#### ORIGINAL RESEARCH



# Efficacy of Postbiotics in a PRP-Like Cosmetic Product for the Treatment of Alopecia Area Celsi: A Randomized Double-Blinded Parallel-Group Study

Fabio Rinaldi · Anna Trink · Daniela Pinto

Received: February 11, 2020 / Published online: April 11, 2020 © The Author(s) 2020

## **ABSTRACT**

Introduction: Alopecia areata (AA), also known as 'area Celsi', is the second most common form of hair loss affecting the scalp. Newly proposed treatments for AA include low-level light therapy, biologics such as Janus kinase inhibitors and autologous platelet-rich plasma (PRP), which is a well-known "elixir" for hair growth. Bioactive peptides developed through biotechnological applications have been used to overcome the limitations of PRP. More recently, the involvement of microbiota in hair growth disorders, in AA in particular, has been reported, and the usefulness of microbial metabolites, i.e. postbiotics, has been suggested.

*Methods*: This study was a randomized double-blinded parallel-group study in which 160 persons of both sexes affected by AA and aged between 18 and 60 years were enrolled. The

subjects were randomly assigned to a treatment group (group 1), receiving the TR-PRP plus-Celsi cosmetic product, and a placebo group (group 2). The SALT (Severity of Alopecia Tool) score was determined in both groups at baseline and after 2 and 3 months of treatment, and the results compared between groups.

**Results**: The subjects in group 1 showed a significant change from baseline in SALT score at 2 months of treatment (61.04%  $\pm$  3.45%; p < 0.0001), with a further improvement at the end of treatment (3 months) (69.56%  $\pm$  4.32%; p < 0.0001). No significant changes from baseline were reported for the subjects in group 2 (T1: 26.45%  $\pm$  3.64%; T3: 27.63%  $\pm$  7.61%).

Conclusions: The results of this study provide further proof of the efficacy of bioactive peptides that mimick the growth factors present in PRP in subjects affected by AA. They also add to our knowledge of the link between microbiota and hair growth disorders, emphasizing the importance of studies on the microbial community and microbial metabolites as a novel therapeutic approach.

**Keywords:** Alopecia areata; Bee bread; Biomimetic peptides; Microbiota; Plantaricin A; Platelet-rich plasma; Postbiotics

**Enhanced Digital Features** To view digital features for this article go to https://doi.org/10.6084/m9.figshare. 11967744.

F. Rinaldi (⊠) · A. Trink · D. Pinto Human Advanced Microbiome Project (HMAP), Giuliani SpA, Milan, Italy e-mail: fabio.rinaldi@studiorinaldi.com

## **Key Summary Points**

## Why carry out this study?

Alopecia areata (AA), also known as 'area Celsi', is the second most common form of hair loss affecting the scalp. Many new treatments for AA have been developed, including autologous platelet-rich plasma (PRP), well-known as an "elixir" for hair growth, bioactive peptides developed through biotechnological applications to overcome the limitations of PRP and microbial metabolites, known as postbiotics.

The aim of the present study was to investigate the efficacy of a topically applied cosmetic product that mimicks PRP and contains postbiotics for the treatment of AA.

## What was learned from the study?

The results provide further proof of the efficacy of bioactive peptides that mimic the growth factors present in PRP in subjects affected by AA.

The results also add to our knowledge of the link between microbiota and hair growth disorders, emphasizing the importance of studies on the microbial community and microbial metabolites as a novel therapeutic approach.

## INTRODUCTION

Alopecia areata (AA), also known as 'area Celsi', is the second most common form of hair loss affecting the scalp. AA is an autoimmune disorder characterized by one or more circular bald patches on the head [1]. About 2% of cases spread to the entire scalp (alopecia totalis) or body (alopecia universalis) [2]. An incidence of > 2% among the general population has been reported, with a lifetime risk of 1.7% for both

men and women [3]. AA is mainly related to genetic, autoimmunity and inflammatory factors [4, 5], with the collapse of the immune privilege of the hair follicle reported to play a pivotal role in the pathogenesis of this autoimmune disorder [6].

First-line treatment for AA includes intralesional corticosteroids for mild cases and topical immunotherapy for extensive disease, but medications such as minoxidil, an antihypertensive vasodilator, and bimatoprost, a prostaglandin analogue, are also used, usually in combination with other treatments [7].

