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Genetic footprint of population fragmentation and contemporary collapse in a freshwater cetacean

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Understanding demographic trends and patterns of gene flow in an endangered species is crucial for devising conservation strategies. Here, we examined the extent of population structure and recent evolution of the critically endangered Yangtze finless porpoise (*Neophocaena asiaeorientalis asiaeorientalis*). By analysing genetic variation at the mitochondrial and nuclear microsatellite loci for 148 individuals, we identified three populations along the Yangtze River, each one connected to a group of admixed ancestry. Each population displayed extremely low genetic diversity, consistent with extremely small effective size (≤ 92 individuals). Habitat degradation and distribution gaps correlated with highly asymmetric gene-flow that was inefficient in maintaining connectivity between populations. Genetic inferences of historical demography revealed that the populations in the Yangtze descended from a small number of founders colonizing the river from the sea during the last Ice Age. The colonization was followed by a rapid population split during the last millennium predating the Chinese Modern Economy Development. However, genetic diversity showed a clear footprint of population contraction over the last 50 years leaving only ~2% of the pre-collapsed size, consistent with the population collapses reported from field studies. This genetic perspective provides background information for devising mitigation strategies to prevent this species from extinction.

Dispersal and gene flow in a meta-population maintain local demographic and genetic variation, thus increasing the probability of species persistence^{1,2}. Persistence of wide-ranging animals occupying fragmented landscapes depends on the matrix quality of the habitat and the ability of individuals to move among habitat patches³, and corridors facilitating this movement across such landscape⁴⁻⁶. Along the Yangtze River (China, Fig. 1), anthropogenic activities of the past 50 years have put intense pressure on the freshwater ecosystem leading to habitat degradation, species range fragmentation and extinction of some emblematic endemic species, such as the Yangtze River dolphin or baiji (*Lipotes vexillifer*)⁷. Today, the Yangtze finless porpoise (*Neophocaena asiaeorientalis asiaeorientalis* or YFP) has become the only surviving freshwater cetacean now found in China and the world's only freshwater porpoise species⁸.

Endemic to the Yangtze River drainage (Fig. 1), the YFP is now primarily restricted to the middle-lower Yangtze Channel and two large appended lakes: Dongting Lake (DT) and Poyang Lake (PY)⁹. The subspecies has occasionally been reported from some of the larger adjacent tributaries though this is now rare¹⁰⁻¹². The amount of river and lake habitat available to this subspecies is relatively small compared to that available to marine populations of finless porpoises, which occur in coastal waters from Japan to the Arabian Sea⁸. However, YFP abundance has suffered from dramatic reductions from 2,500 individuals in 1991¹⁰ to 1,800 in the end of 2006, as estimated by the Yangtze Freshwater Dolphin Expedition in 2006 (YFDE2006)¹³. More recent surveys conducted

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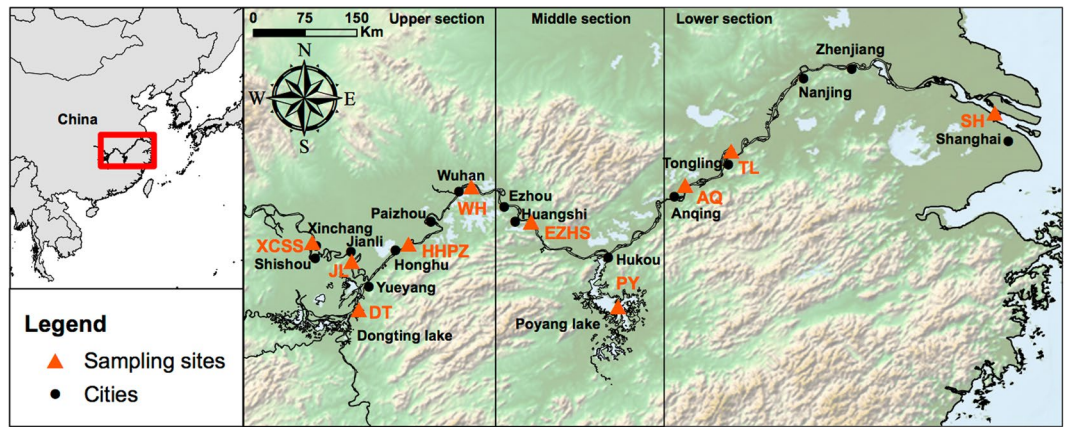


Figure 1. Maps showing the sampling distribution of the Yangtze finless porpoises in the Yangtze River. The top-left insert shows the location of the studied area highlighted by a red rectangle. On the right, the map shows the sampling locations (orange triangles) and their acronyms based on the neighbouring cities. Figure created using ArcGIS 10.3 software using the open source data from the ETOPO1 Global Relief Model⁶³ (<https://www.ngdc.noaa.gov/mgg/global/>).

during the Yangtze Freshwater Dolphin Expedition conducted in 2012 (YFDE2012) reported that populations declined even further to ~1,040 individuals, including ~500 porpoises in the Yangtze main stream, ~450 in PY and ~90 in DT¹⁴. With such rapid range contraction, Mei *et al.*¹⁵ estimated that the YFP may become extinct within the next 60 years or less¹⁴. The YFP was thus recently reclassified as a Critically Endangered sub-species on the IUCN Red List⁹. As a top predator, the survival of the finless porpoise depends heavily on habitat suitability, food availability and maintenance of corridors allowing dispersal between populations. However, with increasing underwater noise from boat traffic and incidental catches in fisheries, food and habitat resources for the species have become increasingly scattered and fragmented, and corridors across the landscape have been compromised by the booming of the Chinese economy over the last decades¹³. Despite these imminent threats, we still do not know how reduction in suitable habitats in the Yangtze River has reduced the number of breeding porpoises and how habitat fragmentation has impacted connectivity between populations and the population structure itself. This information is extremely difficult to quantify using direct observations. On the other hand, population genetic approaches can provide key insights about the current population structure and connectivity and historical population demography by leaving detectable footprints on the genetic diversity and its geographic structure^{16,17}.

Previous phylogeographic analyses based on the non-coding mitochondrial Control Region (mtDNA-CR) of the finless porpoises from Chinese and Japanese waters documented evidence of a demographic expansion following the Last Glacial Maximum (LGM, ~24,000 yrs BP) and the colonization of Yangtze River from the Yellow Sea ~22,000 yrs BP ago^{18,19}. A subsequent study within the Yangtze River by Chen *et al.*²⁰ used mtDNA-CR and nuclear microsatellite loci and revealed evidence of genetic subdivisions within the YFP populations suggestive of population fragmentation. However, the fine scale population structure remained unclear and the processes shaping the genetic variation in the YFP unknown. Changes in connectivity between populations, dynamics of population expansion-contraction and demographic history are potent factors shaping the genetic variation in the YFP, but these were not investigated in further details so far.

