## Vancomycin-resistant Enterococcal Bacteremia in a Hematology Unit: Molecular Epidemiology and Analysis of Clinical Course\*

An increase in vancomycin-resistant enterococcal (VRE) bacteremia in hematooncological patients (n=19) in our institution from 2000 through 2001 led us to analyze the molecular epidemiologic patterns and clinical features unique to our cases. The pulsed field gel electrophoresis of the isolates revealed that the bacteremia was not originated from a single clone but rather showed endemic pattern of diverse clones with small clusters. A different DNA pattern of blood and stool isolates from one patient suggested exogenous rather than endogenous route of infection. *Enterococcus faecium* carrying *vanA* gene was the causative pathogen in all cases. Patients with VRE bacteremia showed similar clinical courses compared with those with vancomycin-susceptible enterococcal (VSE) bacteremia. Vancomycin resistance did not seem to be a poor prognostic factor because of similar mortality (5/8, 62.5%) noted in VSE bacteremia. Initial disease severity and neutropenic status may be major determinants of prognosis in patients with VRE bacteraemia.

Key Words : Vancomycin Resistance; Enterococcus faecium; Enterococcus faecalis; Neutropenia; Bacteremia; Korea; Epidemiology, Molecular; VanB Protein, Enterococcus

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### INTRODUCTION

Vancomycin-resistant enterococci (VRE) have recently emerged as important nosocomial pathogens that mainly affect patients with severe underlying diseases (1, 2). VRE was first reported In Korea in 1992. Since then, however, the incidence of VRE was low until 2000 when an increase in VRE infections began to be reported in many hospitals (3-6). A nationwide survey suggested that the main mode of transmission was interhospital spread (4). Nowadays it is clear that VRE is a major newly established pathogen in Korean hospitals.

At the end of 2000, we recognized a sudden increase in the frequency of hemato-oncological patients with VRE bacteremia (8 patients) in our institution compared with previous years when there was 1 bacteremia in 1997 and 2 each in 1998 and 1999. This trend continued in 2001 in which there were 11 hemato-oncological patients with VRE bacteremia.

We herein conducted a retrospective analysis of the clinical and molecular epidemiologic characteristics of the VRE bacteremic patients who received hematopoietic stem cell transplantation (HSCT) or cytotoxic chemotherapy between January 2000 and December 2001.

## MATERIALS AND METHODS

#### Setting and patients

The Catholic HSCT Center belongs to St. Mary's Hospital, a 600-bed, university-affiliated, tertiary care teaching hospital. The HSCT center deals with more than 250 HSCT per year, which represents more than half of the annual HSCT cases in Korea. It follows that the analysis of clinical data for patients at our institution should provide reliable information about overall trends in hemato-oncological patients in Korea.

We reviewed retrospectively the medical records of all 19 VRE bacteremic patients over a 2-yr period (January 2000 through December 2001) in the Catholic HSCT Center affiliated with the Catholic University of Korea, College of Medicine.

Fever was defined as a single oral temperature of  $\geq 38.3$  °C or a temperature of  $\geq 38.0$  °C for  $\geq 1$  hr. Neutropenia was

defined as a neutrophil count of less than 500 cells/ $\mu$ L, or a count of less than 1,000 cells/ $\mu$ L with a predicted decrease to less than 500 cells/ $\mu$ L.

The term clinically significant VRE bacteremia was defined as at least two blood cultures positive for vancomycin-resistant *Enterococcus faecium* or *E. faecalis* (7).

An empirical antibiotic regimen was given to febrile patients according to the guidelines of the Infectious Diseases Society of America (8).

Identification of isolates, and testing of antibiotic susceptibility

Identification was performed with a conventional Microscan panel (Dade International, West Sacramento, CA, U.S.A.). VRE isolates were tested for antimicrobial susceptibility by the agar dilution method and the results were interpreted according to the Natinal Committee for Clinical Laboratory Standards 2002 breakpoints (9). *Enterococcus faecalis* ATCC 29212 and *Staphylococcus aureus* ATCC 29213 were used for quality control.