Given its typical manifestations and high relapse rates [8], AA is often associated with social and psychological implications [9], and research is ongoing to develop new therapies. Newly proposed treatments include low-level light therapy [10] biologics, such as Janus kinase (JAK) inhibitors [11] and autologous plateletrich plasma (PRP), which is well-known as an "elixir" for hair growth [12]. The efficacy of PRP has been reported in many clinical studies [13–15], but treatment with PRP has a number of limitations, including the absence of a standardized concentration of platelets, variation in the manufacturing of PRP and some legislative overcome these limitations. issues. To researchers have turned to the use of bioactive peptides obtained through the application of modern biotechnology techniques. These peptides mimic the activities of platelet growth factors, have an efficacy similar to PRP treatment and can be easily included in a topical [16, 17] or, in the near future, injectable formulation. More recently, the involvement of microbiota has been reported in hair growth disorders, in AA in particular [18-20], and the usefulness of microbial metabolites, referred to as postbiotics, has been suggested [21].

In the study reported here, we investigated the efficacy of a topically applied gel formulation that mimics the action of PRP and contains postbiotics for the treatment of AA.

## **METHODS**

A total of 160 male and female subjects who were affected by AA and aged between 18 and

60 years were enrolled in this randomized double-blinded parallel-group study. All patients were evaluated at the RS Dermatologic Clinic, Milan, Italy after written informed consent had been given. The study was approved by the Ethical Independent Committee for Clinical, not pharmacological investigation in Genoa (Italy) in June 2016 under the reference number Rif. 2019/06 and all procedures were in accordance with the ethical standards of the 1964 Declaration of Helsinki, as revised in 2013 concerning human rights. All patients provided written informed consent prior to inclusion in the study. The inclusion and exclusion criteria are given in Table 1.

Differential diagnoses of acute telogen effluvium, androgenetic alopecia and cicatricial alopecia in a pattern distribution were considered for all enrolled subjects [22]. Consequently, the affected area of the scalp of all enrolled subjects was also examined by polarized light dermoscopy at ×100 magnification (Molemax HD; Derma Instruments, Vienna, Austria), and the Molemax software tool integrated with the system was used for acquiring the images. A representative image is shown in Fig. 1. Dermatoscopic examination of all patients confirmed the presence of yellow dots and dystrophic hairs, as well as of cadaveric (black dots) hairs, all of which are manifestations typical of AA and occur in 95% of patients at all stages of the disease [23, 24]. These findings were, in some cases, corroborated by histopathological analysis.

Enrolled subjects were randomly assigned to the group receiving the TR-PRP plus-Celsi cosmetic product (group 1) or the placebo group (group 2). All subjects in both groups topically applied about 2 mL of the product/placebo per day (to be applied for least 5 h) for 3 months. The TR-PRP plus-Celsi product was prepared in the form of a semisolid non-ionic gel that contained all of the active ingredients and all of the excipients needed for stabilization and preservation. The main active ingredients of the TR-PRP plus-Celsi gel are biomimetic peptides (copper tripeptide-1, octapeptide-2, oligopeptide-20, acetyl decapeptide-3), postbiotics (plantaricin A [Pln A] and Lactobacillus kunkei bee bread, a fermented product and postbiotic

Table 1 Inclusion/exclusion criteria

Inclusion criteria	Exclusion criteria		
Male or female aged 18–60 years	Known sensitivity to any compound in the Investigational product		
Suffering from AA for at least 3 years	Women who were pregnant or breast feeding or who were planning a pregnancy		
AA with a SALT score between S2 and S5	Serious intercurrent infection or other active disease up to 3 months prior to study entry.		
Condition not responsive to other previous treatments, either systemic, topical or phototherapy	History of concurrent malignancy		
Subjects agreeing to follow the instructions received by the investigator and able to return to the study center at the established times	History of significant alcohol or drug abuse		
Subjects agreeing to not use any drug/cosmetic treatment able to interfere with the study results	Significant psychosocial or psychiatric disorders that may impair the subject's ability to meet the study requirements		
No participation in a similar study at the present time or during the previous 6 months	Significant concurrent medical disorders that may impair the subject's ability to participate ove the whole 1 year of the study		
Not pregnant or breastfeeding	Any other medical condition which the Investigator believed would prevent the participant from taking part in the study		

Table 1 continued

Inclusion criteria	Exclusion criteria
Subjects agreeing to sign the	
informed consent form	

AA Alopecia areata, SALT Severity of Alopecia Tool



Fig. 1 Representative dermoscopic image from enrolled subjects at baseline showing yellow dots (black arrow), black dots (black triangle), dystrophic hairs (white arrow) and exclamation mark (white triangle)

**Table 2** The Severity of Alopecia Tool score

SALT score	Description
S0	No hair loss
S1	< 25% hair loss
S2	25-49% hair loss
S3	50-74% hair loss
S4	75-99% hair loss
S5	100% hair loss

[bee bread]) and *Tropaeolum majus* flower/leaf/stem extract (which provides a gust of oxygen).

