

Supporting Information

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Oncogenic Roles of Laminin Subunit Gamma-2 in Intrahepatic Cholangiocarcinoma via Promoting EGFR Translation

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Supplementary data

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Supplementary Tables

Table S1. Summary of Datasets and Clinical specimens used in this study.

Cohorts	Source	Cases	Datasets or Specimens
Cohort 1	GSE76297	91 iCCA cases; 62 HCC cases.	mRNA array (paired tumor & non-tumor tissues)
Cohort 2	TCGA	36 CCA cases (iCCA, n=31)	RNA sequencing (tumor, n=36; non-tumor, n=9)
Cohort 3	GSE26566	104 CCA cases (iCCA, n=68)	mRNA array (tumor, n=104; non-tumor, n=59; normal bile duct, n=6)
Cohort 4	Shandong cancer hospital	33 iCCA cases; 20 HCC cases.	FFPE tissues of iCCA cases, n=33; FFPE tissues of HCC cases, n=20.
Cohort 5	Ref (<i>Cancer Cell</i> , 2022.)	262 iCCA cases	RNA sequencing (tumor, n=255); Proteome (tumor, n=214).

Table S2. Clinical Characteristics of Cohort 4.

Clinical Variable		iCCA, n=33	HCC, n=20	p-value
Gender	Female	10	1	0.04 ^a
	Male	21	17	
	Missing value	2	2	
Age-year	Median (range)	59(43-77)	53(31-72)	0.02 ^b
ALT (U/L)	Normal (≤ 55)	23	16	0.29 ^a
	Abnormal (> 55)	8	2	
	Missing value	2	2	
AST (U/L)	Normal (≤ 48)	21	16	0.17 ^a
	Abnormal (> 48)	10	2	
	Missing value	2	2	
Albumin (g/L)	Normal (≥ 35)	24	18	0.04 ^a
	Abnormal (< 35)	7	0	
	Missing value	2	2	
AFP (ng/ml)	Normal (≤ 20)	17	10	0.32 ^a
	Abnormal (> 20)	6	8	
	Missing value	10	2	
CA19-9 (U/ml)	Normal (≤ 40)	7	/	/
	Abnormal (> 40)	20	/	
	Missing value	6	/	
CEA (ng/ml)	Normal (≤ 5)	13	/	/
	Abnormal (> 5)	13	/	
	Missing value	7	/	
BCLC stage	0-A	/	12	/
	B-D	/	6	
	Missing value	/	2	
TNM Stage	I-II	10	12	0.07 ^a
	III-IV	18	6	
	Missing value	5	2	
Multi-nodules	No	0	16	$< 0.001^a$
	Yes	7	2	
	Missing value	26	2	
Tumor size	≤ 3 cm	4	9	0.04 ^a
	> 3 cm	20	9	
	Missing value	9	2	

a: Fisher's exact test; b: Unpaired t-test

Table S3. Clinical Characteristics of LAMC2^{high} and LAMC2^{low} group in Cohort 4.

Clinical Variable	IHC staining score			LAMC2 amplification		
	LAMC2 ^{high} Score ≥ 6 n=24	LAMC2 ^{low} Score < 6 n=9	p-value	LAMC2 ^{high} Copies ≥ 4 n=5	LAMC2 ^{low} Copies < 4 n=14	p-value
Gender						
Female	7	3	1.00 ^a	2	5	1.00 ^a
Male	15	6		3	9	
Missing value	2	0		0	0	
Age-year						
Median (range)	59 (43-77)	62 (45-69)	0.91 ^b	53 (43-72)	64 (45-77)	0.14 ^b
ALT (U/L)						
Normal (≤ 55)	16	7	1.00 ^a	4	10	1.00 ^a
Abnormal (>55)	6	2		1	4	
Missing value	2	0		0	0	
Albumin (g/L)						
Normal (≥ 35)	17	7	1.00 ^a	3	12	0.27 ^a
Abnormal (<35)	5	2		2	2	
Missing value	2	0		0	0	
AFP (ng/ml)						
Normal (≤ 20)	11	6	0.14 ^a	3	6	0.50 ^a
Abnormal (>20)	6	0		0	4	
Missing value	7	3		2	4	
CA19-9 (U/ml)						
Normal (≤ 40)	4	3	0.65 ^a	0	5	0.28 ^a
Abnormal (>40)	14	6		4	9	
Missing value	6	0		1	0	
CEA (ng/ml)						
Normal (≤ 5)	11	2	0.38 ^a	1	7	0.28 ^a
Abnormal (>5)	8	5		4	4	
Missing value	5	2		0	3	
TNM Stage						
I-II	7	3	1.00 ^a	0	4	0.52 ^a
III-IV	13	5		4	9	
Missing value	4	1		1	1	
Tumor size (cm)						
≤ 3	3	0	0.53 ^a	0	1	1.00 ^a
>3	14	7		5	10	
Missing value	7	2		0	3	
Overall survival-month						
Median	16.2	29.8	0.15 ^c	9.2	28.2	0.14 ^c
Range	1.8-36.9	3.5-35		3.2-18.4	1.8-35	

a: Fisher's exact test; b: Unpaired t-test; c: Log-rank test

Table S4. Clinical Characteristics of LAMC2^{high} and LAMC2^{low} group in Cohort 5.

Clinical Variable	LAMC2^{high} n=107	LAMC2^{low} n=107	p-value
Gender			
Female	47	47	1.00 ^a
Male	60	60	
Age-year			
Median (range)	65 (39-86)	60 (28-83)	0.05 ^b
ALT (U/L)			
Normal (≤ 55)	90	96	0.31 ^a
Abnormal (> 55)	17	11	
Albumin (g/L)			
Normal (≥ 35)	103	106	0.37 ^a
Abnormal (< 35)	4	1	
AFP (ng/ml)			
Normal (≤ 20)	100	92	0.11 ^a
Abnormal (> 20)	7	15	
CA19-9 (U/ml)			
Normal (≤ 40)	32	66	$< 0.001^a$
Abnormal (> 40)	75	41	
CEA (ng/ml)			
Normal (≤ 5)	70	94	$< 0.001^a$
Abnormal (> 5)	37	13	
TNM Stage			
I-II	64	75	0.15 ^a
III-IV	43	32	
Tumor size (cm)			
≤ 3	16	16	1.00 ^a
> 3	91	91	
Intrahepatic metastasis			
No	65	78	0.08 ^a
Yes	42	29	
Distal metastasis			
No	100	104	0.33 ^a
Yes	7	3	
Overall survival-month			$< 0.001^c$
Median	16.4	23.9	
Range	1.4-52.7	2.4-60.2	

a: Fisher's exact test; b: Unpaired t-test; c: Log-rank test

Table S5. Primers and oligoes used in this study.

