Wiley Journal of Diabetes Research Volume 2025, Article ID 1273073, 8 pages https://doi.org/10.1155/jdr/1273073

Research Article

Association Between Glycemic Control and Complications With Concentration of Urinary Exfoliated Proximal Tubule Kidney Cells in People With Diabetes Mellitus

Henry H. L. Wu, ^{1,2} Venkatesha Bhagavath, ³ Long The Nguyen, ¹ Rajkumar Chinnadurai, ⁴ Ewa M. Goldys, ² Carol A. Pollock, ^{1,5} and Sonia Saad, ¹

Correspondence should be addressed to Henry H. L. Wu; hon.wu@sydney.edu.au

Received 20 June 2024; Accepted 6 January 2025

Academic Editor: Huantian Cui

Copyright © 2025 Henry H. L. Wu et al. Journal of Diabetes Research published by John Wiley & Sons Ltd. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Background: Emerging evidence suggests cell exfoliation could be operating under the control of cell metabolism. It is unclear if there are associations between the concentration of exfoliated kidney proximal tubule cells (PTCs) in urine with glycemic control and complications. Our study is aimed at exploring this.

Methods: Urine samples were collected from 122 adult study participants and stored at -80°C. Exfoliated PTCs were extracted from thawed urine using a validated specific immunomagnetic separation method based on anti-CD13 and anti-SGLT-2 antibodies. The number of PTCs was assessed using brightfield microscopy. Study participants were grouped into those with no diabetes mellitus (DM) and those with DM. Individuals with DM were further subgrouped into those with and without retinopathy. Adjusted Poisson regression analysis was conducted for the DM cohort, investigating associations between demographic, clinical, and biochemical parameters with mean urinary exfoliated PTCs.

Results: The adjusted Poisson regression analysis noted sex to have a significant association with mean number of urinary exfoliated PTCs, with a lower incidence rate in males compared to females (incidence rate ratio (IRR) 0.56, 95% CI 0.35–0.89, p = 0.014). Each 1% increase in glycated haemoglobin (HbA1c) was associated with an increase of 1.03 times in mean exfoliated PTCs (IRR 1.03, 95% CI 1.01–1.04, p = 0.007), and DM patients with retinopathy had an increase of 1.68 times in mean exfoliated PTCs compared to those without retinopathy (IRR 1.68, 95% CI 1.07–2.62, p = 0.024). No significant associations were observed with albuminuria or estimated glomerular filtration rate (eGFR).

Conclusions: Our results indicate increased shedding of PTCs into the urinary tract in patients with poorer glycemic control, particularly those with diabetic retinopathy and in females.

Keywords: cell exfoliation; diabetes mellitus; diabetic kidney disease; proximal tubule cells; retinopathy; urine

1. Introduction

Cell exfoliation is an active biochemical process that has been linked to the homeostasis of mammalian organs. Exfo-

liation occurs when extracellular matrix components that usually tightly connect between cells within a structure break off, with external cells being removed from the epithelial luminal surface [1]. Activity of cell exfoliation is thought to

¹Renal Research Laboratory, Kolling Institute of Medical Research, Royal North Shore Hospital & The University of Sydney, Sydney, Australia

²ARC Centre of Excellence for Nanoscale Biophotonics, School of Biomedical Engineering, The University of New South Wales, Sydney, Australia

³Biostatistics Support and Consultation Services, Northern Sydney Local Health District, Sydney, Australia

⁴Faculty of Biology, Medicine and Health, The University of Manchester, Manchester, UK

 $^{^5}$ Department of Renal Medicine, Royal North Shore Hospital, Northern Sydney Local Health District, Sydney, Australia

be under the control of cell metabolism and is now being deemed a potentially useful indicator of metabolic status [2]. In particular, kidney cell exfoliation into urine has been previously found to have significant links with diabetes mellitus (DM), reflecting metabolic dysregulation [3, 4]. Proximal tubule cell (PTC) exfoliation could be especially relevant in DM given glucose entry into PTCs is insulindependent, making PTCs especially sensitive to hyperglycemia in hyperglycemic conditions [5]. Numerous studies have demonstrated increased shedding of podocytes and extracellular vesicles into urine with poor metabolic control and glomerulosclerosis development in DM [6-8]. However, there are no studies to date that evaluated associations between the number of exfoliated PTCs and glycemic control, alongside related macro and microvascular complications of DM such as cardiovascular disease, diabetic kidney disease (DKD), and in particular retinopathy, which is the most common microvascular complication of DM and typically one of the early complications to manifest in patients with advanced diabetic disease [9]. Our study is aimed at exploring this.

