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Effects of water avoidance stress on peripheral and central responses during bladder filling in the rat: A multidisciplinary approach to the study of urologic chronic pelvic pain syndrome (MAPP) research network study

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# Abstract

Stress plays a role in the exacerbation and possibly the development of functional lower urinary tract disorders. Chronic water avoidance stress (WAS) in rodents is a model with high construct and face validity to bladder hypersensitive syndromes, such as interstitial cystitis/ bladder pain syndrome (IC/BPS), characterized by urinary frequency and bladder hyperalgesia and heightened stress responsiveness. Given the overlap of the brain circuits involved in stress, anxiety, and micturition, we evaluated the effects chronic stress has on bladder function, as well as its effects on regional brain activation during bladder filling. Female Wistar-Kyoto rats were exposed to WAS (10 days) or sham paradigms. One day thereafter, cystometrograms were obtained during titrated bladder dilation, with visceromotor responses (VMR) recorded simultaneously. Cerebral perfusion was assessed during passive bladder distension (20-cmH<sub>2</sub>O) following intravenous administration of [<sup>14</sup>C]-iodoantipyrine. Regional cerebral blood flow was quantified by autoradiography and analyzed in 3dimensionally reconstructed brains with statistical parametric mapping. WAS animals compared to controls demonstrated a decreased pressure threshold and visceromotor threshold triggering the voiding phase. At 20-cmH<sub>2</sub>O, VMR was significantly greater in WAS animals compared to controls. WAS animals showed greater activation in cortical regions of the central micturition circuit, including the posterior cingulate, anterior retrosplenial, somatosensory, posterior insula, orbital, and anterior secondary ("supplementary") motor cortices, as well as in the thalamus, anterior hypothalamus, parabrachial and Barrington nuclei, and striatum. Seed analysis showed increased functional connectivity of WAS compared to control animals of the posterior cingulate cortex to the pontine parabrachial nucleus; of the



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Barrington nucleus to the anterior dorsal midline and ventrobasilar thalamus and somatosensory and retrosplenial cortices; and of the posterior insula to anterior secondary motor cortex. Our findings show a visceral hypersensitivity during bladder filling in WAS animals, as well as increased engagement of portions of the micturition circuit responsive to urgency, viscerosensory perception and its relay to motor regions coordinating imminent bladder contraction. Results are consistent with recent findings in patients with interstitial cystitis, suggesting that WAS may serve as an animal model to elucidate the mechanisms leading to viscerosensitive brain phenotypes in humans with IC/BPS.

# Introduction

Chronic emotional stress plays a role in the exacerbation and possibly the development of functional lower urinary tract disorders (LUTD) [1-5]. These disorders can be viewed as a spectrum of bladder hypersensitivity syndromes sharing the common symptoms of urinary frequency and urgency, with bladder pain syndrome/interstitial cystitis (IC/BPS) patients experiencing pain as an additional symptom. Work over the past decade in chronic pain disorders, including IC/BPS, irritable bowel syndrome, fibromyalgia and vulvodynia, suggests the presence in these conditions of altered sensory perception [6-8]. Specific to IC/BPS, recent work has shown that patients compared to healthy controls show increased functional brain activation in the full bladder state broadly across sites in brain circuits serving sensory perception and pain [9]. The possibility of altered central processing of sensory input is paralleled by epidemiologic studies documenting that a substantial number of IC/BPS patients report stressrelated symptom onset and/or severity, difficulty coping with stressful situations, and symptoms of depression, anxiety, and panic disorder [5,10-15]. However, the relationship between chronic stress, bladder function and brain functional responses remains unclear. The current study addresses this in an animal model by prospectively examining the relationship between chronic stress exposure, bladder function, hyperalgesia and functional brain activation. Our prior work has shown that chronic psychological stress in high-anxiety rats induces urinary frequency, sustained bladder hyperalgesia, tactile hindpaw allodynia and suprapubic hyperalgesia [16,17]. Using this animal model, we now examine the effects chronic psychological stress has on functional bladder dynamics, as well as functional brain responses during bladder filling. We hypothesize that rats exposed to water avoidance stress compared to non-stressed controls will during passive bladder filling show abnormal bladder function and sensitivity, as well as increased activation of brain regions within the micturition circuit.

# Materials and methods

#### Animals

Adult, female Wistar-Kyoto rats (WKY strain 008) were purchased from a commercial vendor (Charles River, Wilmington, MA, USA). Rats were group housed under standard vivaria conditions (lights off from 7 p.m. to 7 a.m.). WAS and control groups were separately housed in groups of two with direct bedding (7090 Teklad sani-chips, Envigo), and ad libitum access to standard lab chow (Laboratory Rodent Diet 5001, Constant Nutrition<sup>®</sup>) and water (internal reverse osmosis systems).

Experimental procedures were approved by the University of Southern California's Institutional Animal Care and Use Committee (protocol #20263) and were conducted in accordance with the National Research Council Guide for the Care and Use of Laboratory Animals [18]. Fourteen rats were randomized to 10-day water avoidance stress (WAS, n = 8, 12.3 +/- 0.4 weeks of age) or handled controls (n = 8, 13.3 +/- 0.4 weeks of age).

### Chronic psychological water avoidance stress

The current study used a validated rat model of chronic psychological stress (chronic water avoidance, WAS) which leads to bladder dysfunction characterized by frequent, small volume voids, bladder hyperalgesia, and increased nociceptive reflex responses to bladder distension [16,17,19]. A standing cube ( $6 \ge 0.5 \text{ cm}^3$ ) was placed in the middle of the water tank ( $25 \ge 0.5 \text{ cm}^3$ ). The water was filled up to 2 cm away from the top. The WAS rat was placed on top of the cube with the water at room temperature for 1 hour every day for 10 consecutive days. The control rat was handled and kept in its housing cage in the experimental room for 1 hour.