## Genotyping

Vancomycin resistance genotypes (*vanA*, *vanB*, *vanC1*, *van-C2/C3*) were tested by amplifying the respective genes by PCR (10), with some modifications. Primers for *vanA* were 5'-GGGAAAACGACAATTGC-3' (175-191) and 5'-ATT-GCCG GCGTAACATG-3' (891-907); for *vanB*, 5'-ATGG-GAAGCCGA TAGTC-3' (173-189) and 5'-CCAGCTCC-TTGCTTTAG-3' (791-807); for *vanC1*, 5'-GGTATCAAG-GAAACCTC-3' (246-272) and 5'-TCGATACTAC CGCC-TTC-3' (1051-1067), and for *vanC2/C3*, 5'-CTCCTACGAT-T CTCTTG-3' (455-486) and 5'-GAATTTCCAGAACGA-GC-3' (869-885). Strains *E. faecium* BM4147 with *vanA* and *ddIE. faecium*, *E. faecalis* V583 (*vanB*, *ddIE. faecalis*), *E. gallinarum* BM 41745 (*vanC1*), and *E. casseliflavus* ATCC 24788 (*vanC2/C3*) served as positive controls.

## Molecular epidemiological typing

Fourteen of the 19 VRE isolates were typed by pulsed field gel electrophoresis (PFGE) with *Sma*I as restriction enzyme, as previously described in detail (6). In addition, we obtained VRE isolates from specimens other than blood (urine and stool) that were available from eight patients without bacteremia (July 2001 through December 2001). These additional non-bacteremic (NB) isolates were also subjected to PFGE to investigate whether they might involve strains identical to or closely related (2-3 fragment differences) to strains in the patients with bacteremia.

PFGE was performed with a CHEF-DR II apparatus (Bio-Rad Korea, Seoul, Korea). The conditions were: initial switch time 0.5 sec, run time 20 hr, final switch time 40 sec. After ethidium bromide staining, patterns were compared and anayzed with Uniband/Map (version 99; UVItech, Cambridge, U.K.).

#### Data collection and analysis

We compared VRE bacteremic patients with a group of hematological patients with vancomycin-susceptible *E. faeci-um* (VSE) bacteremia (n=8) arising during the same period.

Baseline information included age, sex, and underlying disease. A simplified acute physiologic score (SAPS II) was assigned to all patients at the onset of bacteremia (11). We also followed up the patients' record until January 2002, to permit a survival analysis. The starting point of survivial was the date when bacteremia was detected.

Statistical analyses were carried out with SPSS 10.0 (SPSS Korea, Seoul, Korea). If the data were in the category of continous variables, t-test was performed. In case of data with ordinal or nominal scale, nonparametric tests such as chi-square, Mann-Whitney test, etc were done. *P* values <0.05 were considered statistically significant.

## RESULTS

#### Clinical characteristics of patients with VRE bacteremia

Table 1 gives the characteristics of the patients with VRE bacteremia. There were 8 cases of VRE bacteremia in the year 2000, and 11 cases in 2001 (Fig. 1). Eighteen of the 19 patients (94.7%) received broad-spectrum antibiotics prior to detection of the bacteremia. Only eight patients with VRE were able to receive quinupristin/dalfopristin (Q-D) because this was only formally available in our institution from 2001. The other 11 patients received high-dose ampicillin/sulbactam plus gentamicin, or imipenem monotherapy. Four of the 8 patients receiving Q-D died, compared with all the patients who did not receive Q-D (11/11).

# Microbiological characteristics of patients with VRE bacteremia

All 19 VRE isolates were identified as *E. faecium*. All were also *van A* genotype. Their antimicrobial susceptibility profiles showed high-level gentamicin resistance (100%), whereas only 14.3% had high-level resistance to streptomycin. None of them were resistant to Q-D or linezolid.

## Comparison of the clinical characteristics with VSE-bacteremia

Table 2 compares the demographic characteristics of patients with VRE versus those with VSE bacteremia. Simplified acute physiology score (SAPS) II were similar in both groups

