

Article

# Synthesis and Antimicrobial Activity of 4-Substituted 1,2,3-Triazole-Coumarin Derivatives

Priscila López-Rojas <sup>1</sup>, Monika Janeczko <sup>2</sup>, Konrad Kubiński <sup>2</sup>, Ángel Amesty <sup>1,\*</sup>, Maciej Masłyk <sup>2,\*</sup> and Ana Estévez-Braun <sup>1,\*</sup>

- <sup>1</sup> Instituto Universitario de Bio-Orgánica Antonio González (CIBICAN), Departamento de Química Orgánica, Universidad de La Laguna, Avda. Astrofísico Fco. Sánchez 2, 38206 La Laguna, Tenerife, Spain; priscila.quim@gmail.com
- <sup>2</sup> Department of Molecular Biology, The John Paul II Catholic University of Lublin, ul. Konstantynów 1i, 20-708 Lublin, Poland; mjanec@kul.pl (M.J.); kubin@kul.pl (K.K.)
- \* Correspondence: aarnesty@ull.es (Á.A.); maciekm@kul.pl (M.M.); aestebra@ull.es (A.E.-B.)

Received: 18 December 2017; Accepted: 15 January 2018; Published: 18 January 2018

**Abstract:** A new series of coumarin-1,2,3-triazole conjugates with varied alkyl, phenyl and heterocycle moieties at C-4 of the triazole nucleus were synthesized using a copper(I)-catalysed Huisgen 1,3-dipolar cycloaddition reaction of corresponding *O*-propargylated coumarin (**3**) or *N*-propargylated coumarin (**6**) with alkyl or aryl azides. Based on their minimal inhibitory concentrations (MICs) against selected microorganisms, six out of twenty-six compounds showed significant antibacterial activity towards *Enterococcus faecalis* (MIC = 12.5–50 µg/mL). Moreover, the synthesized triazoles show relatively low toxicity against human erythrocytes.

Keywords: triazoles; coumarin; antibacterial activity; Enterococcus faecalis; biofilm

# 1. Introduction

Antimicrobial resistance has been listed by the World Health Organization (WHO) as one of the biggest threats to global health today [1]. The antibiotic resistance crisis has been attributed to the overuse and misuse of these medications, as well as a lack of new drug development by the pharmaceutical industry due to reduced economic incentives and challenging regulatory requirements [2–6]. Over the past decade, it has become apparent that several highly resistant bacterial pathogens have acquired clever mechanisms to negate the effectiveness of numerous therapeutic agents [7]. Staphylococcus aureus is one bacterial pathogen that has emerged as a significant concern to healthcare professionals worldwide. In this sense, isolated strains of S. aureus have exhibited resistance to several classes of antibacterial drugs, including  $\beta$ -lactam antibiotics [8], macrolides [9], fluoroquinolones [10–12], glycopeptides [13] and oxazolidinones [14]. Enterococci were previously considered commensal organisms of little clinical importance but have emerged as serious nosocomial pathogens responsible for e.g. endocarditis and infections of the urinary tract, bloodstream, meninges, wounds and the biliary tract [15]. Recent surveillance data indicate that Enterococcus is the third most commonly isolated nosocomial pathogen (12% of all hospital infections), only behind coagulase-negative Staphylococcus and Staphylococcus aureus [16]. The clinical importance of the genus Enterococcus is directly related to its antibiotic resistance, which contributes to the risk of colonization and infection. *Enterococci* are intrinsically resistant to many commonly used antimicrobial agents (penicillins, ampicillins, cephalosporins, clindamycin) and exhibit native resistance to clinically achievable concentrations of aminoglycosides. Although E. faecalis is naturally resistant to quinupristin-dalfopristin, this combination is highly active against *E. faecium* strains that lack specific resistance determinants. *Enterococci* are tolerant to the (normally) bactericidal activity of cell-wall active agents, such as  $\beta$ -lactam antibiotics and vancomycin. Tolerance implies that the



All the information mentioned above highlights the urgent need to develop novel antibacterial agents devoid of cross-resistance to marketed antibiotics.

The use of privileged structures in drug discovery has proven to be an effective strategy allowing the generation of innovative hits/leads and successful optimization processes [18,19]. Coumarins are considered to be privileged structures due to their broad range of biological properties including anticoagulant [20], anti-neurodegenerative [21], antioxidant [22], anticancer [23] and antimicrobial activities [24–28]. These interesting properties of coumarins can be ascribed to the chemical attributes of the 2*H*-chromen-2-one core; its aromatic ring can establish a series of hydrophobic,  $\pi$ – $\pi$ , CH– $\pi$  and cation– $\pi$  interactions and the two oxygen atoms in the lactone ring can hydrogen-bond to a series of amino acid residues in different classes of enzymes and receptors. Additionally, the double bond in the lactone helps to make the planar system, allows charge delocalization between the carbonyl group of the lactone and the aromatic ring and confers the characteristic fluorescence of this class of compounds.

On the other hand, 1,2,3-triazoles are nitrogen heterocycles capable of forming hydrogen bonds, which improves their solubility and ability to interact with biomolecular targets [29]. The 1,2,3-triazoles are highly stable to metabolic degradation, compared to other compounds containing three adjacent nitrogen (N) atoms [29]. The triazoles have been used for broad therapeutic applications due to their diverse biological activities [30], i.e. antimicrobial [31–33], antiviral [34], anti-inflammatory [35], analgesic [35], anticancer [36–38], antifungal [39] and anticonvulsant [40] activities.

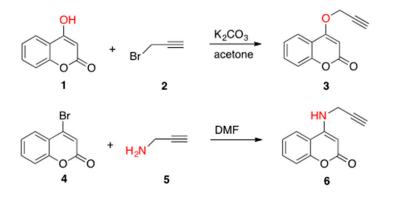
Taking into consideration the antimicrobial activity shown by some coumarins and 1,2,3-triazols mentioned above and as a continuation of our project on searching for new antibacterial molecules [41–45], we envisaged that the linkage of coumarin and 1,2,3-triazole pharmacophores through  $-OCH_2$ - or  $-NCH_2$ - linkers would generate novel hybrid molecules with promising antibacterial activities.

Therefore, we herein report the synthesis and antibacterial activity of coumarin-1,2,3-triazole conjugates with varied alkyl, phenyl and heterocycle moieties at C-4 of the triazole nucleus in order to evaluate their contribution to the antimicrobial activity.

#### 2. Results and Discussion

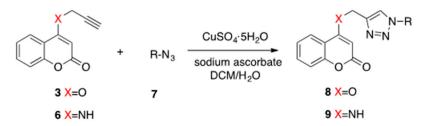
#### 2.1. Chemistry

The required acetylenic dipolarophiles **3** and **6** were obtained as shown in Scheme **1**. Thus, the treatment of 4-hydroxy-coumarin (**1**) with 3-bromoprop-1-yne (**2**) employing potassium carbonate in anhydrous acetone yielded the *O*-propargylated coumarin (**3**) in 55% yield. The *N*-propargylated coumarin (**6**) was obtained in 63% yield from 4-bromo-coumarin (**4**) through nucleophilic substitution with prop-2-yn-1-amine (**5**) in dimethylformamide (DMF).



Scheme 1. Formation of O-propargylated coumarin (3) and N-propargylated coumarin (6).

The 4-substituted 1,2,3-triazole-coumarin derivatives were synthesized using a copper(I)-catalysed Huisgen 1,3-dipolar cycloaddition reaction [46] of the corresponding *O*-propargylated coumarin (**3**) or *N*-propargylated coumarin (**6**) with alkyl or aryl azides (Scheme 2).



Scheme 2. Formation of 4-substituted 1,2,3-triazole-coumarin derivatives 8 and 9.

Azides **7a–7j** were prepared from the corresponding boronic acids and sodium azide, in the presence of CuSO<sub>4</sub> in ethanol (EtOH), at room temperature (Table 1 and Scheme 3) [47,48].

Ar-B(OH)<sub>2</sub> + NaN<sub>3</sub> 
$$\xrightarrow{\text{CuSO}_4}$$
 Ar-N<sub>3</sub>

Scheme 3. Preparation of azides 7a-7j.

Entry	Azide	Ar	Yield <sup>a</sup>
1	7a	Ph	58
2	7b	2-OMe-Ph	61
3	7c 3-OMe–Ph		97
4	7d	4-OMe-Ph	96
5	7e	3-F, 4-OMe–Ph	96
6	7f	4-F–Ph	41
7	7g	3-CF <sub>3</sub> -Ph	31
8	7h	3-NO <sub>2</sub> -Ph	82
9	7i	5-1H-indol	35
10	7j	3-furyl	_ b

Table 1. Synthesized azides 7a–7j.

<sup>a</sup> Isolated yield; <sup>b</sup> It was not isolated but was reacted in situ.

Azides **7k–7m** were obtained from the corresponding alkyl bromide or aryl bromide and sodium azide in DMF (Table 2 and Scheme 4) [49,50].

Table 2. Synthesized azides 7k–7m.

Entry	Azide	R	Yield <sup>a</sup>	
1	7k	(CH <sub>2</sub> ) <sub>10</sub> CH <sub>3</sub>	86	
2	71	(CH <sub>2</sub> ) <sub>13</sub> CH <sub>3</sub>	73	
3	7m	CH <sub>2</sub> Ph	82	

<sup>a</sup> Isolated yield; <sup>b</sup> MW irradiation at 120 °C for 35 min was used.

R-Br + NaN<sub>3</sub> 
$$\xrightarrow{\text{DMF}}$$
 R-N<sub>3</sub>

Scheme 4. Preparation of azides 7k–7m.

Following the reaction shown in Scheme 2 and using azides **7a–7m**, compounds **8a–8m** and **9a–9m** were obtained as illustrated in Figure 1.

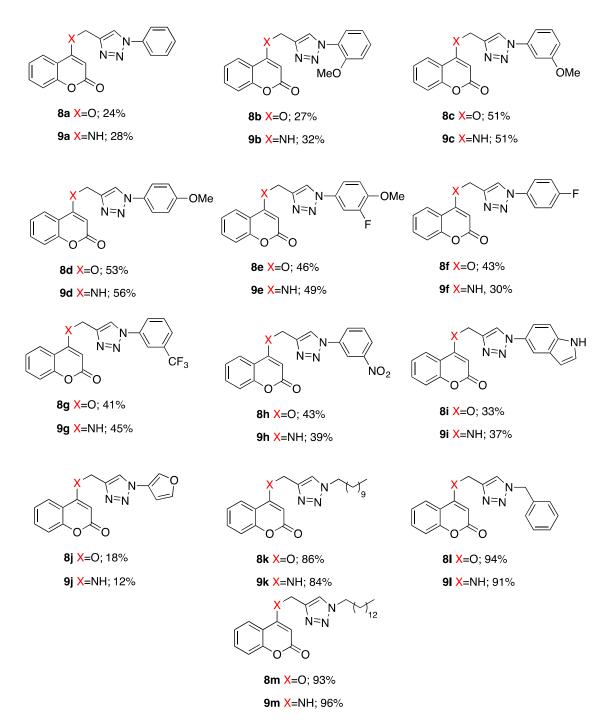


Figure 1. Structures of 4-substituted 1,2,3-triazole-coumarin derivatives (8a-8n) and (9a-9n).

As can be seen, two isosteric series of coumarin derivatives were obtained (X=O, X=NH). Each series presents different substituents at the triazole moiety in order to evaluate their influence on the antimicrobial activity. Thus, coumarin derivatives with an aromatic ring having electron-donating groups or electron-withdrawing groups were prepared (8a–8h, 9a–9h). Coumarin-triazole derivatives with alkyl moieties (8k, 9k, 8l, 9l) and coumarin-indole hybrids (8i, 9i) were synthesized as well. Moderate yields were obtained with aromatic azides while the use of the more stable aliphatic azides (7k, 7l and 7m) led to high yields, in agreement with the more favourable HOMO of the dipole in the 1,3-dipolar cycloaddition. The structures of all adducts were determined by spectroscopic studies. All of them showed the characteristic proton of the triazol ring in the <sup>1</sup>H-NMR spectral region between

 $\delta$  7.63 and 9.31. The hydrogen of the coumarin nucleus was detected as a singlet at  $\delta$  5.18–6.22 and the methylene hydrogens in the oxygenated series appeared as a singlet at  $\delta$  5.35–5.59 and as a doublet at  $\delta$  4.66–4.46 (*J* = 5.6 Hz) in the nitrogenated series.