Primers	Sequence (5'-3')
TB Green real-time PCR	
LAMC2	F: GACAACTGGTAATGGATTCCGC R: TTCTCTGTGCCGGTAAAAGCC
EGFR	F: TTGCCGCAAAGTGTGTAACG R: GTCACCCCTAAATGCCACCG
18S	F: GACTCAACACGGGAAACCTC R: AGCATGCCAGAGTCTCGTTC
LAMC2-mus	F: CAGACACGGGAGATTGCTACT R: CCACGTTCCCCAAAGGGAT
18S-mus	F: GCCGCTAGAGGTGAAATTCTT R: CGTCTTCGAACCTCCGACT
FOS	F: CACTCCAAGCGGAGACAGAC R: AGGTCATCAGGGATCTTGCAG
siRNA target sequence	
siLAMC2 #1	GGUUCUCUUAGUGCUCGAU
siLAMC2 #2	GCAGAAUACAGUGUCCAUA
siBiP #1	GCGUCGGCGUGUUAAGAA
siBiP #2	CCAUGCAGUUGUUACUGUA
siPERK #1	GGAUAGUGACGAAAUGGAA
siPERK #2	GAUACAAGUUUAUCAUCA
Primers for vector construction	
pcDNA3.0-LAMC2-HA	F: CGGGATCCGCCACCATGCCTGCGCTCTGGCTG R: CCGCTCGAGCTGTTGCTCAAGAGCCTGGG
pcDNA3.0-N-LAMC2-HA	F: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC R: GACGTCGTATGGGTACTCGAGGCTGAATGCTCCATGCTCACAG F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-C-LAMC2-HA	R1: GCAAGCTGGACAGACTTCCCTCCTGGAGGTGG F2: GAGGGAAGTCTGTCCAGCTTGCTATAATCAAGTGAAGA R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGG F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-LAMC2 ^{ΔDv} -HA	R1: AGAGCTGCGGACTTCCCTCCTGGAGGTGGC F2: AGGGAAGTCCGCAGCTCTGCAGAATACAGTG R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-LAMC2 ^{ΔDiv} -HA	R1: CCAAACAGGACATATACAGCAGCTGGCTGAATGCCC F2: TGTATATGTCTGTTGGGTACAAGGGGCAATTC R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-LAMC2 ^{ΔDiv} -HA	R1: ATTATAGCAAGCTGGACACTGTTCAACCCAGGGTG F2: CAGTGTCAGCTTGCTATAATCAAGTGAAGATTGAG R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-EGFR-HA	R1: ATTATAGCAAGCTGGACACTGTTCAACCCAGGGTG F2: CAGTGTCAGCTTGCTATAATCAAGTGAAGATTGAG R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGC
pcDNA3.0-BiP-HA	F: TGGTACCGAGCTCGGATCCGCCACCATGCGACCTCCGGGACG R: GACGTCGTATGGGTACTCGAGTGTCCAATAAATCACTGCTTTGTGGC
pcDNA3.0-ERdj1-HA	F: CGGGATCCGCCACCATGAAGCTCTCCCTGGTGGC R: CCCTCGAGCTACAACATCATCTTTAGCGTAGTCTGGGACGTCG
pcDNA3.0-ERdj2-HA	F: CGGGATCCGCCACCATGACGGCTCCTTGCTCCC R: CCGCTCGAGGCTTTTAGCTTGTCTTTCTTTTGACC
p3xflag-cmv-14-EGFR	F: CGGGATCCGCCACCATGGCCGGGCAGCAGTT R: CCGCTCGAGGTCATCATCTTCTTCTTCTCCTCTTCTTC
p3xflag-cmv-14-EGFR-extra	F: CCAAGCTTGCCACCATGCGACCTCCGGGAC R: GCTCTAGATGCTCCAATAAATCACTGCTTTGTGG
p3xflag-cmv-14-EGFR-cyto	F: CCAAGCTTGCCACCATGCGAAGGCGCCACATCGTTC R: GCTCTAGATGCTCCAATAAATCACTGCTTTGTGG

p3xflag-cmv-14-EGFR ^{13Q}	F1: TGAACCGTCAGAATTAAGCTTGCCACCATGCGACCCTCCGGGAC R1: AAGCTGTATTTGCCCTCGGGGTTACATC F2: CCCCAGAGGGCAAATACAGCTTTGGTGCCACCTG R2: AGACCTGGCCCAGTGCATCCGTAGGTG F3: ATGCACTGGGCCAGGTCTTGAAGGCTGTCC R3: CCGGGATCCTCTAGATGCTCCAATAAATTCATGCTTTGTGG
pcDNA3.0-BiP-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: CTTTGTAGTCTTCTGCTGTATCCTCTTCACCACTG F2: CAGCAGAAGACTACAAAGACCATGACGGTGAT R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-BiP-Δ19-124-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: GAATGTATGTTTGGCCCCGCGCCG F2: CGGGCAAACCATACTCAAGTTGATATTGGAGGTGG R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-BiP-ΔNBD-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: GCACAGCTCTATTGTCACTTTTCTTTCAACCACCTGAACGG F2: GAAAACTGACAATAGAGCTGTGCAGAAACTCC R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-BiP-Δ281-419-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: AGGGGACATTTCTGACATCTTTGCCCCGTC F2: GTCAGGAAATGTCCCTTACACTTGGTATTGAAACTG R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-BiP-ΔSBD-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: CTATCTCAAATACATCAAGCAGTACCAGGTCACC F2: CCTGGTACTGCTTGATGTATTGAGATAGATGTGAATGGTATTCTTCGAGTG R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-BiP-Δ501-650-flag	F1: AAGCTTGGTACCGAGCTCGGATCCGCCACCATGAAGCTCTCCCTGGTGCC R1: TTGTAGTCGGTGACTTCAATCTGTGGGACCC F2: CACAGATTGAAGTACCGACTACAAAGACCATGACGGTGATT R2: CCCTCTAGATGCATGCTCGAGCTACAACCTCATCTTTCTTGTGCATCGTCATCC
pcDNA3.0-LAMC2-flag-HA	F: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGCT R: GACGTCGTATGGGTACTCGAGCTTGTGCATCGTCATCCTTGTAGTCGAT
pcDNA3.0-LAMC2-V-flag-HA	F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGCT R1: CTTGTGCATCGTCATCCTTGTAGTCGACTTCCCTCCTGGAGGTGGC F2: GACTACAAGGATGACGATGACAAGTGTGATTGCAATGGGAAGTCCAGG R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT
pcDNA3.0-LAMC2-III-flag-HA	F1: TGGTACCGAGCTCGGATCCGCCACCATGCCTGCGCTCTGGCT R1: CTTGTGCATCGTCATCCTTGTAGTCTGGCTTGCAGCTGCGGG F2: GACTACAAGGATGACGATGACAAGTGTCCCTGTCATAACGGGTTTCA R2: GACGTCGTATGGGTACTCGAGCTGTTGCTCAAGAGCCTGGGTAT
pLKO.1-shLAMC2 #1	F: CCGGGCCCTGCAATTGTAACCTCCAACCTCGAGTTGGAGTTACAATTGCAGGGCTTTTTG R: AATTCAAAAAGCCCTGCAATTGTAACCTCCAACCTCGAGTTGGAGTTACAATTGCAGGGC
pLKO.1-shLAMC2 #2	F: CCGGGCTCACCAAGACTTACACATTCTCGAGAATGTGTAAGTCTTGGTGAGCTTTTTG R: AATTCAAAAAGCTCACCAAGACTTACACATTCTCGAGAATGTGTAAGTCTTGGTGAGC
pLKO.1-sh-mLAMC2 #1	F: CCGGGCTACGTACGGAGAATATAGTCTCGAGACTATATTCTCCGTACGTAGCTTTTTG R: AATTCAAAAAGCTACGTACGGAGAATATAGTCTCGAGACTATATTCTCCGTACGTAGCTTTTTG
pLKO.1-sh-mLAMC2 #2	F: CCGGGCTCAACTGCAATGACAATACTCGAGTATTGTCATTGCAGTTGAGGCTTTTTG R: AATTCAAAAAGCCCTCAACTGCAATGACAATACTCGAGTATTGTCATTGCAGTTGAGGCTTTTTG
pT3-EF1α-myr-AKT-sh-mLAMC2 #1	F1: CCTTAATTAAGAGGGCCTATTTCCCATGATTCTT R1: GCTCTAGAATGAATACTGCCATTTGTCTCGAGGTC F2: AGCAGGCAGAAGTATGCAAAGC R2: GCTCTAGACAATTCCCACTCCTTTCAAGACCTAGAAG
pT3-EF1α-myr-AKT-sh-mLAMC2 #2	F1: CCTTAATTAAGAGGGCCTATTTCCCATGATTCTT R1: GCTCTAGAATGAATACTGCCATTTGTCTCGAGGTC F2: AGCAGGCAGAAGTATGCAAAGC R2: GCTCTAGACAATTCCCACTCCTTTCAAGACCTAGAAG F: GTACAAAAAGCAGGCACGCCACATGCCTGCGCTCTGGC

pT3-EF1 α -C-LAMC2	R: CCACAAC TAGAATGCATCACTGTTGCTCAAGAGCCTGG
pT3-EF1 α -N-LAMC2	F: GTACAAAAAAGCAGGCACGCCACATGCCTGCGCTCTGGC R: CCACAAC TAGAATGCATCAGCTGAATGCTCCATGCTCA
pT3-EF1 α -EGFR ^{L858R}	F: GTACAAAAAAGCAGGCACATGCGACCCTCCGGGAC R: CCACAAC TAGAATGCATCATGCTCCAATAAATTCAGCTTTGTG

Table S6. The detailed usage of plasmids for HDTV mouse models.