## Surgical procedures

On day 11, under isoflurane anesthesia (1–2% in the gas mixture of 30% oxygen and 70% nitrogen) animals received a percutaneous venous catheter, placed in the right external jugular vein.  $\alpha$ -chloralose (40 mg/ml, Sigma Aldrich, St. Louis, MO, USA) was dissolved in 20%  $\beta$ -cyclodextrin (MW 1380, Sigma Aldrich) and administered intravascularly as a bolus at 40 mg/kg. The isoflurane anesthesia was maintained at 1% for 30 minutes to allow the  $\alpha$ -chloralose to take effect. To obtain the visceromotor response (VMR), two fine and insulated silver wire electrodes (0.05 mm diameter, A-M Systems, Everett, WA, USA) with exposed tips were embedded into the left abdominal external oblique muscle (EOM). The sampling rate was set to 1kHz for both electromyography (EMG) channels. A PE-50 catheter was inserted into the bladder through the urethra outlet. The bladder catheter was connected to a 3-way connector with a syringe pump and a pressure sensor to infuse normal saline and obtain a cystometro-gram (CMG).

After all surgical procedures were completed, animals received an infusion of  $\alpha$ -chloralose (15 mg/kg/hr), sufficient to provide continuous conscious sedation to the animals. We chose light sedation using  $\alpha$ -chloralose because of its favorable profile in maintaining cerebral function intact [20] as compared to isoflurane [21,22] or urethane [23]. At the same time,  $\alpha$ -chloralose when given at a low dose (15 mg/kg/hr) allows assessment of bladder urodynamics and the micturition reflex in rats [24].

## Measurement of CMG and VMR recordings

CMG and VMR were obtained 1 hour after discontinuation of isoflurane anesthesia with animals maintained on a heating pad (37.0 °C) under light sedation with  $\alpha$ -chloralose (15 mg/kg/ hr, i.v.). The bladder was infused with normal saline (0.9%, 0.1 mL/min) at room temperature with an open urethral outlet, thus allowing the animal to void. CMG and VMR were recorded by a data acquisition system (MP150; Biopac Systems Inc., Goleta, CA, USA) and stored on a computer for data analysis. CMG and VMR were also obtained during isotonic bladder distention (0  $\rightarrow$  10  $\rightarrow$  20  $\rightarrow$  30  $\rightarrow$  40 cmH<sub>2</sub>O) with urethral occlusion.

During the continuous bladder infusion, the latency of the first void/leak (LFV) was measured as the latency from the beginning of bladder filling to occurrence of the first void/leak. The bladder capacity (mL) was calculated as LFV (min) × infusion rate (0.1 mL/min). The maximum intravesical pressure (IVPmax), and pressure threshold to first void/leak (PT) were obtained. These parameters were calculated on first void to ensure the bladder was completely emptied (not carrying a residual volume) at the beginning of the continuous bladder infusion and CMG measurements. The bladder pressure evoked VMR was referred to as VMR threshold (cmH<sub>2</sub>O). The normalized ratio of VMR threshold/IVPmax was calculated for each animal. During bladder distension, a 25-second duration of VMR area under curve (AUC) was measured at each level of bladder pressure from 0 to 40 cmH<sub>2</sub>O.

## Functional brain mapping

Sixty minutes after completion of the VMR/CMG studies and two hours after discontinuation of the isoflurane, rats were connected via their venous catheter to a cannula extension and a mechanical infusion pump. Light sedation was maintained with  $\alpha$ -chloralose (15 mg/kg/hr, i. v.). Rats received a final isotonic bladder distension (20 cmH<sub>2</sub>O). Forty-five seconds after initiating the distension, and in the absence of any voiding (confirmed by absence of a VMR), [<sup>14</sup>C]-iodoantipyrine (100 µCi/kg, Am. Radiolabelled Chem., St. Louis, MO, USA) was injected as an i.v. bolus, followed immediately by injection of the euthanasia solution (50 mg/kg pentobarbital/3M KCl). This resulted in cardiac arrest within ~5-10s and provided the temporal resolution of the brain mapping method [25].

Brains were flash frozen and serially sectioned (60 coronal 20- $\mu$ m slices, 300- $\mu$ m interslice distance). Autoradiographic images of brain slices along with 12 [<sup>14</sup>C] standards (GE Health-care Life Sciences, Pittsburgh, PA, USA) were digitized. Regional cerebral blood flow related tissue radioactivity (rCBF) was measured by the classic [<sup>14</sup>C]-iodoantipyrine method [26–28]. In this method, there is a strict linear proportionality between tissue radioactivity and CBF when the data is captured within a brief interval (~10 seconds) after the tracer injection [29,30].

#### Statistical analysis

**Urodynamic studies.** For CMG and VMR during bladder infusion, data is expressed as means and standard errors (means  $\pm$  SE). Unpaired t-tests were performed to compare control and WAS groups. A value of P < 0.05 indicated a significant difference between groups. GraphPad Prism5 (GraphPad Software Inc., LaJolla, CA, USA) was used for the statistical analysis and graph demonstration.

**Functional brain mapping.** <u>Image processing:</u> Each 3-D brain was reconstructed from 57 digitized autoradiograms (voxel size:  $40 \times 300 \times 40 \ \mu m^3$ ) using TurboReg [31] and a nonwarping geometric model that includes rotations and translations and nearest-neighbor interpolation. A brain template was created using our prior published methods [32], in which the background and ventricular spaces were thresholded based on their optical density. Spatial normalization consisted of a 12-parameter affine transformation followed by a nonlinear spatial normalization using 3-D discrete cosine transforms. Normalized brains were smoothed with a Gaussian kernel (FWHM = 3 x voxel dimension). Absolute amount of radiotracer of each brain was scaled to a single mean.

<u>Analysis of regional brain activation</u>: Statistical parametric mapping (SPM) [33] was used to identify regions showing significant rCBF changes. Significance was set at P<0.05 at the voxel level and an extent threshold of 100 significant, contiguous voxels to control Type I and Type II errors. In addition, we verified the SPM results with a a region-of-interest analysis. Regions of interest (ROI) were drawn manually for regions within the micturition circuit. Regions were identified by a rat brain atlas [34]. A functional ROI was created by combining the anatomical ROI with its significant SPM cluster by logical conjunction. Mean optical density was extracted for the ROI using the Marsbar toolbox [35]. The directionality of rCBF changes was analyzed with Student's t-tests (P<0.05).

