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Original Article

Chemical compositions of the volatile extracts from seeds of *Dendranthema nankingense* and *Borago officinalis*

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ABSTRACT

Volatile extracts from the seeds of *Dendranthema nankingense* Hand.-Mazz. and *Borago officinalis* L. were prepared using simultaneous distillation and extraction, and analyzed with gas chromatography–mass spectrometry on two capillary gas chromatography columns of different polarity. Ninety-five volatile compounds were identified in *D. nankingense* seeds, with hexanal, benzeneacetaldehyde, borneol, (–)-camphor, and 3-methyl-1-butanol being the predominant species. Sixty-five volatile compounds were identified in *B. officinalis* seeds, with 2-pentanone, 2,3-dihydro-benzofuran, 3-methyl butanal, and hexanal being the most abundant species. Thirty-three compounds, including short-chain aliphatic aldehydes, alcohols, and ketones, were common to both seeds. The volatile composition of both seeds varied significantly depending on their respective origins. The volatile terpenoids borneol and (–)-camphor could be key bioactive contributors to the characteristic flavor and cooling effects of *D. nankingense*. For the first time, coumaran was identified as an abundant species in plant seeds.

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1. Introduction

Dendranthema nankingense Hand.-Mazz. is a perennial herb of the family Compositae, which can be classified into the category of pungent and cool nature according to traditional

Chinese medicine theories. It is harvested in the summer and used to relieve external syndromes induced by hot diseases [1]. Soup cooked with the leaves of *D. nankingense* is very popular in eastern China, as it has heat-clearing and detoxifying effects. People who have drunk the soup have reported

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that they clearly experienced a cooling and refreshing sensation throughout their mouths and throats.

Borago officinalis L. is an annual herbaceous plant of the Boraginaceae family, which is native to Europe and North Africa. This plant is currently cultivated throughout the world and is used for medicinal purposes, and in beverages and salads [2]. *B. officinalis* has been defined as a future “superfood” because of the high level of γ -linolenic acid present in its seeds [3]. It has received considerable attention because of the antioxidant and antiradical activities of its phenolic constituents [4].

Many plants with volatile smelling flavor have been used as traditional herbs or foods in many countries since ancient times [5,6]. The volatiles from plant seeds have also received considerable interest because of their chemical diversity and functional activities. For example, coriander seeds contain a volatile oil, used in a medicinal context for the treatment of indigestion, worms, rheumatism, and pain in the joints [7]. To date, there have been no reports in the literature regarding volatiles from the seeds of *D. nankingense*, although a number of papers have reported the volatile compositions of its leaves [8–10]. Research and development on the seeds of *B. officinalis* has progressed at a steady pace over the past 30 years. Previous studies have primarily focused on γ -linolenic acid and, to a lesser extent, on phenolic compounds. Despite a long history of use of *B. officinalis* seed oil in foods and medicine, there have been few published studies regarding the volatile compounds contained in the oil. To our knowledge, there has only been one paper reporting a very limited number of volatile compounds from *B. officinalis* seeds that were harvested in Tunisia [11]. Thus, nothing is known about the volatile composition of *B. officinalis* seeds from other areas, and how their origin might affect the composition of their volatile components.

This study provides a comprehensive investigation of the volatile components from the seeds of *D. nankingense* and *B. officinalis* harvested in China. Furthermore, the study compares the results with those in the literature for the same family, and discusses the functional applications of herbal seeds in food and medicine.

2. Materials and methods

2.1. Materials and chemicals

Seeds of *D. nankingense* and *B. officinalis* were provided by the Qiutian Seed Co., Ltd. (Nanjing, China) and the Winner Horticultural Seed Co., Ltd. (Shanghai, China), respectively.

Pentane and ether were purchased from CNW (Düsseldorf, Germany) and Sinopharm (Shanghai, China), respectively. Both solvents were distilled prior to use. Anhydrous sodium sulfate was obtained from Lingfeng (Shanghai, China). Methyl nonanoate was supplied by Fluka (Buchs, Switzerland). The *n*-alkanes (C7–C28) were purchased from Aladdin (Shanghai, China). Other standard volatile compounds were bought from Sigma-Aldrich China Co. (Shanghai, China).