Groups	Plasmids (μg)
pT3-EF1α-myr-AKT, pT3-EF1α-YapS127A, pCMV/SB	20, 30, 2
pT3-EF1α-myr-AKT, pT3-EF1α-NICD, pCMV/SB	4, 20, 1
pT3-EF1α-Myc, pCMV/SB	4, 0.16
pT3-EF1α-myr-AKT, NRasV12/pT2-CAGGS, pCMV/SB	4, 4, 0.32
pT3-EF1α-myr-AKT-shCtrl, pT3-EF1α-YapS127A, pCMV/SB	20, 30, 2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pCMV/SB	20, 30, 2
pT3-EF1α-myr-AKT-shLAMC2#m2, pT3-EF1α-YapS127A, pCMV/SB	20, 30, 2
pT3-EF1α-myr-AKT-shCtrl, pT3-EF1α-NICD, pCMV/SB	4, 20, 1
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-NICD, pCMV/SB	4, 20, 1
pT3-EF1α-myr-AKT-shLAMC2#m2, pT3-EF1α-NICD, pCMV/SB	4, 20, 1
pT3-EF1α-myr-AKT-shCtrl, pT3-EF1α-YapS127A, pT3-EF1α-CT, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-CT, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-LAMC2, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-EGFR ^{L858R} , pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shCtrl, pT3-EF1α-YapS127A, pT3-EF1α-CT, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-CT, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-C-LAMC2, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT-shLAMC2#m1, pT3-EF1α-YapS127A, pT3-EF1α-N-LAMC2, pCMV/SB	20, 30, 30, 3.2
pT3-EF1α-myr-AKT, pT3-EF1α-YapS127A, pT3-EF1α-CT, pCMV/SB	6.7, 10, 20, 1.5
pT3-EF1α-myr-AKT, pT3-EF1α-YapS127A, pT3-EF1α-LAMC2, pCMV/SB	6.7, 10, 20, 1.5

Supplementary Figure legends:

Figure S1. A high expression of LAMC2 in iCCA tumor cells. (A) Principal component analysis of iCCA tumors and non-tumors, HCC tumors and non-tumors in Cohort 1. (B) Single-cell RNA-sequencing results of LAMC2 in major cell types and tumor cell types from liver cancer. T, tumor; NT, non-tumor.

Figure S2. Knockdown LAMC2 blocked iCCA cell proliferation, colony formation and migration. (A, B) LAMC2 shRNA knockdown efficiency in protein level (A) and mRNA level (B) in RBE and HUCCT1 cells. (C, D) Relative cell viability (C) and colony formation (D) were examined in RBE and HUCCT1 cells infected with shCtrl and shLAMC2 lentivirus. (E, F) Cell migration was examined in RBE (E) and HUCCT1 (F) cells infected with shCtrl and shLAMC2 lentivirus by wounding healing assay. The remaining wound was measured at the indicated time. (G) Mouse LAMC2 shRNA knockdown efficiency in mRNA level was detected in mouse primary hepatocyte H2.35 cell line. (C, E, F) Two-way ANOVA was used. (D) Student's t-test was used.

Figure S3. ~150kD full-length LAMC2 was the major form in iCCA cells. (A) LAMC2 expression was detected in cell lysate and conditioned medium of RBE and HUCCT1 cells. (B) LAMC2 major form detection. Top: schematic diagram of LAMC2 vector. Bottom: RBE and HUCCT1 cells were transfected with indicated vector LAMC2-flag-HA, collected cell lysate and conditioned medium for western blot analysis. (C) Schematic diagrams of LAMC2 protein and reported cleaved LAMC2 fragments. (D) LAMC2 major form detection. Top: schematic diagram of LAMC2 vectors. Bottom: RBE and HUCCT1 cells were transfected with indicated vectors LAMC2-V-flag-HA or LAMC2-III-flag-HA, collected cell lysate and conditioned medium for western blot analysis. (E) LAMC2 expression detection in cell lysate being collected with scraping method or trypsin method.

Figure S4. Secreted extracellular LAMC2 had no effect on iCCA cell proliferation and colony formation. (A) RBE cells transfected with siLAMC2 + Ctrl vector (indicating LAMC2-low status), or siCtrl + LAMC2-flag-HA vector (indicating LAMC2-high status). Conditioned medium was collected according to the detailed methods in our manuscript. LAMC2 expression in cell lysate and conditioned medium were detected by western blot. The Coomassie blue staining indicated the equal loading of conditioned medium. (B) Relative cell viability and colony formation were examined in LAMC2-high and LAMC2-low RBE cells. (C) RBE cells were incubated with LAMC2-low or LAMC2-high conditioned medium, and then relative cell viability and colony formation were examined. Two-way ANOVA analysis was used for cell viability assay. Student's t-test were used for colony formation assay. NS, not significant.

Figure S5. Mass spectrometry results in RBE cells with LAMC2 silencing. (A) The flow chart of sample preparation for mass spectrometry. (B) Western blot validation of LAMC2-silencing in RBE cells. (C) GSEA analysis with significantly altered proteins ($P < 0.05$) between siCtrl and siLAMC2 samples. Among the top 20 enriched signatures, there were two EGF-related signatures, whose enrichment plotted images were shown. (D) EGFR protein intensities in mass spectrometry data were compared between siCtrl RBE cells and siLAMC2 RBE cells. The cluster analysis of siCtrl and siLAMC2 RBE cells based on 6 molecules in an EGF/EGFR signaling gene set.

Figure S6. Effects of EGF/EGFR on LAMC2 expression and effects of secreted LAMC2 on EGFR signaling. (A) Spearman correlation analysis of LAMC2 protein and EGFR protein in iCCA tumors from Cohort 5. (B) LAMC2 mRNA expression was detected in RBE and HUCCT1 cells transfected with EGFR-flag. (C) RBE and HUCCT1 cells starved 12h and stimulated with or without EGF (100 ng/ml) for 24h. LAMC2 and FOS mRNA expression were detected. (D) LAMC2-low CM and LAMC2-high CM were collected from RBE cells transfected with LAMC2 siRNA + Ctrl vector or control siRNA + LAMC2-flag-HA vector. LAMC2 protein was detected in cell lysate and CM by western blot (left). The Coomassie blue staining indicated the equal loading of CM (right). CM, conditioned medium. (E) RBE cells were incubated with LAMC2-low CM or LAMC2-high CM overnight, then stimulated with or without EGF (100 ng/ml) for 30min. Indicated proteins were detected by western blot analysis.

Figure S7. LAMC2 did not affect EGFR mRNA expression and its protein stability. (A) EGFR mRNA expression was detected in RBE and HUCCT1 cells transfected with LAMC2 siRNAs. (B) EGFR mRNA expression was detected in RBE and HUCCT1 cells transfected with LAMC2-HA. (C) After knocking down LAMC2 with siRNAs, RBE and HUCCT1 cells were treated with cycloheximide (CHX) (20 μ g/ml) at indicated intervals and protein stability of EGFR was detected by western blot. (D) HUCCT1 cells were co-transfected with LAMC2-HA and EGFR-flag, then treated with CHX at indicated intervals. Cell lysates were collected, and EGFR was detected by western blot. (E) After knocking down LAMC2 with shRNAs, HUCCT1 cells were treated with CHX (20 μ g/ml) at indicated intervals and protein stability of EGFR was detected by western blot.

Figure S8. LAMC2 interacted with an undersized EGFR in several non-iCCA cells and Boncat assay was performed. (A) LAMC2 IHC staining results from 21 cancer types in Human Protein Atlas database. (B) Endogenous IP assay with anti-LAMC2 antibody in five different non-iCCA cells. (C) Boncat assay in five different non-iCCA cell lines with LAMC2 silencing. L-

AHA were applied to treat cells for 2 hours.

Figure S9. LAMC2 did not regulate the degradation of nascent EGFR. (A) RBE and HUCCT1 cells were treated with or without tunicamycin (0.5 µg/ml) for 12h, and followed by CHX (20 µg/ml), MG132 (20 µM) or CB-5083 (2 µM) treatment for 12h. The indicated proteins were detected by western blot. (B) RBE and HUCCT1 cells were transfected with EGFR^{13Q}-flag. 36 hours after transfection, cells were treated with CHX (20 µg/ml), Bafilomycin A1 (100 nM) or CB-5083 (2 µM) for 12h. Cell lysates were collected for western blot detection. (C) RBE and HUCCT1 cells were transfected with EGFR-flag. 36 hours after transfection, cells were treated with CHX (20 µg/ml), Bafilomycin A1 (100 nM) or CB-5083 (2 µM) for 12h. Cell lysates were collected for western blot detection. (D) HUCCT1 cells were transfected with EGFR^{13Q}-flag vector and infected with shLAMC2 virus. 36 hours later, cells were treated with CHX (20 µg/ml) at indicated intervals and protein stability of EGFR^{13Q}-flag was detected by western blot. (E) HUCCT1 cells were co-transfected with LAMC2-HA and EGFR^{13Q}-flag. 36 hours later, cells were treated with CHX (20 µg/ml) at indicated intervals and protein stability of EGFR^{13Q}-flag was detected by western blot.