2. Methods

2.1. Study Participant Recruitment and Ethical Considerations. The inclusion criteria for study participant recruitment are adult individuals of either sex aged between 18 and 75 years of age under the care of Royal North Shore Hospital and/or North Shore Private Hospital, Sydney, Australia, with and without Type 1 or 2 DM, and with serum samples obtained within 72h before urine sample collection. Individuals aged <18 years or >75 years were excluded from the study. As per the current Australian Diabetes Society guidelines and for the purposes of this study in grouping study participants into "cases" (i.e., people with DM) and "controls" (i.e., people without DM), DM status is defined by individuals presenting with serum glycated haemoglobin (HbA1c) \geq 6.5%, fasting glucose $\geq 7.0 \text{ mmol/L}$, random glucose $\geq 11.1 \text{ mmol/L}$, and those on a 75-g oral glucose tolerance test displaying fasting glucose $\geq 7.0 \text{ mmol/L}$ or $2 - h \text{ glucose} \geq 11.1 \text{ mmol/L}$ [10]. Study recruitment was conducted prospectively in which participants were only recruited upon their presentation to the departmental clinic, with no prior knowledge nor planning on recruitment of specific individuals. Informed consent was obtained from all study participants. Data collection in this study was carried out in accordance with relevant local guidelines and regulations, and the collection of human data was approved by the human ethics committee at Royal North Shore Hospital and North Shore Private Hospital (Ref: HREC/17/HAWKE/471) and the University of New South Wales, Sydney, Australia (Ref: HC180710).

A total of 122 study participants were recruited. There were 38 individuals with no DM and 84 individuals with DM, of which there were 26 individuals with DM and retinopathy and 58 individuals with DM but without retinopathy.

2.2. Extraction and Quantification of Urinary Exfoliated Proximal Tubule Kidney Cells. Using urine bottles with a capacity of up to 100 mL, spot urine samples were collected

from adult individuals who fulfilled the above criteria and the volume of urine collected (milliliters) was recorded by H.H.L.W. for each study participant. Each collected urine sample was placed on ice immediately after collection for transportation to the Renal Research Laboratory, Kolling Institute of Medical Research, and was centrifuged for 20 min at 4°C to collect exfoliated cells and then washed twice with phosphate-buffered saline (PBS). Prior to extraction of PTCs, the initially processed samples were to be stored at -80°C with PBS and 10% dimethyl sulfoxide (10% DMSO) [11]. Whilst urine may contain a wide range of different cells-podocytes, proximal and distal tubular cells, stem/progenitor cells, extracellular vesicles, epithelial cells, bladder, and collecting duct—it has been previously established that CD13 and sodium-glucose linked transporter-2 (SGLT-2) antibodies are uniquely expressed in PTCs [11]. Specific PTCs expressing CD13, SGLT-2, and angiotensinogen were extracted via a validated specific immunomagnetic separation method based on anti-CD13 and anti-SGLT2 antibodies that our group has previously described (Figure 1) [11] and then quantified in brightfield microscopy.

2.3. Collecting and Summarizing Demographic, Clinical, and Biochemistry Data. Each study participant's demographic information including their age (years), sex and ethnicity, and clinical data encompassing medical history and diagnoses such as the presence of cardiovascular disease and retinopathy was collected from the New South Wales Health Powerchart electronic database. Height (meters) and weight (kilograms) were measured and recorded on the day of urine sample collection, and data in relation to body mass index (BMI) was automatically generated. Biochemistry data recorded for the purposes of this study included HbA1c (%), estimated glomerular filtration rate (eGFR) (mL/min/ 1.73 m²) by the Chronic Kidney Disease Epidemiology Collaboration Equation [12], urinary albumin-to-creatinine ratio (uACR), and serum creatinine (µmol/L). HbA1c, eGFR, and serum creatinine data were obtained from serum tests collected within 72 h of urine sample collection. uACR data was obtained from another urine sample collected on the same day which was sent to New South Wales Health Pathology for analysis. Systolic and diastolic blood pressure (BP) were recorded on the same day using an automated Omron Hem 7142T1 BP Monitor with different cuff sizes available depending on an individual's suitability. The estimated creatinine clearance (milliliters per minute) for each study participant was generated from MDCalc through inputting relevant information into the Cockcroft-Gault equation [13], as it informs us specifically on tubular creatinine clearance [14].

Patient demographics alongside clinical and biochemical data were summarized using appropriate descriptive statistics. Given variables are nonnormally distributed or ordinal, the median and interquartile range (IQR) were used to summarize continuous variables. Proportions with percentages were presented for categorical variables. When comparing between the no DM versus DM groups (i.e., two comparator groups), continuous variables were compared using the

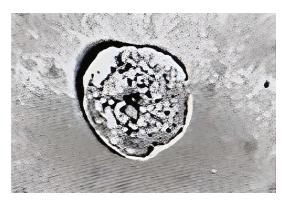


FIGURE 1: Example of a urinary exfoliated proximal tubule kidney cell specifically extracted via a validated specific immunomagnetic separation method based on anti-CD13 and anti-SGLT2 antibodies.

Mann–Whitney U test, and categorical variables were compared using the chi-square test or Fisher's exact test accounting for sparsely distributed data. When comparing between the DM with retinopathy versus the DM without retinopathy versus the no DM group (i.e., three comparator groups), continuous variables were compared using the Kruskal–Wallis H test, and categorical variables were compared using the chi-square test or Fisher's exact test accounting for sparsely distributed data.

