



# HHS Public Access

Author manuscript

*EJC Paediatr Oncol.* Author manuscript; available in PMC 2024 July 02.

Published in final edited form as:

*EJC Paediatr Oncol.* 2024 June ; 3: . doi:10.1016/j.ejcped.2024.100160.

## CAR T cells redirected to B7-H3 for pediatric solid tumors: Current status and future perspectives

Rebecca Epperly\*, Stephen Gottschalk,  
Christopher DeRenzo

Department of Bone Marrow Transplantation and Cellular Therapy, St. Jude Children's Research Hospital, Memphis, TN, USA

### Abstract

Despite intensive therapies, pediatric patients with relapsed or refractory solid tumors have poor outcomes and need novel treatments. Immune therapies offer an alternative to conventional treatment options but require the identification of differentially expressed antigens to direct antitumor activity to sites of disease. B7-H3 (CD276) is an immune regulatory protein that is expressed in a range of malignancies and has limited expression in normal tissues. B7-H3 is highly expressed in pediatric solid tumors including osteosarcoma, rhabdomyosarcoma, Ewing sarcoma, Wilms tumor, neuroblastoma, and many rare tumors. In this article we review B7-H3-targeted chimeric antigen receptor (B7-H3-CAR) T cell therapies for pediatric solid tumors, reporting preclinical development strategies and outlining the landscape of active pediatric clinical trials. We identify challenges to the success of CAR T cell therapy for solid tumors including localizing to and penetrating solid tumor sites, evading the hostile tumor microenvironment, supporting T cell expansion and persistence, and avoiding intrinsic tumor resistance. We highlight strategies to overcome these challenges and enhance the effect of B7-H3-CAR T cells, including advanced CAR T cell design and incorporation of combination therapies.

### Keywords

B7-H3; CAR T cell therapy; Chimeric antigen receptor; Solid tumor; Immunotherapy; Pediatric cancer

---

This is an open access article under the CC BY-NC license (<http://creativecommons.org/licenses/by-nc/4.0/>).

\*Correspondence to: 262 Danny Thomas Place, MS 1130, Memphis, TN 38105, USA. [Rebecca.Epperly@stjude.org](mailto:Rebecca.Epperly@stjude.org) (R. Epperly).

CRedit authorship contribution statement

**Rebecca Epperly:** Writing – original draft, Writing – review & editing. **Stephen Gottschalk:** Writing – review & editing.

**Christopher DeRenzo:** Writing – review & editing.

Declaration of Competing Interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests. R. E., S.G. and C.D. have patent applications in the field of CAR T cell therapy, including B7-H3-specific CAR T cells. S.G. is a member of the Scientific Advisory Board of Be Biopharma and CARGO, and the Data and Safety Monitoring Board (DSMB) of Immatic and has received honoraria from TESSA Therapeutics within the last year.

## 1. Introduction

Pediatric patients with relapsed/refractory solid tumors have dismal outcomes despite attempts at intensive multimodal therapies including chemotherapy, radiation therapy, and surgical resection [1–5]. These patients are urgently in need of novel therapies to improve outcomes. Chimeric antigen receptor (CAR) T cell therapy represents a promising potential strategy for treating pediatric patients with solid tumors. In contrast to CD19-CAR T cell therapy which has produced dramatic responses for pediatric B-ALL [6–10], initial attempts to translate CAR T cell therapy for pediatric solid tumors have demonstrated limited clinical activity [11–16]. However, a recent study with GD2-CAR T cells has demonstrated significant antitumor activity in pediatric patients with neuroblastoma and low disease burden [17]. Identifying target antigens that are differentially expressed in a broad range of pediatric tumors and not in healthy tissues remains a significant challenge. Likewise, for individual pediatric solid tumor types, there may be heterogeneity in antigen expression among individual patients, between metastatic and primary sites, and within an individual tumor [18].

B7-H3 (CD276) is an immunomodulatory protein that has emerged as an attractive target for immunotherapy, due to its expression on a range of malignancies including pediatric solid tumors [4,19–21]. Beyond antigen expression, CAR T cell therapy approaches for pediatric solid tumors must also overcome the challenges of localizing to and penetrating tumor sites, evading a hostile immune microenvironment, optimizing T cell dynamics, and avoiding antigen escape [11,12]. In this review we detail the expression of B7-H3 in pediatric solid tumors and brain tumors, discuss efforts in preclinical B7-H3-targeted CAR (B7-H3-CAR) T cell development, describe the current landscape of pediatric B7-H3-CAR T cell clinical trials, and explore strategies to improve B7-H3-CAR T cell approaches.

## 2. B7-H3 as an antigen for CAR T cell therapy

### 2.1. Physiologic role of B7-H3

B7-H3 is a surface glycoprotein encoded on chromosome 15 which can exist in two isoforms: containing either two or four immunoglobulin-like (Ig) regions, with the four Ig isoform predominating in humans [22–24]. The receptors of B7-H3 have not been definitively described, with conflicting reports existing on the role of potential candidates including TLT-2, IL-20R $\alpha$ , and PLA2R1 [24–30]. Physiologically, B7-H3 plays a role in immunomodulation [30,31]. While initial reports proposed a costimulatory effect [32], subsequent studies have identified associated B7-H3 expression with inhibition of T cell activation and suppression of T cell-mediated antitumor responses [24,27,33]. Proposed mechanisms of immune inhibition include decreased signaling through the PI3K/AKT/mTOR pathway [34], restriction of antigen-presenting cells [35], M2 macrophage polarization [36,37], and inhibition of natural killer cell-mediated lysis [38]. Beyond immune regulation, B7-H3 is proposed to directly support cancer cell invasion, proliferation, angiogenesis, and metabolism [30]. Through both immunologic and non-immunologic roles, B7-H3 has the potential to support tumor progression and immune evasion, while having antitumor effects under some circumstances [31].

Minimal healthy tissue B7-H3 expression has been detected at the protein level by immunohistochemistry (IHC) and at the RNA level in sequencing-based assays [21,39,40]. Low-level expression has been reported in normal stomach [19,40,41], colon [41], salivary gland [40], skin [42], pancreas [19] and liver [19], with variable expression in normal adrenal tissue [19,21,40]. Importantly, initial clinical studies targeting B7-H3 with monoclonal antibodies did not identify concerns for significant on-target off-tumor toxicities [43–45]. Because of this favorable expression profile, a number of B7-H3-targeting therapeutic strategies are being explored, including bispecific antibodies, antibody-drug conjugates, radioimmunotherapy, and cellular therapy [42,45–51]. As B7-H3 continues to be explored as an immunotherapy target, it will be necessary to continue monitoring for off-tumor toxicities and further defining its physiologic role as we seek to understand potential mechanisms of treatment resistance.

## 2.2. B7-H3 expression in pediatric solid tumors

In contrast with low-level normal tissue expression, B7-H3 is highly expressed in pediatric solid tumors [4,11,19,21,39,40]. Pediatric sarcomas have particularly high B7-H3 expression, including osteosarcoma, Ewing sarcoma, rhabdomyosarcoma, and non-rhabdomyosarcoma soft tissue sarcomas [19,21,39,40]. This includes >95% expression reported in samples from patients with osteosarcoma, coinciding with expression in a range of preclinical osteosarcoma models including in vitro assays and in vivo murine and spontaneously-occurring canine models [52–57]. In rhabdomyosarcoma, B7-H3 expression was identified in both alveolar and embryonal histologic subtypes, encompassing *FOXO1* fusion-negative and fusion-positive disease, with a relationship identified between *PAX3--FOXO1* and *B7-H3* expression in functional studies [21,39,58,59].

