Review Article



Colorectal cancer (CRC) as a multifactorial disease and its causal correlations with multiple signaling pathways

Mao-lin Wan^{1,*}, Yu Wang^{2,*}, Zhi Zeng³, Bo Deng⁴, Bi-sheng Zhu⁴, Ting Cao⁵, (b) Yu-kun Li⁶, Jiao Xiao⁷, (b) Qi Han⁴ and Qing Wu⁵

¹Department of Hepatobiliary and Pancreatic Surgery, Xianning Central Hospital, the First Affiliated Hospital of Hubei University of Science and Technology, Xianning, 437000, P.R. China; ²Department of Laboratory Medicine, The Second Affiliated Hospital of University of South China, Hengyang, 421001, P.R. China; ³Department of Pathology, Xianning Central Hospital, the First Affiliated Hospital of Hubei University of Science and Technology, Xianning, 437000, P.R. China; ⁴Department of Oncology, Xianning Central Hospital, the First Affiliated Hospital of Hubei University of Science and Technology, Xianning, 437000, P.R. China; ⁵Department of Oncology, Xianning Central Hospital, the First Affiliated Hospital of Hubei University of Science and Technology, Xianning, 437000, P.R. China; ⁵Department of Digestive Medical, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁶Key Laboratory of Tumor Cellular and Molecular Pathology, College of Hunan Province, Cancer Research Institute, University of South China, Hengyang, Hunan, 421001, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China; ⁷Department of Endocrinology, The Affiliated Nanhua Hospital, University of South China, Hengyang, 421002, P.R. China;

Correspondence: Qi Han (tj_hanqi@hotmail.com) or Qing Wu (nhwuqing@163.com)



Colorectal cancer (CRC) is the third most common malignancy and one of the leading causes of cancer-related death among men worldwide. CRC is a multifactor digestive pathology, which is a huge problem faced not only by clinicians but also by researchers. Importantly, a unique feature of CRC is the dysregulation of molecular signaling pathways. To date, a series of reviews have indicated that different signaling pathways are disordered and have potential as therapeutic targets in CRC. Nevertheless, an overview of the function and interaction of multiple signaling pathways in CRC is needed. Therefore, we summarized the pathways, biological functions and important interactions involved in CRC. First, we investigated the involvement of signaling pathways, including Wnt, PI3K/Akt, Hedgehog, ErbB, RHOA, Notch, BMP, Hippo, AMPK, NF-κB, MAPK and JNK. Subsequently, we discussed the biological function of these pathways in pathophysiological aspects of CRC, such as proliferation, apoptosis and metastasis. Finally, we summarized important interactions among these pathways in CRC. We believe that the interaction of these pathways could provide new strategies for the treatment of CRC.

Introduction

Colorectal cancer (CRC) is the third most common malignancy worldwide and is one of the leading causes of cancer-related death [1]. The high mortality rate is reflective of several factors, including the lack of apparent symptoms in the early stages of CRC and the absence of cancer prevention strategies in developing countries, which causes enormous economic and psychological burden for people worldwide [1,2]. Although surgical resection achieves good clinical efficacy for localized cases, the median 5-year overall survival rate is only approximately 66.4%, which is attributed to the vast majority of CRC cases being medically diagnosed in a late, incurable stage, and the effects of traditional cancer chemotherapy are limited [2].

CRC is a multifactorial digestive pathology and is an enormous problem facing not only clinicians but also scientific researchers [3]. Many investigators believe that inflammation, *Helicobacter pylori* infection, dynamics of gut microbiota, hormonal disorders, gene mutation, epigenetic modification, immune dysfunction and metabolic disturbance are the main and classical risk factors for CRC [4–9]. Several molecular mechanisms, including advanced glycation end product (AGE) [10], aberrant glycosylation

*These authors contributed equally to this work.

Received: 31 January 2020 Revised: 21 February 2020 Accepted: 25 February 2020

Accepted Manuscript online: 09 March 2020 Version of Record published: 20 March 2020



[11], abnormal telomerase activity [12], unfolded protein response (UPR) [13], angiogenesis [14], reactive oxygen species (ROS) production [15], epithelial-mesenchymal transition (EMT) [16], cell apoptosis, proliferation, survival, migration, invasion, self-renewal, differentiation and dedifferentiation reprogramming, are altered to survive host defense or therapeutic insults. However, the dysregulation of these molecular mechanisms may not explain CRC origins and development, suggesting that various genetic and epigenetic events occur at the gene level [17,18].

The function and interaction of molecular pathways have a significant role in multiple cancer types. Previous studies have indicated that CRC progression is mediated by the dysregulation of many signaling pathways, including Wnt [19], PI3K/Akt [20,21], Hedgehog [22], ErbB [23], RHOA [24], Notch [25], BMP [26], Hippo [27], AMPK [28], NF- κ B [29], MAPK [3] and JNK [30]. Moreover, the interaction of these pathways is precise and complicated. In addition, a growing body of research shows that genetic perturbation or epigenetic dysregulation can promote the development of CRC or that the cancer itself can provoke genetic perturbation or epigenetic dysregulation [18]. Vdovikova et al. found that bacteria can target host cell epigenetics to promote carcinogenesis in HCT8 cells [31]. Daeun et al. suggested that β -carotene could inhibit DNMT3A and global DNA methylation levels to decelerate CRC progression [32]. Wu and his colleagues summarized that epigenetics play an important role in CRC progression [33].

In the current review, we summarize recent progress in studying these important potential molecular mechanisms and highlight their impact on CRC in order to reveal an attractive therapeutic strategy for CRC in the near future.

Multiple carcinogenic and anticarcinogenic intracellular pathways in CRC

To explore the molecular pathogenesis of CRC, we summarize recent progress in CRC (Figure 1). The intracellular signaling pathways contributing to carcinogenesis have been elaborated, and the driver genes have been considered promising targets for tumor therapy. Moreover, increasing research on molecular disorders of CRC provides valuable insight into the carcinogenesis of CRC, which may be explained by several molecular mechanisms playing multiple roles at different stages or in different situations during cancer development.

Up-regulated Wnt signaling pathway promotes CRC progression

Wnt signaling can be divided into two types: canonical and noncanonical. In the canonical Wnt pathway, Wnt is engaged with its receptors LRP-5/6 and Frizzled, which also activates Disheveled (DVL), recruiting the complex (Axin, GSK-3 β , CK1, APC) to the receptor, which then impedes cytosolic GSK-3 β rendering it incapable of phosphorylating β -catenin, resulting in the accumulation of unphosphorylated β -catenin in the cytosol [34–36]. Unphosphorylated β -catenin then translocate to the cell nucleus, where it interacts with T cell-specific factor (TCF)/lymphoid enhancer-binding factor (LEF) and coactivators, including Pygopus (Pygo) and Bcl-9, to turn on Wnt target genes such as c-Myc, cyclin D1 and Cdkn1a [34–36].

One of the noncanonical Wnt signaling pathways, the planar cell polarity pathway (PCP), is primitively activated through the interaction between Wnt and Frizzled receptors, along with coreceptors DAAM1/2, to control the activity of small GTPases, including RhoA, which play a role in regulating cytoskeletal remodeling [37]. Rho-associated kinase (ROCK) and myosin are motivated by Rho GTPase to rearrange the cytoskeleton and actin. Activated Rac GTPase and Rac activate the Jun N-terminal kinase (JNK) pathway [24]. Another noncanonical Wnt signaling pathway is the Wnt/Ca²⁺ pathway, which is activated by Wnt5a. Guanine nucleotide binding protein (G-protein) is cleaved into protein β/γ subunits and G-protein alpha-t2 via frizzled FZD2, resulting in Ca²⁺ influx into cytoplasm to promote differentiation. Moreover, Ca²⁺ can promote calmodulin and CaMKII to promote the phosphorylation of T-cell factor and lymphoid enhancer factor, which can repress the canonical Wnt pathway [38–41].

The level of YAP, a key gene in the Hippo pathway, is increased by the β -catenin/TCF4 complexes that bind to the first intron of the YAP gene DNA enhancer element in CRC cells [27]. The first report of the cross-talk between Wnt and Notch was in fruit flies, which indicated that Wnt could be controlled by Notch [27]. The Notch pathway activated by the APC mutant is necessary for the initiation and progression of colonic polyps and subsequent tumors in mice [42]. Kwon and his colleagues [43] found that β -catenin could physically interact with the membrane-bound cytoplasmic tails of Notch. In a previous study, it was identified that the Ras pathway can be up-regulated by enhancing Ras stability in CRC by mutations in APC (WNT pathway). The author found that Ras stability was regulated by Wnt activity, regulating Ras recruitment to the proteasome [44].



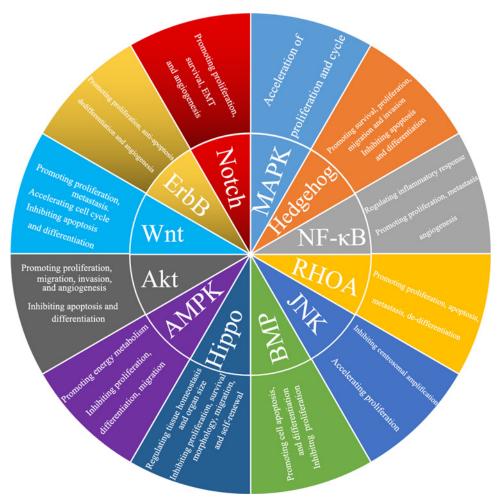


Figure 1. The function of these signaling pathways in CRC

Wnt, PI3K/Akt, Hedgehog, ErbB, Notch, NF-KB and MAPK can promote the carcinogenesis of CRC. BMP, AMPK and Hippo can inhibit the development and progression of CRC. However, RHOA and JNK may play dual roles in CRC.

The PI3K/Akt signaling pathway is enhanced and accelerates carcinogenesis in CRC

The PI3K/Akt signaling pathway is obviously up-regulated in CRC, and inhibition of this pathway may provide a potential therapeutic approach that may induce curable CRC [45,46]. Glucose metabolism is closely related to this signaling pathway in CRC. PI3K, a lipid kinase, promotes the phosphorylation of phosphatidylinositol-4,5-bisphosphate (PIP2) into phosphatidylinositol-3,4,5-trisphosphate (PIP3), in turn facilitating Akt phosphorylation to induce an intracellular signaling cascade [47]. In addition, AKT subtypes have convergent and divergent functions during tumorigenesis. PIP3 is degraded and its signal is terminated by phosphoinositol phosphatases, including PTEN, PIPP (INPP5J) and INPP4B. The PI3K pathway contributes to the growth and transformation of cancer cells and is mediated by proteins called phosphoinositol phosphatases, which are usually dysregulated in human cancers [48].