2.4. Conducting Poisson Regression Analysis. A Poisson regression analysis model was used to determine the significance of the association between diabetic status, other study variables, and the number of urinary exfoliated PTCs. A Poisson regression is a quantitative research methodology specifically used to analyse count data as the dependent variable [15]. The Poisson regression model is the simplest count regression model [15]. Coefficients are exponentiated, since counts must be 0 or greater. A Poisson regression assumes a Poisson distribution, which is often characterized by a substantial positive skew with most cases falling at the low end of the dependent variable's distribution, and a variance that equals the mean. Because count data distributions (i.e., number of urinary exfoliated PTCs) will have a Poisson distribution, a Poisson regression tends to fit these data better than linear regression does (which assumes a normal distribution) [15]. As a result, predictive relationships with a dependent variable (i.e., number of urinary exfoliated PTCs) can be examined as in ordinary linear regression, but without the problems from having the nonnormal distributions that are expected with count data [15].

Adjusted Poisson regression analysis was conducted in this study. The model investigated associations between each included variable of the study and the number of urinary exfoliated PTCs for the DM cohort, adjusting for other covariates within the statistical model. The incidence rate ratio (IRR) along with their corresponding 95% CI and *p* value were calculated to determine the significance of association for each variable and mean urinary exfoliated PTCs. The model incorporated HbA1c, eGFR, BMI, age, sex, ethnicity, urine volume, uACR, serum creatinine, estimated cre-

atinine clearance, systolic and diastolic BP, history of cardiovascular event(s), and incidence of retinopathy as covariates. Statistical analysis was performed with p < 0.05 being the level of significance. Analysis was performed in SPSS (Version 26).

3. Results

In our cohort of 122 study participants, the diabetic group was significantly older compared to the nondiabetic group (63.8 vs. 51.5 years, p < 0.001). Median BMI was higher in the diabetic compared to the nondiabetic group (25.6 vs. 23.2, p = 0.005). Median HbA1c was higher in the diabetic compared to the nondiabetic group (7.4 vs. 6.1%, p = 0.016). Considering all study participants, the range between the highest and lowest recorded HbA1c is 8.6%. Median serum creatinine levels were higher in the diabetic compared to the nondiabetic group (115.5 vs. 96.0 μ mol/L, p = 0.028), reflecting a lower median eGFR in the diabetic compared to the nondiabetic group (52 vs. 66 mL/min/1.73 m², p = 0.042). The proportion of patients with macroalbuminuria was higher in the diabetic compared to the nondiabetic group (36.8 vs. 17.9%, p = 0.044). The proportion of patients with a history of cardiovascular event(s) was more prevalent in the diabetic compared to the nondiabetic group (53.6 vs. 18.4%, p < 0.001). Diabetic retinopathy was observed in 32% of individuals with DM. Median urinary exfoliated PTCs were higher in the diabetic compared to the nondiabetic group (6 vs. 5, p = 0.021). A summary of demographic, clinical, and biochemistry parameters for the diabetic and nondiabetic groups is presented in Table 1.

Subdividing the study participants within the diabetic group into those with DM and retinopathy and those with DM but without retinopathy, whilst also considering those in the nondiabetic group, individuals with DM and retinopathy were older compared to diabetic individuals without retinopathy and individuals without DM (67.5 vs. 66.5 vs. 50.5 years, p < 0.001). BMI was higher in individuals with DM and retinopathy compared to individuals with DM but without retinopathy and individuals without DM (27.1 vs. 25.5 vs. 23.2, p = 0.017). Median HbA1c was higher in the group with DM and retinopathy compared to those with DM but without retinopathy and those without DM (7.6 vs. 7.3 vs. 6.1%, p < 0.001). The proportion of patients with a history of a cardiovascular event(s) was more prevalent in the group with DM and retinopathy compared to those without retinopathy (73.1 vs. 44.8 vs. 18.4%, p < 0.001). Median urinary exfoliated PTCs were higher in those with DM and retinopathy compared to those with DM but without retinopathy and those without DM (8 vs. 6 vs 5, p = 0.002). A summary of demographic, clinical, and biochemistry parameters for study participants with DM and retinopathy, study participants with DM but without retinopathy, and the nondiabetic group is presented in

In order to evaluate associations between a number of urinary exfoliated PTCs, glycemic control, and other covariates, a Poisson regression analysis was conducted for the 84 study participants diagnosed with DM (Table 3). Sex

Table 1: Demographic, clinical, and biochemistry characteristics of the study cohort based on diabetes mellitus status (n = 122).