Seed correlation analysis: Functional ROIs as defined above for the right posterior cingulate cortex, the right Barrington nucleus, as well as the right posterior insula underwent a seed correlation analysis. Seed correlation was visualized in SPM across the whole brain, with color-coded correlation coefficients (P < 0.05, extent threshold > 100 significant, contiguous voxels).

Additional supporting information is also available through the Open Science Framework (OSF; <u>http://osf.io</u>), a data sharing network.

## Results

#### CMG and VMR recordings

During voiding, WAS animals showed a significant decreased PT triggering the voiding phase (P = 0.04, Fig 1D). The VMR evoked by bladder distension in WAS animals appeared at a lower IVP during bladder filling compared to controls (P = 0.01, Fig 1E). The percentage of VMR threshold pressure/IVPmax also showed the VMR to appear significantly earlier in WAS animals compared to controls (P = 0.01, Fig 1F). All these are surrogates for bladder hypersensitivity. The maximum IVP and bladder capacity showed no significant changes between control and WAS groups.

During isotonic bladder distension, VMR AUC was measured from both experimental groups (Fig 2A). At 0 cmH<sub>2</sub>O, both control and WAS groups did not have the appearance of VMR. When the isotonic pressure reached 10 cmH<sub>2</sub>O, few VMRs were evoked in both groups. At 20 cmH<sub>2</sub>O, VMR AUC significantly increased in WAS animals compared to controls (P = 0.03, Fig 2B). At 30 cmH<sub>2</sub>O the VMR AUC remained increased in WAS compared to control animals, with no group difference (ceiling effect) noted at 40 cmH<sub>2</sub>O. In summary, WAS rats compared to control rats showed lower VMR thresholds and pressure thresholds, and greater VMR responses to passive bladder distension.

## Functional brain mapping

Functional brain activation during moderate bladder distension ( $20 \text{ cmH}_2\text{O}$ ) showed significantly greater rCBF (P<0.05, min. cluster size 100 voxels) in WAS compared to control rats, most broadly in posterior cingulate, anterior retrosplenial, somatosensory (primary, secondary) and posterior insular cortices (right > left), and the anterior thalamus (anterior ventral, lateral dorsal, mediodorsal, paraventricular) and lateral habenula. Significant increases in rCBF were also noted in anterior, secondary motor cortex, medial prefrontal (orbital, prelimbic) and caudomedial entorhinal cortices, as well as the dorsomedial striatum, anterior hypothalamus, superior colliculus, median raphe, and parabrachial/Barrington nucleus complex. Significant decreases in rCBF were noted in the dorsal hippocampus, posterior hypothalamus, amygdala and cerebellar cortex (Fig 3). SPM results were confirmed by ROI analysis. Fig 4 shows additional detail for the results of the SPM analysis and definition of the ROIs at the level of the pons. More specifically ROI analysis of Barrington's nucleus and the parabrachial nucleus showed significant group differences (P<0.05, min. cluster size 100 voxels) when ROIs were examined individually or when they were pooled. Significant changes in rCBF as they relate to the micturition circuit are summarized in Fig 5.

Because of the broad and significant group difference noted in the posterior cingulate, this region was used in a seed correlation analysis. This showed significant positive correlation (P<0.005, min. cluster size 100 voxels) with the parabrachial nucleus in WAS animals but not in controls (Fig 6). Seed correlation of the right Barrington nucleus showed a significant positive correlation with midline cortex (mid/posterior cingulate, anterior retrosplenial), as well as somatosensory cortex (primary, secondary) and anterior secondary motor cortex in WAS



**Fig 1. Statistical analysis of cystometrogram and visceromotor reflex (VMR +/- standard error) recordings during bladder infusion.** Shown are the tracings of (A) cystometrogram, (B) IVPmax (maximum intravesical pressure), (C) bladder capacity, (D) pressure threshold to first void/leak, (E) VMR threshold, and (F) VMR threshold/IVPmax for WAS and control animals. \* indicated when voiding happened. The bar on the vertical axis of Figure (A) is a calibration bar, whose length represents a bladder pressure of 40 cmH<sub>2</sub>O.

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# A. VMR during isotonic bladder distension (IBD)

Fig 2. (A) Representative examples of visceromotor reflex (VMR) in the control and WAS animals at each level of bladder pressure between 0 and 40 cmH<sub>2</sub>O. (B) During isotonic bladder distension, VMR AUC (VMR +/- standard error) in WAS significantly increased compared to controls at 20 cmH<sub>2</sub>O (P = 0.03). VMR: visceromotor reflex. AUC: area under the curve. Note different scales used at different pressure levels. WAS: water avoidance stress.

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animals but not in controls. Subcortically WAS rats, but not control animals showed a significant positive functional connectivity to the anterior, dorsal midline thalamus (mediodorsal, paraventricular), ventrobasilar thalamic complex, lateral habenula, dorsolateral striatum, and rostral periaqueductal gray. A significant negative correlation was noted with the posterior insula in WAS rats but not in controls. Seed correlation of the posterior insula showed significant positive correlations with anterior, secondary motor cortex (M2) in WAS animals but not in controls (Fig 6).

#### Discussion

In 2008, the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) initiated the Multi-disciplinary Approach to the Study of Chronic Pelvic Pain (MAPP) Research Network to rigorously study urological chronic pelvic pain syndrome (UCPPS) using an interdisciplinary approach [38]. UCPPS include IC/BPS and chronic prostatitis/chronic pelvic pain

# rCBF (% of global mean CBF)



Fig 3. Differences in functional brain activation during bladder filling of WAS rats compared to controls. Left column: Statistical parametric maps showing significant difference (P < 0.05 for clusters of greater than 100 contiguous significant voxels) in brain activation in sedated WAS rats compared to controls are superimposed on gray-scale coronal sections showing the rCBF distribution of the template brain. **Right column:** ROI analysis shows group mean rCBF (± S.E.) for select brain regions in the micturition circuit, in which rCBF in each animal was normalized by its whole brain mean perfusion. \* P < 0.05, \*\*P < 0.005 for clusters of greater than 100 significant, contiguous voxels. Abbreviations: Cx (cortex), PAG (periaqueductal gray), PB (parabrachial nucleus).

