



Whole-Genome Sequencing and Annotation of *Exiguobacterium* sp. RIT 452, an Antibiotic-Producing Strain Isolated from a Pond Located on the Campus of the Rochester Institute of Technology

Anutthaman Parthasarathy,^a Narayan H. Wong,^a Nicole T. Cavanaugh,^a KayLee K. Steiner,^a Peter C. Wengert,^a Michael A. Savka,^a André O. Hudson^a

^aThomas H. Gosnell School of Life Sciences, Rochester Institute of Technology, Rochester, New York, USA

ABSTRACT *Exiguobacterium* sp. RIT 452 is of biotechnological importance given its potential for antibiotic production. Bactericidal activity was detected using spent medium extract in a disk diffusion assay against *Escherichia coli*. The genome consists of 3,246 protein-coding sequences, including a variety of gene clusters involved in the synthesis of antibacterial compounds.

Exiguobacterium is a genus of Gram-positive soil bacteria widely distributed in the environment, from tropical (1) to polar regions (2). A number of species have been sequenced and were shown to be extremophiles, such as hyperthermophiles, alkaliphiles, halophiles, and psychrophiles (2–4). Other members of this genus have potential applications in the bioremediation of pesticides and metals and other applications in the biotechnology industry (5–11). One strain was found to produce antimicrobial compounds (1). A strain was isolated from the guts of mealworms, and it was shown that the bacterium was able to degrade the plastic polymer polystyrene (12).

We embarked on a project to isolate and identify bacteria that are able to produce bactericidal compounds from a pond located on the campus of the Rochester Institute of Technology (RIT). The bacterium was isolated by directly plating 100 μ l of pond water sample on tryptic soy agar and growing it at 30°C under aerobic conditions. The bacterium was initially identified using PCR amplification and nucleotide sequencing of the 16S rRNA gene variable (V3/V4) regions using the following primers: 5'-CCTACGG GNGGCWGCAG-3' and 5'-GACTACHVGGGTATCTAATCC-3'.

Genomic DNA was isolated from a 5-ml culture grown in tryptic soy broth using the GenElute bacterial genomic DNA isolation kit (Sigma-Aldrich, USA) according to the manufacturer's protocol. For whole-genome sequencing, the genomic DNA was quantified using a NanoDrop spectrophotometer, and the genomic DNA was processed using the Nextera XT (Illumina) library preparation kit for sequencing using the MiSeq Illumina platform at the Rochester Institute of Technology Genomics Facility. Adapter trimming was done using the MiSeq Reporter software using the default parameters

Received 26 September 2018 Accepted 4 October 2018 Published 1 November 2018

Citation Parthasarathy A, Wong NH, Cavanaugh NT, Steiner KK, Wengert PC, Savka MA, Hudson AO. 2018. Whole-genome sequencing and annotation of *Exiguobacterium* sp. RIT 452, an antibiotic-producing strain isolated from a pond located on the campus of the Rochester Institute of Technology. Microbiol Resour Announc 7:e01341-18. <https://doi.org/10.1128/MRA.01341-18>.

Editor John J. Dennehy, Queens College

Copyright © 2018 Parthasarathy et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to André O. Hudson, aohsbi@rit.edu.

TABLE 1 Summary of antiSMASH results for *Exiguobacterium* sp. RIT 452

Cluster no.	Predicted biosynthetic metabolite	Coordinates within the genome	% similarity to known cluster
1	Terpene	69899–90726	33 (with carotenoid_biosynthetic_gene_cluster)
4	Siderophore	505835–519165	
9	Putative antibiotic	897206–908755	26 (with lugdunin_biosynthetic_gene_cluster)
20	Terpene	160337–181161	

