Vet Dermatol 2020; 31: 446-e118

Serum canine thymus and activation-regulated chemokine (TARC/CCL17) concentrations correlate with disease severity and therapeutic responses in dogs with atopic dermatitis

Ryota Asahina* (), Kazunori Ueda†, Yuri Oshima‡, Toshitaka Kanei*, Masahiro Kato§, Masutaka Furue¶, Toshihiro Tsukui§, Masahiko Nagata** () and Sadatoshi Maeda* ()

*Department of Veterinary Medicine, Faculty of Applied Biological Sciences, Gifu University, 1-1 Yanagido, Gifu, 501-1193, Japan †Yokohama Yamate Dog and Cat Medical Center, 27-4 Kashiwaba Naka-ku Yokohama, Kanagawa, 231-0866, Japan ‡Dermatology Services for Dogs and Cats, TANDEM Hirano Bld. 1F, 2-11-14 Hirano Koto-ku, Tokyo, 135-0023, Japan §Animal Life Science Laboratories, Nippon Zenyaku Kogyo Co., Ltd., 1-1 Tairanoue Sasagawa Asaka-machi Koriyama, Fukushima, 963-0196, Japan ¶Department of Dermatology, Graduate School of Medical Science, Kyushu University, 3-1-1 Maidashi Higashi-ku, Fukuoka, 812-8582, Japan **Dermatology Service, Veterinary Specialists Emergency Center, 815 Ishigami Kawaguchi, Saitama, 333-0823, Japan

Correspondence: Sadatoshi Maeda, Department of Veterinary Medicine, Faculty of Applied Biological Sciences, Gifu University, 1-1 Yanagido, Gifu 501-1193, Japan. E-mail: sadat@gifu-u.ac.jp

Background – Thymus and activation-regulated chemokine (TARC/CCL17) has been implicated in the pathogenesis of canine atopic dermatitis (cAD). Serum TARC concentrations are a reliable biomarker for human atopic dermatitis; however, their potential as a biomarker for cAD has not been investigated.

Hypothesis/Objectives – To investigate whether serum TARC concentrations correlate with disease severity and therapeutic responses for cAD.

Animals – Thirty-nine dogs with cAD and 42 healthy dogs were recruited.

Methods and materials – Serum TARC concentrations in dogs with cAD and healthy dogs were measured by sandwich ELISA with anti-canine TARC antibodies. The clinical severity of cAD was scored using the validated Canine Atopic Dermatitis Extent and Severity Index, 4th iteration (CADESI-04). Serum TARC concentrations were compared between dogs with cAD and healthy controls, and their relationship with CADESI-04 was examined. Serum TARC concentrations also were measured in 20 dogs with cAD treated with prednisolone or oclacitinib for four weeks.

Results – Serum TARC concentrations were significantly higher in dogs with cAD than in healthy dogs (P < 0.001). In dogs with cAD, serum TARC concentrations correlated with CADESI-04 scores ($\rho = 0.457$, P < 0.01). Furthermore, serum TARC concentrations significantly decreased in treated dogs with the attenuation of clinical signs (P < 0.001). Changes in serum TARC concentrations before and after treatment correlated with those in CADESI-04 scores ($\rho = 0.746$, P < 0.001).

Conclusions and clinical relevance – Serum TARC concentrations have potential as a clinical and research tool for the objective evaluation of disease severity and therapeutic responses for cAD.

Introduction

Canine atopic dermatitis (cAD) is a chronic inflammatory and pruritic skin disease. It shares many clinical characteristics with its counterpart in humans, such as a genetic predisposition, early age of onset, predilection sites of affected skin, a relationship with epidermal barrier defects and frequent colonization by *Staphylococcus*.¹ Furthermore, dogs with cAD show characteristic laboratory findings, with elevated allergen-specific immunoglobulin (Ig)E concentrations and increased eosinophil counts

Accepted 9 July 2020

Ryota Asahina and Kazunori Ueda are joint first authors. **Sources of Funding:** This study was self-funded. **Conflict of Interest:** No conflicts of interest have been declared. in peripheral blood that may be attributed to excessive amounts of Type 2 helper T (Th2) cytokines.^{2–4} Previous studies reported that concentrations of Th2 cytokines, including interleukin (IL)-4, IL-5 and IL-13, in serum,⁵ peripheral blood mononuclear cells³ and lesional skin,^{6–8} were higher in atopic dogs than in healthy dogs. One study also found elevated numbers of IL-4-expressing helper T cells in the peripheral blood of dogs with cAD.⁹ Therefore, cAD is a Th2-associated inflammatory disease, similar to human atopic dermatitis (AD).

Thymus and activation-regulated chemokine (TARC/ CCL17) is a functional ligand for CC chemokine receptor 4 (CCR4), which is selectively expressed on Th2 cells.^{10,11} The number of CCR4⁺ helper T cells was reported to be elevated in the peripheral blood of dogs with cAD.¹² Furthermore, previous studies demonstrated the preferential transcription of CCR4 and its ligand TARC in the lesional

skin of dogs with cAD, and not in nonlesional and healthy skin.13,14 Immunohistochemical analyses revealed that TARC was expressed in the lesional keratinocytes of cAD.¹¹ In an *in vitro* study using canine keratinocytes, TARC was induced by an agonist of protease-activated receptor-2 that recognized proteases derived from house dust mites (HDM), a major allergen in cAD.¹⁵ Previous studies showed that the transcription of TARC was upregulated, with the greatest increases being observed among the genes examined in the lesional skin of experimentally sensitized dogs with HDM antigens.16,17 In another study using an experimental cAD model, a CCR4 antagonist reduced the infiltration of CCR4⁺ mononuclear cells into skin.¹⁸ Therefore, TARC likely plays a crucial role in the pathogenesis of cAD by facilitating the migration of Th2 cells to lesional skin.

Clinical studies of human AD demonstrated that serum or plasma TARC concentrations correlated with disease severity.^{19–23} Additionally, among several biomarkers, such as serum lactate dehydrogenase (LDH), total IgE concentrations and peripheral eosinophil counts, serum TARC concentrations more strongly correlated with clinical scores.^{24,25} Serum TARC concentrations are currently the most sensitive biomarker for assessing the disease severity of human AD. Furthermore, serum TARC concentrations in patients with human AD decreased with corresponding improvements in clinical signs after treatments with topical agents, including corticosteroids and/ or tacrolimus^{19,22,26} and oral ciclosporin.^{20,25} Accordingly, serum TARC concentrations also have been used as an objective marker to evaluate therapeutic responses in human AD.²⁴

Although increasing evidence implicates TARC in cAD, it currently remains unclear whether serum TARC concentrations have potential as a biomarker for cAD. In the present study, we investigated whether serum TARC concentrations correlate with disease severity and therapeutic responses for cAD using a canine TARC-specific ELISA system.

