

Genome Sequences for Levilactobacillus brevis Autochthonous to Commercial Cucumber Fermentations

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ABSTRACT We report the whole-genome sequences, along with annotations, of 11 Levilactobacillus brevis isolates from commercial cucumber fermentations performed in North Carolina ($n = 9$) and Minnesota ($n = 2$), USA.

evilactobacillus brevis is a lactic acid bacterium of relevance in the production of fermented foods, including kefir, pickled vegetables, and sauerkraut, and is also a common cause of spoilage in beer making ([1](#page-1-0)). L. brevis may be of use in the development of starter cultures for pickling to maintain microbial stability due to its ability to utilize xylose and L-citrulline, which might otherwise provide an energy source for the spoilage-associated organism Lentilactobacillus buchneri [\(2\)](#page-1-1).

We present 11 genome sequences and annotations corresponding to L. brevis strains isolated from commercial cucumber fermentations conducted in 2009 and 2010 in North Carolina and Minnesota, respectively ([3](#page-1-2)). The L. brevis isolates were collected on day 3, 7, or 30 of the commercial fermentations at a collection tank depth of 2 or 8 feet from the brine surface. Isolation day and depth are incorporated into the genome sequence identification number. For example, genome sequence 3.8.25 was generated from a single colonial isolate (number 25) plated from a fermentation brine sample collected on day 3 at a depth of 8 feet.

All isolates were obtained from cucumber fermentation brines that had been spiral plated on Lactobacillus de Man-Rogosa-Sharpe (MRS) agar supplemented with 0.0001% cycloheximide solution and incubated at 30°C ([3](#page-1-2)). Anaerobic growth conditions were maintained via the GasPak EZ system (BD, Franklin Lakes, NJ), which maintains an anaerobic atmosphere with \geq 10% carbon dioxide. Isolated colonies were streaked on MRS agar prior to preparation of frozen stocks in MRS broth supplemented with 1.5% glycerol. Pure cultures were transferred to MRS broth from frozen stocks prior to DNA extraction. Cultures were incubated statically at 30°C. DNA extraction was conducted using the Wizard high-molecular-weight (HMW) extraction kit (Promega, Madison, WI). Bacterial isolates were preliminarily identified using the partial sequence of the 16S rRNA gene as described by Pérez-Díaz et al. [\(3\)](#page-1-2). CosmosID (Rockville, MD) prepared libraries for Illumina reads with the Illumina Nextera XT kit and assessed libraries for quality with a Qubit fluorometer (Thermo Fisher Scientific) prior to whole-genome sequencing. Samples were sequenced on a NextSeq 550 platform (Illumina, San Diego, CA), producing paired-end reads with a maximum length of 150 bases. Raw sequence data were trimmed for adapters and low-quality bases using BBDuk [\(https://sourceforge.net/projects/bbmap\)](https://sourceforge.net/projects/bbmap), applying standard parameters (Phred quality, trimq=22; minimum length, minlen=36).

The initial assembly and annotation were performed in PATRIC [\(4\)](#page-1-3). De novo assemblies were performed with Unicycler version 0.4.8 ([5](#page-1-4)) with a minimum contig cutoff value of 300 bp. Quality assessment of assemblies was performed with QUAST version 5.0.2 [\(6](#page-1-5)), SAMtools version 13 [\(7\)](#page-1-6), and Pilon version 1.23 ([8](#page-1-7)). Assembled genomes were annotated in RASTtk [\(9](#page-1-8)). The closest reference genomes were identified by Mash/MinHash Editor Frank J. Stewart, Montana State University

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The authors declare no conflict of interest.

Received 18 January 2022 Accepted 23 March 2022 Published 12 April 2022

TABLE 1 Accession numbers and genome statistics for 11 Levilactobacillus brevis isolates

^aNC, North Carolina, USA; MN, Minnesota, USA.

employing the PATRIC database ([10\)](#page-1-9). Upon submission to GenBank, assemblies were reannotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) [\(11](#page-1-10)). PGAP identified all isolates as belonging to L. brevis. Default parameters for the software were used.

Data availability. Assemblies were submitted to GenBank under BioProject accession number [PRJNA674638.](https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA674638) Strain identification and accession numbers for the genome annotations and Sequence Read Archive (SRA) data are included in [Table 1.](#page-1-11)

ACKNOWLEDGMENTS

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