

Metastasis-Initiating Cells in Renal Cancer

Mohammed I. Khan^{1,*}, Anna M. Czarnecka¹, Renata Duchnowska¹, Wojciech Kukwa² and Cezary Szczylik¹

¹Molecular Oncology Laboratory, Clinic of Oncology, Military Institute of Medicine, ul. Szaserów 128, 04-141 Warsaw, Poland; ²Department of Otolaryngology, Czerniakowski Hospital, Medical University of Warsaw, ul. Stepinska 19/25, Warsaw, Poland

Abstract: Metastasis is a complex process that propagates cells from the primary or initial site of the cancer occurrence to distant parts of the body. Cancer cells break from the cancer site and circulate through the bloodstream or lymph vessels, allowing them to reach nearly all parts of the body. These circulating tumour cells (CTCs) contain specialized metastasis-initiating cells (MICs) that reside in the biological heterogeneous primary tumour. Researchers have hypothesized that metastasis of renal cell carcinoma is initiated by circulation of MICs in patients' blood and bone marrow. Based on the cancer stem/progenitor cell concept of carcinogenesis, understanding the molecular phenotypes of metastasis-initiating cells (MICs) in renal cancer could play a vital role in developing strategies for therapeutic interventions in renal cancer. Existence of MICs among CTCs in renal carcinoma has not been proven in large scale. However, some studies have reported that specialized markers are found on the surface of circulating cells from the primary tumour. In mice, MICs have been isolated from CTCs using such markers, which have then been transplanted into xenograft model to show whether they give rise to metastasis in different organs. Considering these findings, in this review we have attempted to summarize the studies connected with MICs and their gene expression profiles that are responsible for metastasis in renal cancer.

Keywords: Circulating tumour cells, metastasis-initiating cells, renal cell carcinoma, tumour-initiating cells.

INTRODUCTION

Renal Cell Carcinoma (RCC)

Renal cell carcinoma (RCC) is the most common epithelial malignancy of human adult kidneys, accounting for 3% of all neoplasms. At 40%, RCC has the highest mortality [1]. The most important prognostic factor in RCC is metastatic dissemination of disease. Approximately 30% of patients with RCC will be diagnosed with metastatic disease, and 60% of these patients will die from aggressive disease and metastases [2]. Within Europe, 42,000 patients are diagnosed with RCC, and 25,000 of these die each year.

Immunotherapies such as interferon (INF)- α and interleukin (IL)-2 have been used for decades in metastatic RCC treatment but have provided positive effects in only 10 – 20% of cases [3,4]. For the past ten years, clinical studies have used targeted molecular therapies such as orally administered sorafenib, sunitinib, pazopanib and tivozanib (receptor tyrosine kinase inhibitors), as well as mammalian target of rapamycin (mTOR) inhibitors (everolimus and temsirolimus) for treating patients with RCC. First-, second- and third-line treatments have shown some positive results. However, continuous treatments with these drugs are associated with a high incidence of toxic effects and resistance [5,6]. The targets of these agents are the signalling pathways

responsible for angiogenesis and cancer-cell proliferation. The clinical response is very low, and five-year survival of patients with metastatic RCC is only 10% [7-9].

Tumour-Initiating Cells (TICs) in Renal Cell Carcinoma (RCC)

Increasing evidence shows the existence of small populations of tumour-initiating cells (TICs) or cancer stem cells (CSCs) that reside in the vicinity of the heterogeneous mass of the tumour. TICs have been identified in melanoma [19] and many solid tumours, including brain [10], breast [11], prostate [12], ovarian [13], colon [14], gastric [15], pancreatic [16], head and neck [17] and liver cancers [18]. Like normal stem cells, these specialized tumour regenerating cells share common properties and can be characterized by their ability to self-renew and their capacity to form serially transplantable tumours in immune-deficient mice. On the basis of different protein expression on the cell surface, cells such as CD133, CD105, NCAM, selecting side population (SP) can be easily isolated and identified through aldehyde dehydrogenase 1 (ALDH1) and rhodamine 123 (Rh123) dye activity [20-23] (Fig. 1). During conventional treatments such as radiation and chemotherapy, these cells are not targeted and are responsible for resistance. Afterward, treatment failure results in patient relapse. Currently, there are no efficient treatments for TICs. However, a number of pre-clinical trials have been performed using different therapeutic approaches to halt the proliferation of TICs. These therapies include induction of cell differentiation and blockage of TIC maintenance pathways [24].

*Address correspondence to this author at the Molecular Oncology Laboratory, Clinic of Oncology, Military Institute of Medicine, ul. Szaserów 128, 04-141 Warsaw, Poland; Tel: + 48 - 790763164; Fax: + 48 - 22 - 610 - 30 - 98; E-mails: mkhan@wim.mil.pl, imrankhanbioinfo@gmail.com