Methods and materials

Animals

Thirty-nine client-owned dogs with spontaneous cAD in 17 veterinary hospitals were recruited consecutively for the present study. The age of animals ranged between one and 15 years (median age eight years); there were 22 females (16 neutered) and 17 males (nine neutered). Breeds included Shiba inus (n = 19), mongrels (n = 8), French bulldogs (n = 4) and one each of the following breeds: beagle, Border collie, Labrador retriever, Maltese, miniature dachshund, Samoyed, toy poodle and West Highland white terrier. The diagnosis of cAD was based on the combination of compatible clinical features²⁷ and the fulfilment of at least five of eight criteria.²⁸ Other pruritic skin diseases, such as flea allergy dermatitis, scabies, demodicosis, and bacterial or Malassezia dermatitis, were excluded based on routine dermatological examinations and therapeutic trials. In cases exhibiting year-round clinical signs, an elimination diet trial for a minimum of eight weeks was performed to rule out a cutaneous adverse food reaction. All affected dogs had serological evidence of allergen-specific IgE. Tests for Dermatophagoides farinae (Der f) HDM or Der f 2 were performed on all dogs (Nippon Zenyaku Kogyo, Zenoaq; Fukushima, Japan). In cases in which these tests were negative, other serum allergen-specific IgE tests for environmental allergens, such as Der f HDM, Der f 2, grasses and pollens, were performed (at Animal Allergy Clinical Laboratories; Kanagawa, Japan).²⁹

Forty-two healthy dogs were used as control samples. Dogs were considered to be healthy based on their medical history and a physical examination. The age of animals ranged between one and 13 years (median age 4.5 years); there were 24 females (15 neutered) and 18 males (seven neutered). They originated from a research beagle colony (n = 12) or were privately owned (n = 30). The privately owned dog breeds included Shiba inu (n = 21), golden retriever (n = 5) and one each of the following breeds: cavalier King Charles spaniel, French bulldog, Shetland sheepdog and toy poodle. Research beagles were kept for experimental purposes under a protocol approved by the Animal Care and Use Committee of our institute (#17014). In Japan, ethics committees are not available for private-practice animal hospitals. Even so, this study was conducted in accordance with the ethical codes of the Japan Veterinary Medical Association.³⁰

Clinical score assessment and treatment for cAD

The validated Canine Atopic Dermatitis Extent and Severity Index, 4th iteration (CADESI-04) was used to evaluate the severity of lesions.³¹ Twenty of 39 affected dogs had been treated with oral prednisolone (Predonin, Shionogi & Co., Ltd.; Osaka, Japan; 0.5–1.0 mg/kg once daily for six days, then every other day; n = 11) or oral oclacitinib (Apoquel, Zoetis Inc.; Florham Park, NJ, USA; 0.4–0.6 mg/kg twice daily for 14 days, then once daily; n = 9) for 28 days, according to a previous study.³² CADESI-04 scores were evaluated before and after treatment.

Blood samples

Blood samples were collected from dogs with cAD and client-owned healthy dogs that visited the clinic for routine check-ups or vaccinations; the owners of all dogs consented to sample collection for the purpose of this study. One millilitre blood samples were obtained from each dog and centrifuged to separate serum. For all recruited dogs, any anti-inflammatory agents, including glucocorticoids, ciclosporin, oclacitinib or antihistamines were withdrawn for at least two weeks.³³ Blood collection was performed on all dogs with cAD. Twenty of 39 atopic dogs also underwent blood sampling after four weeks of treatment with oral prednisolone or oclacitinib. Serum samples were stored at -30° C until analysed.

Recombinant canine TARC

The recombinant canine TARC protein was produced using an *Escherichia coli* expression system. The cDNA sequence, excluding the signal sequence, was inserted into an *E. coli* expression vector (pGEX-4T-1, 28954549, GE Healthcare Life Sciences; Little Chalfont, UK). The recombinant canine TARC protein was purified as a fusion protein with glutathione S-transferase (GST); the mature protein and GST protein were cleaved using thrombin (GE Healthcare Life Sciences; 27-0846-01) and only the mature protein was purified.

Anti-canine TARC monoclonal and polyclonal antibodies

The cDNA of canine TARC was inserted into a mammalian expression vector (pcDNA3.1, V79020, Thermo Fisher Scientific; Waltham, MA, USA). Mice (BALB/c) were immunized with plasmid DNA to establish a monoclonal antibody. The recombinant protein was used for monoclonal antibody screening. Rabbits were immunized with the same plasmid DNA that was used to develop the monoclonal antibody in order to generate a polyclonal antibody. After DNA immunization, an increase in the antibody titre was confirmed using a recombinant protein and blood was collected to obtain a polyclonal antibody. The production of anti-canine TARC antibodies using mice and rabbits was approved by the Animal Care and Use Committee of our institute (#Z120720, #Z130413).

Table 1. Breed, age, sex, weight, Canine Atopic Dermatitis Extent and Severity Index, 4th iteration (CADESI-04) scores and serum thymus and
activation-regulated chemokine (TARC) concentrations at baseline and after treatment in dogs with atopic dermatitis

Case	Breed	Age (years)	Sex	Weight (kg)	Baseline		After treatment		
					CADESI-04 score	Serum TARC concentration (pg/mL)	CADESI-04 score	Serum TARC concentration (pg/mL)	Treatment
1	Shiba inu	6	SF	8	41	3,143	19	1,693	Prednisolone
2	Shiba inu	6	СМ	11	50	2,415	25	346	Prednisolone
3	Shiba inu	7	F	8	40	1,591	10	310	Prednisolone
4	Shiba inu	7	СМ	11	7	663	4	309	Prednisolone
5	Shiba inu	8	SF	8	54	2,113	10	189	Oclacitinib
6	Shiba inu	8	SF	10	37	2,447	9	1,532	Prednisolone
7	Shiba inu	11	SF	14	25	4,563	17	3,937	Prednisolone
8	Shiba inu	13	SF	8	105	5,469	53	564	Prednisolone
9	Shiba inu	13	SF	7.2	32	2,135	2	195	Oclacitinib
10	Shiba inu	13	СМ	10	41	1,753	12	183	Prednisolone
11	French bulldog	9	SF	11	65	5,006	35	1,825	Oclacitinib
12	French bulldog	7	SF	10.6	22	3,275	2	251	Oclacitinib
13	French bulldog	11	F	8.2	139	3,173	117	1,376	Oclacitinib
14	Mixed breed	5	Μ	13	43	6,502	7	3,564	Prednisolone
15	Mixed breed	11	Μ	23	48	4,406	15	2,052	Oclacitinib
16	Beagle	12	SF	9	19	953	5	150	Prednisolone
17	Maltese	11	SF	6.5	53	1,558	32	884	Oclacitinib
18	Miniature dachshund	10	SF	3	13	4,060	0	2,708	Oclacitinib
19	Toy poodle	2	СМ	4.2	14	1,338	4	632	Prednisolone
20	West Highland white terrier	9	Μ	8	46	2,195	17	177	Oclacitinib
21	Shiba inu	1	М	8.1	4	2,540			
22	Shiba inu	3	Μ	11	65	7,152			
23	Shiba inu	8	F	10.5	34	2,040			
24	Shiba inu	8	Μ	24	103	6,372			
25	Shiba inu	10	М	9	46	3,626			
26	Shiba inu	11	F	9	15	1,553			
27	Shiba inu	13	F	8	28	1,443			
28	Shiba inu	14	SF	15.2	68	2,153			
29	Shiba inu	15	SF	7.4	69	6,816			
30	Mixed breed	3	SF	5.7	10	3,261			
31	Mixed breed	3	СМ	19	18	1,300			
32	Mixed breed	5	F	9	42	769			
33	Mixed breed	6	СМ	6.8	20	2,057			
34	Mixed breed	6	CM	11	44	5,078			
35	Mixed breed	8	М	8.6	59	1,287			
36	Border collie	10	СМ	17.4	36	3,760			
37	French bulldog	1	SF	8	4	945			
38	Labrador retriever	11	CM	33	24	1,535			
39	Samoyed	7	SF	23	15	2,227			