Figure S10. EGFR extracellular domain interacted with BiP. 293T, RBE and HUCCT1 cells were co-transfected with BiP-HA and different EGFR-flag vectors treated with tunicamycin (0.5 µg/ml). IP was performed with anti-flag beads.

Figure S11. LAMC2 C-terminus together with BiP SBD were important for regulating EGFR translation. (A) Boncat assay was performed in RBE and HUCCT1 cells co-transfected with BiP-flag/EGFR^{13Q}-flag and an intact LAMC2, or N-LAMC2 (disabling the pocket formation). Cells were treated with L-AHA for 12 hours. (B) Boncat assay was performed in RBE and HUCCT1 cells co-transfected with LAMC2-HA /EGFR^{13Q}-flag and an intact BiP, or BiP^{ΔSBD} and BiP^{Δ501-650} (disabling the pocket formation). Cells were treated with L-AHA for 12 hours.

Figure S12. High LAMC2 level was related to poor prognosis in iCCA patients. (A) LAMC2 level in iCCA tumors of Cohort 5 and a median cut-off of LAMC2. (B) Kaplan-Meier survival analysis of iCCA patients in Cohort 5 based on a tertial cut-off of LAMC2. (C) Kaplan-Meier survival analysis of iCCA patients in Cohort 5 based on a quartile cut-off of LAMC2.

Figure S13. BiP promoted EGFR protein translation via its NBD region and ERdj proteins. (A)

Boncat assay in cells co-transfected with EGFR-flag and an intact BiP, or BiP^{ΔNBD}. **(B)** Boncat assay in cells co-transfected with EGFR-flag and BiP-flag, and ERdj1-HA. **(C)** Boncat assay in cells co-transfected with EGFR-flag and BiP-flag, and ERdj2-HA. **(D)** Boncat assay in cells transfected with EGFR-flag with or without silencing BiP, together with silencing PERK. L-AHA were applied to treat cells for 6 hours.

Figure S14. BiP expression, survival analysis and relationship with EGFR signaling activation.

(A) HSPA5 (gene name for BiP) expression levels in Cohorts 1-3. Unpaired Student's t-test was used for Cohort 1 and Cohort 3. Non-parametric t-test was used for Cohort 2. NS, not significant; T, tumor; NT, non-tumor. **(B)** Kaplan-Meier survival analysis of iCCA patients from Cohort 5 based on BiP protein level (Left panel, the median cut-off; Middle panel, the tertial cut-off; Right panel, the quartile cut-off). **(C)** The enrichment analysis of HSPA5-high patients in EGFR signaling activation and non-activation groups subclassified by an EGF/EGFR signaling gene set in Cohort 1 and Cohort 5. The cut off of HSPA5-high and HSPA5-low was based on the median cut-off of HSPA5 in tumor tissues from each cohort.