Likewise in neuroblastoma, B7-H3 was identified in samples from patients with both localized and metastatic disease, with higher levels of B7-H3 identified through IHC or mRNA expression associated with poor prognosis [60,61]. This clinicopathologic observation is supported by preclinical studies demonstrating that knockdown of B7-H3 in neuroblastoma models is associated with tumor cell proliferation [62], and overexpression of B7-H3 can confer chemotherapy resistance [60,63]. Importantly, B7-H3 expression can be retained in patients with GD2-low or negative disease after receipt of prior GD2-targeted immunotherapy [64]. In addition to the extended analyses in neuroblastoma and sarcomas, additional pediatric solid tumors with reported B7-H3 expression include Wilms tumor, malignant peripheral nerve sheath tumor, hepatoblastoma, melanoma, and desmoplastic small round cell tumor [19, 21,39].

## 2.3. B7-H3 as a pan-cancer antigen

Beyond extracranial pediatric solid tumors, B7-H3 is also highly expressed in a range of other malignancies. The majority of pediatric brain tumor samples evaluated express B7-H3 across all subtypes including medulloblastoma, high-grade glioma, diffuse midline glioma (formerly diffuse intrinsic pontine glioma [DIPG]), ependymoma, and atypical teratoid rhabdoid tumor (ATRT) [19,65]. B7-H3 expression exceeds or is comparable to that of other immunotherapy targets of interest in pediatric brain tumors [65]. Increased intensity of B7-H3 IHC has been associated with high-grade features, and increased mRNA expression

associated with shorter overall survival [66]. B7-H3 expression is also noted in adult brain tumors including glioblastoma and anaplastic meningioma [67–69].

Like pediatric solid tumors, adult solid tumors also have marked B7-H3 expression [70]. B7-H3 has been extensively evaluated in prostate, breast, and ovarian cancers, where increased expression is associated with increased risk of recurrence, decreased survival, and treatment resistance [30,71–76]. Notably, in prostate cancer increased B7-H3 expression is associated with biochemical recurrence after radiation therapy and increased androgen receptor activation [72,73]. In non-small cell lung cancer, B7-H3 is associated with advanced stage and metastatic disease [77,78]. Across a range of gastrointestinal cancers including hepatocellular, gastric, pancreatic, esophageal, and colorectal, B7-H3 is expressed and often associated with poor prognosis [40, 70,79–82]. In pancreatic cancer models, increased intensity of B7-H3 expression is seen in metastatic disease and associated with increased pathological stage [80]. In addition to solid tumors, B7-H3 is being explored as a target for hematologic malignancies including acute myeloid leukemia [83,84], anaplastic large cell lymphoma [85], and NK/T cell lymphoma [86]. The spectrum of malignancies which express B7-H3, overall differential expression from healthy tissues, and association with high-risk disease features, support the role of B7-H3 as a pan-cancer antigen and attractive immunotherapeutic target. This potentially broad impact further supports developing B7-H3-targeting therapies for individually rare pediatric tumor types.

### 3. B7-H3-CAR T cell preclinical development

The efficacy of a CAR T cell construct is dependent on the interaction between each component of the CAR design, including antigen recognition domains, structural elements, and costimulatory molecules (Fig. 1). Variations of B7-H3-CAR T cells are undergoing translational development, with many groups exploring additional enhancements to improve CAR T cell function [48].

#### 3.1. CAR design

Traditional CAR T cell antigen recognition domains are based on the single chain variable fragment (scFv) of a monoclonal antibody. For B7-H3, initial products proceeding in clinical development include those based on MGA271 [21,39,41,87] and mAb376.96 [40,88] scFvs. To evaluate the properties of B7-H3-CAR T cells with variable antigen recognition properties, single-chain antibody libraries have been produced to generate additional scFvs for preclinical evaluation [89]. As an alternative to traditional scFvs, nanobody-based libraries have also been explored to generate high-affinity recognition domains [90]. Beyond the antigen recognition domain, key components of the hinge and transmembrane can affect structure and activity of the CAR T cell construct [21,87]. While first generation CAR T cells consisted of antigen recognition and intracellular signaling domains, second generation CAR T cells include the addition of costimulatory molecules. Costimulatory molecules of the immunoglobulin superfamily including CD28 are associated with potent initial antitumor activity, while costimulatory molecules of the tumor necrosis factor receptor superfamily including 4-1BB and OX40 are associated with improved CAR T cell persistence [91]. Both classes of costimulatory molecules are being evaluated in B7-H3-CAR T cell constructs, in

addition to third generation CARs which incorporate multiple costimulatory domains [48]. When incorporating multiple costimulatory domains, the route of costimulation can affect function [21,91].

### 3.2. Functional safety studies

In addition to screening healthy tissue libraries for B7-H3 expression [21], functional preclinical studies support the safety of pursuing B7-H3-CAR T cell therapy. While several preclinical murine studies evaluating human B7-H3-CAR T cell constructs have not identified any unexpected toxicities [21,39,40], models are potentially limited by a lack of cross-reactivity between human and murine B7-H3. While there is 88% homology between human and murine B7-H3 at the amino acid level, many human B7-H3-directed antibodies do not specifically cross react with murine B7-H3 [22,92,93]. Thus, cross-reactivity must be considered both in functional assessment of on-target off-tumor effects and in screening methods. To overcome this barrier, studies evaluating murine CAR T cells in immune competent models have also been performed, and have not unveiled any additional toxicity concerns [65,92]. Beyond murine studies, the MGA271 monoclonal antibody, from which the scFv in several B7-H3-CAR T cell constructs is derived, has been evaluated in a non-human primate model [41] and B7-H3-CAR T cells have been evaluated in naturally occurring spontaneous canine sarcoma. [53,57]. B7-H3-targeted therapies have demonstrated safety in these non-human primate and canine models.

### 3.3. Enhancing function

Due to challenges associated with initial attempts to translate CAR T cell therapy for solid tumors, many groups are developing next-generation B7-H3-CAR T cell constructs with additional enhancements (Fig. 2). A variety of gene editing techniques have been explored to enhance CAR T cell function, safety, and persistence across CAR T cell targets [94]. Hairpin RNA strategies have been explored to knockdown a panel of inhibitory molecules including PD-1, TIM-3, TIGIT, TGF $\beta$ R, IL-10R, and IL-6R to improve antitumor activity of B7-H3-CAR T cells in a cholangiocarcinoma model [95]. B7-H3-CAR T cells engineered to include a PD-1 decoy receptor demonstrated improved persistence in a panel of solid tumor models [96]. An alternative strategy aimed at improving T cell persistence is the inclusion of STAT3- and STAT5-related activation motifs to generate less differentiated B7-H3-CAR T cells, which showed superior activity in breast and ovarian cancer models [97]. In addition, expressing chimeric cytokine receptors to activate STAT-signaling pathways in B7-H3-CAR T cells has improved their effector function in xenograft models [98]. Beyond T cell persistence, CAR T cell design can be enhanced to improve CAR T cell trafficking and direct activity to tumor sites. In brain metastases models, overexpression of CCR2 in B7-H3-CAR T cells improves migration across the blood brain barrier through the CCL2/CCR2 chemokine axis [99]. In an attempt to localize antitumor activity of B7-H3-CAR T cells to tumor sites, a microenvironment regulated system (MRS) was also developed to promote B7-H3-CAR T cell proliferation in the tumor microenvironment in an esophageal carcinoma model [100]. To further enhance targeting specificity for improved safety and expanded applications, B7-H3-targeted CAR T cells have been generated which use alternative intracellular domains such as ZAP-70 in a logic-gated platform, rather than traditional CD3 $\zeta$  sequences [101]. The incorporation of a PDZ binding motif has also been

shown to improve effector cell functionality by enhancing synapse formation [102]. B7-H3 has also been targeted in osteosarcoma models using a switchable CAR T cell system, which offers potential for broad translational efforts [55].

## 4. Pediatric B7-H3-CAR T cell clinical trials

Based on the encouraging preclinical data and safety profile, B7-H3 CAR T cells are now being evaluated in early phase clinical studies. Table 1 outlines “active” and “active, not recruiting” clinical B7-H3 CAR T cell studies for pediatric patients.

