

Diagnostic accuracy of ultrasound superb microvascular imaging for lymph nodes A protocol for systematic review and meta-analysis

Xuejiao Li, MM, Cong Wang, MM, Xiukun Hou, MM st , Ye Tao, MM st

Abstract

Background: As a novel ultrasound technique, superb microvascular imaging can quickly, simply and noninvasively study the microvascular distribution in the tumor and evaluate the microvascular perfusion. Studies suggested that superb microvascular imaging is helpful for the differentiation between benign and malignant lymph nodes. However, the results of these studies have been contradictory. Therefore, the present meta-analysis aimed at determining the accuracy of superb microvascular imaging in the differential diagnosis between benign and malignant lymph nodes.

Methods: We will search PubMed, Web of Science, Cochrane Library, and Chinese biomedical databases from their inceptions to the July 30, 2020, without language restrictions. Two authors will independently carry out searching literature records, scanning titles and abstracts, full texts, collecting data, and assessing risk of bias. Review Manager 5.2 and Stata14.0 software will be used for data analysis.

Results: This systematic review will determine the accuracy of superb microvascular imaging in the differential diagnosis between benign and malignant lymph nodes.

Conclusion: Its findings will provide helpful evidence for the accuracy of superb microvascular imaging in the differential diagnosis between benign and malignant lymph nodes.

Systematic review registration: INPLASY202070133.

Abbreviations: OR = odds ratio, SMI = superb microvascular imaging.

Keywords: lymph node, meta-analysis, superb microvascular imaging

1. Introduction

Lymph nodes belong to important peripheral immune organ, which participates in the immune response of the body.^[1] Lymphadenopathy is very common in clinic.^[2] The etiology mainly includes local or systemic inflammatory reaction, metastasis of malignant tumor to lymph node, lymphoma, and

XL and CW contributed equally to this work and should be considered co-first authors.

This study is supported by Liaoning Natural Science Foundation Project (20170540256).

The authors report no conflicts of interest.

All data generated or analyzed during this study are included in this published article [and its supplementary information files].

Ultrasound department of the First Affiliated Hospital of Dalian Medical University, China.

^{*} Correspondence: Xiukun Hou, Ye Tao, Ultrasound department of the First Affiliated Hospital of Dalian Medical University, No. 222 Zhongshan Road, Xigang District, Dalian City, Liaoning Province 116011, China (e-mails: 365213427@qq.com, fytgzy@163.com).

Copyright © 2020 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Li X, Wang C, Hou X, Tao Y. Diagnostic accuracy of ultrasound superb microvascular imaging for lymph nodes: A protocol for systematic review and meta-analysis. Medicine 2020;99:36(e22034).

Received: 3 August 2020 / Accepted: 4 August 2020

http://dx.doi.org/10.1097/MD.000000000022034

so on.^[3] High frequency ultrasound can not only show the size, shape, and internal echo of lymph nodes, but also show the blood flow distribution.^[4] As a novel ultrasound technique, superb microvascular imaging can quickly, simply and noninvasively study the microvascular distribution in the tumor and evaluate the microvascular perfusion.^[5] The SMI adopts a multidimensional filter to eliminate only the clutter and to preserve lowvelocity flow signals, whereas conventional Doppler systems use a single-dimension filter and, accordingly, can exhibit a loss of low-velocity flow signals that overlap with clutter.^[6] Studies suggested that superb microvascular imaging is helpful for the differentiation between benign and malignant lymph nodes.^[7-10] However, the results of these studies have been contradictory. Therefore, the present meta-analysis aimed at determining the accuracy of superb microvascular imaging in the differential diagnosis between benign and malignant lymph nodes.

2. Materials and methods

This study was conducted in accordance with the PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) guidelines and the protocol was registered in the INPLASY (INPLASY202070133).

2.1. Eligibility criteria

2.1.1. Type of study. This study will only include high quality clinical cohort or case control studies.

2.1.2. *Type of patients.* The patients should be those who had undergone Lymphadenopathy.

Number	Search terms
1	Lymph Node or Node, Lymph or Nodes, Lymph or Lymphadenopathy
2	Pathology
3 4	superb microvascular imaging or superb micro-vascular imaging or SM and 1-3

2.1.3. Intervention and comparison. This study compare SMI with pathology for diagnosing lymph nodes.

2.1.4. Type of outcomes. The primary outcomes include sensitivity, specificity, positive and negative likelihood ratio, diagnostic odds ratio, and the area under the curve of the summary receiver operating characteristic.

2.2. Search methods

PubMed, Web of Science, Cochrane Library, and Chinese biomedical databases will be searched from their inceptions to the July 31, 2020, without language restrictions. The search strategy for PubMed is shown in Table 1. Other online databases will be used in the same strategy.

2.3. Data extraction and quality assessment

Two authors will independently select the trials according to the inclusion criteria, and import into Endnote X9. Then remove duplicated or ineligible studies. Screen the titles, abstracts, and full texts of all literature to identify eligible studies. All essential data will be extracted using previously created data collection sheet by 2 independent authors. Discrepancies in data collection between 2 authors will be settled down through discussion with the help of another author. The following data will be extracted from each included research: the first authors surname, publication year, language of publication, study design, sample size, number of lesions, source of the subjects, instrument, "gold standard," and diagnostic accuracy. The true positives, true negatives, false positives, and false negatives in the fourfold (2 \times 2) tables were also collected. Methodological quality was independently assessed by 2 researchers based on the quality assessment of studies of diagnostic accuracy studies (QUADAS) tool.^[11] The QUADAS criteria included 14 assessment items. Each of these items was scored as "yes" (2), "no" (0), or "unclear"(1). The QUADAS score ranged from 0 to 28, and a score \geq 22 indicated good quality. Any disagreements between 2 investigators will be solved through discussion or consultation by a 3rd investigator.