The best yields were obtained from the *N*-propargylated coumarin (6) and from the aliphatic azides (7k, 7l and 7m).

## 2.2. Biology

Since some coumarins and 1,2,3-triazoles have shown potential as antibacterial drugs [41–45,51], these combined pharmacophores could offer some advantages e.g. in overcoming drug resistance as well as improving their biological potency.

The in vitro antimicrobial activity of the novel coumarin-1,2,3-triazole conjugates was tested against the yeast *Candida albicans*, Gram-positive bacteria *Staphylococcus aureus* and *Enterococcus faecalis* and Gram-negative bacteria *Escherichia coli*, *Klebsiella pneumonia* and *Pseudomonas aeruginosa*. The minimum inhibitory concentrations (MICs) were determined and given in Table 3. As can be seen, most of the coumarin-triazole hybrids did not exhibit considerable activity against the tested microorganisms. The best results were obtained with conjugates **8a**, **8b**, **8f**, **9h** and **9k**, which displayed promising activity against *Enterococcus faecalis* at MICs ranging from 12.5 to 50.0 µg/mL. Compound **8b** having a 2-OMe–Ph group attached at the triazol nucleus and an –OCH<sub>2</sub>– linker was the best of the series, while the corresponding isoster **9b** (–NHCH<sub>2</sub>–) turned out to be 64-fold less active than **8b**. The position of the OMe group in the phenyl ring also plays an important role in the activity, since compounds **8c** (3-OMe–Ph) and **8d** (4-OMe–Ph) showed an 8- and 16-fold lower antibacterial activity, respectively, than **8b**. In the nitrogenated series, compounds **9h** (3-NO<sub>2</sub>–Ph) and **9k** having an undecyl chain showed the best activities.

Compound	Candida albicans	Staphylococcus aureus	Enterococcus faecalis	Escherichia coli	Klebsiella pneumoniae	Pseudomona aeruginosa
8a	1600	400	50	1600	-	1600
8b	800	200	12.5	1600	1600	1600
8c	1600	800	100	1600	800	1600
8d	-	-	200	1600	800	1600
8e	400	400	100	200	800	800
8f	1600	1600	50	1600	400	800
8g	800	1600	100	800	400	800
8ĥ	800	400	400	1600	800	1600
8i	1600	1600	200	1600	1600	1600
8j	1600	800	800	1600	1600	1600
8k	-	-	400	1600	1600	1600
81	-	-	400	1600	1600	800
9a	1600	1600	400	1600	1600	-
9b	-	1600	800	1600	800	1600
9c	200	800	400	1600	1600	800
9d	200	400	100	800	800	800
9e	200	200	100	800	800	800
9f	200	-	1600	1600	1600	-
9g	-	200	-	-	-	-
9h	1600	100	50	800	800	800
9i	1600	1600	100	1600	800	1600
9j	-	800	800	1600	1600	-
9k	1600	200	50	-	-	-
91	1600	-	800	1600	1600	1600
CAM	n.d.	5	5	1.2	5	5
KET	8	n.d.	n.d.	n.d.	n.d.	n.d.

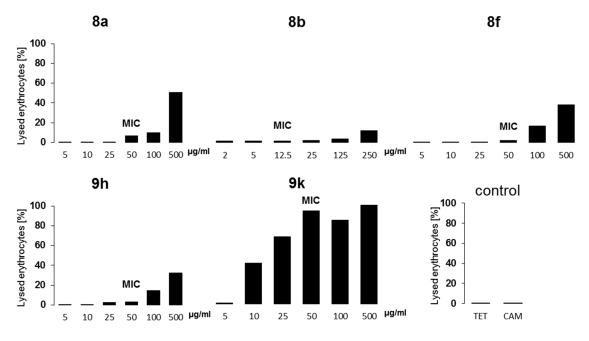
Table 3. Antimicrobial activity (MIC  $\mu$ g/mL) of the synthesized compounds.

no activity; n.d.-not determined, CAM-chloramphenicol; KET-ketoconazole.

In other studies, menthyl 1,4-disubstituted 1,2,3-triazole derivatives of hydroxybenzaldehydes, phenols and bile acids showed a strong inhibitory effect against *E. faecium* with the minimum inhibitory concentration (MIC) values in the range of  $1-3 \mu M$  [52]. Kant and co-workers reported

that 1,2,3-triazole linked chalcone and flavone hybrids showed activity against Gram-positive bacteria (*Staphylococcus aureus*, *Enterococcus faecalis*) and Gram-negative bacteria (*Escherichia coli*, *Pseudomonas aeruginosa*, *Shigella boydii*, *Klebsiella pneumoniae*) with MIC values in the range of 6.25–100  $\mu$ g/mL [53]. In turn, 1,2,4-triazolo[3,4a]phthalazine derivatives showed inhibitory activity against *Staphylococcus aureus* (MIC 16–128  $\mu$ g/mL) [54].

In order to verify if the newly synthesized triazoles could be considered as potential antimicrobial therapeutics, the most active compounds, namely **8a**, **8b**, **8f**, **9h** and **9k** were examined in terms of their haemolytic activity against human erythrocytes. The results are shown in Figure 2.



**Figure 2.** Haemolytic activity of compounds **8a**, **8b**, **8f**, **9h** and **9k**. Tetracycline (TET) and chloramphenicol (CAM) in MIC concentrations were used as a control.

Compounds **8b**, **8f** and **9h** exhibit minimal toxicity towards human blood cells (1.6–3.1% of lysed cells) in MIC. Although compound **8b** appears to be the most active antimicrobial agent, simultaneously it moderately affects the erythrocytes (6.9% of lysed cells) in MIC. The presence of an undecyl chain in the triazole ring (**9k**) results in a drastic increase in the haemolytic activity (94% of lysed cells) in MIC.

## 3. Materials and Methods

## 3.1. Compounds Synthesis

## 3.1.1. General Experimental Procedures

IR spectra were obtained using a Fourier Transform Infrared spectrometer. NMR spectra were recorded in CDCl<sub>3</sub> or DMSO at 500 or 600 MHz for <sup>1</sup>H NMR and 125 or 150 MHz for <sup>13</sup>C-NMR. Chemical shifts are given in ( $\delta$ ) parts per million and coupling constants (*J*) in hertz (Hz). <sup>1</sup>H- and <sup>13</sup>C-spectra were referenced using the solvent signal as an internal standard. Melting points were taken on a capillary melting point apparatus and are uncorrected. Microwave reactions were conducted in sealed glass vessels (capacity 5 mL) using a CEM Discover microwave reactor. HREIMS were recorded using a high-resolution magnetic trisector (EBE) mass analyser. The analytical thin-layer chromatography plates used were Polygram-Sil G/UV254. Preparative thin-layer chromatography was carried out with Analtech (Newark, NJ, USA) silica gel GF plates (20 × 20 cm, 1000 Microns) using appropriate mixtures of ethyl acetate and hexanes. All solvents and reagents were purified by standard techniques reported in [55] or used as supplied from commercial sources. All compounds

were named using the ACD40 Name-Pro program, which is based on IUPAC rules. Azides **7a–7m** were synthesized according to procedures previously described in the literature [47–50,56].

4-(*Prop-2-yn-1-yloxy*)-2*H*-chromen-2-one (**3**). 259  $\mu$ L (2.4 mmol) of propargyl bromide were slowly added to a mixture of 330.9 mg (2.0 mmol) of 4-hydroxycoumarin and 552.8 mg (4.0 mmol) of K<sub>2</sub>CO<sub>3</sub> in 15 mL of acetone. The reaction mixture was refluxed for 8 h until disappearance of the starting coumarin. Then, the solvent was eliminated under reduced pressure, 30 mL of H<sub>2</sub>O were added and the mixture was extracted with AcOEt (3 × 30 mL). The organic phases were collected, washed with H<sub>2</sub>O (20 mL) and brine (20 mL) and dried over anhydrous MgSO<sub>4</sub>. After filtration and elimination of the solvent, the crude extract was purified by silica gel column chromatography using DCM as an eluent and 224.7mg (55%) of compound **3** were obtained as an amorphous white solid. Compound **3** showed identical spectroscopic data to those described in the literature [57].

4-(*Prop-2-yn-1-ylamino*)-2*H*-chromen-2-one (**6**). 47 μL (0.72 mmol) of prop-2-yn-1-amine were slowly added to 200 mg (0.89 mmol) of 4-bromocoumarin in 2 mL of dimethylformamide (DMF) under argon atmosphere. The reaction mixture was stirred at room temperature for 18 h. Then water was added and the *N*-propargylated coumarin precipitated. After filtration, 111.8 mg (63%) of compound (**6**) was obtained as an amorphous white solid. m.p. 223–224 °C; <sup>1</sup>H-NMR (600 MHz, (CDCl<sub>3</sub>) δ 7.57 (1H, t, *J* = 8.1 Hz), 7.46 (1H, d, *J* = 8.1 Hz), 7.37 (1H, d, *J* = 8.1 Hz), 7.30 (1H, t, *J* = 8.1 Hz), 5.46 (1H, s), 5.28 (1H, bs), 4.11 (2H, dd, *J* = 5.5, 2.5 Hz), 2.41 (1H, t, *J* = 2.5 Hz); <sup>13</sup>C-NMR (150 MHz, (CDCl<sub>3</sub>) δ 162.5 (C=O), 153.6 (C), 151.8 (C), 132.0 (CH), 123.6 (CH), 119.9 (CH), 118.1 (CH), 113.9 (C), 85.9 (CH), 77.6 (C), 73.6 (CH<sub>2</sub>), 32.9(CH) ppm; EIMS *m*/*z* 199 ([M<sup>+</sup>], 71); 198 (61); 197 (13); 171 (100); 170 (53); 144 (13); 143 (22); 142 (27); 119 (12); 118 (16); 115 (15); 90 (11); 77 (18); 63 (14); 51 (14); HREIMS 199.0638 (calcd. for C<sub>12</sub>H<sub>9</sub>NO<sub>2</sub> [M<sup>+</sup>] 199.0633); FT-IR (ATR) ν<sub>max</sub> 3325, 3259, 3093, 3074, 2934, 2122, 1807, 1668, 1612, 1552, 1484, 1445, 1389, 1354, 1328, 1271, 1192, 1147, 1045, 983, 937, 865, 811 cm<sup>-1</sup>.

3.1.2. General Procedures for the Preparation of 4-Substituted 1,2,3-Triazole-Coumarin Derivatives

*Method A.* Corresponding boronic acid (0.24 mmol) and 78.5 mg (1.2 mmol) of sodium azide in 1.5 mL of H<sub>2</sub>O were added to a vigorously stirred mixture of 3.4 mg (0.0241 mmol) of Cu<sub>2</sub>O in 0.06 mL of 20% of NH<sub>3</sub> and 0.12 mL of H<sub>2</sub>O. The reaction mixture was stirred for 16 h at room temperature under an oxygen atmosphere. Then, 0.14 mmol of propargylated coumarin (**3** or **6**), 8.11 mg (0.041 mmol) of sodium ascorbate, 1.5 mL of H<sub>2</sub>O and 3 mL of acetone were added. The reaction was left at room temperature for 48h. Then, the reaction mixture was extracted with EtOAc. The aqueous phase was acidified with 5% HCl until pH = 2 and extracted with EtOAc (3 × 15 mL). The organic phases were collected, dried over anhydrous MgSO<sub>4</sub> and after elimination of the solvent, the corresponding residue was purified by silica gel CC or TLC-preparative with DCM or 5% DCM/MeOH.

*Method B.* To a solution of 0.28 mmol of the corresponding azide in 3 mL of DCM, 0.14 mmol of propargylated coumarin (**3** or **6**), 3.6 mg (0.02 mmol) of sodium ascorbate, 1.2 mg (0.004 mmol) of  $CuSO_4 \cdot 5H_2O$  and 3 mL of  $H_2O$ , were added. The reaction mixture was stirred for 48 h at room temperature. The reaction mixture was extracted with EtOAc (3 × 15 mL). The organic phases were collected, dried over anhydrous MgSO<sub>4</sub> and after elimination of the solvent, the corresponding residue was purified by silica gel CC or TLC-preparative with DCM or 5% DCM/MeOH.