Variable	Total $(n = 122)$	No DM $(n = 38)$	DM $(n = 84)$	p value ^a
Age (years), mean (SD)	60.0 (14.3)	51.5 (16.2)	63.8 (11.6)	< 0.001
Sex, n (%)				
Female	36 (29.5)	20 (52.6)	16 (19.1)	< 0.001
Male	86 (70.5)	18 (47.4)	68 (81)	
Ethnicity, <i>n</i> (%)				
0 (White)	68 (55.7)	23 (60.5)	45 (53.6)	0.474
1 (non-White)	54 (44.3)	15 (39.5)	39 (46.4)	
Height (m), median (IQR)	1.69 (0.10)	1.71 (0.19)	1.69 (0.13)	0.439
Weight (kg), median (IQR)	76.2 (18.4)	68.8 (18.3)	72.4 (25.1)	0.011
BMI, median (IQR)	26.4 (5.2)	23.2 (6.1)	25.6 (8.6)	0.005
HbA1c (%), median (IQR)	7.3 (3.2)	6.1 (3.3)	7.4 (3.2)	0.016
eGFR (mL/min/1.73 m ²), median (IQR)	60.4 (32.4)	66 (50)	52 (56.5)	0.042
Urine volume (mL), median (IQR)	71 (15)	77 (48)	68 (65)	0.170
uACR, median (IQR)	23.1 (65.6)	12.8 (90)	80.6 (158.7)	0.068
Serum creatinine (μ mol/L), median (IQR)	107.5 (89)	96 (60)	115.5 (106)	0.028
Urinary exfoliated proximal tubule cells, median (IQR)	6 (5)	5 (7)	6 (6)	0.021
Systolic BP (mmHg), median (IQR)	133.5 (33)	128 (26)	136.5 (34.5)	0.393
Diastolic BP (mmHg), median (IQR)	75 (15)	74.5 (21)	75 (14)	0.353
Albuminuria, n (%)				
Macro	29 (23.8)	14 (36.8)	15 (17.9)	0.044
Micro	50 (41.0)	13 (34.2)	37 (44.1)	
History of CV events, <i>n</i> (%)				
No	70 (57.4)	31 (81.6)	39 (46.4)	< 0.001
Yes	52 (42.6)	7 (18.4)	45 (53.6)	
Retinopathy, n (%)				
No	95 (77.9)	38 (100)	57 (67.9)	< 0.001
Yes	27 (22.1)	0 (0)	27 (32.1)	

Abbreviations: BMI, body mass index; BP, blood pressure; CV, cardiovascular; DM, diabetes mellitus; eGFR, estimated glomerular filtration rate; HbA1c, glycated haemoglobin; IQR, interquartile range; SD, standard deviation; uACR, urine albumin-to-creatinine ratio.

was found to display a significant association with mean number of exfoliated PTCs, with a lower incidence rate for males compared to females (IRR 0.56, 95% CI 0.35–0.89, p=0.014), and significantly lower number of exfoliated PTCs in males (mean 5.58, 95% CI 4.74–6.43) compared to females (mean 10.00, 95% CI 5.83–14.17). HbA1c displayed significant associations with the number of exfoliated PTCs, where each 1% increase in HbA1c is associated with an increase of 1.03 times of mean exfoliated PTCs (IRR 1.03, 95% CI 1.01–1.04, p=0.007). Individuals diagnosed with retinopathy exhibited increased numbers of exfoliated PTCs (IRR 1.68, 95% CI 1.07–2.62, p=0.024), but other variables such as eGFR and serum albumin:creatinine ratio did not demonstrate statistically significant associations with a number of urinary exfoliated PTCs (p>0.05).

4. Discussion

Overall, our results indicate increased shedding of PTCs into the urinary tract in patients with DM compared to those without DM, in those who are female compared to males, in patients with progressively poorer glycemic control, and in patients with DM and retinopathy compared to those without retinopathy.

The relationship between kidney cell exfoliation, glycemic control, and associated diabetic complications has not been explored explicitly in the past. In our study, we have demonstrated a significant increase in the shedding of PTCs into the urinary tract in individuals with progressively poorer glycemic control and patients with DM and retinopathy compared to those without retinopathy. As microvascular endothelial cells are specific targets of intracellular hyperglycemia-induced injury, multiorgan microvascular endothelial cell dysfunction is likely to increase with elevated HbA1c [16-19]. This process is mainly attributed to four hallmark mechanisms—namely, protein kinase C activation, activation of the hexosamine pathway, activation of the polyol pathways, and subsequently the formation of advanced glycation end (AGE) products [20, 21]. Although it is not yet proven, it is likely that worsening endothelial cell

^aContinuous variables were compared between the diabetic and nondiabetic groups using the Mann–Whitney *U* test. Categorical variables were compared using the chi-square test or Fisher's exact test accounting for sparsely distributed data.

Table 2: Demographic, clinical, and biochemistry characteristics of the study cohort in which study participants with diabetes mellitus are grouped based on retinopathy status (n = 122).