2.2. Isolation of volatile oils

The volatile extracts were obtained from the seed using a simultaneous distillation and extraction (SDE) setup equipped

with Likens–Nickerson apparatus connected to a vacuum pump. The seeds (50 g) were initially cooled to 4°C and then ground using a commercial blender for 10 seconds. The seed powder was then mixed with double-distilled water (500 mL), and a solution of methyl nonanoate (50 μ g) in a 1:1.12 (v/v) pentane/ether mixture (1 mL) was added as an internal standard. The mixture was placed in a 2000-mL round-bottom flask, whereas the solvent mixture [50 mL of a 1:1.12 (v/v) pentane/ether] was added to a 250-mL round-bottom flask in the SDE equipment. The sample flask was heated to 52°C for distillation and extraction, respectively. After 1.5 hours of processing, the collected solvent distillate was dried over anhydrous sodium sulfate and concentrated at 42°C using a Vigreux column to a final volume of approximately 1 mL for gas chromatography–mass spectrometry (GC–MS) analysis. The extractions were performed in triplicate.

2.3. GC–MS and GC analysis

GC–MS analysis was carried out using a polar capillary DB–WAX column (polyethylene glycol, 30 m \times 0.25 mm i.d. \times 0.25 μ m film thickness; Agilent, Folsom, CA, USA) and a nonpolar HP–1 column (dimethyl polysiloxane, 30 m \times 0.25 mm i.d. \times 0.25 μ m film thickness; Agilent, USA) on a QP2010 GC–MS system (Shimadzu, Kyoto, Japan). The oven temperature was held at 40°C for 2 minutes, and then increased at a rate of 5°C/min to a final temperature of 250°C. The oven was then held at 250°C for 10 minutes. The temperatures of injector and ion source were set at 230°C and 200°C, respectively. GC–MS was operated at an ionization energy of 70 eV in the electron impact mode over a range of 33–650 amu. Helium was used as a carrier gas at a constant flow rate of 1.6 mL/min. The extract (0.3 μ L) was injected with an autosampler in splitless mode.

Retention indices were calculated according to the Kovats method using *n*-alkanes (C7–C28) as external references. The experimental retention indices of the volatiles were compared with data from the literature [12–14]. To identify the peaks in the mass spectra, we compared our mass spectra with mass spectral libraries (NIST/EPA/NIH 2.0, Wiley 7, and LIBTX) and with literature data [12], and also with authentic reference standards when available. Analyses were performed twice.

GC analysis was performed on a Shimadzu GC2010 system equipped with a flame ionization detector. The column and conditions of the temperatures were identical to those used in the GC–MS analysis. The relative percentage amounts of each identified compounds were calculated with the peak area normalization. Analyses were carried out twice. Approximate concentrations of the volatile compounds were calculated according to the internal standard method using methyl nonanoate.

3. Results and discussion

3.1. Components of volatile oil from the seeds of *D. nankingense*

SDE of *D. nankingense* seeds yielded a yellow oil (2.3%, w/w), from which 95 compounds were isolated and identified. The components are listed in Table 1. In the majority of cases, the

Table 1 – Volatile extract composition of *Dendranthema nankingense* seeds.