4-((1-*Phenyl*-1*H*-1,2,3-*triazol*-4-*yl*)*methoxy*)-2*H*-*chromen*-2-*one* (**8a**). Following the experimental procedure described in method A, from 31.2 mg (0.24 mmol) of phenyl boronic acid and 28.0 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 10.9 mg (24%) of compound **8a** were obtained as an amorphous orange solid [58]. m.p. 193–194 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO) δ 9.09 (1H,s), 7.95 (2H, d, *J* = 7.5 Hz), 7.83 (1H, dd, *J* = 7.9, 1.5 Hz), 7.65–7.63 (3H, m), 7.52 (1H, t, *J* = 7.4 Hz), 7.41 (1H, dd, *J* = 8.3, 0.7 Hz), 7.34 (1H, t, *J* = 7.6 Hz), 6.21 (1H, s), 5.53 (2H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO) δ 164.4 (C=O), 161.6 (C), 152.8 (C), 142.3 (C), 136.5 (C), 132.9 (CH), 129.9 (2CH), 128.9 (CH), 124.2 (CH), 123.4 (CH), 123.1 (CH), 120.3 (2CH), 116.5 (CH), 115.1 (C), 91.5 (CH), 62.8 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 319 ([M<sup>+</sup>], 26); 131 (11);

130 (100); 103 (11); 77 (47); 51 (12); HREIMS319.0967 (calcd. for  $C_{18}H_{13}N_3O_3$  [M<sup>+</sup>] 319.0957); FT-IR (ATR)  $\nu_{max}$  3386, 3146, 3097, 1716, 1621, 1563, 1494, 1460, 1397, 1367, 1337, 1243, 1208, 1186, 1136, 1100, 1029, 925, 835 cm<sup>-1</sup>.

4-((1-(2-*Methoxyphenyl*)-1H-1,2,3-*triazol*-4-*yl*)*methoxy*)-2H-chromen-2-one (**8b**). Following the experimental procedure described in method B, from 42.3 mg (0.28 mmol) of 1-azido-2-methoxybenzene and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 14.2 mg (27%) of compound **8b** were obtained as an amorphous white solid. m.p. 200–201 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ 8.33 (1H, s), 7.84 (2H, dd, J = 7.9, 1.3 Hz), 7.56 (1H, t, J = 8.5 Hz), 7.47 (1H, t, J = 8.5 Hz), 7.32 (1H, dd, J = 8.3, 0.6 Hz), 7.25 (1H, td, J = 7.8, 1.1 Hz), 7.16–7.11 (2H, m), 5.93 (1H, s), 5.43 (2H, s), 3.93 (3H, s) ppm; <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ 165.2 (C=O), 162.8 (C), 153.4 (C), 151.1 (C), 140.8 (C), 132.6 (CH), 130.6 (CH), 126.0 (C), 125.9 (CH), 125.5 (CH), 124.0 (CH), 123.4 (CH), 121.4 (CH), 116.8 (CH), 115.6 (C), 112.4 (CH), 91.2 (CH), 62.8 (CH<sub>2</sub>), 56.2 (CH<sub>3</sub>) ppm; EIMS m/z 349 ([M<sup>+</sup>], 37); 161 (12); 160 (100); 145 (28); 120 (14); 92 (11); 77 (16); HREIMS 349.1050 (calcd. for C<sub>19</sub>H<sub>15</sub>N<sub>3</sub>O<sub>4</sub> [M<sup>+</sup>] 349.1063); FT-IR (ATR)  $\nu_{max}$  3400, 3129, 3093, 3009, 2942, 2841, 2287, 1707, 1617, 1563, 1507, 1493, 1477, 1460, 1410, 1368, 1233, 1186, 1136, 1103, 1052, 1018, 970, 930, 883, 868, 811 cm<sup>-1</sup>.

4-((1-(3-*Methoxyphenyl*)-1H-1,2,3-*triazol*-4-*yl*)*methoxy*)-2H-chromen-2-one (**8**c). Following the experimental procedure described in method B, from 42.3 mg (0.28 mmol) of 1-azido-2-methoxybenzene and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 26.8 mg (51%) of compound **8**c were obtained as an amorphous white solid. m.p. 188–189 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>) δ 8.16 (1H, s), 7.82 (1H, d, *J* = 7.6 Hz), 7.56 (1H, t, *J* = 7.4 Hz), 7.45 (1H, t, *J* = 8.1 Hz), 7.37 (1H, s), 7.33 (1H, d, *J* = 8.1 Hz), 7.30–7.21 (2H, m), 7.00 (1H, d, *J* = 7.8 Hz), 5.91 (1H, s), 5.43 (2H, s), 3.90 (3H, s) ppm;<sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>) δ 165.1 (C=O), 162.7 (C), 160.8 (C), 153.5 (C), 142.2 (C), 137.9 (C), 132.7 (CH), 130.8 (CH), 124.1 (CH), 123.3 (CH), 121.9 (CH), 116.9 (CH), 115.6 (C), 115.2 (CH), 112.7 (CH), 106.7 (CH), 91.4 (CH), 62.7 (CH<sub>2</sub>), 55.8 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 349 ([M<sup>+</sup>], 49); 160 (100); 145 (18); 130 (16); 117 (12); 107 (14); 92 (18); 77 (21); HREIMS 349.1078 (calcd. for C<sub>19</sub>H<sub>15</sub>N<sub>3</sub>O<sub>4</sub> [M<sup>+</sup>] 349.1063); FT-IR (ATR) v<sub>max</sub> 3401, 3148, 3093, 2933, 2838, 1719, 1620, 1610, 1565, 1493, 1456, 1400, 1368, 1337, 1237, 1189, 1158, 1103, 1044, 1004, 931, 833 cm<sup>-1</sup>.

4-((1-(4-*Methoxyphenyl*)-1*H*-1,2,3-*triazol*-4-*yl*)*methoxy*)-2*H*-*chromen*-2-*one* (**8d**). Following the experimental procedure described in method B, from 42.3 mg (0.28 mmol) of 1-azido-4-methoxybenzene and 28.0 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 27.7 mg (53%) of compound **8d** were obtained as an amorphous white solid. m.p. 195–196 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.07 (1H, s), 7.83 (1H, d, *J* = 8.0 Hz), 7.66 (2H, d, *J* = 8.9 Hz), 7.56 (1H, t, *J* = 7.8 Hz), 7.33 (1H, d, *J* = 8.3 Hz), 7.4 (1H, t, *J* = 7.9 Hz), 7.05 (2H, d, *J* = 8.9 Hz), 5.91 (1H, s), 5.42 (2H, s), 3.89 (3H, s) ppm; <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  165.1 (C=O), 162.7 (C), 160.3 (C), 153.5 (C), 142.1 (C), 132.7 (CH), 130.3 (C), 124.1 (CH), 123.3 (CH), 122.5 (2CH), 121.9 (CH), 116.9 (CH), 115.6 (C), 115.0 (2CH), 91.4 (CH), 62.8 (CH<sub>2</sub>), 55.8 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 349 ([M<sup>+</sup>], 29); 161 (12); 160 (100); 145 (13); 92 (12); 77 (13); HREIMS 349.1061 (calcd. for C<sub>19</sub>H<sub>15</sub>N<sub>3</sub>O<sub>4</sub> [M<sup>+</sup>] 349.1063); FT-IR (ATR) v<sub>max</sub> 3140, 3095, 3006, 2942, 2840, 2381, 2055, 1719, 1619, 1564, 1519, 1457, 1400, 1369, 1257, 1240, 1185, 1137, 1102, 1030, 927, 824 cm<sup>-1</sup>.

4-((1-(3-*Fluoro-4-methoxyphenyl*)-1*H*-1,2,3-*triazol-4-yl*)*methoxy*)-2*H*-*chromen*-2-*one* (**8e**). Following the experimental procedure described in method B, from 47.4 mg (0.28 mmol) of 1-azido-3-fluoro-4-methoxybenzene and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 25.3 mg (46%) of compound **8e** were obtained as an amorphous white solid. m.p. 190–191 °C; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>) δ 8.07 (1H, s), 7.82 (1H, dd, *J* = 8.0, 1.0 Hz), 7.60–7.51 (2H, m), 7.48 (1H, d, *J* = 8.8 Hz), 7.33 (1H, d, *J* = 8.3 Hz), 7.24 (1H, t, *J* = 7.9 Hz), 7.11 (1H, t, *J* = 8.7 Hz), 5.91 (1H, s), 5.42 (2H, s), 3.97 (3H, s) ppm; <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>) δ 165.1 (C=O), 162.7 (C), 153.4 (C), 152.3 (*J*<sup>1</sup><sub>C-F</sub> = 249.1 Hz), 148.6 (C,  $J^2_{C-F}$  = 28.7 Hz), 142.4 (C), 132.8 (CH), 124.1 (CH), 123.3 (CH), 121.8 (CH), 117.0 (CH), 116.8 (CH,  $J^3_{C-F}$  = 3.6 Hz), 115.6 (C), 114.0 (CH,  $J^3_{C-F}$  = 1.9 Hz), 110.0 (CH,  $J^2_{C-F}$  = 22.6 Hz), 91.5 (CH), 62.7 (CH<sub>2</sub>), 56.7 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 367 ([M<sup>+</sup>], 27); 179 (12); 178 (100); 163 (21); HREIMS367.0898 (calcd. for

 $C_{19}H_{14}N_3O_4F~[M^+]~367.0968);~FT-IR~(ATR)~\nu_{max}~3488,~3143,~3089,~2976,~2944,~2844,~2361,~1715,~1620,~1565,~1527,~1493,~1451,~1401,~1368,~1328,~1287,~1243,~1233,~1186,~1159,~1133,~1104,~1017,~952,~933,~880,~833,~808~cm^{-1}.$ 

4-((1-(4-*Fluoro-phenyl*)-1*H*-1,2,3-*triazol*-4-*yl*)*methoxy*)-2*H*-*chromen*-2-*one* (**8**f). Following the experimental procedure described in method A, from 34.8 mg (0.24 mmol) of 4-fluorophenyl boronic acid and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 22.1 mg (43%) of compound **8**f were obtained as an amorphous white solid. m.p. 233–235 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  9.07 (1H, s), 8.02–7.98 (2H, m), 7.83 (1H, dd, *J* = 7.9, 1.5 Hz), 7.67 (1H, ddd, *J* = 8.7, 7.4, 1.6 Hz), 7.49 (2H, t, *J* = 8.8 Hz), 7.42 (1H, dd, *J* = 8.3, 0.6 Hz), 7.35 (1H, t, *J* = 7.6 Hz), 6.20 (1H, s), 5.52 (2H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  164.3 (C=O), 161.5 (C), 161.4 (C, *J*<sup>1</sup><sub>C-F</sub> = 245.7 Hz), 152.7 (C), 142.3 (C), 133.0 (C, *J*<sup>4</sup><sub>C-F</sub> = 3.4 Hz), 132.9 (CH), 124.2 (CH), 123.6 (CH), 123.0 (CH), 122.7 (2 CH, *J*<sup>3</sup><sub>C-F</sub> = 8.6 Hz), 116.8 (2 CH, *J*<sup>2</sup><sub>C-F</sub> = 24.1 Hz), 116.5 (CH), 115.1 (C), 91.5 (CH), 62.8 (CH<sub>2</sub>); EIMS *m*/*z* 337 ([M<sup>+</sup>], 23); 149 (11); 148 (100); 95 (25); HREIMS 337.0859 (calcd. for C<sub>18</sub>H<sub>12</sub>N<sub>3</sub>O<sub>3</sub>F [M<sup>+</sup>] 337.0863). FT-IR (ATR) v<sub>max</sub> 3148, 3098, 2384, 2294, 2050, 1718, 1624, 1567, 1516, 1496, 1465, 1454, 1401, 1370, 1233, 1184, 1105, 1051, 1022, 951, 931, 833 cm<sup>-1</sup>.