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syndrome (CP/CPPS). Part of the MAPP network has focused on the validation of rodent models that reflect multiple key characteristics of human UCPPS and provide enhanced clinical significance to mechanistic studies. Strategies for evaluating animal models of UCPPS were developed [39]. The WAS model is one such model which reflects the stress variability seen in the human conditions and manifests the key features of urinary frequency and bladder hyperalgesia experienced by patients [17].





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Recent work by Deutsch et al. [9] demonstrated that female interstitial cystitis subjects compared to healthy controls show increased rCBF during bladder distension in multiple areas of the brain associated with visceral sensory perception. Significant increases were noted in the supplemental motor area (SMA), motor and sensory cortex, the insula, the hippocampus and the middle and posterior cingulate. In light of prior epidemiologic evidence suggesting a significant association of IC/BPS with stress, poor stress coping, and symptoms of anxiety and depression [5,10–15], our study's aim was to prospectively examine in an animal model if chronic stress exposure could result in a functional reorganization of regions within the pain matrix similar to that noted in IC patients.

A rodent model of chronic psychological stress was investigated by CMG recordings of VMR, as well as the brain activity during isotonic bladder distension. WAS animals demonstrated a VMR that appeared earlier and more forcefully than in controls. A decreased PT in WAS animals indicated that the voiding appeared early, indicating bladder hypersensitivity.

Control

Seed ROI



WAS

Fig 6. Seed correlation of functional activity across the whole brain during bladder filling. WAS compared to control rats show greater functional connectivity of (row 1) the posterior cingulate to the parabrachial/Barrington nucleus complex, (row 2) the posterior insula to anterior secondary motor cortex (M2), and (row 3) the Barrington nucleus to the anterior, dorsal midline thalamus (mediodorsal, MD; paraventricular, PVN), as well as the ventrobasilar thalamic complex, primary and secondary somatosensory cortex (S1, S2) and retrosplenial cortex.

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Because the greatest group difference in the CMG studies was observed at  $20 \text{ cmH}_2\text{O}$ , we used this moderate physiologic stimulus during the functional brain mapping.

Bladder distension increased functional brain activation in WAS compared to control rats broadly across the supraspinal micturition circuit (Fig 5) (reviewed in [40,41]). According to this simplified model of supraspinal bladder control, afferent input from the bladder and urethra are conveyed to the periaqueductal gray (PAG) and transmitted further to the insula, where the sensations are mapped. The cingulate cortex is involved in monitoring and evaluating bladder sensations, and may control micturition via its efferent output to the PAG and, indirectly, to the pontine micturition center, including the Barrington's nucleus and the parabrachial nucleus. The prefrontal cortex is involved in attention to bladder sensations and decisions about voiding. In addition, the SMA and the medial primary motor cortex (M1) are regions that control voluntary contraction and relaxation of the pelvic floor muscles via efferent projections to motor neurons in the sacral spinal cord. In our study, significant stress effects were noted in most of the brain regions previously noted by Tai et al. (2009) to be responsive to passive bladder distension. These authors imaging normal, anesthetized rats during bladder filling using fMRI, reported brain activation in motor cortex, cingulate and retrosplenial cortex, primary and secondary somatosensory cortex, the putamen, septum, thalamus, posterior insula, as well as the periaqueductal gray (PAG) [42]. Equivalent brain regions have also been reported to be activated in healthy volunteers [43] and in IC subjects [9] in the context of a full bladder. In addition, our analysis revealed that WAS animals compared to controls demonstrated a significant increase in rCBF in Barrington's nucleus, as well as the adjacent parabrachial nucleus, both representing final relay regions of the micturition circuit.

One of the broadest and most significant group differences in our brain mapping was noted in the posterior cingulate/anterior retrosplenial cortex. Seed analysis for this region revealed that WAS rats compared to controls showed greater functional connectivity of the posterior cingulate to the parabrachial nucleus that functions as a relay of the supraspinal modulation of micturition [44-47]. The function of posterior cingulate cortex remains unclear at present. It is a central node in the default mode network (DMN), with widespread and heterogeneous functional connectivity across the brain, and with proposed roles in attentional control, cognition, pain perception, emotional processing, sensory integration, including bladder filling and urinary urgency [48,49]. Recent work in patients with UCPPS has shown that the posterior cingulate is functionally disconnected at rest from the DMN, and instead shows increased functional connectivity to cerebral regions that play a role in pain, sensory, motor and affective processes, including the insula, putamen, amygdala, and hippocampus [50]. Seed analysis for Barrington's nucleus, a final relay in micturition (reviewed in [41]), showed that WAS animals compared to controls showed a significant and broader positive functional connectivity with the medial thalamus (mediodorsal, paraventricular), associated with limbic modulation [51], as well as with a ventrobasilar thalamic complex, associated with sensory integration [52,53]. Prior work has shown structural connectivity of Barrington's nucleus to this medial thalamic region [54,55], and functional connections to a ventrobasilar thalamic region [56].

A primary brain region implicated in functional neuroimaging of chronic pain is the insula [57-59]. The insula in the rodent is currently understood to have a posterior-to-mid-to-anterior integration of interoceptive information [60]. Primary interoceptive representations of viscerosensory information are believed to be dominantly represented in the posterior insula, while the anterior insula is believed to integrate such visceral inputs with cognitive and affective aspects of subjective awareness. A role for the posterior insula has been proposed in relaying necessary sensory information to motor cortical structures for control of muscles associated with the viscera [61-63]. In our study, examining the posterior insula during bladder filling in sedated animals, WAS rats compared to controls showed greater functional connectivity of the posterior insula to rostral midline motor cortex (M2), as well as somatosensory cortex. Rostral motor cortex in the rodent has been proposed to be the equivalent of the supplementary motor area (SMA) of primates [64-66], and represents a cortical region associated with pelvic floor and gluteal muscle activation [67–71]. Our results are relevant to recent work by Kutch et al. (2015) that suggests an important role for brain motor control in disorders of chronic pelvic pain. Specifically, in male patients with chronic prostatitis/chronic pelvic pain syndrome (CP/CPPS), there are alterations in resting state functional connectivity between the supplementary motor cortex and the posterior insula [72]. Furthermore, women with interstitial cystitis/painful bladder syndrome show altered frequency distributions in intrinsic brain oscillations in viscerosensory (posterior insula), motor regions (including the medial and ventral supplementary motor areas), as well as somatosensory cortex (postcentral gyrus) [73]. Our study suggests that chronic stress may alter functional connectivity between posterior insular cortex and sensorimotor cortex.