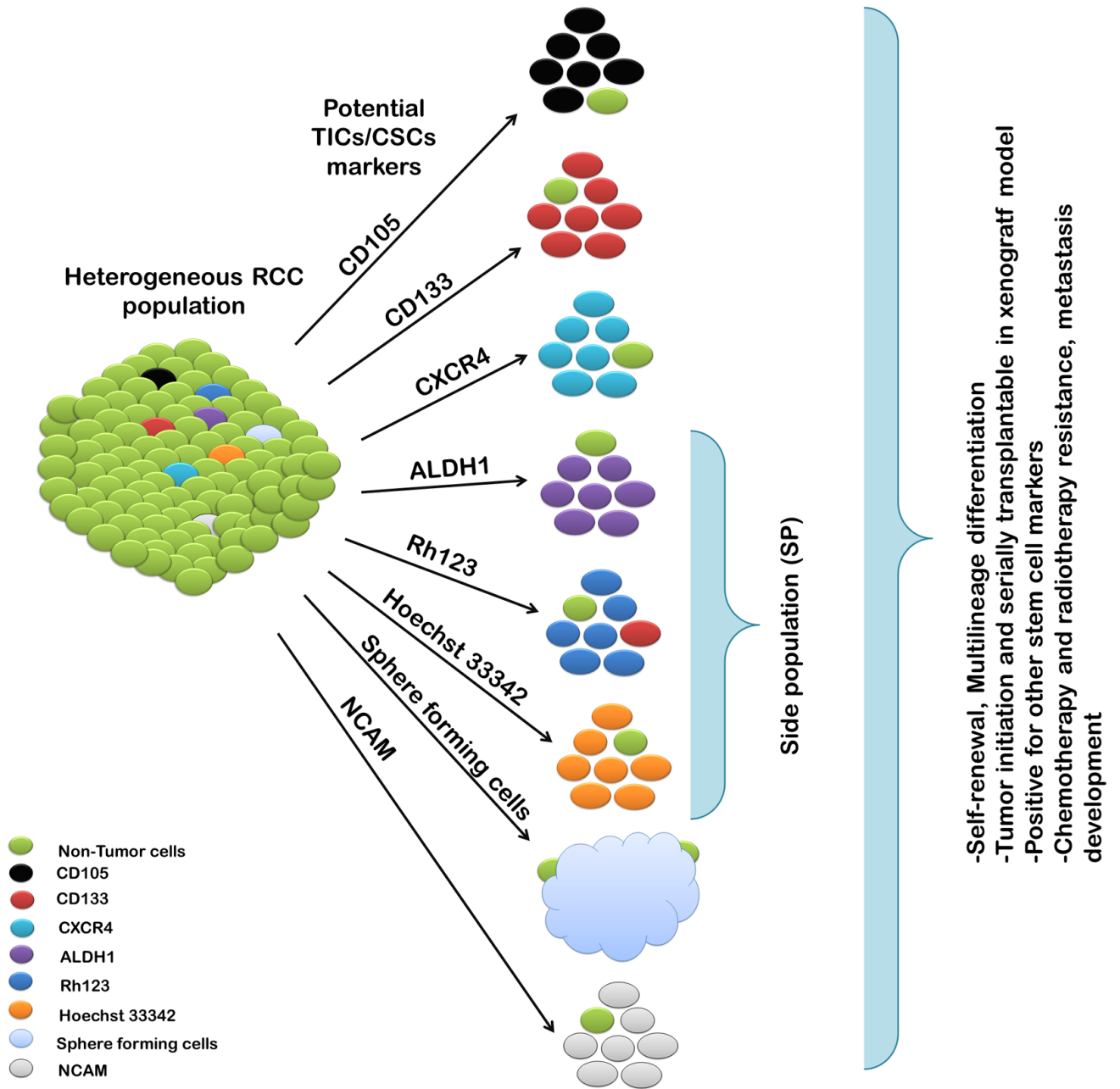


Fig. (1). Diagram showing enrichment methods for tumour initiating cells (TICs) in renal cell carcinoma (RCC) and TIC characteristics.

TICs have been identified and isolated from RCC patients and cell lines using mesenchymal stem cell marker CD105 (endoglin) [25,26]. To evaluate the tumorigenicity of the isolated cells, CD105⁺ (1x10⁶) and CD105⁻ (1x10⁶) cells were transplanted subcutaneously into severely compromised immunodeficient (SCID) mice [25]. CD105⁺ populations showed induced tumours in 100% of cases (10/10). In contrast, 10% tumour incidence (1/10) was observed with CD105⁻ cells. These cells appear to be important in RCC since angiogenesis is critical for tumour growth and development of metastasis, and CD105 is a membrane glycoprotein that is highly expressed in activated endothelial cells associated with angiogenesis. CD105 protein is part of a receptor for TGF-B1 and TGF-B3 that promotes cell

proliferation and differentiation. Moreover, endoglin expression was inversely correlated with Tumour-Node-Metastasis (T-N-M) [27]. Thus, endoglin could be a marker of tumour neovascularization and provide prognostic information in RCC development.

Others have also identified TICs using Hoechst 33342 dye as epithelial side population (SP) in malignant RCC human kidney tissue [28]. SP population cells demonstrated greater potential of colony-forming efficiency. They were enriched for high proliferation in culture conditions and stem cell-like characteristics. Additionally, spheres derived from culturing single cells of RCC cell lines were enriched for cancer stem-like cells [29]. These sphere-forming cells

were able to self-renew in *in vivo* and *in vitro* models and presented higher expression of 'stemness' genes and resistance to chemotherapeutic agents and radiotherapy compared to monolayer-adherent cells. Recently, Huang *et al.* [30] reported cancer stem cell-like cells with SP phenotype using Hoechst 33342 dye in human primary RCC cell lines. Less than 5% SP population has been reported in all studied RCC cell lines. Sorted SP populations have high expression of ABCB1, ABCG2 and ABCC1 proteins and are resistant to chemotherapy and radiotherapy. However, pre-treatment of SP cells with verapamil, an ABC transporter inhibitor, reversed drug resistance, thereby demonstrating that the ABC transporter could be responsible for drug resistance [30,31]. Moreover, cancer-testis antigen and HSP40 family member DNAJB8 were important in maintaining the TIC/CSC phenotype of the SP population in RCC cells, and overexpression of DNAJB8 increased the percentage of SP cells [32].