4-((1-(3-Trifluoromethyl-phenyl)-1H-1,2,3-triazol-4-yl)methoxy)-2H-chromen-2-one (**8g**). Following the experimental procedure described in method B, from 53.2 mg (0.28 mmol) of 1-azido-3-trifluoromethylbenzene and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 24.0 mg (41%) of compound **8g** were obtained as an amorphous white solid. m.p. 192–193 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  9.27 (1H, s), 8.34 (1H, s), 8.32 (1H, d, *J* = 8.2 Hz), 7.93–7.86 (2H, m), 7.85 (1H, dd, *J* = 8.0, 1.5 Hz), 7.68 (1H, ddd, *J* = 8.8, 7.4, 1.6 Hz), 7.43 (1H, d, *J* = 7.7 Hz), 7.37 (1H, t, *J* = 7.6 Hz), 6.22 (1H, s), 5.56 (2H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  164.3 (C=O), 165.1 (C), 152.7 (C), 142.5 (C), 136.9 (C), 132.8 (CH), 131.3 (CH), 130.5 (C,  $J^2_{C-F}$  = 29.6 Hz), 125.4 (CH,  $J^3_{C-F}$  = 4.1 Hz), 124.2 (CH), 124.1 (CH), 123.5 (C,  $J^1_{C-F}$  = 276.4 Hz), 123.7 (CH), 123.0 (CH), 116.9 (CH,  $J^3_{C-F}$  = 4.5 Hz), 116.4 (CH), 115.0 (C), 91.5 (CH), 62.7 (CH<sub>2</sub>);EIMS *m*/*z* 387 ([M<sup>+</sup>], 12); 386 (42); 358 (23); 357 (45); 329 (10); 199 (12); 198 (100); 159 (26); 145 (50); HREIMS 387.0846 (calcd. for C<sub>19</sub>H<sub>12</sub>N<sub>3</sub>O<sub>3</sub>F<sub>3</sub> [M<sup>+</sup>] 387.0831); FT-IR (ATR) v<sub>max</sub> 3531, 3409, 1685, 1618, 1559, 1069, 972 cm<sup>-1</sup>.

4-((1-(3-Nitro-phenyl)-1H-1,2,3-triazol-4-yl)methoxy)-2H-chromen-2-one (**8h**). Following the experimental procedure described in method B, from 46.6 mg (0.28 mmol) of 1-azido-3-nitrobenzene and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 23.1 mg (43%) of compound **8g** were obtained as an amorphous white solid. m.p. 215–217 °C; <sup>1</sup>H-NMR (600 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  9.31 (1H, s), 8.78 (1H, s), 8.45 (1H, d, *J* = 7.8 Hz), 8.36 (1H, d, *J* = 7.9 Hz), 7.93 (1H, t, *J* = 8.1 Hz), 7.85 (1H, d, *J* = 7.8 Hz), 7.67 (1H, t, *J* = 7.6 Hz), 7.42 (1H, d, *J* = 8.3 Hz), 7.36 (1H, t, *J* = 7.5 Hz), 6.21 (1H, s), 5.56 (2H, s) ppm; <sup>13</sup>C-NMR (150 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  164.4 (C=O), 161.6 (C), 152.8 (C), 148.6 (C), 142.8 (C), 137.1 (C), 133.0 (CH), 131.7 (CH), 126.4 (CH), 123.9 (CH), 123.4 (CH), 123.1 (CH), 116.5 (CH), 115.2 (CH), 115.1 (C), 91.6 (CH), 62.8 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 364 ([M<sup>+</sup>], 59); 176 (11); 175 (100); 162 (23); 145 (14); 129 (92); 128 (37); 121 (14); 120 (39); 92 (18); 77 (11); 76 (37); HREIMS 364.0820 (calcd. for C<sub>18</sub>H<sub>12</sub>N<sub>4</sub>O<sub>5</sub> [M<sup>+</sup>] 364.0808); FT-IR (ATR) v<sub>max</sub> 3431, 3146, 3091, 2925, 1713, 1617, 1565, 1531, 1493, 1464, 1406, 1352, 1245, 1185, 1138, 1104, 1045, 1009, 980, 952, 929, 830 cm<sup>-1</sup>.

4-((1-(1H-Indole-5-yl)-1H-1,2,3-triazol-4-yl)methoxy)-2H-chromen-2-one (**8i**). Following the experimental procedure described in method B, from 44.9 mg (0.28 mmol) of 5-azido-1H-indol and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 18.1 mg (33%) of compound **8i** were obtained as an amorphous white solid. m.p. 191–193 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  11.46 (1H, s), 8.99 (1H, s), 8.05 (1H, s), 7.84 (1H, d, *J* = 7.1 Hz), 7.67 (1H, t, *J* = 7.2 Hz), 7.63–7.57 (2H, m), 7.53–7.51 (1H, m), 7.42 (1H, d, *J* = 8.3 Hz), 7.35 (1H, t, *J* = 7.5 Hz), 6.59 (1H, s), 6.22 (1H, s), 5.52 (2H, s); <sup>13</sup>C NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO) 164.4 (C=O), 161.6 (C), 152.8 (C), 141.8 (C), 135.6 (C), 132.9 (CH), 129.3 (C), 127.8 (CH), 127.6 (C), 124.3 (CH), 123.7 (CH), 123.1 (CH), 116.5 (CH), 115.1 (C), 114.4 (CH), 112.4 (CH), 112.3 (CH), 102.0 (CH), 91.4 (CH), 62.9 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 358 ([M<sup>+</sup>], 28); 169 (100); 168 (19); 162 (30); 121 (13); 120 (39); 116 (28);

92 (14); HREIMS 358.1068 (calcd. for  $C_{20}H_{14}N_4O_3$  [M<sup>+</sup>] 358.1066); FT-IR (ATR)  $\nu_{max}$  3399, 3299, 1699, 1685, 1616, 1563, 1494, 1403, 1350, 1327, 1228, 1097, 1048, 1025, 993 cm<sup>-1</sup>.

4-((1-(*Furan-3-yl*)-1*H*-1,2,3-*triazol-4-yl*)*methoxy*)-2*H*-*chromen-2-one* (**8j**). Following the experimental procedure described in method A, from 34.8 mg (0.24mmol) of 3-furylboronic acid and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 8.1 mg (18%) of compound **8j** were obtained as an amorphous white solid. m.p. 189–190 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.88 (1H,s), 8.48 (1H, s), 7.91 (1H, t, *J* = 1.9 Hz), 7.80 (1H, dd, *J* = 7.9, 1.5 Hz), 7.67 (1H, t, *J* = 7.6Hz), 7.42 (1H, dd, *J* = 8.3, 0.7 Hz), 7.35 (1H, t, *J* = 7.8 Hz), 7.16 (1H, dd, *J* = 2.0, 0.9 Hz), 6.19 (1H, s), 5.51 (2H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  164.4 (C=O), 161.6 (C), 152.8 (C), 144.9 (CH), 141.9 (C), 134.3 (CH), 132.9 (CH), 125.9 (C), 124.3 (CH), 124.2 (CH), 123.0 (CH), 116.5 (CH), 115.1 (C), 105.3 (CH), 91.5 (CH), 62.7 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 309 ([M<sup>+</sup>], 59); 279 (25); 198 (21); 159 (23); 120 (100); 94 (18); 65 (13); HREIMS 309.0662 (calcd. for C<sub>16</sub>H<sub>11</sub>N<sub>3</sub>O<sub>4</sub> [M<sup>+</sup>] 309.0671). FT-IR (ATR)  $\nu_{max}$  3514, 3399, 3084, 2924, 2387, 2094, 2064, 1992, 1696, 1619, 1606, 1563, 1492, 1450, 1410, 1365, 1270, 1248, 1193, 1105, 1053, 1023, 942, 911, 871, 839 cm<sup>-1</sup>.

4-((1-Undecyl-1H-1,2,3-triazol-4-yl)methoxy)-2H-chromen-2-one (**8k**). Following the experimental procedure described in method B, from 56.1 mg (0.28mmol) of 1-azido-undecane and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 51.4 mg (86%) of compound **8k** were obtained as an amorphous white solid. m.p. 144–145 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.78 (1H, dd, *J* = 13.2, 6.5 Hz), 7.75 (1H, s), 7.54 (1H, t, *J* = 7.5 Hz), 7.30 (1H, d, *J* = 7.9 Hz), 7.24 (1H, t, *J* = 7.6 Hz), 5.87 (1H, s), 5.34 (1H, s), 4.41 (2H, t, *J* = 7.3 Hz), 1.96 (2H, t, *J* = 7.2 Hz), 1.38–1.23 (16H, m), 0.87 (3H, t, *J* = 6.9 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  165.1 (C=O), 162.7 (C), 153.4 (C), 141.4 (C), 132.6 (CH), 124.0 (CH), 123.4 (CH), 123.2 (CH), 116.8 (CH), 115.5 (C), 91.2 (CH), 62.8 (CH<sub>2</sub>), 50.7 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 30.3 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 29.0 (CH<sub>2</sub>), 26.6 (CH<sub>2</sub>), 22.7 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 397 ([M<sup>+</sup>], 18); 236 (17); 209 (15); 208 (100); 68 (21); 57 (16); 55 (15); HREIMS 397.2351 (calcd. for C<sub>23</sub>H<sub>31</sub>N<sub>3</sub>O<sub>3</sub> [M<sup>+</sup>] 397.2365); FT-IR (ATR) v<sub>max</sub> 3134, 3075, 2918, 2849, 1725, 1623, 1610, 1566, 1492, 1458, 1424, 1381, 1328, 1272, 1248, 1187, 1154, 1138, 1106, 1059, 1031, 979, 931, 882, 851, 814 cm<sup>-1</sup>.

4-((1-Benzyl-1H-1,2,3-triazol-4-yl)methoxy)-2H-chromen-2-one (81). Following the experimental procedure described in method B, from 40.8 mg (0.28 mmol) of benzylazide and 28 mg (0.14 mmol) of *O*-propargylated coumarin (3), 47.0 mg (94%) of compound 81 were obtained as an amorphous white solid. m.p. 210–211 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (1H, dd, *J* = 7.9, 1.5 Hz), 7.63 (1H, s), 7.56–7.52 (1H, t, *J* = 7.6 Hz), 7.47–7.36 (3H, m), 7.35–7.28 (3H, m), 7.23 (1H, t, *J* = 7.6 Hz), 5.84 (s, 1H), 5.59 (s, 2H), 5.31 (2H, d, *J* = 3.8 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  165.1 (C=O), 162.5 (C), 153.7 (C), 142.1 (C), 134.4 (C), 132.6 (CH), 129.5 (2CH), 129.2 (CH), 128.4 (2CH), 124.0 (CH), 123.3 (CH), 123.2 (CH), 117.0 (CH), 115.8 (C), 91.5 (CH), 62.9 (CH<sub>2</sub>), 54.6 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 333 ([M<sup>+</sup>], 39); 172 (13); 144 (62); 104 (11); 92 (14); 91 (100); HREIMS 333.1116 (calcd. for C<sub>19</sub>H<sub>15</sub>N<sub>3</sub>O<sub>3</sub>[M<sup>+</sup>] 333.1113); FT-IR (ATR)  $\nu_{max}$  3075, 1722, 1625, 1569, 1498, 1459, 1421, 1377, 1331, 1277, 1250, 1187, 1141, 1111, 1063, 1035, 983, 933, 885, 849, 814 cm<sup>-1</sup>.

4-((1-*Tetradecyl*-1*H*-1,2,3-*triazol*-4-*yl*)*methoxy*)-2*H*-*chromen*-2-*one* (**8m**). Following the experimental procedure described in method B, from 68.0 mg (0.28 mmol) of 1-azido-tetradecane and 28 mg (0.14 mmol) of *O*-propargylated coumarin (**3**), 61.3 mg (93%) of compound **8m** were obtained as an amorphous white solid. m.p. 146–147 °C; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.79 (1H, dt, *J* = 7.9, 1.5 Hz), 7.72 (1H, s), 7.54 (1H, tt, *J* = 9.1, 1.7 Hz), 7.31 (1H, dd, *J* = 8.4, 1.5 Hz), 7.24 (1H, t, *J* = 7.6 Hz), 5.87 (1H, s), 5.35 (2H, s), 4.41 (2H, t, *J* = 7.3 Hz), 1.96 (2H, t, *J* = 7.1 Hz), 1.38–1.23 (22H, m), 0.88 (3H, t, *J* = 6.9 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  165.1 (C=O), 162.8 (C), 153.4 (C), 141.4 (C), 132.6 (CH), 124.0 (CH), 123.4 (CH), 123.3 (CH), 116.8 (CH), 115.6 (C), 91.2 (CH), 62.8 (CH<sub>2</sub>), 50.8 (CH<sub>2</sub>), 32.0 (CH<sub>2</sub>), 31.0 (CH), 30.4 (CH<sub>2</sub>), 29.9 (CH<sub>2</sub>), 29.8 (CH<sub>2</sub>), 29.7 (2CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 29.1 (CH<sub>2</sub>), 26.6 (CH<sub>2</sub>), 22.8 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 439 ([M<sup>+</sup>], 11); 278 (26); 251 (19); 250 (100); 215 (20); 120 (10); 71 (14); 70 (12); 68 (23); 57 (22); 56 (10); 55 (19); HREIMS 439.2852 (calcd.

for  $C_{26}H_{37}N_3O_3$  [M<sup>+</sup>] 439.2835); FT-IR (ATR)  $\nu_{max}$  3135, 3075, 2918, 2849, 1817, 1726, 1624, 1567, 1468, 1423, 1381, 1328, 1273, 1248, 1185, 1154, 1139, 1107, 1058, 979, 931, 882, 851, 814 cm<sup>-1</sup>.