In light of the greater prevalence and severity and symptom expression of IC/BPS in women as compared to men, the focus of our study was on female rats [74]. Past brain mapping in healthy volunteers, however, has suggested sex differences during "attempted

micturition", with increased responses in women than in men in the right thalamus and in subregions of the right prefrontal cortex [43], with others reporting greater activity of the right insula and operculum in women than in men during a filled bladder condition [75]. Such findings have been hypothesized to reflect sex differences of a 'salience network' [36], part of a larger 'interoceptive network' [43]. In anesthetized rats, visceromotor responses to bladder distention show more vigorous reflex responses in females than in males [76]. Our own work in mapping brain function during visceral pain in awake rats has also noted sex differences in the thalamus, amygdala and insula during acute colorectal distension [77]. Future studies will need to examine if such sex differences in brain responses are also noted during bladder filling in the animal model. A limitation of our study is that since we did not include a "no bladder distension" group, we cannot determine if the significant differences in rCBF noted in response to WAS are specific to the physiologic challenge of bladder filling, or if they are also apparent in the baseline condition. Collectively, our physiologic and brain mapping results suggest a hypersensitivity in WAS animals, as well as increased engagement of portions of the micturition circuit responsive to urgency and the perception of bladder fullness, including viscerosensory perception and its relay to motor regions coordinating imminent bladder contraction. A subset of the brain regions demonstrating group differences in our study are wellknown to serve multiple functions. In particular, regions including the cingulate, insula, and periaqueductal gray are described not only within the micturition circuit, but also as core regions within the pain neuromatrix [78], as well as in circuits mediating fear, mood and stress [79,80]. This is consistent with our understanding of the multiple sensory and affective/motivational aspects of pain [81].

#### **Author Contributions**

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#### References

- Lim JR, Bak CW, Lee JB. Comparison of anxiety between patients with mixed incontinence and those with stress urinary incontinence. Scand J Urol Nephrol. 2007; 41: 403–406. <u>https://doi.org/10.1080/ 00365590701303702</u> PMID: 17853035
- Macaulay AJ, Stern RS, Holmes DM, Stanton SL. Micturition and the mind: psychological factors in the aetiology and treatment of urinary symptoms in women. Br Med J (Clin Res Ed). 1987; 294: 540–543.
- 3. Mayer EA, Naliboff BD, Chang L, Coutinho SVV. Stress and irritable bowel syndrome. Am J Physiol Gastrointest Liver Physiol. 2001; 280: G519–524. PMID: <u>11254476</u>
- Rogers R, Bachmann G, Jumadilova Z, Sun F, Morrow JD, Guan Z, et al. Efficacy of tolterodine on overactive bladder symptoms and sexual and emotional quality of life in sexually active women. Int Urogynecol J Pelvic Floor Dysfunct. 2008; 19: 1551–1557. https://doi.org/10.1007/s00192-008-0688-6 PMID: 18685795

- Rothrock NE, Lutgendorf SK, Kreder KJ, Ratliff T, Zimmerman B. Stress and symptoms in patients with interstitial cystitis: a life stress model. Urology. 2001; 57: 422–427. PMID: 11248609
- Cagnie B, Coppieters I, Denecker S, Six J, Danneels L, Meeus M. Central sensitization in fibromyalgia? A systematic review on structural and functional brain MRI. Semin Arthritis Rheum. 2014; 44: 68–75. https://doi.org/10.1016/j.semarthrit.2014.01.001 PMID: 24508406
- Hampson JP, Reed BD, Clauw DJ, Bhavsar R, Gracely RH, Haefner HK, et al. Augmented central pain processing in vulvodynia. J Pain. 2013; 14: 579–589. <u>https://doi.org/10.1016/j.jpain.2013.01.767</u> PMID: 23578957
- Mayer EA, Gupta A, Kilpatrick LA, Hong JY. Imaging brain mechanisms in chronic visceral pain. Pain. 2015; 156 Suppl 1: S50–63.
- Deutsch G, Deshpande H, Frolich MA, Lai HH, Ness TJ. Bladder Distension Increases Blood Flow in Pain Related Brain Structures in Subjects with Interstitial Cystitis. J Urol. 2016; 196: 902–910. https:// doi.org/10.1016/j.juro.2016.03.135 PMID: 27018508
- Bullones Rodriguez MA, Afari N, Buchwald DS, National Institute of D, Digestive, Kidney Diseases Working Group on Urological Chronic Pelvic P. Evidence for overlap between urological and nonurological unexplained clinical conditions. J Urol. 2013; 189: S66–74. <u>https://doi.org/10.1016/j.juro.2012.11</u>. 019 PMID: 23234637
- Lutgendorf SK, Kreder KJ, Rothrock NE, Ratliff TL, Zimmerman B. Stress and symptomatology in patients with interstitial cystitis: a laboratory stress model. J Urol. 2000; 164: 1265–1269. PMID: 10992377
- Nickel JC, Shoskes D, Irvine-Bird K. Clinical phenotyping of women with interstitial cystitis/painful bladder syndrome: a key to classification and potentially improved management. J Urol. 2009; 182: 155– 160. https://doi.org/10.1016/j.juro.2009.02.122 PMID: 19447429
- Nickel JC, Tripp DA, Pontari M, Moldwin R, Mayer R, Carr LK, et al. Interstitial cystitis/painful bladder syndrome and associated medical conditions with an emphasis on irritable bowel syndrome, fibromyalgia and chronic fatigue syndrome. J Urol. 2010; 184: 1358–1363. https://doi.org/10.1016/j.juro.2010. 06.005 PMID: 20719340
- Nickel JC, Tripp DA, Pontari M, Moldwin R, Mayer R, Carr LK, et al. Psychosocial phenotyping in women with interstitial cystitis/painful bladder syndrome: a case control study. J Urol. 2010; 183: 167– 172. https://doi.org/10.1016/j.juro.2009.08.133 PMID: 19913812
- Rothrock NE, Lutgendorf SK, Kreder KJ, Ratliff TL, Zimmerman B. Daily stress and symptom exacerbation in interstitial cystitis patients. Urology. 2001; 57: 122.
- Lee UJ, Ackerman AL, Wu A, Zhang R, Leung J, Bradesi S, et al. Chronic psychological stress in highanxiety rats induces sustained bladder hyperalgesia. Physiol Behav. 2015; 139: 541–548. <u>https://doi.org/10.1016/j.physbeh.2014.11.045</u> PMID: 25449389
- Smith AL, Leung J, Kun S, Zhang R, Karagiannides I, Raz S, et al. The effects of acute and chronic psychological stress on bladder function in a rodent model. Urology. 2011; 78: 967 e961–967.
- National Research Council (US) Committee for the Update of the Guide for the Care and Use of Laboratory Animals. Guide for the Care and Use of Laboratory Animals. Washington, DC: National Academies Press. 2011.
- Ackerman AL, Jellison FC, Lee UJ, Bradesi S, Rodriguez LV. Glt1 glutamate receptor mediates the establishment and perpetuation of chronic visceral pain in an animal model of stress-induced bladder hyperalgesia. Am J Physiol Renal Physiol. 2015; 310(7):F628–F636. <u>https://doi.org/10.1152/ajprenal.</u> 00297.2015 PMID: 26697981
- Baek K, Shim WH, Jeong J, Radhakrishnan H, Rosen BR, Boas D, et al. Layer-specific interhemispheric functional connectivity in the somatosensory cortex of rats: resting state electrophysiology and fMRI studies. Brain Struct Funct. 2016; 221: 2801–2815. <u>https://doi.org/10.1007/s00429-015-1073-0</u> PMID: 26077581
- Kalthoff D, Po C, Wiedermann D, Hoehn M. Reliability and spatial specificity of rat brain sensorimotor functional connectivity networks are superior under sedation compared with general anesthesia. NMR Biomed. 2013; 26: 638–650. https://doi.org/10.1002/nbm.2908 PMID: 23303725
- Magnuson ME, Thompson GJ, Pan WJ, Keilholz SD. Time-dependent effects of isoflurane and dexmedetomidine on functional connectivity, spectral characteristics, and spatial distribution of spontaneous BOLD fluctuations. NMR Biomed. 2014; 27: 291–303. <u>https://doi.org/10.1002/nbm.3062</u> PMID: 24449532
- Martin C, Martindale J, Berwick J, Mayhew J. Investigating neural-hemodynamic coupling and the hemodynamic response function in the awake rat. Neuroimage. 2006; 32: 33–48. https://doi.org/10. 1016/j.neuroimage.2006.02.021 PMID: 16725349

