

New Role for Grb10 Signaling in the Pancreas

Andrew Ward

After surgically removing the canine pancreas, Joseph von Mering and Oskar Minkowski linked the pancreas with diabetes for the first time (1). Their observation led to the use of purified pancreatic extracts to successfully treat diabetic patients some 30 years later, then to the identification and application of subcutaneous insulin delivery, as well as more recent islet transplantation therapies (2). Despite these life-saving advances, diabetes is still a global health care problem. However, there remains much to learn about the mechanisms involved in glucose homeostasis. Insulin receptor signaling is the central molecular pathway and remains the subject of intense research activity. Mouse knockout models have played a pivotal role in bridging the gap between our understanding of the relevant molecular mechanisms and glucose homeostasis in the whole animal (3–5).

In this issue, Zhang et al. (6) report on the pancreas-specific disruption of the *Grb10* signaling adaptor protein gene. This builds on a long tradition of mouse knockout experiments targeting components of the insulin-signaling pathway. Seminally, germline or “global” disruption of the insulin receptor (*Insr*) gene revealed a subtle effect of *Insr* signaling on fetal growth (7) as well as perinatal lethality due to acute diabetic ketoacidosis (8,9). The physiological effects of gene disruption can be difficult to interpret when they involve such a severe phenotype and/or when signaling is altered within multiple insulin-sensitive tissues. To circumvent these problems “conditional” or tissue-specific mouse knockouts can be generated using Cre-lox technology, an approach used to great effect within the glucose homeostasis field. In a series of elegant experiments, the effects of *Insr* ablation have been analyzed separately in several tissues, including skeletal muscle, white adipose tissue (WAT), liver, brain, and pancreas (rev. in 3–5). This work revealed roles for *Insr* signaling in noncanonical insulin-responsive tissues, such as liver, brain, and endocrine pancreas that were more prominent than had previously been appreciated (4). A number of genes acting downstream of the *Insr* have also been the subject of mouse knockout experiments. Broadly speaking, knockouts resulting in impaired signaling, such as those for insulin receptor substrate-1 and -2 and *Akt2*, were associated with insulin resistance or diabetes, whereas knockouts that disrupted an inhibitor of insulin signaling (e.g., *PTP1B*) led to increased insulin sensitivity.

These experiments also revealed myriad subtleties due to, for instance, redundancies between related factors and to the relative importance of individual factors in specific tissues (3–5).

The *Grb10* signaling adaptor functions to inhibit signaling through receptor tyrosine kinases including the *Insr* and insulin-like growth factor type 1 receptor (*Igf1r*) (10). *Grb10* germline knockout mice have elevated *Insr/Igf1r* downstream signaling, at least in skeletal muscle and WAT, without an increase in circulating insulin levels and have an enhanced ability to clear a glucose load from the circulation (11–13) (Table 1). Expression of *Grb10* is widespread during fetal development but more restricted postnatally, including in both canonical (muscle and WAT) and noncanonical (pancreas and brain) insulin-responsive tissues (11,12). *Grb10* is known to inhibit both fetal and placental growth, such that *Grb10* knockout mice are at birth ~30% heavier than their wild-type sibs (14). In adulthood, *Grb10* knockout mice have increased muscle mass and reduced adipose compared with wild types (11,12,15). This “antidiabetic” phenotype of lean body proportions with an enhanced ability to clear blood glucose is very interesting but needs to be better understood.

In the article by Zhang et al. (6), the first report of a conditional *Grb10* knockout, *Grb10* was abolished in pancreas by crossing a floxed *Grb10* allele with transgenic mice that express Cre-recombinase under the control of a pancreas-specific *Pdx1* gene promoter. Global *Grb10* knockout resulted in significantly increased growth of many tissues, including pancreas (11–15). Zhang et al. (6) confirm that *Grb10* is expressed in pancreatic islets of adult mice and show that pancreas-specific *Grb10* knockout resulted in a substantial increase in pancreas tissue weight. This observation is consistent with the established role for *Grb10* as an inhibitor of tissue growth (14,16) and indicates that *Grb10* participates in a local growth control mechanism, consistent with its intracellular signaling function. More work will be required to uncover the relative importance of *Grb10* in regulating pancreas growth during development versus tissue maintenance in adulthood. Under control of the *Pdx1* promoter, Cre-recombinase is expressed from the earliest stages of pancreas development (17), resulting in deletion of the *Grb10* floxed allele in both exocrine and endocrine tissue. Expression of *Grb10* is not readily detected in exocrine pancreas (6,11). However, in a separate study, knockdown of *Grb10* levels in adult mouse pancreas using viral delivery of a short hairpin RNA targeting *Grb10* resulted in increased apoptosis of both endocrine and exocrine tissue (18), supporting a role for *Grb10* in promoting cell survival in both compartments. Discrepancies between the two studies (6,18) will need to be resolved but could be due to differences in the techniques used and the timing of *Grb10* knockout or knockdown.