This pathway can also phosphorylate NF- κ B to facilitate cell survival, which phosphorylates and degrades I κ B (an inhibitor of NF- κ B). Therefore, liberated NF- κ B translocates to the nucleus to enhance cell survival and thus induce carcinogenesis. NF- κ B not only enhances survival but also induces angiogenesis [49], which also plays a significant role in CRC progression induced by PI3K/Akt/eNOS [50]. In addition, PI3K/Akt can promote the phosphorylation of MDM2, an oncogenic protein, which can promote the ubiquitination of p53. p53 is a significant tumor suppressor gene that accelerates cellular apoptosis in response to DNA damage from ionizing radiation or stress stimulation [51]. In CRC, the Fas/Fas-ligand system is an important downstream gene-expression cascade for the PI3K/Akt signaling pathway, which induces cellular apoptosis. Fas and Fas-ligand, members of the tumor necrosis factor receptor family, can recruit the Fas-associated death domain to induce cell apoptosis by activating the caspase cascade [52,53]. In



addition, PI3K/Akt can also enhance cell survival by phosphorylating Bad, a proapoptotic protein, to degrade it, which can increase the level and activity of the antiapoptotic proteins Bcl-xl and Bcl2 [54]. p27Kip1 (p27), a significant regulator of cyclin-dependent kinase 2, induces the G1/S transition, which can cooperate with p130Rb2 (p130) to inhibit the cellular cycle at the G1/S transition [55,56]. In addition, PI3K/Akt can obviously inhibit the expression of p27 and p130 to accelerate the cell cycle by inhibiting forkhead protein [57]. Furthermore, PI3K/Akt can deactivate glycogen synthase kinase 3 (GSK3) to increase the levels of cyclin D1 and myc, accelerating the cell cycle to upregulate proliferation [58].

Furthermore, the PI3K/Akt pathway can inhibit the Hippo pathway by promoting the phosphorylation of YAP to accelerate colon cancer cell proliferation [59]. In addition, the PI3K/Akt pathway can activate mTOR to promote protein synthesis, resulting in cell metabolism, growth and proliferation [46,60]. Other work has implicated interactions between PI3K/Akt signaling and BMP in colon cancers [61]. Taken together, these results suggest that this pathway contributes to CRC development.

The ErbB signaling pathway plays an oncologic role in CRC

The receptor tyrosine kinase superfamily is mainly distributed on the surface of CRC and breast cancer cells and plays a significant role in nearly every aspect of cell biology. The ErbB receptor family has four subtypes, including ErbB1 (EGFR), ErbB2 (HER2), ErbB3 (HER3) and ErbB4 (HER4) [62].

In a recent study, EGFR was overexpressed in 60–80% of colon cancers [62,63]. In addition, a standardized IHC test was used for the clinical trial analysis to separate CRC patients who expressed EGFR in at least 1% of the cancer cells [64,65]. EGFR triggers a molecular cascade that results in activation of the MAPK and PI3K signaling pathways, which can promote the proliferation, apoptosis inhibition, dedifferentiation and angiogenesis of CRC cells [66].

Park and his colleagues found that 47.4% (65/137) of patients with CRC exhibited overexpression of HER2, which was closely associated with poor prognosis [67]. Activation of HER2 is predominantly present in CRC cells, mediating differentiation, proliferation and apoptosis [68]. Furthermore, constant amplification of HER2 can mediate chemoresistance by activating ERK1/2 signaling [69]. HER2 is the preferred partner of other ErbBs, and the formation of dimers or complexes with HER3 and HER4 is involved in the initiation, development and progression of cancer [70].

HER3, as an impaired kinase in the receptor tyrosine kinase superfamily, is phosphorylated only by dimerization with another ErbB receptor [70,71]. This most often happens to HER2, which is the most carcinogenic member of the family [72]. Baiocchi showed a 69.7% HER3 response rate in patients with CRC, resulting in lymph vascular invasion [73]. In addition, HER3 is expressed in many tumors that express HER2 [74]. Therefore, the HER2/HER3 axis may play a significant role in the aberrant growth of CRC cells [75].

HER4, activated by heparin-binding EGF-like growth factor, neuregulins and betacellulin, can activate the PI3-kinase and Shc pathways to promote cell proliferation and metastasis but inhibit differentiation [76]. Daekee Lee found that inhibition of HER3 leads to increased caspase-3-mediated apoptosis in cancers, and absence of HER3 or HER4 leads to CRC cell apoptosis, which may be caused by the HER3-HER4 heterodimer-dependent AKT pathway. Furthermore, they observed that activation of PI3K/AKT is the primary mechanism promoting CRC growth in an EGFR–HER3 heterodimer-dependent manner [17].

In mouse CRC cells that have both WNT and Ras activating mutations, Christopher observed that ectopic HER4 expression enhances unanchored growth *in vitro* and the establishment of xenografts *in vivo*. Moreover, HER4 knockout disrupted the WNT-induced growth of CRC cells. These results suggest that high levels of HER4 coexist with activated WNT signaling to promote carcinogenesis of mouse and human colon cells. Furthermore, ectopic expression of HER4 is also correlated with EGFR pathway activation in human colorectal cancer [19]. Interestingly, EGFR can also activate the Ras–Raf–MEK–ERK signaling pathway in colon cancer, which accelerates the progression and development of colon cancer [3].

The dual function of the RHOA signaling pathway in CRC development

Many investigators believe that the RHOA signaling pathway, in its carcinogenic role, has a complex interaction with many cancers via proliferation, apoptosis, metastasis, dedifferentiation and polarization [77–79], although a recent study suggests that it has totally different function in CRC than it does in other cancers [80]. In previous studies, many investigators showed that activation of RHOA can promote the proliferation and growth of CRC [77,81]. Nevertheless, Paulo Rodrigues and his colleagues found that decreases in the levels of RHOA have a strong correlation with poor patient prognosis [80,82], and inactivation of RHOA induced obviously faster proliferation, metastasis, dedifferentiation and polarization *in vivo* and *in vitro* [80]. These data suggest that there are two different theories



about the activation state of RHOA: one is that activation of RHOA directly increases the proliferation and growth of cancer cells [77,81]. The other, in opposition, is that inactivation of RHOA depends on Rho-associated protein kinase (ROCK) and diaphanus-related formin 1 (DIAPH1) to up-regulate canonical Wnt signaling to maintain constant proliferation and growth [80]. In general, the suggestion that inactivation of RHOA can promote carcinogenesis is presumably because canonical Wnt signaling is a master regulator of carcinogenesis in CRC, and this pathway can induce constant proliferation and growth [83,84].

The Notch signaling pathway plays an oncogenous role in the development and progression of CRC

The Notch pathway is activated in multiple cancer types as a result of multiple genetic changes, such as point mutations, gene amplification, chromosomal translocations and other epigenetic modifications [85]. It interacts with four receptors, Notch 1/2/3/4, to regulate cell proliferation, differentiation, apoptosis and stem cell maintenance [86]. Different studies in CRC patients have hinted that the Notch pathway is implicated in the development of CRC, and a series of mechanisms have been proposed to demonstrate this regulation [87,88]. First, the Notch intracellular domain (NICD) is cleaved from the receptor by an γ -secretase complex [87], and the cleaved NICD translocates from the cytoplasm to the nucleus [88]. Second, NICD interacts with recombining binding protein suppressor of hairless (RBPJ), resulting in the activation of oncogenes, such as hairy enhancer of split (HES) family proteins, CDKN1A (also known as p21), HES-related proteins (HEY), Notch-regulated ankyrin repeat protein, cyclin D1/3, c-myc and HER2 [89,90]. Many investigators have demonstrated that the Notch pathway is up-regulated in patients with CRC, which ultimately induces cancer cell proliferation, survival, EMT and angiogenesis [87,91,92]. Notch ligand expression is increased in several solid tumors, including CRC and pancreatic cancer. In addition, there appears to be a close interaction between the Notch and Ras signaling pathways, as Ras activating mutations have been shown to activate Notch signaling, which is required for Ras-mediated transformation [93].

Bone morphogenetic protein (BMP)/SMAD pathways as important tumor suppressor pathways in CRC

It is generally accepted that BMP4, a member of the transforming growth factor- β (TGF- β) superfamily, plays a significant role during embryonic development that regulates cell apoptosis, proliferation and differentiation [26]. Regarding the function of BMP4 in cancers, BMP4 has been indicated to mediate the differentiation of cancer stem cells and inhibit the cancer growth of CRC [94].

BMP, a secreted glycoprotein, activates and binds to BMPR1A (ALK3), BMPR1B (ALK6) and BMPRII receptors by activating Drosophila (SMAD) 1/5/8 and phosphorylated Mothers against decapentaplegic [95]. Subsequently, activated p-SMAD 1/5/8 translocates from the cytoplasm to the nucleus by forming a complex with SMAD4, and ultimately, it regulates transcription and expression of target genes such as inhibitor of DNA binding (ID) [96] and CCL15 [97]. Importantly, the BMP pathway is down-regulated in the majority of colon cancer cases [98].

A recent report indicated that β -catenin can be inhibited by BMP via SMAD4 to down-regulate the expression of *c-myc* and *Axin2* in microdissected small intestinal adenomas of tamoxifen-treated mice [98]. Another study sufficiently suggests that BMP and WNT appear to be interconnected via the PI3k/Akt pathway [99,100].

The Hedgehog signaling (HH) pathway plays a tumorigenic role in CRC

HH signaling was first detected in *Drosophila melanogaster* and was demonstrated to play a significant role in mediating proliferation, establishing cellular fate and regulating embryonic development [101]. The HH family is a group of proteins implicated in many cellular functions: cell survival, proliferation, apoptosis, differentiation, migration and invasion [102]. Hedgehog (SHH, IHH or DHH) binds to the receptor PTCH1, which is a direct target of Hedgehog signaling [103]. This interaction can relieve PTCH1-mediated repression of SMO, which can subsequently activate a downstream signaling pathway, leading to the translocation of GLI1 and GLI2 (GLI family zinc-finger transcription factors) from the cytoplasm to the nucleus [104], where they regulate the transcription of several target genes, including platelet-derived growth factor (*PDGF*) [105] and *SNAIL* [106].

Hh signaling is involved in some types of solid tumors, especially basal cell skin cancer and medulloblastoma. Notably, the HH signaling pathway plays a key role in the progression and repair of colon epithelial cells, and a previous study indicated that it was activated in CRC. Various studies have reported up-regulation of HH ligand SHH, HH receptor PTCH1, and hh-associated transmembrane receptor Smoh in hyperplastic polyps, adenomas, and adenocarcinomas of the colon [93]. Exogenous SHH can promote the growth of colon cells in primary mouse



models, suggesting that shh-triggered signals may promote the development of CRC. The study also showed that the expression level of Shh mRNA in CRC cells was significantly higher than that in normal colon cells [107].