Supplementary Figures

Figure S1

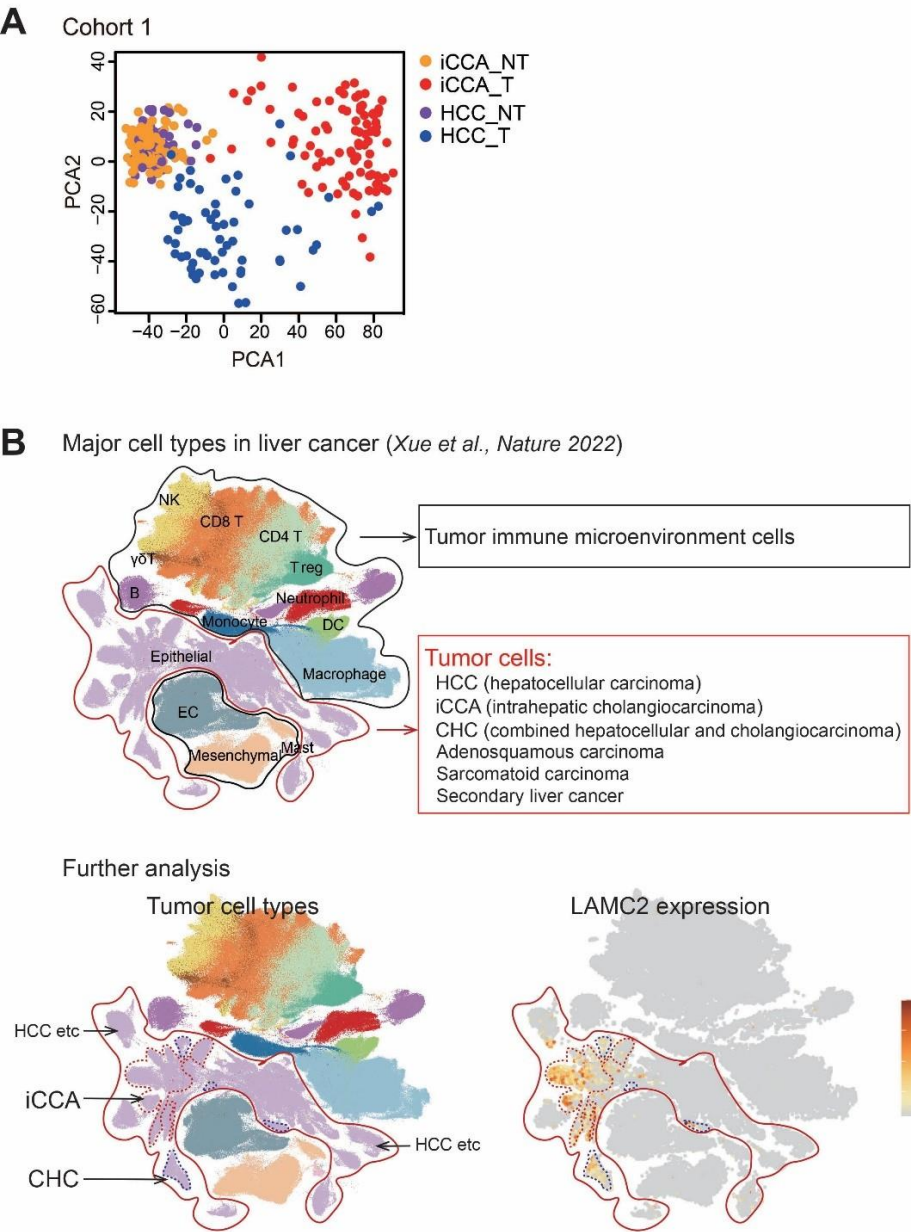


Figure S2

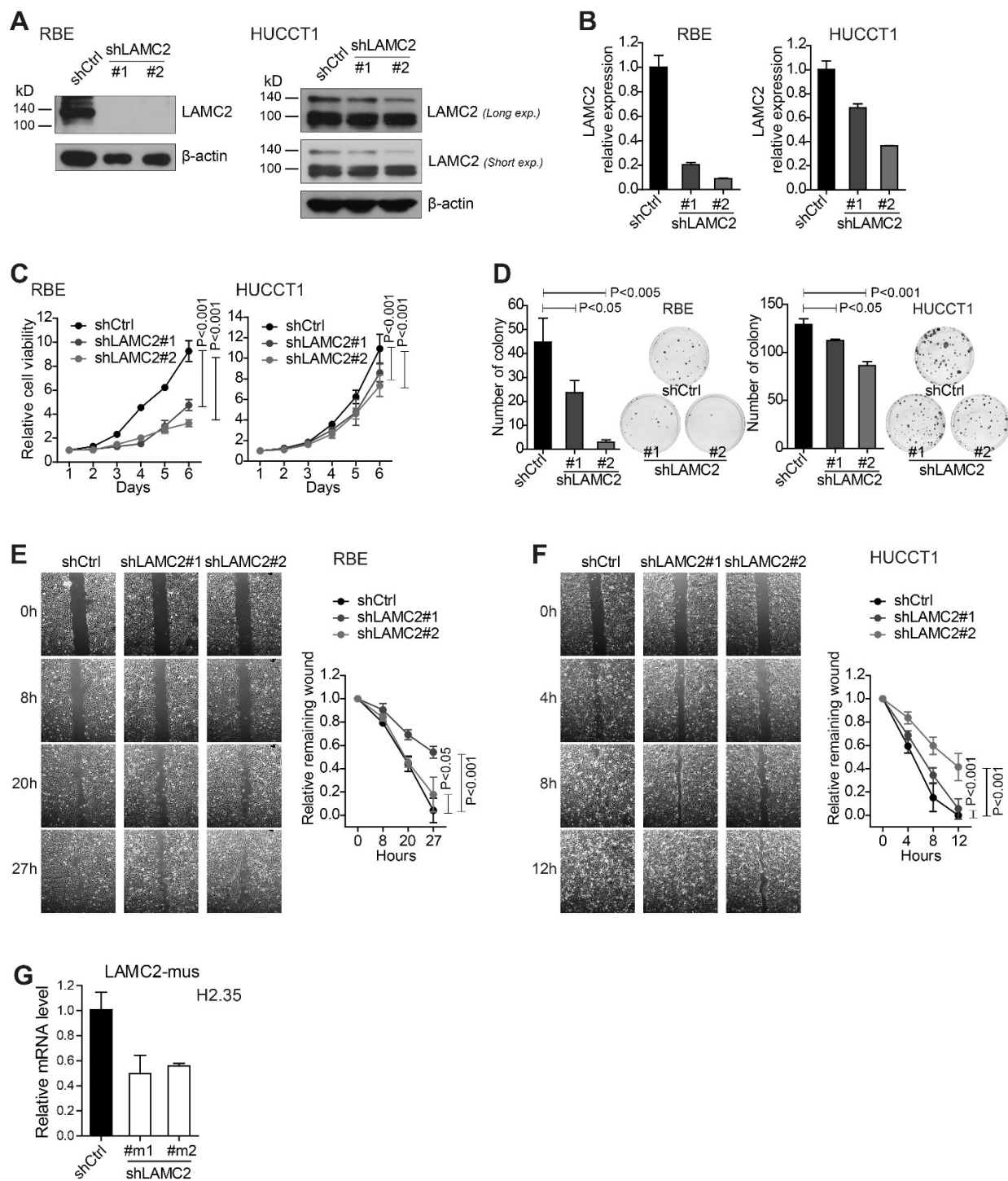


Figure S3

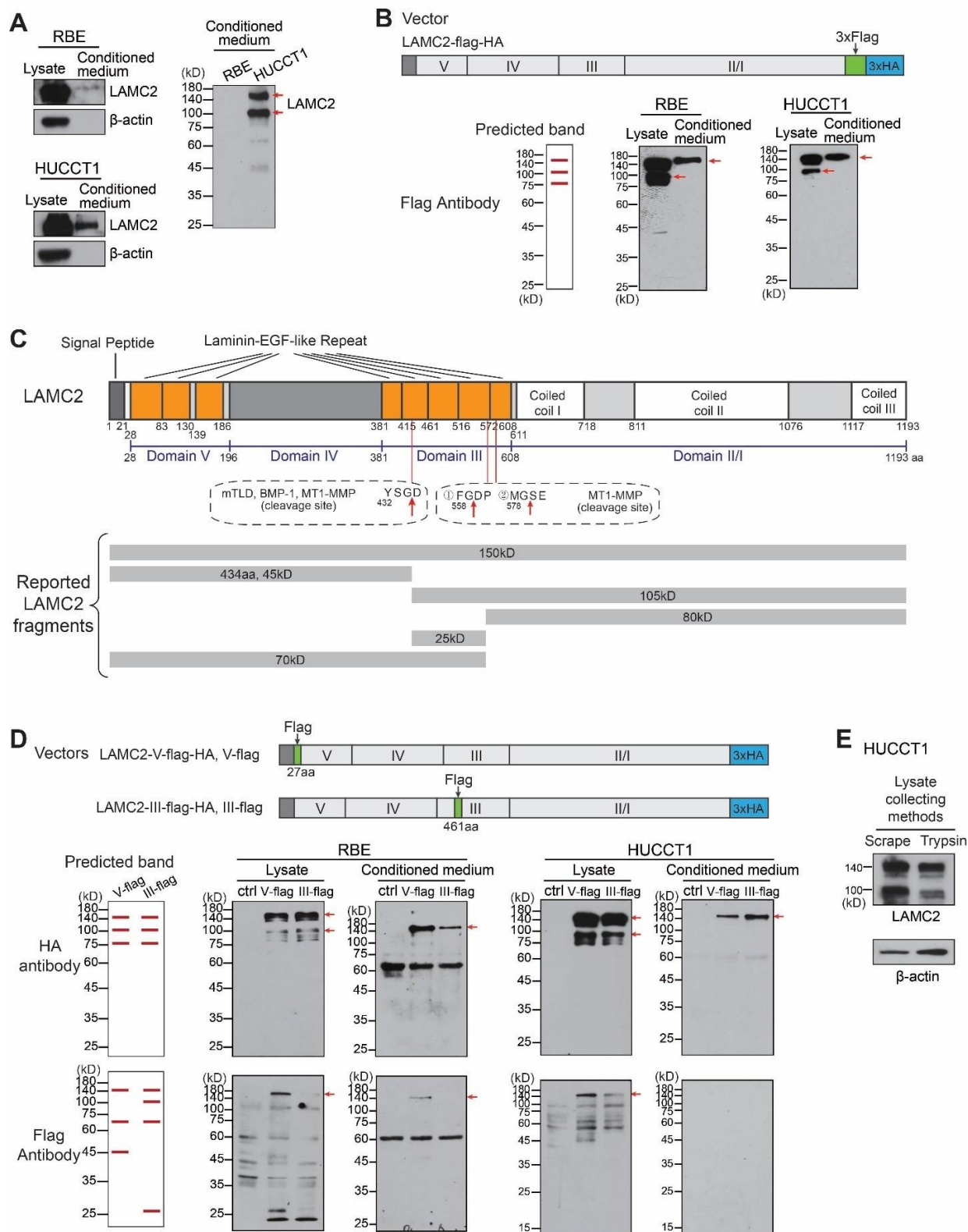


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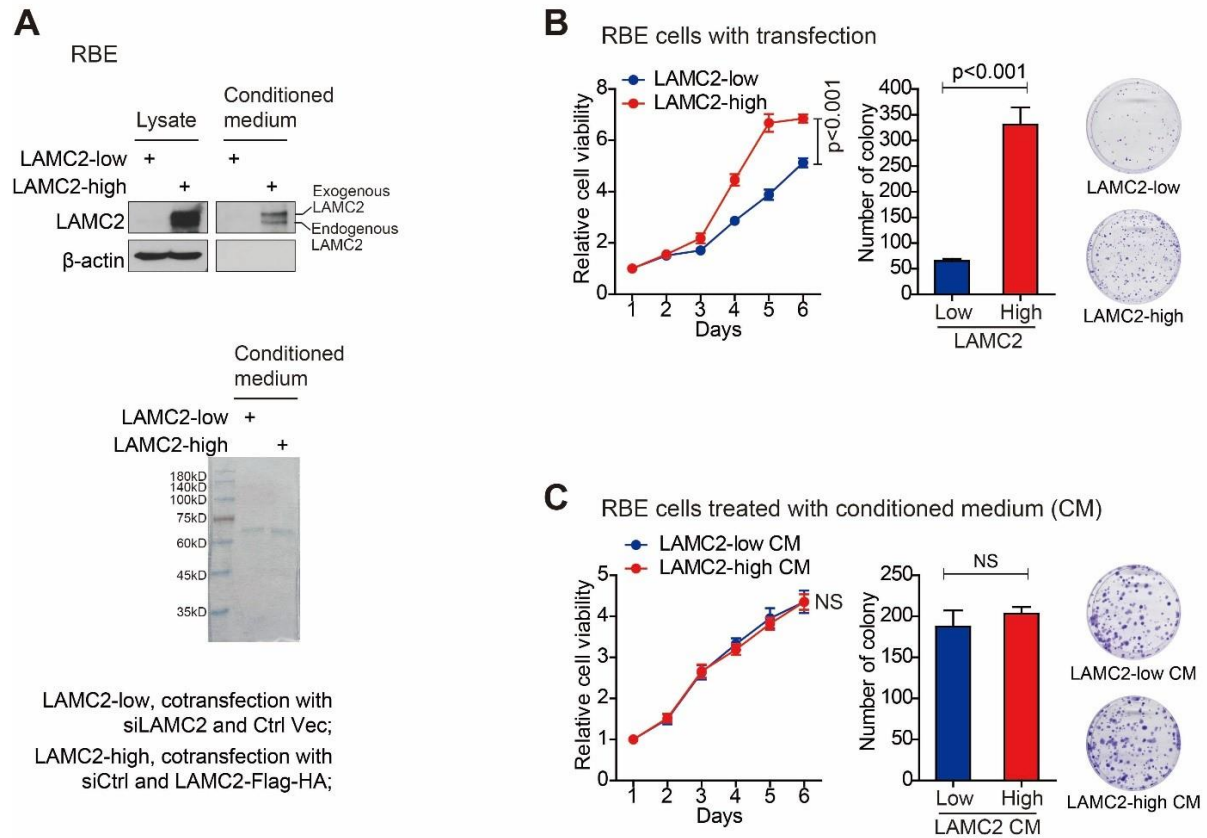