### 4.1. Pediatric solid tumor clinical studies

Systemic CAR T cell infusion is being studied for pediatric solid tumors (Table 1). In general, these studies are designed as basket trials, which enroll patients across a range of tumor types. Enrollment criteria vary regarding requirements for confirming B7-H3 antigen positivity and whether there is a focus on specific tumor types. The CAR T cell constructs being translated vary in the method and route of costimulation. They include plans for both single- and multiple-antigen targeting strategies, including the co-expression of CD19-CARs in B7-H3-CAR T cells with the goal of improving their expansion and persistence. In general, these studies evaluate a systemic infusion of B7-H3-CAR T cells after administration of lymphodepleting chemotherapy (i. e. fludarabine and cyclophosphamide) (Fig. 1). These first-in-human studies are ongoing, and current data available in abstract form notes a tolerable safety profile with limited antitumor activity [103].

### 4.2. Pediatric primary central nervous system (CNS) clinical studies

For primary central nervous system (CNS) tumors, preclinical data favors locoregional CAR T cell administration [65,104,105]. This approach is being translated in active clinical studies, where B7-H3-CAR T cells are injected into the ventricular space or tumor resection cavity [106,107]. In contrast to systemic infusion with preceding lymphodepleting chemotherapy, these local approaches are administered as serial infusions without lymphodepleting chemotherapy (Table 1). Recently, three patients with DIPG were reported who had received intracranial B7-H3-CAR T cells, which were tolerated without dose limiting toxicity at the first dose level and associated with evidence of local immune activation [87]. One participant demonstrated clinical and radiographic response.

### 4.3. Lessons from adult B7-H3-CAR T cell clinical experience

Parallel studies in adult solid and CNS tumors have the potential to generate important safety and efficacy data regarding B7-H3-CAR T cell therapy [20]. Early evidence includes a report of partial response after intratumoral B7-H3-CAR T cell injection for a single patient with basal cell carcinoma [108]. Transient clinical activity has also been reported in an adult patient with glioblastoma multiforme after local B7-H3-CAR T cell administration [68]. Beyond brain tumors, adult studies are evaluating local administration for other solid tumors, including intraperitoneal delivery for ovarian tumors [40]. While data in adults can provide proof of principle for clinical activity, pediatric patients have unique physiologic considerations and tumor biology. Due to the urgent need to improve treatments for

pediatric patients with relapsed/refractory tumors, it is imperative that novel products be simultaneously studied directly in the pediatric population.

## 5. Strategies to improve B7-H3-CAR T cell therapy

Beyond antigen selection, there are several hurdles to establishing effective CAR T cell therapy for pediatric patients with solid tumors. These include CAR T cell homing to and penetrating tumor sites, overcoming the hostile tumor microenvironment, and having adequate T cell expansion and persistence [5,12]. As described above, investigators are pursuing enhanced CAR T cell design [94] and exploring locoregional delivery methods [68,87,104] to begin addressing these hurdles. Moving forward, additional strategies seek to evaluate B7-H3 CAR T cells in combination therapies, as part of multi-antigen targeting approaches, and through alternative immune effector cells (Fig. 2).

### 5.1. Combination therapies for immune and antigen modulation

Given the challenges of effectively targeting solid tumors with CAR T cells, combined immune therapy approaches have the potential to enhance T cell activity and improve penetration to a hostile tumor microenvironment. One approach is the combination of B7-H3-CAR T cells with oncolytic adenovirus (ADV) to deliver immune regulators. In a glioblastoma model, CXCL11-armed oncolytic ADV injected to the tumor site improves infiltration of B7-H3-CAR T cells and decreases proportions of inhibitory immune cells [109]. Also in glioblastoma models, interleukin-7-loaded oncolytic ADV has been co-administered with B7-H3-CAR T cells, demonstrating improved T cell proliferation [110]. B7-H3-CAR T cell therapy can also be enhanced by modulating surface B7-H3 expression in tumor targets. The combination of a pan-histone deacetylase inhibitor (SAHA) has been shown to upregulate B7-H3 expression on solid tumors, in addition to downregulating CTLA-4 and TET2, leading to improved B7-H3 CAR T cells in preclinical models [111]. In a drug library screen, ingenol-3-angelate was also identified to increase B7-H3 expression through PKC $\alpha$  activation, which enhanced B7-H3-CAR T cell function [112]. Screening efforts have also identified small molecules which enhance B7-H3-CAR T cell activity through direct antitumor mechanisms. For example, the hedgehog signaling inhibitor JK184 directly induced tumor apoptosis and had synergistic antitumor effects in combination with B7-H3-CAR T cells [113]. A recent study has highlighted that local treatment of solid tumors in xenograft models simulates robust immunogenic cell death, enhancing B7-H3-CAR T cell expansion, persistence, and antitumor activity [114]. Local radiation by itself has also been shown to increase B7-H3 expression and promote a more favorable immune microenvironment [115,116].

### 5.2. Multi-antigen targeting

Targeting multiple antigens with CAR T cells has the potential to overcome key resistance mechanisms. Strategies which are dependent on the presence of both antigens for CAR T cell activation (AND gates) can improve specificity, overcome low antigen density, and help discern between tumor and normal tissues [117]. Strategies that allow for activation in the presence of either one or both antigens (OR gates) aim to address antigen heterogeneity and prevent resistance through development of, or selection for, antigen-loss variants. For

neuroblastoma, B7-H3 has been combined with other common antigens. GD2/B7-H3 CAR T cells developed in a SynNotch system sought to improve specificity [118], and ALK/B7-H3-CAR T cells overcame low antigen density [119]. An OR gated strategy targeting GPC2 or B7H3 retained activity despite heterogeneous antigen expression [120]. Beyond neuroblastoma, CAR T cells targeting both CD70 and B7-H3 had activity against a range of solid tumors [121] and GPC3-CARs also secreting B7-H3-specific T cell engagers had activity in hepatocellular carcinoma [122]. In an osteosarcoma model, dual B7-H3- and CXCR2-specific CAR T cells had enhanced antitumor activity [57]. The inclusion of B7-H3 or EpCAM targeting in a ROR1-CAR T cell construct focused antitumor activity to tumor sites, avoiding bone marrow toxicity observed with single-targeting ROR1 CAR T cells [123]. While multi-antigen targeting holds promise to improve CAR T cell activity and limit toxicity, there is significant complexity to the design of these constructs. Comprehensive preclinical and correlative studies will be necessary to establish the best methods of dual targeting, which may vary by antigen and tumor type.

### 5.3. Alternative immune effector cell sources

Because of the challenges of T cell expansion and persistence in the tumor environment, efforts are underway to identify ideal subsets of T cells for adoptive cellular therapy and explore alternative immune effector cell sources. Both  $\gamma\delta$  T cells and natural killer (NK) cells are subsets that exist at the intersection between the innate and adaptive arms of the immune system [124]. Benefits of these cell sources include natural tropism for tumor tissues, potential for intrinsic antitumor activity, and lack of alloreactivity supporting potential allogeneic approaches [124]. Engineering these immune cells to express B7-H3-CARs can add an additional layer of specificity. The antitumor effect of V $\gamma$ 9 V $\delta$ 2 T cells was enhanced with the addition of a B7-H3-CAR in glioma models [125]. Similarly, B7-H3-CAR NK cells generated from the NK-92 line have demonstrated activity in melanoma and non-small cell lung cancer models [126,127]. Activity of B7-H3-CAR NK cells evaluated in a glioblastoma model were more resilient in a hostile immune environment through the addition of a TGF- $\beta$  dominant negative receptor [128]. These examples highlight the potential benefits of exploring alternative immune effector cell sources for engineered cell therapy.

