggesting that the heterogeneity of the origin of TICs is not completely chaotic. TIC's epigenome includes not only genetic- and epigeneticabnormalities, but also established epigenetic status of genes inherited from NOCs. In reviewing previous studies, we found much evidence supporting that the status of many tumor-related "epigenetic abnormalities" in TICs is consistent with that of the corresponding NOC of the same differentiation hierarchy, suggesting that they may not be true epigenetic abnormalities. So, we speculate that the established statuses of genes that control NOC's migration, adhesion and colonization capabilities, cell-cycle quiescence, expression of drug transporters, induction of mesenchymal formation, overexpression of telomerase, and preference for glycolysis can be inherited to TICs through epigenetic memory and be manifested as their aggressive biology. TICs of different origins can maintain different degrees of innate stemness from NOC, which may explain why malignancies with stem cell phenotypes are usually more aggressive.

Keywords

tumorigenesis, heterogeneity, tumor stem cell, tumor-initiating cell, epigenetic abnormality

Introduction

Currently, there are two dimensions for understanding tumorigenesis and tumor biology. The first one is genetic abnormality that is defined as any alteration in the DNA sequence that causes disease. In the past 50 years, a large number of tumor-related genetic variants were identified, which has brought about tremendous changes in the cognition of tumorigenesis and treatment. The second one is epigenetic abnormality, also called as epigenetic mutation (epimutation), that is defined as abnormal transcriptional repression of active genes and/or abnormal activation of usually repressed genes caused by errors in epigenetic gene repression.^{1,2} Later, it was further extended as epigenetic plasticity, referring to a "plastic" state that allows stochastic oncogene activation.³ Functionally, epigenetic abnormalities are thought to be equivalent to genetic mutations that lead to neoplastic transformation. In addition, some researchers have

Corresponding Author:

Jiliang Feng, Clinical-Pathology Center, Capital Medical University Affiliated Beijing Youan Hospital, No.8, Xitoutiao, Youanmenwai Street, FengTai District, Beijing 100069, China. Email: jiliangfeng@ccmu.edu.cn



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE

and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

¹Clinical-Pathology Center, Capital Medical University Affiliated Beijing Youan Hospital, Beijing, China

²Medical Imaging Department, Capital Medical University Affiliated Beijing Youan Hospital, Beijing, China

proposed that, just like genetic changes, epigenetic changes in tumors are gradually accumulated in the process of transformation and evolution.⁴

Most tumors are clonal. That is, a normal cell gains the advantage of clonal proliferation by accumulating adequate driver mutations, thus becomes a tumor-initiating cell (TIC). All tumor parenchymal cells that make up the tumor mass are descendants of this TIC.⁵ Cell division is probably the most well-known way of introducing mutations, accumulating mutations, and passing mutations on to offspring. During selection under stress conditions, cells at any stage of differentiation may accumulate enough driver mutations to transform. Although mutations disturb the epigenome of normal cells and contribute to the formation of TIC's epigenome, TIC and its counterpart normal cells of the same differentiation hierarchy may have greatest degree of epigenetic similarity. In this work, we define the normal cells at the same hierarchy of differentiation as TIC as normal origin cells (NOCs).

Interestingly, there is a phenomenon that tumors derived from less-differentiated NOCs or TSCs usually have a higher risk of vascular involvement, lymphatic and distant metastasis. This regularity is first seen in lymphohematopoietic malignancies. For example, the biological behavior of leukemia stem cells and hematopoietic malignancies with hematopoietic stem cell phenotypes is generally more aggressive.⁶ The biological behavior of lymphoma or leukemia derived from mature cell types, such as B-cell chronic lymphocytic leukemia/small lymphocytic lymphoma (B-CLL/SLL), mantle cell lymphoma, plasma cell lymphoma, and follicular lymphoma, is relatively inert.^{7,8} In recent years, the influence of cellular origin on tumor biology has been observed in a wide range of somatic tumors.⁹⁻¹¹ In a given system, cells of different lineages and stages of differentiation have distinct biomarker expression profiles, which have been formed over a long period of species evolution and are strictly programmed. Accordingly, the tracing of tumors' cellular origins is achieved based on the conserved expression profiles of these biomarkers in tumors. Given that the cellular origin of tumors is an independent indicator that associated with tumor biology, we speculate that there may be other mechanism that contributes to the linear relationship between cellular origin and the biological behavior of tumors.

Limitations in Understanding of the Cellular Origin of Tumors May Lead to Systemic Deficiencies in the Identification of Tumor-Associated Epigenetic Abnormalities

Main Models of Tumorigenesis

Currently, there are two main hypotheses about the origin of a TIC, and the principles of genetic variation and clonal selection are still the foundations of the hypotheses. The first is the stochastic model (or clonal evolution model), which proposes that as long as adequate driver mutations are accumulated, any proliferating cell may gain the advantage of clonal proliferation and become a TIC. The second is the tumor stem cell (TSC) model, which proposes that a normal cell escapes regulation and gives rise to a stem cell-like counterpart, a TSC.¹² Some researchers have suggested that TSCs are just the transformed products of normal stem cells or progenitor cells.¹³ In addition, many scholars believe that TSCs may derive from a few tumor-initiating cells, which are reprogrammed differentiated cells after acquiring initial cancer-causing mutations.¹⁴ Recently, some excellent reviews have summarized the historical developments and new advances in tumorigenesis models.^{15,16} These models incorporated non-genetic determinants into stochastic model and TSC models and highlighted the tumorigenic epigenetic changes via cellular reprogramming or microenvironmental stress. All of these models are reasonable hypotheses to describe tumorigenesis and tumor heterogeneity, but each model is insufficient to explain how the linear relationship between the cellular origin and the biological behavior of tumors are formed.

The Multi-Lineage Cellular Origin of TIC in a Given System

Genetic abnormalities in oosperm (a representative totipotent stem cell) are usually inherited from the parent's germ cells and these genetic abnormalities will be carried by all progeny cells that make up the organism. Whole genome sequencing and RNA sequencing revealed that germline mutations in cancerpredisposing genes were very high in pediatric patients with cancers.¹⁷ Somatic mutations are most often brought on by problems that occur during cell division. In embryos, fetuses, and newborns, stem cells are in an active state of proliferation, which puts cells at risk of accumulating enough mutations to transform under tumorigenic factors.¹⁸ So, it is easy to understand that some tumors with stem cell characteristics, such as neuroblastoma, Wilm's tumor, and hepatoblastoma, occur preferentially in infants and young children.¹⁹ Since adult normal organs consist mainly of mature parenchymal cells, it was once thought that tumors were formed due to the dedifferentiation of mature cells. In the past decade, normal parenchymal cells of different differentiation hierarchies have been identified in almost all adult organs in the context of different diseases.²⁰⁻²³ These findings provide necessary evidence for us to understand the multiple cellular origins of certain tumor entities. Clues to the multi-lineage cellular origins of tumors have emerged in colorectal cancer,¹¹ breast cancer,¹² hematopoietic malignancies,²⁰ and other malignancies (Table 1).

By analyzing the cumulative process of driver mutations in cell differentiation and self-renewal, a model was developed to describe the heterogeneity of the origin of TSCs (Figure 1A). Cell division is the cytological basis for genetic laws and biological variation. Through asymmetric or symmetric division, two basic modes of cell division, the fate of stem cells is determined towards differentiation or self-renewal.²⁴ In either mode, genetic abnormalities in DNA can be accumulated and transmitted to progeny cells. Stem cells have the

Col Count:4Cancer Types	Cells of Origin	Species	Reference
Prostate cancer	Luminal and basal progenitors, and more differentiated cell types	Mouse models	Wang, Z., Mitrofanova, A., Bergren, S. et al. Lineage analysis of basal epithelial cells reveals their unexpected plasticity and supports a cell-of-origin model for prostate cancer heterogeneity. Nat Cell Biol.2013; 15, 274-283.
Hepatocellular carcinoma	Hepatic progenitors, immature and mature cells	Human	Feng J, Zhu R, Feng D, Yu L, Zhao D, Wu J, Yuan C, Chen J, Zhang Y, Zheng X. Prediction of Early Recurrence of Solitary Hepatocellular Carcinoma after Orthotopic Liver Transplantation. Sci Rep. 20199 (1):15855.
Invasive bladder carcinoma Bladder cancers	Basal urothelial stem cells Luminal-, basal-, mesenchymal-, and neuroendocrine-cells	Human Mouse model and human samples	 Shin, K., Lim, A., Odegaard, J. et al. Cellular origin of bladder neoplasia and tissue dynamics of its progression to invasive carcinoma. Nat Cell Biol.2014; 16, 469-478. Sfakianos JP, Daza J, Hu Y, Anastos H, Bryant G, Bareja R, Badani KK, Galsky MD, Elemento O, Faltas BM, Mulholland DJ. Epithelial plasticity can generate multi-lineage phenotypes in human and murine bladder cancers. Nat Commun. 2020; 11 (1):2540.
Intestinal tumors Intestinal crypts and adenomas	Crypt stem cells, crypt base columnar cells, paneth cell and its progenitor Different subpopulations of intestinal stem cells		 Vermeulen L, Morrissey E, van der Heijden M. et al. Defining stem cell dynamics in models of intestinal tumor initiation. <i>Science (New York,</i> <i>N.Y.)</i> 2013; 342: 995-998. Kozar S, Morrissey E, Nicholson AM. et al. Continuous clonal labeling reveals small numbers of functional stem cells in intestinal crypts and adenomas. <i>Cell Stem Cell</i>.2013; 13, 626-633.
Acute leukemias, squamous-cell carcinoma, adenocarcinoma	Precursor (stem) cells, subpopulation of stem cells	Human	Garraway, L., Sellers, W. Lineage dependency and lineage-survival oncogenes in human cancer. <i>Nat Rev Cancer</i> .2006; 6: 593-602.
Lung carcinoma Lung adenocarcinoma, squamous cell carcinoma and small cell lung cancer	Molecular phenotypes of almost all parenchymal cell types during lung development are described; Basal cells, bronchioalveolar stem cells,	Human Human and mouse model	Nikolić MZ, Sun D, Rawlins EL. Human lung development: recent progress and new challenges. <i>Development</i> . 2018; 145 (16): 163485.
Lung squamous cell carcinoma	Clara and alveolar type II cells; Classical, primitive, basal and secretory subtype correspond to normal cell types	Human	 Testa U, Castelli G, Pelosi E. Lung Cancers: Molecular Characterization, Clonal Heterogeneity and Evolution, and Cancer Stem Cells. <i>Cancers (Basel)</i>. 2018; 10 (8):248. Wilkerson M.D., Yin X., Hoadley K.A. et al. Lung squamous cell carcinoma mRNA expression subtypes are reproducible, clinically important, and correspond to normal cell types. <i>Clin. Cancer Res.</i> 2010; 16:4864-4875.
Breast cancers	Stem cells and progenitor cells Various stem cells and progenitors of the mammary gland	Human Human	 Sims, A., Howell, A., Howell, S. et al. Origins of breast cancer subtypes and therapeutic implications. Nat Rev Clin Oncol.2007; 4: 516- 525. Taurin S, Alkhalifa H. Breast cancers, mammary stem cells, and cancer stem cells, characteristics, and hypotheses. Neoplasia.
Hematopoietic malignancies	A tree-like hierarchy of oligo-, bi-, and unipotent progenitors	Human	 2020; 22 (12):663-678. Velten L, Haas SF, Raffel S. et al. Human haematopoietic stem cell lineage commitment is a continuous process. Nat Cell Biol. 2017; 19:271-281.