- Zeng J, Pan C, Jiang C, Lindstrom S. Cause of residual urine in bladder outlet obstruction: an experimental study in the rat. J Urol. 2012; 188: 1027–1032. https://doi.org/10.1016/j.juro.2012.04.101 PMID: 22819110
- Holschneider DP, Maarek JM, Harimoto J, Yang J, Scremin OU. An implantable bolus infusion pump for use in freely moving, nontethered rats. Am J Physiol Heart Circ Physiol. 2002; 283: H1713–1719. https://doi.org/10.1152/ajpheart.00362.2002 PMID: 12234827
- Goldman H, Sapirstein LA. Brain blood flow in the conscious and anesthetized rat. Am J Physiol. 1973; 224: 122–126. PMID: 4566847
- Patlak CS, Blasberg RG, Fenstermacher JD. An evaluation of errors in the determination of blood flow by the indicator fractionation and tissue equilibration (Kety) methods. J Cereb Blood Flow Metab. 1984; 4: 47–60. https://doi.org/10.1038/jcbfm.1984.7 PMID: 6363433
- 28. Sakurada O, Kennedy C, Jehle J, Brown JD, Carbin GL, Sokoloff L. Measurement of local cerebral blood flow with iodo [14C] antipyrine. Am J Physiol. 1978; 234: H59–66. PMID: 623275
- Jones SC, Korfali E, Marshall SA. Cerebral blood flow with the indicator fractionation of [14C]iodoantipyrine: effect of PaCO2 on cerebral venous appearance time. J Cereb Blood Flow Metab. 1991; 11: 236–241. https://doi.org/10.1038/jcbfm.1991.55 PMID: 1900067
- Van Uitert RL, Levy DE. Regional brain blood flow in the conscious gerbil. Stroke. 1978; 9: 67–72. PMID: 341424
- Thevenaz P, Ruttimann UE, Unser M. A pyramid approach to subpixel registration based on intensity. IEEE Trans Image Process. 1998; 7: 27–41. https://doi.org/10.1109/83.650848 PMID: 18267377
- Nguyen PT, Holschneider DP, Maarek JM, Yang J, Mandelkern MA. Statistical parametric mapping applied to an autoradiographic study of cerebral activation during treadmill walking in rats. Neuroimage. 2004; 23: 252–259. https://doi.org/10.1016/j.neuroimage.2004.05.014 PMID: 15325372
- **33.** Friston K and the Methodology group at the Wellcome Dept.of Cognitive Neurology,. Statistical Parametric Mapping (SPM99). http://wwwfilionuclacuk/spm/. 1999.
- 34. Paxinos G, Watson C. The Rat Brain in Stereotactic Coordinates. New York: Elsevier Academic Press. 2007.
- **35.** Brett M, Anton J-L, Valabregue R, Poline J-B. Region of interest analysis using an SPM toolbox; June, 2002; Sendai, Japan. NeuroImage.
- Griffiths D. Functional imaging of structures involved in neural control of the lower urinary tract. Handb Clin Neurol. 2015; 130: 121–133. <u>https://doi.org/10.1016/B978-0-444-63247-0.00007-9</u> PMID: 26003241
- de Groat WC, Yoshimura N. Anatomy and physiology of the lower urinary tract. Handb Clin Neurol. 2015; 130: 61–108. https://doi.org/10.1016/B978-0-444-63247-0.00005-5 PMID: 26003239
- Clemens JQ, Mullins C, Kusek JW, Kirkali Z, Mayer EA, Rodriguez LV, et al. The MAPP research network: a novel study of urologic chronic pelvic pain syndromes. BMC Urol. 2014; 14: 57. https://doi.org/ 10.1186/1471-2490-14-57 PMID: 25085007
- Lai H, Gereau RWt, Luo Y, O'Donnell M, Rudick CN, Pontari M, et al. Animal Models of Urologic Chronic Pelvic Pain Syndromes: Findings From the Multidisciplinary Approach to the Study of Chronic Pelvic Pain Research Network. Urology. 2015; 85: 1454–1465. https://doi.org/10.1016/j.urology.2015.03.007 PMID: 26099889
- Griffiths D, Derbyshire S, Stenger A, Resnick N. Brain control of normal and overactive bladder. J Urol. 2005; 174: 1862–1867. https://doi.org/10.1097/01.ju.0000177450.34451.97 PMID: 16217325
- **41.** de Groat WC, Wickens C. Organization of the neural switching circuitry underlying reflex micturition. Acta Physiol (Oxf). 2013; 207: 66–84.
- Tai C, Wang J, Jin T, Wang P, Kim SG, Roppolo JR, et al. Brain switch for reflex micturition control detected by FMRI in rats. J Neurophysiol. 2009; 102: 2719–2730. https://doi.org/10.1152/jn.00700. 2009 PMID: 19741099
- Kuhtz-Buschbeck JP, Gilster R, van der Horst C, Hamann M, Wolff S, Jansen O. Control of bladder sensations: an fMRI study of brain activity and effective connectivity. Neuroimage. 2009; 47: 18–27. https://doi.org/10.1016/j.neuroimage.2009.04.020 PMID: 19371782
- Lumb BM, Morrison JF. An excitatory influence of dorsolateral pontine structures on urinary bladder motility in the rat. Brain Res. 1987; 435: 363–366. PMID: 3427465
- Kruse MN, Noto H, Roppolo JR, de Groat WC. Pontine control of the urinary bladder and external urethral sphincter in the rat. Brain Res. 1990; 532: 182–190. PMID: 2282512
- 46. Chen SY, Wang SD, Cheng CL, Kuo JS, De Groat WC, Chai CY. Glutamate activation of neurons in CV-reactive areas of cat brain stem affects urinary bladder motility. Am J Physiol. 1993; 265: F520– 529. PMID: 7902000