2.4. Statistical analysis

The STATA version 14.0 (Stata Corp, College Station, TX, USA) and Meta-Disc version 1.4 (Universidad Complutense, Madrid, Spain) softwares were used for meta-analysis. We calculated the pooled summary statistics for sensitivity, specificity, positive and negative likelihood ratio, and diagnostic odds ratio with their 95% confidence intervals. The summary receiver operating characteristic curve and corresponding area under the curve were obtained. The threshold effect was assessed using Spearman correlation coefficients. The Cochrans Q-statistic and *I* test were used to evaluate potential heterogeneity between studies. If

significant heterogeneity was detected (Q test P < .05 or I test > 50%), a random effects model or fixed effects model was used. We also performed sub group and meta-regression analyses to investigate potential sources of heterogeneity. To evaluate the influence of single studies on the overall estimate, a sensitivity analysis was performed. We conducted Beggs funnel plots and Eggers linear regression tests to investigate publication bias.

2.5. Ethics and dissemination

We will not obtain ethic documents because this study will be conducted based on the data of published literature. We expect to publish this study on a peer-reviewed journal.

3. Discussion

The vascular morphology and distribution of lymph nodes are closely related to their nature, which can be used as an important supplementary sign in their differential diagnosis.^[12,13] SMI can clearly and completely show the shape and distribution of vascular network of lymph node without injection of contrast agent.^[14] Studies suggested that superb microvascular imaging is helpful for the differentiation between benign and malignant lymph nodes. However, the results of these studies have been contradictory. To gain clarity, in this study, we will perform a systematic review to summarize high-quality studies and to provide evidence on the evidence-based medical support for clinical practice.

Author contributions

Conceptualization: Ye Tao and Xiukun Hou.

- Data curation: Cong Wang and Xuejiao Li.
- Methodology: Cong Wang and Xuejiao Li.
- Writing original draft: Cong Wang and Xuejiao Li.
- Writing review & editing: Cong Wang, Xuejiao Li, and Xiukun Hou.

References

- [1] Eleonora Melzi, Mara S, Rocchi , et al. Immunophenotyping of sheep paraffin-embedded peripheral lymph nodes. Front Immunol 2018; 9:2892.
- [2] Quentin Hurlot, Judith Fillaux, Camille Laurent, et al. A case report of isolated lymphadenopathy revealing localized leishmanial lymphadenopathy in an asthenic 25-year-old man. Medicine (Baltimore) 2016;95: e3932.
- [3] Stephan Lang, Benjamin , Kansy . Cervical lymph node diseases in children. GMS Curr Top Otorhinolaryngol Head Neck Surg 2014;13: Doc08.
- [4] Kim-Cuong T, Nguyen , Camila Pachêco-Pereira, et al. Comparison of ultrasound imaging and cone-beam computed tomography for examination of the alveolar bone level: a systematic review. PLoS One 2018;13: e0200596.
- [5] Hata T, Koyanagi A, Yamanishi T, et al. Superb microvascular imaging with Doppler luminance using an 18-MHz probe to visualize fetal intraabdominal blood vessels and organ microvasculature. J Perinat Med 2020;48:184–8.
- [6] Jiang ZZ, Huang YH, Shen HL, et al. Clinical applications of superb microvascular imaging in the liver, breast, thyroid, skeletal muscle, and carotid plaques. J Ultrasound Med 2019;38:2811–20.
- [7] Zuhal Bayramoglu, Emine Caliskan, Zeynep Karakas, et al. Diagnostic performances of superb microvascular imaging, shear wave elastography and shape index in pediatric lymph nodes categorization: a comparative study. Br J Radiol 2018;91:20180129.
- [8] Liu Lijuan, Xu Xiaohong, Deng Dongli, et al. Application of Superb Microvascular Imaging in the identification of cervical lymph nodes. J Guangdong Med Univ 2020;38:94–6.

- [9] Lei yongbiao, Jiang Haiyu, Tang hailing, et al. Application of Superb Microvascular Imaging in differential diagnosis of cervical lymph node lesions. Chin Med Imaging Technol 2016;32:655–8.
- [10] Yang Junwei, Liu Zhimin, Ouyang Lichang, et al. Analysis of clinical application value of Superb Microvascular Imaging in the diagnosis of cervical lymph node lesions. Modern Med Imaging 2018;27:2614–6.
- [11] Whiting PF, Weswood ME, Rutjes AW, et al. Evaluation of quadas, a tool for the quality assessment of diagnostic accuracy studies. BMC Med Res Methodol 2006;6:9.
- [12] Jerome W, Breslin , Ying Yang, et al. Lymphatic vessel network structure and physiology. Compr Physiol 2018;9:207–99.
- [13] Mohammad Jafarnejad, Matthew C, Woodruff, et al. Modeling lymph flow and fluid exchange with blood vessels in lymph nodes. Lymphat Res Biol 2015;13:234–47.
- [14] Ryoo I, Suh S, You SH, et al. Usefulness of microvascular ultrasonography in differentiating metastatic lymphadenopathy from tuberculous lymphadenitis. Ultrasound Med Biol 2016;42: 2189–95.