Subjects visited the clinic on three different occasions: at the randomization visit (baseline [T0]), after 2 months of treatment (T1; 60 days) and at the end of the treatment period at month 3 (T2; 90 days). The Severity of Alopecia Tool (SALT) score was used to assess the efficacy of the treatment. Digital photographs were taken

at each visit. The AA SALT score was assessed according to the guidelines of the National Alopecia Areata Foundation (Table 2) [25], with S0 indicating no hair loss; S1, < 25% hair loss; S2, 25–49% hair loss; S3, 50–74% hair loss; S4, 75–99% hair loss; S5, 100% hair loss.

The efficacy of the treatment was assessed as the change in SALT score relative to baseline, as follows:  $100 \times (\text{baseline SALT score} - \text{SALT score})$  score at T1 or T2)/baseline SALT score. The percentage of hair regrowth was also calculated according to a 6-level scale, with A0 indicating no change or further loss of hairs; A1, 1–24% regrowth; A2, 25–49% regrowth; A3, 50–74% regrowth; A4, 75–99% regrowth; A5, 100% regrowth.

At the end of the study (T2), each subject completed a questionnaire on the perceived efficacy of the treatment and product compliance.

Data were collected, and a two-sample Student's t test was used for statistical analysis. P values of < 0.05 were considered to indicate clinical significance.

## RESULTS

A total of 160 persons (Table 3) suffering from AA (SALT score S2–S5) were enrolled in the study and randomly assigned to the TR-PRP plus-Celsi group (group 1) or the placebo group

**Table 3** Baseline demographic characteristics of the subjects randomized to the two study groups

Demographic characteristics	Group 1 $(N = 80)^a$	Group 2 $(N = 80)^a$	
Men	44.00 (55.00%)	37 (47.50%)	
Women	36.00 (45.00%)	43.00 (52.50%)	
Age (years)	$51.84 \pm 9.54$	$53.12 \pm 6.18$	
Number of patches	$3.54 \pm 1.63$	$3.79 \pm 2.01$	

Values in table are presented as a number with the percentage in parenthesis or as the mean  $\pm$  standard deviation (SD)

<sup>a</sup> Group 1 subjects received the TR-PRP plus-Celsi cosmetic product as treatment; group 2 subjects received placebo

(group 2). The subjects in the two randomized groups were found to have comparable demographic characteristics.

All of the enrolled subjects had not been responsive to earlier treatments, either systemic, topical or phototherapy. The last treatment had been at least 1 year earlier.

The percentage of hair loss relative to baseline was calculated as the sum of five scalp areas (vertex, right profile, left profile, posterior aspect of scalp). The results of these calculations relative to baseline (T0) at 2 (T1) and 3 months (T2) of treatment are reported in Table 4. The subjects in group 1 showed a significant (p < 0.0001) change from the baseline SALT 2 months score of treatment at  $(61.04\% \pm 3.45\%)$ , with a further improvement observed at the end of the treatment (T2, 3 months) (69.56%  $\pm$  4.32%; p < 0.0001). No significant changes from baseline to T1 and T2 were observed for the subjects in group 2 (T1:  $26.45\% \pm 3.64\%$ ; T2:  $27.63\% \pm 7.61\%$ ).

Grading of the overall improvement in the subjects in group 1 and 2, respectively, is reported in Table 5. In group 1, 47.50% of subjects achieved a complete regression (A5) and, in line with results from previous studies [16], 13.75% achieved a partial regression (A3); only 6.25% of subjects in group 1 reported no response at all (Table 5). In contrast, only 5% of the subjects in group 2 reported a complete regression (Table 5).

No adverse effects were reported from the subjects in both groups, and all subjects were in good compliance with the tested products.

An explicative dermoscopic image after treatment is shown in Fig. 2. Figure 3 shows

**Table 5** Grading of overall improvement in group I and group II subjects

Overall improvement	Group 1 (N)	Group 2 (N)		
A0 (no hair regrowth)	5	29		
A1 (1-24%)	6	34		
A2 (25–49%)	5	8		
A3 (50-74%)	11	5		
A4 (75–99%)	15	0		
A5 (100%)	38	4		



Fig. 2 Representative dermoscopic image from enrolled subjects at baseline (T0)

explicative photographic images of the effect of the TR-PRP plus-Celsi treatment on hair regrowth in three subjects in group I.