Figure I. A. In a given system, TICs of different tumors or their subtypes can be mapped to different differentiation hierarchies of normal stem cells. Clonal proliferation is characteristic of both benign and malignant tumors. Before malignant transformation, normal cells usually undergo a precancerous phase. Here, we hypothesized that when a normal cell accumulates an average of three driver mutations, it becomes a precancerous cell or a clonal proliferating benign TIC. Further, when an average of four drive mutations is accumulated, a malignant transformation occurs. By differentiation and self-renewal, any proliferating cells can introduce driver mutations to become precancerous cells. Precancerous cells carrying a small number of driver mutations may be histologically indistinguishable from normal cells, but they may exhibit atypia as the driver mutations are further accumulated. However, precancerous cells did not gain the advantage of clonal proliferation and eventually entered the apoptotic program. In developing and adult organs, the differentiation hierarchies of parenchymal cells exist. Through self-renewal and/or differentiation, proliferating cells at any stage of differentiation can accumulate adequate drive mutations to become TICs. Therefore, TICs can be mapped to different differentiation hierarchies of normal stem cells in a given system. B. The stemness of TIC of different origin may be parallel to that of their corresponding NOC. Normal stem cells are characterized by their stemness properties. As normal stem cells differentiate towards maturity, their stemness gradually decreases and eventually disappears. Although stochastic genetic or epigenetic mutations drive transformation, they cannot cause earth-shaking changes in the epigenome of TIC. The physiological characteristics of NOC, especially its stemness, can be inherited (perhaps not entirely) to its corresponding TIC and are presented by the aggressive biology of TIC. TIC of different origins can inherit different degrees of stemness from NOC, which may explain why a malignancy derived from a less differentiated NOC usually have a higher risk of vascular involvement, lymphatic and distant metastasis.

highest potential for proliferation and a much longer life span compared with their progeny and therefore have a greater opportunity to accumulate mutations through self-renewal or differentiation.²⁵ However, mature cells can accumulate mutations and pass them on to their descendants through only limited rounds of self-renewal. Generally, the surveillance mechanisms of the human body can identify the illegitimate changes.²⁶ In normal conditions, if the surveillance system identifies genetic alterations, the repair mechanisms, such as p53-dependent pathway, may correct the errors and the cell will survive. If the alterations cannot be corrected, cells with a malignant potential (precancerous cells) can be removed by apoptosis mechanisms.²⁷ In other cases, if the body cannot recognize and repair the illegitimate changes, they will be carried by precancerous cells and be passed to all offspring through self-renewal and differentiation. As long as these mutation carrying cells do not lose their ability to differentiate, they may enter the apoptotic program. Otherwise, they can survive. Under selective pressures, proliferating cells of any differentiation stage can be at risk of introducing and accumulating mutations, it is reasonable to infer that the proliferating cells at any differentiation stage can accumulate adequate driver mutations to achieve the advantage of clonal proliferation. Thus, not only pluripotent stem cells, blasts/progenitor cells, and lineage-committed progenitor cells,²⁸ but also mature cells²⁹ can be qualified as candidates for the origin of TSCs. Given the common feature of clonal proliferation of benign and malignant tumors, this model can be used to explain the heterogeneity of cellular origins of both in a given system.

For both TIC and its corresponding NOC, all the biological characteristics are determined by their respective epigenomes, that is, all epigenetic programs that regulate the expression of all genes within the genome.³⁰ TIC's epigenome includes not only epigenetic-abnormalities, secondary epigenetic-abnormalities (epigenetic aberrance due to genetic changes),³¹ but also the established epigenetic status of genes inherited from NOC. Of which, secondary epigenetic-abnormalities are the most wellknown aspect that contributes to the malignant phenotype of tumors. While thousands of genetic alterations have been identified in different tumors, several studies have revealed that most of the genetic changes in tumors are passenger mutations, with only a few driver mutations (four on averages) sufficient to convert a normal cell into a cancer cell.^{32,33} Clearly, limited genetic variation is not sufficient to cause the great heterogeneity within a given tumor, let alone explain the laws behind it.³⁴ Naturally, the relationship between epigenetic factors and tumorigenesis and biological behavior of tumor cells has attracted researchers' attention and become a research hotspot.

Which Normal Cell Should Be Used as a Control to Identify the Tumor-Associated Epigenetic Abnormality?

So far, there is no consensus on explaining how epigenetic abnormalities occur in tumors. Cell's stochastic epigenetic

regulatory changes and response to tumorigenic factors or microenvironmental changes are two major hypotheses.^{35,36} In practice, the identification of epigenetic abnormalities in tumors was commonly based on the differences in epigenetic status of certain gene between tumor bulks/cells and normal control, mostly tissue bulks from the excised organs.^{37,38} Normal tissues, non-tumor tissues under the background of chronic diseases, and tumor tissues all have parenchymal and mesenchymal heterogeneous composition,³⁹ which suggests that the method of identifying epigenetic abnormalities of tumors with tissue masses has systemic defects. Ideally, the identification of epigenetic abnormalities in certain tumors or tumor subtypes should be based on differences in epigenetic status of genes between TIC and corresponding NOC, rather than on normal or pericancerous tissue bulks. Otherwise, it will not be possible to reliably distinguish whether the differential epigenetic event is the innate epigenetic state of the gene inherited from the NOC or the real epigenetic abnormality.

Epigenetic Similarities Between Tumors and Their Corresponding NOCs

The definition of TIC emphasizes the cellular origin of tumors, while the definition of TSC emphasizes the cells with the characteristics of stem cells in the tumor mass. In some early and even current studies, researchers did not make a strict distinction between TSCs and TICs.⁴⁰ We now know that although cells with stem cell characteristics may not represent the real TICs, comparing the epigenetic status of genes of interest between TSCs and the corresponding normal stem cells can still help reveal the epigenetic regulation of the biological behavior of tumor cells. Known epigenetic events are of 3 types: methylation of DNA and RNA, histone modification (acetylation, methylation, and phosphorylation), and the expression of non-coding RNA.⁴¹ The epigenetic similarity between TSC and NOC may provide some clues for our understanding of the formation of tumor biology.

DNA Methylation

In mammals, DNA methylation is the main form of epigenetic regulation that controls embryonic development, cell differentiation, and lineage specification. Stem cells and differentiated cells have distinct global DNA methylation profiles. Compared to the differentiated cells, the global DNA of the stem cells is usually in a hypomethylated state. With differentiation, the cell's global DNA methylation level increases.^{42,43} Interestingly, although DNA hypomethylation can be observed in various tumors, global DNA hypomethylation is preferentially distributed in high-grade cancers and in tumors with a stem cell phenotype.^{44,46} In liver, global DNA hypomethylation is significantly detected in HCCs with stem cell phenotypes^{47,48} and hepatoblastoma⁴⁹ than in well-

differentiated HCC and adjacent tissues. In well-differentiated thyroid neoplasms, global hypermethylation, rather than global hypomethylation was found, which is consistent with that of normal mature thyroid epithelium.⁵⁰ In the lymphopoietic system, Kulis and his colleagues found extensive overlap of methylated regions between multiple B cell neoplasms and their corresponding NOCs.⁵¹ In other systems, preference distribution of the global DNA hypomethylation in tumors with stem/progenitor cell characteristics was also observed.⁵²⁻⁵⁴

Coupled with global DNA hypomethylation, another hallmark of many tumors is local hypermethylation, which mainly occurs on the promoter CpG islands of tumor suppressor genes. Many scholars have proposed that methylation of promoters of these genes was stochastically acquired during tumorigenesis.^{55,56} With the help of genome-wide DNA methylation scans, many researchers have evaluated the alterations of the DNA methylation landscape in various human tumors. In their studies, although considerable differential CpG island methylation was found, only a small proportion of them were candidate drivers for tumorigenesis.⁵⁷⁻⁶¹ Furthermore, results from several independent groups have shown that a large number of TSCs-specific methylation and their occurrence sites overlap with those of normal stem cells.^{55,62-65} A study revealed that abnormal methylation patterns in prostate cancer were associated with dynamic patterns of DNA methylation from the normal basal prostate epithelial cell differentiation towards luminal cell.⁶⁶ This makes people wonder whether the abnormal methylation of prostate cancer is formed via the epigenetic inheritance from the basal cell during the malignant transformation. Di Domenico analyzed genome-wide DNA methylation data of 125 Pancreatic Neuroendocrine Tumors (PanNETs). Based on epigenetic similarities, they found that PanNETs cluster in pancreatic α -like cell, β -like cell, and intermediate tumors.⁶⁷ Shen observed that DNA methylation patterns of embryonal carcinoma (EC) resembles that of embryonic stem cells, while non-EC non-seminomatous germ cell tumors adopt DNA methylation patterns resembling somatic and extraembryonal lineages.⁶⁸ Smith and colleagues found that small cell neuroendocrine carcinomas of lung, prostate, and bladder are enriched for a transcriptional signature shared by epithelial adult stem cells. Further, DNA methyltransferase expression correlated with adult stem cell signature status was enriched in these small cell neuroendocrine carcinomas, and a highly consistent DNA methylation profile across the carcinomas was found. Most interestingly, adult stem cell signatures associated with the small cell neuroendocrine carcinomas were not significantly affected by other molecular signatures.⁶⁹ Recently, Giacopelli analyzed genome-wide DNA methylation in 649 acute myeloid leukemia (AML) patients. They revealed that epitypes of AML subtypes showed developmental arrest at discrete stages of myeloid differentiation, revealing epitypes that retain arrested hematopoietic stemcell-like phenotypes. Further, they found that patients in epitypes with stem-cell-like methylation features showed poor overall survival along with up-regulated stem cell gene expression signatures.⁷⁰ In many tumors or TSCs, there are signs of de novo DNA methylation, involving essential DNA methyltransferase (Dnmt), especially Dnmt3a and Dnmt3b. Professor Meissner's research shows that DNA methylation profiles of 15 cancer types is consistent with de novo DNA methylation of extraembryonic ectoderm, and are regulated by Dnmt3a and Dnmt3b, indicating that they can use similar mechanisms.⁵⁵

In addition to CpG islands, there are a large number of DNA methylation changes outside of CpG islands, such as the methylation of "CpG island shores" in tumors. CpG island shore methylation is strongly related to gene expression, and it is highly conserved in mouse cells, discriminating tissue types regardless of species of origin. Irizarry and colleagues found a broad overlap between colon cancer-related methylation and stem cell differentiation-related methylation in CpG island shores.⁷¹ Doi and colleagues reported that their target methylated regions in CpG island shores for pluripotent stem cells, embryonic stem cells, and fibroblasts differentiation largely overlap those of the previously reported aberrant methylation in cancers.⁷² Based on the DNA methylation profiles and CpG sites, Tang and colleagues develop a classifier that can accurately identify the primary site of tumor.⁷³