CD133 has been investigated as a putative stem cell marker in many solid tumours, including RCC [21,33]. A small population of CD133⁺ cells have been found in RCC and characterized as renal resident progenitor cells [26, 34]. CD133⁺ cells were able to differentiate into endothelial and epithelial cells. Therefore, a low number of CD133⁺ cells in RCC might be an early event in stem cell differentiation and possibly in malignant transformation. In addition, when undifferentiated CD133⁺ cells from RCC were subcutaneously transplanted alone into SCID in mice, the mice did not give rise to tumour formation, suggesting CD133⁺ are not TICs. However, when co-transplanted with renal carcinoma cells, CD133⁺ progenitors fostered tumour engraftment, growth and development. The tumours formed *in vivo* demonstrated that CD133⁺ derived endothelial cells were able to form functional vessels enriched in human HLA class I connected with the mouse vasculature. This showed that CD133 can contribute to the growth factor stimulation necessary for angiogenesis [34]. The expression level of *CD133* in RCC patients biopsies were not correlated with clinical pathology and prognostic significance [35].

The chemokine receptor CXCR4 is a putative stem cell marker, and its increased expression has been recently reported in renal carcinoma [36,37]. Two RCC cell lines derived from the primary and metastatic site were used to demonstrate that high expression of CXCR4 is associated with a more tumorigenic cell line [38]. RCC cell lines derived from the metastatic site were found enriched in CXCR4⁺ cells and capable of forming larger spheres *in vitro*, generating more tumours in mice when compared with CXCR4⁻ cells [24]. These results demonstrated that CXCR4⁺ cells as a TIC subpopulation the differences in CXCR4 expression reveal differences in the TICs content in RCC cell lines. Moreover, CXCR4⁺ cells exhibited greater resistance to tyrosine kinase inhibitors and expressed other 'stemness' associated genes. Surgical biopsies of the RCC patients confirmed that the high expression of *CXCR4* has significant prognostic value and therapeutic importance [35].

The ability of stem cells to efflux dye such as Rhodamine 123 (Rh123) can be used to analyse and isolate these cells with progenitor characteristics [39,40]. Based on Rh123 staining, RCC cells were characterized as Rh123^{high} and Rh123^{low} cells in a recent study [39]. Serially transplanted

Rh123^{high} cells into SCID mice were able to form tumours in all cases (12/12 cases). However, no visible tumours were observed for Rh123^{low} cells (1/12 cases) [39]. In addition, the Rh123^{high} cells showed higher differentiating potential and increased survival capability against radiotherapy compared to the Rh123^{low} cells. This finding indicates that TICs might exist in Rh123^{high} populations in RCC cells, which is opposite the results reported in other cancers [41,42]. This could explain the different biological characteristics of RCC compared to other cancer tissues.

Other markers have been analysed in RCC. Podeshakked *et al.* [22] observed neural cell adhesion molecule-NCAM as a putative marker for malignant renal stem/progenitor in Wilms' tumour. However, this marker has not been investigated in RCC. The intracellular aldehyde dehydrogenase 1 (ALDH1) functions to catalyse the oxidation of aldehyde and maintain cellular homeostasis. Recent studies have shown that normal and cancer cells with high ALDH1 activity have great potential to function as TICs in metastatic RCC cell lines [43]. ALDH1 activity analysis revealed that SP populations from metastatic RCC cell lines have higher levels of activity for ALDH1 compared to NSP (non-side population). ALDH1⁺ cells also showed higher sphere-forming ability, self-renewal, tumorigenic and expressed higher mRNA levels of stem cell associated genes [43].

In summary, CD105, CXCR4 and ALDH1 activity could be suitable targets for identifying tumour- initiating cell markers in RCC [24, 25, 43]. (See Table 1.)

Metastasis-Initiating Cells (MICs) as Circulating Tumour Cells (CTCs) in RCC

High cancer-associated mortality is mostly due to tumour metastasis. The term metastasis is used to indicate the successful dissemination of tumour cells from the primary tumour site to other parts of the body. Recent findings indicate that circulating tumour cells (CTCs) could contribute to metastasis [44]. The discovery of novel biomarkers for identifying CTCs in many human cancers has provided a novel way of understanding tumorigenesis at the cellular level. The mechanism of the formation of metastasis is comprised of three events: (1) A group of cancer cells detach from the primary tumour. (2) The CTCs circulate to distant parts of the body *via* the bloodstream or lymphatic system and adapt to the new environment. (3) The cells then settle in the new location to proliferate and colonize, giving rise to a metastatic tumour. However, this process is highly inefficient, as numerous disseminated cells die during migration, and some remain dormant for several years [44]. Because metastasis occurs through CTCs, examining these cells could lead to future research directions and help realize the cells' clinical potential in RCC treatment and diagnosis.