## 6. Conclusions

Targeting B7-H3 with CAR T cells presents a promising strategy for treating pediatric patients with solid tumors. B7-H3 represents an ideal target antigen due to diffuse tumor expression including a range of pediatric solid tumors and limited healthy tissue expression. Several B7-H3-CAR T cell products have demonstrated preclinical activity. Early phase clinical studies in pediatric patients are underway, evaluating systemic administration for extracranial solid tumors and locoregional administration for primary CNS tumors. Despite encouraging preclinical data, CAR T cell therapy for solid tumors, including for B7-H3, still faces significant barriers including localizing T cells to tumor sites, overcoming the immune suppressive microenvironment, avoiding intrinsic tumor resistance mechanisms, and promoting functional T cell persistence. Thus, producing safe and effective B7-H3-CAR T cell therapy to target pediatric solid tumors will likely require enhanced CAR T cell



design and thoughtful combinatorial approaches. Insights gained from correlative analyses on early phase clinical studies will be key in prioritizing next steps to advance this treatment approach.

## Acknowledgements

Figs. 1 and 2 were designed with [www.BioRender.com](http://www.BioRender.com), for which we have a license.

## Funding

R.E. is supported by a New Investigator Award from the American Society for Transplantation and Cellular Therapy. C.D. and S.G. are supported by the United States Department of Health and Human Services, National Institutes of Health, National Cancer Institute (R01CA262589), the Osteosarcoma Institute, and the Assisi Foundation of Memphis. R.E., S.G. and C.D. are supported by the American Lebanese Syrian Associated Charities. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

## References

- [1]. National cancer institute, NCCR\*Explorer: an interactive website for NCCR cancer statistics., Vol, National Cancer Institute, Bethesda, MD, 2023.
- [2]. Ward E, DeSantis C, Robbins A, Kohler B, Jemal A, Childhood and adolescent cancer statistics, 2014, *CA Cancer J. Clin* 64 (2014) 83–103. [PubMed: 24488779]
- [3]. Spraker-Perlman HL, et al. , Factors influencing survival after recurrence in osteosarcoma: a report from the Children’s Oncology Group, *Pediatr. Blood Cancer* 66 (2019) e27444. [PubMed: 30255612]
- [4]. Bottino C, Vitale C, Dondero A, Castriconi R, B7-H3 in pediatric tumors: far beyond neuroblastoma, *Cancers (Basel)* 15 (2023).
- [5]. Evdokimova V, Gassmann H, Radvanyi L, Burdach SEG, Current state of immunotherapy and mechanisms of immune evasion in ewing sarcoma and osteosarcoma, *Cancers (Basel)* 15 (2022).
- [6]. Maude S, et al. , Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia, *N. Engl. J. Med* 378 (2018) 439–448. [PubMed: 29385370]
- [7]. Gardner R, et al. , Intent-to-treat leukemia remission by CD19 CAR T cells of defined formulation and dose in children and young adults, *Blood* 129 (2017) 3322–3331. [PubMed: 28408462]
- [8]. Talleur A, et al. , Preferential expansion of CD8+ CD19-CAR T cells postinfusion and the role of disease burden on outcome in pediatric B-ALL, *Blood Adv.* (2022).
- [9]. Talleur AC, Myers R, Annesley C, Shalabi H, Chimeric antigen receptor T-cell therapy: current status and clinical outcomes in pediatric hematologic malignancies, *Hematol. Oncol. Clin. North Am* 36 (2022) 701–727. [PubMed: 35780062]
- [10]. Curran KJ, et al. , Toxicity and response after CD19-specific CAR T-cell therapy in pediatric/young adult relapsed/refractory B-ALL, *Blood* 134 (2019) 2361–2368. [PubMed: 31650176]
- [11]. Kulczycka M, Derlatka K, Tasior J, Lejman M, Zawitkowska J, CAR T-cell therapy in children with solid tumors, *J. Clin. Med* 12 (2023).
- [12]. Wagner J, Wickman E, DeRenzo C, Gottschalk S, CAR T Cell therapy for solid tumors: bright future or dark reality? *Mol. Ther* 28 (2020) 2320–2339. [PubMed: 32979309]
- [13]. Straathof K, et al. , Antitumor activity without on-target off-tumor toxicity of GD2-chimeric antigen receptor T cells in patients with neuroblastoma, *Sci. Transl. Med* 12 (2020).
- [14]. Hegde M, et al. , Tumor response and endogenous immune reactivity after administration of HER2 CAR T cells in a child with metastatic rhabdomyosarcoma, *Nat. Commun* 11 (2020) 3549. [PubMed: 32669548]
- [15]. Ahmed N, et al. , Human epidermal growth factor receptor 2 (HER2) -specific chimeric antigen receptor-modified T cells for the immunotherapy of HER2-positive sarcoma, *J. Clin. Oncol* 33 (2015) 1688–1696. [PubMed: 25800760]

- [16]. Park JR, et al. , Adoptive transfer of chimeric antigen receptor re-directed cytolytic T lymphocyte clones in patients with neuroblastoma, *Mol. Ther* 15 (2007) 825–833. [PubMed: 17299405]
- [17]. Del Bufalo F, et al. , GD2-CART01 for relapsed or refractory high-risk neuroblastoma, *N. Engl. J. Med* 388 (2023) 1284–1295. [PubMed: 37018492]
- [18]. Bertacca I, Pegoraro F, Tondo A, Favre C, Targeted treatment of solid tumors in pediatric precision oncology, *Front. Oncol* 13 (2023) 1176790. [PubMed: 37213274]
- [19]. Modak S, Kramer K, Gultekin SH, Guo HF, Cheung NK, Monoclonal antibody 8H9 targets a novel cell surface antigen expressed by a wide spectrum of human solid tumors, *Cancer Res.* 61 (2001) 4048–4054. [PubMed: 11358824]
- [20]. Li G, Wang H, Wu H, Chen J, B7-H3-targeted CAR-T cell therapy for solid tumors, *Int. Rev. Immunol* 41 (2022) 625–637. [PubMed: 35855615]
- [21]. Nguyen P, et al. , Route of 41BB/41BBL costimulation determines effector function of B7-H3-CAR.CD28zeta T cells. *Mol. Ther. Oncolyt* 18 (2020) 202–214.
- [22]. Sun M, et al. , Characterization of mouse and human B7-H3 genes, *J. Immunol* 168 (2002) 6294–6297. [PubMed: 12055244]
- [23]. Steinberger P, et al. , Molecular characterization of human 4Ig-B7-H3, a member of the B7 family with four Ig-like domains, *J. Immunol* 172 (2004) 2352–2359. [PubMed: 14764704]
- [24]. Vigdorovich V, et al. , Structure and T cell inhibition properties of B7 family member, B7-H3, *Structure* 21 (2013) 707–717. [PubMed: 23583036]
- [25]. Hashiguchi M, et al. , Triggering receptor expressed on myeloid cell-like transcript 2 (TLT-2) is a counter-receptor for B7-H3 and enhances T cell responses, *Proc. Natl. Acad. Sci. U. S. A* 105 (2008) 10495–10500. [PubMed: 18650384]
- [26]. Kobori H, et al. , Enhancement of effector CD8+ T-cell function by tumour-associated B7-H3 and modulation of its counter-receptor triggering receptor expressed on myeloid cell-like transcript 2 at tumour sites, *Immunology* 130 (2010) 363–373. [PubMed: 20141543]
- [27]. Leitner J, et al. , B7-H3 is a potent inhibitor of human T-cell activation: No evidence for B7-H3 and TREML2 interaction, *Eur. J. Immunol* 39 (2009) 1754–1764. [PubMed: 19544488]
- [28]. Cao S, et al. , A membrane protein display platform for receptor interactome discovery, *Proc. Natl. Acad. Sci. U. S. A* 118 (2021).
- [29]. Husain B, et al. , A platform for extracellular interactome discovery identifies novel functional binding partners for the immune receptors B7-H3/CD276 and PVR/CD155, *Mol. Cell Proteom* 18 (2019) 2310–2323.
- [30]. Koumprentziotis IA, et al. , New emerging targets in cancer immunotherapy: the role of B7-H3, *Vaccin. (Basel)* 12 (2024).
- [31]. Wang L, Kang FB, Shan BE, B7-H3-mediated tumor immunology: friend or foe? *Int. J. Cancer* 134 (2014) 2764–2771. [PubMed: 24013874]
- [32]. Chapoval AI, et al. , B7-H3: a costimulatory molecule for T cell activation and IFN-gamma production, *Nat. Immunol* 2 (2001) 269–274. [PubMed: 11224528]
- [33]. Chen C, et al. , Induced expression of B7-H3 on the lung cancer cells and macrophages suppresses T-cell mediating anti-tumor immune response, *Exp. Cell Res* 319 (2013) 96–102. [PubMed: 22999863]
- [34]. Shao L, et al. , B7-H3 on breast cancer cell MCF7 inhibits IFN-gamma release from tumour-infiltrating T cells, *Pathol. Res. Pract* 224 (2021) 153461. [PubMed: 34265738]
- [35]. Schneider T, et al. , Non-small cell lung cancer induces an immunosuppressive phenotype of dendritic cells in tumor microenvironment by upregulating B7-H3, *J. Thorac. Oncol* 6 (2011) 1162–1168. [PubMed: 21597388]
- [36]. Kang FB, Wang L, Li D, Zhang YG, Sun DX, Hepatocellular carcinomas promote tumor-associated macrophage M2-polarization via increased B7-H3 expression, *Oncol. Rep* 33 (2015) 274–282. [PubMed: 25370943]
- [37]. Gao Y, et al. , LncRNA NEAT1 sponges miR-214 to regulate M2 macrophage polarization by regulation of B7-H3 in multiple myeloma, *Mol. Immunol* 117 (2020) 20–28. [PubMed: 31731055]