## DISCUSSION

Various treatment options are available to clinicians for the management of hair growth

Table 4 Percentage changes in baseline Severity of Alopecia Tool score in group 1 and group 2 subjects

Percentage changes in baseline SALT score	After 2 months of treatment (T1)	After 3 months of treatment (T2)		Statistical analysis (two-sample Student's <i>t</i> test) <sup>a</sup>		
			T0 vs.	T0 vs. T2	T1 vs. T2	
Group I	$61.04 \pm 3.45$	$69.56 \pm 4.32$	< 0.0001	< 0.0001	< 0.0001	
Group II	$26.45 \pm 3.64$	$27.63 \pm 7.61$	0.040	0.715	0.956	

Values in table are presented as the mean  $\pm$  SD

<sup>&</sup>lt;sup>a</sup> T0, Baseline; T1, after 2 months (60 days) of treatment; T2, after 90 days (end of treatment period)

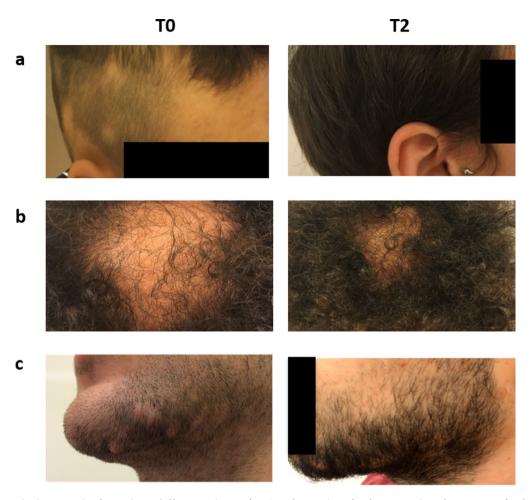


Fig. 3 Digital photographs from three different subjects (a, b, c). Baseline (T0), 3 months of treatment (T2)

disorders (AA, androgenetic alopecia [AGA], lichen planopilaris) [7, 26, 27] Autologous PRP is a relatively new therapeutic approach for patients suffering from alopecias that has achieved a high degree of compliance among patients [15, 28, 29]. It differs from other treatments in terms of its mechanism of action, which is based on the actions of the growth factors contained in the formulation, such as growth platelet-derived factor. vascular endothelial growth factor and transforming growth factor, among others, in regulating cell migration, proliferation, remodeling of the extracellular matrix and promotion endothelial permeability [30].

Platelet growth factors can interact with both dermal papilla cells and cells from the bulge area, with the effect of activating the proliferative phase of the hair [31–35].

A large number of biomimetic peptides have been developed to overcome the limitations of autologous PRP [36]. Short chains of 10–15 amino acids act similarly to natural growth factors by mimicking their structure or activity, or both [37]. Such peptides possess a higher stability and specificity, are less expensive than PRP and can be easily included in a topical formulation. These benefits have been shown in many applications, ranging from skin rejuvenation [37, 38] to wound-healing [38] and hair growth [39–44].

In our study, we reported the efficacy of a PRP-like cosmetic gel containing postbiotics for the treatment of AA. Of the 160 subjects enrolled in the study, a significant improvement in hair regrowth was recorded in the majority of the 80 subjects treated with the active product. These results are in line with those from our previous studies in which we demonstrated the efficacy of a pool of peptides derived by biotechnology that mimic platelet growth factors in AA [16] and AGA [17]. These peptides include copper tripeptide-1, oligopeptide-20, acetyl decapeptide-3 and octapeptide-2. Copper tripeptide-1 (copper-glycyl-L-histidyl-Llysine [GHKCu]) [45] stimulates stem cells in the hair follicle [46, 47], possesses a remodelling activity [48-51] and stimulates metalloproteinases [**45**, 52-54] and angiogenesis [48, 55-57]. Oligopeptide-20 (8H-Cys-Arg-Lys-Ile-Pro-Asn-Gly-Tyr-Asp-Thr-Leu-OH) involved in hair growth mechanisms bv increasing the synthesis of collagen and glycosaminoglycans [58], thereby preserving hair follicles from aging [59]. Acetyl decapeptide-3 acts as a biomimetic of basic fibroblast growth factor and has been reported to stimulate hair growth [60, 61]. Octapeptide-2 (Thr-Ala-Glu-Glu-His-Glu-Val-Met) is a biomimetic of the hair growth stimulator thymosin-b4 [40, 41]; most interestingly, it also possesses a strong antimicrobial activity [62].