The clinical relevance of disseminated CTCs in the blood stream of RCC patients remains a controversial subject, as the biology of these cells has been poorly understood [45]. Investigations in patients with different tumours have assessed the prognostic value of disseminated epithelial cells in bone marrow and peripheral blood detected by immunocytochemistry using anti-cytokeratin antibodies [46-48]. Only a few studies in RCC patients have demonstrated the possibility of cytokeratin-positive (CK+) cells as markers for CTCs investigated independently in peripheral blood and

Table 1. Tumour-initiating cell (TIC) markers and function in RCC.

TICs' Identification Markers	Function in RCC	References
CD105 (Endoglin)	Proliferation and differentiation in endothelial cells	Bussolati <i>et al.</i> [25]
CD133 (Prominin-1)	Angiogenesis	Bruno <i>et al.</i> [34]
CXCR4 (Chemokine receptor type 4)	Maintaining TICs and spread metastasis	Gassenmaier <i>et al.</i> [24]
ALDH1 activity	Maintain cellular homeostasis	Ueda <i>et al.</i> [43]
Hoechst 33342 staining	ABC activity	Addle <i>et al.</i> [28]
Rhodamine 123 staining	ABC activity	Lu <i>et al.</i> [39]
Sphere formation	Tumour initiation	Zhong <i>et al.</i> [29]

bone marrow [49,50]. An increase in the number of CK+ cells in RCC patients correlates with advanced tumour stage in RCC [51]. Bluemke *et al.* found two types of CTCs in the peripheral blood of RCC patients: cytokeratin (CK+) expressing cells and cells without cytokeratin expression, which are large with tumour-like morphology [49]. Only CK+ cells statistically correlated with the poor overall survival of RCC patients. Similar results have been reported in earlier studies [52,53].

In one study conducted with non-metastatic RCC patients, no prognostic relevance was found for disseminated cytokeratin-positive (CK+) cells in the bone marrow [54]. However, in another study, Buchner *et al.* [50] found the prognostic values of disseminated CK+ cells in the bone marrow of patients with metastatic RCC indicated that these cells played an important role in tumour spread in metastatic RCC. The researchers also found that immunocytochemical detection of these cells could be useful in the assessment of clinical outcomes in patients. However, further characterization of the cells is necessary to evaluate their malignant potential as a relevant therapeutic target for novel systematic therapy. Recently, El-Heiliebi *et al.* used a new blood filtration technique to distinguish circulating non-hematologic cells (CNHCs) as CTCs in the blood of RCC patients based on cytomorphological criteria [55].

Gene Expression Pattern in the Development of Metastasis RCC

Gene expression analysis has been widely adopted in clinical and laboratory research to discern genomic background and identify genes that might serve as a prognostic biomarker in metastatic RCC. Identifying such genes could serve to delineate novel targets for the invention of specific anti-cancer drugs. In a recent study, gene expression profiling was performed by comparing microarrays on samples from metastatic and primary RCC patients with other patients' normal kidney tissue [56]. There were 95 gene sets significantly up-regulated in metastatic RCC. The majority of up-regulated genes in these sets were those responsible for DNA replication, cell cycle control, apoptosis and cell mortality. For instance, genes from the minichromosome maintenance gene family (*MCM2*, *MCM4*,

MCM6 and *MCM7*), *AURKA*-coding for a kinase, involved cell cycle regulation and tumour metastasis progression, as well as *FEN1* in DNA replication. Other researchers used a different approach to analyse the gene expression profiles based on stepwise progression and metastasis in RCC [57]. Samples from three different progressive sites of RCC were compared—Kidney tissue (N), through early tumour stage (T1) to distant metastasis (M). *Caveolin 1*, *annexin A4* and *lysyl oxidase* were continuously deregulated on progression, indicating that these genes might be important in driving the tumour toward increased malignancy and metastasis [57].

In a study of 58 cases, Vasselli *et al.* identified a gene signature between the primary tumour and one with metastasis RCC [58]. Forty-five genes were consistently up- or down-regulated and associated with survival using the Cox proportional hazards model. In addition, vascular cell adhesion molecule 1 (*VCAM-1*) was revealed to be the most likely biomarker predicting survival of RCC patients. The higher expression for the *VCAM-1* gene was significantly associated with longer survival compared to low *VCAM-1* expression. Others have reported higher expression of *CYR1* and *LDB2* genes associated with disease-free survival with higher expression in tumours that metastasized after 24 months compared to tumours that metastasized earlier [59]. In a different approach, two cell lines derived from a matched primary tumour and adrenal metastasis from the same RCC patients were analysed using similar technology [60]. *EGFR*, *cadherin-6* and *vimentin* expression were higher in the metastatic cell line. Furthermore, those highly expressed genes were associated with growth factors, cell division, signal transduction and cell-cell adhesion function. Bockhorn *et al.* analysed the gene expression of cells shedding from two RCC cell lines with different metastatic potential [61]. Their analysis revealed that 23 deregulated genes were involved in metastasis. However, only *caveolin-1*, *CD44* and *α 3-integrin* expression were down regulated in cells shed from both cell lines. These expressions might be important in faster migration of the tumour cells and give shed cells a metastatic advantage.