4-(((1-Phenyl-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9a**). Following the experimental procedure described in method A, from 31.2 mg (0.24 mmol) of phenyl boronic acid and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 13.8 mg (28%) of compound **9a** were obtained as an amorphous white solid. m.p. 194–195 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.82 (1H, s), 8.31 (1H, t, *J* = 5.6 Hz), 8.10 (1H, dd, *J* = 8.0, 1.0 Hz), 7.90 (2H, d, *J* = 7.6 Hz), 7.61–7.56 (3H, m), 7.48 (1H, t, *J* = 7.4 Hz), 7.36–7.30 (2H, m), 5.30 (1H, s), 4.64 (2H, d, *J* = 5.6 Hz) ppm; <sup>13</sup>C-NMR (125 MHz,(CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.5 (C=O), 153.1 (C),153.0 (C), 144.6 (C), 136.6 (C), 132.0 (CH), 130.0 (CH), 128.7 (CH), 123.4 (CH), 122.6 (CH), 121.5 (CH), 120.0 (CH), 117.0 (CH), 114.5 (C), 82.5 (CH), 37.8 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 318([M<sup>+</sup>], 64); 290 (21); 289 (44); 261 (16); 198 (15); 159 (22); 130 (100); 77 (56); HREIMS 318.1117 (calcd. for C<sub>18</sub>H<sub>14</sub>N<sub>4</sub>O<sub>2</sub> [M<sup>+</sup>] 318.1117); FT-IR (ATR)  $\nu_{max}$  3500, 3297, 3138, 3082, 2940, 1707, 1609, 1557, 1503, 1480, 1146, 1377, 1340, 1321, 1188, 1054, 954, 922, 861, 823 cm<sup>-1</sup>.

4-(((1-(2-*methoxyphenyl*)-1H-1,2,3-*triazol*-4-*yl*)*methyl*)*amino*)-2H-*chromen*-2-*one* (**9b**). Following the experimental procedure described in method B, from 68.0 mg (0.28 mmol) of 1-azido-2-methoxybenzene and 28 mg (0.14 mmol) of N-propargylated coumarin (**6**), 16.9 mg (32%) of compound **9b** were obtained as an amorphous white solid. m.p. 187–189 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.36 (1H, s), 8.09 (1H, t, *J* = 5.5 Hz), 8.01 (1H, dd, *J* = 8.1, 1.2 Hz), 7.54 (1H, dd, *J* = 7.9, 1.6 Hz), 7.51 (1H, dd, *J* = 11.3, 4.2 Hz), 7.44 (1H, t, *J* = 7.9 Hz), 7.27–7.20 (m, 3H), 7.06 (1H, td, *J* = 7.7, 1.0 Hz, ), 5.30 (1H, s), 4.56 (2H, d, *J* = 5.7 Hz), 3.77 (3H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.2 (C=O), 153.0 (C), 152.8 (C), 151.4 (C), 142.9 (C), 131.7 (CH), 130.4 (CH), 125.7 (C), 125.4 (CH), 125.1 (CH), 123.1 (CH), 122.4 (CH), 120.8 (CH), 116.7 (CH), 114.4 (C), 113.0 (CH), 82.5 (CH), 56.0 (CH<sub>3</sub>), 37.5 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 348 ([M<sup>+</sup>], 20); 320 (17); 319 (20); 160 (100); 159 (20); 145 (11); 120 (11); 77 (19); HREIMS 348.1228 (calcd. for C<sub>19</sub>H<sub>16</sub>N<sub>4</sub>O<sub>3</sub> [M<sup>+</sup>] 348.1222); FT-IR (ATR)  $\nu_{max}$  3524, 3318, 3173, 3085, 2944, 2847, 2376, 1649, 1607, 1555, 1505, 1476, 1446, 1384, 1327, 1290, 1260, 1244, 1197, 1120, 1044, 1017, 957, 937, 865 cm<sup>-1</sup>.

4-(((1-(3-*Methoxyphenyl*)-1*H*-1,2,3-*triazo*l-4-*y*)*methyl*)*amino*)-2*H*-*chromen*-2-*one* (**9c**). Following the experimental procedure described in method B, from 42.3 mg (0.28 mmol) of 1-azido-3-methoxybenzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 26.9 mg (51%) of compound **9c** were obtained as an amorphous white solid. m.p. 246–248 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.71 (1H, s), 8.08 (1H, t, *J* = 5.5 Hz), 8.02 (1H, dd, *J* = 8.1, 1.2 Hz), 7.52 (1H, t, *J* = 7.8 Hz), 7.44–7.36 (3H, m), 7.27 (1H, d, *J* = 0.8 Hz), 7.24 (1H, t, *J* = 7.7 Hz), 6.97 (1H, dt, *J* = 6.8, 2.5 Hz), 5.25 (1H, s), 4.57 (2H, d, *J* = 5.6 Hz), 3.78 (3H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.2 (C=O), 160.1 (C), 153.0 (C), 152.7 (C), 144.3 (C), 137.5 (C), 131.7 (CH), 130.6 (CH), 123.1 (CH), 123.0 (CH), 121.5 (CH), 116.7 (CH), 114.4 (C), 114.2 (CH), 111.9 (CH), 105.7 (CH), 82.5 (CH), 55.5 (CH<sub>3</sub>), 37.7 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 348 ([M<sup>+</sup>], 40); 319 (25); 291 (10); 198 (11); 161 (13); 160 (100); 159 (30); 123 (11); 107 (24); 92 (19); 77 (26); HREIMS 348.1221 (calcd. for C<sub>19</sub>H<sub>16</sub>N<sub>4</sub>O<sub>3</sub> [M<sup>+</sup>] 348.1222); FT-IR (ATR) v<sub>max</sub> 3297, 3143, 3085, 3000, 2936, 2830, 1708, 1609, 1558, 1483, 1446, 1373, 1322, 1246, 1192, 1158, 1142, 1118, 1048, 954, 922, 859, 846, 819 cm<sup>-1</sup>.

4-(((1-(4-*Methoxyphenyl*)-1*H*-1,2,3-*triazo*l-4-*y*])*methyl*)*amino*)-2*H*-*chromen*-2-*one* (9d). Following the experimental procedure described in method B, from 42.3 mg (0.28 mmol) of 1-azido-4-methoxybenzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (6), 29.6 mg (56%) of compound 9d were obtained as an amorphous white solid. m.p. 246–248 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.58 (1H, s), 8.07 (1H, t, *J* = 5.4 Hz), 8.01 (1H, dd, *J* = 8.1, 1.1 Hz), 7.74–7.69 (2H, m), 7.52 (1H, t, *J* = 7.8 Hz), 7.24 (2H, dd, *J* = 14.4, 7.6 Hz), 7.07–7.01 (2H, m), 5.25 (1H, s), 4.55 (2H, d, *J* = 5.6 Hz), 3.75 (3H, s) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.2 (C=O), 159.2 (C), 153.0 (C), 152.8 (C), 144.1 (C), 131.7 (CH), 130.0 (C), 123.1 (CH), 122.4 (CH), 121.6 (2CH), 121.3 (CH), 116.7 (CH), 114.7 (2CH), 114.4 (C), 82.5 (CH), 55.4 (CH<sub>3</sub>), 37.7 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 348 ([M<sup>+</sup>], 21); 320 (14); 319 (22); 161 (12); 160 (100); 159 (26); 123 (10); 77 (15); HREIMS 348.1225 (calcd. for C<sub>19</sub>H<sub>16</sub>N<sub>4</sub>O<sub>3</sub> [M<sup>+</sup>] 348.1222); FT-IR (ATR) v<sub>max</sub> 3528, 3285,

3137, 3081, 3067, 3003, 2945, 2924, 2829, 2310, 2051, 1700, 1609, 1556, 1517, 1446, 1378, 1307, 1247, 1187, 1140, 1055, 1041, 989, 953, 920, 860, 822 cm<sup>-1</sup>.

4-(((1-(3-Fluoro-4-methoxyphenyl)-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9e**). Following the experimental procedure described in method B, from 47.4 mg (0.28 mmol) of 1-azido-3-fluoro-4-methoxybenzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 27.3 mg (49 %) of compound **9e** were obtained as an amorphous white solid. m.p. 204–205 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.76 (1H, s), 8.29 (1H, t, *J* = 5.7 Hz), 8.09 (1H, dd, *J* = 8.1, 1.2 Hz), 7.87 (1H, dd, *J* = 12.1, 2.6 Hz), 7.72 (1H, ddd, *J* = 8.9, 2.5, 1.4 Hz), 7.60 (1H, t, *J* = 7.8 Hz), 7.39–7.30 (3H, m), 5.28 (1H, s), 4.62 (2H, d, *J* = 5.6 Hz), 3.90 (3H, s); <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  162.4 (C=O), 153.7 (C), 153.4 (C), 151.6 (C, *J*<sup>1</sup><sub>C-F</sub> = 247.8 Hz), 147.7 (C, *J*<sup>2</sup><sub>C-F</sub> = 20.3 Hz), 144.9 (C), 132.7 (CH), 129.9 (C, *J*<sup>3</sup><sub>C-F</sub> = 9.0 Hz), 124.1 (CH), 122.8 (CH), 122.1 (CH), 117.4 (CH), 117.0 (CH, *J*<sup>3</sup><sub>C-F</sub> = 2.8 Hz), 114.9 (C), 114.7 (CH), 109.2 (CH, *J*<sup>2</sup><sub>C-F</sub> = 22.5 Hz), 82.8 (CH), 56.7 (CH<sub>3</sub>), 38.0 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 366 ([M<sup>+</sup>], 30); 338 (16); 337 (22); 198 (15); 179 (13); 178 (100); 159 (22); HREIMS 366.1138 (calcd. for C<sub>19</sub>H<sub>15</sub>N<sub>4</sub>O<sub>3</sub>F [M<sup>+</sup>] 366.1128); FT-IR (ATR) v<sub>max</sub> 3301, 3133, 3078, 3009, 2936, 2849, 1699, 1609, 1557, 1518, 1477, 1446, 1377, 1317, 1184, 1123, 1082, 1053, 953, 921, 884, 859, 817 cm<sup>-1</sup>.

4-(((1-(4-Fluoro-phenyl)-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9f**). Following the experimental procedure described in method A, from 34.8 mg (0.24mmol) of 4-fluoro-phenyl boronic acid and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 15.2 mg (30 %) of compound **9f** were obtained as an amorphous white solid. m.p. 212–213 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.79 (1H, s), 8.32 (1H, t, *J* = 6.0 Hz), 8.16 (1H, d, *J* = 7.8 Hz), 7.97–7.91 (2H, m), 7.61 (1H, t, *J* = 8.4 Hz), 7.45 (2H, t, *J* = 8.8 Hz), 7.35–7.30 (2H, m), 5.30 (1H, s), 4.6 (2H, d, *J* = 5.5 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.4 (C=O), 153.1 (C), 153.0 (C), 144.7 (C), 133.1 (C), 132.0 (CH), 123.4 (CH), 122.6 (CH), 122.3 (2CH, *J*<sup>3</sup><sub>C-F</sub> = 9.1 Hz), 121.7 (C), 116.9 (CH), 116.7 (2CH, *J*<sup>2</sup><sub>C-F</sub> = 23.9 Hz), 114.5 (C), 82.6 (CH), 37.4 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 336 ([M<sup>+</sup>], 54); 308 (19); 307 (36); 198 (15); 159 (27); 148 (100); 95 (38); HREIMS 336.1030 (calcd. for C<sub>18</sub>H<sub>13</sub>N<sub>4</sub>O<sub>2</sub>F [M<sup>+</sup>] 336.1023); FT-IR (ATR) v<sub>max</sub> 3537, 3417, 3307, 3143, 3084, 2454, 2288, 2167, 2051, 1985, 1707, 1610, 1558, 1541, 1516, 1481, 1447, 1377, 1230, 1183, 1051, 993, 957, 920, 825 cm<sup>-1</sup>.