Two other main active components of the TR-PRP plus-Celsi cosmetic product are Pln A and Lactobacillus kunkeei-fermented bee bread (bee bread); both can be considered to be "postbiotics". Postbiotics are class molecules with health-promoting effects that are derived from microorganisms, usually probiotics [63]. More specifically, they are substances (metabolites, enzymes, bioactive peptides, short-chain fatty acids, antimicrobial peptides, polysaccharides, cell surface proteins, vitamins, plasmalogens and organic acids) produced by beneficial bacteria as a metabolic product [64] that possess several functional properties, mainly antimicrobial, antioxidant and immunomodulatory in nature [63]. It has been hypothesized that these substances are responsible for probiotic efficacy. Therefore, compared to probiotics, postbiotics can interact easier with both the microbiome and human cells of the host [65]. Other advantages derived from the use of postbiotics are (1) no need for survival or propagation; (2) greater absorption, distribution and extraction potential than probiotics; (3) a safer profile [66].

Pln A and bee bread are obtained by fermenting a raw matrix with lactic acid bacteria. PlnA is an antimicrobial peptide with a pheromone-like activity, produced as the result of quorum sensing between two lactic acid bacteria [67]. It possesses proliferative and woundhealing activities and is also able to induce key mediators of the proliferation, migration and differentiation of epithelial cells [67, 68]. Bee bread is the fermented endproduct of bee-collected pollen [69, 70]. In vitro tests on human keratinocytes (patent bee bread) have demonstrated the immunomodulatory effect of bee bread. Both of of these postbiotics have a strong antioxidant activity [68-70]. Regarding the endproducts of microbial metabolism, postbiotics can also be positively sensed by the resident microbiota of the host, modulating the network pathways of both the microbiota and the host.

In an earlier study on microbial dysbiosis on the scalp of AA subjects, we hypothesized a correlation between the presence of some microbial strains in the perifollicular region of AA subjects and hypoxia [18]. This hypothesis has led to novel therapeutic approaches aimed at resolving microbial dysbiosis, including via oxygenation. In this context, the TR-PRP plus-Celsi cosmetic product also contains *Tropaeolum majus* flower/leaf/stem extract, which is a natural active ingredient derived from the nasturtium flower that boosts oxygenation by significantly increasing the activity of hypoxia-inducible factor 1-alpha [71].

## CONCLUSION

The results of this study provide further proof of the efficacy of bioactive peptides that mimic the growth factors present in PRP in subjects affected by AA. They also add to our knowledge of the link between microbiota and hair growth disorders, emphasizing the importance of studies on the microbial community and microbial metabolites as a novel therapeutic approach.

## **ACKNOWLEDGEMENTS**

We thank the participants of the study.

*Funding.* This study and the journal's Rapid Service Fee were supported by Giuliani SpA.

Authorship. All named authors meet the International Committee of Medical Journal Editors (ICMJE) criteria for authorship for this manuscript, take responsibility 3for the integrity of the work as a whole, and have given final approval for the version to be published.

**Disclosures.** Fabio Rinaldi and Anna Trink serve as a consultant for Giuliani S.p.A. Daniela Pinto is employed by Giuliani S.p.A.

Compliance with Ethics Guidelines. The study was approved by the Ethical Independent Committee for Clinical, not pharmacological investigation in Genoa (Italy) in June 2016 under reference number Rif. 2019/06, and all procedures were in accordance with the ethical standards of the 1964 Declaration of Helsinki, as revised in 2013, concerning human rights. All patients provided written informed consent prior to inclusion in the study.

**Data Availability.** The datasets during and/ or analyzed during the current study are available from the corresponding author on reasonable request.

*Open Access.* This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License, which permits any non-commercial use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <a href="http://creativecommons.org/licenses/by-nc/4.0/">http://creativecommons.org/licenses/by-nc/4.0/</a>.

## REFERENCES

- 1. Odom RB, Davidsohn IJ, William D, Henry JB, Berger TG. Clinical diagnosis by laboratory methods. In: Elston, Dirk M, editors. Andrews' diseases of the skin: clinical dermatology. Philadelphia/Amsterdam: Saunders/Elsevier; 2006, p. 637.
- 2. Darwin E, Hirt PA, Fertig R, Doliner B, Delcanto G, Jimenez JJ. Alopecia areata: review of epidemiology, clinical features, pathogenesis, and new treatment options. Int J Trichol. 2018;10(2):51–60.
- 3. Dainichi T, Kabashima K. Alopecia areata: What's new in epidemiology, pathogenesis, diagnosis, and therapeutic options? J Dermatol Sci. 2017;86(1): 3–12
- Pratt CH, King LE Jr, Messenger AG, Christiano AM, Sundberg JP. Alopecia areata. Nat Rev Dis Primers. 2017;3:17011. https://doi.org/10.1038/nrdp.2017. 11.
- 5. Paus R, Bertolini M. The role of hair follicle immune privilege collapse in alopecia areata: status and perspectives. J Investig Dermatol Symp Proc. 2013;16(1):S25–7.
- Azzawi S, Penzi LR, Senna MM. Immune privilege collapse and alopecia development: is stress a factor. Skin Appendage Disord. 2018;4(4):236–44.
- 7. Gupta AK, Carviel J, Abramovits W. Treating alopecia areata: current practices versus new directions. Am J Clin Dermatol. 2017;18(1):67–75.
- 8. Harries MJ, Sun J, Paus R, King LE Jr. Management of alopecia areata. BMJ. 2010;341:c3671.
- 9. Nardi AE. Psychological impact of alopecia: alopecia may lead to social anxiety. BMJ. 2005;331(7524):1084.
- 10. Darwin E, Arora H, Hirt PA, Wikramanayake TC, Jimenez JJ. A review of monochromatic light devices for the treatment of alopecia areata. Lasers Med Sci. 2018;33(2):435–44.
- 11. Damsky W, King BA. JAK inhibitors in dermatology: the promise of a new drug class. J Am Acad Dermatol. 2017;76(4):736–44.