CM, castrated male; F, female; M, male; SF, spayed female.

ELISA

The anti-TARC monoclonal antibody with carbonate buffer (0.25 µg/ mL) was immobilized on ELISA plates (Thermo Scientific Nunc, 439454, Thermo Fisher Scientific) in 100 $\mu L,$ and incubated at 4°C overnight. ELISA plates were washed using PBST (Phosphate buffered saline containing 0.05% Tween-20), 100 µL of sera diluted two-fold with a blocking buffer (four-fold Block Ace in PBST, UK-B80, Yukijirushi; Tokyo, Japan) was added and then plates were kept at room temperature for 2 h. After washing with washing buffer, the anti-TARC polyclonal antibody, 400-fold diluted with blocking buffer, was added in 100 μL increments and the plates were kept at room temperature for 2 h. After washing, horseradish peroxidase (HRP)conjugated goat anti-rabbit IgG (656120, Invitrogen; Carlsbad, CA, USA), 2,000-fold diluted with blocking buffer, was added to the plates, which were then kept at room temperature for 2 h. After washing, 3,3-,5,5-tetramethylbenzidine (3405-100TAB, Sigma-Aldrich, St Louis, MO, USA) solution was added to the plates in 100 μL increments and plates were then kept at room temperature for 10 min. To stop the reaction, 50 μL of 2N H_2SO_4 was added to

the plates. Plates were read at a dual wavelength of 450–570 nm with a micro plate leader (model 680, 168-1000, Bio-Rad; Hercules, CA, USA). The assay was performed in duplicate on the same day. Diluted recombinant TARC protein from 98 to 50,000 pg/mL, instead of sera, was added to the plate in duplicate to prepare a calibration curve. Optical density values were increased proportionally with concentrations of the recombinant canine TARC ($R^2 = 0.9961$, P < 0.001; Figure S1). The workflow of ELISA is shown in Figure S2.

Statistical analysis

Serum TARC concentrations were measured as continuous variables. The results of serum TARC concentrations were represented as medians with interquartile ranges (IQR: 25–75th percentiles) owing to the non-normal data distribution. Differences in serum TARC concentrations as the dependent variables were analysed using the Wilcoxon–Mann–Whitney U-test for comparisons between the independent variables of group – dogs with cAD and healthy dogs. The Wilcoxon signed rank test was used to compare paired serum TARC concentrations before and after treatment. Similarly,

Table 2. Breed, age, sex,	weight and serum thy	mus and activation-regulated ch	hemokine (TARC) concentrations in h	ealthy dogs
---------------------------	----------------------	---------------------------------	-------------------------------------	-------------

Case	Breed	Age (years)	Sex	Weight (kg)	Serum TARC concentration (pg/mL)
1	Shiba inu	0.6	Μ	6.4	1,330
2	Shiba inu	1	CM	7.9	180
3	Shiba inu	1	CM	13	205
4	Shiba inu	2	SF	11.2	183
5	Shiba inu	2	SF	13.2	416
6	Shiba inu	2	SF	11.5	117
7	Shiba inu	3	SF	8.7	283
8	Shiba inu	4	M	8	249
9	Shiba inu	6	F	9.2	0
10	Shiba inu	6	SF	8.5	150
11	Shiba inu	6	M	10.5	2,243
12	Shiba inu	6	M	10.2	123
13	Shiba inu	7	F	6.6	0
14	Shiba inu	7	SF	12.9	409
14	Shiba inu	7	SF	9.9	192
16	Shiba inu	8	SF	12.3	164
17	Shiba inu	8	M	8.7	343
18	Shiba inu	9	SF	10.3	201
19	Shiba inu	11	CM	10.3	157
20	Shiba inu	12	SF	9.7	614
20 21	Shiba inu	12	CM	9.7	146
22		2	M	10	164
22	Beagle	3	F	10	245
23 24	Beagle	3	F	10	178
	Beagle	3	F	10	
25 26	Beagle	3		10	194
	Beagle		M		132
27	Beagle	3	M	10	149
28	Beagle		M F	10	124
29	Beagle	4	F	10	189
30	Beagle	4		10	229
31	Beagle	4	M	10	281
32	Beagle	5	M	10	322
33	Beagle	9	F	10	146
34	Golden retriever	3	CM	33.6	346
35	Golden retriever	4	SF	28	205
36	Golden retriever	8	SF	24.5	250
37	Golden retriever	9	SF	29	172
38	Golden retriever	10	SF	28.7	127
39	Cavalier King Charles spaniel	5	SF	8.5	0
40	French bulldog	1	F	12.8	391
41	Shetland sheepdog	8	CM	13.5	122
42	Toy poodle	5	CM	3.4	82

CM, castrated male; F, female; M, male; SF, spayed female.

CADESI-04 scores were collected as continuous variables. The relationship between serum TARC concentrations and CADESI-04 scores was analysed by the Spearman rank correlation test. A value of P < 0.05 was considered to be significant. Statistical analyses were performed using PRISM 8 software (GraphPad; San Diego, CA, USA).