- Mallory BS, Roppolo JR, de Groat WC. Pharmacological modulation of the pontine micturition center. Brain Res. 1991; 546: 310–320. PMID: 1676929
- Nardos R, Gregory WT, Krisky C, Newell A, Nardos B, Schlaggar B, et al. Examining mechanisms of brain control of bladder function with resting state functional connectivity MRI. Neurourol Urodyn. 2014; 33: 493–501. https://doi.org/10.1002/nau.22458 PMID: 23908139
- Tadic SD, Griffiths D, Schaefer W, Cheng CI, Resnick NM. Brain activity measured by functional magnetic resonance imaging is related to patient reported urgency urinary incontinence severity. J Urol. 2010; 183: 221–228. https://doi.org/10.1016/j.juro.2009.08.155 PMID: 19913803
- Martucci KT, Shirer WR, Bagarinao E, Johnson KA, Farmer MA, Labus JS, et al. The posterior medial cortex in urologic chronic pelvic pain syndrome: detachment from default mode network-a resting-state study from the MAPP Research Network. Pain. 2015; 156: 1755–1764. https://doi.org/10.1097/j.pain. 00000000000238 PMID: 26010458
- Vertes RP, Linley SB, Hoover WB. Limbic circuitry of the midline thalamus. Neurosci Biobehav Rev. 2015; 54: 89–107. https://doi.org/10.1016/j.neubiorev.2015.01.014 PMID: 25616182
- Jung SC, Kim JH, Choi IS, Cho JH, Bae YC, Lee MG, et al. Corticothalamic modulation on formalininduced change of VPM thalamic activities. Neuroreport. 2004; 15: 1405–1408. PMID: 15194862
- Monconduit L, Bourgeais L, Bernard JF, Le Bars D, Villanueva L. Ventromedial thalamic neurons convey nociceptive signals from the whole body surface to the dorsolateral neocortex. J Neurosci. 1999; 19: 9063–9072. PMID: 10516323
- Otake K, Nakamura Y. Single neurons in Barrington's nucleus projecting to both the paraventricular thalamic nucleus and the spinal cord by way of axon collaterals: a double labeling study in the rat. Neurosci Lett. 1996; 209: 97–100. PMID: 8761991
- 55. Valentino RJ, Pavcovich LA, Hirata H. Evidence for corticotropin-releasing hormone projections from Barrington's nucleus to the periaqueductal gray and dorsal motor nucleus of the vagus in the rat. J Comp Neurol. 1995; 363: 402–422. https://doi.org/10.1002/cne.903630306 PMID: 8847408
- 56. Robbins MT, Uzzell TW, Aly S, Ness TJ. Characterization of thalamic neuronal responses to urinary bladder distention, including the effect of acute spinal lesions in the rat. J Pain. 2006; 7: 218–224. https://doi.org/10.1016/j.jpain.2005.10.012 PMID: 16516828
- Apkarian AV, Bushnell MC, Treede RD, Zubieta JK. Human brain mechanisms of pain perception and regulation in health and disease. European Journal of Pain: Ejp. 2005; 9: 463–484. <u>https://doi.org/10. 1016/j.ejpain.2004.11.001</u> PMID: 15979027
- Peyron R, Laurent B, Garcia-Larrea L. Functional imaging of brain responses to pain. A review and meta-analysis (2000). Neurophysiol Clin. 2000; 30: 263–288. PMID: <u>11126640</u>
- 59. Sadler KE, Kolber BJ. Urine trouble: Alterations in brain function associated with bladder pain. J Urol. 2016.
- 60. Craig AD. Significance of the insula for the evolution of human awareness of feelings from the body. Ann N Y Acad Sci. 2011; 1225: 72–82. https://doi.org/10.1111/j.1749-6632.2011.05990.x PMID: 21534994
- Cauda F, D'Agata F, Sacco K, Duca S, Geminiani G, Vercelli A. Functional connectivity of the insula in the resting brain. Neuroimage. 2011; 55: 8–23. https://doi.org/10.1016/j.neuroimage.2010.11.049 PMID: 21111053
- Deen B, Pitskel NB, Pelphrey KA. Three systems of insular functional connectivity identified with cluster analysis. Cereb Cortex. 2011; 21: 1498–1506. https://doi.org/10.1093/cercor/bhq186 PMID: 21097516
- Levinthal DJ, Strick PL. The motor cortex communicates with the kidney. J Neurosci. 2012; 32: 6726– 6731. https://doi.org/10.1523/JNEUROSCI.0406-12.2012 PMID: 22573695
- Neafsey EJ, Bold EL, Haas G, Hurley-Gius KM, Quirk G, Sievert CF, et al. The organization of the rat motor cortex: a microstimulation mapping study. Brain Res. 1986; 396: 77–96. PMID: <u>3708387</u>
- Rouiller EM, Moret V, Liang F. Comparison of the connectional properties of the two forelimb areas of the rat sensorimotor cortex: support for the presence of a premotor or supplementary motor cortical area. Somatosens Mot Res. 1993; 10: 269–289. PMID: 8237215
- Sievert CF, Neafsey EJ. A chronic unit study of the sensory properties of neurons in the forelimb areas of rat sensorimotor cortex. Brain Res. 1986; 381: 15–23. PMID: 3530375
- Zhang H, Reitz A, Kollias S, Summers P, Curt A, Schurch B. An fMRI study of the role of suprapontine brain structures in the voluntary voiding control induced by pelvic floor contraction. Neuroimage. 2005; 24: 174–180. https://doi.org/10.1016/j.neuroimage.2004.08.027 PMID: 15588608
- Di Gangi Herms AM, Veit R, Reisenauer C, Herms A, Grodd W, Enck P, et al. Functional imaging of stress urinary incontinence. Neuroimage. 2006; 29: 267–275. https://doi.org/10.1016/j.neuroimage. 2005.07.018 PMID: 16150613