4-(((1-(3-Trifluoromethylphenyl)-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9g**). Following the experimental procedure described in method B, from 53.2 mg (0.28 mmol) of 1-azido-3-trifluoromethylbenzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 26.2 mg (45 %) of compound **9g** were obtained as an amorphous white solid. m.p. 231–232 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  9.00 (1H, s), 8.32 (1H, t, *J* = 5.7 Hz), 8.29–8.24 (2H, m), 8.09 (1H, dd, *J* = 8.1, 1.2 Hz), 7.88–7.80 (2H, m), 7.60 (1H, dd, *J* = 15.6, 1.4 Hz), 7.34 (2H, ddd, *J* = 9.2, 8.2, 1.0 Hz), 5.29 (1H, s), 4.66 (2H, d, *J* = 5.7 Hz) ppm; <sup>13</sup>C-NMR (125MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.5 (C=O), 153.1 (C), 153.0 (C), 145.0 (C), 137.1 (C), 132.0 (CH), 131.4 (CH), 130.5 (C, *J*<sup>2</sup><sub>C-F</sub> = 32.5 Hz), 125.2(CH, *J*<sup>3</sup><sub>C-F</sub> = 3.1 Hz), 123.9 (CH), 123.6 (C, *J*<sup>1</sup><sub>C-F</sub> = 272.5 Hz), 123.4 (CH), 122.6 (CH), 121.8 (CH), 117.0 (CH), 116.7 (CH, *J*<sup>3</sup><sub>C-F</sub> = 3.8 Hz), 114.5 (C), 82.6 (CH), 37.8 (CH<sub>2</sub>); EM-IE *m*/*z* 386 ([M<sup>+</sup>], 72); 358 (26); 357 (50); 198 (100); 159 (29); 145 (49); HREIMS 386.0977 (calcd. for C<sub>19</sub>H<sub>13</sub>N<sub>4</sub>O<sub>2</sub>F<sub>3</sub> [M<sup>+</sup>] 386.0991); FT-IR (ATR) v<sub>max</sub> 3425, 3318, 3142, 3085, 2942, 1701, 1610, 1557, 1540, 1482, 1448, 1377, 1342, 1321, 1298, 1266, 1248, 1172, 1142, 1110, 1070, 1046, 955, 923, 896, 861, 819 cm<sup>-1</sup>.

4-(((1-(3-Nitrophenyl)-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9h**). Following the experimental procedure described in method B, from 46.6 mg (0.28 mmol) of 1-azido-3-nitrobenzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 21.2 mg (39 %) of compound **9h** were obtained as an amorphous white solid. m.p. 240–241 °C; <sup>1</sup>H-NMR (600 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  9.06 (1H, s), 8.73 (1H, t, *J* = 1.9 Hz), 8.42 (1H, d, *J* = 7.8 Hz), 8.32 (2H, dd, *J* = 8.2, 2.1 Hz), 8.10 (1H, d, *J* = 7.9 Hz), 7.93–7.83 (1H, m), 7.61 (1H, t, *J* = 7.4 Hz), 7.37–7.27 (2H, m), 5.29 (1H, s), 4.67 (2H, d, *J* = 5.4 Hz) ppm; <sup>13</sup>C-NMR (150 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.4 (C=O), 153.1 (C), 153.0 (C), 148.6 (C), 145.2 (C), 137.1 (C), 132.0 (CH), 131.5 (CH), 126.0 (CH), 123.4 (CH), 123.1 (CH), 122.5 (CH), 122.0 (CH), 116.9 (CH), 114.7 (CH), 114.5 (C), 82.61 (CH), 37.7 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 363 ([M<sup>+</sup>], 48); 335 (31); 334 (69); 333 (100); 286 (23); 242

(30); 198 (28); 197 (31); 175 (40); 161 (38); 159 (31); 129 (40); 92 (27); 76 (30); 65 (27); HREIMS 363.0983 (calcd. for  $C_{18}H_{13}N_5O_4$  [M<sup>+</sup>] 363.0968); FT-IR (ATR)  $\nu_{max}$  3426, 3137, 3106, 3082, 2927, 1707, 1614, 1561, 1531, 1481, 1448, 1352, 1315, 1269, 1189, 1142, 1119, 1043, 1001, 960, 925, 863, 818 cm<sup>-1</sup>.

4-(((1-(1*H*-Indole-5-*y*l)-1*H*-1,2,3-*triazol*-4-*y*l)*methyl*)*amino*)-2*H*-chromen-2-one (**9i**). Following the experimental procedure described in method B, from 44.9 mg (0.28 mmol) of 5-azido-1*H*-indole and 28.0 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 20.7 mg (37 %) of compound **9i** were obtained as an amorphous white solid. m.p. 236–237 °C;<sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO) δ 11.42 (1H, bs), 8.72 (1H, s), 8.29 (1H, t, *J* = 5.6 Hz), 8.11 (1H, d, *J* = 7.2 Hz), 8.00 (1H, s), 7.63–7.53 (3H, m), 7.50 (1H, s), 7.33 (2H, dd, *J* = 12.4, 7.8 Hz), 6.55 (1H, s, *J* = 2.1 Hz), 5.34 (1H, s), 4.63 (2H, d, *J* = 5.6 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)δ162.4 (C=O), 161.5 (C), 153.1 (C), 153.0(C), 144.1 (C), 135.5 (C), 132.0 (CH), 129.5 (C), 127.7 (CH), 127.6 (C), 123.4 (CH), 122.6 (CH), 121.9 (CH), 116.9 (CH), 114.5 (C), 114.2 (CH), 112.3 (CH), 111.9 (CH), 101.9 (CH), 82.5 (CH), 37.9 (CH<sub>2</sub>); EIMS *m*/*z* 357 ([M<sup>+</sup>], 27); 329 (14); 328 (23); 170 (16); 169 (100); 168 (30); 156 (23); 132 (21); 116 (57); 115 (16); 89 (19); HREIMS 357.1217 (calcd. for C<sub>20</sub>H<sub>15</sub>N<sub>5</sub>O<sub>2</sub> [M<sup>+</sup>] 357.1226); FT-IR (ATR) v<sub>max</sub> 3522, 3397, 3324, 1682, 1616, 1562, 1486, 1354, 1092, 1053, 886 cm<sup>-1</sup>.

4-(((1-(*Furan-3-yl*)-1*H*-1,2,3-*triazol-4-yl*)*methyl*)*amino*)-2*H*-*chromen-2-one* (**9j**). Following the experimental procedure described in method A, from 21.3 mg (0.24 mmol) of 3-furyl boronic acid and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 5.4 mg (12%) of compound **9j** were obtained as an amorphous white solid. m.p. 198–199 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.58 (1H, s), 8.42 (1H, d, *J* = 0.8 Hz), 8.30 (1H, t, *J* = 5.6 Hz), 8.08 (1H, d, *J* = 6.9 Hz), 7.86 (1H, t, *J* = 1.8 Hz), 7.60 (1H, t, *J* = 7.8 Hz), 7.32 (2H, dd, *J* = 8.1, 4.6 Hz), 7.12 (1H, dd, *J* = 2.0, 0.8 Hz), 5.27 (1H, s), 4.62 (2H, d, *J* = 5.6 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.4 (C=O), 153.1 (C), 153.0 (C), 144.8 (CH), 144.3 (C), 133.9 (CH), 132.0 (CH), 126.0 (C), 123.4 (CH), 122.6 (CH), 122.2 (CH), 117.0 (CH), 114.5 (C), 105.1 (CH), 82.5 (CH), 37.7 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 308 ([M<sup>+</sup>], 99); 120 (67); 93 (28); 91 (12); 66 (23); 65 (100); 58 (17); HREIMS 308.1000 (calcd. for C<sub>16</sub>H<sub>12</sub>N<sub>4</sub>O<sub>3</sub> [M<sup>+</sup>] 308.0988); FT-IR (ATR) v<sub>max</sub> 3293, 3134, 3083, 2924, 2853, 2285, 1706, 1610, 1557, 1480, 1446, 1378, 1323, 1262, 1230, 1191, 1143, 1118, 1089, 1039, 1017, 955, 921, 867, 821 cm<sup>-1</sup>.

4-(((1-Undecyl-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9k**). Following the experimental procedure described in method B, from 56.1 mg (0.28 mmol) of 1-azido-undecane and 28 mg (0.14 mmol) of N-propargylated coumarin (**6**), 50.3 mg (84%) of compound **9k** were obtained as an amorphous white solid. m.p. 154–156 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.08–7.93 (3H, m), 7.51 (1H, t, *J* = 7.7 Hz), 7.25 (1H, s), 7.22 (1H, t, *J* = 7.9 Hz), 5.18 (1H, s), 4.46 (2H, d, *J* = 5.6 Hz), 4.24 (2H, t, *J* = 7.0 Hz), 1.80–1.59 (2H, m), 1.14 (16H, s), 0.78 (3H, t, *J* = 6.9 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.1 (C=O), 153.0 (C), 152.7 (C), 143.1 (C), 131.6 (CH), 123.1 (CH), 122.8 (CH), 122.3 (CH), 116.7 (CH), 114.4 (C), 82.3 (CH), 49.2 (CH<sub>2</sub>), 37.7 (CH<sub>2</sub>), 31.0 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 28.6 (CH<sub>2</sub>), 28.4 (CH<sub>2</sub>), 28.1 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>), 21.8 (CH<sub>2</sub>), 13.6 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 396 ([M<sup>+</sup>], 88); 395 (16); 368 (40); 367 (100); 339 (15); 297 (18); 283 (17); 235 (20); 227 (37); 199 (23); 198 (21); 162 (19); 57 (19); 55 (23); HREIMS 396.2504 (calcd. for C<sub>23</sub>H<sub>32</sub>N<sub>4</sub>O<sub>2</sub> [M<sup>+</sup>] 396.2525); FT-IR (ATR)  $\nu_{max}$  3523, 3399, 3325, 3127, 3069, 2955, 2920, 2849, 1679, 1607, 1553, 1481, 1465, 1446, 1376, 1096, 1055 cm<sup>-1</sup>.

4-(((1-Benzyl-1H-1,2,3-triazol-4-yl)methyl)amino)-2H-chromen-2-one (**9**1). Following the experimental procedure described in method B, from 56.1 mg (0.28 mmol) of azidomethyl-benzene and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 45.5 mg (91%) of compound **9**1 were obtained as an amorphous white solid. m.p. 217–219 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.03 (2H, d, *J* = 8.7 Hz), 7.96 (1H, dd, *J* = 8.0, 1.2 Hz), 7.50 (1H, t, *J* = 7.8 Hz), 7.31–7.19 (7H, m), 5.50 (2H, s), 5.19 (1H, s), 4.47 (2H, d, *J* = 5.8 Hz) ppm; <sup>13</sup>C-NMR (125 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.2 (C=O), 153.0 (C), 152.8 (C), 143.5 (C), 135.9 (C), 131.7 (CH), 128.5 (2CH), 127.9 (CH), 127.7 (2CH), 123.2 (CH), 123.1 (CH), 122.3 (CH), 116.7 (CH), 114.3 (C), 82.4 (CH), 52.7 (CH<sub>2</sub>), 37.6 (CH<sub>2</sub>) ppm; EIMS *m*/*z* 332 ([M<sup>+</sup>], 41); 303 (14); 213 (18); 144 (13); 91 (100); 65 (11); HREIMS 332.1279 (calcd. for C<sub>19</sub>H<sub>16</sub>N<sub>4</sub>O<sub>2</sub> [M<sup>+</sup>] 332.1273); FT-IR (ATR)  $\nu_{max}$  3283,

3141, 3075, 2922, 2852, 2366, 2323, 1697, 1608, 1557, 1482, 1446, 1380, 1322, 1262, 1222, 1188, 1124, 1058, 955, 920, 861, 827 cm<sup>-1</sup>.