Results

Animals and clinical data

Forty-two healthy dogs (male:female ratio 18:24; median age 4.5 years; median weight 10.0 kg) and 39 dogs with cAD (17:22; 8 years; 9.0 kg) were recruited. No significant differences were observed in age, sex or weight between healthy and atopic dogs (data not shown). In both groups, Shiba inus accounted for approximately half of the cases (21 of 42 healthy dogs, 19 of 39 atopic dogs). Index scores in 39 atopic dogs ranged between 4 and 139 (median 40) on the CADESI-04 scale, which has a

maximum score of $180.^{31}$ The severity of cAD included mild (CADESI-04 index 0–34, 17 dogs), moderate (35–59, 15 dogs) or severe (≥60, seven dogs) according to the CADESI-04 severity categories.³¹ CADESI-04 scores of all healthy dogs were 0. Twenty dogs with cAD were treated with prednisolone (11 dogs) or oclacitinib (nine dogs). The median CADESI-04 score was 41 (range 7–139) before treatment and decreased to 11 (range 0–117) after treatment. Breed, age, sex, weight, treatment and CADESI-04 scores at baseline and after treatment in dogs with cAD are listed in Table 1. The breed, age, sex and weight of healthy dogs are shown in Table 2.

Serum TARC concentrations in dogs with cAD and healthy dogs

Serum TARC concentrations were >10-fold higher in dogs with cAD than in healthy dogs. The median serum TARC concentration of dogs with cAD was 2,227 pg/mL (IQR



Figure 1. Serum thymus and activation-regulated chemokine (TARC) concentrations in dogs with atopic dermatitis (cAD) (n = 39) and healthy dogs (n = 42).

Serum TARC concentrations were significantly higher in dogs with atopic dermatitis than in healthy controls (P < 0.001). The lines within the boxes indicate the median serum TARC concentrations and the upper and lower boundaries of the boxes represent the 25th and 75th percentiles, respectively. ***P < 0.001 by the Wilcoxon–Mann–Whitney U-test.

1,556–3,910), whereas that of healthy dogs was 183 pg/ mL (IQR 146–273). Serum TARC concentrations were significantly higher in dogs with cAD than in healthy dogs (P < 0.001; Figure 1).

Relationship between serum TARC concentrations and CADESI-04 scores in dogs with cAD

In order to assess the utility of serum TARC concentrations as a marker for clinical severity, we examined the relationship between serum TARC concentrations and CADESI-04 scores. Serum TARC concentrations in dogs with cAD correlated with CADESI-04 scores ($\rho = 0.457$, P < 0.01; Figure 2).

Serum TARC concentrations before and after treatment in dogs with cAD

We measured serum TARC concentrations in 20 dogs with cAD before and after systemic treatment with oral prednisolone or oclacitinib. In all 20 dogs, serum TARC concentrations significantly decreased after four weeks of treatment in accordance with the amelioration of skin lesions (P < 0.001). Serum TARC concentrations decreased from 2,431 pg/mL (IQR 1,713–4,147) before treatment to 598 pg/mL (IQR 237–1,726) after treatment (Figure 3).



Figure 2. Relationship between serum thymus and activation-regulated chemokine (TARC) concentrations and Canine Atopic Dermatitis Extent and Severity Index, 4th iteration (CADESI-04) scores in dogs with atopic dermatitis (n = 39).

Serum TARC concentrations in dogs positively correlated with CADESI-04 scores ($\rho = 0.457$, P < 0.01). **P < 0.01 by the Spearman rank correlation test.

Relationship between changes in serum TARC concentrations and CADESI-04 scores

In order to assess the utility of serum TARC concentrations as a marker for treatment responses, we examined the relationship between changes in TARC concentrations and CADESI-04 scores after treatment. Changes in serum TARC concentrations correlated with those in CADESI-04 scores (p = 0.746, P < 0.001; Figure 4).

Discussion

To the best of the authors' knowledge, this is the first study to investigate the relationship between serum TARC concentrations and the disease severity of cAD. To evaluate the usefulness of serum TARC concentrations as a biomarker for cAD, we established a sandwich ELISA system for quantifying canine TARC. Previous studies demonstrated that plasma and serum TARC concentrations in human patients with AD correlated with disease severity.^{19–23} Serum TARC concentrations were found to be markedly higher than plasma TARC concentrations in patients with AD because platelets in serum released large amounts of TARC.¹⁹ Furthermore, serum TARC concentrations were stable independent of the freeze-thaw process, whereas those in plasma contaminated with platelets were elevated after freeze-thaw due to the release of TARC.³⁴ A meta-analysis proposed serum TARC concentrations as the most reliable biomarker for the disease severity of human AD.35 Therefore, herein we used serum samples to quantify canine TARC.

The results obtained showed that serum TARC concentrations were >10-fold higher in dogs with cAD than in healthy controls, which is consistent with previous findings on human AD.^{19,22} The median serum TARC concentrations in dogs with cAD and healthy controls were 2,227 pg/mL (IQR 1,556–3,910) and 183 pg/mL (IQR 146–273), respectively. These concentrations closely corresponded to previously reported values from human



Figure 3. Serum thymus and activation-regulated chemokine (TARC) concentrations before and after treatment in dogs with atopic dermatitis (cAD) (n = 20).

Serum TARC concentrations showed a significant decrease after four weeks of treatment with either oral prednisolone (n = 11) or oclacitinib (n = 9) (P < 0.001). ***P < 0.001 by the Wilcoxon–Mann–Whitney U-test.

counterparts in the literature with an average serum TARC concentration of 1,480 pg/mL (IQR 640-3,540) in patients with AD and 250 pg/mL (IQR 240-260) in healthy controls.²⁵ However, a previous study also showed that a high serum TARC concentration was not specific to human AD. An elevated serum TARC concentration also has been reported in human patients with other skin diseases, such as bullous pemphigoid, scabies, polymorphic prurigo, cutaneous T-cell lymphoma, drug eruption, pustular dermatosis and other internal disorders.²⁴ Based on these findings, serum TARC concentrations have not been solely used to establish a diagnosis of human AD. Thus, additional studies that evaluate TARC concentrations in dogs with other diseases, besides cAD, are needed to assess the effect of other inflammatory or neoplastic skin conditions on TARC concentrations.

In the present study, serum TARC concentrations in two healthy dogs were >1,000 pg/mL (cases 1 and 11; Table 2). This anomaly may be partially due to an age-related difference in TARC concentrations. One of the two dogs with a high serum TARC concentration (Case 1) was only 7 months old; the mean age of the healthy controls with lower serum TARC concentrations was 4.5 years. A previous study on healthy human subjects reported that serum TARC concentrations were higher in infants (<1



Figure 4. Relationship between changes in serum thymus and activation-regulated chemokine (TARC) concentrations and Canine Atopic Dermatitis Extent and Severity Index, 4th iteration (CADESI-04) scores (n = 20).