No.	Retention time on DB-WAX (min)	Compounds	RIs		Identification and MS ions (EI) ^a	Total area (%)	Approximate concentration (µg/mL extract) ^b
			DB-WAX	HP-1			
1	2.63	3-Methyl butanal	924	<700	++++; M+ 86 (16.67), 44	4.5	B
2	3.10	2-Ethylfuran	962	<700	++++; M+ 96 (32.46), 81	0.3	C
3	3.45	Pentanal	985	<700	++++; M+ 86 (6.59), 44	1.7	C
4	4.06	α -Pinene	1021	921	++++; M+ 136 (7.67), 93	0.2	D
5	4.78	2-Methylbutanoic acid ethyl ester	1059	841	++++; M+ 130 (2.03), 57	0.2	D
6	4.91	Camphene	1065	942	+++; M+ 136 (15.98), 93	0.4	C
7	5.41	Hexanal	1086	779	++++; M+ 100 (1.21), 44	19.8	A
8	5.59	2-Methyl-1-propanol	1093	<700	++++; M+ 74 (15.32), 43	1.1	C
9	5.81	β -Pinene	1102	967	++++; M+ 136 (8.83), 93	0.6	C
10	5.96	3-Pentanol	1110	<700	++++; M+ 88 (0.65), 59	0.4	C
11	6.19	Sabinene	1120	966	+++; M+ 136 (12.19), 93	1.9	C
12	6.27	2-Pentanol	1124	<700	++++; M+ 88 (1.03), 45	1.0	C
13	6.61	2-Butylfuran	1139	880	+++; M+ 124 (13.46), 81	0.1	D
14	6.84	1-Butanol	1148	<700	++++; M+ 74 (1.86), 56	0.1	D
15	7.25	2-Ethyl-hexanal	1165	937	+++; M+ 128 (2.17), 57	0.2	D
16	7.36	β -Myrcene	1169	984	+++; M+ 136 (5.24), 41	0.5	C
17	7.62	α -Terpinene	1178	1007	++++; M+ 136 (42.98), 121	0.1	D
18	7.80	2-Heptanone	1185	872	++++; M+ 114 (6.60), 43	0.3	C
19	8.11	D-limonene	1195	1032	++++; M+ 136 (23.41), 68	1.4	C
20	8.22	3-Methyl-2-butenal	1199	759	+++; M+ 84 (100.00), 55 (77.16)	0.1	D
21	8.30	1,8-Cineole	1202	1020	++++; M+ 154 (37.13), 43	1.0	C
22	8.45	3-Methyl-1-butanol	1208	721	++++; M+ 88 (0.61), 55	5.2	B
23	8.71	(E)- 2-Hexenal	1219	829	+++; M+ 98 (22.10), 41	0.4	C
24	9.12	2-Pentyl-furan	1236	981	+++; M+ 138 (18.35), 81	3.4	B
25	9.38	(Z)-Ocimene	1246	1025	+++; M+ 136 (5.87), 93	0.2	D
26	9.59	1-Pentanol	1254	759	++++; M+ 88 (1.16), 42	1.8	C
27	9.74	Styrene	1259	875	+++; M+ 104 (100.00), 103 (48.72)	0.5	C
28	10.06	β -Cymene	1271	998	+++; M+ 134 (26.37), 119	0.6	C
29	10.35	Terpinolen	1281	1078	+++; M+ 136 (62.05), 93	0.4	C
30	10.62	Octanal	1290	980	++++; M+ 128 (1.20), 43	0.3	C
31	11.07	2-Ethenyl-2-butenal	1307	n.d.	++; M+ 96 (43.07), 67	0.1	D
32	11.50	(Z)-2-Heptenal	1324	932	++++; M+ 112 (6.85), 41	0.8	C
33	11.66	2-Methyl-3-octanone	1331	1068	+++; M+ 142 (4.55), 43	0.1	D
34	11.92	6-Methyl-5-hepten-2-one	1341	n.d.	+++; M+ 126 (11.16), 43	0.1	D
35	12.32	1-Hexanol	1356	862	++++; M+ 102 (0.58), 56	3.4	B
36	13.12	(Z)- 3-Hexen-1-ol	1385	835	+++; M+ 100 (3.35), 67	0.1	D
37	13.38	Nonanal	1394	1083	++++; M+ 142 (1.01), 57	0.3	C
38	13.42	3-Octanol	1395	988	+++; M+ 130 (2.38), 59	0.1	D
39	13.70	(E)-3-Octen-2-one	1406	1014	+++; M+ 126 (14.38), 43	0.2	D
40	13.96	Thujone	1417	1101	+++; M+ 152 (6.57), 110	0.2	D
41	14.08	2-Octanol	1422	995	+++; M+ 130 (0.63), 45	0.1	D

EI = electron impact; GC–MS = gas chromatography–mass spectrometry; RI = retention index.

^a The reliability of the proposed identification has been indicated according to the following: +++++, mass spectrum and retention indices both on polar and nonpolar columns agreed with those of the standards, mass spectral libraries and literature; +++, mass spectrum and retention indices both on polar and nonpolar columns agreed with mass spectral libraries and the literature; ++, mass spectrum and retention index on a polar column agreed with the values from the mass spectral libraries and the literature. MS ions (EI) were given by GC-MS on a DB-WAX column. They consisted of molecular ion M+ (relative abundance) and the base peak ion, or the peak ion with the second-highest relative abundance if the molecular ion was the base peak.