In this study, we address these above questions by re-analysing the previously published data set of Chen *et al.*²⁰ that combined fast evolving microsatellite loci together with more slowly evolving sequences from the mtDNA-CR. Specifically, in contrast with previous studies, we used a statistical population genetic framework in order to (i) resolve the fine-scale population structure using a combination of individual-based multivariate and Bayesian clustering approaches designed for weak genetic structure when it exists; (ii) estimate past and contemporaneous effective population sizes and connectivity among populations; and (iii) reconstruct the demographic history best fitting with the genetic diversity observed in the populations of the YFP using a quantitative model-based inferential population genetic framework relying on Approximate Bayesian Computation (ABC) approaches^{16,17,21–23}. Determining and quantifying to which extent populations in the YFP are fragmenting, losing connectivity, and the magnitudes of the demographic trends are critical knowledge for designing conservation and management plans. For example, we still do not understand whether population fragmentation and decline have been triggered only recently by human activities during the past 50 years or if these patterns were initiated earlier by more long-term ecological and evolutionary processes and exacerbated lately during the Anthropocene.

Results

Genetic structure and diversity of the Yangtze finless porpoises. The final data set consisted of 148 individuals sampled along the Yangtze River and the two main lakes (Dongting Lake (DT) and Poyang Lake (PY), Fig. 1) genotyped for 11 microsatellite loci and sequenced for a 597 base-pairs fragment of the hyper-variable region 1 (see Table 1 and materials and methods). The clustering of the microsatellite data using the Bayesian clustering algorithm of *STRUCTURE*^{24–26} provided consistent results over 10 replicated runs performed for each number *K* of cluster tested (Fig. 2a). The probability of the data greatly increased when two genetic clusters were

	Total	PY	TL	XCSS	Admix
Microsatellite loci					
$N_{mic.} \pm SD$ (max)	148	57.2 ± 0.32 (58)	16.0 ± 0.4 (17)	16.4 ± 0.36 (17)	48.1 ± 1.3 (53)
NA	—	1.4%	4.8%	3.7%	9.3%
$A_r \pm SE$	—	4.82 ± 0.40	5.19 ± 0.52	5.25 ± 0.46	5.93 ± 0.45
$pA \pm SE$	—	0.38 ± 0.09	0.55 ± 0.21	0.48 ± 0.12	0.69 ± 0.18
$H_o \pm SD$	—	0.60 ± 0.06	0.70 ± 0.047	0.69 ± 0.05	0.72 ± 0.04
$H_e \pm SD$	—	0.62 ± 0.05	0.62 ± 0.05	0.63 ± 0.04	0.68 ± 0.03
$F_{IS} \pm SD$	—	0.041 ± 0.03 ^{ns}	-0.123 ± 0.06 ^{ns}	-0.112 ± 0.03 ^{ns}	-0.057 ± 0.03 ^{ns}
M_{GW}^\dagger	—	0.51***	0.46**	0.44**	0.53**
MtDNA control region					
N_{mtDNA}	129	56	17	16	37
S	7	2	1	0	5
Singleton	4	0	0	0	3
Shared P.	3	2	1	0	2
#hap.	7	3	2	1	5
Hd	0.57	0.53	0.22	0	0.62
π (per site, %)	0.112	0.096	0.037	0	0.150
θ_w (per site, %)	0.216	0.073	0.050	0	0.201
D [‡]	-1.08 ^{ns}	0.54 ^{ns}	-0.49 ^{ns}	0	-0.64 ^{ns}

Table 1. Genetic variation at the 11 microsatellites and mtDNA control region loci for each distinct population inferred from the *STRUCTURE* analysis. $N_{mic.}$, microsatellite average sample size (max); NA: average proportion of missing data per locus, A_r : allelic richness (estimated for a sample size of 26 individuals); pA : Private allelic richness (estimated for a sample size of 3 individuals); H_o and H_e : observed and expected heterozygosity; F_{IS} : Inbreeding coefficient; M_{GW} , M of Garza and Williamson³⁴; N_{mtDNA} , MtDNA sample size; S, number of segregating sites; *Singleton*: rare mutation observed only in one sequence among all; *Share P.*, shared polymorphism (mutation observed in at least two or more sequences) also known as parsimony informative site; #hap, number of haplotypes; Hd, haplotype diversity; π , nucleotide diversity; θ_w , theta from S or Theta-Watterson; D, Tajima's D. [†]The significance level of the M_{GW} statistic was evaluated in DIYABC⁷⁸ using 1×10^6 coalescent simulations under a scenario of constant effective population size. The P-value indicate the proportion of simulations which provide a value below the observed one. [‡]The significance of D values was estimated using 10 000 coalescent simulations in DNAsp⁷³. ns: not significant (p-value > 0.05); *p-value ≤ 0.05; **p-value ≤ 0.01; ***p-value ≤ 0.001.

modelled instead of one and showed the highest values on average over 10 replicates (Fig. S1). At $K > 2$, the increase in probability decreased in average; however some runs displayed the highest probability of all runs at $K = 3$ and 4 (Fig. S1). A visual inspection of the individual clustering for each K value (Fig. 2a) revealed that porpoises from the PY split from the other individuals of the Yangtze river at $K = 2$, suggesting that these porpoises are highly differentiated from the others. When higher K values were tested ($K = 3$ and 4) porpoises from PY, XCSS and TL localities were all identified as differentiated genetic units, while the porpoises from the in-between regions consisted of an admixed group sharing genetic ancestry with the three other populations (Fig. 2a,d). The three individuals at the mouth of the Yangtze River close to Shanghai city (SH) also seemed to depart from the other groups at $K = 4$, but the low sample size ($n = 3$) preclude any definitive conclusions. No further subdivision was observed beyond $K = 4$ (result not shown).

The results from *STRUCTURE* were further validated using two multivariate approaches that do not rely on model assumptions²⁷: a Discriminant Analysis of Principal Components (*DAPC*)²⁸ and a Principal Component Analysis (PCA)^{27,29}. Both methods showed a genetic structure consistent with the results of *STRUCTURE* (Figs 2b,c and S2). The *DAPC* provided a clear-cut discrimination of the four groups identify by *STRUCTURE* (Fig. 2c). The first discriminant function (DF) discriminated XCSS and PY and the second DF TL and SH. The other porpoises recognized as admixed in *STRUCTURE* were located at the intersection of the four other groups. The proportions of successful reassignment (based on the discriminant functions) of individuals to their original clusters was high (Fig. 2b): 100% for XCSS and SH, 94.8% for PY, 94.1% for TL, and 94.3% for the admixed porpoises. These large values indicate clear-cut clusters. Finally, the PCA provided a similar picture as the *DAPC* and *STRUCTURE*, but with more overlap among the groups (Fig. S2).

None of the identified populations displayed significant departures from Hardy-Weinberg expectations as shown by the F_{IS} values (Table 1). Porpoises from the admixed and TL groups showed the highest level of microsatellite genetic diversities and XCSS and PY the lowest, as estimated with the values of allelic richness (A_r), private allelic richness (pA) and expected heterozygosity (H_e) (Table 1 and Fig. S3). Only the two extremes groups, PY and Admix, showed a significant difference in A_r and H_e (Wilcoxon signed-ranked test p-value < 0.05). The mitochondrial genetic diversity followed a similar trend with the admixed group showing the highest haplotype and nucleotide diversity followed by TL and PY (Table 1 and Fig. 2d). We found only one haplotype fixed in the XCSS group.