Figure S5

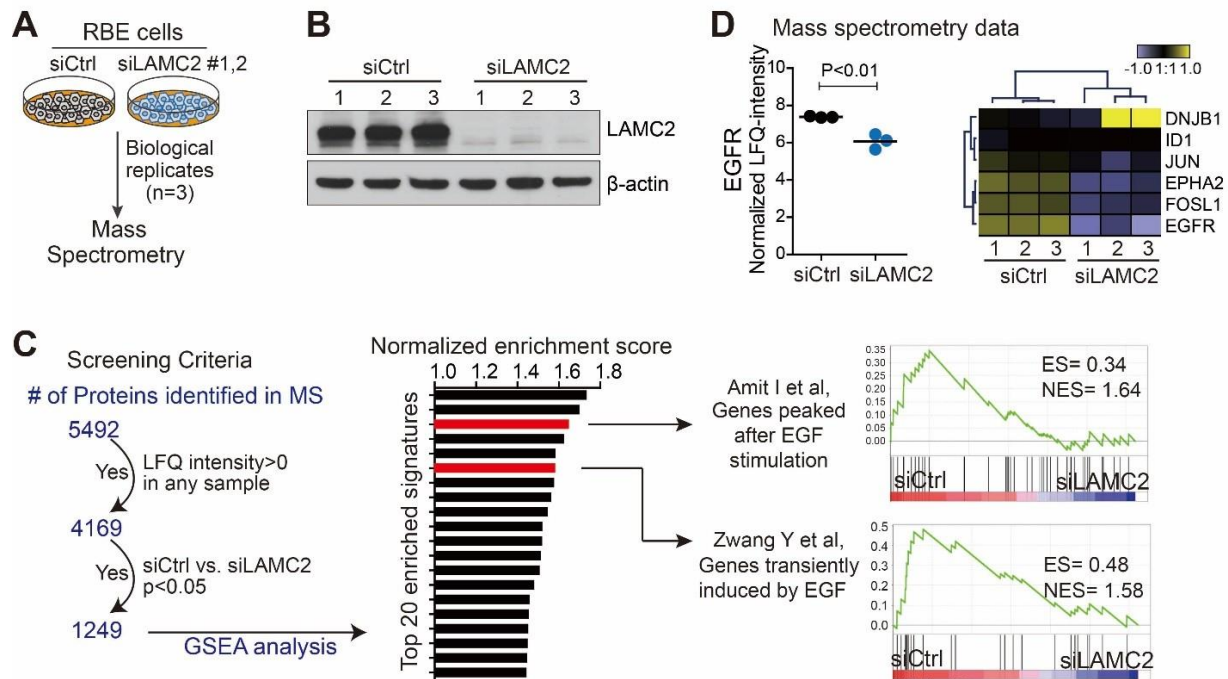


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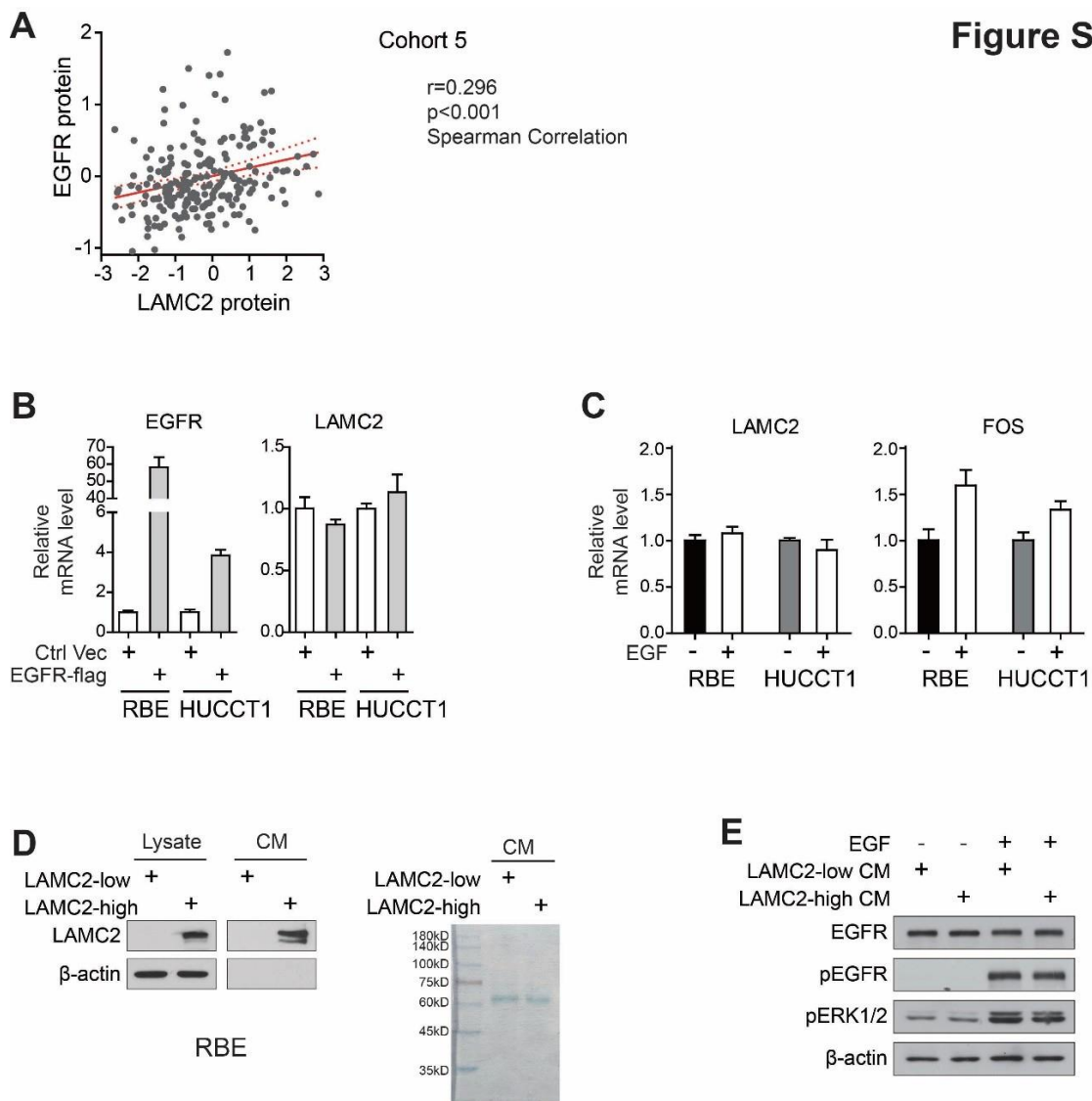