Variable	No DM (n = 38)	DM without retinopathy (n = 58)	DM with retinopathy (n = 26)	p value ^a
Age (years), median (IQR)	50.5 (29)	67.5 (14)	66.5 (24)	< 0.001
Sex, n (%)				
Female	20 (52.6)	13 (22.4)	3 (11.5)	< 0.001
Male	18 (47.4)	45 (77.6)	23 (88.5)	
Ethnicity, n (%)				
0 (White)	23 (60.5)	30 (51.7)	15 (57.7)	0.680
1 (non-White)	15 (39.5)	28 (48.3)	11 (42.3)	
Height (m), median (IQR)	1.71 (0.19)	1.69 (0.14)	1.70 (0.12)	0.719
Weight (kg), median (IQR)	68.8 (18.3)	71.4 (25.7)	75.4 (24.7)	0.025
BMI, median (IQR)	23.2 (6.1)	25.5 (9.2)	27.1 (8.5)	0.017
HbA1c (%), median (IQR)	6.1 (3.3)	7.3 (1.1)	7.6 (1.2)	< 0.001
eGFR (mL/min/1.73 m ²), median (IQR)	66 (50)	57 (60)	51.8 (48)	0.110
Urine volume (mL), median (IQR)	77 (48)	60 (41)	62 (47)	0.192
uACR, median (IQR)	12.8 (90)	8.2 (83.5)	18.7 (94.6)	0.068
Serum creatinine (μ mol/L), median (IQR)	96 (60)	111 (111)	125 (107)	0.088
Urinary exfoliated proximal tubule cells, median (IQR)	5 (7)	6 (6)	8 (5)	0.002
Systolic BP (mmHg), median (IQR)	128 (26)	132 (40)	139 (21)	0.372
Diastolic BP (mmHg), median (IQR)	74.5 (21)	73 (13)	77 (20)	0.188
Albuminuria, n (%)				
Macro	14 (36.8)	10 (17.2)	5 (19.2)	0.248
Micro	13 (34.2)	25 (43.1)	12 (46.2)	
History of CV events, n (%)				
No	31 (81.6)	32 (55.2)	7 (26.9)	< 0.001
Yes	7 (18.4)	26 (44.8)	19 (73.1)	

Abbreviations: BMI, body mass index; BP, blood pressure; CV, cardiovascular; DM, diabetes mellitus; eGFR, estimated glomerular filtration rate; HbA1c, glycated haemoglobin; IQR, interquartile range; uACR, urine albumin-to-creatinine ratio.

^aContinuous variables were compared between the three groups using the Kruskal–Wallis H test. Categorical variables were compared using the chi-square test or Fisher's exact test accounting for sparsely distributed data.

dysfunction alongside increased cellular inflammation, oxidative stress, and apoptotic processes eventually leads to local vascular structural changes, where increased activity of cell exfoliation occurs as part of dysregulated autophagy [2, 22]. Such changes at a larger, systemic scale in organs embedded with microvasculature such as the kidneys may occur as microvascular disease progresses with DM. The intercellular junctions and desmosomes which connect between different cell types within a structure may gradually loosen and break off leading to increased exfoliation of cells into the extracellular spaces [1]. This could be the pathway in which kidney cell exfoliation is likely affected by the severity of endothelial cell dysfunction with systematically worsened microvascular disease in DM [23]. This mechanism may explain for the increased number of exfoliated PTCs into urine with poorer glycemic control as observed from our analysis. It is also plausible that links between the number of exfoliated PTCs into urine and retinopathy, the most common microvascular complication of DM, are explained by this cascade of pathophysiological events. Further studies

are required to determine the causality of the associations observed in our study.

Otherwise, previous investigation, comparing the ability to grow urinary exfoliated kidney cells in culture between people with DM and its associated macro and microvascular complications versus healthy and diabetic individuals free of complications, has been conducted. A seminal study by Detrisac et al. [3] found few cultivable exfoliated cells in healthy adults and diabetic patients free of complications, but significantly greater amounts of cultivable exfoliated kidney cells in individuals with diabetic retinopathy and/or advanced DKD. Nevertheless, Detrisac et al. [3] did not quantitate the cells in that study and did not specifically assess PTCs. Considering our data displayed more exfoliated PTCs in individuals with elevated HbA1c, it is not surprising that more exfoliated cells in the advanced diabetic group grew in culture previously.

Whilst there are reasonable physiological postulations to explain for the associations between a number of urinary exfoliated PTCs, elevated HbA1c, and retinopathy, it is

Table 3: Poisson regression model evaluating the association between covariates and the number of urinary exfoliated proximal tubule cells in study participants with diabetes mellitus (n = 84).