Histone Modification

A histone modification is a covalent post-translational modification to histone proteins that can impact gene expression by altering chromatin structure or recruiting histone modifiers.⁷⁴ Histone modification includes methylation, phosphorylation, acetylation, ubiquitylation, and sumoylation of histone proteins of which histone methylation is one of the most important modifications associated with either transcriptional repression or activation. Activating (H3K4me3) and repressive (H3K27me3) histone methylations are representative events for the transcription and repression of gene expression, respectively.⁷⁵ Concomitant H3K4me3 (activating) and H3K27me3 (repressive) methylation patterns mark untranscribed lineage-specific gene loci, termed bivalent domains. Bivalent domains are usually found in embryonic stem cells and determine whether a cell will remain unspecified or will eventually differentiate.^{71,76-78} What is striking is the bivalent domains were detected in glioblastoma stem cells,⁷⁵ ovarian TSC-like side-population cells,⁸⁰ and stem cell-like prostate cancer cells,⁸¹ rather than in corresponding normal astrocytes, non-side-population ovarian cancer cells, and nonstem-like prostate cancer cells, respectively. Histone methvlation can occur at various sites in histone proteins, primarily on lysine and arginine residues. Histone methyltransferases catalyze histone methylations that are the important epigenetic marks regulating gene expression and cell fate. Jambhekar et al. well summarized the histone methyltransferases that regulate the histone methylation in animal development. Upregulation of SET1, MLL, SET7/9, and SymD3 for H3K4 methylation, and EZH1, EZH2, or G9a for H3K27 methylation that helps establish bivalent chromatin domains in early embryonic systems can be observed in various TSCs.⁸²

Non-coding RNAs

Non-coding RNAs (ncRNAs) are functional RNA molecules that are not translated into proteins. However, ncRNAs play important roles in the regulation of gene expression. In numerous types of embryonic and adult stem cells and progenitors, the expression of ncRNAs is often tissue- or even cell type-specific, emphasizing their involvement in defining space, time, and developmental stages in gene regulation.^{83,84} Based on the length, ncRNAs can be divided into two major classes, small and long ncRNAs (lncRNAs). Small ncRNAs are further divided into microRNAs (miRNAs), short interfering RNAs (siRNAs), and piwi-interacting RNAs (piRNAs). Here, we briefly review the impact of miRNAs and lncRNAs on tumorigenesis.

Tordonato had compared the transcription of microRNAs between breast stem cells/progenitor cells and breast TSCs. They found that the expression of Let-7, miR-200 family, miR-22, miR-205 was consistent between normal stem cells and TSCs.⁸⁴ Divisato summarized the expression of micro-RNAs in various other TSCs and embryonic stem cells (ESCs). They found that in addition to the microRNAs mentioned above, epigenetic status of other microRNAs such as miR-451, miR-21, miR-29, miR-320, miR-30, miR-122, miR-155, miR-302, C19MC miRNA cluster, Mir-23a-24-27a cluster, and MiR-125a/b family between ESCs and TSCs/ cancer progenitor cells of colorectal cancer, breast cancer, follicular thyroid tumor, lung adenocarcinoma, multiple myeloma, ovarian teratocarcinoma, glioma, gastric cancer, retinoblastoma, human non-small-cell lung cancer, hepatocellular carcinoma, and undifferentiated embryonal sarcoma of the liver was consistent.⁸⁵ So far, comparative study of IncRNA between normal stem cell and TSCs is just emerging, the consistence of the epigenetic status of HOTAIR and Linc-ROR between TSCs of a variety of human tumors and ESCs/ adult stem cells was revealed.⁸⁴⁻⁸⁶ Differences in non-coding RNAs expression profiles between TSCs and non-TSCs have led many investigators to associate them with the conversion of epigenetic status from normal stem cells to differentiated cells.⁸⁷⁻⁹⁰ All these evidences prompt us to consider that the expression of these key ncRNAs in TSCs are not stochastic epigenetic events, but the epigenetic inheritance from NOCs.

Key Genes and Signaling Pathways

The essence of genetic mutations is to affect cell fate by changing the status of related signaling pathways. In terms of tumor genetics, a large number of mutations involving in the disruption of many pivotal signaling pathways, such as the Notch, Hedgehog, Wnt/ β -catenin, JAK/STAT, NF- κ B

pathways, as well as genes, such as HMGA2, Bcl-2, Bmi-1, Thy1, C-myc, Oct-4, and HIF1- α , that regulate normal stem cell self-renewal, differentiation, proliferation, and survival have been identified.^{87,91-95} It is worth noting that there is now a large body of evidence linking the status of these signaling pathways in TSC to that of normal stem or progenitor cells (Table 2).⁹⁶⁻¹⁰⁶ Different teams have compared epigenetic modulations in these signaling pathways between ESC, adult stem cells, progenitor cells, and TSCs. High similarities in DNA methylation, histone modification, and ncRNAs expression pattern that regulate these key pathways and genes between TSCs and normal stem cells¹⁰⁷⁻¹¹⁰ also point to our consideration that epigenetic inherence from NOCs plays an important role in shaping the epigenome and neoplastic behavior of TICs.

Hippo signaling has been a field of very active research in recent years. Abnormal activation of YAP (Yes-associated protein), the main effector molecule of Hippo pathway, is closely related to the occurrence of various tumors. Normally, YAP activation plays an important role in promoting cell proliferation and maintaining the stemness of stem cells and progenitor cells.¹¹¹ YAP was not expressed in pericancerous tissues and mature cells.¹¹² Although YAP is overexpressed or enriched in the nucleus of tumor cells, especially TSCs, such as hepatocellular carcinoma, gastric cancer, colon cancer, prostate cancer, and ovarian cancer,^{111,113} YAP is low or not expressed in well-differentiated tumors.¹¹⁴⁻¹¹⁶ These evidences suggest that YAP gene can be activated not only by genetic mutation, but also by epigenetic inheritance from NOC in tumors. Further, in vitro, YAP has been shown to promote dedifferentiation of mature hepatocytes into progenitors,¹¹⁷ which can imply that YAP activation, whether through genetic mutation or epigenetic inheritance, may be equivalent in determining the biological behavior of TIC.

Similarities in Biological Behavior Between TICs and Their Corresponding NOCs

The epigenetic events of cells will eventually be transformed into specific biological functions. The exploration of cellular biology and its conversion in the process of stem cell differentiation can help us understand how the physiological function of NOC manifests as the aggressive biological behavior of TIC. Figure 1B shows the gradual decrease in stemness and the increase in adhesion and colonization ability of stem cells during the differentiation, which may reflect the stemness characteristics of TICs of different cellular origins. We will elaborate below.

Cell Dynamics

The biological characteristics of normal cells are closely related to their differentiation status. Along the direction of stem cell differentiation, stemness of cells successively decreases

Comparison	of consistency	of epigene

Col Count:4 Putative NOCs		TSCs/Cancers	References
Signaling Pathways			
Notch	Intestinal stem cells	Mammary stem cells; intestinal cancer stem cells	87,92
Hedgehog	Hematopoietic stem cells	Basal-cell carcinoma; ovarian stem cells	87
Wnt/β-catenin	Intestinal stem cells	Intestinal cancer stem cells; multiple cancer stem cells	87,92,96
NF-κB	Embryonic stem cells and adult stem cells	Breast, prostate, ovarian and pancreatic cancer stem cells	96
JAK/STAT	Embryonic stem cells and adult stem cells	Stem-like cells of the breast, prostate, blood, and glia tumors; Liver, colon, prostate cancer, and leukemia stem cells	91,93,96
NANOG	Embryonic stem cells and adult stem cells	Hepatic, prostate, colorectal, and brain cancer stem cells	96
Genes			
TP53	Embryonic stem cells and multiple adult stem cell	Breast cancer stem cells	95
RBI (inactivity)	Embryonic stem cells and adult stem cell	Small cell, large cell, adenocarcinoma with neuroendocrine differentiation; Retinoblastoma	69,97
HMGA2	Embryonic stem cells and undifferentiated cells	Many types of cancer stem cells	98
Bcl-2	Stem cell and immature cell	Many types of cancer stem cells	87,99
Bmi-I	Neural stem cell and nondifferentiated cells	Glioma stem cell; glioblastoma stem cell	87,100
C-myc	Epithelial adult stem/progenitor cells	Multiple epithelial cancer stem cells; glioma cancer stem cells, breast cancer stem cells	69,94,96
HIFI-α	Neural stem cell	Glioblastoma stem cell, glioma stem cell	101
Thyl	Early downstream progenitor of HSC	Acute myeloid leukemia	102
OCT-4	HSCs and their downstream progenitors Adult stem cell	Pancreatic cancer stem cells; leukemia stem cells; multiple tumor stem cells	96,103,104
FSHR	Ovarian stem/progenitor cellsPara Run-on-	ovarian cancer stem cells	105
Lsd I	Pluripotent stem cell	Teratocarcinoma, embryonic carcinoma, seminoma	106

Table 2. Comparison of consistency of epigenetic status of important pathways and genes between TSC and putative NOC.

and finally disappears in mature cells. In terms of cell dynamics, it is mainly manifested as increased cell adhesion and colonization ability and decreased cell proliferation and migration ability.¹¹⁸ Cell migration capacity is a prerequisite for tumor invasion and metastasis. Previously, some scholars have noted that tumor invasion and metastasis may be associated with stem cell homing.^{119,120} Stem cell homing is a controlled recruitment of stem cells that leads to trans-endothelial and directional migration, which is strictly regulated by welldirected epigenetic programs in organism development, tissue regeneration and repair.¹²¹ During differentiation, epigenetic reprogramming of stem cell is accompanied by the conversion of cellular biology. With differentiation until maturity, the epigenetic programs that drive stem cell homing are shut down, and the cells eventually lose their ability to migrate and then colonize. For example, in the context of chronic cholecystitis, stem cells in the biliary system proliferate to form new glands to compensate for the loss of the epithelial layer.¹²² These new glands can appear in or through the muscle layer to form the Rokitansky-Aschoff sinuses, which may be misdiagnosed as cancerous infiltrations.¹²³ The

formation of the Rokitansky-Aschoff sinuses is the result of tissue regeneration and repair after bile duct epithelium injury and is a typical biological manifestation of biliary stem cell activation, differentiation, migration, and colonization. Well-programmed cell kinetics can contribute to embryonic development and tissue regeneration and repair, while unconstrained cell kinetics can lead to damage of tissues and organs, especially in cancers. Cholangiocarcinomas are malignancies characterized by intensive local invasion and high incidence of distant metastasis. These tumors often occur in the context of repeated damage and repair of epithelial cells caused by chronic inflammation or biliary obstruction. Cholangiocarcinoma is a heterogeneous tumor of multilineage origin and some of its subtypes can express stem cell biomarker CD133. These CD133 positive cholangiocarcinomas showed significantly higher rates of nodal metastasis and positive surgical margin status. In vitro, CD133 positive tumor cells had a higher invasive ability compared with CD133 negative cells.¹²⁴ In another work, hypermethylation of multiple CpG sites at genes associated with the stem cell phenotype in some highly aggressive cholangiocarcinomas

was observed.¹²⁵ Zarco et al summarized findings on the mechanisms of cellular migration that overlap between neural progenitor cells (NPCs) and brain tumor stem cells (BTSCs). They found that signal pathways (JNK, PKA/Akt, PI3K/AKT/ MAPK, CXCL12/CXCR4, Slit/Robo) and proteins (sem-aphorins, pleiotrophin, integrins, NCAM, cadherins, matrix metalloproteinases) that participate NPCs and BTSCs migration are highly similar.¹²⁶ Placental trophoblast cells are highly invasive, able to migrate and invade the uterine wall and vasculature without being rejected by the immune system, which resembles the biological behavior of choriocarcinoma.¹²⁷ These evidences lead us to speculate that TSC may be harnessing normal stem cell mechanisms and thus being endowed with greater invasiveness.