	Age (yr)	Gender	Diagnosis	PFGE type*	Date of isolation (dd/mm/yy)	Room Number	Antibiotics given until bacteremia (days)	Antibiotics given for bacteremia (days)	Outcome
1	34	М	ALL	А	02/05/00	BMT	Cef (7), Gly (7), FQ (21)	SAM/GEN (15)	Expired
2	30	F	CML	J	08/08/00	BMT	Cef (11), Gly (10), Ag (15), AMB (18), FQ (35)	IPM (24)	Expired
3	19	М	ALL	D	31/10/00	1317	Cef (6), Gly (21), Ag (18), AMB (13), FQ (7)	SAM/GEN (2)	Expired
4	41	F	AML	I	02/11/00	1309	Cef (5), Gly (8), Ag (10), AMB (8), FQ (11)	SAM/GEN (38)	Expired
5	41	F	CML	E1	18/12/00	BMT	Cef (10), Gly (21), Ag (15), AMB (8), FQ (19)	SAM/GEN (3)	Expired
6	19	F	ALL	E3	09/01/01	1309	Cef (8), Gly (10), Ag (13), AMB (5), FQ (13)	Q-D (11)	Expired
7	59	М	MM	F	09/02/01	1209	Cef (8), Gly (7), AMB (2)	Q-D (23)	Survived
8	23	F	ALL	E2	04/04/01	BMT	Cef (6), Gly (10), Ag (4), AMB (4), FQ (10)	IPM (1)	Expired
9	16	М	AML	В	19/04/01	1302	Cef (2), Gly (12), Ag (14), AMB (12), FQ (15)	Q-D (5)	Survived
10	34	М	ALL	C1	09/05/01	1206	Cef (5), Gly (2), FQ (20)	Q-D (18)	Expired
11	22	F	SAA	G	16/05/01	1203	Cef (20), Gly (30), Ag (31), AMB (15)	IPM (8)	Expired
12	40	F	AML	C2	15/06/01	1309	Cef (4), Gly (3), Ag (4), FQ (13)	Q-D (14)	Survived
13	35	F	ALL	C3	22/11/01	1308	Cef (8), Gly (1), Ag (8), FQ (16)	Q-D (20)	Expired
14	16	М	ALL	Н	10/12/01	BMT	Cef (9), Gly (2), Ag (9), AMB (2)	IPM (1)	Expired
15	53	М	MDS	NT	26/02/00	1316	Cef (8), Gly (10), Ag (11), AMB (8), FQ (51)	Q-D (7)	Expired
16	37	М	AML	NT	26/03/00	BMT	Cef (3), Gly (5), Ag (5), AMB (2), FQ (28)	SAM/GEN (11)	Expired
17	32	F	AML	NT	09/07/00	BMT	Cef (1), Gly (1), Ag (1), FQ (14)	SAM/GEN (20)	Expired
18	53	FΕ	Breast Cance	r NT	26/05/01	1206	None	IPM (7)	Expired
19	16	М	ALL	NT	07/06/01	1307	Cef (16), Gly (16), Ag (24), AMB (11), FQ (12)	Q-D (11)	Survived

Table 1. Characteristics of patients with VRE bacteremia

VRE, vancomycin-resistant enterococci; AML, acute myelogenous leukemia; ALL, acute lymphocytic leukemia; CML, chronic myelogenous leukemia; SAA, severe aplastic anemia; MDS, Myelodysplastic syndrome; MM, Multiple myeloma; BMT, bone marrow transplantation room; NT, Not typed. Cef, 3rd generation cephalosporin; Gly, glycopeptide; Ag, aminoglycoside; FQ, fluoroquinolone; Q-D, quinupristin/dalfopristin; SAM/GEN, high dose ampicillin/sulbactam plus gentamicin; IPM, imipenem; AMB, amphotericin B. \*PFGE type: See Fig. 2.



Fig. 1. Summary of monthly occurrence of vancomycin-resistant *Enterococcus* (VRE) bacteremia from January 2000 to December 2001.

(42.6 $\pm$ 12.8 in VSE vs. 41.4 $\pm$ 13.7 in VRE, *p*>0.05, respectively), indicating that initial disease severity at the time of bacteremia did not differ significantly. Age, gender ratio, and other demographic characteristics were also similar.

The VSE group had a lower median number of days to bacteremia and fewer days of hospitalization than the VRE group, but the mortality of the two groups did not differ significantly (62.5% for VSE vs. 78.9% for VRE, p>0.05, respectively). More patients with VRE received antibiotics before bacteremia than those with VSE (94.7% vs. 62.5%, p<0.05, respectively). The median days survived for VSE was not statistically different from that for VRE as shown in Table 2 (log rank 1.55, p=0.2130).

## Characteristics of patients with VRE bacteremia who survived and of those who died

Table 3 compares patients with VRE bacteremia who survived (n=6) with those who died (n=15). Age, number of previous admissions, days to detection, total days of hospitalization, and neutropenic days did not differ significantly between the two groups. But SAPS II was significantly higher in those that died than in those that survived. While all the patients who survived recovered from the neutropenia, 40% of those who died did not.