Variable	Poisson regression model IRR (95% CI)	p value
HbA1c per 1% increase	1.03 (1.01-1.04)	0.007
eGFR per 1 mL/min/1.73 m ² increase	1.01 (0.99–1.03)	0.156
BMI per 1 unit increase	1.04 (0.99–1.09)	0.171
Age per 1 year increase	1.01 (0.99-1.03)	0.230
Sex		
Female	0^a	
Male	0.56 (0.35-0.89)	0.014
Ethnicity		
0 (White)	0^a	
1 (non-White)	1.25 (0.85-1.86)	0.261
Estimated creatinine clearance per 1 mL/min increase	0.30 (0.09-1.06)	0.061
Urine volume per 1 mL increase	1.001 (1.00-1.01)	0.354
uACR per 1 unit increase	1.001 (1.00-1.003)	0.100
Serum creatinine per 1 μ mol/L increase	1.00 (0.99–1.001)	0.827
Systolic BP per 1 mmHg increase	0.99 (0.98–1.01)	0.848
Diastolic BP per 1 mmHg increase	0.99 (0.98–1.01)	0.272
History of CV event		
No	0^{a}	
Yes	0.80 (0.54-1.18)	0.263
Retinopathy		
No	0^{a}	
Yes	1.68 (1.07-2.62)	0.024

Abbreviations: BMI, body mass index; BP, blood pressure; CV, cardiovascular; eGFR, estimated glomerular filtration rate; HbA1c, glycated haemoglobin; IRR, incidence rate ratio (i.e., ratio of mean urinary exfoliated proximal tubule cells); uACR, urine albumin-to-creatinine ratio. aReference category used in the estimation of the IRR in the Poisson regression model.

surprising that such significant associations between a number of urinary exfoliated PTCs and kidney function in the Poisson regression analysis were not observed. This is despite serum creatinine and the proportion of study participants with macroalbuminuria being significantly increased and eGFR being significantly decreased in the diabetic group compared to the nondiabetic group. PTCs make up the majority of the kidney mass, and PTC histopathology is a hallmark representation of multiple kidney diseases (including DKD) and correlates with kidney function decline [24, 25]. Previous animal and translational studies have established increased shedding of podocytes and extracellular vesicles into urine with poor glycemic control and glomerulosclerosis development in DM [6, 8, 26]. In particular, an increased number of exfoliated podocyte microparticles across the DKD spectrum is observed, with this being shown to prognosticate DKD progression [7, 8, 27]. Numerous exosomal enzymes found in exfoliated kidney cells, such as glycogen synthase kinase-3 beta (GSK-3 β) which is mostly detected in higher levels in podocytes but also in PTCs with DM and DKD severity, have been shown to display greater prognostic accuracy for DKD progression when compared with albuminuria [4, 28]. Multiple studies noted exosomal WT-1 from urine correlates with the severity of proteinuria, the extent of glomerular damage, and the rate of kidney function decline in patients with DM [29, 30]. Considering our findings, the exact mechanisms that explain for the associations between the activity of PTC exfoliation into urine and DKD progression may be more complex than previously thought, and further study is needed.

There are no published studies at present that investigated the role of sex differences in affecting the number of urinary exfoliated PTCs. Our Poisson regression analysis results demonstrated increased shedding of PTCs into the urinary tract in females compared to males. Whilst our analysis may potentially reflect the sex-associated links to urinary PTC exfoliation in both nondiabetic and diabetic individuals, a more demographically and ethnically diverse sample is required to validate this early hypothesis. Furthermore, additional studies should be conducted to address whether the number of urinary exfoliated PTCs is affected by different therapies including conventional Western medicine, as well as herbal medicines and natural products used by many patients for treatment of DM [31–34].

Reflecting on the methodology of our study in garnering these results, there are strengths of our study methodology to acknowledge. We managed to perform a validated longterm cryopreservation protocol and reliably demonstrated noninvasive isolation of assessable exfoliated PTCs from 122 human urine samples using immunomagnetic extraction with CD13, SGLT-2 antibodies, and magnetic beads applied in consistent concentrations. The reliability and reproducibility of this protocol are important steps to allow for reliable identification and quantification of urinary exfoliated PTCs in individuals with varying degrees of glycemic control to fulfill the main aim of our study. The Poisson regression model considered many relevant covariates in its analysis outside of glycemic control, including renal function parameters such as uACR and serum creatinine, and relevant comorbidities such as a history of cardiovascular event(s). Based on current understanding, each of the included demographic, clinical, and biochemistry parameters could play a role in affecting cellular metabolism and hence the activity of PTC exfoliation into urine.

There are limitations to our study approach in quantifying PTC concentration in urine. It is likely that poor glycemic control increases diuresis, and hence, the absolute increase in the number of exfoliated PTCs may well be underestimated by our approach. Adjusting for the concentration of urine volume depends on an individual's water intake, and this is challenging to objectively control for given human factors. It would be similarly difficult to determine the concentration of different cell types and molecules that may be exfoliated in urine alongside PTCs. Another consideration relates to the collection of spot urine samples versus a "timed" urine sample collected across different timepoints. Diurnal variation of metabolic activity may impact on the

degree of PTC exfoliation which may influence results. This is challenging to control in practice considering the multitude of human and practical factors which can affect the reliability of urine samples collected over different timepoints. These limitations should be subject to further investigation going forward.

In summary, our findings provide a novel observation that suggests the presence of significant associations between the number of exfoliated PTCs into urine with progressively poorer glycemic control and diabetic retinopathy. Whilst a convincing association with DKD is yet to be demonstrated, additional research is required to explore the pathophysiological mechanisms that may underpin an intricate relationship that exists between the activity of PTC exfoliation into urine and metabolic changes that occur in diabetes.

Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Ethics Statement

The collection of human data was approved by the human ethics committee at Royal North Shore Hospital and North Shore Private Hospital (Ref: HREC/17/HAWKE/471) and the University of New South Wales, Sydney, Australia (Ref: HC180710).

Conflicts of Interest

The authors declare no conflicts of interest.

Author Contributions

Conceptualization: Henry H. L. Wu and Sonia Saad; methodology: Henry H. L. Wu, Long The Nguyen, and Sonia Saad; formal analysis and investigation: Henry H. L. Wu, Venkatesha Bhagavath, and Rajkumar Chinnadurai; writing—original draft and preparation: Henry H. L. Wu; writing—review and editing: Long The Nguyen, Rajkumar Chinnadurai, Ewa M. Goldys, Carol A. Pollock, and Sonia Saad; funding acquisition: Ewa M. Goldys, Carol A. Pollock, and Sonia Saad; resources: Henry H. L. Wu, Ewa M. Goldys, Carol A. Pollock, and Sonia Saad; supervision: Carol A. Pollock and Sonia Saad.

Funding

Funding for this study is supported by the Diabetes Australia Research Trust.

Acknowledgments

The authors would like to acknowledge the funding support from the Diabetes Australia Research Trust for this study.

References

- [1] D. B. Lee, E. Huang, and H. J. Ward, "Tight junction biology and kidney dysfunction," *Physiology*, vol. 290, no. 1, pp. F20–F34, 2006.
- [2] B. Kaeffer, "Survival of exfoliated epithelial cells: a delicate balance between anoikis and apoptosis," *BioMed Research International*, vol. 2011, Article ID 22131811, 2011.
- [3] C. J. Detrisac, R. K. Mayfield, J. A. Colwell, A. J. Garvin, and D. A. Sens, "In vitro culture of cells exfoliated in the urine by patients with diabetes mellitus," *The Journal of Clinical Investigation*, vol. 71, no. 1, pp. 170–173, 1983.
- [4] T. Marumo, J. Hoshino, W. Kawarazaki et al., "Methylation pattern of urinary DNA as a marker of kidney function decline in diabetes," *BMJ Open Diabetes Research and Care*, vol. 8, no. 1, 2020.
- [5] R. Pereira-Moreira and E. Muscelli, "Effect of insulin on proximal tubules handling of glucose: a systematic review," *Journal of Diabetes Research*, vol. 2020, Article ID 32377524, 2020.
- [6] A. T. Petermann, J. Pippin, R. Krofft et al., "Viable podocytes detach in experimental diabetic nephropathy: potential mechanism underlying glomerulosclerosis," *Nephron Experimental Nephrology*, vol. 98, no. 4, pp. e114–e123, 2004.
- [7] Y. Lytvyn, F. Xiao, C. R. Kennedy et al., "Assessment of urinary microparticles in normotensive patients with type 1 diabetes," *Diabetologia*, vol. 60, no. 3, pp. 581–584, 2017.
- [8] Y. Lu, D. Liu, Q. Feng, and Z. Liu, "Diabetic nephropathy: perspective on extracellular vesicles," Frontiers in Immunology, vol. 11, p. 943, 2020.
- [9] M. J. Fowler, "Microvascular and macrovascular complications of diabetes," *Clinical Diabetes*, vol. 26, no. 2, pp. 77–82, 2008.
- [10] Australian Diabetes Society, "HbA1c for diagnosis of diabetes mellitus," 2023, https://www.diabetessociety.com.au/ guideline/hba1c-for-diagnosis-of-diabetes-mellitus-may-2023/.
- [11] S. B. Mahbub, L. T. Nguyen, A. Habibalahi et al., "Non-invasive assessment of exfoliated kidney cells extracted from urine using multispectral autofluorescence features," *Scientific Reports*, vol. 11, no. 1, Article ID 10655, 2021.
- [12] L. A. Inker, N. D. Eneanya, J. Coresh et al., "New creatinineand cystatin C-based equations to estimate GFR without race," *New England Journal of Medicine*, vol. 385, no. 19, pp. 1737–1749, 2021.
- [13] D. W. Cockcroft and H. Gault, "Prediction of creatinine clearance from serum creatinine," *Nephron*, vol. 16, no. 1, pp. 31–41, 1976.
- [14] W. M. Michels, D. C. Grootendorst, M. Verduijn, E. G. Elliott, F. W. Dekker, and R. T. Krediet, "Performance of the Cockcroft-Gault, MDRD, and new CKD-EPI formulas in relation to GFR, age, and body size," *Clinical Journal of the American Society of Nephrology*, vol. 5, no. 6, pp. 1003–1009, 2010.
- [15] J. D. Elhai, P. S. Calhoun, and J. D. Ford, "Statistical procedures for analyzing mental health services data," *Psychiatry Research*, vol. 160, no. 2, pp. 129–136, 2008.
- [16] J. R. Singleton, A. G. Smith, J. W. Russell, and E. L. Feldman, "Microvascular complications of impaired glucose tolerance," *Diabetes*, vol. 52, no. 12, pp. 2867–2873, 2003.
- [17] W. B. Horton and E. J. Barrett, "Microvascular dysfunction in diabetes mellitus and cardiometabolic disease," *Endocrine Reviews*, vol. 42, no. 1, pp. 29–55, 2021.