Resistance to Chemicals and Cell Death, Induction of Angiogenesis, Promotion Inflammation, Avoiding Immune Destruction, and Maintaining the Ouiescent States

Many key characteristics of normal stem cells are often reported in tumors, especially in those tumors with stem cell phenotypes or TSCs. For example, like normal stem cells, cells of many types of tumors can express drug transporters to resist chemical toxicity.¹²⁸⁻¹³⁰ As normal stem cells, TSCs in chronic myeloid leukemia and solid tumors can remain quiescent and resistant to drug damage.⁴¹ In stem cells and many types of tumor cells, telomerase is activated to maintain the cell's ability to continue dividing and to resist cell death.^{131,132} Like normal stem cells, cells of many types of tumors can induce angiogenesis.^{133,134} Although neoplastic angiogenesis is conspicuously different from angiogenesis in normal conditions, in the past decade, many studies showed that some genes and pathways were involved in the neoplastic angiogenesis of gliomas, breast cancer, melanoma, hepatocellular carcinoma, ovarian cancer, mesothelial sarcomas, etc. are very similar to that of their respective normal stem.^{135,136} In vitro, purified TSCs also showed stronger ability of inducing angiogenesis than non-TSCs.¹³⁷ Normal mature somatic cells do not recruit inflammatory cells except for pathological conditions such as autoimmune diseases and tissue damage, as well as cellular senescence. However, in embryonic development and organogenesis, immunocytes can be recruited by almost all types of stem cells and are closely related to stem cell renewal, differentiation, proliferation, migration, angiogenesis induction, and protecting stem cells against immune clearance.¹³⁸⁻¹⁴⁰ Cancer-related inflammation is the recruitment of immunocytes to the tumor microenvironment. These immunocytes have many functions such as promotion of tumor growth, immunosuppression, angiogenesis, and metastasis.¹⁴¹ Previous studies have suggested that tumor-related inflammation may be a borrowing of a similar mechanism from normal stem cells by TSC to promote angiogenesis, metastasis and escape immune clearance.^{142,143}

Energy Metabolism

The energy supply of normal human cells usually comes in two forms, namely glycolysis and oxidative phosphorylation. Most of the quiescent stem cell populations rely on glycolysis to provide ATP. With the differentiation of stem cells, energy metabolism shifts toward oxidative phosphorylation to support the increasing energy demands.¹⁴⁴ Both types of energy supply can be detected in tumor cells. Depending on tumor type, degree of differentiation, or cellular origin, the type of energy supply that tumor cells prefer to use varies. In some well-differentiated tumors,145,146 energy is mainly obtained through oxidative phosphorylation, while in tumors with stem cell phenotype or poor-differentiated cancers, such as low differentiated prostate cancer, glioblastoma, triple-negative breast cancer, and colorectal cancer, energy is mainly generated through glycolysis.¹⁴⁷⁻¹⁵⁰ Embryonal carcinoma can be induced by retinoic acid to differentiate toward neurons and ectoderm lineages. In differentiated tumor cells, a metabolic transition from glycolysis to oxidative phosphorylation was observed.¹⁵¹

As discussed above, a large amount of evidence to demonstrate the similarity in many core biological properties and epigenetic regulation between multiple tumors and their putative NOCs has been obtained. Collectively, we suggest that the established epigenetic states of genes in NOCs can be inherited to TICs. Many neoplastic biology of TIC may be a translation of innate epigenetics inherited from NOC rather than random epigenetic abnormalities. From pluripotent stem cells, blasts/progenitor cells, lineage committed stem cells to mature cells, the stemness of the cells decreases sequentially.¹⁵² Parallel to this, TICs of different origins can inherit different degrees of stemness that are determined by the established epigenetic states of genes of NOCs, which may explain why malignancies that derived from lessdifferentiated NOCs usually have a higher risk of vascular involvement, lymphatic and distant metastasis. In vitro experiments have shown that TICs with stem cell phenotypes have the ability to differentiate, and their differentiated progeny generally have reduced or disappeared tumorigenic and invasive capabilities.^{153,154} Clearly, this phenomenon cannot be explained by random genetic- or epigeneticabnormalities, but is similar to the conversion of physiological functions that occurs during normal stem cell differentiation.

Innate Post-Transcriptional Regulation of NOC May Be Transferred to TIC via Epigenetic Memory

Propagation of the chromatin landscape across cell divisions is central to epigenetic memory of cells. Understanding the intergenerational transmission of chromatin status in selfrenewing cells might shed some light on how the epigenetics of NOC is passed on to TIC. In the nucleus, DNA

wrapped around histones form nucleosomes that allow for compaction of DNA and assist in gene regulation. There are many post-translational modifications on histones, including phosphorylation, acetylation, methylation, ubiquitination, and ADP-ribosomal modification, with cell-specific sites, degrees, and patterns.¹⁵⁵ On newly synthesized DNA during chromatin replication, nucleosomes are assembled from new naive histones and old modified histones. Reverón-Gómez et al revealed that H3K4me3, H3K36me3, H3K79me3, and H3K27me3 positional information is reproduced with high accuracy on newly synthesized DNA through histone recycling. Further, they found that de novo methylation to restore H3K4me3 and H3K27me3 levels occurs across the cell cycle with mark- and locus-specific kinetics.¹⁵⁶ Their work suggests that accurate parental histone recycling preserves positional information and allows post-translational modifications transmission to daughter cells.

Just like normal cell self-renewal, cell division is also a way for NOC to transform into TIC. In most cases, a small number of tumorigenic driver mutations cannot cause subversive changes in tumor epigenetics. Under the premise of obtaining the advantage of clonal proliferation through mutation, TIC might still maintain the innate post-translational modifications from NOC through epigenetic memory. The cellular biology translated from the innate epigenetic state of key genes and pathways in TIC can merely be a continuation of their inherent biological functions in NOC, and is manifested as the aggressive biology of TIC. Compared to the TIC transformed from the differentiated normal cell, TIC from stem cell can maintain stronger stemness. Perhaps it is this stemness that makes TIC derived from stem cells more competitive and survival advantage. In this regards, it should follow Darwinian principles.

The Cellular Origin of TIC can Determine Both Tumor Parenchyma and Stroma Heterogeneity

The heterogeneity within each individual tumor is called intratumoral heterogeneity, which starts with a single TIC and becomes greater as the disease progresses.¹⁵⁷ Intratumoral heterogeneity is thought to play a significant role in treatment resistance and failure. Recent few years, several mathematical models were put forward to describe intratumoral heterogeneity. These models well demonstrated the intratumoral heterogeneity caused by accumulated genetic and epigenetic mutations, and microenvironment stress.^{15,158} Different from previous models, we currently attempted to discuss the influence of the intrinsic stemness of TIC on parenchymal and stromal heterogeneity of a tumor, and to describe the epigenetic regularity behind tumor heterogeneity.

For the epigenetic causes of intratumoral heterogeneity, cell differentiation hierarchies and tumor microenvironment are two major concerns.¹⁵⁹ The former is also the core idea of the TSC model. However, ignoring the cellular origin of tumors may lead to an improper estimation of intratumoral

heterogeneity in an individual. Figure 2 shows the relationship between the cellular origins of tumors and the heterogeneity of the parenchymal cells within tumors. In a given system, TICs of different cellular origins possess different capacities for differentiation or self-renewal. Tumors that originate from less-differentiated stem cell types might have a greater degree of intratumoral heterogeneity because they could have had potential to produce offspring at more hierarchies of differentiation. At the other extreme, if tumors originate from mature cells, acquisition of genetic or epigenetic mutations via self-renewal may be the only way contributing to their intratumoral heterogeneity. If intratumoral heterogeneity is driven solely by random genetic- or epigenetic-alterations, then finding rules for parenchymal heterogeneity within a tumor is challenging. The high epigenetic similarity between TICs and their NOCs strongly suggests the importance of tracing the origin of tumors more precisely in practice, since numerous studies have shown that normal lineage-specific molecules and pathways can be effective targets for tumor therapy. In addition, precancerous cells derived from less-differentiated NOCs might develop into more aggressive malignancies due to their inherent stemness.

Tumor cells and their microenvironment are a complete functional system. Most of the previous studies have focused on the influence of microenvironment on the acquisition of genetic and epigenetic abnormalities of TIC and the formation of tumor biology,¹⁶⁰ which may be one-sided for understanding the relationship between tumor microenvironment and tumor parenchyma. Normal stem cells can recruit other cells, such as myofibroblasts/fibroblasts, mesenchymal stem/ progenitor cells, endothelial cells, vascular cells, adipocytes, and immune cells. All of these cells and their secreted paracrine factors, as well as the extracellular matrix, make up the stem cell microenvironment and play significant roles in fate decision of stem cells, tissue development, and homeostasis.¹⁶¹ The stem cell microenvironment is diverse and dynamic, and the information it senses and conveys reflects the difference in tissue types and anatomic sites, as well as the state of differentiation.

Many parallels have been drawn between the tumor microenvironment and physiological microenvironment, including those observed in embryonic development, organogenesis, and inflammation/wound healing. A study from Lacina revealed various biological aspects of interaction between CSCs of melanoma and cancer-associated fibroblasts with multiple parallels to interaction of normal epidermal stem cells and their niche.¹⁶² Through fueling STAT3, MAPK, and Akt signaling, elevated expression of IL-6 by mesenchymal stem cells is associated with cancer cell proliferation, angiogenesis and metastasis in neuroblastoma, breast, colon, pancreatic, esophageal, gastric, hepatic, pancreatic, non-small cell lung, renal cancer, and multiple myeloma.¹⁶³ Tissue-resident macrophages and regulatory T cells (Tregs) are well-known regulators involved in maintaining stem cell



Figure 2. The influence of cellular origin on the heterogeneity of the parenchymal cells within a tumor. Intratumoral heterogeneity begins with a TIC and increases with the expansion and evolution of tumors. In this figure, we assume that TICs from different sources have the same genetic- and epigenetic-mutation background and evolve by obtaining two additional driver mutations (in reality, the types and quantities of them are diverse). All cells contained in a dotted box represent heterogeneous cell compositions within the tumor derived from a TIC. In a given tumor entity, TIC can originate from multiple cell types. Tumors derived from less-differentiated NOCs can have a greater degree of intratumoral heterogeneity, because they can produce offspring with the same genetic- or epigenetic mutations at more hierarchies of differentiation. For TICs derived from terminally differentiated cells, acquisition of genetic- or epigenetic mutations via self-renewal may be the only way to generate heterogeneous cells. Targeting TSCs are expected to be a potential therapeutic strategy for tumor eradication, and it is a common practice to use biomarkers of normal stem cells to distinguish TSCs. The diversity of the cellular origin of a given tumor and its subtypes suggests that tracing the cellular origin of tumors and killing TICs is the basis for tumor eradication.

homeostasis. Interestingly, induced macrophages and Tregs are prominent in the tumor microenvironment, where they expand in response to IL-6, VEGF, CXCL12, IL-10, and TGF- β and are capable of producing a number of immunosuppressive signals.¹⁶⁴ Tumor angiogenesis is essential for tumor growth and metastasis. Vascular endothelial cells from the local environment and bone marrow-derived endothelial progenitor cells are recruited into tumor niches by similar proangiogenic factors in embryonic development and organogenesis to initiate and promote angiogenesis.¹³³⁻¹³⁶ The extracellular matrix (ECM) is composed of a variety of proteins, polysaccharides, and proteoglycans that make up the basement membrane and the interstitium. Normal stem cells can directly or indirectly recruit ECM to maintain stemness and avoid immune clearance. In liver, abundant fibrous stroma is usually deposited in those tumors with stem cell phenotypes, such as neuroendocrine carcinoma, hepatoblastoma, scirrhous hepatocellular carcinoma, and fibrolaminar carcinoma, rather than well-differentiated hepatocellular carcinoma and adenoma.¹⁶⁵ As such, it cannot be ruled out that the components of a TIC's microenvironment, if not all the components, can be proactively recruited by TICs, which may be dominated by hereditary epigenetic mechanisms. Furthermore, differences in the origin of TICs may determine the differences in tumor microenvironmental components associated with treatment or prognosis, including inflammatory cell subsets, fibroblasts, and secreted products (such as cytokines and chemokines) as well as non-cellular components of the extracellular matrix, which has important implications for understanding and evaluating the effectiveness and drug resistance of therapies that targeting tumor stroma, and will be discussed in the next section.