The Hippo signaling pathway involved in the anticarcinogenesis of CRC

The Hippo signaling pathway is a significant signaling pathway in the regulation of stem cell proliferation, morphology, survival, migration, self-renewal, migration, tissue homeostasis and organ size [108–110]. Recently, Hippo signaling has been considered one of the most significant signaling pathways in tumorigenesis, as it inhibits the development of tumors though multiple components of this pathway, including fat storage-inducing transmembrane protein, large tumor suppressor kinase (LATS), macrophage stimulating (MST), taffazin (TAZ), Yes-associated protein 1 (YAP1) and transcriptional enhancer associated domain (TEAD) [111]. In brief, MST increased by the transmembrane protein of FAT upregulates LATS to phosphorylate YAP, which prevents the translocation of YAP from the cytoplasm to the nucleus. Ultimately, it attenuates the ability of the YAP/TAZ complex to interact with TEAD [112–114], resulting in the inactivation of oncogenes such as β -catenin, k-ras, and Akt/mTOR in colon tumor initiation and progression [115–117].

In CRC, the Hippo signaling pathway is suppressed, and the expression of YAP is increased, which induces the migration, invasion, proliferation and EMT of colon cancer cells [16]. The interaction between $Ck1\delta/\epsilon$ and Dvl is impeded by TAZ, which blocks Dvl phosphorylation mediated by Wnt3a and finally represses Wnt/ β -catenin signaling. Furthermore, dysregulation of MST and LATS also blocks the membrane translocation of TAZ to block the pathophysiological effect of Wnt3a [118]. In addition, YAP1/KLF5 can bind to the promoter of Achaete scute-like 2 (Ascl2), which is a Wnt signaling target, and transcriptionally enhance its expression, resulting in increased self-renewability of CRC progenitor cells [119]. In colon-derived cell lines, inhibition of YAP obviously decreased the effect of Notch and Wnt signaling to reduce proliferation and survival ability. In Mst1/2-deficient cells, the expression of Hes1 and Hey1 (key regulatory genes in the Notch pathway) was largely enhanced [120]. Moreover, YAP overexpression induced Notch ectopic activation to inhibit cell differentiation [121].

The function and mechanism of AMPK, a key energy metabolism pathway, in CRC

AMPK, a serine/threonine kinase, is characterized by sensitivity to changes in the ratio of AMP/ATP; AMPK acts as a cellular energy sensor, regulating various cell progression processes such as CRC cell survival, proliferation, differentiation, migration and metabolism [122,123].

At the molecular level, the major effects of AMPK are predominantly exerted through the regulation of oxidative phosphorylation and the interaction of other signaling pathways. In particular, activation of AMPK can activate p53, which induces autophagy and apoptosis and decelerates the cell cycle. Phosphorylated AMPK inhibits the phosphorylation of mTOR to inhibit cell proliferation and protein synthesis by mediating tuberous sclerosis 1 and 2 (TSC1/2) [20,21], inactivating Rag GTPases [124] and up-regulating the expression of REDD1 [125]. Furthermore, activation of AMPK negatively regulates receptor tyrosine kinase pathways, such as the ErbB2 and EGFR pathways, further targeting the downstream protein factors mTOR and ERK [23]. Inactivated AMPK can also inhibit insulin receptor substrate-1 (IRS1), which is an activating factor for the IGF1/insulin signaling axis and subsequent PI3K/Akt/mTOR signaling, which can inhibit neoplastic activity [126]. AMPK activation by metformin recruits anti-angiogenic and anti-inflammatory factors, such as interleukin-1 β (IL-1 β), tumor necrosis factor α (TNF α), IL-6, nuclear factor κ light-chain-enhancer of activated B-cells (NF- κ B) and hypoxia inducible factor-1 α (HIF-1 α), diminishing the effect of vascular endothelial growth factor (VEGF) [127,128]. AMPK, as an upstream regulator of the Hippo pathway, can also promote the phosphorylation of YAP to inhibit its effects on colon cancer cells, including proliferation, apoptosis inhibition, glucose uptake and glycolysis [28].

On the other hand, AMPK displays obvious metabolic effects that are associated with malignancy, such as the development of a lipogenic phenotype. Phosphorylation of AMPK can facilitate the phosphorylation of acetyl-coenzyme A carboxylase-1 (ACC-1) to decrease the levels of fatty acid synthase (FASN), sterol-regulatory element-binding protein (SREBP-1c) and stearoyl coenzyme A desaturase-1 (SCD-1) [127,129], thereby impeding lipogenesis, a process needed for cancer cells to meet the excess nutritional requirements of sustained cancer cell growth and subsequent cancer cell proliferation. Moreover, the Warburg effect, as a hallmark of cancer metabolism, affects glucose metabolism to further influence amino acid and lipid metabolism. Elise et al. indicated that resveratrol could reverse the Warburg effect by activating the AMPK pathway in CRC [130]. This result indicated that many signaling pathways might influence the Warburg effect by directly or indirectly activating the AMPK pathway. Therefore, whether other pathways can independently affect metabolism requires the exclusion of the effect of AMPK in future studies.



The NF-KB signaling pathway, a central mediator between inflammation and cancer, promotes the development of CRC

The NF- κ B signaling pathway has central roles in innate and adaptive cellular immunity and serves as an important link between inflammation and cell survival. In the inactivated state, a large portion of NF- κ B is bound to inhibitor of NF- κ B (IkB) proteins to form dimers in the cytosol; NF- κ B signaling is divided into two pathways: the canonical pathway and the noncanonical pathway [29].

In the canonical NF- κ B signaling pathway, NF- κ B can be activated by multiple inflammatory stimuli, such as lipopolysaccharide (LPS), IL-1 α/β , TNF- α and many medicines. Stimulus-specific receptors release these various inflammatory stimuli to convert into protein phosphorylation signals. The activation signal of the receptor is transmitted mainly by I κ B kinase (IKK), which promotes I κ B inhibitory protein degradation and phosphorylation. The activated IKK complex consists of two catalytic subunits (IKK α and IKK β) and a regulatory component (IKK γ /NF- κ B essential modulator (NEMO)). Normally, the activation of IKK is mainly mediated by IKK β . Finally, IKK activates I κ B to promote NF- κ B translocation into the nucleus, where it activates target genes, such as chemokines, cytokines and antiapoptotic genes [131]. The noncanonical NF- κ B pathway is primarily mediated by B cell activating factor (BAFF), CD40 ligand and lymphotoxin $\beta/$ (LT β), which are members of the TNF family. NF- κ B-inducing kinase (NIK) can promote the activation of generic IKK α . Subsequently, the p100 protein processing process begins. By processing p100, nuclear translocation of p52/NF- κ B2 complexes induces downstream gene expression to mediate secondary lymphoid organ development and to regulate adaptive immune responses [29].

Constitutional NF- κ B activation in cancer cells enhances the level of growth factor genes such as hepatocyte growth factor (HGF), BMP, and granulocyte colony-stimulating factor (G-CSF) [29]. Furthermore, NF- κ B, a significant transcription factor, binds to the cyclin D1, D2, and D3 promoters to accelerate the cancer cell cycle [134]. In addition, cancer metastasis can be promoted by activating NF- κ B by increasing gelatinases (MMP-2 and MMP-9) [135]. NF- κ B also inhibits expression of epithelial markers E-cadherin and desmoplakin and induces expression of mesenchymal marker vimentin [136]. Tong et al. found that apigenin, a Chinese medicine monomer, inhibits the activation of NF- κ B to inhibit Snail expression, which further indicated that the EMT cascade was one of the downstream targets of the NF- κ B pathway [137]. On the other hand, NF- κ B repression may be an effective anti-angiogenic therapy for CRC, especially in the case of NF- κ B activation. Microarray and proteomic array analyses identified that several chemokines were associated with increased angiogenesis and constitutive NF- κ B activation of cancer cells [138]. Inhibition of NF- κ B could repress these chemokines, such as MCP-1, VEGF, IL-8 and Gro- α [138–141]. Moreover, PI3K promoted the transcription of MDM2 and degradation of p21 in an NF- κ B-dependent manner, which can promote colon cancer cell migration [142].

Ras/Raf/MEK/MAPK/ERK signaling pathway dysregulation promotes the progression of CRC

The RAS/mitogen-activated protein kinase (MAPK) signaling pathway has been widely studied in mammalian cells and plays a significant role in human cancer by promoting cell proliferation and cell cycle progression. Approximately 30% of tumors have tumorigenic mutations in the RAS gene. Activated RAS mediates RAF activation and subsequently promotes RAF phosphorylation and mitogen-activated protein kinase (MEK) activation, which promotes the phosphorylation and activation of MAPK/extracellular signal-related kinase (ERK) [143].

In CRC, gene mutations in the MAPK pathway are activating. Ganoderma lucidum polysaccharide (GLP) increases the expression of JNK via the MAPK pathway and induces the apoptosis of HCT-116 cells. GLP-induced apoptosis of human CRC cells is associated with mitochondrial and MAPK pathway activation [144]. However, in another report, activation of PIK3CA mutations decreased sensitivity to MEK inhibitors, while PTEN mutations appeared to lead to complete resistance. Furthermore, dual inhibition of the PI3K/AKT and RAF/MEK/ERK pathways also appears to completely inhibit the effect of the downstream mTOR pathway [145]. Interestingly, Wnt signaling was found to be a potential mediator of resistance to the MEK1/2 inhibitor in cancers with BRAF mutation [146], which may be induced by cell migration-inducing and hyaluronan-binding protein (CEMIP) [147]. Furthermore, another study indicated that AKT and Wnt pathways were slightly attenuated by the k-Ras inhibitors ABT263 and axitinib in k-Ras mutant CRC cells [148], which further suggested that Ras signaling could interact with AKT and Wnt pathways in CRC.

The C-Jun N-terminal kinase (JNK) signaling pathway has a dual effect on CRC progression

The proto-oncoprotein c-Jun is a component of the AP-1 transcription factor, and its activity is enhanced in a variety of cancer types [30]. A significant mechanism by which AP-1 is activated is phosphorylation of c-Jun n-terminal kinases



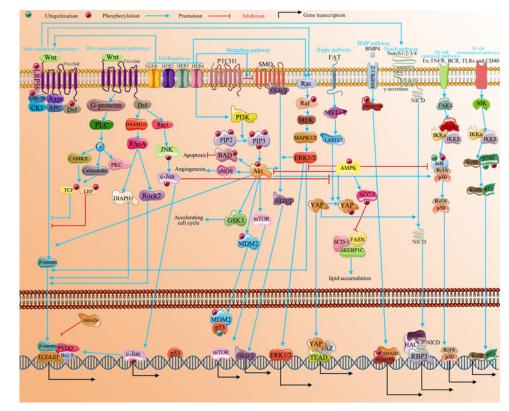


Figure 2. The correlation of multiple signaling pathways in CRC

The development and progression of CRC is mediated by multiple signaling pathways, including Wnt, PI3K/Akt, Hedgehog, ErbB, RHOA, Notch, BMP, AMPK, Hippo, NF-kB, MAPK and JNK. These signaling pathways do not function singly but interact with each other. AMPK, BMP and Hippo mainly play anticancer roles in CRC progression. Other pathways are involved in CRC as carcinogenic factors. These pathways interact not only in the cytoplasm but also in the nucleus. These interactions regulate the transcription of downstream genes leading to cancer cell proliferation, apoptosis inhibition, migration, invasion and dedifferentiation.