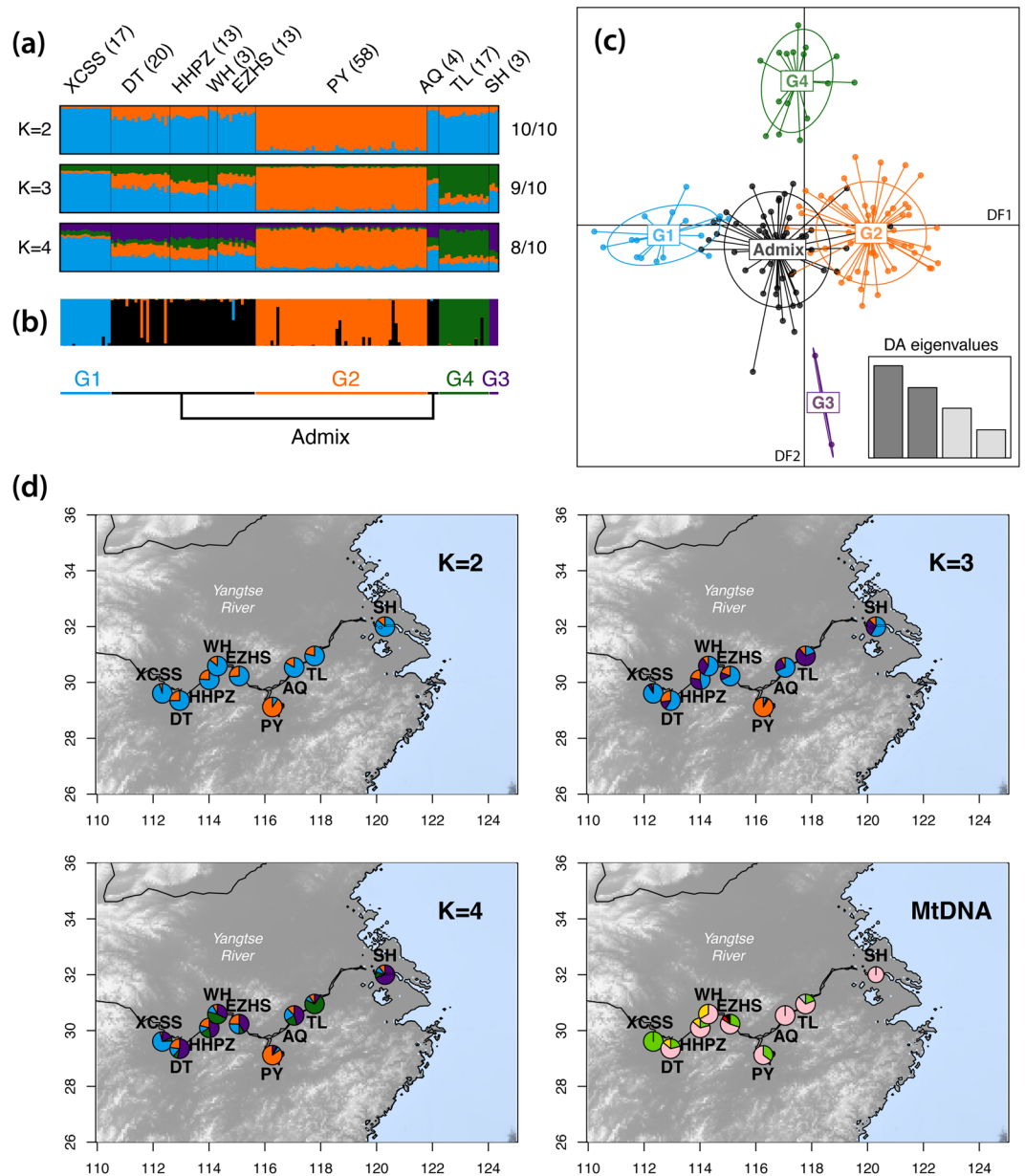


Figure 2. (a) Population structure estimated using the Bayesian clustering approach of *STRUCTURE*. Each individual is represented by a vertical line divided into K segments showing the admixture proportions from each cluster. Sample size in each locality is shown between brackets. Numbers on the right side of the barplot show the number of time this result was found out of the 10 replicates. (b) DAPC cluster membership probability plot of the 148 individuals. (c) Scatter plot showing the first two discriminant functions (DFs) of the DAPC. (d) Geographical distribution of the *STRUCTURE* admixture proportions and mitochondrial haplotype frequencies per localities. The mtDNA map is modified from Chen *et al.*²⁰. Panel (a) was created using CLUMPAK⁵⁹, panel (b,c) using R v.3.4.0⁶⁶, and panel (d) using R v.3.4.0⁶⁶, the package MARMAP v.0.9.5⁶², and the open source ETOPO1 Global Relief Model⁶³ (<https://www.ngdc.noaa.gov/mgg/global/>).

All populations showed significant differences in allelic frequencies for the microsatellite loci with F_{ST} values ranging from 0.023 to 0.070 (Table 2). Notably F_{ST} values between XCSS, PY and TL were relatively high (>0.05), while F_{ST} values were intermediate between the Admix group and each distinct population (between 0.02 and 0.03, Table 2). For the mitochondrial locus (Table 2), F_{ST} values were all significant except one (Admix vs. TL), and were especially strong between XCSS and all other groups, due to the fact that one haplotype is fixed in this population (Fig. 2d).

Contemporary effective population sizes and migration rates. We used two methods for estimating contemporary effective population size (N_e) in each population from the microsatellite data: *NeEstimator*³⁰ based on linkage disequilibrium among loci within population and *ONESAMP*³¹ relying on an Approximate Bayesian

(Tables S4, S5 and Fig. S5) shows that 61.4% of the simulated dataset under this SC1 were correctly identified using the ABC-LDA procedure, leading to an average Type-I error rate (false negative) of 4.3% ranging from 1.7% to 9.7% and a total prior error rate (*i.e.* the average misclassification error)^{37,38} of 38.6% (62.9% using the ABC-RF). Simulations under the nine other competing scenarios led to a Type-II error rate of <12.9% incorrect assignment to SC1 (false positives) and a power (87.1%) to discriminate the best scenario from the others (Table S4).