### Molecular epidemiological characteristics of patients with VRE bacteremia

As illustrated in Fig. 1, the occurrence of VRE bacteremia appeared initially to be sporadic. However, its frequency began to increase from April 2001 and reached its peak through June 2001, which suggested us that there might have been a cluster of outbreaks in our institution. We therefore performed PFGE in order to determine whether the VRE bac-

	VSE bacteremia	VRE bacteremia
Median age (range) (yr)	45.5 (18-58)	34 (16-59)
Gender (male:female)	6:2	9:10
SAPS II	$42.6 \pm 12.8$	$41.4 \pm 13.7$
Underlying diagnosis		
AML	1	5
ALL	3	8
CML	3	2
SAA	1	1
Others	0	3
Type of treatment		
Chemotherapy	7	12
HSCT	1	7
Conservative care	0	0
Duration of neutropenia (days)	$23.4 \pm 11.6$	$27.4 \pm 17.9$
Hospitalization days	$33.4 \pm 15.6^{*}$	$56.2 \pm 28.2$
Median days to bacteremia since admission (range)	15.0 (1-22)*	27.0 (1-105)
Hospital days after detection of bacteremia	22.9±13.3	20.7±16.7
Median days survived after bacteremia (range)	45 (8-481)	26 (1-356)
Number of previous admission	$5.3 \pm 4.7$	$3.7 \pm 2.9$
Therapeutic administration of antibiotics before bacteremia	5/8 (62.5%)*	18/19 (94.7%)
Mortality	5/8 (62.5%)	15/19 (78.9%)

Table 2. Characteristics of patients with VRE bacteremia (n=19) and VSE bacteremia (n=8)

VSE, vancomycin-susceptible enterococci; SAPS, Simplified Acute Physiologic Score; HSCT, Haematopoietic stem cell transplantation; Others, multiple myeloma (1), myelodysplastic syndrome (1), breast cancer (1). \*: significantly different (*p*<0.05) from VRE bacteraemia group.

#### teremias from 2000 to 2001 were clonally related.

Most of the PFGE fingerprint patterns differed from one another, but there were two very small clusters of clones (C1-3 and E1-3, in Fig. 2 and Table 1). C1 was isolated on 9 May 2001, and C2 appeared about one month later (15 June 2001). As already suggested in Fig. 1, an outbreak did occur during the period May through June 2002, but it was very small (a mini-outbreak). Another closely related strain (C3) emerged 5 months later (22 November 2001) in a room adjacent to where C2 had appeared (room numbers 1308 and 1309, respectively, Table 1). There also occurred another mini-outbreak from December 2000 (E1) through January 2001 (E3). Three months later, a closely-related strain (E2) emerged in the same ward as E1. However the PFGE type E strains did not appear again (up to the end of 2001).

After examining the clonality of VRE bacteremia, we performed additional PFGE analyses, this time of NB isolates, in order to search for a possible source of transmission from patients without bacteremia. As shown in Fig. 3, we found three NB strains which were clonally related to those from patients with VRE bacteremia. The urine isolate (25 July 2001) from patient NB8 (lane 31 in Fig. 3) was virtually identical to PFGE type F which was isolated on 9 February 2001. The stool isolate (30 August 2001) from patient NB7

Table 3. Comparison between patients with VRE bacteremia who survived and those who died

	Survived (n=4)	Expired (n=15)	р
Median age (range) (yr)	28 (16-59)	34 (16-53)	NS
SAPS II	$33.0 \pm 2.5$	$43.6 \pm 14.6$	0.017
Number of previous admission	1.0±1.1	4.5±2.8	0.03
Median onset of day to bacteremia since admission	22 (1-39) (range)	27 (13-105)	NS
Hospitalization days	$45.5 \pm 14.6$	$59.1 \pm 30.6$	NS
Duration of neutropenia (days)	18.3±13.1	29.8±18.7	NS
Recovery from neutropenia	4 (100%)	6 (40.0%)	0.014



Fig. 2. Pulsed field gel electrophoresis (PFGE) of isolates from patients with VRE. 1-14, case numbers; A-J, PFGE genotypes (See Table I).

(lane 30) was closely related to PFGE type B (19 April 2001, lane 9), and the urine and stool isolates (28 December 2001) from patient NB3 (lane 21 and 22) were probably related to PFGE type C2. These patterns are summarized in Fig. 4.

The strains in lane 15 (blood) and 16 (stool) were from the same patient as lane 13 (C3), but the PFGE patterns of the blood and stool isolates differed from each other.