Molecular differences between tumour subtypes of RCC have been largely unknown. In a study of 112 RCC and normal kidney samples, researchers distinguished the gene

expression pattern between three major types of RCC: clear cell RCC (ccRCC), papillary RCC (pRCC) and chromophobe RCC (chRCC) [62]. Gene expression was widely correlated with RCC subtype. Down regulation of genes was reported in pRCC and chRCC compared with ccRCC. These genes were responsible for metabolism (*GAPD*), angiogenesis (*ANGPTL4* and *VEGF*), cell adhesion (*COL3A1* and *FNI*) and immune response (*IGHG3* and *HLA-DRBI*). Up-regulation was reported in genes for osteopontin (*SPPI*) and retinol binding protein 4 (*RBP4*). Moreover, primary tumour gene expression delineated 12 such genes that were associated with tumour metastasis formation and patient survival. Among them, the most significantly deregulated genes were the human high-mobility group gene (*HMGAI*), mitochondrial dienyol-CoA reductase (*DECRI*), GTP-binding protein (*RAGB*) and genes belonging to the gene families *COL5A1*, *SLC13A3*, *SLC29A2*, *IGFBP3* and *GUCY2C* [62]. In summary, these finding could serve to discriminate clinically meaningful sub classification of RCC and expression pattern approaches to describe metastasis formation in various RCC sub-types.

Genome expression analysis has been widely implicated in cancer research. Extracting relevant biological insight from such genome data sets remains a major challenge. In recent years, the Gene Set Enrichment Analysis (GSEA) method has been purposed to incorporate knowledge from databases with gene expression data set [63]. Maruschke *et al.* used the GSEA approach to analyse the expression profile in ccRCC by comparing poorly and well-differentiated tumour tissue samples [64]. Using comparative analysis, the researchers selected 16 expression gene sets out of 120. Genes in these sets were involved in cell motility, signalling, proliferation and metastasis and were gene products in the building of extracellular matrix and as cell surface markers.

CONCLUSION

In conclusion, this short review of the current literature has shown that the activities of CD105, CXCR4 and ALDH1 represent the most prominent markers for identifying tumour-initiating cells in RCC. These cells are also associated with metastasis initiation and progression. In addition, cells expressing these markers share many common characteristics of the TIC/CSC phenotype, having strong proliferation potential, self-renewal ability, resistance against conventional therapy and the formation of tumours in *in vivo* studies. Gene expression profiling helps to define TICs and might help develop targeted therapeutics with refined diagnostic data for RCC cure. However, current described gene expression studies cannot be employed into clinical application. Therefore, additional confirmation is needed, and future studies might be helpful to identify relevance with the clinical response in RCC.

CONFLICT OF INTEREST

The authors indicate no potential conflict of interest.

ACKNOWLEDGEMENTS

This research was supported by the Military Institute of Medicine statutory founding 1/1744 (101). CS, AMC was supported by the National Science Centre UMO-2011/01/B/NZ5/02822 and 2011/01/B/NZ4/01602 projects. CS, AMC, IMK were supported by the Foundation for Polish Science

TEAM project TEAM/2010-6/8. AMC was also supported by the Ministry of Science and Higher Education "Juventus" grant. The authors acknowledge the support of the Scribendi, Inc. for professional editing and proofreading of this manuscript.

REFERENCES

- [1] Chow TF, Youssef YM, Lianidou E, Romaschin AD, Honey RJ, Stewart R, *et al.* Differential expression profiling of microRNAs and their potential involvement in renal cell carcinoma pathogenesis. *Clinical biochemistry*. 2010; 43(1-2): 150-8. Epub 2009/08/04.
- [2] Ljungberg B. Prognostic factors in renal cell carcinoma. *Der Urologe Ausg A*. 2004; 43 Suppl 3: S119-20. Epub 2004/06/05.
- [3] Leibovich BC, Han KR, Bui MH, Pantuck AJ, Dorey FJ, Figlin RA, *et al.* Scoring algorithm to predict survival after nephrectomy and immunotherapy in patients with metastatic renal cell carcinoma: a stratification tool for prospective clinical trials. *Cancer*. 2003; 98(12): 2566-75. Epub 2003/12/12.
- [4] Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, *et al.* Sorafenib in advanced clear-cell renal-cell carcinoma. *The New England journal of medicine*. 2007; 356(2): 125-34. Epub 2007/01/12.
- [5] Buczek M, Escudier B, Bartnik E, Szczylik C, Czarnecka A. Resistance to tyrosine kinase inhibitors in clear cell renal cell carcinoma: From the patient's bed to molecular mechanisms. *Biochimica et biophysica acta*. 2013. Epub 2013/10/19.
- [6] Czarnecka AM, Solarek W. The activity of tyrosine kinase inhibitors on clear cell renal cell carcinoma tumor initiating cells in hypoxic microenvironment. In: BJUI, editor. *The 11th International Kidney Cancer Symposium 5-6 October 2012*; Chicago, Illinois, USA: BJU International; 2012. p. 1.
- [7] Motzer RJ, Hutson TE, Cella D, Reeves J, Hawkins R, Guo J, *et al.* Pazopanib versus sunitinib in metastatic renal-cell carcinoma. *The New England journal of medicine*. 2013; 369(8): 722-31. Epub 2013/08/24.
- [8] Sternberg CN, Hawkins RE, Wagstaff J, Salman P, Mardiak J, Barrios CH, *et al.* A randomised, double-blind phase III study of pazopanib in patients with advanced and/or metastatic renal cell carcinoma: final overall survival results and safety update. *Eur J Cancer*. 2013; 49(6): 1287-96. Epub 2013/01/17.
- [9] Rini BI, Escudier B, Tomczak P, Kaprin A, Szczylik C, Hutson TE, *et al.* Comparative effectiveness of axitinib versus sorafenib in advanced renal cell carcinoma (AXIS): a randomised phase 3 trial. *Lancet*. 2011; 378(9807): 1931-9. Epub 2011/11/08.
- [10] Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, *et al.* Identification of a cancer stem cell in human brain tumors. *Cancer research*. 2003; 63(18): 5821-8. Epub 2003/10/03.
- [11] Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100(7): 3983-8. Epub 2003/03/12.
- [12] Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ. Prospective identification of tumorigenic prostate cancer stem cells. *Cancer research*. 2005; 65(23): 10946-51. Epub 2005/12/03.
- [13] Bapat SA, Mali AM, Koppikar CB, Kurrey NK. Stem and progenitor-like cells contribute to the aggressive behavior of human epithelial ovarian cancer. *Cancer research*. 2005; 65(8): 3025-9. Epub 2005/04/19.
- [14] Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, *et al.* Identification and expansion of human colon-cancer-initiating cells. *Nature*. 2007; 445(7123): 111-5. Epub 2006/11/24.
- [15] Haraguchi N, Utsunomiya T, Inoue H, Tanaka F, Mimori K, Barnard GF, *et al.* Characterization of a side population of cancer cells from human gastrointestinal system. *Stem Cells*. 2006; 24(3): 506-13. Epub 2005/10/22.
- [16] Li C, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V, *et al.* Identification of pancreatic cancer stem cells. *Cancer research*. 2007; 67(3): 1030-7. Epub 2007/02/07.
- [17] Prince ME, Sivanandan R, Kaczorowski A, Wolf GT, Kaplan MJ, Dalerba P, *et al.* Identification of a subpopulation of cells with cancer stem cell properties in head and neck squamous cell carcinoma. *Proceedings of the National Academy of Sciences of the United States of America*. 2007; 104(3): 973-8. Epub 2007/01/11.