The second ABC step tested for occurrence of simple changes in effective population size (N_e) during the divergence of the three populations under six nested competing scenarios (Fig. 4bi, Tables S2 and S3). The scenario assuming a bottleneck in the ancestral population prior to population split (SC3) outcompeted the other five scenarios (*i.e.*, no change (SC1) or simple decline or expansion in the ancestral or daughter populations, SC2, SC4-6), with a probability of 78% and no overlap in 95%CI for the ABC-LDA, and $69.8 \pm 1.9\%$ under the ABC-RF (Fig. 4bi and Table S6). The sensitivity analysis showed that simulations generated with SC3 were more difficult to identify, with an average Type-I error rate of 12.4% (ranging between 1 and 21% error, depending on the scenario), and a total prior error rate of 62.7% under the ABC-LDA and 41.4% under the ABC-RF (Fig. S6 and Table S6). Nevertheless, the Type-II error rate (false positive) was only 7.2% and the power to discriminate this scenario from the others was 92.8%. Furthermore, the assessment of goodness-of-fit of this bottleneck scenario (SC3) to the data showed very good performance, as the simulations using this scenario and posterior distributions for each parameter, were able to reproduce all but one observed summary statistic, in contrast to all other scenarios (Tables S6 and S7).

The final step (Fig. 4bii, Tables S2 and S3) tested whether a recent demographic collapse in each population within the last 5 generations (or 50 years as reported in the literature¹⁵) could have produced a detectable genetic footprint. The scenario involving a recent collapse (SC2) in each population received a significantly higher probability (ABC-LDA 78.2%; ABC-RF: $69.2 \pm 2.0\%$) compared with the alternative scenario of constant N_e since the population split (Fig. 4bii, Fig. S6, Tables S8 and S9). Both Type-I and Type-II error rates were <16% indicating adequate power and sensitivity of our ABC analysis.

The model parameters estimated under the final best demographic scenario (SC2 in Fig. 4bii, Tables S10 and S11) suggest that the ancestral population would have been large ($N_{anc2} = 18,700$ individuals; 95%CI: [3,480 – 19,700]), and that a small fraction would have founded the Yangtze River populations $\sim 3,400$ 95%CI: [1300 – 41,000] years ago (T_{exp2}) and expanded to reach an effective size of about 5,660 individuals (N_{exp2} ; 95%CI: [2,900 – 9,840]). The three daughter populations would have then split from each other $\sim 1,030$ years ago (T_{isol2} ; 95%CI: [214 – 4,380]), and reached an effective size of about 2,000 individuals after their split. Each of these daughter populations would have gone through a significant decline leaving less than 2% of their pre-collapse size during the last 50 years (see Tables 3, S10 and S11).

Discussion

Our study shows that the present-day genetic diversity of finless porpoise in the Yangtze River (YFP) has been strongly influenced by an initial founder event that took place several thousand years ago, followed by a relatively recent split into 3 populations (XCSS, TL, PY), and a recent demographic collapse within the last 50 years. Indeed, consistent with previous studies^{18,19}, the ABC genetic inferences showed that a few individuals coming from a large ancestral population, likely a marine population of *Neophocaena asiaeorientalis sunameri* in the Yellow Sea, colonized the Yangtze River within the last 41 kyrs, and most likely during the Last Ice Age. The subsequent population split into three populations occurred between 200 and 5,000 years BP. This suggests that the population split was triggered before the intensification of human activities of the last 50 years in the Yangtze River. This event may thus have been related to the colonization process itself during the post-glacial period¹⁹ and/or other environmental or human-related factors. Consistent with the hypothesis of population split driven by post-glacial changes, episodes of contraction and expansion of the Yangtze River mainstream and the adjacent lakes occurred during the Holocene period³⁹. Interestingly, the Yangtze River mainstream and the appendage lakes, including the Poyang and Dongting lakes, retracted and shrank significantly during the late Holocene (from *ca.* 3000 years BP to now)^{39–41}. These environmental changes might have promoted the split of the ancestral population after the colonization of the Yangtze River.

Each population harboured a genetic footprint of dramatic population reduction. This especially affected the genetic diversity at the microsatellite markers which displayed very small values of the M_{GW} statistic (Table 1) characteristic of significant recent decline³⁴. This is also indicated by the ABC analysis supporting a scenario describing a drastic population reduction in each population within the last five generations (Fig. 4bii, SC2). According to this scenario, the reduction in effective size would have been massive since their current sizes would be less than $\sim 2\%$ of their pre-collapsed sizes (Fig. 4, Tables 3, S10 and S11). These genetic inferences are in line with the field estimates^{14,15,42}, reporting a continuous decline of the YFP since the 1980s^{10–15,43,44}. Half of the census populations in the main stem of the Yangtze River would have been lost within the past 15 years, with the abundance dropping from 2,500 porpoises in 1991¹⁰ to 1,225 in 2006¹³. With the 400 porpoises in the Poyang Lake and the 100 to 150 porpoises of the Dongting Lake, the total census size in the Yangtze River and the two adjacent lakes observed in 2006 was only 1,800 individuals. The most recent estimates from the YFDE2012 showed that the porpoises in the main stem of the Yangtze would have been reduced again by half with 505 individuals reported the middle and lower reaches of the Yangtze River and approximately 450 porpoises in PY, and 90 porpoises in DT¹⁴.

These very small census populations sizes (N) are consistent with the very small contemporary effective population size (N_e) estimated from genetic data (≤ 92 individuals) in each population, with the XCSS population being the smallest of all three differentiated populations (between 14 and 22 individuals, Table 3). Comparably low estimates have been reported in other cetacean species, such as the southern Iberian ecotype of harbour porpoise (*Phocoena phocoena meridionalis*) in European waters ($N_e \leq 80$ ^{17,45}) or the coastal ecotype of bottlenose

dolphins (*Tursiops truncatus*) in European waters ($N_e \leq 77^{46}$). However only highly endangered populations, such as the Borneo Orangutans (*Pongo pygmaeus*)⁴⁷ or the Canadian woodland caribou (*Rangifer tarandus*)⁴⁸, have N_e values as low as those observed in the XCSS population. Extremely low N_e clearly translate the very low genetic diversity observed in each population of the YFP and imply very low numbers of breeding individuals in each population of the Yangtze River and the adjacent lakes⁴⁹. However, drawing a more direct link between N_e and N is actually difficult considering our study design. Previous studies have shown that no direct relationship can be expected between N_e and N when generations are overlapping, sampling spans several years, includes multiple cohorts and age classes, and when immigration may occur^{50–52}.

The three YFP populations – XCSS, TL and PY – have not exchanged migrants over the last few generations according to our genetic estimates (Fig. 3 and Table S1). However, all three are or have been recently connected to the admixed group. We observed genetic evidence of unidirectional gene flow from the admixed group to XCSS and TL. This recent migration in the middle section of the Yangtze and XCSS in the upper section may be rare and/or no longer occurring based on the observed fixation of the mitochondrial haplotype in the XCSS population (Table 1 and Fig. 2d). This is further supported by the YFDE2006 and YFDE2012 surveys that reported increasing gaps in the distribution of the species in the upper section of the Yangtze River¹⁴. In contrast to the two other populations in the main stream river, gene flow was in the opposite direction in the Poyang Lake, from PY to the Admixed group (Fig. 3 and Table S1). This is consistent with field observations reporting groups of porpoises from PY moving to the main river stem in the morning and back to the lake in the afternoon⁴³. This result also supports previous assertion¹⁴ that immigration of porpoises from PY to the river may dampen population decline in the Yangtze River. Unfortunately, such migration is likely insufficient given the observed ongoing decline^{14,42}. In principle, the admixed group in the middle section of the Yangtze River could serve as a bridge connecting the three differentiated populations. However, our gene flow estimates do not support this (Fig. 3) and are consistent with the increasing observation of gaps in the distribution of the YFP and the loss of connectivity between populations.