Changes in serum TARC concentrations before and after treatment with either oral prednisolone (n = 11) or oclacitinib (n = 9) positively correlated with the CADESI-04 scores ($\rho = 0.746$, P < 0.001). ***P < 0.001 by the Spearman rank correlation test.

year old) than in children (>2-5 and 6-16 years old) and adults (>16 years old).³⁶ The same study also found that to achieve the same diagnostic accuracy of human AD, different cut-off values were required for different age groups.³⁶ Because serum TARC concentrations in healthy human subjects differ among infants, children and adults, healthy thresholds also vary according to age, based on current evidence.²⁴ Therefore, further studies are needed to investigate age-related differences in serum TARC concentrations in healthy and affected dogs in order to establish appropriate reference ranges. Additionally, it is important to note that one (Case 1) of these two outliers subsequently developed the typical clinical signs of cAD, suggesting that TARC concentrations became elevated before the onset of cAD. This hypothesis is supported by previous findings indicating the potential of serum TARC concentrations to detect subclinical dermal inflammation or the early stages of human AD.37,38 Because the remaining outlier (Case 11) was lost to follow-up, we were unable to confirm whether this subject eventually developed cAD or another disease that may have caused an elevated TARC concentration. Although it may be of clinical significance, it was not possible to confirm the utility of TARC concentrations in predicting the onset of cAD in the present study. Thus, further studies are necessary to evaluate that serum TARC concentrations would be a good marker to predict onset of cAD, which may lead to prophylactic administration of anti-inflammatory agents.

The present results showed that serum TARC concentrations correlated with CADESI-04 scores, suggesting the potential of TARC as a useful objective measure for the disease severity of cAD, similar to human AD.^{20–22} In previous studies on human AD, among available markers, including total IgE concentrations, peripheral eosinophil counts and serum LDH concentrations, serum TARC more strongly correlated with clinical scores.^{24,25} In

Asahina et al.

studies on cAD, although total IgE concentrations were significantly higher in dogs with cAD than in healthy controls, they did not correlate with clinical severity according to the Canine Atopic Dermatitis Lesion Index.² Furthermore, the lesional severity scores of dogs with cAD did not correlate with the serum concentrations of other cytokines, including IL-17, IL-31 and macrophage migration inhibitory factor.³⁹ Therefore, serum TARC concentrations appear to be a more accurate marker than previously studied biomarkers and correspond well with the lesional severity of cAD.

In the present study, we also investigated whether serum TARC concentrations reflected therapeutic responses. To the best of our knowledge, a serum biomarker has not yet been used to assess spontaneous cAD before and after treatment in individual subjects. We demonstrated that serum TARC concentrations decreased with corresponding improvements in clinical signs in all dogs treated with prednisolone or oclacitinib, both of which are first-line treatments in the management of cAD exacerbations. We also found a strong correlation between changes in TARC concentrations and CADESI-04 scores after treatment. The present results show the potential of serum TARC concentrations as an objective marker for evaluating therapeutic responses in cAD treated with prednisolone or oclacitinib.

The comparison of TARC concentrations before and after treatment with prednisolone and oclacitinib would have been clinically interesting. However, because dogs were not randomized into each group, there was a risk of allocation bias leading to inaccurate interpretation. A previous study using a mouse model of allergic dermatitis reported that oclacitinib reduced the TARC concentration in skin.⁴⁰ In this model, oclacitinib also suppressed IL-31, $\mathsf{TNF-}\alpha$ and TSLP, although abrupt withdrawal led to a rapid increase of these cytokines. 40,41 TNF- $\!\alpha$ has been shown to be the mediator inducing TARC release from canine keratinocytes.^{42,43} Thus, oclacitinib may reduce serum TARC concentrations by inhibiting TNF- α in keratinocytes. Common treatment regimens for cAD also may include the use of topical glucocorticoids, oral ciclosporin or lokivetmab injections, which were not evaluated in the present study. In human patients with AD, serum TARC concentrations decreased after the treatments with topical glucocorticoids^{19,22,26} or oral ciclosporin.^{20,25} Additionally, an in vitro study revealed that IL-31 induced TARC in human keratinocytes.44 These studies suggest that serum TARC has the potential to act as a marker for evaluating therapeutic responses in cAD treated with other drugs, although the generalizability of our results cannot be assumed for these drugs. Therefore, specific interactions between serum TARC concentrations and other treatment modalities require further investigation.

In conclusion, serum TARC concentrations correlated with the lesional severity scores of cAD, indicating that serum TARC concentrations are an objective marker for assessing disease severity and therapeutic responses in cAD. The assessment of disease severity and efficacy of treatment for cAD has relied on traditional markers susceptible to various confounders. The use of TARC as a biomarker will allow standardized evaluations not only in clinical settings, but also importantly in research settings, thereby facilitating the cross-comparison of trials and its use as a surrogate end-point. Further studies are needed to confirm the potential of serum TARC concentrations as an objective and predictive marker in the treatment monitoring of cAD.

Acknowledgements

The authors would like to thank Takahiro Akita (Akita Animal Hospital), Daisuke Itsukaichi (Poplar Animal Hospital), Chiaki Kitanaka (SENA Animal Hospital Rakuhoku Animal Wellness Center), Jin Kozakai (Ai Animal Hospital Kashima), Yuko Machida (AZ Animal Hospital), Hisanori Mutoh (Miyaki Animal Clinic), Hiromi Oboso (Cookie Animal Hospital), Taichi Oshima (Taichi Animal Hospital) and Akihiro Sugeno (Sugeno Animal Hospital) for collecting blood samples.

References

- 1. Marsella R, De Benedetto A. Atopic dermatitis in animals and people: an update and comparative review. *Vet Sci* 2017; 4: 37.
- Chaudhary SK, Singh SK, Kumari P, et al. Alterations in circulating concentrations of IL-17, IL-31 and total IgE in dogs with atopic dermatitis. *Vet Dermatol* 2019; 30: 383-e114.
- Hayashiya S, Tani K, Morimoto M, et al. Expression of T helper 1 and T helper 2 cytokine mRNAs in freshly isolated peripheral blood mononuclear cells from dogs with atopic dermatitis. *J Vet Med A Physiol Pathol Clin Med* 2002; 49: 27–31.
- Lian TM, Halliwell RE. Allergen-specific IgE and IgGd antibodies in atopic and normal dogs. *Vet Immunol Immunopathol* 1998; 66: 203–223.
- Majewska A, Gajewska M, Dembele K, et al. Lymphocytic, cytokine and transcriptomic profiles in peripheral blood of dogs with atopic dermatitis. *BMC Vet Res* 2016; 12: 174.
- Nuttall TJ, Knight PA, McAleese SM, et al. Expression of Th1, Th2 and immunosuppressive cytokine gene transcripts in canine atopic dermatitis. *Clin Exp Allergy* 2002; 32: 789–795.
- Olivry T, Dean GA, Tompkins MB, et al. Toward a canine model of atopic dermatitis: amplification of cytokine–gene transcripts in the skin of atopic dogs. *Exp Dermatol* 1999; 8: 204–211.
- Schlotter YM, Rutten VP, Riemers FM, et al. Lesional skin in atopic dogs shows a mixed Type-1 and Type-2 immune responsiveness. *Vet Immunol Immunopathol* 2011; 143: 20–26.
- Martins GC, de Oliveira Melo Júnior OA, Botoni LS, et al. Clinical-pathological and immunological biomarkers in dogs with atopic dermatitis. *Vet Immunol Immunopathol* 2018; 205: 58–64.
- Iio A, Motohashi T, Kunisada T, et al. Preferential gene transcription of T helper 2 cytokines in peripheral CCR4(+) CD4(+) lymphocytes in dogs. *Vet Dermatol* 2014; 25: 199–e150.
- Maeda S, Tsukui T, Saze K, et al. Production of a monoclonal antibody to canine thymus and activation-regulated chemokine (TARC) and detection of TARC in lesional skin from dogs with atopic dermatitis. *Vet Immunol Immunopathol* 2005; 103: 83–92.
- Maeda S, Ohmori K, Yasuda N, et al. Increase of CC chemokine receptor 4-positive cells in the peripheral CD4 cells in dogs with atopic dermatitis or experimentally sensitized to Japanese cedar pollen. *Clin Exp Allergy* 2004; 34: 1,467–1,473.
- Maeda S, Fujiwara S, Omori K, et al. Lesional expression of thymus and activation-regulated chemokine in canine atopic dermatitis. *Vet Immunol Immunopathol* 2002; 88: 79–87.
- Maeda S, Okayama T, Omori K, et al. Expression of CC chemokine receptor 4 (CCR4) mRNA in canine atopic skin lesion. *Vet Immunol Immunopathol* 2002; 90: 145–154.
- Maeda S, Maeda S, Ohno K, et al. Protease-activated receptor-2 induces proinflammatory cytokine and chemokine gene expression in canine keratinocytes. *Vet Immunol Immunopathol* 2013; 153: 17–25.