### DISCUSSION

In this study, all of 19 isolates were *E. facium* of *vanA* genotype; other vancomycin-resistant genotypes such as *vanB* or *vanC* were not found. This feature is consistent with other instances of VRE in Korea. In a Korean tertiary care hospital with 1,550 beds all of 330 VRE isolates were *vanA* (6). It follows that *vanA* is the major genotype in Korea although a case of *vanB* has also been reported (12). The 5th Korean Nation-



Fig. 3. PFGE analysis of VRE isolates from bacteremic patients and from non-bacteremic (NB) patients. M, molecular weight markers; lane 1-14: blood isolates from bacteremic patients (see Table I and Fig. 2); lane 15: blood isolate identical with that of lane 13; lane 16: stool isolates from the same person as lane 13; lanes 17-31: isolates from NB patients; lanes 17 & 18: urine and stool isolates from patient NB1, respectively; lanes 19 & 20: patient NB2; lanes 21 & 22: patient NB3; lanes 23 & 24: patient NB4; lanes 25 & 26: patient NB5; lanes 27 & 28: patient NB6; lanes 29 & 30: patient NB7; lane 31: urine isolate from patient NB8. Lines connecting lanes 7 and 31, lanes 9 and 30, lanes 12 and 21-22 indicate clonally related pairs and groups.



Fig. 4. Summary of epidemiological patterns of four main clones (B, C, E, F) circulating in our institution during 2 yr (2000-2001).

wide Surveillance of Antimicrobial Resistance also revealed an increase in vancomycin-resistant *E. faecium* in the period 1997-2001 (3).

We used patients with VSE for comparison with the VRE group, as we thought that a control group that simply did not have VRE bacteremia would not be suitable because it would differ in many respects: disease severity as well as clinical characteristics would be quite different from those with VRE, and it would be hard to adjust for these variables. On the other hand, the initial severity of disease as represented by SAPS II was not significantly different in VSE group from that of the VRE bacteremia group. The well-established risk factors for VRE acquisition are neutropenia, prolonged hospitalization, prior use of broad-spectrum antibiotics, and hypoalbuminemia (13-17). These factors, with the exception of days of hospitalization, did not differ significantly between our VRE and VSE groups. However, as the number of patients with VSE (n=8) was very small, and less than half of those with VRE (n=19), we think the variables predictive of the acquisition of VRE should be evaluated in a larger study.

The influence of vancomycin-resistance on mortality is controversial. Some authors have insisted on its importance (18-20), whereas others are of the contrary opinion (21, 22). Since the mortality of our patients with VSE did not differ from those with VRE, it is unlikely that vancomycin-resistance itself affects the prognosis. Our findings suggest that enterococcal infection itself, or its degree of virulence, rather than vancomycin-resistance, is a determinant of prognosis.

Several authors have suggested that Q-D and linezolid would both be effective in treating VRE (23-27). In our study, the patients who received Q-D appeared to do better than those receiving other antimicrobials, though the number of cases (n=8) was too small to be sure.

Among the 19 patients with VRE bacteremia, expired patients had significantly higher SAPS II values than those who survived, and 60% of the latter did not recover from neutropenia. Although further cases are needed to establish the predictors of outcome in VRE bacteremia, we think that initial disease severity, and host immunity as reflected in neutropenia, could be major determinants.

Since its first isolation in 1992, VRE seemed to be rare until 1997 after which an abrupt increase in VRE infections was reported by many hospitals in Korea from 1998 onwards (3-6). One study of VRE infection in a Korean tertiary teaching hospital revealed that the increase in VRE was probably due to the increased use of oral vancomycin to treat Clostridium difficile infections, and clonal spread (6). Another nationwide study in Korea also suggested that interhospital spread contributed to the rapid increase of VRE in many hospitals (4). The findings of our study suggested that spread by a few clones among heterogenous ones may have contributed to the increase in VRE bacteremia. However, it could not be the only mechanism because the PFGE analysis demonstrated a high degree of diversity among the isolates, and bacteremia of endogenous origin could not be excluded. As reported earlier, the hemato-oncological disease itself has been recognized as an important risk factor for the acquisition of VRE (12, 28, 29). In addition to the lack of host immunity, breakdown of the gastrointestinal mucosal barrier and subsequent translocation of bowel flora could play a major role in the acquisition of VRE bacteremia (30-32). In other words, endogenous infection by translocation as well as exogenous transmission from other patients (33, 34) or from fomites (35) could have been another major source of VRE bacteremia in our patients. However, there was also a perplexing but nevertheless important observation arguing against the endogenous route. As illustrated in Fig. 3, blood and stool isolates from one bacteremic patient yielded completely different DNA band patterns (lanes 15 and 16, respectively). If the patient's bacteremia was from her own bowel flora, the DNA patterns of blood and stool isolates should have been identical or at least closelyrelated. This finding suggested that the VRE, at least in this patient, was not of endogenous origin but came from someone or somewhere else. However, the possibility that the patient may be colonized with more than one strain or that there was transfer of the vanA gene could not be excluded.