Additional populations of YFP may exist in the Yangtze mainstream as the YFP is known to occur upstream and downstream from our study area (Fig. 1). For example, the three porpoises sampled around Shanghai city (SH) seem to belong to another differentiated unit (Fig. 2), but no definitive conclusions can be drawn at this time due to low sample size ($n = 3$). Nevertheless, the present study provides a representative view of the population genetic structure, connectivity and demographic trends that can help to define priority areas where conservation measures need to be taken.

Drastic reduction in population abundance has left a clearly detectable genetic footprint on the genetic diversity of the YFP and coincides with the loss of connectivity between populations as well as the intensification of human activities along the Yangtze River over the last 50 years. To ensure that connectivity between populations is maintained, mitigation of human impacts need to include the entire river catchment⁴⁴. For example, restricting fishing and sand-mining activities in the mouth area of Poyang Lake (Hukou, Fig. 1) could restore the lake-river migration of the YFP. Modification of current *in situ* reserves could improve connectivity between Ezhou and Zhenjiang (Fig. 1). Likewise, more active measures including a whole year fishing ban in the *in situ* natural reserves could certainly help¹⁴ and possible translocation of isolated individuals in the hope of increasing breeding opportunities could increase genetic diversity.

Material and Methods

Samples collection and DNA extraction. A total of 153 Yangtze finless porpoises were sampled between 1998 and 2011 across the distribution range (Fig. 1), including 3 from Shanghai (SH), 17 from Tongling (TL), 5 from Anqing (AQ), 15 from Ezhou-Huangshi (EZHS), 3 from Wuhan (WH), 14 from Honghu-Paizhou (HHPZ), 2 from Jianli (JL), 16 from Xingchang-Shishou (XCSS), 20 from Dongting Lake (DT) and 58 from Poyang Lake (PY). A detailed description of the sampling procedure and genomic DNA extraction is provided in Chen *et al.*²⁰. Briefly, blood samples ($n = 113$) were drawn from the caudal vein of live porpoises, immediately preserved in an Acid-Citrate-Dextrose solution, and stored in liquid nitrogen. Tissue samples ($n = 40$, mainly muscle) were obtained from dead porpoises and preserved in 80% alcohol. Samples from the Yangtze mainstream and Dongting Lake were collected from accidentally killed or stranded individuals over the past decade. Fifty-eight blood samples from PY were collected from live animals during three field surveys conducted in early spring of 2009 ($n = 28$), 2010 ($n = 17$) and 2011 ($n = 13$), under a special permit from the Poyang Lake Fishery and Fishing Administration Office of Jiangxi Province. Gender was identified by visual inspection of the genital parts. In total, 76 males and 77 females were sampled. The sampling was conducted in accordance with the Regulations of the People's Republic of China for the Implementation of Wild Aquatic Animal Protection promulgated in 1993 by the Food and Agriculture Organization of the United Nations (FAO, FAOLEX No. LEX-FAOC011943, <http://www.fao.org/faolex/results/details/en/?details=LEX-FAOC011943>), adhering to all ethical guidelines and legal requirements in China.

Total genomic DNA was isolated from blood samples using the Whole Genome DNA Extraction Kit (SBS, Shanghai Inc.) following the manufacturer's instructions. For tissue samples, total genomic DNA was extracted using a standard proteinase K digestion and phenol/chloroform extraction protocol⁵³.

Microsatellite and mitochondrial data set. Microsatellite and mitochondrial (mtDNA) data have been previously obtained and described by Chen *et al.*²⁰. All 153 porpoises were genotyped at 11 polymorphic microsatellite loci (YFSSR1, YFSSR42, YFSSR59, YFSSR5, YFSSR40 from *N. p. asiaorientalis*^{54,55}, NP391, NP404, NP409, NP464, NP428 from *N. phocaenoides*^{56,57}, and PPHO130 from *Phocoena phocoena*⁵⁸). The genotyping protocol and quality checks have been described in Chen *et al.*²⁰. In the subsequent analyses of the microsatellites, we only kept individuals for which we had at least 50% of the locus available (i.e. at least 5 microsatellite loci and the mtDNA or 6 microsatellite loci). Those filters excluded of 5 individuals, leading to a final data set of 148

individuals. The mtDNA data includes a 597 base-pairs fragment of the hyper-variable region 1 of the control region successfully sequenced for 129 individuals (see Chen *et al.*²⁰ for details on the PCR and sequencing procedures). The microsatellite and mtDNA-CR data are available in Supplementary Dataset S1.

Population genetic structure. We used the Bayesian model-based clustering of *STRUCTURE* v2.3.4²⁶ to estimate the admixture proportions for each individual to each cluster identified in the microsatellite data. *STRUCTURE* cluster individual multilocus genotypes into K groups while minimizing departures from Hardy-Weinberg and Linkage Equilibria. We used the admixture *Locprior* model with correlated allele frequencies designed to detect weak signals of genetic structure without introducing bias or forcing the clustering²⁶. We conducted a series of independent runs with different value for K from 1 to 5. Each run used 1×10^6 iterations after a burn-in of length 2×10^5 . To assess that convergence of the Monte Carlo Markov Chains (MCMCs) had been reached, we performed 10 independent replicates for each K and checked the consistency of results using *CLUMPAK*⁵⁹. We assessed which K value best fit with the data using (1) the likelihood of each K , following *STRUCTURE*'s user manual; (2) its rate of change with increasing K ⁶⁰; and (3) visual inspection of newly created clusters with increasing K ⁶¹. Post-processing of the results, including generation of barplots, was conducted using *CLUMPAK*⁵⁹. The geographic distribution of each group (Fig. 2) was mapped using the R statistical package *MARMAP* v.0.9.5⁶² and ETOPO dataset⁶³.

The genetic structure in the microsatellite data was further inspected using a Discriminant Analysis of Principal Components (DAPC)²⁸ and a Principal Component Analysis (PCA)^{27,29}. These exploratory methods does not rely on any model assumptions and provides a complementary validation of the structure depicted by *STRUCTURE*⁶⁴. These analysis were conducted using *adegenet* 2.0.1 package⁶⁵ for R⁶⁶ on centred genetic data (i.e. set to a mean allele frequency of zero), with missing data replaced by the mean, following the authors' recommendations. The PCA was used to display the individual multilocus genotypes into a reduced multidimensional space defined by the first two principal components (PCs), colour-coding each individual according to the clustering identified by *STRUCTURE*, in order to assess the congruence. The DAPC method identifies genetic clusters by optimizing the difference between predefined groups and minimizing the variation within those groups²⁸. It first reduces the number of variables using a PCA and then maximises the differences between groups using a Discriminant Analysis. The DAPC was performed with prior information on groups using the clusters defined by *STRUCTURE*. The number of PCs to retain and the reliability of the DAPC were determined using the cross-validation approach present in the *adegenet* 2.0.1 package⁶⁵ for R⁶⁶. As a result of this cross-validation step, a total of 60 PCs and 4 discriminant functions were retained to describe the relationship between the clusters. This number of PCs captured 80% of the total variation and provided the highest percent of correctly predicted subsamples with the lowest error. Finally, the score of each individual for the first two discriminant functions (DFs) were plotted as scatter plot in R and the memberships probability of each individual to the clusters defined by the DAPC were plotted as barplot in R.