4-(((1-*Tetradecyl*-1*H*-1,2,3-*triazol*-4-*yl*)*methyl*)*amino*)-2*H*-*chromen*-2-*one* (**9m**). Following the experimental procedure described in method B, from 68.0 mg (0.28 mmol) of 1-azido-tetradecane and 28 mg (0.14 mmol) of *N*-propargylated coumarin (**6**), 63.5 mg (96%) of compound **9m** were obtained as an amorphous white solid. m.p. 156–157 °C; <sup>1</sup>H-NMR (500 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  8.23 (1H, t, *J* = 5.7 Hz), 8.07 (1H, s), 8.05 (1H, d, *J* = 8.1 Hz), 7.58 (1H, d, *J* = 7.5 Hz), 7.33–7.29 (2H, m), 5.24 (1H, s), 4.53 (2H, d, *J* = 5.7 Hz), 4.31 (2H, t, *J* = 7.0 Hz), 1.75 (2H, t, *J* = 7.3 Hz), 1.23–1.13 (22H, m), 0.85 (3H, t, *J* = 6.9 Hz) ppm; <sup>13</sup>C-NMR (150 MHz, (CD<sub>3</sub>)<sub>2</sub>SO)  $\delta$  161.8 (C=O), 153.5 (C), 153.3 (C), 143.7 (C), 132.4 (CH), 123.8 (CH), 123.5 (CH), 122.9 (CH), 117.4 (CH), 114.9 (C), 82.8 (CH), 49.7 (CH<sub>2</sub>), 31.7 (CH<sub>2</sub>), 30.1 (CH<sub>2</sub>), 29.5 (2CH<sub>2</sub>), 29.4 (2CH<sub>2</sub>), 29.3 (2CH<sub>2</sub>), 29.1 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 26.2 (CH<sub>2</sub>), 22.5 (CH<sub>2</sub>), 14.4 (CH<sub>3</sub>) ppm; EIMS *m*/*z* 438 ([M<sup>+</sup>], 92); 410 (55); 409 (100); 367 (37); 227 (51); 199 (44); 198 (60); 162 (39); 57 (42); 55 (47); HREIMS 438.2978 (calcd. for C<sub>26</sub>H<sub>38</sub>N<sub>4</sub>O<sub>2</sub> [M<sup>+</sup>] 438.2995); FT-IR (ATR) v<sub>max</sub> 3320, 3127, 3070, 2918, 2848, 2416, 2167, 2051, 1983, 1658, 1606, 1550, 1467, 1377, 1322, 1258, 1202, 1150, 1118, 1055, 966, 938, 867, 835 cm<sup>-1</sup>.

## 3.2. Microbial Strains

Staphylococcus aureus (ATCC 6538), Enterococcus faecalis (PCM 2673), Escherichia coli (ATCC 8739), Klebsiella pneumoniae (PCM1, Pseudomonas aeruginosa (PCM 2562) and yeast Candida albicans (ATCC 10231) were obtained from the Department of Molecular Biology, The John Paul II Catholic University of Lublin, Poland.

#### 3.3. MIC Determination

The in vitro antimicrobial studies were carried out with the microbroth dilution method against test organisms, as described previously [59,60]. The bacterial strains were inoculated in Mueller Hinton Broth medium (Biocorp, Warsaw, Poland) and the *Candida* strain was inoculated in Sabouraud Dextrose liquid medium (Biocorp, Poland) and incubated at 37 °C and at 30 °C, respectively, with vigorous shaking (200 rpm) for 24 h. Bacterial cell suspensions at initial inoculums of  $5 \times 10^5$  in Mueller-Hinton liquid medium and adequate yeast suspensions at initial inoculums of  $3 \times 10^3$  cfu/mL in Sabouraud Dextrose Broth were exposed to the examined compound at relevant concentrations (range 0.001–2 mg/mL) for 24 h at 37 °C for the bacteria and for 48 h at 30 °C in the case of the fungi. Simultaneously, the standard antibiotics, chloramphenicol for antibacterial activity and ketoconazole for antifungal activity (as a positive control), were tested against the pathogens. The MIC was the lowest concentration of the compounds that inhibited the visible growth of the microorganism. The experiments were performed in triplicate.

#### 3.4. Haemolytic Assay

Haemolytic properties of the selected compounds were determined according to the method described previously [45]. The human blood samples were centrifuged at  $500 \times g$  for 10 min at 4 °C and the supernatant was discarded. Next, the erythrocytes were resuspended with PBS buffer (10 mM phosphate, pH 7.5; 150 mM NaCl) and centrifuged as previously. The washing procedure was repeated until a transparent supernatant was obtained. The washed erythrocytes were finally resuspended in PBS buffer to a final concentration of 2%. Simultaneously, appropriate concentrations (5, 10, 25, 50, 100 and 500 µg/mL for **8a**, **8f**, **9h** and **9k**, or 2, 5, 12.5, 50, 125 and 250 µg/mL for **8b**) of the examined compounds were prepared in a final volume of 50 mL DMSO. The compounds prepared in this way were mixed with 450 mL of 2% erythrocyte suspension and incubated for 1 h at 37 °C. Then, the samples were centrifuged at  $5000 \times g$  for 10 min and absorbance at wavelength 415 nm was measured.

# 4. Conclusions

In conclusion, twenty-eight coumarin-triazole conjugates were synthesized through a copper(I)catalysed Huisgen 1,3-dipolar cycloaddition reaction of the corresponding *O*-propargylated coumarin (3) or *N*-propargylated coumarin (6) with alkyl or aryl azides. Five of them (8a, 8b, 8f, 9h and 9k) displayed promising activity against *Enterococcus faecalis* at MICs ranging from 12.5 to 50.0  $\mu$ g/mL. Compound 8b having a 2-OMe-Ph group attached at the triazol nucleus and an –OCH<sub>2</sub>– linker was the best of the series. The most active compounds showed minimal toxicity towards human blood cells.

**Supplementary Materials:** The following are available online: <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compounds 6, 8a–8m and 9a–9m.

**Acknowledgments:** We gratefully acknowledge the financial support from the Spanish MINECO SAF 2015-65113-C2-1-R to A.E.B. This project is also co-funded by the European Regional Development Fund (FEDER). PLR thanks to Spanish MINECO for a pre-doctoral grant (FPU-Program).

**Author Contributions:** Ana Estévez-Braun, Ángel Amesty, and Maciej Masłyk conceived and designed the experiments; Priscila López-Rojas, Monika Janeczko, Konrad Kubiński, and Ángel Amesty performed the experiments; Ana Estévez-Braun, Ángel Amesty and Maciej Masłyk wrote the paper.

Conflicts of Interest: The authors declare no conflict of interest.

# References

- 1. Holpuch, A. UN meeting tackles the "fundamental threat" of antibiotic-resistant superbugs. *Guardian* 2016.
- 2. Viswanathan, V.K. Off-label abuse of antibiotics by bacteria. *Gut Microbes* 2014, 5, 3–4. [CrossRef] [PubMed]
- 3. Read, A.F.; Woods, R.J. Antibiotic resistance management. *Evol. Med. Public Health* **2014**, 2014, 147. [CrossRef] [PubMed]
- 4. The antibiotic alarm. *Nature* **2013**, *495*, 151.
- 5. Lushniak, B.D. Antibiotic resistance: A public health crisis. *Public Health Rep.* **2014**, *129*, 314–316. [CrossRef] [PubMed]
- 6. Michael, C.A.; Dominey-Howes, D.; Labbate, M. The antimicrobial resistance crisis: Causes, consequences, and management. *Front. Public Health* **2014**, *2*, 145. [CrossRef] [PubMed]
- Laxminarayan, R.; Duse, A.; Wattal, C.; Zaidi, A.K.; Wertheim, H.F.; Sumpradit, N.; Vlieghe, E.; Hara, G.L.; Gould, I.M.; Goossens, H.; et al. Antibiotic resistance-the need for global solutions. *Lancet Infect. Dis.* 2013, 13, 1057–1098. [CrossRef]
- 8. Chambers, H.F. Community-associated MRSA—Resistance and virulence converge. *N. Engl. J. Med.* 2005, 352, 1485–1487. [CrossRef] [PubMed]
- 9. Moran, G.J.; Krishnadasan, A.; Gorwitz, R.J.; Fosheim, G.E.; McDougal, L.K.; Carey, R.B.; Talan, D.A.; Group, E.I.N.S. Methicillin-resistant S. aureus infections among patients in the emergency department. *N. Engl. J. Med.* **2006**, *355*, 666–674. [CrossRef] [PubMed]
- Frazee, B.W.; Lynn, J.; Charlebois, E.D.; Lambert, L.; Lowery, D.; Perdreau-Remington, F. High prevalence of methicillin-resistant Staphylococcus aureus in emergency department skin and soft tissue infections. *Ann. Emerg. Med.* 2005, 45, 311–320. [CrossRef] [PubMed]
- 11. Fridkin, S.K.; Hageman, J.C.; Morrison, M.; Sanza, L.T.; Como-Sabetti, K.; Jernigan, J.A.; Harriman, K.; Harrison, L.H.; Lynfield, R.; Farley, M.M.; et al. Methicillin-resistant Staphylococcus aureus disease in three communities. *N. Engl. J. Med.* **2005**, *352*, 1436–1444. [CrossRef] [PubMed]
- 12. Moran, G.J.; Amii, R.N.; Abrahamian, F.M.; Talan, D.A. Methicillin-resistant Staphylococcus aureus in community-acquired skin infections. *Emerg. Infect. Dis.* **2005**, *11*, 928–930. [CrossRef] [PubMed]
- 13. Hiramatsu, K. Vancomycin-resistant Staphylococcus aureus: A new model of antibiotic resistance. *Lancet Infect. Dis.* **2001**, *1*, 147–155. [CrossRef]
- Wilson, P.; Andrews, J.A.; Charlesworth, R.; Walesby, R.; Singer, M.; Farrell, D.J.; Robbins, M. Linezolid resistance in clinical isolates of Staphylococcus aureus. *J. Antimicrob. Chemother.* 2003, *51*, 186–188. [CrossRef] [PubMed]
- 15. Hollenbeck, B.L.; Rice, L.B. Intrinsic and acquired resistance mechanisms in enterococcus. *Virulence* **2012**, *3*, 421–433. [CrossRef] [PubMed]