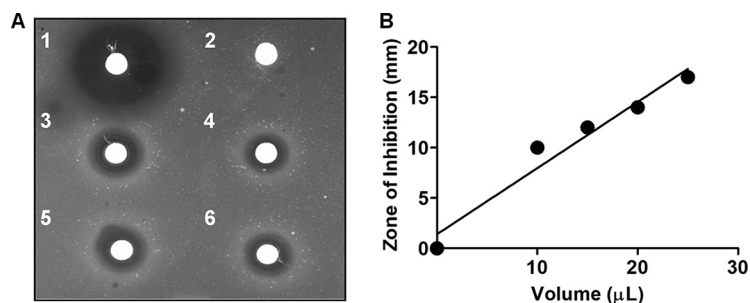


FIG 1 (A) Disk diffusion assay using ethyl acetate extract of spent medium of *Exiguobacterium* sp. RIT 452 against *Escherichia coli* ATCC 25922. (1) Tetracycline, 20 μ l (10 mg/ml); (2) methanol, 20 μ l; and (3, 4, 5, and 6) 25 μ l, 10 μ l, 20 μ l, and 15 μ l of RIT452 extract, respectively. (B) Diameter of the zone of inhibition (ZOI) showing the positive correlation between ZOI and volume.

(sequences with >90% sequence identity to adapter sequences were trimmed). The trimmed reads were subsequently assembled *de novo* with Unicycler version 0.3.0b (13). An assembly of 1.76 million Illumina paired-end reads generated 27 contigs with a total length of 3,217,892 bp, an N_{50} value of 693,695 bp, and a GC content of 47.86%. The National Center for Biotechnology Information (NCBI) Prokaryotic Genome Annotation Pipeline (PGAP) predicted 3,246 protein-coding sequences, 5 rRNAs, and 67 tRNAs (14, 15).

A scan of the genome using the antibiotics and secondary metabolite analysis shell (antiSMASH4.0) webserver showed evidence that the bacterium has 27 gene clusters potentially encoding pathways for the synthesis of secondary metabolites, including carotenoids, other terpenes, and possibly antibiotics (16). A summary of the results highlights 4 of the 27 clusters (Table 1). With regard to the production of bactericidal compounds, the antiSMASH *in silico* analysis was corroborated by a disk diffusion inhibitory assay against *Escherichia coli* ATCC 25922 using ethyl acetate extract from *Exiguobacterium* sp. RIT 452 (Fig. 1).

Data availability. This whole-genome project for *Exiguobacterium* sp. RIT 452 has been deposited in GenBank under accession number [QXJB000000000](https://www.ncbi.nlm.nih.gov/GenBank/QXJB000000000). The version described in this paper is the first version, QXJB01000000 (BioProject number [PRJNA489292](https://www.ncbi.nlm.nih.gov/BioProject/PRJNA489292); BioSample number [SAMN09954399](https://www.ncbi.nlm.nih.gov/BioSample/SAMN09954399)).

ACKNOWLEDGMENTS

A.P., N.H.W., N.T.C., K.K.S., P.C.W., M.A.S., and A.O.H. acknowledge ongoing support from the Gosnell School of Life Science (GSoS) at the Rochester Institute of Technology (RIT) and the College of Science (COS). N.T.C. was also supported by a 2017 COS Summer Undergraduate Research Fellowship.