Genetic diversity and differentiation. We compared microsatellite genetic diversity between populations using the allelic richness (A_r) and private A_r (pA_r), computed with *ADZE*⁶⁷, and the observed and expected heterozygosity (H_o and H_e) computed with *GenAlEx* v6.5⁶⁸. Departures from Hardy-Weinberg were tested using 10^4 permutations with *FSTAT* v2.9.3.2⁶⁹ and quantified using F_{IS} and F_{ST} ⁷⁰ in *GenAlEx* v6.5⁶⁸. We applied a Bonferonni correction to correct for multiple tests.

Mitochondrial genetic diversity was estimated for each of the genetically distinct groups identified in the present study. Variation among sequences was measured using the number of segregating sites (S), number of singletons and shared polymorphisms, number of haplotypes, haplotype diversity (H_d) and two estimators of population genetic diversity, π based on the average number of pairwise differences⁷¹ and θ_w based on the number of polymorphic sites⁷². All statistics were calculated using *DNASP* v5.10.01⁷³. We used the F_{ST} statistics estimated from the average number of differences within and between populations⁷⁴. Significance was tested with 1,000 permutations of Hudson's nearest neighbour distance *Snn* statistics, which measures how often the nearest neighbour of a sequence (in sequence space) is from the same population⁷⁵.

Contemporary effective population sizes. We used *NeEstimator* v2.01³⁰ as a first approach to estimate contemporary N_e based on linkage disequilibrium (LD) between loci, filtering out rare alleles with a frequency $P_{crit} \leq 0.02$ that could bias N_e estimate⁷⁶. The second method implemented in *ONESAMP* v1.2³¹ also use LD as summary statistics among others in an Approximate Bayesian Computation (ABC) to estimate N_e , considering uniform prior between 2 and 500 of N_e .

Contemporary gene flow between populations. We estimated contemporary effective migration rate (m) between populations using *BayesAss* v3.0.3³². Preliminary runs were performed to adjust the mixing parameters of the MCMC and ensure proposal acceptance rates between 20% and 60% following authors' recommendations. We then performed 12 independent runs with different seeds, a burn-in of 5×10^6 iterations followed by 2×10^8 iterations, and a sampling parameter values every 2000 iterations. Convergence of the MCMCs was checked by comparing the traces of each run using *Tracer* v1.5⁷⁷ and by evaluating the Effective Sample Sizes (ESSs) of each parameter, keeping only runs where $ESS \geq 200$. Model fitting to the data was assessed using the Bayesian Deviance Index using the R-script of Meirmans³³. Runs that converged were combined to estimate the mean, median and 95% Highest Probability Density interval for each parameter in *Tracer* v1.5. The effective number migrants ($N_e \times m$) per generation between populations was obtained by combining N_e and m estimates.

Genetic inference of population demographic history. We used *DIYABC* v2.1⁷⁸ to estimate the M_{GW} value and conduct 1×10^6 coalescent simulations to produce a null distribution against which the observed value could be compared. The P -value indicates the proportion of simulations which provide a value below the observed one. For the mtDNA-CR, we used the Tajima's D^{35} and tested for significant departure from a null expectations using 10,000 coalescent simulations in DNAsp⁷³.

Next, we investigated the demographic history best describing the genetic diversity of the combined microsatellite and mtDNA markers using a coalescent-based ABC approach²². We subdivided our workflow in two nested parts (Fig. 4): identify the most likely population tree topology for our dataset among 10 plausible scenarios describing different potential population branching (Fig. 4a); and then test for evidence of changes in effective population size in the ancestral and daughter populations (Fig. 4b). This later part was further subdivided in two steps, (i) first testing for simple changes in effective population size (N_e) with six nested competing scenarios, and (ii) then testing whether adding the known population decline observed in each population over the last 50 yrs in the best scenario improved significantly the model fit to the data (Fig. 4b, Tables S2 and S3).

For each part (Fig. 4), an ABC analysis was conducted using the program *DIYABC* v2.1.0⁷⁸ applying the following steps (Fig. S4): (1) coalescent simulations of 1×10^6 pseudo-observed datasets (PODs) for each competing scenario and calculation of summary statistics (SS) describing the observed genetic variation for each POD; (2) select the best model by estimating the posterior probability (PP) of each scenario using two approaches: the "standard" procedure relying on a logistic regression on 1% PODs producing SS values closest to the observed ones after a Linear Discriminant Analysis (ABC-LDA) as a pre-processing step³⁶ and the recently introduced Random Forest (ABC-RF) procedure^{37,38}; (3) evaluate the confidence in scenario choice by estimating the type-I and type-II error rates based on simulated PODs using the ABC-LDA, as well as the prior error rate from the of the ABC-LDA and ABC-RF^{37,38}; (4) estimate the marginal posterior distribution of each parameter based on the best model including (among other) N_e and times of population size changes and splits (T); and finally, (5) evaluate the goodness-of-fit of the fitted model to the data. Details are provided in Fig. 4 and in the supplementary materials (Appendix S1, Fig. S4, Tables S2 and S3).

Data accessibility. All data generated during this study are included in this article (and its Supplementary Information files).

