



Back to the Origin: $bla_{OXA-204}$ and bla_{NDM-1} Genes in *Shewanella* spp. from a Tunisian River

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he environmental bacteria Xanthomonas spp. and Shewanella spp. are progenitors of the bla_{NDM} and $bla_{OXA-48-like}$ carbapenemase genes, respectively (1, 2). The most likely hypothesis is that these genes have been mobilized in the environment on mobile genetic elements, and subsequently transferred to Gram-negative bacteria (GNB) of clinical importance. The pathogens (primarily Escherichia coli and Klebsiella pneumoniae) that acquired these carbapenemase genes then emerged as a major One Health issue and are currently reported not only in clinical settings but also in animals and the environment (3). In January 2019, two carbapenem-resistant (CP-R) Shewanella spp. were isolated from fresh water sampled at two different locations along the El Hammam Oued river in Sousse, Tunisia, as part of a pilot study on multidrug-resistant GNB in aquatic environments. This river collects urban treated wastewater but no hospital effluents. The two isolates resistant to ertapenem (MIC ≥32 mg/L) were whole-genome sequenced using short-read (NovaSeq-6000, Illumina) and long-read (MinION, Oxford Nanopore) technologies (all genomic seguences were deposited in DDBJ/EMBL/GenBank under the accession number PRJNA830355). Genomes were assembled with Unicycler and analyzed using tools of the Center for Genomic Epidemiology (https://www.genomicepidemiology.org/).

S. xiamenensis 8A possessed a 4,959,567 bp chromosome and one nontypeable 216,160 bp plasmid (p8A) showing poor or fragmented homologies with published sequences. Except for $bla_{OXA-538}$, all resistance genes—conferring resistances to beta-lactams, aminoglycosides, sulfonamides, trimethoprim, rifampicin and erythromycin—were located on p8A (Fig. 1A). Specifically, p8A carried the bla_{PER-1} extended-spectrum beta-lactamase (ESBL) and the $bla_{OXA-204}$ carbapenemase genes. $bla_{OXA-204}$ has already been reported in one *S. xiamenensis* strain from water in Portugal (4), flanked by the peptidase C15 and lysR gene as intrinsically found on the *Shewanella* spp. chromosome (5). Here, the $bla_{OXA-204}$ gene was surrounded by an IS*Ecp1* insertion sequence and a truncated lysR gene, as reported in the *K. pneumoniae* KP49 isolated from a Tunisian patient (KP027886) (6). This indicates that $bla_{OXA-204}$ in p8A is not intrinsic but has transited through another bacterial host before being reintegrated in *S. xiamenensis* p8A via a transposition event.

Shewanella bicestrii 10B possessed a 4,717,299 bp chromosome carrying a bla_{OXA-48-like} and the aadA2 resistance genes. The strain additionally presented a nontypeable 160,879 bp plasmid (p10B) sharing conserved regions with the pSA70-3 and pKC-Na-R1 plasmids, respectively, isolated from Shewanella putrefasciens and Shewanella algae (Fig. 1B) (7). p10B presented genes conferring resistance to beta-lactams, aminoglycosides, sulfonamides, trimethoprim, chloramphenicol, quinolones, and rifampicin, but also to mercury and biocides. CP-R was conferred by the bla_{NDM-1} gene; bla_{NDM-1}, together with the bla_{OXA-10}, sul1, and aadA1 genes, was surrounded by two IS91-like that might have mediated its insertion/excision (8). A similar genetic organization was observed in pSA70-3 with only the left

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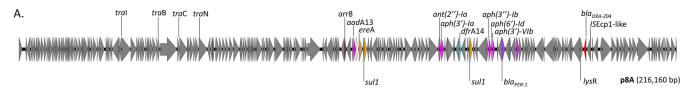
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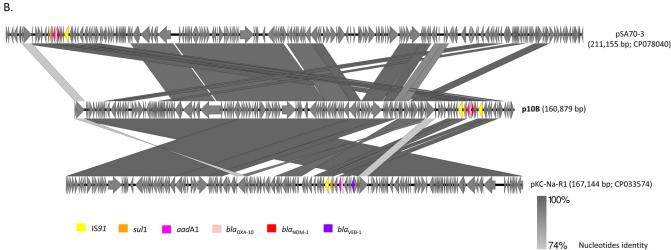


FIG 1 Schematic representation of the p8A plasmid (A). Comparative schematic representation of the p10B, pSA70-3 (CP078040), and pKC-Na-R1 (CP033574) plasmids. Genetic elements were analyzed by BLASTn and visualized using Easyfig 2.2.5_win. The window size used for BLASTn analysis was 2,000 bp.

IS91-like missing (7), while the element was more fragmented in pKC-Na-R1, where only the sul1-IS91-like and aadA1- bla_{OXA-10} genes were conserved upstream the bla_{VEB-1} ESBL gene.

Environmental bacteria are progenitors of carbapenemases (1, 2), but reacquisition of these genes by environmental bacteria is rarely reported. This study showed that the $bla_{OXA-204}$ and bla_{NDM-1} genes, which are highly prevalent in Tunisian hospital settings and effluents (9, 10), can spread back to environmental bacteria. In both cases, resistance genes moved to a *Shewanella* spp. nontypeable plasmid after IS-mediated acquisition, a mechanism that might be more efficient than plasmid transfer for bacterial genera that are not closely related.

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