[18] M. T. Coughlan, M. E. Cooper, and J. M. Forbes, "Renal microvascular injury in diabetes: RAGE and redox signaling," *Anti*oxidants & Redox Signaling, vol. 9, no. 3, pp. 331–342, 2007.

- [19] T. M. Curtis, T. A. Gardiner, and A. W. Stitt, "Microvascular lesions of diabetic retinopathy: clues towards understanding pathogenesis?," Eye, vol. 23, no. 7, pp. 1496–1508, 2009.
- [20] A. G. Huebschmann, J. G. Regensteiner, H. Vlassara, and J. E. B. Reusch, "Diabetes and advanced glycoxidation end products," *Diabetes Care*, vol. 29, no. 6, pp. 1420–1432, 2006.
- [21] Y. Li, Y. Liu, S. Liu et al., "Diabetic vascular diseases: molecular mechanisms and therapeutic strategies," *Therapy*, vol. 8, no. 1, p. 152, 2023.
- [22] B. Levine and D. J. Klionsky, "Development by self-digestion: molecular mechanisms and biological functions of autophagy," *Developmental Cell*, vol. 6, no. 4, pp. 463–477, 2004.
- [23] J. A. Gailit, D. A. Colflesh, I. L. Rabiner, J. A. Simone, and M. S. Goligorsky, "Redistribution and dysfunction of integrins in cultured renal epithelial cells exposed to oxidative stress," *American Journal of Physiology-Renal Physiology*, vol. 264, no. 1, pp. F149–F157, 1993.
- [24] S. Mackensen-Haen, A. Bohle, J. Christensen, M. Wehrmann, H. Kendziorra, and F. Kokot, "The consequences for renal function of widening of the interstitium and changes in the tubular epithelium of the renal cortex and outer medulla in various renal diseases," *Clinical Nephrology*, vol. 37, no. 2, pp. 70–77, 1992.
- [25] A. A. Eddy, "Overview of the cellular and molecular basis of kidney fibrosis," *Kidney International Supplements*, vol. 4, no. 1, pp. 2–8, 2014.
- [26] H. H. L. Wu, E. M. Goldys, C. A. Pollock, and S. Saad, "Exfoliated kidney cells from urine for early diagnosis and prognostication of CKD: the way of the future?," *International Journal of Molecular Sciences*, vol. 23, no. 14, p. 35886957, 2022.
- [27] A. Fukuda, A. Minakawa, M. Kikuchi et al., "Urinary podocyte mRNAs precede microalbuminuria as a progression risk marker in human Type 2 diabetic nephropathy," *Scientific Reports*, vol. 10, no. 1, Article ID 18209, 2020.
- [28] X. Liang, P. Wang, B. Chen et al., "Glycogen synthase kinase 3β hyperactivity in urinary exfoliated cells predicts progression of diabetic kidney disease," *Kidney International*, vol. 97, no. 1, pp. 175–192, 2020.
- [29] T. P. Kelder, M. E. Penning, H. W. Uh et al., "Quantitative polymerase chain reaction-based analysis of podocyturia is a feasible diagnostic tool in preeclampsia," *Hypertension*, vol. 60, no. 6, pp. 1538–1544, 2012.
- [30] A. Kalani, A. Mohan, M. M. Godbole et al., "Wilm's tumor-1 protein levels in urinary exosomes from diabetic patients with or without proteinuria," *PloS One*, vol. 8, no. 3, Article ID e60177, 2013.
- [31] S. E. Kahn, M. E. Cooper, and S. Del Prato, "Pathophysiology and treatment of Type 2 diabetes: perspectives on the past, present, and future," *Lancet*, vol. 383, no. 9922, pp. 1068– 1083, 2014.
- [32] H. Wei, S. Liu, Y. Liao et al., "A systematic review of the medicinal potential of mulberry in treating diabetes mellitus," *The American Journal of Chinese Medicine*, vol. 46, no. 8, pp. 1743–1770, 2018.

[33] Q. Chen, L. Zhu, Y. Tang, Z. Zhao, T. Yi, and H. Chen, "Preparation-related structural diversity and medical potential in the treatment of diabetes mellitus with ginseng pectins," *Annals of the New York Academy of Sciences*, vol. 1401, no. 1, pp. 75–89, 2017.

[34] K. W. Ng, Z. J. Cao, H. B. Chen, Z. Z. Zhao, L. Zhu, and T. Yi, "Oolong tea: a critical review of processing methods, chemical composition, health effects, and risk," *Critical Reviews in Food Science and Nutrition*, vol. 58, no. 17, pp. 2957–2980, 2018.