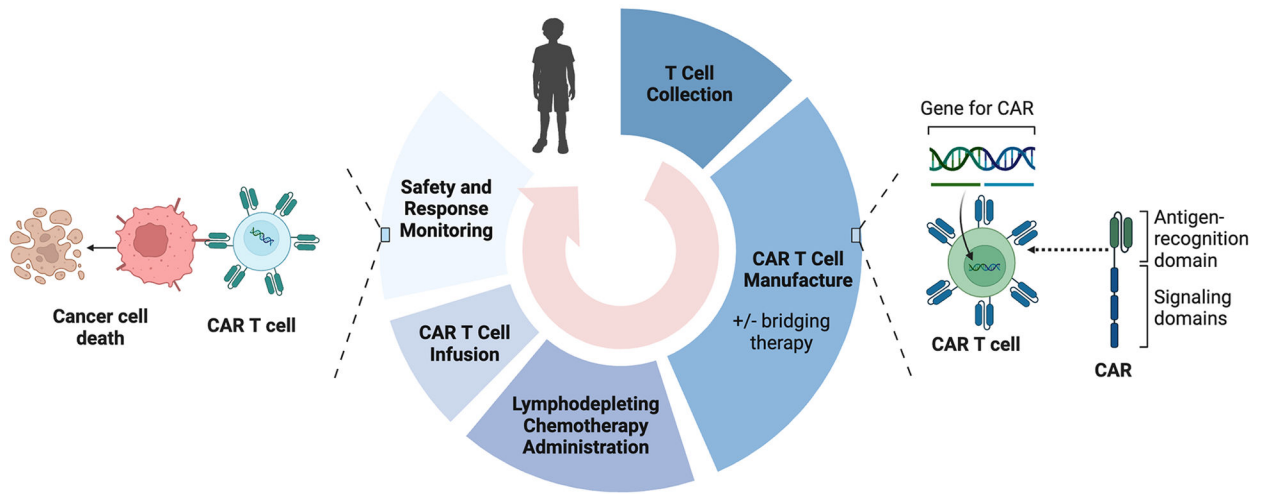
- [38]. Castriconi R, et al. , Identification of 4Ig-B7-H3 as a neuroblastoma-associated molecule that exerts a protective role from an NK cell-mediated lysis, *Proc. Natl. Acad. Sci. U. S. A* 101 (2004) 12640–12645. [PubMed: 15314238]
- [39]. Majzner RG, et al. , CAR T cells targeting B7-H3, a pan-cancer antigen, demonstrate potent preclinical activity against pediatric solid tumors and brain tumors, *Clin. Cancer Res* 25 (2019) 2560–2574. [PubMed: 30655315]
- [40]. Du H, et al. , Antitumor responses in the absence of toxicity in solid tumors by targeting B7-H3 via chimeric antigen receptor T cells, *Cancer Cell* 35 (2019) 221–237. e228.
- [41]. Loo D, et al. , Development of an Fc-enhanced anti-B7-H3 monoclonal antibody with potent antitumor activity, *Clin. Cancer Res* 18 (2012) 3834–3845. [PubMed: 22615450]
- [42]. Kendsersky NM, et al. , The B7-H3-targeting antibody-drug conjugate m276-SL-PBD is potently effective against pediatric cancer preclinical solid tumor models, *Clin. Cancer Res* 27 (2021) 2938–2946. [PubMed: 33619171]
- [43]. Powderly J, et al. , Interim results of an ongoing Phase I, dose escalation study of MGA271 (Fc-optimized humanized anti-B7-H3 monoclonal antibody) in patients with refractory B7-H3-expressing neoplasms or neoplasms whose vasculature expresses B7-H3, *J. Immunother. Cancer* 3 (2015) 1–2. [PubMed: 25648675]
- [44]. Kramer K, et al. , Compartmental intrathecal radioimmunotherapy: results for treatment for metastatic CNS neuroblastoma, *J. Neurooncol* 97 (2010) 409–418. [PubMed: 19890606]
- [45]. Modak S, et al., Intraperitoneal Radioimmunotherapy for Desmoplastic Small Round Cell Tumor: Results of a PHASE I Study (Clinicaltrials. Gov Identifier [NCT01099644](https://clinicaltrials.gov/ct2/show/study/NCT01099644)), S52-S52, in: *PEDIATRIC BLOOD & CANCER*, Vol. 64, WILEY 111 RIVER ST, HOBOKEN 07030–5774, NJ USA, 2017.
- [46]. Liu C, et al. , Targeting the immune checkpoint B7-H3 for next-generation cancer immunotherapy, *Cancer Immunol. Immunother* 71 (2022) 1549–1567. [PubMed: 34739560]
- [47]. Brignole C, et al. , Antitumor activity of the investigational B7-H3 antibody-drug conjugate, vobramitamab duocarmazine, in preclinical models of neuroblastoma, *J. Immunother. Cancer* 11 (2023).
- [48]. Rasic P, et al. , Targeting B7-H3-a novel strategy for the design of anticancer agents for extracranial pediatric solid tumors treatment, *Molecules* 28 (2023).
- [49]. Fan R, et al. , Engineering MMP-2 activated nanoparticles carrying B7-H3 bispecific antibodies for ferroptosis-enhanced glioblastoma immunotherapy, *ACS Nano* 17 (2023) 9126–9139. [PubMed: 37097811]
- [50]. Huang C, et al. , Combination therapy with B7H3-redirected bispecific antibody and Sorafenib elicits enhanced synergistic antitumor efficacy, *Theranostics* 10 (2020) 10498–10512. [PubMed: 32929362]
- [51]. Scribner JA, et al. , Preclinical development of MGC018, a duocarmycin-based antibody-drug conjugate targeting B7-H3 for solid cancer, *Mol. Cancer Ther* 19 (2020) 2235–2244. [PubMed: 32967924]
- [52]. Zhang Q, et al. , B7-H3 targeted CAR-T cells show highly efficient anti-tumor function against osteosarcoma both in vitro and in vivo, *BMC Cancer* 22 (2022) 1124. [PubMed: 36320072]
- [53]. Zhang S, et al. , B7-H3 specific CAR T cells for the naturally occurring, spontaneous canine sarcoma model, *Mol. Cancer Ther* 21 (2022) 999–1009. [PubMed: 35405743]
- [54]. Talbot LJ, et al. , A novel orthotopic implantation technique for osteosarcoma produces spontaneous metastases and illustrates dose-dependent efficacy of B7-H3-CAR T cells, *Front. Immunol* 12 (2021) 691741. [PubMed: 34211478]
- [55]. Hidalgo L, et al. , Switchable CAR T cell strategy against osteosarcoma, *Cancer Immunol. Immunother* 72 (2023) 2623–2633. [PubMed: 37062034]
- [56]. Murty S, et al. , PET reporter gene imaging and ganciclovir-mediated ablation of chimeric antigen receptor T cells in solid tumors, *Cancer Res.* 80 (2020) 4731–4740. [PubMed: 32958548]
- [57]. Cao JW, et al. , Canine osteosarcoma as a solid tumor model for evaluation of dual valent B7H3-CXCR2 CAR T cell therapy, *J. Immunol* 210 (2023), 159.122–159.122.