References

- Marko, P. B. & Hart, M. W. The complex analytical landscape of gene flow inference. *Trends Ecol. Evol.* **26**, 448–456 (2011).
- Epps, C. W. & Keyghobadi, N. Landscape genetics in a changing world: disentangling historical and contemporary influences and inferring change. *Mol. Ecol.* **24**, 6021–6040 (2015).
- Fahrig, L. Effects of Habitat Fragmentation on Biodiversity. *Annu. Rev. Ecol. Syst.* **34**, 487–515 (2003).
- Hale, M. L. *et al.* Impact of landscape management on the genetic structure of red squirrel populations. *Science* **293**, 2246–2248 (2001).
- Mech, S. G. & Hallett, J. G. Evaluating the effectiveness of corridors: a genetic approach. *Conserv. Biol.* **5**, 467–474 (2001).
- Carroll, C., McRae, B. H. & Brookes, A. Use of linkage mapping and centrality analysis across habitat gradients to conserve connectivity of gray wolf populations in western North America. *Conserv. Biol.* **26**, 78–87 (2012).
- Turvey, S. T. *et al.* First human-caused extinction of a cetacean species? *Biol. Lett.* **3**, 537–540 (2007).
- Gao, A. & Zhou, K. Geographical variation of external measurements and three subspecies of *Neophocaena phocaenoides* in Chinese waters. *Acta Theriol. Sin.* **15**, 81–92 (1995).
- Wang, D., Turvey, S. T., Zhao, X. & Mei, Z. *Neophocaena asiaorientalis ssp.* The IUCN Red List of Threatened Species e.T43205774A45893487 (accessed on 2017-10-02), <https://doi.org/10.2305/IUCN.UK.2013-1.RLTS.T43205774A45893487.en> (2013).
- Zhang, X. *et al.* The population of finless porpoise in the middle and lower reaches of Yangtze river. *Acta Theriol. Sin.* **13**, 260–270 (1993).
- Yang, J., Xiao, W., Kuang, X., Wei, Z. & Liu, R. Studies on the distribution, population size and the active regularity of *Lipotes vexillifer* and *Neophocaena phocaenoides* in Dongting Lake and Boyang Lake. *Resour. Env. Yangtze Valley* **9**, 444–450 (2000).
- Xiao, W. & Zhang, X. Distribution and population size of Yangtze finless porpoise in Poyang Lake and its branches. *Acta Theriol. Sin.* **22**, 7–14 (2002).
- Zhao, X. *et al.* Abundance and conservation status of the Yangtze finless porpoise in the Yangtze River, China. *Biol. Conserv.* **141**, 3006–3018 (2008).
- Mei, Z. *et al.* The Yangtze finless porpoise: On an accelerating path to extinction? *Biol. Conserv.* **172**, 117–123 (2014).
- Mei, Z. *et al.* Accelerating population decline of Yangtze finless porpoise (*Neophocaena asiaorientalis asiaorientalis*). *Biol. Conserv.* **153**, 192–200 (2012).
- Fontaine, M. C. *et al.* History of expansion and anthropogenic collapse in a top marine predator of the Black Sea estimated from genetic data. *P. Natl. Acad. Sci. USA* **109**, E2569–76 (2012).
- Fontaine, M. C. *et al.* Postglacial climate changes and rise of three ecotypes of harbour porpoises, *Phocoena phocoena*, in western Palearctic waters. *Mol. Ecol.* **23**, 3306–3321 (2014).
- Yang, G., Guo, L., Bruford, M. W., Wei, F. & Zhou, K. Mitochondrial phylogeography and population history of finless porpoises in Sino-Japanese waters. *Biol. J. Linn. Soc.* **95**, 193–204 (2008).
- Lin, W. *et al.* Phylogeography of the finless porpoise (genus *Neophocaena*): testing the stepwise divergence hypothesis in the northwestern Pacific. *Sci. Rep.* **4**, 6572 (2014).
- Chen, M. *et al.* Genetic Diversity and Population Structure of the Critically Endangered Yangtze Finless Porpoise (*Neophocaena asiaorientalis asiaorientalis*) as Revealed by Mitochondrial and Microsatellite DNA. *Int. J. Mol. Sci.* **15**, 11307–11323 (2014).
- Beaumont, M. A. Approximate Bayesian Computation in Evolution and Ecology. *Annu. Rev. Ecol. Syst.* **41**, 379–406 (2010).
- Csilléry, K., Blum, M. G. B., Gaggiotti, O. E. & Francois, O. Approximate Bayesian Computation (ABC) in practice. *Trends Ecol. Evol.* **25**, 410–418 (2010).
- Cornuet, J.-M., Ravignin, V. & Estoup, A. Inference on population history and model checking using DNA sequence and microsatellite data with the software *DIYABC* (v1.0). *BMC Bioinform.* **11**, 401 (2010).
- Pritchard, J. K., Stephens, M. & Donnelly, P. Inference of population structure using multilocus genotype data. *Genetics* **155**, 945–959 (2000).
- Falush, D., Stephens, M. & Pritchard, J. K. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* **164**, 1567–1587 (2003).