Conclusions

Mutations are indeed critical to tumorigenesis and are associated with clinical behavior. However, in a given tumor, not all of the established physiological processes are disrupted. Since the acquisition of the clonal proliferation advantage is the prerequisite for judging tumor transformation, whether possess molecular characteristics of normal stem cells are not indispensable for judging TICs. In spite of dramatically phenotypic changes can be observed in very few tumor cases, making their exact origin difficult to trace, in general, multilineage cellular origin of TIC can better explain the cellular origin of the vast majority of tumors.

Mutations can alter almost all types of physiological processes including differentiation, self-renewal, proliferation, metabolism, DNA mismatch repair, or cell-cycle transition. Differentiation arrest and reprogramming caused by mutation in tumors have been provoking scientists' great interests. Differentiation arrest was initially observed in lymphoid and hematopoietic tumors and subsequently in somatic tumors as well. The existence of TSC differentiation hierarchies in solid tumors suggests that cell-differentiation arrest hypothesis does not apply to all tumors. Broadly speaking, reprogramming can be understood as alteration of the status of certain signaling pathways and genes by genetic or epigenetic mutations. Specifically, reprogramming refer to the mechanisms by which expression of certain transcription factors, such as Yamanaka transcription factors (OSKM), renders somatic cells capability to acquire pluripotency or totipotency-like features.¹⁶⁶ Today, some literature suggests that cell reprogramming can play an important role in tumorigenesis. In vitro, overexpression of MYC induced epigenetic reprogramming of mammary epithelial cells into a stem cell-like state with metastatic capacity.¹⁶⁷ Using intestinal organoids, Heuberger showed that concomitant activation of Notch signaling and ablation of p53 induced reprogramming of specified intestinal epithelial cells into a regenerative cell state that makes them susceptible for tumorigenesis.¹⁶⁸ These studies on epigenetic reprogramming highlight the impact of the differentiation state on cell biological behavior and may be used to explain tumorigenesis of some tumors. However, in low-grade tumor types, such as well-differentiated HCC and thyroid tumors, multiple stem cell transcription factors and biomarkers are completely absent, indicating that cell reprogramming is not a universal mechanism for tumorigenesis in all tumor types.47,50

Traditionally, the understanding of tumor heterogeneity mostly focused on the complexity of tumor parenchymal cell composition and the disorder of signal regulation due to the introduction of genetic or epigenetic mutations or the stress of the microenvironment. Increased tumor heterogeneity of this kind is indeed strongly associated with the more aggressive biological characteristics of tumor cells, therapeutic resistance, and poor clinical outcomes for patients. However, the ultimate goal of tumor heterogeneity exploration is to find information or phenomena for clinical diagnosis and treatment. Therefore, it may be more practical to discover the underlying rules behind tumor heterogeneity.

Today, advances in large-scale genome sequencing have enabled the identification of characteristic genetic signatures in many tumors, which has become a key prerequisite for targeted drug therapy. However, genetic abnormality-targeted drugs require strict matching of mutated gene types, which can benefit only a limited group of patients. In contrast to the randomness of genetic and epigenetic mutations, the regularity of epigenetic determinants inherited from NOCs may help us to develop broad-spectrum anti-tumor drugs, as these determinants may be shared by the TICs of tumors of different systems at similar stages of differentiation. In recent years, the drugs targeting programmed cell death protein-1 (PD1)/ programmed cell death protein-1 ligand 1 (PD-L1) pathway have achieved promising effects in various tumor types. PD1 and PD-L1 act as important negative immune regulators. Combining PD1 receptors on T cells with their corresponding ligands PD-L1 on normal stem cells can inhibit T cell activation and prevent the stem cells from immune killing.^{169,170} In some tumors, this mechanism is borrowed by tumor cells and lead to tumor cell immune escape.^{171,172} The purpose of PD1/PD-L1 immunotherapy is to restore the anti-tumor activity of T cells by blocking this pathway.

As in the normal tissues, the negative modulation pathway of PD1/PD-L1 exists in tumors of multiple systems, which indicates that hereditary epigenetics-based drugs have the potential to become tumor-type agnostic therapies. Even though PD-1/PD-L1 blockade have brought new hope to tumor therapy, PD-1/PD-L1 inhibitors have only benefited nearly 20% of tumor patients. Detecting the expression rates of PD-L1 (tumor cells) and PD1 (lymphocytes) in biopsy tissues is the main method to determine drug response at present, but the therapeutic effect based on this method is sometimes not ideal.¹⁷³ Although different mechanisms, such as false positive or false negative detection, irrelevant PD-1T cell expression and tumor suppression immune destruction, or lack of TNF- γ -mediated inflammation have been used to explain the limitations of this treatment, the lack of PD-L1 expression in TIC determined by its cellular origin may be one of the reasons that cannot be ignored, because in normal cells, the expression of PD-L1, as well as some tumor-associated molecules, such as histone deacetylase, DNA methyltransferases, and vascular endothelial growth factor receptors, are closely related to their differentiation stage.¹⁷⁴⁻¹⁷⁸ If TICs can express PD-L1, PD-L1 blockade may produce the best therapeutic effect. If the target molecule is only expressed in the differentiated progeny cells of TIC, the same therapy can lead to a reduction in tumor volume, but cannot eliminate TIC from the source and cause relapse. It is well established that the cytotoxic T-lymphocyte associated protein 4 (CTLA-4) pathway is another complementary immunosuppression mechanism to PD1/PDL1 pathway that allows cancer cells to escape host immunity.¹⁴³ Notably, both PD1/PDL1 and CTLA-4 pathways are critical for embryonic development, organogenesis, and physiological homeostasis in adults because they can prevent somatic cells from being destroyed by cell-mediated immune responses.^{139,140,179,180} Understanding the epigenetic status of these pathway in the differentiation hierarchies of normal stem cells and tracking the cellular origin of tumors more precisely may lead to increased efforts to develop combination immunotherapy strategies or to determine which population of patients can benefit from the immunotherapies.¹⁸¹

Along with the effectiveness of PD-1/PD-L1 blockers, their side effects, including skin toxicity, colitis, pneumonia, liver

toxicity, etc. have emerged.¹⁸² However, it is not clear which cancer patients who use PD-1/PD-L1 blockers have potential side effects. Many tumors have a background of chronic inflammatory diseases, such as chronic hepatitis, pneumonia, and enteritis. In severe cases, stem cells can proliferate significantly to compensate for the loss of cells in the organs, and at the same time, the stem cells will recruit PD1 + lymphocytes to avoid killing themselves. Therefore, while activating the immune system to kill tumor cells, blockers in PD-1/PD-L1 pathway may also lead to the killing of normal stem cells, thus triggering or aggravating the acute inflammatory response.

There are some limitations to this work. First, based on the available literature, we only compared the similarity of partial epigenetic regulation between tumors and normal origin cells in limited systems. Systematic comparative studies of epigenetic profiles between hierarchical descendants of normal stem cells and tumors and their subtypes in different tissues are necessary. Second, although the term of epigenetic abnormality is widely used, it is not clear how many reported differential epigenetic events/epigenetic abnormalities in tumors are stochastic epimutations rather than hereditary epigenetic events from NOCs. Given the dynamic regulation of gene expression by epigenetic mechanisms, epigenomic analysis of single cells of different differentiation stages will provide more accurate evidence for understanding the global patterns of epigenetic regulation in cancers. Third, genetic and epigenetic determinants are likely to interact in creating a TIC's hallmark. It is unclear to what extent they synergistically endow TICs with a malignant phenotype. Finally, in the in vitro system, genetic manipulation can help differentiated cells to regain stemness, which have not been proven in vivo. Whether epimutations can cause dedifferentiation in vivo is unknown.

In a given system, the cellular origin of TIC can not only determine the heterogeneity between tumors or tumor subtypes, but also determine the heterogeneity of the parenchyma and stroma within the tumor, which may be dominated by the innate epigenetic status of genes and pathways from NOC through epigenetic memory. Although tumor heterogeneity owing to the genetic or epigenetic determinants raises diagnosis and treatment challenges, tumor heterogeneity is not always a random and unpredictable phenomenon. An in-depth understanding of the global patterns of both genetic and epigenetic aspects of tumors will enable us to accurately depict the essential characteristics of tumors, which is expected to open a new path for tumor classification, prognosis assessment, individualized treatment, and new target discovery in the future.

Acknowledgments

We would like to thank Professor Gaosheng Huang (Department of Pathology, Fourth Military Medical University) and Chunguang Li (Department of Pathology, the 251st Hospital of the PLA). Their profound insights and spirit of exploration in medicine have always influenced us. We also thank the editors and three anonymous reviewers for their insightful and constructive comments.

Author Contributions

J.L. Feng contributed the central idea, analyzed most of the data, and wrote the initial draft of the paper. D.W. Zhao and F.D. Lv contributed to refining the ideas, carrying out additional analyses. Z.Y. Yuan contributed to drawing the figures. All authors discussed the results and revised the manuscript.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by the Scientific Research Project of Beijing You-an Hospital, CCMU (No. YNKTTS20180110).

Ethical Statement

This is a review work that does not involve ethical approval and consent to participate. All authors approved the manuscript and publication.

ORCID iD

Jiliang Feng b https://orcid.org/0000-0002-2027-9737

References

- Flavahan WA, Gaskell E, Bernstein BE. Epigenetic plasticity and the hallmarks of cancer. *Science*. 2017;357(6348):eaal2380.
- Banno K, Kisu I, Yanokura M, et al. Epimutation and cancer: A new carcinogenic mechanism of Lynch syndrome. *Int J Oncol.* 2012;41:793-797.
- McCarrey JR, Lehle JD, Raju SS, Wang Y, Nilsson EE, Skinner MK. Tertiary epimutations - a novel aspect of epigenetic transgenerational inheritance promoting genome instability. *PLoS One.* 2016;11:e0168038.
- Takeshima H, Ushijima T. Accumulation of genetic and epigenetic alterations in normal cells and cancer risk. *NPJ precis oncol.* 2019;3(3):7.
- Singh SK, Hawkins C, Clarke ID, et al. Identification of human brain tumour initiating cells. *Nature*. 2004;432(7015):396-401.
- Krivtsov AV, Figueroa ME, Sinha AU, et al. Cell of origin determines clinically relevant subtypes of MLL-rearranged AML. *Leukemia*. 2013;27(4):852-860.
- Peinert S, Seymour JF. Indolent lymphomas other than follicular and marginal zone lymphomas8 Kurtin PJ. Indolent lymphomas of mature B lymphocytes. *Hematol Oncol Clin North Am*. 2008; 22(54):903769-994090.
- 8. Kurtin PJ. Indolent lymphomas of mature B lymphocytes. Hematol Oncol Clin North Am. 2009;23(4):769-790.