- [58]. Timpanaro A, et al. , Surfaceome profiling of cell lines and patient-derived xenografts confirm FGFR4, NCAM1, CD276, and highlight AGRL2, JAM3, and L1CAM as surface targets for rhabdomyosarcoma, *Int. J. Mol. Sci* 24 (2023).
- [59]. Kanayama T, et al. , Reduced B7-H3 expression by PAX3-FOXO1 knockdown inhibits cellular motility and promotes myogenic differentiation in alveolar rhabdomyosarcoma, *Sci. Rep* 11 (2021) 18802. [PubMed: 34552155]
- [60]. Pulido R, Nunes-Xavier CE, Hopes on immunotherapy targeting B7-H3 in neuroblastoma, *Transl. Oncol* 27 (2023) 101580. [PubMed: 36327699]
- [61]. Gregorio A, et al. , Small round blue cell tumours: diagnostic and prognostic usefulness of the expression of B7-H3 surface molecule, *Histopathology* 53 (2008) 73–80. [PubMed: 18613926]
- [62]. Zhang H, et al. , Survival association and cell cycle effects of B7H3 in neuroblastoma, *J. Korean Neurosurg. Soc* 63 (2020) 707–716. [PubMed: 32580265]
- [63]. Tan WQ, Chen G, Ye M, Jia B, Artemether regulates chemosensitivity to doxorubicin via regulation of B7-H3 in human neuroblastoma cells, *Med. Sci. Monit* 23 (2017) 4252–4259. [PubMed: 28866709]
- [64]. Dondero A, et al. , Multiparametric flow cytometry highlights B7-H3 as a novel diagnostic/therapeutic target in GD2neg/low neuroblastoma variants, *J. Immunother. Cancer* 9 (2021).
- [65]. Haydar D, et al. , Cell-surface antigen profiling of pediatric brain tumors: B7-H3 is consistently expressed and can be targeted via local or systemic CAR T-cell delivery, *Neuro Oncol.* 23 (2021) 999–1011. [PubMed: 33320196]
- [66]. Maachani UB, et al. , B7-H3 as a prognostic biomarker and therapeutic target in pediatric central nervous system tumors, *Transl. Oncol* 13 (2020) 365–371. [PubMed: 31887631]
- [67]. Tang X, et al. , Bioactivity and safety of B7-H3-targeted chimeric antigen receptor T cells against anaplastic meningioma, *Clin. Transl. Immunol* 9 (2020) e1137.
- [68]. Tang X, et al. , Administration of B7-H3 targeted chimeric antigen receptor-T cells induce regression of glioblastoma, *Signal Transduct. Target Ther* 6 (2021) 125. [PubMed: 33767145]
- [69]. Nehama D, et al. , B7-H3-redirected chimeric antigen receptor T cells target glioblastoma and neurospheres, *EBioMedicine* 47 (2019) 33–43. [PubMed: 31466914]
- [70]. Picarda E, Ohaegbulam KC, Zang X, Molecular pathways: targeting B7-H3 (CD276) for human cancer immunotherapy, *Clin. Cancer Res* 22 (2016) 3425–3431. [PubMed: 27208063]
- [71]. Li S, et al. , B7-H3 specific CAR-T cells exhibit potent activity against prostate cancer, *Cell Death Discov.* 9 (2023) 147. [PubMed: 37149721]
- [72]. Mendes AA, et al. , Association of B7-H3 expression with racial ancestry, immune cell density, and androgen receptor activation in prostate cancer, *Cancer* 128 (2022) 2269–2280. [PubMed: 35333400]
- [73]. Parker AS, et al. , Evaluation of B7-H3 expression as a biomarker of biochemical recurrence after salvage radiation therapy for recurrent prostate cancer, *Int. J. Radiat. Oncol. Biol. Phys* 79 (2011) 1343–1349. [PubMed: 20598810]
- [74]. Roth TJ, et al. , B7-H3 ligand expression by prostate cancer: a novel marker of prognosis and potential target for therapy, *Cancer Res.* 67 (2007) 7893–7900. [PubMed: 17686830]
- [75]. Zhang Y, et al. , Targeting radiation-resistant prostate cancer stem cells by B7-H3 CAR T cells, *Mol. Cancer Ther* 20 (2021) 577–588. [PubMed: 33653946]
- [76]. Zang X, et al. , Tumor associated endothelial expression of B7-H3 predicts survival in ovarian carcinomas, *Mod. Pathol* 23 (2010) 1104–1112. [PubMed: 20495537]
- [77]. Mao Y, et al. , B7-H1 and B7-H3 are independent predictors of poor prognosis in patients with non-small cell lung cancer, *Oncotarget* 6 (2015) 3452–3461. [PubMed: 25609202]
- [78]. Liu J, et al. , Targeting B7-H3 via chimeric antigen receptor T cells and bispecific killer cell engagers augments antitumor response of cytotoxic lymphocytes, *J. Hematol. Oncol* 14 (2021) 21. [PubMed: 33514401]
- [79]. Sun F, Yu X, Ju R, Wang Z, Wang Y, Antitumor responses in gastric cancer by targeting B7H3 via chimeric antigen receptor T cells, *Cancer Cell Int.* 22 (2022) 50. [PubMed: 35101032]
- [80]. Yamato I, et al. , Clinical importance of B7-H3 expression in human pancreatic cancer, *Br. J. Cancer* 101 (2009) 1709–1716. [PubMed: 19844235]