Figure S7

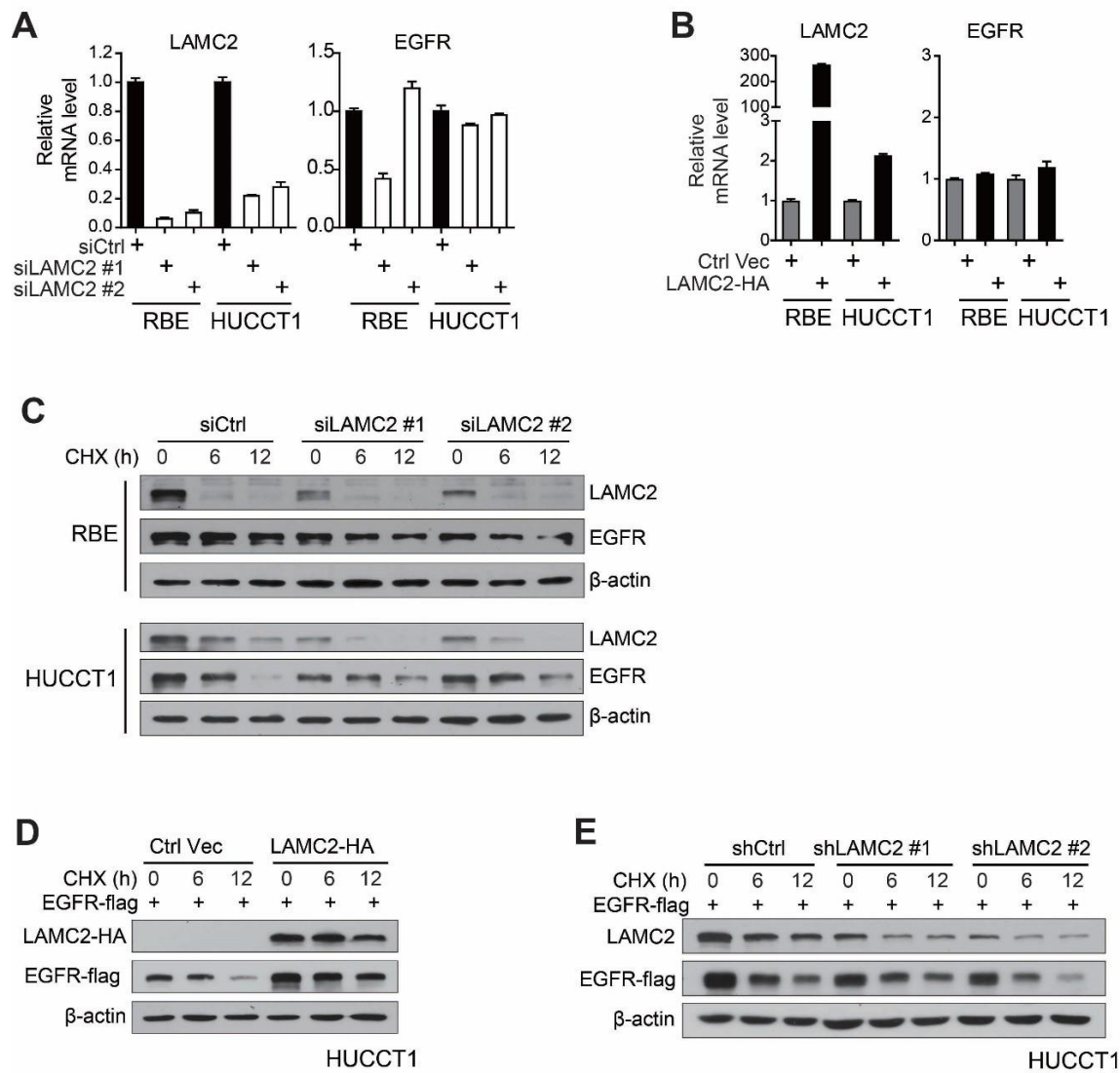


Figure S8

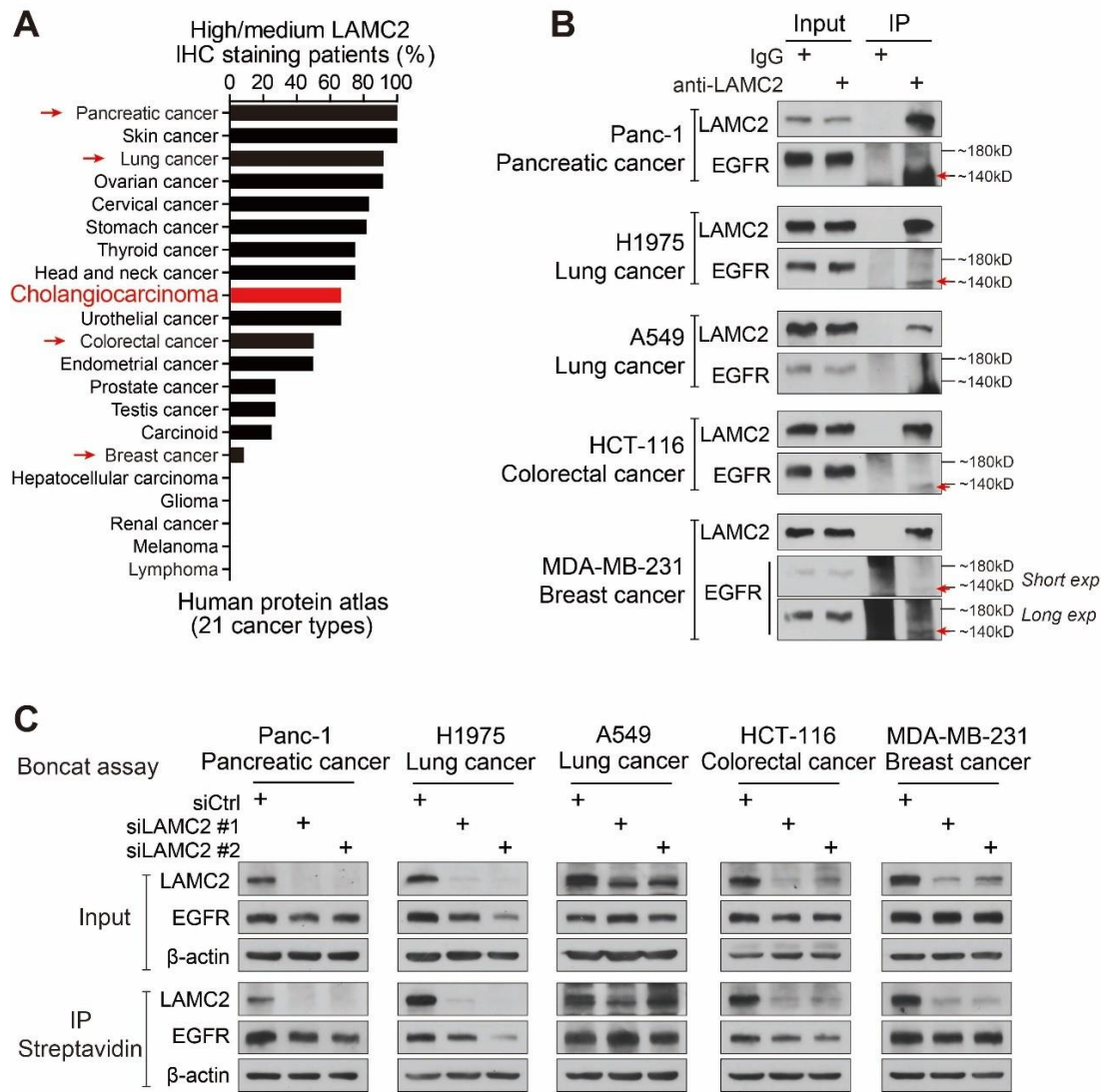


Figure S9

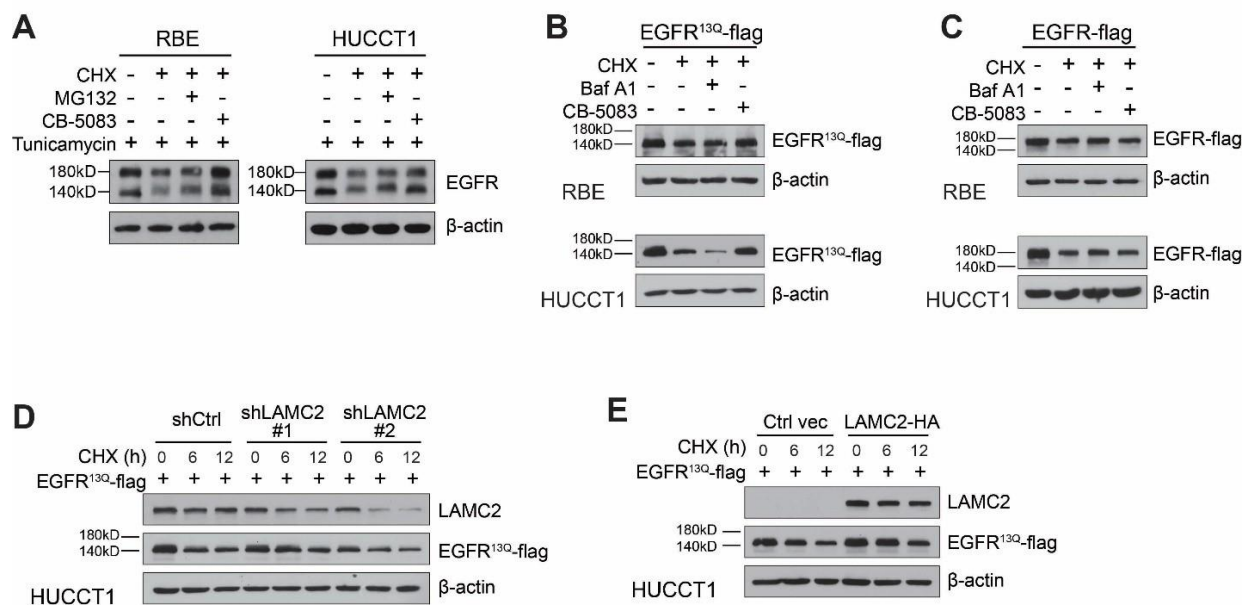


Figure S10

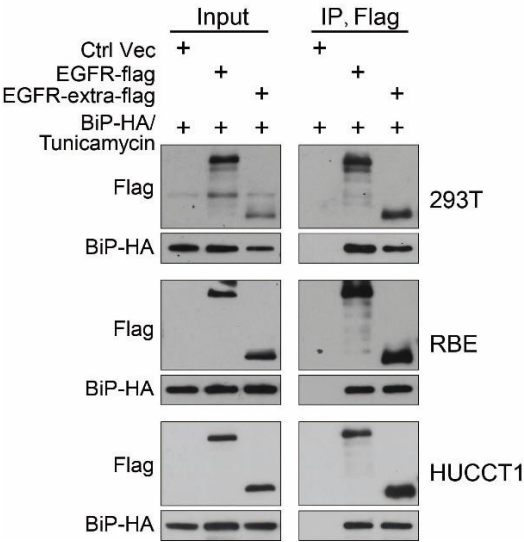


Figure S11

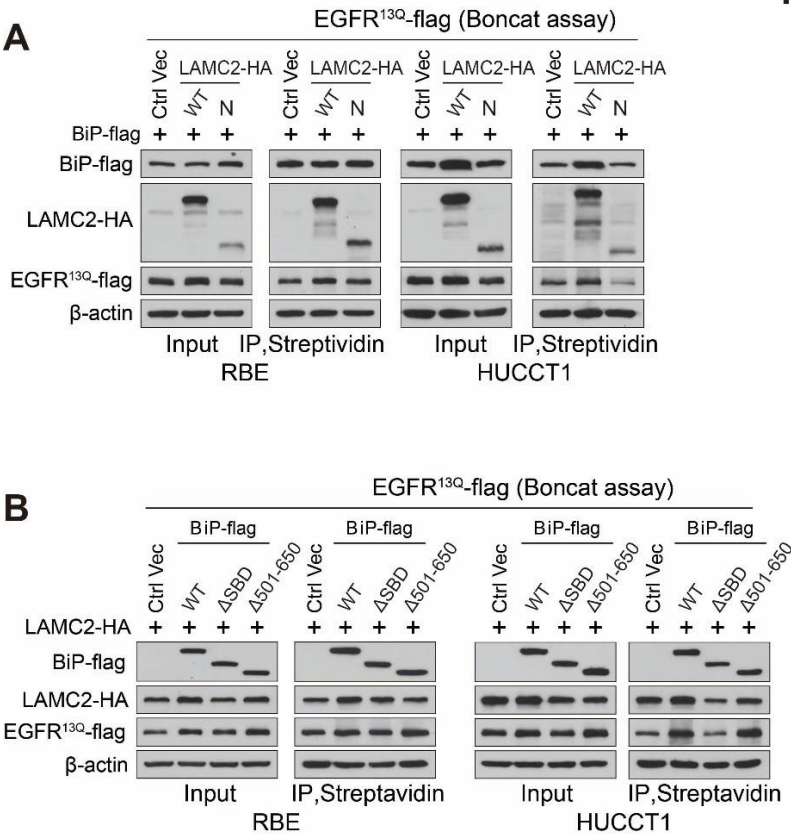