- Marsella R, Olivry T, Maeda S. Cellular and cytokine kinetics after epicutaneous allergen challenge (atopy patch testing) with house dust mites in high-IgE beagles. *Vet Dermatol* 2006; 17: 111–120.
- Olivry T, Mayhew D, Paps JS, et al. Early activation of Th2/Th22 Inflammatory and pruritogenic pathways in acute canine atopic dermatitis skin lesions. *J Invest Dermatol* 2016; 136: 1,961– 1,969.
- Murray C, Ahrens K, Devalaraja M, et al. Use of a canine model of atopic dermatitis to investigate the efficacy of a CCR4 antagonist in allergen-induced skin inflammation in a randomized study. *J Invest Dermatol* 2016; 136: 665–671.
- Fujisawa T, Fujisawa R, Kato Y, et al. Presence of high contents of thymus and activation-regulated chemokine in platelets and elevated plasma levels of thymus and activation-regulated chemokine and macrophage-derived chemokine in patients with atopic dermatitis. *J Allergy Clin Immunol* 2002; 110: 139–146.
- Hijnen D, De Bruin-Weller M, Oosting B, et al. Serum thymus and activation-regulated chemokine (TARC) and cutaneous T cell- attracting chemokine (CTACK) levels in allergic diseases: TARC and CTACK are disease-specific markers for atopic dermatitis. *J Allergy Clin Immunol* 2004; 113: 334–340.
- Jahnz-Rozyk K, Targowski T, Paluchowska E, et al. Serum thymus and activation-regulated chemokine, macrophage-derived chemokine and eotaxin as markers of severity of atopic dermatitis. *Allergy* 2005; 60: 685–688.
- Kakinuma T, Nakamura K, Wakugawa M, et al. Thymus and activation-regulated chemokine in atopic dermatitis: Serum thymus and activation-regulated chemokine level is closely related with disease activity. J Allergy Clin Immunol 2001; 107: 535–541.
- Morita A, Kikuoka S, Horikawa T, et al. Evaluation of human thymus and activation-regulated chemokine concentrations in blood using a new sandwich ELISA based on monoclonal antibodies. *Clin Chim Acta* 2002; 322: 67–75.
- Kataoka Y. Thymus and activation-regulated chemokine as a clinical biomarker in atopic dermatitis. *J Dermatol* 2014; 41: 221– 229.
- Kou K, Aihara M, Matsunaga T, et al. Association of serum interleukin-18 and other biomarkers with disease severity in adults with atopic dermatitis. *Arch Dermatol Res* 2012; 304: 305–312.
- Yasukochi Y, Nakahara T, Abe T, et al. Reduction of serum TARC levels in atopic dermatitis by topical anti-inflammatory treatments. *Asian Pac J Allergy Immunol* 2014; 32: 240–245.
- Terada Y, Nagata M, Murayama N, et al. Clinical comparison of human and canine atopic dermatitis using human diagnostic criteria (Japanese Dermatological Association, 2009): proposal of provisional diagnostic criteria for canine atopic dermatitis. *J Dermatol* 2011; 38: 784–790.
- Favrot C, Steffan J, Seewald W, et al. A prospective study on the clinical features of chronic canine atopic dermatitis and its diagnosis. *Vet Dermatol* 2010; 21: 23–31.
- Okayama T, Matsuno Y, Yasuda N, et al. Establishment of a quantitative ELISA for the measurement of allergen-specific IgE in dogs using anti-IgE antibody cross-reactive to mouse and dog IgE. *Vet Immunol Immunopathol* 2011; 139: 99–106.
- Yuki M, Aoyama R, Nakagawa M, et al. A clinical investigation on serum Amyloid A concentration in client-owned healthy and diseased cats in a primary care animal hospital. *Vet Sci* 2020; 7: E45.
- Olivry T, Saridomichelakis M, Nuttall T, et al. Validation of the Canine Atopic Dermatitis Extent and Severity Index (CADESI)-4, a simplified severity scale for assessing skin lesions of atopic dermatitis in dogs. *Vet Dermatol* 2014; 25: 77–85, e25.