(JNKs) on the amino terminus of c-Jun [149]. Phosphorylation of c-Jun can form a ternary complex with TCF4 (the HMG-box transcription factor) and β -catenin. Furthermore, TCF4 could also interact on the promoter to activate c-Jun in a β -catenin-dependent manner [150]. In addition, using the Apc^{Min} mouse CRC model, inhibition of phosphorylated c-Jun [151] or c-Jun colon-specific inactivation mice attenuated the number, shape and size of cancer as well as increased their lifespan [152]. Transcriptional cooperation between TCF4 and c-Jun is activated by β -catenin and relies on TCF and AP-1 c-jun promoter elements, indicating that the interaction of phospho-c-Jun-TCF4 may promote transcription by recruiting β -catenin to the proximal transcriptional initiation site of AP-1 elements [150].

However, JNK1 is located upstream of Stat3. The JNK1-Stat3 pathway was first found to inhibit centrosomal amplification of human HCT116 cells, suggesting that it has another anticancer mechanism [150,153]. The CRC pathogenesis study conducted by Bai et al. [154] found that ZBP-89 induces apoptosis mediated by JNK activation by inhibiting JNK dephosphorylation.

Several studies have indicated that the Hippo pathway is mediated by JNK. Bleomycin can activate JNK to mediate the nuclear translocation of Yki, which enhances the expression of Upd and activates Jak-Stat signaling to accelerate cell proliferation [155,156]. Furthermore, JNK activates YAP1 in response to DNA damage [157]. JNK enhances the activation of YAP1 by inhibiting LATS by promoting the binding of Ajuba (the mammalian homologue of Jub) and LATS [158]. However, no studies have been conducted to establish a connection between the JNK signaling pathway and the Hippo pathway in the mammalian colon, which might indicate an interaction involving both the JNK pathway and the Hippo pathway in CRC tumorigenesis. In general, these results suggest that the JNK pathway may play a dual role in the carcinogenesis of CRC.



The importance of communication among these pathways

The linear progression of the PI3K/Akt, Hedgehog, ErbB, RHOA, Notch, BMP, Hippo, AMPK, NF-κB, MAPK and JNK pathways is summarized in Figure 2. Nevertheless, a growing number of studies on a variety of tissues suggest that these important signaling pathways participate in cross-talk or share crucial junctions of interaction [159,160].

Wht signaling has been reported to control the expression of YAP [27] and to regulate Gli3 from the HH pathway [102]. Furthermore, it also interacts with Notch [102], Rho [24], JNK [30] and Ras signaling [44]. PI3K/Akt [17], Wnt [19] and Ras [3] signaling can be regulated by ErbB. RAS mutations mediate the stability of the RAS protein, resulting in RAS proteins existing primarily in the GTP binding form, making them structurally active and immune to exogenous growth factors such as EGF. Thus, the mutant KRAS protein that makes up the active conformation may make tumor cells resistant to ErbB-targeted drugs, including cetuximab or panizumab [161]. Activation of the PI3K/AKT pathway can promote the NF-KB pathway to accelerate the development and progression of CRC [162]. Furthermore, k-Ras can also increase the level of NF-KB to promote carcinogenesis in CRC [163]. Activating mutations of k-Ras can increase Gli1 via the RAF/MEK/ERK and PI3K/AKT pathways [164,165]. The relationship between Wnt/β-catenin and Hedgehog/Gli was observed in CRC. These pathways are controlled by common factors, including GSK3β, p53, PTEN and KRAS. β-catenin could be increased by Gli1 via Snail and Wnt. In addition, Gli1 could also be enhanced by β -catenin though c-myc, which binds to the coding region of Gli1 mRNA to increase stability [166]. Furthermore, a previous study indicated that SMO can inhibit the activity and nuclear translocation of β -catenin, which does not rely on the function of Gli [167]. Moreover, HH can also interact with Notch by activating Hes-1 [168]. Activated AMPK can not only inhibit the activation of ErbB [23], PI3K/Akt [20,21] and Ras [23] signaling but can also promote the biological function of NF-κB [127,128]. Likewise, Hippo can be activated by AMPK [28] but inactivated by both PI3K/Akt [59] and JNK [158]. The synergistic effect on Hes-1 is switched on by Notch and HH signaling [168]. The Notch pathway can also be up-regulated by Wnt [25] and down-regulated by Hippo [121]. BMP and Wnt appear to be interconnected via the PI3k/Akt pathway [100]. Furthermore, Smad promotes EMT through Wnt, Ras, HH and Notch [169]. BMPs are also regulated by NF- κ B, which indicates the importance and complexity of pathway interactions [132,133]. In addition, a recent report noted that k-Ras up-regulated ERK levels to inhibit BMP4 [132,133,170].

Therefore, the interaction among these pathways has a significant role in the formation and development of cancer. Feedback loops are also essential factors in carcinogenesis, which indicates that a slight move in one part may affect the situation as a whole.

Conclusion

This paper reviews the pathophysiological functions, characteristics and interactions of multiple signaling pathways in CRC. A large number of pathophysiological studies have confirmed that dysregulation of signaling pathways plays a significant role in CRC, promoting cell proliferation and migration but inhibiting cell differentiation and apoptosis through multiple interactions and feedback loops [159,160].

Over the past decade, many pathways been shown to play a role in tumorigenesis, driving the development of targeted therapies. Studies have shown that the roles of these pathways are not isolated but are interconnected, with changes in one pathway leading to changes in the other [171]. By focusing on molecular cross-talk, it is possible to develop more effective therapeutic strategies.

Recent studies have suggested that the accumulation of mutations that lead to tumor heterogeneity might not act as bystanders but rather may play an active role in establishing a unique biological phenotype for each cancer patient. By understanding the interplay of pathways, we can develop effective global strategies for multiple cancers. The formation, development and progression of CRC is mediated by the dysregulation of multiple signaling pathways, including Wnt [19], PI3K/Akt [20,21], HH [22], ErbB [23], RHOA [24], Notch [25], BMP [26], Hippo [27], AMPK [28], NF-κB [29], MAPK [3] and JNK [30].

Hence, it is necessary to conduct in-depth research on the relationship between the basic etiology, inducing factors and clinical treatment of CRC to develop new therapeutic strategies with more scientific and clinical value [3]. However, if we treat CRC by targeting a single gene, we diverge from the optimal path to accurately treating CRC. Because the causes of CRC are so diverse, multidrug combinations known as 'cocktail therapy' will promote more systematic treatment of the disease. Finally, the concept of epigenetics has been widely applied in the occurrence and development of CRC, and investigating these changes will help us explore new treatment options and improve the early diagnosis and treatment of CRC [172].



10

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

The authors declare that there are no sources of funding to be acknowledged.

Abbreviations

AGE, advanced glycation end product; CRC, colorectal cancer; EMT, epithelial-mesenchymal transition; ROS, reactive oxygen species; UPR, unfolded protein response.