Figure S12

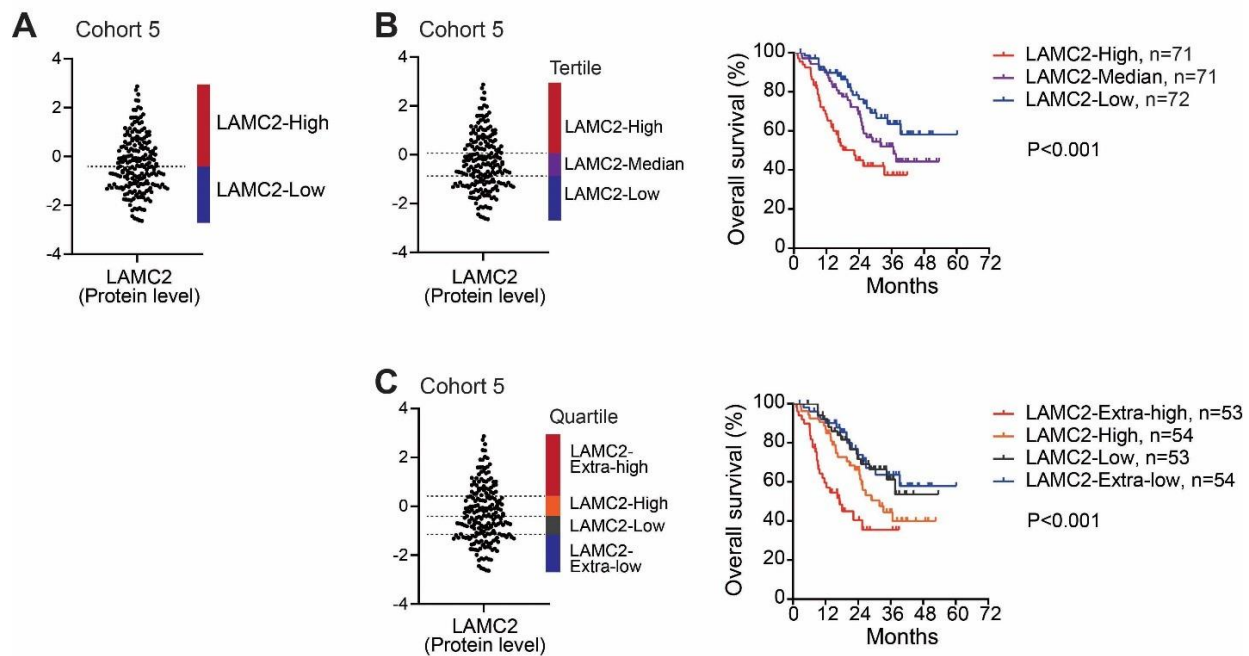


Figure S13

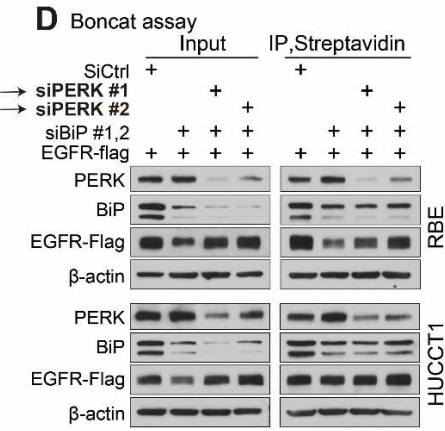
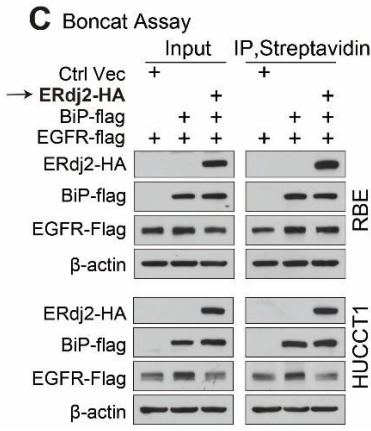
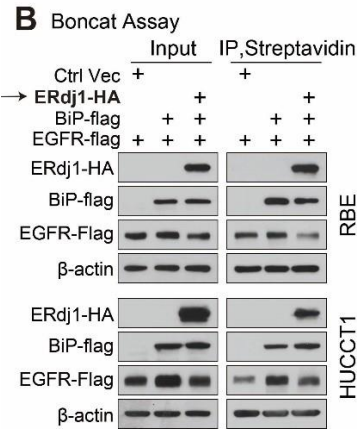
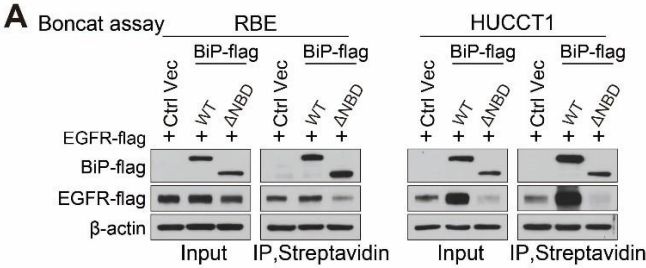


Figure S14