- 16. World Health Organization (WHO). *Antimicrobial Resistance: Global Report on Surveillance;* WHO: Geneva, Switzerland, 2014.
- 17. Kristich, C.J.; Rice, L.B.; Arias, C.A. Enterococcal infection—Treatment and antibiotic resistance. In *Enterococci: From Commensals to Leading Causes of Drug Resistant Infection*; Gilmore, M., Clewell, D., Ike, Y., Eds.; Massachusetts Eye and Ear Infirmary: Boston, MA, USA, 2014.
- Hueso-Falcón, I.; Amesty, Á.; Anaissi-Afonso, L.; Lorenzo-Castrillejo, I.; Machín, F.; Estévez-Braun, A. Synthesis and biological evaluation of naphthoquinone-coumarin conjugates as topoisomerase II inhibitors. *Bioorg. Med. Chem. Lett.* 2017, 27, 484–489. [CrossRef] [PubMed]
- Jiménez-Alonso, S.; Pérez-Lomas, A.L.; Estévez-Braun, A.; Muñoz Martinez, F.; Chávez Orellana, H.; Ravelo, A.G.; Gamarro, F.; Castanys, S.; López, M. Bis-pyranobenzoquinones as a new family of reversal agents of the multidrug resistance phenotype mediated by P-glycoprotein in mammalian cells and the protozoan parasite Leishmania. *J. Med. Chem.* 2008, *51*, 7132–7143. [CrossRef] [PubMed]
- 20. Weigt, S.; Huebler, N.; Strecker, R.; Braunbeck, T.; Broschard, T.H. Developmental effects of coumarin and the anticoagulant coumarin derivative warfarin on zebrafish (Danio rerio) embryos. *Reprod. Toxicol.* **2012**, *33*, 133–141. [CrossRef] [PubMed]
- 21. Jameel, E.; Umar, T.; Kumar, J.; Hoda, N. Coumarin: A privileged scaffold for the design and development of antineurodegenerative agents. *Chem. Biol. Drug. Des.* **2016**, *87*, 21–38. [CrossRef] [PubMed]
- Torres, F.C.; Brucker, N.; Andrade, S.F.; Kawano, D.F.; Garcia, S.C.; Poser, G.L.; Eifler-Lima, V.L. New insights into the chemistry and antioxidant activity of coumarins. *Curr. Top. Med. Chem.* 2014, 14, 2600–2623. [CrossRef] [PubMed]
- 23. Kaur, M.; Kohli, S.; Sandhu, S.; Bansal, Y.; Bansal, G. Coumarin: A promising scaffold for anticancer agents. *Anticancer. Agents Med. Chem.* **2015**, *15*, 1032–1048. [CrossRef] [PubMed]
- 24. Widelski, J.; Popova, M.; Graikou, K.; Glowniak, K.; Chinou, I. Coumarins from angelica lucida L.—Antibacterial activities. *Molecules* **2009**, *14*, 2729–2734. [CrossRef] [PubMed]
- Ouahouo, B.M.; Azebaze, A.G.; Meyer, M.; Bodo, B.; Fomum, Z.T.; Nkengfack, A.E. Cytotoxic and antimicrobial coumarins from Mammea africana. *Ann. Trop. Med. Parasitol.* 2004, *98*, 733–739. [CrossRef] [PubMed]
- Walasek, M.; Grzegorczyk, A.; Malm, A.; Skalicka-Woźniak, K. Bioactivity-guided isolation of antimicrobial coumarins from Heracleum mantegazzianum Sommier & Levier (Apiaceae) fruits by high-performance counter-current chromatography. *Food Chem.* 2015, *186*, 133–138. [PubMed]
- 27. Tsassi, V.B.; Hussain, H.; Meffo, B.Y.; Kouam, S.F.; Dongo, E.; Schulz, B.; Greene, I.R.; Krohn, K. Antimicrobial coumarins from the stem bark of Afraegle paniculata. *Nat. Prod. Commun.* **2010**, *5*, 559–561. [PubMed]
- 28. Baraza, L.D.; Neser, W.; Jackson, K.C.; Fredrick, J.B.; Dennis, O.; Wairimu, K.R.; Keya, A.O.; Heydenreich, M. Antimicrobial Coumarins from the Oyster Culinary-Medicinal Mushroom, Pleurotus ostreatus (Agaricomycetes), from Kenya. *Int. J. Med. Mushrooms* **2016**, *18*, 905–913. [CrossRef] [PubMed]
- Vatmurge, N.S.; Hazra, B.G.; Pore, V.S.; Shirazi, F.; Chavan, P.S.; Deshpande, M.V. Synthesis and antimicrobial activity of beta-lactam-bile acid conjugates linked via triazole. *Bioorg. Med. Chem. Lett.* 2008, 18, 2043–2047. [CrossRef] [PubMed]
- Agalave, S.G.; Maujan, S.R.; Pore, V.S. Click chemistry: 1,2,3-triazoles as pharmacophores. *Chem. Asian J.* 2011, 6, 2696–2718. [CrossRef] [PubMed]
- Prasad, D.J.; Ashok, M.; Karegoudar, P.; Poojary, B.; Holla, B.S.; Kumari, N.S. Synthesis and antimicrobial activities of some new triazolothiadiazoles bearing 4-methylthiobenzyl moiety. *Eur. J. Med. Chem.* 2009, 44, 551–557. [CrossRef] [PubMed]
- 32. Holla, B.S.; Gonsalves, R.; Shenoy, S. Studies on some N-bridged heterocycles derived from bis-[4-amino-5-mercapto-1,2,4-triazol-3-yl] alkanes. *Farmaco* **1998**, *53*, 574–578. [CrossRef]
- Turan-Zitouni, G.; Kaplancikli, Z.A.; Yildiz, M.T.; Chevallet, P.; Kaya, D. Synthesis and antimicrobial activity of 4-phenyl/cyclohexyl-5-(1-phenoxyethyl)-3-[*N*-(2-thiazolyl)acetamido]thio-4H-1,2,4-triazole derivatives. *Eur. J. Med. Chem.* 2005, 40, 607–613. [CrossRef] [PubMed]
- Masuda, K.; Toga, T.; Hayashi, N. Synthesis of 3-morpholino-N-ethoxycarbonyl sydnonimine-5-<sup>14</sup>C (sin-10-<sup>14</sup>C). J. Labell. Compd. 1975, 11, 301–304. [CrossRef]
- 35. Almasirad, A.; Tabatabai, S.A.; Faizi, M.; Kebriaeezadeh, A.; Mehrabi, N.; Dalvandi, A.; Shafiee, A. Synthesis and anticonvulsant activity of new 2-substituted-5-[2-(2-fluorophenoxy)phenyl]-1,3,4-oxadiazoles and 1,2,4-triazoles. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 6057–6059. [CrossRef] [PubMed]

- 36. Holla, B.S.; Poojary, K.N.; Rao, B.S.; Shivananda, M.K. New bis-aminomercaptotriazoles and bis-triazolothiadiazoles as possible anticancer agents. *Eur. J. Med. Chem.* **2002**, *37*, 511–517. [CrossRef]
- Shivarama Holla, B.; Veerendra, B.; Shivananda, M.K.; Poojary, B. Synthesis characterization and anticancer activity studies on some Mannich bases derived from 1,2,4-triazoles. *Eur. J. Med. Chem.* 2003, *38*, 759–767. [CrossRef]
- Pingaew, R.; Mandi, P.; Nantasenamat, C.; Prachayasittikul, S.; Ruchirawat, S.; Prachayasittikul, V. Design, synthesis and molecular docking studies of novel N-benzenesulfonyl-1,2,3,4-tetrahydroisoquinoline-based triazoles with potential anticancer activity. *Eur. J. Med. Chem.* 2014, *81*, 192–203. [CrossRef] [PubMed]
- Manclús, J.J.; Moreno, M.J.; Plana, E.; Montoya, A. Development of monoclonal immunoassays for the determination of triazole fungicides in fruit juices. *J. Agric. Food Chem.* 2008, *56*, 8793–8800. [CrossRef] [PubMed]
- 40. Amir, M.; Shikha, K. Synthesis and anti-inflammatory, analgesic, ulcerogenic and lipid peroxidation activities of some new 2-[(2,6-dichloroanilino)phenyl]acetic acid derivatives. *Eur. J. Med. Chem.* **2004**, *39*, 535–545. [CrossRef] [PubMed]
- 41. Peña, R.; Martín, P.; Feresin, G.E.; Tapia, A.; Machín, F.; Estévez-Braun, A. Domino synthesis of embelin derivatives with antibacterial activity. *J. Nat. Prod.* **2016**, *79*, 970–977. [CrossRef] [PubMed]
- 42. Casero, C.; Machín, F.; Méndez-Álvarez, S.; Demo, M.; Ravelo, Á.; Pérez-Hernández, N.; Joseph-Nathan, P.; Estévez-Braun, A. Structure and antimicrobial activity of phloroglucinol derivatives from Achyrocline satureioides. *J. Nat. Prod.* **2015**, *78*, 93–102. [CrossRef] [PubMed]
- 43. Peña, R.; Jiménez-Alonso, S.; Feresin, G.; Tapia, A.; Méndez-Alvarez, S.; Machín, F.; Ravelo, Á.; Estévez-Braun, A. Multicomponent synthesis of antibacterial dihydropyridin and dihydropyran embelin derivatives. *J. Org. Chem.* **2013**, *78*, 7977–7985. [CrossRef] [PubMed]
- 44. Casero, C.; Estévez-Braun, A.; Ravelo, A.G.; Demo, M.; Méndez-Álvarez, S.; Machín, F. Achyrofuran is an antibacterial agent capable of killing methicillin-resistant vancomycin-intermediate Staphylococcus aureus in the nanomolar range. *Phytomedicine* **2013**, *20*, 133–138. [CrossRef] [PubMed]
- 45. Janeczko, M.; Demchuk, O.M.; Strzelecka, D.; Kubiński, K.; Masłyk, M. New family of antimicrobial agents derived from 1,4-naphthoquinone. *Eur. J. Med. Chem.* **2016**, 124, 1019–1025. [CrossRef] [PubMed]
- Rostovtsev, V.V.; Green, L.G.; Fokin, V.V.; Sharpless, K.B. A stepwise huisgen cycloaddition process: copper(I)-catalyzed regioselective "ligation" of azides and terminal alkynes. *Angew. Chem. Int. Ed. Engl.* 2002, 41, 2596–2599. [CrossRef]
- 47. Yang, H.; Li, Y.; Jiang, M.; Wang, J.; Fu, H. General copper-catalyzed transformations of functional groups from arylboronic acids in water. *Chem. Eur. J.* **2011**, *17*, 5652–5660. [CrossRef] [PubMed]
- 48. Tao, C.-Z.; Cui, X.; Li, J.; Liu, A.-X.; Liu, L.; Guo, Q.-X. Copper-catalyzed synthesis of aryl azides and 1-aryl-1, 2, 3-triazoles from boronic acids. *Tetrahedron Lett.* **2007**, *48*, 3525–3529. [CrossRef]
- 49. Yuhong, J.; Dalip, K.; Rajender, S.V. Revisiting nucleophilic substitution reactions: Microwave-Assisted synthesis of azides, thiocyanates, and sulfones in an aqueous medium. *J. Org. Chem.* **2006**, *71*, 6697–6700.
- 50. Díaz, L.; Bujons, J.; Casas, J.; Llebaria, A.; Delgado, A. Click chemistry approach to new N-substituted aminocyclitols as potential pharmacological chaperones for Gaucher disease. *J. Med. Chem.* **2010**, *53*, 5248–5255. [CrossRef] [PubMed]
- 51. Zhou, C.H.; Wang, Y. Recent researches in triazole compounds as medicinal drugs. *Curr. Med. Chem.* **2012**, *19*, 239–280. [CrossRef] [PubMed]
- 52. Khaligh, P.; Salehi, P.; Bararjanian, M.; Aliahmadi, A.; Khavasi, H.R.; Nejad-Ebrahimi, S. Synthesis and in Vitro Antibacterial Evaluation of Novel 4-Substituted 1-Menthyl-1,2,3-triazoles. *Chem. Pharm. Bull.* **2016**, *64*, 1589–1596. [CrossRef] [PubMed]
- 53. Kant, R.; Kumar, D.; Agarwal, D.; Gupta, R.D.; Tilak, R.; Awasthi, S.K.; Agarwal, A. Synthesis of newer 1,2,3-triazole linked chalcone and flavone hybrid compounds and evaluation of their antimicrobial and cytotoxic activities. *Eur. J. Med. Chem.* **2016**, *113*, 34–49. [CrossRef] [PubMed]
- Zhang, Q.R.; Xue, D.Q.; He, P.; Shao, K.P.; Chen, P.J.; Gu, Y.F.; Ren, J.L.; Shan, L.H.; Liu, H.M. Synthesis and antimicrobial activities of novel 1,2,4-triazolo [3,4-a] phthalazine derivatives. *Bioorg. Med. Chem. Lett.* 2014, 24, 1236–1238. [CrossRef] [PubMed]
- 55. Perrin, D.D.; Armarego, W.L.F. Purification of Laboratory Chemicals, 3rd ed.; Pergamon Press: Oxford, UK, 1988.
- 56. Wan, X.; Wang, D.; Liu, S. Fluorescent pH-sensing organic/inorganic hybrid mesoporous silica nanoparticles with tunable redox-responsive release capability. *Langmuir* **2010**, *26*, 15574–15579. [CrossRef] [PubMed]

- 57. Yu, B.; Qi, P.P.; Shi, X.J.; Huang, R.; Guo, H.; Zheng, Y.C.; Yu, D.Q.; Liu, H.M. Efficient synthesis of new antiproliferative steroidal hybrids using the molecular hybridization approach. *Eur. J. Med. Chem.* **2016**, *117*, 241–255. [CrossRef] [PubMed]
- 58. Gupta, A.; Jamatia, R.; Mahato, M.; Pal, A.K. Metalloprotein-inspired ruthenium polymeric complex: A highly efficient catalyst in parts per million level for 1,3-dipolar huisgen's reaction in aqueous medium at room temperature. *Indust. Engin. Chem. Res.* **2017**, *56*, 2375–2382. [CrossRef]
- Janeczko, M.; Kazimierczuk, Z.; Orzeszko, A.; Niewiadomy, A.; Krol, E.; Szyszka, R.; Maslyk, M. In Search of the Antimicrobial Potential of Benzimidazole Derivatives. *Pol. J. Microbiol.* 2016, 65, 359–364. [CrossRef] [PubMed]
- Janeczko, M.; Masłyk, M.; Kubiński, K.; Golczyk, H. Emodin, a natural inhibitor of protein kinase CK2, suppresses growth, hyphal development, and biofilm formation of Candida albicans. *Yeast* 2017, *34*, 253–265. [CrossRef] [PubMed]

Sample Availability: Samples of the compounds 8a–l, and 9a–l are available from the authors.



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).