References

- 1 Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R.L., Torre, L.A. and Jemal, A. (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **68**, 394–424, https://doi.org/10.3322/caac.21492
- 2 Huang, Y., Zhao, M., Yin, J. et al. (2019) Pulmonary metastasis in newly diagnosed colon-rectal cancer: a population-based nomogram study. Int. J. Colorectal Dis. 34, 867–878, https://doi.org/10.1007/s00384-019-03270-w
- 3 Li, K., Guo, Q., Yang, J. et al. (2017) FOXD3 is a tumor suppressor of colon cancer by inhibiting EGFR-Ras-Raf-MEK-ERK signal pathway. Oncotarget 8, 5048
- 4 Eaden, J.A., Abrams, K.R. and Mayberry, J.F. (2001) The risk of colorectal cancer in ulcerative colitis: a meta-analysis. *Gut* **48**, 526–535, https://doi.org/10.1136/gut.48.4.526
- 5 Shahanavaj, K., Gil-Bazo, I., Castiglia, M. et al. (2015) Cancer and the microbiome: potential applications as new tumor biomarker. *Expert Rev. Anticancer Ther.* **15**, 317–330, https://doi.org/10.1586/14737140.2015.992785
- 6 Westbrook, A.M., Wei, B., Braun, J. and Schiestl, R.H. (2009) Intestinal mucosal inflammation leads to systemic genotoxicity in mice. *Cancer Res.* 69, 4827–4834, https://doi.org/10.1158/0008-5472.CAN-08-4416
- 7 Zitvogel, L., Ma, Y., Raoult, D., Kroemer, G. and Gajewski, T.F. (2018) The microbiome in cancer immunotherapy: Diagnostic tools and therapeutic strategies. *Science* 359, 1366–1370, https://doi.org/10.1126/science.aar6918
- 8 Williams, C., Dileo, A., Niv, Y. and Gustafsson, J.Å. (2016) Estrogen receptor beta as target for colorectal cancer prevention. *Cancer Lett.* **372**, 48–56, https://doi.org/10.1016/j.canlet.2015.12.009
- 9 Ponzetto, A. and Figura, N. (2019) Colon Cancer Risk and VacA Toxin of Helicobacter pylori. *Gastroenterology* **156**, 2356, https://doi.org/10.1053/j.gastro.2018.11.083
- 10 Wang, P., Lu, Y.C., Li, Y.F., Wang, L. and Lee, S.C. (2018) Advanced Glycation End Products Increase MDM2 Expression via Transcription Factor KLF5. J. Diabetes Res. 2018, 3274084, https://doi.org/10.1155/2018/3274084
- 11 Hung, J.S., Huang, J., Lin, Y.C. et al. (2014) C1GALT1 overexpression promotes the invasive behavior of colon cancer cells through modifying 0-glycosylation of FGFR2. Oncotarget 5, 2096–2106, https://doi.org/10.18632/oncotarget.1815
- 12 Kuranaga, N., Shinomiya, N. and Mochizuki, H. (2001) Long-term cultivation of colorectal carcinoma cells with anti-cancer drugs induces drug resistance and telomere elongation: an in vitro study. *BMC Cancer* **1**, 10, https://doi.org/10.1186/1471-2407-1-10
- 13 Sustic, T., van Wageningen, S., Bosdriesz, E. et al. (2018) A role for the unfolded protein response stress sensor ERN1 in regulating the response to MEK inhibitors in KRAS mutant colon cancers. *Genome Med.* **10**, 90, https://doi.org/10.1186/s13073-018-0600-z
- 14 Velatooru, L.R., Vakamullu, S., Penugurti, V. and Reddy, S.P. (2019) Alpinoid c analog inhibits angiogenesis and induces apoptosis in COL0205cell line. *Chem. Biol. Interact.* **308**, 1–10, https://doi.org/10.1016/j.cbi.2019.05.009
- 15 Wu, D., Zhou, W.Y., Lin, X.T., Fang, L. and Xie, C.M. (2019) Bufalin induces apoptosis via mitochondrial ROS-mediated caspase-3 activation in HCT-116 and SW620 human colon cancer cells. *Drug Chem Toxicol.* **42**, 444–450, https://doi.org/10.1080/01480545.2018.1512611
- 16 Sun, M., Song, H., Wang, S. et al. (2017) Integrated analysis identifies microRNA-195 as a suppressor of Hippo-YAP pathway in colorectal cancer. *J. Hematol. Oncol.* **10**, 79, https://doi.org/10.1186/s13045-017-0445-8
- 17 Lee, D., Yu, M., Lee, E. et al. (2009) Tumor-specific apoptosis caused by deletion of the ERBB3 pseudo-kinase in mouse intestinal epithelium. *J. Clin. Invest.* **119**, 2702–2713, https://doi.org/10.1172/JCl36435
- 18 Visone, R., Bacalini, M.G., Franco, S.D. et al. (2019) DNA methylation of shelf, shore and open sea CpG positions distinguish high microsatellite instability from low or stable microsatellite status colon cancer stem cells. *Epigenomics-UK* **11**, 587–604, https://doi.org/10.2217/epi-2018-0153
- 19 Williams, C.S., Bernard, J.K., Michelle, D.B. et al. (2015) ERBB4 is over-expressed in human colon cancer and enhances cellular transformation. *Carcinogenesis* **36**, 710–718, https://doi.org/10.1093/carcin/bgv049
- 20 Pernicova, I. and Korbonits, M. (2014) Metformin-mode of action and clinical implications for diabetes and cancer. *Nat. Rev. Endocrinol.* **10**, 143–156, https://doi.org/10.1038/nrendo.2013.256
- 21 Bridges, H.R., Jones, A.J., Pollak, M.N. and Hirst, J. (2014) Effects of metformin and other biguanides on oxidative phosphorylation in mitochondria. *Biochem. J.* 462, 475–487, https://doi.org/10.1042/BJ20140620
- 22 Alvarez-Medina, R., Cayuso, J., Okubo, T., Takada, S. and Marti, E. (2008) Wnt canonical pathway restricts graded Shh/Gli patterning activity through the regulation of Gli3 expression. *Development* **135**, 237–247, https://doi.org/10.1242/dev.012054
- 23 Zhang, H.H. and Guo, X.L. (2016) Combinational strategies of metformin and chemotherapy in cancers. *Cancer Chemother. Pharmacol.* **78**, 13–26, https://doi.org/10.1007/s00280-016-3037-3



- 24 Topol, L., Jiang, X., Choi, H., Garrettbeal, L., Carolan, P.J. and Yang, Y. (2003) Wnt-5a inhibits the canonical Wnt pathway by promoting GSK-3–independent β-catenin degradation. J. Cell Biol. 162, 899–908, https://doi.org/10.1083/jcb.200303158
- 25 Axelrod, J.D., Matsuno, K., Artavanis-Tsakonas, S. and Perrimon, N. (1996) Interaction between Wingless and Notch signaling pathways mediated by dishevelled. *Science* 271, 1826–1832, https://doi.org/10.1126/science.271.5257.1826
- 26 Irshad, S., Bansal, M., Guarnieri, P. et al. (2017) Bone morphogenetic protein and Notch signalling crosstalk in poor-prognosis, mesenchymal-subtype colorectal cancer. J. Pathol. 242, 178–192, https://doi.org/10.1002/path.4891
- 27 Konsavage, W.M., Kyler, S.L., Rennoll, S.A., Ge, J. and Yochum, G.S. (2012) Wnt/β-catenin signaling regulates Yes-associated protein (YAP) gene expression in colorectal carcinoma cells. J. Biol. Chem. 287, 11730–11739, https://doi.org/10.1074/jbc.M111.327767
- 28 Wenqi, W., Zhen-Dong, X., Xu, L. et al. (2015) AMPK modulates Hippo pathway activity to regulate energy homeostasis. Nat. Cell Biol. 17, 490–499
- 29 Karin, M., Cao, Y., Greten, F.R. and Li, Z.W. (2002) NF-kappaB in cancer: from innocent bystander to major culprit. Nat. Rev. Cancer 2, 301–310, https://doi.org/10.1038/nrc780
- 30 Robert, E. and Wagner, E.F. (2011) AP-1: a double-edged sword in tumorigenesis. *Hepatology* 3, 1470–1472
- 31 Vdovikova, S., Gilfillan, S., Wang, S., Dongre, M. and Hurtado, A. (2018) Modulation of gene transcription and epigenetics of colon carcinoma cells by bacterial membrane vesicles. *Sci. Rep-U.K.* **8**, 7434
- 32 Kim, D., Kim, Y. and Kim, Y. (2019) Effects of β-carotene on Expression of Selected MicroRNAs, Histone Acetylation, and DNA Methylation in Colon Cancer Stem Cell. J. Cancer Prevention 24, 224–232, https://doi.org/10.15430/JCP.2019.24.4.224
- 33 Wu, R., Wang, L., Yin, R. et al. (2020) Epigenetics/epigenomics and prevention by curcumin of early stages of inflammatory-driven colon cancer. *Mol. Carcinog.* 59, 227–236, https://doi.org/10.1002/mc.23146
- 34 Wu, D. and Pan, W. (2010) GSK3: a multifaceted kinase in Wnt signaling. Trends Biochem. Sci. 35, 161, https://doi.org/10.1016/j.tibs.2009.10.002
- 35 Bilić, J., Huang, Y.L., Davidson, G. et al. (2007) Wnt Induces LRP6 Signalosomes and Promotes Dishevelled-Dependent LRP6 Phosphorylation. *Science* **316**, 1619, https://doi.org/10.1126/science.1137065
- 36 Gordon, M.D. and Nusse, R. (2006) Wht signaling: multiple pathways, multiple receptors, and multiple transcription factors. J. Biol. Chem. 281, 22429–22433, https://doi.org/10.1074/jbc.R600015200
- 37 Kohn, A.D. and Moon, R.T. (2005) Wht and calcium signaling: beta-catenin-independent pathways. *Cell Calcium* **38**, 439–446, https://doi.org/10.1016/j.ceca.2005.06.022
- 38 Okamoto, M., Udagawa, N., Uehara, S. et al. (2014) Noncanonical Wnt5a enhances Wnt/β-catenin signaling during osteoblastogenesis. Sci. Rep. 4, 4493
- 39 Holmen, S.L., Salic, A., Zylstra, C.R., Kirschner, M.W. and Williams, B.O. (2002) A novel set of Wnt-Frizzled fusion proteins identifies receptor components that activate beta -catenin-dependent signaling. J. Biol. Chem. 277, 34727–34735, https://doi.org/10.1074/jbc.M204989200
- 40 Macdonald, B.T. and He, X. (2012) Frizzled and LRP5/6 receptors for Wnt/β-catenin signaling. *CSH Perspect. Biol.* 4, S107
- 41 Jernigan, K.K., Cselenyi, C.S., Thorne, C.A. et al. (2010) Gbetagamma activates GSK3 to promote LRP6-mediated beta-catenin transcriptional activity. *Sci. Signal* **3**, a37, https://doi.org/10.1126/scisignal.2000647
- 42 van Es, J.H., van Gijn, M.E., Riccio, O. et al. (2005) Notch/gamma-secretase inhibition turns proliferative cells in intestinal crypts and adenomas into goblet cells. *Nature* 435, 959–963, https://doi.org/10.1038/nature03659
- 43 Kwon, C., Cheng, P., King, I.N. et al. (2011) Notch post-translationally regulates beta-catenin protein in stem and progenitor cells. *Nat. Cell Biol.* **13**, 1244–1251, https://doi.org/10.1038/ncb2313
- 44 Jeong, W.J., Yoon, J., Park, J.C. et al. (2012) Ras stabilization through aberrant activation of Wnt/beta-catenin signaling promotes intestinal tumorigenesis. *Sci. Signal* **5**, a30, https://doi.org/10.1126/scisignal.2002242
- 45 Chen, J. (2008) Is Src the key to understanding metastasis and developing new treatments for colon cancer? *Nat. Clin. Pract. Gastroenterol. Hepatol.* **5**, 306
- 46 Garcia-Echeverria, C. and Sellers, W.R. (2008) Drug discovery approaches targeting the PI3K/Akt pathway in cancer. *Oncogene* 27, 5511, https://doi.org/10.1038/onc.2008.246
- 47 Zhao, L. and Vogt, P.K. (2008) Class I PI3K in oncogenic cellular transformation. Oncogene 27, 5486, https://doi.org/10.1038/onc.2008.244
- 48 Rodgers, S.J., Ferguson, D.T., Mitchell, C.A. and Ooms, L.M. Regulation of PI3K effector signalling in cancer by the phosphoinositide phosphatases. *Biosci. Rep.* **37**, R20160432, https://doi.org/10.1042/BSR20160432
- 49 Kjetil, B., Ida, G.D., Hans-Christian, A., Eivind, H. and Maelandsmo, G.M. (2010) Activation of NF-kappaB by extracellular S100A4: analysis of signal transduction mechanisms and identification of target genes. *Int. J. Cancer* **123**, 1301–1310
- 50 Jin, S., Yun, H.J., Jeong, H.Y. et al. (2015) Widdrol, a sesquiterpene isolated from Juniperus chinensis, inhibits angiogenesis by targeting vascular endothelial growth factor receptor 2 signaling. *Oncol. Rep.* **34**, 1178–1184, https://doi.org/10.3892/or.2015.4075
- 51 Trotman, L.C. and Pandolfi, P.P. (2003) PTEN and p53: who will get the upper hand? *Cancer Cell* **3**, 97–99, https://doi.org/10.1016/S1535-6108(03)00022-9
- 52 Whiteside, T.L. (2007) The role of death receptor ligands in shaping tumor microenvironment. *Immunol. Invest.* **36**, 25–46, https://doi.org/10.1080/08820130600991893
- 53 Zhu, Q., Liu, J.Y., Xu, H.W. et al. (2005) Mechanism of counterattack of colorectal cancer cell by Fas/Fas ligand system. *World J. Gastroenterol.* **11**, 6125–6129, https://doi.org/10.3748/wjg.v11.i39.6125
- 54 Pastorino, J.G., Tafani, M. and Farber, J.L. (1999) Tumor necrosis factor induces phosphorylation and translocation of BAD through a phosphatidylinositide-3-OH kinase-dependent pathway. J. Biol. Chem. 274, 19411–19416, https://doi.org/10.1074/jbc.274.27.19411
- 55 Kiyokawa, H., Kineman, R.D., Manova-Todorova, K.O. et al. (1996) Enhanced growth of mice lacking the cyclin-dependent kinase inhibitor function of p27 (Kip1). *Cell* **85**, 721–732, https://doi.org/10.1016/S0092-8674(00)81238-6