- [18] Yang ZF, Ngai P, Ho DW, Yu WC, Ng MN, Lau CK, *et al.* Identification of local and circulating cancer stem cells in human liver cancer. *Hepatology*. 2008; 47(3): 919-28. Epub 2008/02/16.
- [19] Fang D, Nguyen TK, Leishear K, Finko R, Kulp AN, Hotz S, *et al.* A tumorigenic subpopulation with stem cell properties in melanomas. *Cancer research*. 2005; 65(20): 9328-37. Epub 2005/10/19.
- [20] Baccelli I, Trumpp A. The evolving concept of cancer and metastasis stem cells. *The Journal of cell biology*. 2012; 198(3): 281-93. Epub 2012/08/08.
- [21] Bussolati B, Brossa A, Camussi G. Resident stem cells and renal carcinoma. *International journal of nephrology*. 2011; 2011: 286985. Epub 2011/06/08.
- [22] Pode-Shakked N, Metsuyanim S, Rom-Gross E, Mor Y, Fridman E, Goldstein I, *et al.* Developmental tumorigenesis: NCAM as a putative marker for the malignant renal stem/progenitor cell population. *Journal of cellular and molecular medicine*. 2009; 13(8B): 1792-808. Epub 2010/02/27.
- [23] Khan MI, Czarnecka AM, Krol M, Zdanowski R, Sobocinska A, Lewicki S, *et al.* Analysis of Tumour Initiating Cells (TICs) populations in primary and metastatic cell lines from clear cell Renal Cell Carcinoma (ccRCC). In: Singh SM, editor. 4th International Conference on Stem Cells and Cancer (ICSCC-2013): Proliferation, Differentiation and Apoptosis; 19-22 October 2013; Mumbai, India: ICSCCB; 2013. p. 73-4.
- [24] Gassenmaier M, Chen D, Buchner A, Henkel L, Schiemann M, Mack B, *et al.* CXCR4 Chemokine Receptor 4 is Essential for Maintenance of Renal cell Carcinoma-Initiating Cells and Predicts Metastasis. *Stem Cells*. 2013; 31(8): 1467-76. Epub 2013/05/01.
- [25] Bussolati B, Bruno S, Grange C, Ferrando U, Camussi G. Identification of a tumor-initiating stem cell population in human renal carcinomas. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 2008; 22(10): 3696-705. Epub 2008/07/11.
- [26] Khan MI, Czarnecka AM, Szczylik C. Detection of CD105+ and CD133+ sub-populations (cancer initiating cells) in SKMT-R2, SKMT-R3 and 786-O human primary renal cancer cell lines. In: Wiersdorff V, editor. The 4th EMBO meeting; 22-25 September 2012; Nice, France: The 4th EMBO meeting; 2012. p. 212-3.
- [27] Sandlund J, Hedberg Y, Bergh A, Grankvist K, Ljungberg B, Rasmuson T. Endoglin (CD105) expression in human renal cell carcinoma. *BJU international*. 2006; 97(4): 706-10. Epub 2006/03/16.
- [28] Addla SK, Brown MD, Hart CA, Ramani VA, Clarke NW. Characterization of the Hoechst 33342 side population from normal and malignant human renal epithelial cells. *American journal of physiology Renal physiology*. 2008; 295(3): F680-7. Epub 2008/07/11.
- [29] Zhong Y, Guan K, Guo S, Zhou C, Wang D, Ma W, *et al.* Spheres derived from the human SK-RC-42 renal cell carcinoma cell line are enriched in cancer stem cells. *Cancer letters*. 2010; 299(2): 150-60. Epub 2010/09/18.
- [30] Huang B, Huang YJ, Yao ZJ, Chen X, Guo SJ, Mao XP, *et al.* Cancer stem cell-like side population cells in clear cell renal cell carcinoma cell line 769P. *PLoS one*. 2013; 8(7): e68293. Epub 2013/07/23.
- [31] Golebiewska A, Brons NH, Bjerkvig R, Niclou SP. Critical appraisal of the side population assay in stem cell and cancer stem cell research. *Cell stem cell*. 2011; 8(2): 136-47. Epub 2011/02/08.
- [32] Nishizawa S, Hirohashi Y, Torigoe T, Takahashi A, Tamura Y, Mori T, *et al.* HSP DNAJB8 controls tumor-initiating ability in renal cancer stem-like cells. *Cancer research*. 2012; 72(11): 2844-54. Epub 2012/05/04.
- [33] Czarnecka AM, Matak D, Solarek W, Khan MI, Szczylik C. Hypoxia response regulates clear cell renal cell carcinoma tumor initiating cells. In: BJUI, editor. 12th International Kidney Cancer Symposium; 25-26 October 2013; Chicago, Illinois, USA: BJU International; 2013. p. 12.
- [34] Bruno S, Bussolati B, Grange C, Collino F, Graziano ME, Ferrando U, *et al.* CD133+ renal progenitor cells contribute to tumor angiogenesis. *The American journal of pathology*. 2006; 169(6): 2223-35. Epub 2006/12/07.
- [35] D'Alterio C, Cindolo L, Portella L, Polimeno M, Consales C, Riccio A, *et al.* Differential role of CD133 and CXCR4 in renal cell carcinoma. *Cell Cycle*. 2010; 9(22): 4492-500. Epub 2010/12/04.