- Feng J, Zhu R, Chang C, et al. CK19 and Glypican 3 expression profiling in the prognostic indication for patients with HCC after surgical resection. *PLoS One.* 2016;11(3):e0151501.
- Poudel P, Nyamundanda G, Patil Y, Cheang MCU, Sadanandam A. Heterocellular gene signatures reveal luminal-A breast cancer heterogeneity and differential therapeutic responses. *NPJ Breast Cancer*. 2019;5:21.
- Bormann F, Rodríguez-Paredes M, Lasitschka F, et al. Cell-oforigin DNA methylation signatures are maintained during colorectal carcinogenesis. *Cell Rep.* 2018;23(11):3407-3418.
- Taurin S, Alkhalifa H. Breast cancers, mammary stem cells, and cancer stem cells, characteristics, and hypotheses. *Neoplasia*. 2020;22(12):663-678.
- Zhao Y, Feng F, Zhou YN. Stem cells in gastric cancer. World J Gastroenterol. 2015;21(1):112-123.
- Hou SX, Singh SR. Stem-cell-based tumorigenesis in adult Drosophila. *Curr Top Dev Biol.* 2017;121:311-337.
- Vendramin R, Litchfield K, Swanton C. Cancer evolution: Darwin and beyond. *EMBO J.* 2021;40(18):e108389.
- Kim J. Cellular Reprogramming to Model and Study Epigenetic Alterations in Cancer. *Stem Cell Res.* 2020;49:102062.
- Wong M, Mayoh C, Lau LMS, et al. Whole genome, transcriptome and methylome profiling enhances actionable target discovery in high-risk pediatric cancer. *Nat Med.* 2020;26(11):1742-1753.
- Futreal PA, Coin L, Marshall M, et al. A census of human cancer genes. *Nat Rev Cancer*. 2004;4(3):177-183.
- Davenport KP, Blanco FC, Sandler AD. Pediatric malignancies. Surg Clin. 2012;92(3):745-767. x.
- Renardy M, Jilkine A, Shahriyari L, Chou CS. Control of cell fraction and population recovery during tissue regeneration in stem cell lineages. *J Theor Biol.* 2018;445:33-50.
- Velten L, Haas SF, Raffel S, et al. Human haematopoietic stem cell lineage commitment is a continuous process. *Nat Cell Biol.* 2017;19:271-281.
- 22. Chen F, Fine A. Stem cells in lung injury and repair. *Am J Pathol.* 2016;186:2544-2550.
- Chen F, Jimenez RJ, Sharma K, et al. Broad Distribution of Hepatocyte Proliferation in Liver Homeostasis and Regeneration. *Cell Stem Cell* 2020;26(1):27-33.e4.
- Venkei ZG, Yamashita YM. Emerging mechanisms of asymmetric stem cell division. J Cell Biol. 2018;217(11):3785-3795.
- Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature*. 2001;414(6859):105-111.
- 26. Klein G. Cancer, apoptosis, and nonimmune surveillance. *Cell Death Differ*. 2004;11(1):13-17.
- 27. Chen J. The cell-cycle arrest and apoptotic functions of p53 in tumor initiation and progression. *Cold Spring Harb Perspect Med.* 2016;6(3):a026104.
- Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. Nat Rev Cancer. 2005;5(4):275-284.
- Flores-Téllez, Villa-Trevio S, Pia-Vázquez C. Road to stemness in hepatocellular carcinoma. World J Gastroenterol. 2017; 23(37):6750-6776.
- Altucci L, Stunnenberg HG, Altucci L, et al. Time for epigenetics. *Int J Biochem Cell Biol*. 2009;41(1):2-3.

- Vilkin A, Niv Y. Association between hMLH1 hypermethylation and JC virus (JCV) infection in human colorectal cancer (CRC). *Clin Epigenet*. 2010;2(1):1-5.
- Martincorena I, Raine KM, Gerstung M, et al. Universal patterns of selection in cancer and somatic tissues. *Cell*. 2017;171(5): 1029-1041.
- Park JW, Lee JK, Sheu KM, et al. Reprogramming normal human epithelial tissues to a common, lethal neuroendocrine cancer lineage. *Science*. 2018;362:91-95.
- Martínez-Jiménez F, Muiños F, Sentís I, et al. A compendium of mutational cancer driver genes. *Nat Rev Cancer*. 2020;20(10): 555-572.
- 35. Vittoria P, Luca F, Alessio Z. Tumorigenic cell reprogramming and cancer plasticity: interplay between signaling, microenvironment, and epigenetics. *Stem Cell Int.* 2018;2018:1-16.
- Toh TB, Lim JJ, Chow EK. Epigenetics of hepatocellular carcinoma. *Clin Transl Med.* 2019;8:13.
- Dolci M, Favero C, Toumi W, et al. Human endogenous retroviruses long terminal repeat methylation, transcription, and protein expression in human colon cancer. *Front Oncol.* 2020; 10:569015.
- Moss J, Magenheim J, Neiman D, et al. Comprehensive human cell-type methylation atlas reveals origins of circulating cell-free DNA in health and disease. *Nat Commun.* 2018;9:5068.
- 39. Chirshev E, Hojo N, Bertucci A, et al. Epithelial/mesenchymal heterogeneity of high-grade serous ovarian carcinoma samples correlates with miRNA let-7 levels and predicts tumor growth and metastasis. *Mol Oncol.* 2020;14(11):2796-2813.
- Rossi F, Noren H, Jove R, Beljanski V, Grinnemo KH. Differences and similarities between cancer and somatic stem cells: Therapeutic implications. *Stem Cell Res Ther.* 2020;11(1):489.
- 41. Piña-Sánchez P, Chávez-González A, Ruiz-Tachiquín M, et al. Cancer biology, epidemiology, and treatment in the 21st century: Current status and future challenges from a biomedical perspective. *Cancer Control.* 2021;28.
- 42. Schmidt CS, Bultmann S, Meilinger D, et al. Global DNA hypomethylation prevents consolidation of differentiation programs and allows reversion to the embryonic stem cell state. *PLoS One* 2012;7(12):e52629.
- Nishino K, Toyoda M, Yamazaki-Inoue M, et al. DNA methylation dynamics in human induced pluripotent stem cells over time. *PLoS Genet*. 2011;7:e1002085.
- 44. Ehrlich M. DNA hypomethylation in cancer cells. *Epigenomics*. 2009;1(2):239-259.
- 45. Hoffman RM. Is DNA methylation the new guardian of the genome? *Mol Cytogenet*. 2017;10:11.
- Stefansson OA, Moran S, Gomez A, et al. A DNA methylationbased definition of biologically distinct breast cancer subtypes. *Mol Oncol.* 2015;9:555-568.
- 47. Shen J, Wang S, Zhang YJ, Kappil M, Wu HC, Kibriya MG, et al. Genome-wide DNA methylation profiles in hepatocellular carcinoma. *Hepatology*. 2012;55:1799-1808.
- Wang XQ, Ng RK, Ming X, et al. Epigenetic regulation of pluripotent genes mediates stem cell features in human hepatocellular carcinoma and cancer cell lines. *PLoS One.* 2013;8:e72435.

- 49. Rumbajan JM, Maeda T, Souzaki R, et al. Comprehensive analyses of imprinted differentially methylated regions reveal epigenetic and genetic characteristics in hepatoblastoma. *BMC Cancer*. 2013;13:608-619.
- 50. Keelawat S, Thorner PS, Shuangshoti S, et al. Detection of global hypermethylation in well-differentiated thyroid neoplasms by immunohistochemical (5-methylcytidine) analysis. J Endocrinol Invest. 2015;38:725-732.
- Kulis M, Merkel A, Heath S, et al. Whole-genome fingerprint of the DNA methylome during human B cell differentiation. *Nat Genet.* 2015;47:746-756.
- Hadjimichael C, Chanoumidou K, Papadopoulou N, Arampatzi P, Papamatheakis J, Kretsovali A. Common stemness regulators of embryonic and cancer stem cells. *World J Stem Cells*. 2015;7: 1150-1184.
- Berry NB, Bapat SA. Ovarian cancer plasticity and epigenomics in the acquisition of a stem-like phenotype. J Ovarian Res. 2008;1:8.
- Easwaran H, Johnstone SE, Van Neste L, et al. A DNA hypermethylation module for the stem/progenitor cell signature of cancer. *Genome Res.* 2012;22:837-849.
- 55. Smith ZD, Shi J, Gu H, et al. Epigenetic restriction of extraembryonic lineages mirrors the somatic transition to cancer. *Nature*. 2017;549:543-547.
- LandanCohen GNM, Cohen NM, Mukamel Z, et al. Epigenetic polymorphism and the stochastic formation of differentially methylated regions in normal and cancerous tissues. *Nat Genet*. 2012;44:1207-1214.
- Park YJ, Claus R, Weichenhan D, Plass C. Genome-wide epigenetic modifications in cancer. *Prog Drug Res.* 2011;67: 25-49.
- Sipos F, Mũzes G, Patai ÁV, et al. Genome-wide screening for understanding the role of DNA methylation in colorectal cancer. *Epigenomics*. 2013;5(5):569-581.
- Carvalho RH, Haberle V, Hou J, et al. Genome-wide DNA methylation profiling of non-small cell lung carcinomas. *Epi*genetics Chromatin. 2012;5(1):9.
- 60. Shen J, Wang S, Siegel AB, et al. Genome-wide expression of MicroRNAs is regulated by DNA methylation in hepatocarcinogenesis. *Gastroenterol Res pract*. 2015;2015:230642.
- 61. Fujiwara K, Ghosh S, Liang P, Morien E, Soma M, Nagase H. Genome-wide screening of aberrant DNA methylation which associated with gene expression in mouse skin cancers. *Mol Carcinog.* 2015;54(3):178-188.
- 62. Hoadley KA, Yau C, Hinoue T, et al. Cell-of-origin patterns dominate the molecular classification of 10,000 tumors from 33 types of cancer. *Cell*. 2018;173:291-e6.
- 63. Forn M, Díez-Villanueva A, Merlos-Suárez A, et al. Overlapping DNA methylation dynamics in mouse intestinal cell differentiation and early stages of malignant progression. *PLoS One*. 2015;10:e0123263.
- 64. Maschietto M, Rodrigues TC, Kashiwabara AY, et al. DNA methylation landscape of hepatoblastomas reveals arrest at early stages of liver differentiation and cancer-related alterations. *Oncotarget*. 2017;8:97871-97889.