- 56 Liang, J. and Slingerland, J.M. (2003) Multiple roles of the PI3K/PKB (Akt) pathway in cell cycle progression. Cell Cycle 2, 339–345, https://doi.org/10.4161/cc.2.4.433
- 57 Martinez-Gac, L., Alvarez, B., Garcia, Z., Marques, M., Arrizabalaga, M. and Carrera, A.C. (2004) Phosphoinositide 3-kinase and Forkhead, a switch for cell division. *Biochem. Soc. Trans.* 32, 360–361, https://doi.org/10.1042/bst0320360
- 58 Sears, R., Nuckolls, F., Haura, E., Taya, Y., Tamai, K. and Nevins, J.R. (2000) Multiple Ras-dependent phosphorylation pathways regulate Myc protein stability. *Genes Dev.* 14, 2501–2514, https://doi.org/10.1101/gad.836800
- 59 Juan, L.X., Sun-Hee, L., Hee, P.M. and Soo Mi, K. (2013) Regulation of YAP through an Akt-dependent process by 3, 3'-diindolylmethane in human colon cancer cells. *Int. J. Oncol.* **43**, 1992–1998
- 60 Wullschleger, S., Loewith, R. and Hall, M.N. (2006) TOR signaling in growth and metabolism. *Cell* **124**, 471–484, https://doi.org/10.1016/j.cell.2006.01.016
- 61 Chen, X., Liao, J., Lu, Y., Duan, X. and Sun, W. (2011) Activation of the PI3K/Akt pathway mediates bone morphogenetic protein 2-induced invasion of pancreatic cancer cells Panc-1. *Pathol. Oncol. Res.* **17**, 257–261, https://doi.org/10.1007/s12253-010-9307-1
- 62 Soung, Y.H., Lee, J.W., Kim, S.Y. et al. (2006) Somatic mutations of the ERBB4 kinase domain in human cancers. Int. J. Cancer **118**, 1426, https://doi.org/10.1002/ijc.21507
- 63 Elrayes, B.F. and Lorusso, P.M. (2004) Targeting the epidermal growth factor receptor. Brit J Cancer 91, 418, https://doi.org/10.1038/sj.bjc.6601921
- 64 Cunningham, D., Humblet, Y., Siena, S. et al. (2016) Cetuximab monotherapy and cetuximab plus irinotecan in irinotecan-refractory metastatic colorectal cancer. J. Evidence-Based Med. 351, 337–345
- 65 Saltz, L.B., Meropol, N.J., Loehrer, Sr., P.J., Needle, M.N., Kopit, J. and Mayer, R.J. (2004) Phase II trial of cetuximab in patients with refractory colorectal cancer that expresses the epidermal growth factor receptor. *J. Clin. Oncol.* **22**, 1201–1208, https://doi.org/10.1200/JC0.2004.10.182
- 66 Yarden, Y. and Sliwkowski, M.X. (2001) Untangling the ErbB signalling network. *Nat. Rev. Mol. Cell Biol.* **2**, 127–137, https://doi.org/10.1038/35052073
- 67 Dong, I.P., Kang, M.S., Oh, S.J. et al. (2007) HER-2/neu overexpression is an independent prognostic factor in colorectal cancer. *Int. J. Colorectal Dis.* 22, 491
- 68 Berg, K.C.G., Eide, P.W., Eilertsen, I.A. et al. (2017) Multi-omics of 34 colorectal cancer cell lines a resource for biomedical studies. *Mol. Cancer* 16, 116, https://doi.org/10.1186/s12943-017-0691-y
- 69 Yonesaka, K., Zejnullahu, K., Okamoto, I. et al. (2011) Activation of ERBB2 signaling causes resistance to the EGFR-directed therapeutic antibody cetuximab. Eur. J. Cancer 47, 86r–99r, https://doi.org/10.1016/S0959-8049(11)72627-2
- 70 Holbro, T., Beerli, R.R., Maurer, F., Koziczak, M., Barbas, C.F. and Hynes, N.E. (2003) The ErbB2/ErbB3 heterodimer functions as an oncogenic unit: ErbB2 requires ErbB3 to drive breast tumor cell proliferation. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 8933–8938, https://doi.org/10.1073/pnas.1537685100
- 71 Guy, P.M., Platko, J.V., Cantley, L.C., Cerione, R.A. and Carraway, III, L.K. (1994) Insect cell-expressed p180erbB3 possesses an impaired tyrosine kinase activity. *Proc. Natl. Acad. Sci. U.S.A.* **91**, 8132–8136, https://doi.org/10.1073/pnas.91.17.8132
- 72 Kim, H., Vijapurkar, U., Hellyer, N., Bravo, D. and Koland, J. (1998) Signal transduction by epidermal growth factor and heregulin via the kinase-deficient ErbB3 protein. *Biochem. J.* **334**, 189–195, https://doi.org/10.1042/bj3340189
- 73 Baiocchi, G., Lopes, A., Coudry, R.A. et al. (2009) ErbB family immunohistochemical expression in colorectal cancer patients with higher risk of recurrence after radical surgery. Int. J. Colorectal Dis. 24, 1059, https://doi.org/10.1007/s00384-009-0702-6
- 74 Chow, N.H., Chan, S.H., Tzai, T.S., Ho, C.L. and Liu, H.S. (2001) Expression Profiles of ErbB Family Receptors and Prognosis in Primary Transitional Cell Carcinoma of the Urinary Bladder. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **7**, 1957–1962
- 75 Kapitanović, S., Radosević, S., Kapitanović, M. et al. (1997) The expression of p185 (HER-2/neu) correlates with the stage of disease and survival in colorectal cancer. *Gastroenterology* **112**, 1103–1113, https://doi.org/10.1016/S0016-5085(97)70120-3
- 76 Carpenter, G. (2003) ErbB-4: mechanism of action and biology. Exp. Cell Res. 284, 66–77, https://doi.org/10.1016/S0014-4827(02)00100-3
- 77 Sahai, E. and Marshall, C.J. (2002) ROCK and Dia have opposing effects on adherens junctions downstream of Rho. *Nat. Cell Biol.* **4**, 408–415, https://doi.org/10.1038/ncb796
- 78 Leve, F. and Morgadodíaz, J.A. (2012) Rho GTPase signaling in the development of colorectal cancer. J. Cell. Biochem. **113**, 2549–2559, https://doi.org/10.1002/jcb.24153
- 79 Olson, M.F., Ashworth, A. and Hall, A. (1995) An essential role for Rho, Rac, and Cdc42 GTPases in cell cycle progression through G1. *Science* **269**, 1270, https://doi.org/10.1126/science.7652575
- 80 Rodrigues, P., Macaya, I., Bazzocco, S. et al. (2014) RHOA inactivation enhances Wnt signaling and promotes colorectal cancer. Nat. Commun. 5, 5458, https://doi.org/10.1038/ncomms6458
- 81 Wang, H., Zhao, G., Liu, X. et al. (2010) Silencing of RhoA and RhoC expression by RNA interference suppresses human colorectal carcinoma growth in vivo. J. Exp. Clin. Cancer Res. 29, 1–7, https://doi.org/10.1186/1756-9966-29-123
- 82 Arango, D., Laiho, P., Kokko, A. et al. (2005) Gene-expression profiling predicts recurrence in Dukes' C colorectal cancer. Gastroenterology 129, 874–884, https://doi.org/10.1053/j.gastro.2005.06.066
- 83 Van de Wetering, M., Sancho, E., Verweij, C. et al. (2002) The beta-catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. *Cell* **111**, 241–250
- Sansom, O.J., Reed, K.R., Hayes, A.J. et al. (2004) Loss of Apc in vivo immediately perturbs Wnt signaling, differentiation, and migration. *Genes Dev.* 18, 1385–1390, https://doi.org/10.1101/gad.287404
- 85 Miele, L. (2006) Notch signaling. Clin Cancer Res. 12, 1074–1079, https://doi.org/10.1158/1078-0432.CCR-05-2570
- 86 Koch, U. and Radtke, F. (2007) Notch and cancer: a double-edged sword. *Cell. Mol. Life Sci.* 64, 2746