^b A: 500–1000; B: 100–500; C: 10–100; and D: 1–10.

amounts of identified compounds were very low (< 1%), and aldehydes (43.5%), terpenoids (25.3%), and alcohols (17.6%) were identified as the predominant chemical classes (Fig. 1). The five most abundant compounds, representing approximately 43% of the total amount of volatile compounds in the raw seeds, were hexanal (19.8%), 3-methyl-1-butanol (5.2%), (–)-camphor (5.8%), benzeneacetaldehyde (6.4%), and borneol (6.2%). Similarly, hexanal, benzeneacetaldehyde, and 3-methyl-1-butanol were identified as the major volatile

components in pine and grape seeds [15,16]. In comparison, hexanal and 3-methyl-1-butanol were not found in the leaf oils of *D. nankingense* [8–10]. The volatile compounds in the leaf harvested in Jiangsu province were borneol (26.9%), bornyl acetate (18.6%), germacrene D (6.06%), and α -sesquiphellandrene (6.06%) [10]. The most abundant compounds in the leaf oil from Anhui province were camphor (11.6%), carvophyllene oxide (6.7%), β -santalol (14.4%), and bisabolol oxide A (17.9%) [8].

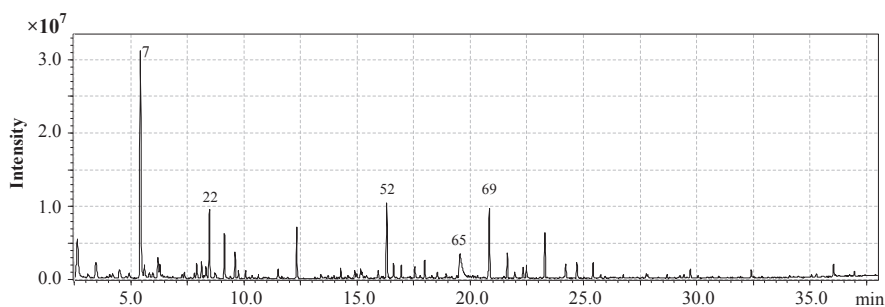


Fig. 1 – GC–MS chromatogram (DB-WAX) of the volatile extract from the seed of *Dendranthema nankingense*, highlighting the most abundant compounds. Peak number as in [Table 1](#). GC–MS = gas chromatography–mass spectrometry.

Interestingly, the seeds contained significant quantities of volatile terpenoids such as camphor and borneol, which are commonly found in the leaves and flowers of *D. nankingense* [8–10]. It appears that these two volatiles could be present throughout the entire life cycle of *D. nankingense*, providing the characteristic flavor and functional components of the plant. The relative amount of borneol in the leaf oil of *D. nankingense* harvested in Jiangsu province [10] is 4.5 times greater than that in our study. The sum of the relative amounts of borneol and camphor in the leaf oil of *D. nankingense* harvested in Anhui province [8] is very similar to that in this study. Camphor and borneol are collectively regarded as the key bioactive components of a number of traditional Chinese medicines, including *Amomum villosum* Lour seeds and the two Compositae herbs, *Chrysanthemum morifolium* Ramat and *Flos Chrysanthemi Indici* [17,18]. Camphor and borneol have also been identified as the main components in *Tanacetum vulgare*, which is another Compositae plant that grows wild in Lithuania [19]. Furthermore, camphor was identified as a major volatile component in *D. indicum*, which is a Chinese herbal medicine [20]. The relative amount of camphor in the *D. nankingense* seeds was similar to that found in coriander (*Coriandrum sativum* L.) seeds harvested in Atlantic Canada [21]. Camphor is also a key compound in several commercial and potentially commercial essential oils from the eucalyptus species and other myrtaceae plants [22]. Borneol has a pungent, camphor-like odor and provides a burning taste sensation, somewhat reminiscent of mint. It can be synthesized by reduction of camphor or from pinene [23]. Natural borneol, however, only consists of *d*-borneol, whereas synthetic borneol contains both *d*-borneol and isoborneol. Borneol is a common ingredient in many traditional Chinese herbal formulas, where it is more commonly referred to by its Chinese name, bingpian. It occurs not only in the essential oils of numerous medicinal plants, but also in the volatile fractions of edible oil seeds [24]. Borneol induces a variety of physiological responses, including stimulation of the digestive system by increasing the production of gastric juices; toning the heart and improving circulation; treating bronchitis, coughs, and colds; reducing swelling; and relieving stress; it can also be used as a tonic to promote relaxation and reduce fatigue [25]. Furthermore, a number of studies have shown that borneol could improve nasal and oral bioavailability by enhancing the permeability of the blood–brain barrier and improving the distribution of drugs in the brain [26]. It is also