- 12. Garg S, Manchanda S. Platelet-rich plasma-an 'Elixir' for treatment of alopecia: personal experience on 117 patients with review of literature. Stem Cell Investig. 2017;4:64.
- Giordano S, Romeo M, di Summa P, et al. A metaanalysis on the evidence of platelet-rich plasma for androgenetic alopecia. Int J Trichology. 2018;10: 1–10.
- 14. Sorbellini E, Trink A, Rinaldi F. Experimental clinical assessment of the use of platelet-rich plasma in dermatology and rationale for its use in the treatment of non-scarring alopecia. Presented at the 35th La Medicina Estetica 4 October 2011. 43.
- 15. Trink A, Sorbellini E, Bezzola P, et al. A randomized, double-blind, placebo- and active-controlled, half-head study to evaluate the effects of platelet-rich plasma on alopecia areata. Br J Dermatol. 2013;169: 690–4.
- Rinaldi F, Marzani B, Pinto D, Sorbellini E. Randomized controlled trial on a PRP-like cosmetic, biomimetic peptides based, for the treatment of alopecia areata. J Dermatol Treat. 2019;30(6): 588–93.
- 17. Rinaldi F, Marzani B, Pinto D, Sorbellini E. Efficacy of a cosmetic product mimicking PRP in androgenetic alopecia. J Transl Sci. 2019;5:1–5.
- Pinto D, Sorbellini E, Marzani B, Rucco M, Giuliani G, Rinaldi F. Scalp bacterial shift in Alopecia areata. PLoS ONE. 2019;14(4):e0215206.
- Ho BS, Ho EXP, Chu CW, et al. Microbiome in the hair follicle of androgenetic alopecia patients. PLoS ONE. 2019;14(5):e0216330.
- Migacz-Gruszka K, Branicki W, Obtulowicz A, Pirowska M, Gruszka K, Wojas-Pelc A. What's new in the pathophysiology of alopecia areata? The possible contribution of skin and gut microbiome in the pathogenesis of alopecia–Big opportunities, big challenges, and novel perspectives. Int J Trichol. 2019;11:185–8.
- 21. Rinaldi F, Pinto D, Giuliani G. Postbiotic evolution in dermatology. EC Microbiology. 2020;16(3):01–4.
- Finner AM. Alopecia areata: clinical presentation, diagnosis, and unusual cases. Dermatol Ther. 2011;24(3):348–54.
- Ross EK, Vincenzi C, Tosti A. Videodermoscopy in the evaluation of hair and scalp disorders. J Am Acad Dermatol. 2006;55:799–806.
- 24. Inui S, Nakajima T, Nakagawa K, Itami S. Clinical significance of dermoscopy in alopecia areata:

- analysis of 300 cases. Int J Dermatol. 2008;47: 688–93.
- 25. Olsen E, Hordinsky M, McDonald-Hull S, et al. Alopecia areata investigational assessment guidelines. National Alopecia Areata Foundation. J Am Acad Dermatol. 1999;40:242–6.
- 26. Kaliyadan F, Nambiar A, Vijayaraghavan S. Androgenetic alopecia: an update. Indian J Dermatol Venereol Leprol. 2013;79:613–25. https://doi.org/10.4103/0378-6323.116730.
- 27. Errichetti E, Figini M, Croatto M, Stinco G. Therapeutic management of classic lichen planopilaris: a systematic review. Clin Cosmet Investig Dermatol. 2018;11:91–102.
- 28. Sohrab K, Tahir K, Anam E, Tahir JA. Role of autologous platelet-rich plasma (PRP) in limited alopecia areata in local population. J Pak Assoc Dermatol. 2016;26(2):107–11.
- 29. Borhan R, Gasnier C, Reygagne P. Autologous platelet rich plasma as a treatment of male androgenetic alopecia: study of 14 cases. J Clin Exp Dermatol Res. 2015;6:292.
- 30. Pavlovic V, Ciric M, Jovanovic V, Stojanovic P. Platelet rich plasma: a short overview of certain bioactive components. Open Med (Wars). 2016;11(1):242–7.
- 31. Gentile P, Garcovich S, Bielli A, et al. The effect of platelet-rich plasma in hair regrowth: a randomized placebo-controlled trial. Stem Cells Transl Med. 2015;4:1317–23.
- 32. Pai VV, Bhandari P, Shukla P. Topical peptides as cosmeceuticals. Indian J Dermatol Venereol Leprol. 2017;83:9–18.
- 33. B€ohlen P, Esch F, Baird A, et al. Acidic fibroblast growth factor (FGF) from bovine brain: amino-terminal sequence and comparison with basic FGF. Embo J. 1985;4:1951–6.
- 34. Kawano M, Komi-Kuramochi A, Asada M, et al. Comprehensive analysis of FGF and FGFR expression in the skin: fGF18 is highly expressed in hair follicles and capable of inducing anagen from telogen stage hair follicles. J Invest Dermatol. 2005;124:877–85.
- 35. Kimura-Ueki M, Oda Y, Oki J, et al. Hair cycle resting phase is regulated by cyclic epithelial FGF18 signaling. J Invest Dermatol. 2012;132:1338–45.
- 36. Giordano S, Romeo M, di Summa P, et al. A metaanalysis on evidence of platelet-rich plasma for androgenetic alopecia. Int J Trichol. 2018;10:1–10.

- 37. Gazitaeva ZI, Drobintseva AO, Chung Y, Polyakova VO, Kvetnoy IM. Cosmeceutical product consisting of biomimetic peptides: antiaging effects in vivo and in vitro. Clin Cosmet Investig Dermatol. 2017;10:11–6.
- 38. Lima TN, Pedriali Moraes CA. Bioactive peptides: applications and relevance for cosmeceuticals. Cosmetics. 2018;5:21.
- 39. Pyo HK, Yoo HG, Won CH, et al. The effect of tripeptide-copper complex on human hair growth in vitro. Arch Pharm Res. 2007;30:834–9.
- 40. Lee WJ, Sim HB, Jang YH, et al. Efficacy of a complex of 5-aminolevulinic acid and glycyl-histidyllysine peptide on hair growth. Ann Dermatol. 2016;28:438–43.
- 41. Philp D, Nguyen M, Scheremeta B, et al. Thymosin beta4 increases hair growth by activation of hair follicle stem cells. Faseb J. 2004;18:385–7.
- 42. Philp D, St-Surin S, Cha HJ, et al. Thymosin beta 4 induces hair growth via stem cell migration and differentiation. Ann N Y Acad Sci. 2007;1112: 95–103.
- 43. Ito C, Saitoh Y, Fujita Y, et al. Decapeptide with fibroblast growth factor (FGF)-5 partial sequence inhibits hair growth suppressing activity of FGF-5. J Cell Physiol. 2003;197:272–83.
- 44. Bassino E, Zanardi A, Gasparri F, et al. Effects of the biomimetic peptide Sh-Polypeptide 9 (CG-VEGF) on cocultures of human hair follicle dermal papilla cells and microvascular endothelial cells. Exp Dermatol. 2016;25:237–9.
- 45. Pickart L, Downey D, Lovejoy S, et al. Gly-l-his-l-lys:copper(II)—a human plasma factor with super-oxide dismutase-like and wound-healing properties. In: Rotilio G, editor. Superoxide and superoxide dismutase. Amsterdam: Elsevier; 1986. p. 555–8.
- 46. Uno H, Kurata S. Chemical agents and peptides affect hair growth. J Invest Dermatol. 1993;101: 143S–7S.
- 47. Chen P, Cescon M, Bonaldo P. Lack of collagen VI promotes wound-induced hair growth. J Invest Dermatol. 2015;135:2358–67.
- 48. Miller DM, DeSilva D, Pickart L, et al. Effects of glycyl-histidyl-lysyl chelated Cu(II) on ferritin dependent lipid peroxidation. Adv Exp Med Biol. 1990;264:79–84.
- 49. Vinci C, Caltabiano V, Santoro AM, et al. Copper addition prevents the inhibitory effects of