- Gadeyne C, Little P, King VL, et al. Efficacy of oclacitinib (Apoquel®) compared with prednisolone for the control of pruritus and clinical signs associated with allergic dermatitis in clientowned dogs in Australia. *Vet Dermatol* 2014; 25: 512–518, e86.
- Katayama M, Igarashi H, Tani K, et al. Effect of multiple oral dosing of fluconazole on the pharmacokinetics of cyclosporine in healthy beagles. J Vet Med Sci 2008; 70: 85–88.
- Zhao X, Delgado L, Weiner R, et al. Influence of pre-analytical factors on thymus- and activation-regulated chemokine quantitation in plasma. *J Circ Biomark* 2015; 4: 10.
- Thijs J, Krastev T, Weidinger S, et al. Biomarkers for atopic dermatitis: a systematic review and meta-analysis. *Curr Opin Allergy Clin Immunol* 2015; 15: 453–460.
- Fujisawa T, Nagao M, Hiraguchi Y, et al. Serum measurement of thymus and activation-regulated chemokine/CCL17 in children with atopic dermatitis: elevated normal levels in infancy and agespecific analysis in atopic dermatitis. *Pediatr Allergy Immunol* 2009; 20: 633–641.
- Furue M, Matsumoto T, Yamamoto T, et al. Correlation between serum thymus and activation-regulated chemokine levels and stratum corneum barrier function in healthy individuals and patients with mild atopic dermatitis. *J Dermatol Sci* 2012; 66: 60–63.
- Shoda T, Futamura K, Kobayashi F, et al. Expression of thymus and activation-regulated chemokine (TARC) by human dermal cells, but not epidermal keratinocytes. *J Dermatol Sci* 2014; 76: 90–95.
- Gow DJ, Jackson H, Forsythe P, et al. Measurement of serum macrophage migration inhibitory factor (MIF) and correlation with severity and pruritus scores in client owned dogs with atopic dermatitis. *Vet Dermatol* 2019; 30: 115–e32.
- Fukuyama T, Ehling S, Cook E, et al. Topically administered janus-kinase inhibitors tofacitinib and oclacitinib display impressive antipruritic and anti-inflammatory responses in a model of allergic dermatitis. *J Pharmacol Exp Ther* 2015; 354: 394–405.
- Fukuyama T, Ganchingco JR, Bäumer W. Demonstration of rebound phenomenon following abrupt withdrawal of the JAK1 inhibitor oclacitinib. *Eur J Pharmacol* 2017; 794: 20–26.
- Banovic F, Olivry T, Bäumer W, et al. Diluted sodium hypochlorite (bleach) in dogs: antiseptic efficacy, local tolerability and in vitro effect on skin barrier function and inflammation. *Vet Dermatol* 2018; 29: 6–e5.
- Shibata S, Maeda S, Kondo N, et al. Identification of the signaling pathway of TNF-alpha-induced CCL17/TARC transcription in a canine keratinocyte cell line. *Vet Immunol Immunopathol* 2011; 139: 90–98.
- Dillon SR, Sprecher C, Hammond A, et al. Interleukin 31, a cytokine produced by activated T cells, induces dermatitis in mice. *Nat Immunol* 2004; 5: 752–760.

Supporting Information

Additional Supporting Information may be found in the online version of this article.

Figure S1. A standard curve of the sandwich ELISA with serial dilutions of purified recombinant canine TARC ranging from 98 to 50,000 pg/mL.

Figure S2. The workflow of the sandwich ELISA assay.

Résumé

Contexte – La TARC/CCL17 (thymus and activation-regulated chemokine) a été impliquée dans la pathogénie de la dermatite atopique canine (cAD). Les concentrations sériques de TARC sont un biomarqueur fiable chez l'homme atopique ; cependant, leur potentiel en tant que biomarqueur pour cAD n'a pas été étudié.

Hypothèses/Objectifs – Etudier si les concentrations sériques de TARC corrèlent avec la sévérité de la maladie et les réponses thérapeutiques pour cAD.

Sujets – Trente – neuf chiens avec cAD et 42 chiens sains ont été inclus.

Matériels et méthodes – Les concentrations sériques en TARC chez les chiens avec cAD et les chiens sains ont été mesurées par ELISA avec anticorps anti-TARC canin. La sévérité clinique de cAD a été notée par CADESI-04 (Canine Atopic Dermatitis Extent and Severity Index, 4th iteration). Les concentrations de TARC sériques ont été comparées entre les chiens avec cAD et les chiens sains contrôles, et leur relation avec CADESI-04 a été étudiée. Les concentrations sériques de TARC ont aussi été mesurées chez 20 chiens avec cAD traités par prednisolone ou oclacitinib pendant quatre semaines.

Résultats – Les concentrations sériques de TARC étaient significativement plus élevées chez les chiens avec cAD que chez les chiens sains (P < 0.001). Chez les chiens avec cAD, les concentrations sériques de TARC corrélaient avec les scores de CADESI-04 ($\rho = 0.457$, P < 0.01). En outre, les concentrations sériques de TARC diminuaient significativement chez les chiens traités avec atténuation des signes cliniques (P < 0.001). Les changements de concentrations sériques de TARC avant et après traitement corrélaient avec ceux des scores CADESI-04 ($\rho = 0.746$, P < 0.001).

Conclusions et importance clinique – Les concentrations sériques de TARC ont le potentiel d'être un outil clinique et de recherche pour l'évaluation objective de l'intensité de la maladie et de la réponse thérapeutique de la CAD.

RESUMEN

Introducción – el timo y la quimiocina regulada por activación (TARC/CCL17) se han implicado en la patogénesis de la dermatitis atópica canina (cAD). Las concentraciones séricas de TARC son un biomarcador fiable en la dermatitis atópica humana; sin embargo, no se ha investigado su potencial como biomarcador en cAD.

Hipótesis/Objetivos – investigar si las concentraciones séricas de TARC se correlacionan con la gravedad de la enfermedad y las respuestas terapéuticas en cAD.

Animales – se incluyeron treinta y nueve perros con cAD y 42 perros sanos.

Métodos y materiales – Las concentraciones séricas de TARC en perros con cAD y perros sanos se midieron mediante ELISA sándwich con anticuerpos anti-TARC caninos. La gravedad clínica de la cAD se valoró utilizando el índice validado de extensión y gravedad de la dermatitis atópica canina, cuarta versión (CADESI-04). Se compararon las concentraciones de TARC en suero entre perros con cAD y controles sanos, y se examinó su relación con CADESI-04. También se midieron las concentraciones séricas de TARC en 20 perros con cAD tratados con prednisolona u oclacitinib durante cuatro semanas.

Resultados – las concentraciones séricas de TARC fueron significativamente más altas en perros con cAD que en perros sanos (P < 0,001). En perros con cAD, las concentraciones séricas de TARC se correlacionaron con las puntuaciones de CADESI-04 ($\rho = 0,457$, P < 0,01). Además, las concentraciones séricas de TARC disminuyeron significativamente en los perros tratados con la disminución de los signos clínicos (P < 0,001). Los cambios en las concentraciones séricas de TARC antes y después del tratamiento se correlacionaron con los de los valores de CADESI-04 ($\rho = 0,746$, P < 0,001).

Conclusiones y relevancia clínica – las concentraciones séricas de TARC tienen potencial como herramienta clínica y de investigación para la evaluación objetiva de la gravedad de la enfermedad y las respuestas terapéuticas en cAD.

Zusammenfassung

Hintergrund – Dem Thymus und Activation-regulated Chemokine (TARC/CCL17) wird in der Pathogenese der caninen atopischen Dermatitis (cAD) eine Rolle zugeschrieben. Serum TARC Konzentrationen sind ein verlässlicher Biomarker bei der atopischen Dermatitis des Menschen; ihr Potential als Biomarker für die cAD wurde jedoch noch nicht untersucht.