- [36] Zagzag D, Krishnamachary B, Yee H, Okuyama H, Chiriboga L, Ali MA, *et al.* Stromal cell-derived factor-1alpha and CXCR4 expression in hemangioblastoma and clear cell-renal cell carcinoma: von Hippel-Lindau loss-of-function induces expression of a ligand and its receptor. *Cancer research*. 2005; 65(14): 6178-88. Epub 2005/07/19.
- [37] Gelmini S, Mangoni M, Serio M, Romagnani P, Lazzeri E. The critical role of SDF-1/CXCR4 axis in cancer and cancer stem cells metastasis. *Journal of endocrinological investigation*. 2008; 31(9): 809-19. Epub 2008/11/11.
- [38] Djafarzadeh R, Noessner E, Engelmann H, Schendel DJ, Notohamiprodjo M, von Lutichau I, *et al.* GPI-anchored TIMP-1 treatment renders renal cell carcinoma sensitive to FAS-mediated killing. *Oncogene*. 2006; 25(10): 1496-508. Epub 2005/11/02.
- [39] Lu J, Cui Y, Zhu J, He J, Zhou G, Yue Z. Biological characteristics of Rh123 stem-like cells in a side population of 786-O renal carcinoma cells. *Oncology letters*. 2013; 5(6): 1903-8. Epub 2013/07/09.
- [40] Challen GA, Little MH. A side order of stem cells: the SP phenotype. *Stem Cells*. 2006; 24(1): 3-12. Epub 2006/02/02.
- [41] Wagner-Souza K, Diamond HR, Ornellas MH, Gomes BE, Almeida-Oliveira A, Abdelhay E, *et al.* Rhodamine 123 efflux in human subpopulations of hematopoietic stem cells: comparison between bone marrow, umbilical cord blood and mobilized peripheral blood CD34+ cells. *International journal of molecular medicine*. 2008; 22(2): 237-42. Epub 2008/07/19.
- [42] Touil Y, Zulfiani T, Wolowczuk I, Kuranda K, Prochazkova J, Andrieux J, *et al.* The PI3K/AKT signaling pathway controls the quiescence of the low-Rhodamine123-retention cell compartment enriched for melanoma stem cell activity. *Stem Cells*. 2013; 31(4): 641-51. Epub 2013/01/29.
- [43] Ueda K, Ogasawara S, Akiba J, Nakayama M, Todoroki K, Sanada S, *et al.* Aldehyde dehydrogenase 1 identifies cells with cancer stem cell-like properties in a human renal cell carcinoma cell line. *PLoS one*. 2013; 8(10): e75463. Epub 2013/10/12.
- [44] Jacob K, Sollier C, Jabado N. Circulating tumor cells: detection, molecular profiling and future prospects. *Expert review of proteomics*. 2007; 4(6): 741-56. Epub 2007/12/11.
- [45] Gradilone A, Iacovelli R, Cortesi E, Raimondi C, Gianni W, Nicolazzo C, *et al.* Circulating tumor cells and "suspicious objects" evaluated through CellSearch(R) in metastatic renal cell carcinoma. *Anticancer research*. 2011; 31(12): 4219-21. Epub 2011/12/27.
- [46] Braun S, Pantel K, Muller P, Janni W, Hepp F, Kantenich CR, *et al.* Cytokeratin-positive cells in the bone marrow and survival of patients with stage I, II, or III breast cancer. *The New England journal of medicine*. 2000; 342(8): 525-33. Epub 2000/02/24.
- [47] Partridge M, Brakenhoff R, Phillips E, Ali K, Francis R, Hooper R, *et al.* Detection of rare disseminated tumor cells identifies head and neck cancer patients at risk of treatment failure. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2003; 9(14): 5287-94. Epub 2003/11/14.
- [48] Wollenberg B, Walz A, Kolbow K, Pauli C, Chaubal S, Andratschke M. Clinical relevance of circulating tumour cells in the bone marrow of patients with SCCHN. *Onkologie*. 2004; 27(4): 358-62. Epub 2004/09/07.
- [49] Bluemke K, Bilkenroth U, Meyer A, Fuessel S, Lautenschlaeger C, Goebel S, *et al.* Detection of circulating tumor cells in peripheral blood of patients with renal cell carcinoma correlates with prognosis. *Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology*. 2009; 18(8): 2190-4. Epub 2009/08/08.
- [50] Buchner A, Riesenberger R, Kotter I, Hofstetter A, Stief C, Oberneder R. Frequency and prognostic relevance of disseminated tumor cells in bone marrow of patients with metastatic renal cell carcinoma. *Cancer*. 2006; 106(7): 1514-20. Epub 2006/03/07.
- [51] Bilkenroth U, Taubert H, Riemann D, Rebmann U, Heynemann H, Meyer A. Detection and enrichment of disseminated renal carcinoma cells from peripheral blood by immunomagnetic cell separation. *International journal of cancer Journal international du cancer*. 2001; 92(4): 577-82. Epub 2001/04/17.
- [52] Ashida S, Okuda H, Chikazawa M, Tanimura M, Sugita O, Yamamoto Y, *et al.* Detection of circulating cancer cells with von hippel-lindau gene mutation in peripheral blood of patients with renal cell carcinoma. *Clinical cancer research : an official journal*