While there have been several reports of a single clone dominating a VRE outbreak (29, 36, 37), many molecular epidemiologic studies have shown that most of the VRE in a given hospital is clonally diverse (14, 39-42). In our case, the VRE of our bacteremic patients was clonally diverse with two small clusters of mini-outbreaks (E1-3 and C1-3 in Fig. 2), which suggested that VRE already became endemic in our institution. We also found three strains from non-bacteremic patients were clonally related to those in bacteremic patients (PFGE types B, C2, and F), and each of these strains appeared sporadically at intervals of 4-6 months. Whereas strains of type E (found only in bacteremic patients), B and F did not appear again after August 2001, type C strains continued to emerge up to the end of the study. Based on these findings, we assume that there were multiple clones of VRE coexisting with at least two primary clones of sporadic patterns in our unit, as also postulated in another instance (43).

Though both exogenous and endogenous modes of infection could be major causes of bacteremia in our patients, the evidence for mini-outbreaks in the midst of clonal diversity strongly suggests that strict precautions are crucial to prevent nosocomial transmission of VRE (44, 45).

In summary, patients with VRE (all were *vanA E. faecium*) bacteremia showed heterogenous molecular epidemiologic pattern with some small clusters, but vancomycin-resistance did not seem to be a prognostic factor because of the similar mortality in the VSE group. We consider that greater attention should be paid to the prevention and management of VRE bacteremia because it is now one of the important established pathogens in hemato-oncological patients in Korean hospitals.

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## REFERENCES

- Cetinkaya Y, Falk P, Mayhall CG. Vancomycin-resistant enterococci. Clin Microbiol Rev 2000; 13: 686-707.
- Vergis EN, Hayden MK, Chow JW, Snydman DR, Zervos MJ, Linden PK, Wagener MM, Schmitt B, Muder RR. *Determinants of vancomycin resistance and mortality rates in enterococcal bacteremia: a prospective multicenter study. Ann Intern Med* 2001; 135: 484-92.
- 3. Lee K, Jang SJ, Lee HJ, Ryoo N, Kim M, Hong SG, Chong Y; The Korean Nationwide Surveillance of Antimicrobial Resistance Group. Increasing prevalence of vancomycin-resistant Enterococcus faecium, expanded-spectrum cephalosporin-resistant Klebsiella pneumoniae, and imipenem-resistant Pseudomonas aeruginosa in Korea: KONSAR Study in 2001. J Korean Med Sci 2004; 19: 8-14.
- 4. Lee K, Lee HS, Jang SJ, Park AJ, Lee MH, Song WK, Chong Y; Members of Korean Nationwide Surveillance of Antimicrobial Resistance Group. Antimicrobial resistance surveillance of bacteria in 1999 in Korea with a special reference to resistance of enterococci to vancomycin and Gram-negative bacilli to third generation cephalosporin, imipenem, and fluoroquinolone. J Korean Med Sci 2001; 16: 262-70.
- Lee WG, Jernigan JA, Rasheed JK, Anderson GJ, Tenover FC. Possible horizontal transfer of the vanB2 gene among genetically diverse strains of vancomycin resistant Enterococcus faecium in a Korean hospital. J Clin Microbiol 2001; 39: 1165-8.
- 6. Shin JW, Yong D, Kim MS, Chang KH, Lee K, Kim JM, Chong Y. Sudden increase of vancomycin-resistant enterococcal infections in a Korean tertiary care hospital: possible consequences of increased use of oral vancomycin. J Infect Chemother 2003; 9: 62-7.
- Garner JS, Jarvis WR, Emori TG, Horan TC, Hughes JM. CDC definitions for nosocomial infections, 1988. Am J Infect Control 1988; 16: 128-40.