REFERENCES

- Shanthakumar SP, Duraisamy P, Vishwanath G, Selvanesan BC, Ramaraj V, Vasantharaj David B. 2015. Broad spectrum antimicrobial compounds from the bacterium *Exiguobacterium mexicanum* MSSRF59. *Microbiol Res* 178:59–65. <https://doi.org/10.1016/j.micres.2015.06.007>.
- Vishnivetskaya TA, Chauhan A, Layton AC, Pfiffner SM, Huntemann M, Copeland A, Chen A, Kyrpidis NC, Markowitz VM, Palaniappan K, Ivanova N, Mikhailova N, Ovchinnikova G, Andersen EW, Pati A, Stamatis D, Reddy TB, Shapiro N, Nordberg HP, Cantor MN, Hua XS, Woyke T. 2014. Draft genome sequences of 10 strains of the genus *Exiguobacterium*. *Genome Announc* 2:e01058-14. <https://doi.org/10.1128/genomeA.01058-14>.
- Vishnivetskaya TA, Lucas S, Copeland A, Lapidus A, Glavina del Rio T, Dalin E, Tice H, Bruce DC, Goodwin LA, Pitluck S, Saunders E, Brettin T, Detter C, Han C, Larimer F, Land ML, Hauser LJ, Kyrpidis NC, Ovchinnikova G, Kathariou S, Ramaley RF, Rodrigues DF, Hendrix C, Richardson P, Tiedje JM. 2011. Complete genome sequence of the thermophilic bacterium *Exiguobacterium* sp. AT1b. *J Bacteriol* 193:2880–2881. <https://doi.org/10.1128/JB.00303-11>.
- Castro-Severyn J, Remonsellez F, Valenzuela SL, Salinas C, Fortt J, Aguilar P, Pardo-Esté C, Dorador C, Quatrini R, Molina F, Aguayo D, Castro-Nallar E, Saavedra CP. 2017. Comparative genomics analysis of a new *Exiguobacterium* strain from Salar de Huasco reveals a repertoire of stress-related genes and arsenic resistance. *Front Microbiol* 8:456. <https://doi.org/10.3389/fmicb.2017.00456>.
- Kumar A, Singh V, Kumar R. Characterization of an alkaliphile, *Exiguobacterium* sp. and its application in bioremediation, p 115. *In* Proceedings of the International Conference on Extremophiles. Brest, France.
- López L, Pozo C, Rodelas B, Calvo C, Juárez B, Martínez-Toledo MV, González-López J. 2005. Identification of bacteria isolated from an oligotrophic lake with pesticide removal capacities. *Ecotoxicology* 14: 299–312. <https://doi.org/10.1007/s10646-003-6367-y>.
- Petrova MA, Mindlin SZ, Gorlenko ZM, Kaliaeva ES, Soina VS, Bogdanova ES. 2002. Mercury-resistant bacteria from permafrost sediments and

- prospects for their use in comparative studies of mercury resistance determinants. *Genetika* 38:1569–1574.
8. Okeke BC. 2008. Bioremoval of hexavalent chromium from water by a salt tolerant bacterium, *Exiguobacterium* sp. GS1. *J Ind Microbiol Biotechnol* 35:1571–1579. <https://doi.org/10.1007/s10295-008-0399-5>.
 9. Pattanapitpaisal P, Mabbett AN, Finlay JA, Beswick AJ, Paterson-Beedle M, Essa A, Wright J, Tolley MR, Badar U, Ahmed N, Hobman JL, Brown NL, Macaskie LE. 2002. Reduction of Cr(VI) and bioaccumulation of chromium by gram positive and gram negative microorganisms not previously exposed to Cr-stress. *Environ Technol* 23:731–745. <https://doi.org/10.1080/09593332308618367>.
 10. Takebe F, Hara I, Matsuyama H, Yumoto I. 2007. Effects of H₂O₂ under low- and high-aeration-level conditions on growth and catalase activity in *Exiguobacterium oxidotolerans* T-2-2T. *J Biosci Bioeng* 104:464–469. <https://doi.org/10.1263/jbb.104.464>.
 11. Yumoto I, Hishinuma-Narisawa M, Hirota K, Shingyo T, Takebe F, Nodasaka Y, Matsuyama H, Hara I. 2004. *Exiguobacterium oxidotolerans* sp. nov., a novel alkaliphile exhibiting high catalase activity. *Int J Syst Evol Microbiol* 54:2013–2017. <https://doi.org/10.1099/ijs.0.63129-0>.
 12. Yang Y, Yang J, Wu WM, Zhao J, Song Y, Gao L, Yang R, Jiang L. 2015. Biodegradation and mineralization of polystyrene by plastic-eating mealworms: part 2. Role of gut microorganisms. *Environ Sci Technol* 49:12087–12093. <https://doi.org/10.1021/acs.est.5b02663>.
 13. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595.
 14. Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:D851–D860. <https://doi.org/10.1093/nar/gkx1068>.
 15. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
 16. Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de Los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. *Nucleic Acids Res* 45:W36–W41. <https://doi.org/10.1093/nar/gkx319>.