- of the American Association for Cancer Research. 2000; 6(10): 3817-22. Epub 2000/10/29.
- [53] Shimazui T, Yoshikawa K, Uemura H, Kawamoto R, Kawai K, Uchida K, *et al.* Detection of cadherin-6 mRNA by nested RT-PCR as a potential marker for circulating cancer cells in renal cell carcinoma. *International journal of oncology*. 2003; 23(4): 1049-54. Epub 2003/09/10.
- [54] Buchner A, Riesenberger R, Kotter I, Crispin A, Hofstetter A, Oberneder R. Detection and prognostic value of cytokeratin positive tumor cells in bone marrow of patients with renal cell carcinoma. *The Journal of urology*. 2003; 170(5): 1747-51. Epub 2003/10/09.
- [55] El-Heliebi A, Kroneis T, Zohrer E, Haybaeck J, Fischereder K, Kappel-Kettner K, *et al.* Are morphological criteria sufficient for the identification of circulating tumor cells in renal cancer? *Journal of translational medicine*. 2013; 11(1): 214. Epub 2013/09/21.
- [56] Maruschke M, Hakenberg OW, Koczan D, Zimmermann W, Stief CG, Buchner A. Expression profiling of metastatic renal cell carcinoma using gene set enrichment analysis. *International journal of urology : official journal of the Japanese Urological Association*. 2013. Epub 2013/05/03.
- [57] Jones J, Otu H, Spentzos D, Kolia S, Inan M, Beecken WD, *et al.* Gene signatures of progression and metastasis in renal cell cancer. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2005; 11(16): 5730-9. Epub 2005/08/24.
- [58] Vasselli JR, Shih JH, Iyengar SR, Maranchie J, Riss J, Worrell R, *et al.* Predicting survival in patients with metastatic kidney cancer by gene-expression profiling in the primary tumor. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100(12): 6958-63. Epub 2003/06/05.
- [59] Wozniak MB, Le Calvez-Kelm F, Abedi-Ardekani B, Byrnes G, Durand G, Carreira C, *et al.* Integrative genome-wide gene expression profiling of clear cell renal cell carcinoma in Czech Republic and in the United States. *PloS one*. 2013; 8(3): e57886. Epub 2013/03/26.
- [60] Ohno Y, Izumi M, Tachibana M, Kawamura T, Yoshioka K, Aoyagi T, *et al.* Characterization and gene expression analysis of novel matched primary and metastatic renal cell carcinoma cell lines. *Oncology reports*. 2008; 20(3): 501-9. Epub 2008/08/13.
- [61] Bockhorn M, Roberge S, Sousa C, Jain RK, Munn LL. Differential gene expression in metastasizing cells shed from kidney tumors. *Cancer research*. 2004; 64(7): 2469-73. Epub 2004/04/03.
- [62] Sultmann H, von Heydebreck A, Huber W, Kuner R, Buness A, Vogt M, *et al.* Gene expression in kidney cancer is associated with cytogenetic abnormalities, metastasis formation, and patient survival. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2005; 11(2 Pt 1): 646-55. Epub 2005/02/11.
- [63] Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, *et al.* Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences of the United States of America*. 2005; 102(43): 15545-50. Epub 2005/10/04.
- [64] Maruschke M, Reuter D, Koczan D, Hakenberg OW, Thiesen HJ. Gene expression analysis in clear cell renal cell carcinoma using gene set enrichment analysis for biostatistical management. *BJU international*. 2011; 108(2 Pt 2): E29-35. Epub 2011/03/26.