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- Hughes WT, Armstrong D, Bodey GP, Bow EJ, Brown AE, Calandra T, Feld R, Pizzo PA, Rolston KV, Shenep JL, Young LS. 2002 guidelines for the use of antimicrobial agents in neutropenic patients with cancer. Clin Infect Dis 2002: 34: 730-51.
- National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial susceptibility testing; Twelfth informational supplement; Natinal Committee for Clinical Laboratory Standards, Wayne, PA, 2002.
- Dutka-Malen S, Evers S, Courvalin P. Detection of glycopeptide resistance genotypes and identification to the species level of clinically relevant enterococci by PCR. J Clin Microbiol 1995; 32: 24-7.
- Le Gall JR, Lemeshow S, Saulnier F. A new simplified acute physiology score (SAPS II) based on a European/North American multicenter study. JAMA. 1993; 270: 2957-63.
- Chadwick PR, Oppenheim BA, Fox A, Woodford N, Morgenstern GR, Scarffe JH. Epidemiology of an outbreak due to glycopeptide resistant Enterococcus faecium on a leukaemia unit. J Hosp Infect 1996; 34: 171-82.
- Boyce JM, Opal SM, Chow JW, Zervos MJ, Potter-Bynoe G, Sherman CB, Romulo RL, Fortna S, Medeiros AA. Outbreak of multidrugresistant Enterococcus faecium with transferable vanB class vancomycin resistance. J Clin Microbiol 1994; 32: 1148-53.
- 14. Boyle JF, Soumakis SA, Rendo A, Herrington JA, Gianarkis DG, Thurberg BE, Painter BG. Epidemiologic analysis and genotypic characterization of a nosocomial outbreak of vancomycin-resistant enterococci. J Clin Microbiol 1993; 31: 1280-5.
- Harbarth S, Cosgrove S, Carmeli Y. Effects of antibiotics on nosocomial epidemiology of vancomycin-resistant Enterococci. Antimicrob Agents Chemother 2002; 46: 1619-28.
- 16. Nourse C, Murphy H, Byrne C, O'Meara A, Breatnach F, Kaufmann M, Clarke A, Butler K. Control of a nosocomial outbreak of vancomycin resistant Enterococcus faecium in a paediatric oncology unit: risk factors for colonisation. Eur J Pediatr 1998; 157: 20-7.
- Suntharam N, Lankford MG, Trick WE, Peterson LR, Noskin GA. Risk factors for acquisition of vancomycin resistant enterococci among haematology oncology patients. Diagn Microbiol Infect Dis 2002; 43: 183-8.
- Bach PB, Malak SF, Jurcic J, Gelfand SE, Eagan J, Little C, Sepkowitz KA. Impact of infection by vancomycin resistant Enterococcus on survival and resource utilization for patients with leukemia. Infect Control Hosp Epidemiol 2002; 23: 471-4.
- Linden PK, Pasculle AW, Manez R, Kramer DJ, Fung JJ, Pinna AD, Kusne S. Differences in outcomes for patients with bacteremia due to vancomycin-resistant Enterococcus faecium or vancomycin-susceptible E. faecium. Clin Infect Dis 1996; 22: 663-70.
- Stosor V, Peterson LR, Postelnick M, Noskin GA. Enterococcus faecium bacteremia: does vancomycin resistance make a difference? Arch Intern Med 1998; 158: 522-7.
- Garbutt JM, Ventrapragada M, Littenberg B, Mundy LM. Association between resistance to vancomycin and death in cases of Enterococcus faecium bacteremia. Clin Infect Dis 2000; 30: 466-72.
- 22. Wells CL, Juni BA, Cameron SB, Mason KR, Dunn DL, Ferrien P, Rhame FS. Stool carriage, clinical isolation, and mortality during an outbreak of vancomycin-resistant enterococci in hospitalized medi-

cal and/or surgical patients. Clin Infect Dis 1995; 21: 45-50.