widely used for the treatment of anxiety and pain, and as an anesthetic, and can be used in combination with *Gardenia* for the treatment of stroke in traditional Chinese medicines [27]. As a fragrance ingredient, borneol has been widely used in cosmetics, fine fragrances, and cleaners and detergents. If further studies elucidate new functions and bioactive mechanisms, demand for natural borneol and camphor may increase in the future. With this in mind, *D. nankingense* seeds or its essential oil may serve as an important source of natural camphor and borneol. Of the other terpenoids identified in the volatiles, 1,8-cineol, linalool, thujone, chrysanthenone, spathulenol, ocimene, camphene, and caryophyllene were identified as the major constituents in other Compositae plants [28–30]. Caryophyllene, caryophyllene oxide, and 4-terpineol were also found in the leaf of *D. nankingense* [6].

3.2. Components of volatile oil from the seeds of *B. officinalis*

SDE of *B. officinalis* seeds afforded a pale, tawny-colored oil (yield 2.2%, w/w). The color of the volatile oil was very similar to that reported by Mhamdi et al [11]; however, the SDE yield was 22 times greater than that obtained with the hydro-distillation method used by Mhamdi et al [11]. Furthermore, the observed composition of the volatile mixture from *B. officinalis* seeds differed considerably between the two studies, and we identified 65 compounds, which is four times as many volatile compounds than Mhamdi et al [11] identified. The volatile compounds are listed in [Table 2](#) ([Fig. 2](#)). Surprisingly, in comparison with the qualitative results reported by Mhamdi et al [11], only three compounds—hexanol, linalool, and phenol—coincided. Aldehydes (35.4%), heterocyclic compounds (25.2%), and ketones (22.8%) were identified as the major chemical classes, and 3-methyl butanal (10.8%), 2-pentanone (18.0%), hexanal (10.8%), and 2,3-dihydro-benzofuran (16.1%) were the most abundant compounds. These four main compounds represented approximately 55% of the total amount of volatile compounds in the raw seeds. By comparison, the most abundant volatiles in the borage seeds harvested in Tunisia were β -caryophyllene (26%), *p*-cymen-8-ol (19.7%), decanal (10.4%), and (*Z*)-3-hexenol (6.3%). Hexanal, a common plant volatile compound with a grassy odor, was not detected in the Tunisia seeds. These results suggest that the volatiles of *B. officinalis* seeds might vary significantly depending on their origin, harvest maturity, and storage time.

Table 2 – Volatile extract composition of *Borago officinalis* seeds.