- Seseke S, Baudewig J, Kallenberg K, Ringert RH, Seseke F, Dechent P. Voluntary pelvic floor muscle control—an fMRI study. Neuroimage. 2006; 31: 1399–1407. https://doi.org/10.1016/j.neuroimage. 2006.02.012 PMID: 16574434
- Kuhtz-Buschbeck JP, van der Horst C, Wolff S, Filippow N, Nabavi A, Jansen O, et al. Activation of the supplementary motor area (SMA) during voluntary pelvic floor muscle contractions—an fMRI study. Neuroimage. 2007; 35: 449–457. https://doi.org/10.1016/j.neuroimage.2006.12.032 PMID: 17293127
- Asavasopon S, Rana M, Kirages DJ, Yani MS, Fisher BE, Hwang DH, et al. Cortical activation associated with muscle synergies of the human male pelvic floor. J Neurosci. 2014; 34: 13811–13818. <a href="https://doi.org/10.1523/JNEUROSCI.2073-14.2014">https://doi.org/10.1523/JNEUROSCI.2073-14.2014</a> PMID: 25297107
- Kutch JJ, Yani MS, Asavasopon S, Kirages DJ, Rana M, Cosand L, et al. Altered resting state neuromotor connectivity in men with chronic prostatitis/chronic pelvic pain syndrome: A MAPP: Research Network Neuroimaging Study. Neuroimage Clin. 2015; 8: 493–502. <u>https://doi.org/10.1016/j.nicl.2015.05.013 PMID: 26106574</u>
- Kilpatrick LA, Kutch JJ, Tillisch K, Naliboff BD, Labus JS, Jiang Z, et al. Alterations in resting state oscillations and connectivity in sensory and motor networks in women with interstitial cystitis/painful bladder syndrome. J Urol. 2014; 192: 947–955. https://doi.org/10.1016/j.juro.2014.03.093 PMID: 24681331
- 74. Clemens JQ, Meenan RT, Rosetti MC, Gao SY, Calhoun EA. Prevalence and incidence of interstitial cystitis in a managed care population. J Urol. 2005; 173: 98–102; discussion 102. https://doi.org/10. 1097/01.ju.0000146114.53828.82 PMID: 15592041
- **75.** Blok BF, Sturms LM, Holstege G. Brain activation during micturition in women. Brain. 1998; 121 (Pt 11): 2033–2042.
- 76. Ness TJ, Lewis-Sides A, Castroman P. Characterization of pressor and visceromotor reflex responses to bladder distention in rats: sources of variability and effect of analgesics. J Urol. 2001; 165: 968–974. PMID: <u>11176524</u>
- 77. Wang Z, Guo Y, Bradesi S, Labus JS, Maarek JM, Lee K, et al. Sex differences in functional brain activation during noxious visceral stimulation in rats. Pain. 2009; 145: 120–128. <u>https://doi.org/10.1016/j.pain.2009.05.025 PMID: 19560270</u>
- Legrain V, Iannetti GD, Plaghki L, Mouraux A. The pain matrix reloaded: a salience detection system for the body. Prog Neurobiol. 2011; 93: 111–124. <u>https://doi.org/10.1016/j.pneurobio.2010.10.005</u> PMID: 21040755
- 79. Duval ER, Javanbakht A, Liberzon I. Neural circuits in anxiety and stress disorders: a focused review. Ther Clin Risk Manag. 2015; 11: 115–126. https://doi.org/10.2147/TCRM.S48528 PMID: 25670901
- Kim JJ, Jung MW. Neural circuits and mechanisms involved in Pavlovian fear conditioning: a critical review. Neurosci Biobehav Rev. 2006; 30: 188–202. https://doi.org/10.1016/j.neubiorev.2005.06.005 PMID: 16120461
- Melzack R. Evolution of the neuromatrix theory of pain. The Prithvi Raj Lecture: presented at the third World Congress of World Institute of Pain, Barcelona 2004. Pain Pract. 2005; 5: 85–94. https://doi.org/ 10.1111/j.1533-2500.2005.05203.x PMID: 17177754