- interleukin 1-beta on rat pancreatic islets. Diabetologia. 1995;38:39–45.
- 50. McCormack MC, Nowak KC, Koch RJ. The effect of copper tripeptide and tretinoin on growth factor production in a serum-free fibroblast model. Arch Facial Plast Surg. 2001;3:28–32.
- 51. Simeon A, Monier F, Emonard H, et al. Expression and activation of matrix metalloproteinases in wounds: modulation by the tripeptide-copper complex glycyl-L-histidyl-L-lysine- Cu2þ. J Invest Dermatol. 1999;12:957–64.
- 52. Simeon A, Wegrowski Y, Bontemps Y, et al. Expression of glycosaminoglycans and small proteoglycans in wounds: modulation by the tripeptide-copper complex glycyl-L-histidyl- L-lysine-Cu(2b). J Invest Dermatol. 2000;115:962–8.
- 53. Simeon A, Emonard H, Hornebeck W, et al. The tripeptidecopper complex glycyl-L-histidyl-L-lysine-Cu2b stimulates matrix metalloproteinase-2 expression by fibroblast cultures. Life Sci. 2000;18: 2257–665.
- 54. Raju KS, Alessandri G, Gullino PM. Characterization of a chemoattractant for endothelium induced by angiogenesis effectors. Cancer Res. 1984;44: 1579–84.
- 55. Pickart L. New metal peptide complexes and derivatives used for stimulating growth of hair in warm-blooded animals, especially humans. US Patent 5,120,831. Alexandria: United States Patent and Trademark Office.
- 56. Compositions for stimulating hair growth containing cupric complexes of peptide derivatives including glycyl-l-histidyl-l-lysine n-octyl ester. US Patent 5,177,061.
- 57. New glycyl-histidyl-lysyl copper compounds used in stimulating hair growth; US Patent 5,214,032.
- 58. Metal-peptide compositions and methods for stimulating hair growth, US Patent 5,550,183.
- 59. Matsumura H, Mohri Y, Binh NT, et al. Hair follicle aging is driven by transepidermal elimination of stem cells via COL17A1 proteolysis. Science. 2016. https://doi.org/10.1126/science.aad4395.
- 60. Lin WH, Xiang LJ, Shi HX, et al. Fibroblast growth factors stimulate hair growth through b-catenin and Shh expression in C57BL/6 mice. Biomed Res Int. 2015;2015:730139.
- 61. Yoon SY, Kim K-T, Jo SJ, et al. Induction of hair growth by insulin-like growth factor-1 in 1,763 mhz radiofrequency-irradiated hair follicle cells. PLoS ONE. 2011;6:e28474.

- 62. Tang YQ, Yeaman MR, Selsted ME. Antimicrobial peptides from human platelets. Infect Immun. 2002;70(12):6524–33.
- 63. Toalaa JE, Garcia Varelab R, Garciac HS, Harod V. Postbiotics: an evolving term within the functional foods field. Trends Food Sci Technol. 2018;75: 105–14.
- 64. Tsilingiri K, Rescigno M. Postbiotics: what else? Beneficial Microb. 2013;4(1):101–7.
- 65. Shenderov BA. Metabiotics: novel idea or natural development of probiotic conception. Microbial Ecol Health Dis. 2013;24:1–8.
- Tsilingiri K, Barbosa T, Penna G, Caprioli F, Sonzogni A, Viale G. Probiotic and postbiotic activity in health and disease: comparison on a novel polarized ex vivo organ culture model. Gut. 2012;61: 1007–15.
- 67. Pinto D, Marzani B, Minervini F, et al. Plantaricin A synthesized by *Lactobacillus plantarum* induces in vitro proliferation and migration of human

- keratinocytes and increases the expression of TGF- $\beta$ 1, FGF7, VEGF-A and IL-8 genes. Peptides. 2011;32(9):1815–24.
- 68. Marzani B, Pinto D, Minervini F, et al. The antimicrobial peptide pheromone Plantaricin A increases antioxidant defenses of human keratinocytes and modulates the expression of filaggrin, involucrin, β-defensin 2 and tumor necrosis factor-α genes. Exp Dermatol. 2012;21(9):665–71. https://doi.org/10.1111/j.1600-0625.2012.01538.x.
- 69. Di Cagno R, Filannino P, Cantatore V, Gobbetti M. Novel solid-state fermentation of bee-collected pollen emulating the natural fermentation process of bee bread. Food Microbiol. 2019;82:218–30.
- 70. Giuliani G, Gobbetti M, Di Cagno R, Filannino P, Cantatore V, Mascolo A, Marzani B. WO2020016770 (A1) Microbiological process for the production of bee bread. 2020.
- 71. SILAB. Oxygenskin®, Reactivates the skin's oxygenation mechanisms. Saint-Viance: SILAB. https://www.silab.fr/produit-75-oxygeskin\_usa.html