- [81]. Xuan Y, et al. , Targeting CD276 by CAR-T cells induces regression of esophagus squamous cell carcinoma in xenograft mouse models, *Transl. Oncol* 14 (2021) 101138. [PubMed: 34052626]
- [82]. Yue G, et al. , CD276 suppresses CAR-T cell function by promoting tumor cell glycolysis in esophageal squamous cell carcinoma, *J. Gastrointest. Oncol* 12 (2021) 38–51. [PubMed: 33708423]
- [83]. Fan S, et al. , B7-H3 chimeric antigen receptor-modified T cell shows potential for targeted treatment of acute myeloid leukaemia, *Eur. J. Med. Res* 28 (2023) 129. [PubMed: 36941687]
- [84]. Lichtman EI, et al. , Preclinical evaluation of B7-H3-specific chimeric antigen receptor T cells for the treatment of acute myeloid leukemia, *Clin. Cancer Res* 27 (2021) 3141–3153. [PubMed: 33531429]
- [85]. Zi Z, Zhao H, Wang H, Ma X, Wei F, B7-H3 chimeric antigen receptor redirected T cells target anaplastic lymphoma kinase-positive anaplastic large cell lymphoma, *Cancers (Basel)* 12 (2020).
- [86]. Zheng M, et al. , Efficacy of B7-H3-redirection BiTE and CAR-T immunotherapies against extranodal nasal natural killer/T cell lymphoma, *Transl. Oncol* 13 (2020) 100770. [PubMed: 32298986]
- [87]. Vitanza NA, et al. , Intraventricular B7-H3 CAR T cells for diffuse intrinsic pontine glioma: preliminary first-in-human bioactivity and safety, *Cancer Discov.* 13 (2023) 114–131. [PubMed: 36259971]
- [88]. Fauci JM, et al. , Monoclonal antibody-based immunotherapy of ovarian cancer: targeting ovarian cancer cells with the B7-H3-specific mAb 376.96, *Gynecol. Oncol* 132 (2014) 203–210. [PubMed: 24216048]
- [89]. Birley K, et al. , A novel anti-B7-H3 chimeric antigen receptor from a single-chain antibody library for immunotherapy of solid cancers, *Mol. Ther. Oncol* 26 (2022) 429–443.
- [90]. Li D, et al. , Camel nanobody-based B7-H3 CAR-T cells show high efficacy against large solid tumours, *Nat. Commun* 14 (2023) 5920. [PubMed: 37739951]
- [91]. Weinkove R, George P, Dasyam N, McLellan AD, Selecting costimulatory domains for chimeric antigen receptors: functional and clinical considerations, *Clin. Transl. Immunol* 8 (2019) e1049.
- [92]. Kontos F, et al. , B7-H3: an attractive target for antibody-based immunotherapy, *Clin. Cancer Res* 27 (2021) 1227–1235. [PubMed: 33051306]
- [93]. Seaman S, et al. , Eradication of tumors through simultaneous ablation of CD276/B7-H3-positive tumor cells and tumor vasculature, *e508, Cancer Cell* 31 (2017) 501–515. e508. [PubMed: 28399408]
- [94]. Dimitri A, Herbst F, Fraietta JA, Engineering the next-generation of CAR T-cells with CRISPR-Cas9 gene editing, *Mol. Cancer* 21 (2022) 78. [PubMed: 35303871]
- [95]. Qiao Y, et al. , Enhancement of CAR-T cell activity against cholangiocarcinoma by simultaneous knockdown of six inhibitory membrane proteins, *Cancer Commun. (Lond.)* 43 (2023) 788–807. [PubMed: 37282786]
- [96]. Huang B, et al. , B7-H3 specific T cells with chimeric antigen receptor and decoy PD-1 receptors eradicate established solid human tumors in mouse models, *Oncoimmunology* 9 (2020) 1684127. [PubMed: 32002297]
- [97]. Zhang X, et al. , Highly proliferative and hypodifferentiated CAR-T cells targeting B7-H3 enhance antitumor activity against ovarian and triple-negative breast cancers, *Cancer Lett.* 572 (2023) 216355. [PubMed: 37597651]
- [98]. Bell M LS, Sejdiu BI, Ibanez J, Shi H, Sun X, Meng X, Nguyen P, Sutton M, Wagner J, KC A, Langfitt D, Patil SL, Tan H, Pandey RV, Li Y, Yuan Z-F, Anido AA, Ho M, Sheppard H, Vogel P, Yu J, Peng J, Chi H, Babu MM, Krenciute G, Gottschalk S, Modular chimaeric cytokine receptors with leucine zippers enhance the antitumour activity of CAR T cells via JAK/STAT signalling. *Nat. Biomed. Eng. Accept. Publ* (2023).
- [99]. Li H, et al. , Targeting brain lesions of non-small cell lung cancer by enhancing CCL2-mediated CAR-T cell migration, *Nat. Commun* 13 (2022) 2154. [PubMed: 35443752]
- [100]. Wang L, et al. , A novel microenvironment regulated system CAR-T (MRS.CAR-T) for immunotherapeutic treatment of esophageal squamous carcinoma, *Cancer Lett.* 568 (2023) 216303. [PubMed: 37422126]

- [101]. Tousley AM, et al. , Co-opting signalling molecules enables logic-gated control of CAR T cells, *Nature* 615 (2023) 507–516. [PubMed: 36890224]
- [102]. Chockley PJ, Ibanez-Vega J, Krenciute G, Talbot LJ, Gottschalk S, Synapse-tuned CARs enhance immune cell anti-tumor activity, *Nat. Biotechnol* 41 (2023) 1434–1445. [PubMed: 36732477]
- [103]. Pinto N, et al. , STRIVE-02: A first-in-human phase 1 trial of systemic B7H3 CAR T cells for children and young adults with relapsed/refractory solid tumors, *J. Clin. Oncol* 40 (2022).
- [104]. Theruvath J, et al. , Locoregionally administered B7-H3-targeted CAR T cells for treatment of atypical teratoid/rhabdoid tumors, *Nat. Med* 26 (2020) 712–719. [PubMed: 32341579]
- [105]. Tang X, et al. , B7-H3 as a novel CAR-T therapeutic target for glioblastoma, *Mol. Ther. Oncolyt* 14 (2019) 279–287.
- [106]. Vitanza NA, et al. , Locoregional CAR T cells for children with CNS tumors: clinical procedure and catheter safety, *Neoplasia* 36 (2023) 100870. [PubMed: 36599192]
- [107]. Akel S, et al. , Preparation of cryopreserved chimeric antigen receptor T cells for the locoregional delivery to the neural axis, *Cytotherapy* (2023).
- [108]. Hu G, et al. , Case report: B7-H3 CAR-T therapy partially controls tumor growth in a basal cell carcinoma patient, *Front. Oncol* 12 (2022) 956593. [PubMed: 36059640]
- [109]. Wang G, et al. , CXCL11-armed oncolytic adenoviruses enhance CAR-T cell therapeutic efficacy and reprogram tumor microenvironment in glioblastoma, *Mol. Ther* 31 (2023) 134–153. [PubMed: 36056553]
- [110]. Huang J, et al. , Interleukin-7-loaded oncolytic adenovirus improves CAR-T cell therapy for glioblastoma, *Cancer Immunol. Immunother* 70 (2021) 2453–2465. [PubMed: 33543339]
- [111]. Lei X, et al. , A pan-histone deacetylase inhibitor enhances the antitumor activity of B7-H3-specific CAR T cells in solid tumors, *Clin. Cancer Res* 27 (2021) 3757–3771. [PubMed: 33811153]
- [112]. Lee HW, et al. A high-content screen identified ingenol-3-angelate as an enhancer of B7-H3-CAR T cell activity by increasing B7-H3 protein expression on the target cell surface via protein kinase C alpha activation. *bioRxiv*, 2023.2005. 2026.542130 (2023).
- [113]. Zhang Z, et al. , A drug screening to identify novel combinatorial strategies for boosting cancer immunotherapy efficacy, *J. Transl. Med* 21 (2023) 23. [PubMed: 36635683]
- [114]. Wang Y, et al. , Stressed target cancer cells drive nongenetic reprogramming of CAR T cells and solid tumor microenvironment, *Nat. Commun* 14 (2023) 5727. [PubMed: 37714830]
- [115]. Ventin M, et al. , B7-H3-targeted CAR T cell activity is enhanced by radiotherapy in solid cancers, *Front. Oncol* 13 (2023) 1193963. [PubMed: 37483496]
- [116]. Wang T, et al. , Preconditioning of radiotherapy enhances efficacy of B7-H3-CAR-T in treating solid tumor models, *Life Sci.* (2023) 122024. [PubMed: 37574043]
- [117]. Roybal KT, et al. , Precision tumor recognition by T cells with combinatorial antigen-sensing circuits, *Cell* 164 (2016) 770–779. [PubMed: 26830879]
- [118]. Moghimi B, et al. , Preclinical assessment of the efficacy and specificity of GD2-B7H3 SynNotch CAR-T in metastatic neuroblastoma, *Nat. Commun* 12 (2021) 511. [PubMed: 33479234]
- [119]. Halliwell E, et al. , Targeting of low ALK antigen density neuroblastoma using AND logic-gate engineered CAR-T cells, *Cytotherapy* 25 (2023) 46–58. [PubMed: 36396552]
- [120]. Tian M, et al. , An optimized bicistronic chimeric antigen receptor against GPC2 or CD276 overcomes heterogeneous expression in neuroblastoma, *J. Clin. Invest* 132 (2022).
- [121]. Yang M, et al. , Tandem CAR-T cells targeting CD70 and B7-H3 exhibit potent preclinical activity against multiple solid tumors, *Theranostics* 10 (2020) 7622–7634. [PubMed: 32685008]
- [122]. Cao G, et al. , GPC3-targeted CAR-T cells secreting B7H3-targeted BiTE exhibit potent cytotoxicity activity against hepatocellular carcinoma cell in the in vitro assay, *Biochem. Biophys. Rep* 31 (2022) 101324. [PubMed: 36032401]
- [123]. Srivastava S, et al. , Logic-gated ROR1 chimeric antigen receptor expression rescues T cell-mediated toxicity to normal tissues and enables selective tumor targeting, *e488, Cancer Cell* 35 (2019) 489–503. e488. [PubMed: 30889382]