No.	Retention time on DB-WAX (min)	Compounds	RIs		Identification and MS ions (EI) ^a	Total area (%)	Approximate concentration (µg/mL extract) ^b
			DB-WAX	HP-1			
1	2.64	3-Methyl butanal	925	<700	++++; M+ 86 (16.67), 44	10.8	B
2	3.50	2-Pentanone	987	<700	++++; M+ 86 (20.02), 43	18.0	A
3	5.00	2,3-Pentanedione	1069	<700	+++; M+ 100 (16.32), 43	0.7	C
4	5.41	Hexanal	1086	779	++++; M+ 100 (1.21), 44	10.8	B
5	5.97	3-Pentanol	1110	<700	++++; M+ 88 (0.65), 59	0.4	B
6	6.28	2-Pentanol	1124	<700	++++; M+ 88 (1.03), 45	1.2	C
7	6.62	2-Butyl furan	1139	880	+++; M+ 124 (13.46), 81	0.3	C
8	6.85	1-Butanol	1148	<700	++++; M+ 74 (1.86), 56	0.1	D
9	7.81	2-Heptanone	1185	872	++++; M+ 114 (6.60), 43	0.9	C
10	7.90	Heptanal	1188	881	++++; M+ 114 (2.01), 70	1.2	C
11	8.24	Dodecane	1200	1199	++++; M+ 170 (4.12), 57	0.2	D
12	8.71	(E)-2-Hexenal	1219	829	+++; M+ 98 (22.10), 41	0.7	C
13	9.12	2-Pentyl-furan	1236	981	+++; M+ 138 (18.35), 81	1.9	C
14	9.60	1-Pentanol	1254	759	++++; M+ 88 (1.16), 42	1.3	C
15	9.74	Styrene	1259	875	+++; M+ 104 (100.00), 103 (48.72)	0.3	C
16	9.93	Methyl-pyrazine	1267	796	+++; M+ 94 (100.00), 67 (47.31)	0.5	C
17	10.51	2-Octanone	1287	964	+++; M+ 128 (6.18), 43	0.7	C
18	10.63	Octanal	1290	980	++++; M+ 128 (1.20), 43	0.8	C
19	10.87	1-Hydroxy-2-propanone	1299	<700	+++; M+ 74 (11.62), 43	0.3	C
20	10.96	1-Octen-3-one	1302	959	++++; M+ 126 (0.96), 55	0.5	C
21	11.39	2,5-Dimethyl-pyrazine	1320	885	+++; M+ 108 (93.21), 42	0.3	C
22	11.50	(Z)-2-Heptenal	1325	932	+++; M+ 112 (6.85), 41	0.8	C
23	11.57	2,6-Dimethyl-pyrazine	1327	882	++++; M+ 108 (100.00), 42 (90.15)	0.3	C
24	11.67	2-Methyl-3-octanone	1331	1074	+++; M+ 142 (4.55), 43	0.2	D
25	12.32	1-Hexanol	1356	862	++++; M+ 102 (0.58), 56	0.9	C
26	13.07	2-Ethyl-6-methyl-pyrazine	1383	976	+++; M+ 122 (65.87), 121	0.2	D
27	13.25	2-Nonanone	1390	1072	+++; M+ 142 (8.94), 43	0.5	C
28	13.38	Nonanal	1394	1083	++++; M+ 142 (1.01), 57	1.6	C
29	13.70	(E)-3-Octen-2-one	1406	1014	+++; M+ 126 (14.38), 43	0.3	C
30	14.26	(E)-2-Octenal	1430	1034	+++; M+ 126 (3.29), 41	0.8	C
31	14.88	1-Octen-3-ol	1454	981	++++; M+ 128 (0.60), 57	1.6	C
32	14.96	Acetic acid	1457	<700	++++; M+ 60 (76.08), 43	2.7	B
33	15.21	Furfural	1467	802	++++; M+ 96 (100.00), 95 (99.23)	1.8	C
34	15.76	(Z)-6-Nonenal	1488	1007	+++; M+ 140 (0.85), 41	0.2	D
35	15.93	2-Decanone	1494	1173	+++; M+ 156 (5.11), 58	0.4	C
36	16.53	Pyrrrole	1518	748	++++; M+ 67 (100.00), 39 (64.74)	0.3	C
37	16.60	Benzaldehyde	1521	929	++++; M+ 106 (96.18), 77	0.8	C
38	16.93	2-Nonenal	1535	1144	+++; M+ 140 (3.96), 43	0.5	C
39	17.33	Linalool	1552	1088	++++; M+ 154 (1.24), 71	0.2	D
40	17.53	1-Octanol	1560	1069	++++; M+ 130 (0.34), 56	0.4	C
41	17.88	5-Methyl-2-furancarboxaldehyde	1574	939	+++; M+ 110 (100.00), 109 (79.03)	0.2	D

EI = electron impact; GC–MS = gas chromatography–mass spectrometry; RI = retention index.

^a The reliability of the proposed identification has been indicated according to the following: +++++, mass spectrum and retention indices both on polar and nonpolar columns agreed with those of the standards, mass spectral libraries and literature; +++, mass spectrum and retention indices both on polar and nonpolar columns agreed with mass spectral libraries and the literature; ++, mass spectrum and retention index on a polar column agreed with the mass spectral libraries and the literature. MS ions (EI) were given by GC-MS on a DB-WAX column. They consisted of molecular ion M+ (relative abundance) and the base peak ion, or the peak ion with the second-highest relative abundance if the molecular ion was the base peak.

^b A: 500–1000; B: 100–500; C: 10–100; and D: 1–10.

The percentage of hexanal can increase with increasing harvest and storage time of the seeds, and is significantly affected by the amount of lipoxygenase present in the seeds. This phenomenon has been observed in both pine and rice seeds [15,31]. 2,3-Dihydro-benzofuran, also known as coumaran, has been observed as a minor component in several studies involving green tea, Rooibos tea, cotton honey, some fruits, and many plant leaves. It was also found to be a volatile marker compound in 10 honeys originating from unifloral *Salvia officinalis* L., which contained 4.1–6.8% of coumaran [32].