Tiere – Neununddreißig Hunde mit cAD und 42 gesunde Hunde wurden rekrutiert.

Methoden und Materialien – Serum TARC Konzentrationen bei Hunden mit cAD und bei gesunden Hunden wurden mittels Sandwich ELISA mit anti-caninen TARC Antikörpern gemessen. Der klinische Schweregrad der cAD wurde mittels validiertem Canine Atopic Dermatitis Extent and Severity Index, 4te Ausgabe (CADESI-04) beurteilt. Die Serum TARC Konzentrationen wurden zwischen den Hunden mit cAD und den gesunden Kontrollen verglichen und ihre Beziehung mittels CADESI-04 untersucht. Es wurden auch die TARC Konzentrationen bei 20 Hunden mit cAD, die mit Prednisolon oder Oclacitinib vier Wochen lang behandelt worden waren, gemessen.

Ergebnisse – Die Serum TARC Konzentrationen waren bei Hunden mit cAD signifikant höher als bei den gesunden Hunden (P < 0,001). Bei Hunden mit cAD korrelierten die Serum TARC Konzentrationen mit den CADESI-04 Werten ($\rho = 0.746$, P < 0.001). Weiters nahmen die Serum TARC Konzentrationen bei behandelten Hunden mit Abnahme der klinischen Zeichen ab (P < 0,001). Veränderungen bei den Serum TARC Konzentrationen vor und nach der Behandlung korrelierten mit denen der CADESI-04 Werte ($\rho = 0.746$, P < 0.001).

Schlussfolgerungen und klinische Bedeutung – Die Serum TARC Konzentrationen haben Potential als klinisches und wissenschaftliches Werkzeug zur objektiven Evaluierung der Schwere der Erkrankung und der therapeutischen Resonanz bei cAD.

要約

背景 – 胸腺および活性化調節ケモカイン(TARC / CCL17)は、大アトピー性皮膚炎(cAD)の病因に関 与している。血清TARC濃度は、ヒトアトピー性皮膚炎の信頼できるバイオマーカーである。ただし、 cADに対するバイオマーカーとしての可能性は調査されていない。

仮説/目的-本研究の目的は、血清TARC濃度が疾患の重症度およびcADの治療反応と相関するかどうかを 調査することであった。

供試動物 - cAD犬39頭と健常犬42頭を募った。

材料と方法 - cAD犬および健常犬の血清TARC濃度を、抗犬TARC抗体を用いたサンドイッチELISAで測定した。 cADの臨床的重症度は、検証済みの犬アトピー性皮膚炎の程度および重症度指数、4thインテレーション (CADESI-04) を使用してスコア化された。cAD犬および健常犬間の血清TARC濃度を比較し、CADESI-04との関係を調査した。血清TARC濃度も、プレドニゾロンまたはオクラシチニブで4週間治療されたcADの犬20頭で測定した。

結果 – 血清TARC濃度は、健常犬よりもcAD犬で有意に高かった(P<0.001)。 cAD犬では、血清TARC濃 度がCADESI-04スコアと相関していた(q= 0.457、P<0.01)。さらに、治療された犬の血清TARC濃度 は、臨床徴候の減衰とともに有意に減少した(P<0.001)。治療前後の血清TARC濃度の変化は、CADE-SI-04スコアの変化と相関していた(q= 0.746、P<0.001)。

結論と臨床的関連性 – 血清TARC濃度は、cADの疾患の重症度および治療反応を客観的に評価する臨床お よび研究ツールとしての可能性を秘めている。

摘要

背景 — 胸腺和活化调节趋化因子(TARC/CCL17)与犬异位性皮炎(cAD)的发病机制有关。血清TARC浓度是 人类特应性皮炎的可靠生物标志物;然而,尚未有研究报道其作为cAD生物标志物的潜力。

假设/目的 – 研究血清TARC浓度是否与cAD的疾病严重程度和治疗反应相关。

动物 — 招募了39只cAD犬和42只健康犬。

方法和材料 — 通过夹心ELISA和抗犬TARC抗体测定cAD犬和健康犬的血清TARC浓度。使用犬异位性皮炎 程度和严重指数第4版(CADESI-04)对cAD的临床严重程度进行评分。比较cAD犬和健康对照犬的血清TARC 浓度,并检查其与CADESI-04的关系。还测定了泼尼松龙或奥拉替尼治疗4周的20只cAD犬,测定了血清 TARC浓度。

结果 — cAD犬血清TARC浓度显著高于健康犬(P<0.001)。对于cAD犬,血清TARC浓度与CADESI-04评分相 关(ρ=0.457, P<0.01)。此外,治疗犬的血清TARC浓度显著降低,临床症状减轻(P<0.001)。治疗前后血清 TARC浓度变化与CADESI-04评分相关(ρ=0.746, P<0.001)。

结论和临床相关性 — 检测血清TARC浓度,有可能作为客观评价疾病严重程度和cAD治疗反应的临床和研究工具。

Resumo

Contexto – A quimiocina regulada pelo timo e ativação (TARC / CCL17) tem sido implicada na patogênese da dermatite atópica canina (DAC). As concentrações séricas de TARC são um biomarcador confiável para dermatite atópica humana; no entanto, seu potencial como biomarcador para CAD não foi investigado.

Hipótese/Objetivos – Investigar se as concentrações séricas de TARC se correlacionam com a gravidade da doença e as respostas terapêuticas para DAC.

Animais – Trinta e nove cães com DAC e 42 cães saudáveis foram recrutados.

Métodos e materiais – As concentrações séricas de TARC em cães com DAC e em cães saudáveis foram medidas por ELISA sanduíche com anticorpos anti-TARC caninos. A gravidade clínica da DAC foi avaliada usando o índice de extensão e gravidade da dermatite atópica canina validado, 4ª iteração (CADESI-04). As concentrações séricas de TARC foram comparadas entre cães com DAC e controles saudáveis, e sua relação com CADESI-04 foi avaliada. As concentrações séricas de TARC também foram mensuradas em 20 cães com DAC tratados com prednisolona ou oclacitinib por quatro semanas.

Resultados – As concentrações séricas de TARC foram significativamente maiores em cães com DAC do que em cães saudáveis (P < 0,001). Em cães com DAC, as concentrações séricas de TARC se correlacionaram com as pontuações de CADESI-04 ($\rho = 0,457$, P < 0,01). Além disso, as concentrações séricas de TARC diminuíram significativamente nos cães tratados, com a atenuação dos sinais clínicos (P < 0,001). Mudanças nas concentrações séricas de TARC antes e depois do tratamento correlacionaram-se com as dos escores CADESI-04 ($\rho = 0,746$, P < 0,001).

Conclusões e relevância clínica – As concentrações séricas de TARC têm potencial como uma ferramenta clínica e de pesquisa para a avaliação objetiva da gravidade da doença e das respostas terapêuticas para DAC.