Importantly, Zhang et al. (6) show that loss of pancreatic *Grb10* resulted in increased β -cell mass, with an associated increase in the number of insulin secretory granules,

From the Department of Biology and Biochemistry and Centre for Regenerative Medicine, University of Bath, Bath, U.K.

Corresponding author: Andrew Ward, bssaw@bath.ac.uk

DOI: 10.2337/db12-1044

© 2012 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. See <http://creativecommons.org/licenses/by-nc-nd/3.0/> for details.

See accompanying original article, p. 3189.

TABLE 1

Phenotypic comparison of adult mice with either global or pancreas-specific *Grb10* knockout alleles illustrates differences in body composition, insulin signaling, and glucose-regulated metabolism, relative to wild-type controls

	Global <i>Grb10</i> knockout	Pancreas-specific <i>Grb10</i> knockout
Body weight	↑	→
Food intake	→	→
Adipose	↓	→
Skeletal muscle	↑	n.d.
Pancreas weight	↑	↑
β-Cell mass	n.d.	↑
Insulin levels	→	↑
Insulin sensitivity	↑	n.d.
Glucose clearance	↑	↑
Insr/Igf1r signaling in skeletal muscle and WAT	↑	n.d.
Insr/Igf1r signaling in islets	n.d.	↑
References	11–13, 15	6

Pancreas weight is increased in both models. Global knockouts have insulin levels appropriate for their body weight, and enhanced glucose clearance is associated with increased insulin sensitivity and enhanced Insr/Igf1r signaling in peripheral tissues. Pancreas-specific knockouts also exhibit enhanced glucose clearance, but in this case it is associated with increased insulin levels and secretion. →, no change; ↑, increased; ↓, decreased; n.d., not determined.

increased insulin secretion, and improved glucose tolerance, but without a significant change in insulin tolerance. These favorable changes in pancreatic β-cell physiology were replicated in mice challenged with a high-fat diet; moreover, pancreas-specific *Grb10* knockout ameliorated the effects of streptozotocin-induced diabetes. This fuels the suggestion that inhibition of *Grb10* might offer a means of increasing β-cell mass in type 1 and type 2 diabetes. In this context, it is interesting to compare the outcomes of pancreas-specific with global *Grb10* knockouts (Table 1). Mice lacking *Grb10* in all tissues had increased lean tissue mass, with no significant change in circulating insulin levels, despite having an enlarged pancreas, and exhibited improvements in both glucose tolerance and insulin sensitivity (11,12). Collectively, the global and tissue-specific knockout experiments indicate a role for *Grb10* in coordinating endocrine pancreas function with that of the canonical insulin-sensitive tissues, suggesting there may be additive therapeutic benefits from targeting *Grb10* function at both sites. However, if therapeutic molecules are to be developed, then a greater understanding is needed of the signaling pathways that *Grb10* acts on in vivo. Zhang et al. (6) provide evidence of increased Insr/Igf1r signaling in islets lacking *Grb10* expression but also point out that this is not necessarily the cause of the increased β-cell mass. The recently established link between *Grb10* and mammalian target of rapamycin signaling is a promising advance (19,20), and data reported by Zhang et al. (6) showing increased mammalian target of rapamycin signaling in *Grb10* knockout islets is an early indication that the link has physiological significance.

The conditional *Grb10* knockout mice have illuminated the pancreatic role of *Grb10* (6) and will undoubtedly continue to aid in unraveling the intricacies of *Grb10* signaling function, allowing key questions to be addressed, including

the following: What are the tissue-specific roles of *Grb10* in the regulation of Insr signaling? What are the relative contributions of altered signaling versus body proportions to the physiological changes seen in *Grb10* knockout mice?