To the best of our knowledge, our study is the first to identify coumaran as a major (> 10%) volatile component in plants. Thus, *B. officinalis* seeds may serve as a potential source for the production of coumaran.

Thirteen heterocyclic compounds, including derivatives of pyrazine, pyrrole, and furan, were identified in the volatiles of *B. officinalis* seeds. Pyrazine, pyrrole, and furan derivatives are widely distributed in processed food items, such as roasted sorghum, wines, and roasted almond [33–35]. Four of the alky pyrazines detected—methyl-pyrazine, 2,5-dimethyl-

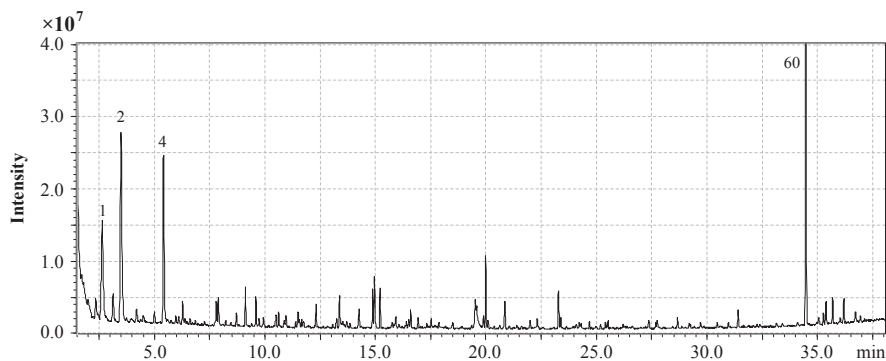


Fig. 2 – GC–MS chromatogram (DB–WAX) of the volatile extract from the seed of *Borago officinalis*, highlighting the most abundant compounds. Peak number as in Table 2. GC–MS = gas chromatography–mass spectrometry.

pyrazine, 2,6-dimethyl-pyrazine, and 2-ethyl-6-methyl-pyrazine—have also been found in roasted soybean oils, and may contribute to the characteristic toasted, nut- and peanut-like burnt aroma [36,37]. Of these four compounds, 2-methyl-pyrazine, 2,5-dimethyl-pyrazine, and 2-ethyl-6-methyl-pyrazine are synthetically accessible via the reaction between L-ascorbic acid and L-threonine or L-serine.

3.3. Similarities between volatile oils obtained from *D. nankingense* and *B. officinalis* seeds

The volatile oil yields from both types of seeds were very similar, and 33 of the volatile compounds were common to both seeds, with the majority of these compounds being aliphatic aldehydes and alcohols. Of these compounds, hexanal, 1-hexanol, nonanal, and 1-octen-3-ol have also been reported in poppy seeds and groats [38,39]. Hexanal is particularly abundant in both seeds. High levels of hexanal have been found in a number of plant seeds, including pine seeds, coriander seeds, rice seeds, and *Trichosanthes kirilowii* seeds [15,23,31,40]. All four of these volatiles play important roles as food flavor ingredients in baked goods, frozen dairy products, candies, and nonalcoholic beverages.

Both seeds contained seven C8 compounds with a mushroom flavor, and 1-octen-3-ol, (E)-3-octen-2-one, and octanal were common to them both. 1-Octen-3-ol, also known as “mushroom alcohol,” is a volatile alcohol produced by many edible fungi species [41]. Furthermore, it is a component of the volatile oil and plays an important role in the odor profiles of several plants, especially in members of the Lamiaceae family.

4. Conclusion

A detailed investigation of the volatile extracts of two seeds (*D. nankingense* and *B. officinalis*) prepared by SDE revealed a total of 127 compounds. The major volatiles found in the *D. nankingense* seeds were hexanal, benzeneacetaldehyde, borneol, (–)-camphor, and 3-methyl-1-butanol. These compounds, especially borneol and (–)-camphor, can be used as quality markers for *D. nankingense*. The major volatiles found in the *B. officinalis* seeds were 2-pentanone, coumaran,

3-methyl butanal, and hexanal. Coumaran was the most abundant volatile component in *B. officinalis* seeds.

Conflicts of interest

All authors have no conflicts of interest.

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