26. Hubisz, M. J., Falush, D., Stephens, M. & Pritchard, J. K. Inferring weak population structure with the assistance of sample group information. *Mol. Ecol. Resour.* **9**, 1322–1332 (2009).
27. Jombart, T., Pontier, D. & Dufour, A.-B. Genetic markers in the playground of multivariate analysis. *Heredity* **102**, 330–341 (2009).
28. Jombart, T., Devillard, S. & Balloux, F. Discriminant analysis of principal components: a new method for the analysis of genetically structured populations. *BMC Genet.* **11**, 94 (2010).
29. Patterson, N., Price, A. L. & Reich, D. Population Structure and Eigenanalysis. *PLoS Genet.* **2**, e190 (2006).
30. Do, C. *et al.* NeEstimatorv2: re-implementation of software for the estimation of contemporary effective population size (N_e) from genetic data. *Mol. Ecol. Resour.* **14**, 209–214 (2014).
31. Tallmon, D. A., Koyuk, A., Luikart, G. & Beaumont, M. A. ONESAMP: a program to estimate effective population size using approximate Bayesian computation. *Mol. Ecol. Resour.* **8**, 299–301 (2008).
32. Wilson, G. A. & Rannala, B. Bayesian inference of recent migration rates using multilocus genotypes. *Genetics* **163**, 1177–1191 (2003).
33. Meirmans, P. G. Nonconvergence in Bayesian estimation of migration rates. *Mol. Ecol. Resour.* **14**, 726–733 (2014).
34. Garza, J. C. & Williamson, E. G. Detection of reduction in population size using data from microsatellite loci. *Mol. Ecol.* **10**, 305–318 (2001).
35. Tajima, F. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* **123**, 585–595 (1989).
36. Estoup, A. *et al.* Estimation of demo-genetic model probabilities with Approximate Bayesian Computation using linear discriminant analysis on summary statistics. *Mol. Ecol. Resour.* **12**, 846–855 (2012).
37. Pudlo, P. *et al.* Reliable ABC model choice via random forests. *Bioinformatics* **32**, 859–866 (2016).
38. Fraimout, A. *et al.* Deciphering the routes of invasion of *Drosophila suzukii* by means of ABC random forest. *Mol. Biol. Evol.* **34**, 980–996 (2017).
39. Wu, L., Li, F., Zhu, C., Li, L. & Li, B. Holocene environmental change and archaeology, Yangtze River Valley, China: Review and prospects. *Geosci. Front.* **3**, 875–892 (2012).
40. Zhang, X. Y., Cai, S. M. & Sun, S. C. Evolution of Dongting Lake since Holocene. *J. Lake Sci.* **60**, 13–21 (1994).
41. Fang, J.-Q. Lake evolution during the last 3000 years in China and its implications for environmental change. *Quat. Res.* **39**, 175–185 (1993).
42. Dong, L. *et al.* Yangtze finless porpoises along the main channel of Poyang Lake, China: Implications for conservation. *Mar. Mamm. Sci.* **31**, 612–628 (2015).
43. Wei, Z., Wang, D., Zhang, Q., Wang, K. & Kuang, X. Population size, behavior, movement pattern and protection of Yangtze finless porpoise at balijiang section of the Yangtze River. *Resour. Env. Yangtze Valley* **11**, 427–432 (2002).
44. Huang, S.-L. *et al.* Saving the Yangtze finless porpoise: Time is rapidly running out. *Biol. Conserv.* **20**, 40–46 (2017).
45. Fontaine, M. C. *et al.* Genetic and historic evidence for climate-driven population fragmentation in a top cetacean predator: the harbour porpoises in European water. *Proc. R. Soc. B* **277**, 2829–2837 (2010).
46. Louis, M. *et al.* Habitat-driven population structure of bottlenose dolphins, *Tursiops truncatus*, in the North-East Atlantic. *Mol. Ecol.* **23**, 857–874 (2014).
47. Goossens, B. *et al.* Genetic Signature of Anthropogenic Population Collapse in Orang-utans. *PLoS Biol.* **4**, e25 (2006).
48. Weckworth, B. V. *et al.* Preferred habitat and effective population size drive landscape genetic patterns in an endangered species. *Proc. R. Soc. B* **280**, 20131756 (2013).
49. Waples, R. S. Genetic estimates of contemporary effective population size: to what time periods do the estimates apply? *Mol. Ecol.* **14**, 3335–3352 (2005).
50. Palstra, F. P. & Fraser, D. J. Effective/census population size ratio estimation: a compendium and appraisal. *Ecol. Evol.* **2**, 2357–2365 (2012).
51. Hare, M. P. *et al.* Understanding and Estimating Effective Population Size for Practical Application in Marine Species Management. *Conserv. Biol.* **25**, 438–449 (2011).
52. Waples, R. S. & England, P. R. Estimating Contemporary Effective Population Size on the Basis of Linkage Disequilibrium in the Face of Migration. *Genetics* **189**, 633–644 (2011).
53. Sambrook, J. & Russell, D. W. *Molecular Cloning: A Laboratory Manual*. (CSHL Press, 2012).
54. Zheng, J. S. *et al.* Development and characterization of polymorphic microsatellite loci in the endangered Yangtze finless porpoise (*Neophocaena phocaenoides asiaorientalis*). *Conserv. Genet.* **9**, 1007–1009 (2008).
55. Zhou, Z., Zheng, J. S., Chen, M. M., Zhao, Q. Z. & Wang, D. Genetic evaluation and development prognosis on *ex situ* conserved Yangtze finless porpoise living in Tian-E-Zhou National Natural Reserve. *Acta Theriol. Sin.* **36**, 403–411 (2012).
56. Chen, L., Bruford, M. & Yang, G. Isolation and characterization of microsatellite loci in the finless porpoise (*Neophocaena phocaenoides*). *Mol. Ecol. Notes* **7**, 1129–1131 (2007).
57. Chen, L. & Yang, G. Development of tetranucleotide microsatellite loci for the finless porpoise (*Neophocaena phocaenoides*). *Conserv. Genet.* **9**, 1033–1035 (2008).
58. Rosel, P. E., France, S. C., Wang, J. Y. & Köcher, T. D. Genetic structure of harbour porpoise *Phocoena phocoena* populations in the northwest Atlantic based on mitochondrial and nuclear markers. *Mol. Ecol.* **8**, S41–S4 (1999).
59. Kopelman, N. M., Mayzel, J., Jakobsson, M., Rosenberg, N. A. & Mayrose, I. Clumpak: a program for identifying clustering modes and packaging population structure inferences across K. *Mol. Ecol. Resour.* **15**, 1179–1191 (2015).
60. Evanno, G., Regnaut, S. & Goudet, J. Detecting the number of clusters of individuals using the software structure: a simulation study. *Mol. Ecol.* **14**, 2611–2620 (2005).
61. Vercken, E. *et al.* Glacial refugia in pathogens: European genetic structure of another smut pathogens on *Silene latifolia* and *Silene dioica*. *PLoS Pathog.* **6**, e1001229 (2010).
62. Pante, E. & Simon-Bouhet, B. marmap: A package for importing, plotting and analyzing bathymetric and topographic data in R. *PLoS One* **8**, e73051 (2013).
63. Amante, C. & Eatkins, B. W. ETOPO1 1 Arc-Minute Global Relief Model: Procedures, Data Sources and Analysis. NOAA Technical Memorandum NESDIS NGDC-24. National Geophysical Data Center, NOAA, <https://doi.org/10.7289/V5C8276M> [accessed 2017-10-02] (2009).
64. Francois, O. & Durand, E. Spatially explicit Bayesian clustering models in population genetics. *Mol. Ecol. Resour.* **10**, 773–784 (2010).
65. Jombart, T. adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics* **24**, 1403–1405 (2008).
66. R Core Team. R: A Language and Environment for Statistical Computing. *R Foundation for Statistical Computing* (2016).
67. Szpiech, Z. A., Jakobsson, M. & Rosenberg, N. A. ADZE: a rarefaction approach for counting alleles private to combinations of populations. *Bioinformatics* **24**, 2498–2504 (2008).
68. Peakall, R. & Smouse, P. E. GenALEX 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. *Bioinformatics* **28**, 2537–2539 (2012).
69. Goudet, J. FSTAT, A program to estimate and test gene diversities and fixation indices (version 2.9.3). Available at <http://www2.unil.ch/popgen/softwares/fstat.htm> [accessed 2017-10-02] (2001).
70. Weir, B. S. & Cockerham, C. C. Estimating F-statistics for the analysis of population structure. *Evolution* **1358–1370** (1984).
71. Tajima, F. Evolutionary relationship of DNA sequences in finite populations. *Genetics* **105**, 437–460 (1983).
72. Watterson, G. A. On the number of segregating sites in genetical models without recombination. *Theor. Popul. Biol.* **7**, 256–276 (1975).

73. Librado, P. & Rozas, J. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* **25**, 1451–1452 (2009).
74. Hudson, R. R., Boos, D. D. & Kaplan, N. L. A statistical test for detecting geographic subdivision. *Mol. Biol. Evol.* **9**, 138–151 (1992).
75. Hudson, R. R. A new statistic for detecting genetic differentiation. *Genetics* **155**, 2011–2014 (2000).
76. Waples, R. S. & Do, C. Linkage disequilibrium estimates of contemporary N_e using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. *Evol. Appl.* **3**, 244–262 (2010).
77. Rambaut, A. Tracersv1.5 Available from <http://tree.bio.ed.ac.uk/software/tracer/> [accessed 2017-07-05] (2007).
78. Cornuet, J.-M. *et al.* DIYABCv2.0: a software to make approximate Bayesian computation inferences about population history using single nucleotide polymorphism, DNA sequence and microsatellite data. *Bioinformatics* **30**, 1187–1189 (2014).

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Author Contributions

M.C.F., J.Z., D.W. designed the study; M.C., J.Z., Z.M., Y.H., K.W., M.W., Q.Z., D.W. conducted the field expeditions and collected the samples; M.C. conducted the laboratory experiments and collected the data; M.C.F., Y.B.C. and F.L. analysed the data; M.C.F. interpreted the results and wrote the manuscript with help from Y.B.C., M.C., J.Z. and D.W., and final approval by all co-authors.

Additional Information

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