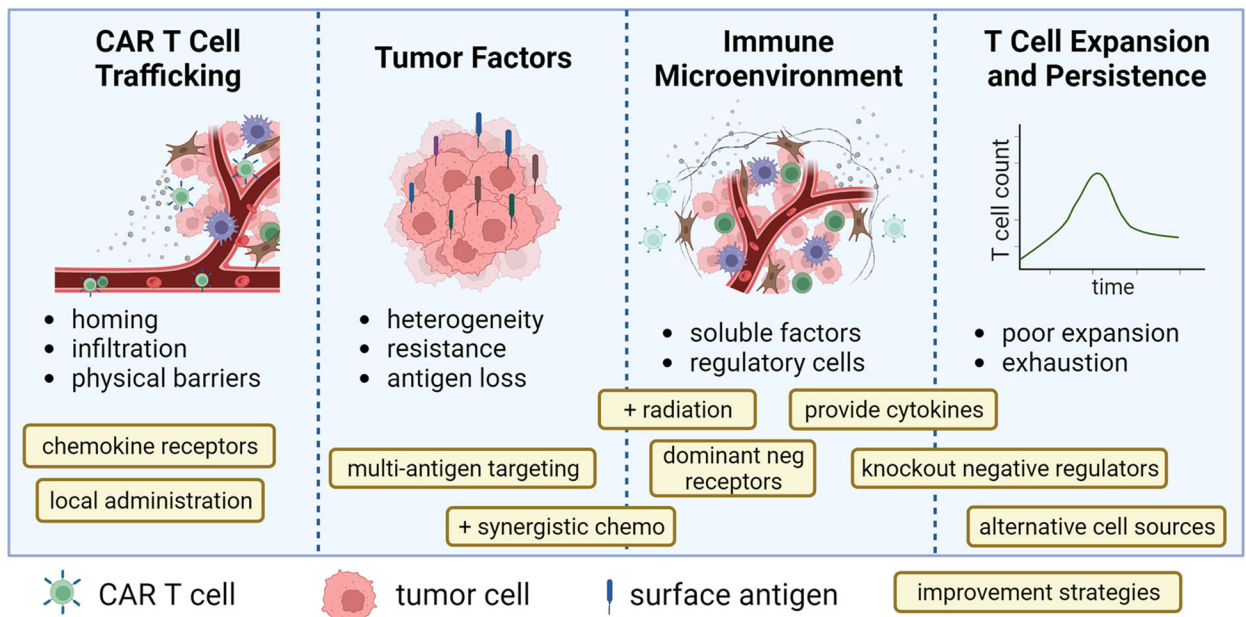
- [124]. Morandi F, Yazdanifar M, Cocco C, Bertaina A, Airoidi I, Engineering the bridge between innate and adaptive immunity for cancer immunotherapy: focus on gammadelta T and NK cells, *Cells* 9 (2020).
- [125]. Wang Y, et al. , B7H3-targeting chimeric antigen receptor modification enhances antitumor effect of Vgamma9Vdelta2 T cells in glioblastoma. *J. Transl. Med* 21 (2023) 672. [PubMed: 37770968]
- [126]. Grote S, et al. , In vitro evaluation of CD276-CAR NK-92 functionality, migration and invasion potential in the presence of immune inhibitory factors of the tumor microenvironment, *Cells* 10 (2021).
- [127]. Yang S, et al. , Targeting B7-H3 immune checkpoint with chimeric antigen receptor-engineered natural killer cells exhibits potent cytotoxicity against non-small cell lung cancer, *Front. Pharm* 11 (2020) 1089.
- [128]. Chaudhry K, et al. , Co-transducing B7H3 CAR-NK cells with the DNR preserves their cytolytic function against GBM in the presence of exogenous TGF-beta, *Mol. Ther. Methods Clin. Dev* 27 (2022) 415–430. [PubMed: 36381305]



**Fig. 1.** CAR T cell therapy strategy for pediatric patients with solid tumors. Inserts highlight CAR T cell generation and CAR T cell tumor cell killing. For additional details see text.



## Challenges to B7-H3-CAR T Cell Therapy



**Fig. 2.** Strategies to Enhance B7-H3-CAR T Cell Therapy. Primary challenges facing chimeric antigen receptor (CAR) T cell therapy for solid tumors; Neg: negative; +: combination therapy. For additional details see text.

Table 1

## Pediatric B7-H3 CAR T Cell Clinical Trials.

NCT/status	Institution/Sponsor	CAR Design	Indication	Age*	Administration
<b>Extracranial Solid Tumors</b>					
NCT04483778 <i>Active, not recruiting</i>	Seattle Children's Hospital	Arm A: B7-H3.4-1BB $\zeta$ Arm B: B7-H3.4-1BB $\zeta$ + CD19.4-1BB $\zeta$	Recurrent/refractory non-primary CNS solid tumor	26	Systemic infusion
NCT04897321 <i>3CAR Recruiting</i>	St. Jude Children's Research Hospital	B7-H3.CD28 $\zeta$ +41BBL	Relapsed/refractory B7-H3 positive solid tumor	21	Systemic infusion
NCT04433221 <i>Recruiting</i>	Shenzhen Geno-Immune Medical Institute	multi-antigen specific	Stage III/IV or recurrent sarcomas	1-75	CAR T cells in combination therapy
NCT04637503 <i>4SCAR-T Recruiting</i>	Shenzhen Geno-Immune Medical Institute	B7-H3, GD2, and/or PSMA-targeted	Relapsed/refractory neuroblastoma	1-65	Systemic infusion
NCT05562024 <i>Recruiting</i>	PersonGen BioTherapeutics (Suzhou)	B7-H3 (TAA-06)	B7-H3 positive recurrent/ refractory neuroblastoma	1	Systemic infusion
<b>Primary Central Nervous System tumors</b>					
NCT04185038 <i>BrainChild-03 Recruiting</i>	Seattle Children's Hospital	B7-H3.4-1BB $\zeta$ /EGFRt	Recurrent/ refractory CNS tumors, DIPG, or DMG	1-26	-Local administration Arm A: tumor resection cavity (supratentorial) Arms B and C: ventricular -repeat weekly infusions
NCT05768880 <i>BrainChild-04 Recruiting</i>	Seattle Children's Hospital	B7-H3, EGFR806, HER2, and IL13-zetakine	Recurrent/ refractory CNS tumors, DIPG, or DMG	1-26	-Local administration -repeat weekly infusions
NCT05835687 <i>Loc3CAR Recruiting</i>	St. Jude Children's Research Hospital	B7-H3.CD28 $\zeta$ +41BBL	Arm A: B7-H3 positive relapsed/ refractory non-brainstem primary CNS tumors Arm B: brainstem high-grade neoplasms	21	-Local administration -repeat weekly infusions

Child (birth-17), and Recruiting OR Active, not recruiting, as of September 25, 2023

\* years; DIPG = diffuse intrinsic pontine glioma; DMG = diffuse midline glioma