- 65. Aran D, Abu-Remaileh M, Levy R, et al. Embryonic stem cell (ES)specific enhancers specify the expression potential of ES genes in cancer. In: Mitchell JA, ed, *PLoS Genet.* 12; 2016:e1005840.
- 66. Long MD, Dhiman VK, Affronti HC, Hu Q, Liu S, Smiraglia DJ. Dynamic patterns of DNA methylation in the normal prostate epithelial differentiation program are targets of aberrant methylation in prostate cancer. *Sci Rep.* 2021;11:11405.
- Di Domenico A, Pipinikas CP, Maire RS, et al. Epigenetic landscape of pancreatic neuroendocrine tumours reveals distinct cells of origin and means of tumour progression. *Commun Biol.* 2020;3(1):740.
- Shen H, Shih J, Hollern DP, et al. Integrated molecular characterization of testicular germ cell tumors. *Cell Rep.* 2018; 23(11):3392-3406.
- 69. Smith BA, Balanis NG, Nanjundiah A, et al. A human adult stem cell signature marks aggressive variants across epithelial cancers. *Cell Rep.* 2018;24(12):3353-3366. e5.
- Giacopelli B, Wang M, Cleary A, et al. DNA methylation epitypes highlight underlying developmental and disease pathways in acute myeloid leukemia. *Genome Res.* 2021;31(5):747-761.
- Irizarry RA, Ladd-Acosta C, Wen B, et al. The human colon cancer methylome shows similar hypo- and hypermethylation at conserved tissue-specific CpG island shores. *Nat Genet*. 2009; 41:178-186.
- 72. Doi A, Park IH, Wen B, et al. Differential methylation of tissueand cancer-specific CpG island shores distinguishes human induced pluripotent stem cells, embryonic stem cells and fibroblasts. *Nat Genet*. 2009;41:1350-1353.
- Tang W, Wan S, Yang Z, Teschendorff AE, Zou Q. Tumor origin detection with tissue-specific miRNA and DNA methylation markers. *Bioinformatics*. 2018;34(3):398-406.
- Isagawa T, Nagae G, Shiraki N, et al. DNA methylation profiling of embryonic stem cell differentiation into the three germ layers. *PLoS One*. 2011;6:e26052.
- Kouzarides T. Chromatin modifications and their function. *Cell.* 2007;128:693-705.
- 76. Ohm JE, McGarvey KM, Yu X, et al. A stem cell-like chromatin pattern may predispose tumor suppressor genes to DNA hypermethylation and heritable silencing. *Nat Genet*. 2007;39(2):237-242.
- Bernstein BE, Mikkelsen TS, Xie X, et al. A bivalent chromatin structure marks key developmental genes in embryonic stem cells. *Cell*. 2006;125(2):315-326.
- Huang J, Fan T, Yan Q, et al. Lsh, an epigenetic guardian of repetitive elements. *Nucleic Acids Res.* 2004;32(17):5019-5028.
- Lin B, Lee H, Yoon JG, et al. Global analysis of H3K4me3 and H3K27me3 profiles in glioblastoma stem cells and identification of SLC17A7 as a bivalent tumor suppressor gene. *Oncotarget*. 2015;6:5369-5381.
- Chapman-Rothe N, Curry E, Zeller C, et al. Chromatin H3K27me3/H3K4me3 histone marks define gene sets in highgrade serous ovarian cancer that distinguish malignant, tumoursustaining and chemo-resistant ovarian tumour cells. *Oncogene*. 2013;32:4586-4592.
- Zhao JC, Yu J, Runkle C, et al. Cooperation between Polycomb and androgen receptor during oncogenic transformation. *Genome Res.* 2012;22:322-331.

- Jambhekar A, Dhall A, Shi Y. Roles and regulation of histone methylation in animal development. *Nat Rev Mol Cell Biol.* 2019;20(10):625-641.
- Maa B, Dca B. Role of lncRNAs in stem cell maintenance and differentiation- ScienceDirect. *Curr Top Dev Biol.* 2020;138: 73-112.
- 84. Tordonato C, Di Fiore PP, Nicassio F. The role of non-coding RNAs in the regulation of stem cells and progenitors in the normal mammary gland and in breast tumors. *Front Genet.* 2015;6:72.
- 85. Divisato G, Piscitelli S, Elia M, Cascone E, Parisi S. MicroRNAs and Stem-like Properties: The complex regulation underlying stemness maintenance and cancer development. *Biomolecules*. 2021;11(8):1074.
- Hu S, Shan G. LncRNAs in stem cells. *Stem Cells Int.* 2016; 2016:2681925.
- DeSano JT, Xu L. MicroRNA regulation of cancer stem cells and therapeutic implications. *AAPS J.* 2009;11(4):682-692.
- Papagiannakopoulos T, Kosik KS. MicroRNAs: Regulators of oncogenesis and stemness. *BMC Med.* 2008;6:15.
- Mathieu J, Ruohola-Baker H. Regulation of stem cell populations by microRNAs. *Adv Exp Med Biol*. 2013;786:329-351.
- Hernandez-Vargas H, Sincic N, Ouzounova M, Herceg Z. Epigenetic signatures in stem cells and cancer stem cells. *Epigenomics*. 2009;1:261-280.
- Matsui WH. Cancer stem cell signaling pathways. *Medicine*. 2016;95(1 suppl 1):S8-S19.
- Tarayrah L, Chen X. Epigenetic regulation in adult stem cells and cancers. *Cell Biosci.* 2013;3(1):41.
- Herrera SC, Bach EA. JAK/STAT signaling in stem cells and regeneration: From Drosophila to vertebrates. *Development*. 2019;146(2):dev167643.
- 94. Varlakhanova NV, Cotterman RF, Devries WN, et al. Myc maintains embryonic stem cell pluripotency and self-renewal. *Differentiation*. 2010;80(1):9-19.
- 95. Arsic N, Gadea G, Lagerqvist EL, et al. The p53 Isoform Δ133p53β promotes cancer stem cell potential. *Stem Cell Reports*. 2015;4(4):531-540.
- Huang T, Song X, Xu D, et al. Stem cell programs in cancer initiation, progression, and therapy resistance. *Theranostics*. 2020;10(19):8721-8743.
- Zhang J, Benavente CA, McEvoy J, et al. A novel retinoblastoma therapy from genomic and epigenetic analyses. *Nature*. 2012;481:329-334.
- Mansoori B, Mohammadi A, Ditzel HJ, et al. HMGA2 as a critical regulator in cancer development. *Genes.* 2021;12(2):269.
- 99. Keyvani-Ghamsari S, Khorsandi K, Rasul A, Zaman MK. Current understanding of epigenetics mechanism as a novel target in reducing cancer stem cells resistance. *Clin Epigenet*. 2021;13(1):120.
- 100. Chatoo W, Abdouh M, David J, et al. The polycomb group gene Bmi1 regulates antioxidant defenses in neurons by repressing p53 pro-oxidant activity. *J Neurosci.* 2009;29(2):529-542.
- 101. Seidel S, Garvalov BK, Wirta V, et al. A hypoxic niche regulates glioblastoma stem cells through hypoxia inducible factor 2α. *Brain.* 2010;133:983-995.

- Batlle E, Clevers H. Cancer stem cells revisited. *Nat Med.* 2017; 23:1124-1134.
- 103. Picot T, Kesr S, Wu Y, et al. Potential role of OCT4 in leukemogenesis. *Stem Cell Dev.* 2017;26:1637-1647.
- 104. Trosko JE. Review Paper: Cancer stem cells and cancer nonstem cells: From adult stem cells or from reprogramming of differentiated somatic cells. *Vet Pathol.* 2009;46(2):176-193.
- 105. Perales-Puchalt A, Svoronos N, Rutkowski MR, et al. Folliclestimulating hormone receptor is expressed by most ovarian cancer subtypes and is a safe and effective immunotherapeutic target. *Clin Cancer Res.* 2017;23:441-453.
- 106. Wang J, Lu F, Ren Q, et al. Novel histone demethylase LSD1 inhibitors selectively target cancer cells with pluripotent stem cell properties. *Cancer Res.* 2011;71(23):7238-7249.
- 107. Chen HF, Wu KJ. Endothelial transdifferentiation of tumor cells triggered by the Twist1-Jagged1-KLF4 Axis: Relationship between cancer stemness and angiogenesis. *Stem Cells Int.* 2015; 2016:6439864.
- Spangle JM, Roberts TM. Epigenetic regulation of RTK signaling. J Mol Med. 2017;95:791-798.
- 109. Yoshida GJ. Emerging roles of Myc in stem cell biology and novel tumor therapies. *J Exp Clin Cancer Res.* 2018;37:173.
- 110. Koukourakis MI, Kakouratos C, Kalamida D, et al. Hypoxiainducible proteins HIF1α and lactate dehydrogenase LDH5, key markers of anaerobic metabolism, relate with stem cell markers and poor post-radiotherapy outcome in bladder cancer. *Int J Radiat Biol.* 2016;92:353-363.
- 111. Chen YA, Lu CY, Cheng TY, Pan SH, Chen HF, Chang NS. WW domain-containing proteins yap and taz in the hippo pathway as key regulators in stemness maintenance, tissue homeostasis, and tumorigenesis. *Front Oncol.* 2019;9:60.
- 112. Moya IM, Castaldo SA, Van den Mooter L, et al. Peritumoral activation of the Hippo pathway effectors YAP and TAZ suppresses liver cancer in mice. *Science*. 2019;366(6468): 1029-1034.
- 113. Pearson JD, Huang K, Pacal M, et al. Binary pan-cancer classes with distinct vulnerabilities defined by pro- or anti-cancer YAP/ TEAD activity. *Cancer Cell*. 2021;39(8):1115-1134.
- 114. Shelke P, Sarode GS, Sarode SC, Maniyar N, Prajapati G, Patil S. Expression of yes-associated protein in oral squamous cell carcinoma. *J Contemp Dent Pract.* 2019;20(8):887-892.
- 115. Noh MG, Kim SS, Hwang EC, Kwon DD, Choi C. Yesassociated protein expression is correlated to the differentiation of prostate adenocarcinoma. *J Pathol Trans Med.* 2017; 51(4):365-373.
- Zanconato F, Cordenonsi M, Piccolo S. YAP/TAZ at the roots of cancer. *Cancer Cell*. 2016;29(6):783-803.
- 117. Yimlamai D, Christodoulou C, Galli GG, et al. Hippo pathway activity influences liver cell fate. *Cell*. 2014;157(6):1324-1338.
- 118. Moore MAS. Cytokine and chemokine networks influencing stem cell proliferation, differentiation, and marrow homing. J Cell Biochem. 2002;85:29-38.
- 119. Lee N, Barthel SR, Schatton T. Melanoma stem cells and metastasis: Mimicking hematopoietic cell trafficking? *Lab Invest.* 2014;94:13-30.