- 87 Gopalakrishnan, N., Sivasithamparam, N.D. and Devaraj, H. (2014) Synergistic association of Notch and NFκB signaling and role of Notch signaling in modulating epithelial to mesenchymal transition in colorectal adenocarcinoma. *Biochimie* **107**, 310–318, https://doi.org/10.1016/j.biochi.2014.09.020
- 88 Kopan, R. and Ilagan, M.X.G. (2009) The Canonical Notch Signaling Pathway: Unfolding the Activation Mechanism. Cell 137, 216–233, https://doi.org/10.1016/j.cell.2009.03.045
- Allenspach, E.J., Maillard, I., Aster, J.C. and Pear, W.S. (2002) Notch Signaling in Cancer. *Curr. Mol. Med.* **6**, 466–476
- 90 Brzozowa, M., Wojnicz, R., Kowalczyk-Ziomek, G. and Helewski, K. (2013) The Notch ligand Delta-like 4 (DLL4) as a target in angiogenesis-based cancer therapy? *Contemporary Oncol.* 17, 234–237
- 91 Ç, D., Coşkunsu, D., Kiliçoğlu, Ö, Ergönül, Ö and Irrgang, J.J. (2014) Colocalization of β-catenin with Notch intracellular domain in colon cancer: a possible role of Notch1 signaling in activation of CyclinD1-mediated cell proliferation. *Mol. Cell. Biochem.* **396**, 281–293
- 92 Khazaei, M., Kalantari, E., Saeidi, H. et al. (2015) Gamma-secretase inhibitor does not modulate angiogenesis in colon adenocarcinoma in obese mice. Bratisl. Lek. Listy **116**, 248
- 93 Saif, M.W. and Chu, E. (2010) Biology of Colorectal Cancer. Cancer J. 16, 196–201, https://doi.org/10.1097/PP0.0b013e3181e076af
- 94 Lombardo, Y., Scopelliti, A., Cammareri, P. et al. (2011) Bone Morphogenetic Protein 4 Induces Differentiation of Colorectal Cancer Stem Cells and Increases Their Response to Chemotherapy in Mice. *Gastroenterology* 140, 297–309, https://doi.org/10.1053/j.gastro.2010.10.005
- 95 Schmierer, B. and Hill, C.S. (2007) TGFbeta-SMAD signal transduction: molecular specificity and functional flexibility. Nat. Rev. Mol. Cell Biol. 8, 970–982, https://doi.org/10.1038/nrm2297
- 96 Scoville, D.H., Sato, T., He, X.C. and Li, L. (2008) Current view: intestinal stem cells and signaling. *Gastroenterology* **134**, 849–864, https://doi.org/10.1053/j.gastro.2008.01.079
- 97 Itatani, Y., Kawada, K., Fujishita, T. et al. (2013) Loss of SMAD4 from colorectal cancer cells promotes CCL15 expression to recruit CCR1+ myeloid cells and facilitate liver metastasis. *Gastroenterology* 145, 1064–1075, https://doi.org/10.1053/j.gastro.2013.07.033
- 98 Karagiannis, G.S., Afaloniati, H., Karamanavi, E., Poutahidis, T. and Angelopoulou, K. (2016) BMP pathway suppression is an early event in inflammation-driven colon neoplasmatogenesis of uPA-deficient mice. *Tumour Biol.* 37, 2243–2255, https://doi.org/10.1007/s13277-015-3988-8
- 99 Freeman, T.J., Smith, J.J., Chen, X. et al. (2012) Smad4-mediated signaling inhibits intestinal neoplasia by inhibiting expression of beta-catenin. Gastroenterology 142, 562–571, https://doi.org/10.1053/j.gastro.2011.11.026
- 100 Tian, Q., He, X.C., Hood, L. and Li, L. (2005) Bridging the BMP and Wnt pathways by PI3 kinase/Akt and 14-3-3zeta. *Cell Cycle* 4, 215–216, https://doi.org/10.4161/cc.4.2.1412
- 101 Rubin, L.L. and de Sauvage, F.J. (2006) Targeting the Hedgehog pathway in cancer. *Nat. Rev. Drug Discov.* 5, 1026–1033, https://doi.org/10.1038/nrd2086
- 102 McMahon, A.P., Ingham, P.W. and Tabin, C.J. (2003) Developmental roles and clinical significance of hedgehog signaling. *Curr. Top. Dev. Biol.* 53, 1–114, https://doi.org/10.1016/S0070-2153(03)53002-2
- 103 Lombardo, Y., Scopelliti, A., Cammareri, P. et al. (2011) Bone morphogenetic protein 4 induces differentiation of colorectal cancer stem cells and increases their response to chemotherapy in mice. *Gastroenterology* **140**, 297–309, https://doi.org/10.1053/j.gastro.2010.10.005
- 104 Gerling, M., Büller, N.V., Kirn, L.M. et al. (2016) Stromal Hedgehog signalling is downregulated in colon cancer and its restoration restrains tumour growth. *Nat. Commun.* **7**, 12321, https://doi.org/10.1038/ncomms12321
- 105 Bian, Y.H., Huang, S.H., Yang, L., Ma, X.L., Xie, J.W. and Zhang, H.W. (2007) Sonic hedgehog-Gli1 pathway in colorectal adenocarcinomas. World J Gastroentero 13, 1659–1665, https://doi.org/10.3748/wjg.v13.i11.1659
- 106 Varnat, F., Duquet, A., Malerba, M. et al. (2009) Human colon cancer epithelial cells harbour active HEDGEHOG-GLI signalling that is essential for tumour growth, recurrence, metastasis and stem cell survival and expansion. *EMBO Mol. Med.* 1, 338–351, https://doi.org/10.1002/emmm.200900039
- 107 Monzo, M., Moreno, I., Artells, R. et al. (2006) Sonic hedgehog mRNA expression by real-time quantitative PCR in normal and tumor tissues from colorectal cancer patients. *Cancer Lett.* 233, 123, https://doi.org/10.1016/j.canlet.2005.03.001
- 108 Pan, D. (2007) Hippo signaling in organ size control. Genes Dev. 21, 886-897, https://doi.org/10.1101/gad.1536007
- 109 Zhao, B., Tumaneng, K. and Guan, K.L. (2011) The Hippo pathway in organ size control, tissue regeneration and stem cell self-renewal. *Nat. Cell Biol.* **13**, 877–883, https://doi.org/10.1038/ncb2303
- 110 Johnson, R. and Halder, G. (2014) The two faces of Hippo: targeting the Hippo pathway for regenerative medicine and cancer treatment. *Nat. Rev. Drug Discov.* **13**, 63–79, https://doi.org/10.1038/nrd4161
- 111 Cho, S.Y., Gwak, J.W., Shin, Y.C. et al. (2018) Expression of Hippo pathway genes and their clinical significance in colon adenocarcinoma. *Oncol. Lett.* **15**, 4926–4936
- 112 Heidary Arash, E., Shiban, A., Song, S. and Attisano, L. (2017) MARK4 inhibits Hippo signaling to promote proliferation and migration of breast cancer cells. *EMBO Rep.* **18**, 420, https://doi.org/10.15252/embr.201642455
- 113 Felleybosco, E. and Stahel, R. (2014) Hippo/YAP pathway for targeted therapy. Transl. Lung Cancer Res. 3, 75-83
- 114 Silva, E., Tsatskis, Y., Gardano, L., Tapon, N. and Mcneill, H. (2006) The Tumor-Suppressor Gene fat Controls Tissue Growth Upstream of Expanded in the Hippo Signaling Pathway. *Curr. Biol.* **16**, 2081–2089, https://doi.org/10.1016/j.cub.2006.09.004
- 115 Nussinov, R., Tsai, C.J., Jang, H., Korcsmáros, T. and Csermely, P. (2016) Oncogenic KRAS signaling and YAP1/β-catenin: Similar cell cycle control in tumor initiation. Semin. Cell Dev. Biol. 58, 79–85, https://doi.org/10.1016/j.semcdb.2016.04.001
- 116 Cai, J., Maitra, A., Anders, R.A., Taketo, M.M. and Pan, D. (2015) β-Catenin destruction complex-independent regulation of Hippo–YAP signaling by APC in intestinal tumorigenesis. *Gene Dev* 29, 1493–1506, https://doi.org/10.1101/gad.264515.115
- 117 Sylvester, K.G. and Colnot, S. (2014) Hippo/YAP, beta-catenin, and the cancer cell: a "menage a trois" in hepatoblastoma. *Gastroenterology* **147**, 562–565, https://doi.org/10.1053/j.gastro.2014.07.026