ACKNOWLEDGMENTS

No potential conflicts of interest relevant to this article were reported.

The author thanks Professors Geoff Holman and David Tosh of the University of Bath for helpful comments on the manuscript.

REFERENCES

1. von Mering J, Minkowski O. Diabetes mellitus nach Pankreasextirpation. Arch. Exp. Path. Pharmacol. 1890;26:37 [in German]
2. Bretzel RG. What is the cause of (Type 1) diabetes mellitus – How can we cure this disease? J Mol Med 2002;80:3–4
3. Biddinger SB, Kahn CR. From mice to men: insights into the insulin resistance syndromes. Annu Rev Physiol 2006;68:123–158
4. Okamoto H, Accili D. In vivo mutagenesis of the insulin receptor. J Biol Chem 2003;278:28359–28362
5. Saltiel AR, Kahn CR. Insulin signalling and the regulation of glucose and lipid metabolism. Nature 2001;414:799–806
6. Zhang J, Zhang N, Liu M, et al. Disruption of growth factor receptor-binding protein 10 in the pancreas enhances β-cell proliferation and protects mice from streptozotocin-induced β-cell apoptosis. Diabetes 2012;61:3189–3198
7. Louvi A, Accili D, Efstratiadis A. Growth-promoting interaction of IGF-II with the insulin receptor during mouse embryonic development. Dev Biol 1997;189:33–48
8. Accili D, Drago J, Lee EJ, et al. Early neonatal death in mice homozygous for a null allele of the insulin receptor gene. Nat Genet 1996;12:106–109
9. Joshi RL, Lamothe B, Cordonnier N, et al. Targeted disruption of the insulin receptor gene in the mouse results in neonatal lethality. EMBO J 1996;15:1542–1547
10. Holt LJ, Siddle K. *Grb10* and *Grb14*: enigmatic regulators of insulin action—and more? Biochem J 2005;388:393–406
11. Smith FM, Holt LJ, Garfield AS, et al. Mice with a disruption of the imprinted *Grb10* gene exhibit altered body composition, glucose homeostasis, and insulin signaling during postnatal life. Mol Cell Biol 2007;27:5871–5886
12. Wang L, Balas B, Christ-Roberts CY, et al. Peripheral disruption of the *Grb10* gene enhances insulin signaling and sensitivity in vivo. Mol Cell Biol 2007;27:6497–6505
13. Holt LJ, Lyons RJ, Ryan AS, et al. Dual ablation of *Grb10* and *Grb14* in mice reveals their combined role in regulation of insulin signaling and glucose homeostasis. Mol Endocrinol 2009;23:1406–1414
14. Charalambous M, Smith FM, Bennett WR, Crew TE, Mackenzie F, Ward A. Disruption of the imprinted *Grb10* gene leads to disproportionate overgrowth by an *Igf2*-independent mechanism. Proc Natl Acad Sci USA 2003;100:8292–8297
15. Holt LJ, Turner N, Mokbel N, et al. *Grb10* regulates the development of fiber number in skeletal muscle. FASEB J 2012;26:3658–3669
16. Shiura H, Nakamura K, Hikichi T, et al. Paternal deletion of *Meg1/Grb10* DMR causes maternalization of the *Meg1/Grb10* cluster in mouse proximal Chromosome 11 leading to severe pre- and postnatal growth retardation. Hum Mol Genet 2009;18:1424–1438
17. Herrera PL. Adult insulin- and glucagon-producing cells differentiate from two independent cell lineages. Development 2000;127:2317–2322
18. Doiron B, Hu W, Norton L, DeFronzo RA. Lentivirus shRNA *Grb10* targeting the pancreas induces apoptosis and improved glucose tolerance due to decreased plasma glucagon levels. Diabetologia 2012;55:719–728
19. Hsu PP, Kang SA, Rameseder J, et al. The mTOR-regulated phosphoproteome reveals a mechanism of mTORC1-mediated inhibition of growth factor signaling. Science 2011;332:1317–1322
20. Yu Y, Yoon SO, Poullogiannis G, et al. Phosphoproteomic analysis identifies *Grb10* as an mTORC1 substrate that negatively regulates insulin signaling. Science 2011;332:1322–1326