- 120. Schulenburg A, Blatt K, Cerny-Reiterer S, et al. Cancer stem cells in basic science and in translational oncology: can we translate into clinical application? *J Hematol Oncol.* 2015;8:16.
- 121. Park S, Jang H, Kim BS, Hwang C, Jeong GS, Park Y. Directional migration of mesenchymal stem cells under an SDF-1α gradient on a microfluidic device. In: Leipzig ND, ed, *PLoS One.* 2017;12(9):e0184595.
- 122. Dorantes-Heredia R, Chablé-Montero F, Henson DE, Albores-Saavedra J. Rokitansky-aschoff sinuses mimicking adenocarcinoma of the gallbladder. *Am J Surg Pathol.* 2013;37:1269-1274.
- 123. Galivo FH, Dorrell C, Grompe MT, Zhong Y, Streeter PR, Grompe M. Novel surface markers directed against adult human gallbladder. *Stem Cell Res.* 2015;15(1):172-181.
- 124. Leelawat K, Thongtawee T, Narong S, et al. Strong expression of CD133 is associated with increased cholangiocarcinoma progression. *World J Gastroenterol.* 2011;17:1192-1198.
- 125. Sriraksa R, Zeller C, Dai W, et al. Aberrant DNA methylation at genes associated with a stem cell-like phenotype in cholangiocarcinoma tumors. *Cancer Prev Res.* 2013;6:1348-1355.
- 126. Zarco N, Norton E, Quiñones-Hinojosa A, Guerrero-Cázares H. Overlapping migratory mechanisms between neural progenitor cells and brain tumor stem cells. *Cell Mol Life Sci.* 2019;76(18):3553-3570.
- 127. Soundararajan R, Rao AJ. Trophoblast 'pseudo-tumorigenesis': Significance and contributory factors. *Reprod Biol Endocrinol*. 2004;2:15.
- 128. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell*. 2011;144(5):646-674.
- 129. Radpour R. Tracing and targeting cancer stem cells: New venture for personalized molecular cancer therapy. *World J Stem Cell.* 2017;9:169-178.
- Abbott BL. ABCG2 (BCRP): A cytoprotectant in normal and malignant stem cells. *Clin Adv Hematol Oncol.* 2006;4(1):63-72.
- 131. Lanza RP, Cibelli JB, Blackwell C, et al. Extension of cell lifespan and telomere length in animals cloned from senescent somatic cells. *Science*. 2000;288(5466):665-669.
- 132. Huang X, Ju Z. Mouse Model: Telomeres and telomerase in stem cell and cancer. In: Hiyama K, eds. *Telomeres and Telomerase in Cancer: Cancer Drug Discovery and Development*. Totowa, New Jersey: Humana Press; 2009.
- 133. Fang S, Salven P. Stem cells in tumor angiogenesis. J Mol Cell Cardiol. 2011;50:290-295.
- 134. Rahbarghazi R, Nassiri SM, Khazraiinia P, et al. Juxtacrine and paracrine interactions of rat marrow-derived mesenchymal stem cells, muscle-derived satellite cells, and neonatal cardiomyocytes with endothelial cells in angiogenesis dynamics. *Stem Cells Dev.* 2012;22(6):855-865.
- 135. Bao S, Wu Q, Sathornsumetee S, et al. Stem cell-like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. *Cancer Res.* 2006;66:7843-7848.
- 136. Yao XH, Ping YF, Bian XW. Contribution of cancer stem cells to tumor vasculogenic mimicry. *Protein Cell*. 2011;2:266-272.
- 137. Momparler RL. Cancer epigenetics. Oncogene. 2003;22:6479-6483.
- 138. Need EF, Atashgaran V, Ingman WV, Dasari P. Hormonal regulation of the immune microenvironment in the mammary gland. *J Mammary Gland Biol Neoplasia*. 2014;19:229-239.

- Utermöhlen O, Krönke M. Survival of priceless cells: active and passive protection of embryonic stem cells against immune destruction. *Arch Biochem Biophys.* 2007;462:273-277.
- 140. Lin VTG, Pruitt HC, Samant RS, Shevde LA. Developing cures: Targeting ontogenesis in cancer. *Trends Cancer*. 2017;3(2): 126-136.
- 141. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature*. 2008;454(7203):436-444.
- 142. Briede I, Strumfa I, Vanags A, Gardovskis J. The association between inflammation, epithelial mesenchymal transition and stemness in colorectal carcinoma. *J Inflamm Res.* 2020;13:15-34.
- 143. Asano Y, Kashiwagi S, Goto W, et al. Prediction of treatment responses to neoadjuvant chemotherapy in triple-negative breast cancer by analysis of immune checkpoint protein expression. J Transl Med. 2018;16(1):87.
- 144. Cao Y, Guo WT, Tian S, et al. miR-290/371-Mbd2-Myc circuit regulates glycolytic metabolism to promote pluripotency. *EMBO J.* 2015;34:609-623.
- 145. DeWaal D, Nogueira V, Terry AR, et al. Hexokinase-2 depletion inhibits glycolysis and induces oxidative phosphorylation in hepatocellular carcinoma and sensitizes to metformin. *Nat Commun.* 2018;9:446.
- 146. Riester M, Xu Q, Moreira A, Zheng J, Michor F, Downey RJ. The Warburg effect: Persistence of stem-cell metabolism in cancers as a failure of differentiation. *Ann Oncol.* 2018;29: 264-270.
- 147. Ji Y, Yang C, Tang Z, et al. Adenylate kinase hCINAP determines self-renewal of colorectal cancer stem cells by facilitating LDHA phosphorylation. *Nat Commun.* 2017;8:15308.
- 148. Yuen CA, Asuthkar S, Guda MR, Tsung AJ, Velpula KK. Cancer stem cell molecular reprogramming of the Warburg effect in glioblastomas: A new target gleaned from an old concept. *CNS Oncol.* 2016;5:101-108.
- Thapar R, Titus M. Recent advances in metabolic profiling and imaging of prostate cancer. *Curr Metabolomics*. 2014;2:53-69.
- 150. Kim S, Kim DH, Jung W-H, Koo JS. Metabolic phenotypes in triple-negative breast cancer. *Tumor Biol.* 2013;34:1699-1712.
- 151. Kelly GM, Gatie MI. Mechanisms regulating stemness and differentiation in embryonal carcinoma cells. *Stem Cells Internat.* 2017;2017:3684178.
- 152. Jeong Y, Mangelsdorf DJ. Nuclear receptor regulation of stemness and stem cell differentiation. *Exp Mol Med.* 2009;41: 525-537.
- 153. Yang ZF, Ho DW, Ng MN, et al. Significance of CD90+ cancer stem cells in human liver cancer. *Cancer Cell*. 2008;13(2): 153-166.
- 154. Nurwidya F, Murakami A, Takahashi F, Takahashi K. Lung cancer stem cells: Tumor biology and clinical implications. *Asia Pac J Clin Oncol.* 2012;8:217-222.
- 155. Stewart-Morgan KR, Petryk N, Groth A. Chromatin replication and epigenetic cell memory. *Nat Cell Biol.* 2020;22(4):361-371.
- 156. Reverón-Gómez N, González-Aguilera C, et al. Accurate recycling of parental histones reproduces the histone modification landscape during DNA replication. *Mol Cell.* 2018;1872(2): 239-249. e5.

- 157. Dagogo-Jack I, Shaw AT. Tumour heterogeneity and resistance to cancer therapies. *Nat Rev Clin Oncol.* 2018;15(2):81-94.
- 158. Shakiba N, Fahmy A, Jayakumaran G, et al. Cell competition during reprogramming gives rise to dominant clones. *Science*. 2019;364(6438):eaan0925.
- Vitale I, Shema E, Loi S, Galluzzi L. Intratumoral heterogeneity in cancer progression and response to immunotherapy. *Nat Med.* 2021;27:212-224.
- 160. Park TS, Donnenberg VS, Donnenberg AD, Zambidis ET, Zimmerlin L. Dynamic interactions between cancer stem cells and their stromal partners. *Curr Pathobiol Rep.* 2014;2(1): 41-52.
- 161. Mesa KR, Rompolas P, Greco V. The Dynamic Duo: Niche/Stem Cell Interdependency. *Stem Cell Reports*. 2015;4(6):961-966.
- 162. Lacina L, Plzak J, Kodet O, et al. Cancer microenvironment: What can we learn from the stem cell niche. *Int J Mol Sci.* 2015; 16(10):24094-24110.
- 163. Bharti R, Dey G, Mandal M. Cancer development, chemoresistance, epithelial to mesenchymal transition and stem cells: A snapshot of IL-6 mediated involvement. *Cancer Lett.* 2016; 375(1):51-61.
- 164. Murgai M, Giles A, Kaplan R. Physiological, tumor, and metastatic niches: opportunities and challenges for targeting the tumor microenvironment. *Crit Rev Oncog.* 2015;20:301-314.
- 165. Nagtegaal ID, Odze RD, Klimstra D, Paradis V, Rugge M, Schirmacher P, et al. WHO Classification of Tumours of the Digestive System. 5th ed. Lyon: International Agency for Research on Cancer; 2019:216-217.
- 166. Taguchi J, Shibata H, Kabata M, et al. DMRT1-mediated reprogramming drives development of cancer resembling human germ cell tumors with features of totipotency. *Nat Commun.* 2021;12(1):5041.
- 167. Poli V, Fagnocchi L, Fasciani A, et al. MYC-driven epigenetic reprogramming favors the onset of tumorigenesis by inducing a stem cell-like state. *Nat Commun.* 2018;9(1):1024.
- 168. Heuberger J, Grinat J, Kosel F, et al. High Yap and Mll1 promote a persistent regenerative cell state induced by Notch signaling and loss of p53. *Proc Natl Acad Sci Unit States Am.* 2021; 118(22):e2019699118.
- 169. Rong Z, Wang M, Hu Z, et al. An effective approach to prevent immune rejection of human ESC-derived allografts. *Cell Stem Cell*. 2014;14(1):121-130.

Nomenclature

Abbreviations

- TIC tumor-initiating cell
- NOC normal origin cell
- TSC tumor stem cell

- 170. Dempke WCM, Fenchel K, Uciechowski P, Chevassut T. Targeting developmental pathways: the achilles heel of cancer? Oncology. 2017;93(4):213-223.
- 171. Castagnoli L, Cancila V, Cordoba-Romero SL, et al. WNT signaling modulates PD-L1 expression in the stem cell compartment of triple-negative breast cancer. *Oncogene*. 2019;38: 4047-4060.
- 172. Wang C, Li Y, Jia L, et al. CD276 expression enables squamous cell carcinoma stem cells to evade immune surveillance. *Cell Stem Cell*. 2021;28(9):1597-1613. e7.
- 173. El-Khoueiry AB, Sangro B, Yau T, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): An open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet*. 2017;389(10088):2492-2502.
- 174. Tang DG. Understanding cancer stem cell heterogeneity and plasticity. *Cell Res.* 2012;22:457-472.
- 175. Nordström L, Andersson E, Kuci V, et al. DNA methylation and histone modifications regulate SOX11 expression in lymphoid and solid cancer cells. *BMC Cancer*. 2015;15:273.
- 176. Toh TB, Lim JJ, Chow EKH. Epigenetics in cancer stem cells. Mol Cancer. 2017;16:29.
- 177. Topper MJ, Vaz M, Chiappinelli KB, et al. Epigenetic therapy ties myc depletion to reversing immune evasion and treating lung cancer. *Cell*. 2017;171:1284-1300.
- 178. Tsai H-C, Li H, Van Neste L, et al. Transient low doses of DNAdemethylating agents exert durable antitumor effects on hematological and epithelial tumor cells. *Cancer Cell*. 2012;21:430-446.
- 179. Duraiswamy J, Kaluza KM, Freeman GJ, Coukos G. Dual blockade of PD-1 and CTLA-4 combined with tumor vaccine effectively restores T-cell rejection function in tumors. *Cancer Res.* 2013;73(12):3591-3603.
- 180. Hombach AA, Görgens A, Chmielewski M, et al. Superior therapeutic index in lymphoma therapy: CD30 + CD34 + hematopoietic stem cells resist a chimeric antigen receptor T-cell attack. *Mol Ther.* 2016;24(8):1423-1434.
- 181. Sharma P, Allison JP. Dissecting the mechanisms of immune checkpoint therapy. *Nat Rev Immunol.* 2020;20:75-76.
- 182. Londoño MC, Reig M, RETOINMUNO Multidisciplinary Group. Multidisciplinary clinical approach to cancer patients with immune-related adverse events induced by checkpoint inhibitors. *Cancers*. 2020;12(11):3446.
- HCC hepatocellular carcinoma
- DNA deoxyribonucleic acid
- CpG cytosine-phosphor-guanine
- CD cluster of differentiation
- PD-1 programmed cell death-1
- PD-L programmed cell death-ligand 1