- 118 Varelas, X., Miller, B.R., SONG, S. et al. (2010) The Hippo pathway regulates Wnt/beta-catenin signaling. *Dev. Cell* **18**, 579–591, https://doi.org/10.1016/j.devcel.2010.03.007
- 119 Wei, X., Ye, J., Shang, Y. et al. (2017) Ascl2 activation by YAP1/KLF5 ensures the self-renewability of colon cancer progenitor cells. *Oncotarget* **8**, 109301–109318, https://doi.org/10.18632/oncotarget.22673
- 120 Dawang, Z., Yongyou, Z., Hongtan, W. et al. (2011) Mst1 and Mst2 protein kinases restrain intestinal stem cell proliferation and colonic tumorigenesis by inhibition of Yes-associated protein (Yap) overabundance. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 1312–1320
- 121 Camargo, F.D., Gokhale, S., Johnnidis, J.B. et al. (2007) YAP1 Increases Organ Size and Expands Undifferentiated Progenitor Cells. Curr. Biol. 17, 2054–2060, https://doi.org/10.1016/j.cub.2007.10.039
- 122 Sansal, I. and Sellers, W.R. (2004) The biology and clinical relevance of the PTEN tumor suppressor pathway. J. Clin. Oncol. 22, 2954–2963, https://doi.org/10.1200/JC0.2004.02.141
- 123 Li, Y.K., Xie, Y.J., Wu, D.C., Long, S.L., Tang, S. and Mo, Z.C. (2018) NogoB receptor in relevant carcinoma: Current achievements, challenges and aims (Review). Int. J. Oncol. 53, 1827–1835
- 124 Kalender, A., Selvaraj, A., Kim, S.Y. et al. (2010) Metformin, independent of AMPK, inhibits mTORC1 in a rag GTPase-dependent manner. *Cell Metab.* **11**, 390–401, https://doi.org/10.1016/j.cmet.2010.03.014
- 125 Ben, S.I., Regazzetti, C., Robert, G. et al. (2011) Metformin, independent of AMPK, induces mTOR inhibition and cell-cycle arrest through REDD1. *Cancer Res.* **71**, 4366–4372
- 126 Lei, Y., Yi, Y., Liu, Y. et al. (2017) Metformin targets multiple signaling pathways in cancer. *Chin. J. Cancer* **36**, 289–297, https://doi.org/10.1186/s40880-017-0184-9
- 127 Morales, D.R. and Morris, A.D. (2015) Metformin in cancer treatment and prevention. *Annu. Rev. Med.* 66, 17–29, https://doi.org/10.1146/annurev-med-062613-093128
- 128 Gadducci, A., Biglia, N., Tana, R., Cosio, S. and Gallo, M. (2016) Metformin use and gynecological cancers: A novel treatment option emerging from drug repositioning. *Crit. Rev. Oncol. Hematol.* **105**, 73–83, https://doi.org/10.1016/j.critrevonc.2016.06.006
- 129 Park, E.J., Grabińska, K.A., Guan, Z. and Sessa, W.C. (2016) NgBR is essential for endothelial cell glycosylation and vascular development. *EMBO Rep.* **17**, 167, https://doi.org/10.15252/embr.201540789
- 130 Saunier, E., Antonio, S., Regazzetti, A. et al. Resveratrol reverses the Warburg effect by targeting the pyruvate dehydrogenase complex in colon cancer cells. *Sci Rep-U.K* **7**, 6945, https://doi.org/10.1038/s41598-017-07006-0
- 131 Ghosh, S. and Karin, M. (2002) Missing Pieces in the NF-κB Puzzle. Cell 109, S81–S96, https://doi.org/10.1016/S0092-8674(02)00703-1
- 132 Hanada, T. and Yoshimura, A. (2002) Regulation of cytokine signaling and inflammation. *Cytokine Growth Factor Rev.* **13**, 413–421, https://doi.org/10.1016/S1359-6101(02)00026-6
- 133 Goldring, M., Otero, M.K., Ijiri, K. and Li, Y. (2008) Defining the roles of inflammatory and anabolic cytokines in cartilage metabolism. Ann. Rheum. Dis. 67, 75–82, https://doi.org/10.1136/ard.2008.098764
- 134 Joyce, D., Albanese, C., Steer, J., Fu, M., Bouzahzah, B. and Pestell, R.G. (2001) NF-KB and cell-cycle regulation: the cyclin connection. *Cytokine Growth Factor Rev.* **12**, 73–90
- 135 Egeblad, M. and Werb, Z. (2002) New functions for the matrix metalloproteinases in cancer progression. *Nat. Rev. Cancer* 2, 161–174, https://doi.org/10.1038/nrc745
- 136 Tang, F.Y., Pai, M.H. and Chiang, E.P.I. (2012) Consumption of high-fat diet induces tumor progression and epithelial–mesenchymal transition of colorectal cancer in a mouse xenograft model. *J. Nutr. Biochem.* **23**, 1302–1313, https://doi.org/10.1016/j.jnutbio.2011.07.011
- 137 Tong, J., Shen, Y. et al. (2019) Apigenin inhibits epithelial-mesenchymal transition of human colon cancer cells through NF-KB/Snail signaling pathway. *Biosci. Rep.*, https://doi.org/10.1042/BSR20190452
- 138 Kei, S., Shin, M., Yohko, H. et al. (2009) Constitutive NF-kappaB activation in colorectal carcinoma plays a key role in angiogenesis, promoting tumor growth. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **15**, 2248
- 139 Bates, R.C., Deleo, M.J. and Mercurio, A.M. (2004) The epithelial-mesenchymal transition of colon carcinoma involves expression of IL-8 and CXCR-1-mediated chemotaxis. *Exp. Cell Res.* 299, 315–324, https://doi.org/10.1016/j.yexcr.2004.05.033
- 140 Jing, M., Qiang, W., Teng, F., Jing-Dong Jackie, H. and Ye-Guang, C. (2007) MCP-1 mediates TGF-beta-induced angiogenesis by stimulating vascular smooth muscle cell migration. *Blood* **10**, 987–994
- 141 Richmond, A. (2002) Nf-kappa B, chemokine gene transcription and tumour growth. Nat. Rev. Immunol. 2, 664–674, https://doi.org/10.1038/nri887
- 142 Jana, A., Krett, N.L., Guzman, G. et al. (2017) NFkB is essential for activin-induced colorectal cancer migration via upregulation of PI3K-MDM2 pathway. *Oncotarget* 8, 37377–37393, https://doi.org/10.18632/oncotarget.16343
- 143 McCubrey, J.A., Steelman, L.S., Chappell, W.H. et al. (2007) Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. *Biochim. Biophys. Acta* **1773**, 1263–1284, https://doi.org/10.1016/j.bbamcr.2006.10.001
- 144 Zengenni, L., Youjin, Y., Yutong, G., Rencai, W., Qiulong, H. and Xingyao, X. (2014) Chemical characterization and antitumor activities of polysaccharide extracted from Ganoderma lucidum. *Int. J. Mol. Sci.* **15**, 9103
- 145 Wee, S., Jagani, Z., Xiang, K.X. et al. (2009) PI3K pathway activation mediates resistance to MEK inhibitors in KRAS mutant cancers. *Cancer Res.* **69**, 4286–4293, https://doi.org/10.1158/0008-5472.CAN-08-4765
- 146 Anna, S., Tentler, J.J., Pitts, T.M. et al. (2013) Rational combination of a MEK inhibitor, selumetinib, and the Wnt/calcium pathway modulator, cyclosporin A, in preclinical models of colorectal cancer. *Clin. Cancer Res.* **19**, 4149–4162
- 147 Duong, H.Q., Nemazanyy, I., Rambow, F. et al. (2018) The endosomal protein CEMIP links Wnt signaling to MEK1-ERK1/2 activation in Selumetinib-resistant intestinal organoids. *Cancer Res.* 2017–3149
- 148 Wang, G., Huang, Y. et al. (2019) KRAS-mutant colon cancer cells respond to combined treatment of ABT263 and axitinib. Biosci. Rep. 39, 7434
- 149 Davis, R.J. (2000) Signal transduction by the JNK group of MAP kinases. Cell 103, 239–252, https://doi.org/10.1016/S0092-8674(00)00116-1

14



- 150 Nateri, A.S., Bradley, S.D. and Axel, B. (2005) Interaction of phosphorylated c-Jun with TCF4 regulates intestinal cancer development. *Nature* **437**, 281–285, https://doi.org/10.1038/nature03914
- 151 Mailhot, B., Morlat-Thérias, S., Bussière, P.O. et al. (1999) Amino-terminal phosphorylation of c-Jun regulates stress-induced apoptosis and cellular proliferation. *Nat. Genet.* **21**, 326–329
- 152 el Marjou, F., Janssen, K.J.C.B., Li, M. et al. (2010) Tissue-specific and inducible Cre-mediated recombination in the gut epithelium. *Genesis* **39**, 186–193, https://doi.org/10.1002/gene.20042
- 153 Lu, Y.C., Wang, P., Wang, J., Ma, R. and Lee, S.C. (2019) PCNA and JNK1-Stat3 pathways respectively promotes and inhibits diabetes-associated centrosome amplification by targeting at the ROCK1/14-3-3sigma complex in human colon cancer HCT116 cells. *J. Cell. Physiol.* **234**, 11511–11523, https://doi.org/10.1002/jcp.27813
- 154 Bai, L., Yoon, S.O., King, P.D. and Merchant, J.L. (2004) ZBP-89-induced apoptosis is p53-independent and requires JNK. *Cell Death Differen*. **11**, 663–673
- 155 Staley, B.K. and Irvine, K.D. (2010) Warts and Yorkie Mediate Intestinal Regeneration by Influencing Stem Cell Proliferation. *Current Biol. Cb* 20, 1580–1587, https://doi.org/10.1016/j.cub.2010.07.041
- 156 Sun, G. and Irvine, K.D. (2011) Regulation of Hippo signaling by Jun kinase signaling during compensatory cell proliferation and regeneration, and in neoplastic tumors. *Dev. Biol.* **350**, 139–151, https://doi.org/10.1016/j.ydbio.2010.11.036
- 157 Tomlinson, V., Gudmundsdottir, K., Luong, P., Leung, K.Y., Knebel, A. and Basu, S. (2010) JNK phosphorylates Yes-associated protein (YAP) to regulate apoptosis. *Cell Death. Dis.* **1**, e29, https://doi.org/10.1038/cddis.2010.7
- 158 Gongping, S. and Irvine, K.D. (2013) Ajuba family proteins link JNK to Hippo signaling. Sci. Signal 6, a81
- 159 Geissler, K. (2012) Pathways involved in Drosophila and human cancer development: the Notch, Hedgehog, Wingless, Runt, and Trithorax pathway. *Ann. Hematol.* **91**, 645–669, https://doi.org/10.1007/s00277-012-1435-0
- 160 Lin, G.L. and Hankenson, K.D. (2011) Integration of BMP, Wnt, and notch signaling pathways in osteoblast differentiation. J. Cell. Biochem. 112, 3491–3501, https://doi.org/10.1002/jcb.23287
- 161 Konstantinopoulos, P.A., Karamouzis, M.V. and Papavassiliou, A.G. (2007) Post-translational modifications and regulation of the RAS superfamily of GTPases as anticancer targets. *Nat. Rev. Drug Discov.* **6**, 541–555, https://doi.org/10.1038/nrd2221
- 162 Zhang, L.L., Mu, G., Ding, Q. et al. (2015) Phosphatase and Tensin Homolog (PTEN) Represses Colon Cancer Progression through Inhibiting Paxillin Transcription via PI3K/AKT/NF-κB Pathway. J. Biol. Chem. **290**, 15018–15029, https://doi.org/10.1074/jbc.M115.641407
- 163 Simões, A.E.S., Pereira, D.M., Gomes, S.E. et al. (2015) Aberrant MEK5/ERK5 signalling contributes to human colon cancer progression via NF-KB activation. Cell Death Dis. 6, e1718, https://doi.org/10.1038/cddis.2015.83
- 164 Mazumdar, T., Devecchio, J., Agyeman, A., Shi, T. and Houghton, J.A. (2011) The GLI genes as the molecular switch in disrupting Hedgehog signaling in colon cancer. *Oncotarget* **2**, 638–645, https://doi.org/10.18632/oncotarget.310
- 165 Varnat, F., Siegl-Cachedenier, I., Malerba, M., Gervaz, P. and Altaba, A.R.I. (2010) Loss of WNT-TCF addiction and enhancement of HH-GLI1 signalling define the metastatic transition of human colon carcinomas. *EMBO Mol. Med.* **2**, 440–457, https://doi.org/10.1002/emmm.201000098
- 166 Song, L., Li, Z., Liu, W. and Zhao, M. Crosstalk between Wnt/β-catenin and Hedgehog/Gli signaling pathways in colon cancer and implications for therapy. *Cancer Biol Ther.* 16, 1–7, https://doi.org/10.4161/15384047.2014.972215
- 167 Arimura, S., Matsunaga, A., Kitamura, T., Aoki, K., Aoki, M. and Taketo, M.M. (2009) Reduced Level of Smoothened Suppresses Intestinal Tumorigenesis by Down-Regulation of Wnt Signaling. *Gastroenterology* **137**, 629–638, https://doi.org/10.1053/j.gastro.2009.04.059
- 168 Wall, D.S., Mears, A.J., Mcneill, B. et al. (2009) Progenitor cell proliferation in the retina is dependent on Notch-independent Sonic hedgehog/Hes1 activity. J. Cell Biol. **184**, 101–112, https://doi.org/10.1083/jcb.200805155
- 169 Jonas, F., Theresa, V. and Antonio, G.D.H. (2010) Transcriptional crosstalk between TGF-β and stem cell pathways in tumor cell invasion: role of EMT promoting Smad complexes. *Cell Cycle* **9**, 2363–2374
- 170 Duerr, E.M., Mizukami, Y., Moriichi, K., Gala, M. and Chung, D.C. (2012) Oncogenic KRAS regulates BMP4 expression in colon cancer cell lines. *Am. J. Physiol. Gastrointestinal Liver Physiol.* **302**, G1223–G1230
- 171 van den Brink, G.R., Bleuming, S.A., Hardwick, J.C. et al. (2004) Indian Hedgehog is an antagonist of Wnt signaling in colonic epithelial cell differentiation. *Nat. Genet.* **3**, 277–282, https://doi.org/10.1038/ng1304
- 172 Yang, Z.-h., Dang, Y.-Q. and Ji, G. (2019) Role of epigenetics in transformation of inflammation into colorectal cancer. *World J. Gastroenterol.* 23, 2863–2877, https://doi.org/10.3748/wjg.v25.i23.2863