- Goff DA, Sierawski SJ. Clinical experience of quinupristin-dalfopristin for the treatment of antimicrobial-resistant gram-positive infections. Pharmacotherapy 2002; 22: 748-58.
- Linden PK, Moellering RC Jr, Wood CA, Rehm SJ, Flaherty J, Bompart F, Talbot GH; Synercid Emergency-Use Study Group. *Treatment* of vancomycin-resistant Enterococcus faecium infections with quinupristin/dalfopristin. Clin Infect Dis 2001; 33: 1816-23.
- Moellering RC. Quinupristin/dalfopristin: therapeutic potential for vancomycin-resistant enterococcal infections. J Antimicrob Chemother 1999; 44 (Suppl): S25-30.
- 26. Moellering RC, Linden PK, Reinhardt J, Blumberg EA, Bompart F, Talbot GH. The efficacy and safety of quinupristin/dalfopristin for the treatment of infections caused by vancomycin-resistant Enterococcus faecium. Synercid Emergency-Use Study Group. J Antimicrob Chemother 1999; 44: 251-61.
- Winston DJ, Emmanouilides C, Kroeber A, Hindler J, Bruckner DA, Territo MC, Busuttil RW. *Quinupristin/Dalfopristin therapy for infections due to vancomycin-resistant Enterococcus faecium. Clin Infect Dis 2000; 30: 790-7.*
- 28. Roghmann MC, McCarter RJ, Brewrink J, Cross AS, Morris JG. Clostridium difficile infection is a risk factor for bacteremia due to vancomycin resistant enterococci (VRE) in VRE-colonized patients with acute leukemia. Clin Infect Dis 1997; 25: 1056-9.
- 29. Timmers G, van der Zwet WC, Simoons-Smit IM, Savelkoul PH, Meester HH, Vandenbroucke-Grauls CM, Huijgens PC. Outbreak of vancomycin-resistant Enterococcus faecium in a haematology unit: risk factor assessment and successful control of the epidemic. Br J Haematol 2002; 116: 826-33.
- Edmond MB, Ober JF, Weinbaum DL, Pfaller MA, Hwang T, Sanford MD, Wenzel RP. Vancomycin resistant Enterococcus faecium bacteremia: risk factors for infection. Clin Infect Dis 1995; 20: 1126-33.
- Falk PS, Winnike J, Woodmansee C, Desai M, Mayhall CG. Outbreak of vancomycin resistant enterococci in a burn unit. Infect Control Hosp Epidemiol 2000; 21: 575-82.
- 32. Kuehnert MJ, Jernigan JA, Pullen AL, Rimland D, Jarvis WR. Association between mucositis severity and vancomycin resistant enterococcal bloodstream infection in hospitalized cancer patients. Infect Control Hosp Epidemiol 1999; 20: 660-3.
- 33. Handwerger S, Raucher B, Altarac D, Monka J, Marchione S, Singh KV, Murray BE, Wolff J, Walters B. Nosocomial outbreak due to Enterococcus faecium highly resistant to vancomycin, penicillin, and gentamicin. Clin Infect Dis 1993; 16: 750-5.
- Noskin GA, Stosor V, Cooper I, Peterson LR. Recovery of vancomycin resistant enterococci on fingertips and environmental surfaces. Infect Control Hosp Epidemiol 1995; 16: 577-81.
- 35. Porwancher R, Sheth A, Remphrey S, Taylor E, Hinkle C, Zervos M. Epidemiological study of hospital-acquired infection with vancomycin-resistant Enterococcus faecium: possible transmission by an electronic ear-probe thermometer. Infect Control Hosp Epidemiol 1997; 18: 771-4.
- 36. Dunne WM Jr, Wang W. Clonal dissemination and colony morphotype variation of vancomycin-resistant Enterococcus faecium iso-

lates in metropolitan Detroit, Michigan. J Clin Microbiol 1997; 35: 388-92.

- Fridkin SK, Yokoe DS, Whitney CG, Onderdonk A, Hooper DC. Epidemiology of a dominant clonal strain of vancomycin-resistant Enterococcus faecium at separate hospitals in Boston, Massachusetts. J Clin Microbiol 1998; 36: 965-70.
- Bonten MJ, Hayden MK, Nathan C, van Voorhis J, Matushek M, Slaughter S, Rice T, Weinstein RA. *Epidemiology of colonization of* patients and environment with vancomycin-resistant enterococci. Lancet 1996; 348: 1615-9.
- Chow JW, Kuritza A, Shlaes DM, Green M, Sahm DF, Zervos MJ. Clonal spread of vancomycin-resistant Enterococcus faecium between patients in three hospitals in two states. J Clin Microbiol 1993; 31: 1609-11.
- 40. Morris JG Jr, Shay DK, Hebden JN, McCarter RJ Jr, Perdue BE, Jarvis W, Johnson JA, Dowling TC, Polish LB, Schwalbe RS. Enterococcci resistant to multiple antimicrobial agents including vancomycin: establishment of endemicity in a university medical cen-

ter. Ann Intern Med 1995; 123: 250-9.

- 41. Nourse C, Byrne C, Murphy H, Kaufmann ME, Clarke A, Butler K. Eradication of vancomycin resistant Enterococcus faecium from a pediatric oncology unit and prevalence of colonization in hospitalized and community-based children. Epidemiol Infect 2000;124: 53-9.
- Stosor V, Kruszynski J, Suriano T, Noskin GA, Peterson LR. Molecular epidemiology of vancomycin-resistant Enterococci: a 2-year perspective. Infect Control Hosp Epidemiol 1999; 20: 653-9.
- Montecalvo MA, de Lencastre H, Carraher M, Gedris C, Chung M, VanHorn K, Wormser GP. Natural history of colonization with vancomycin resistant Enterococcus faecium. Infect Control Hosp Epidemiol 1995; 16: 680-5.
- 44. Garner JS. Guideline for isolation precautions in hospitals. Infect Control Hosp Epidemiol 1996; 17: 53-80.
- 45. CDC. Recommendations for preventing the spread of vancomycin resistance: recommendations of the Hospital Infection Control Practices Advisory Committee. MMWR 1995; 44: 1-13.

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