

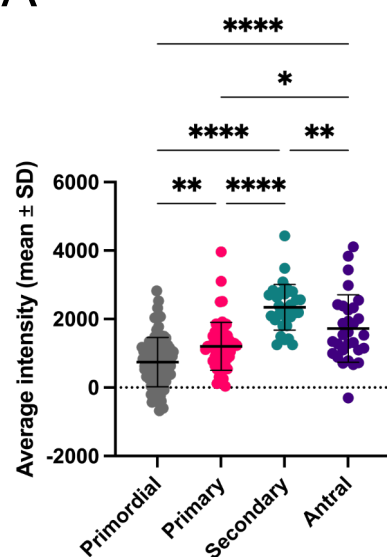
Additional File 1

Coordinated regulation of chromatin modifiers reflects organised epigenetic programming in mouse oocytes

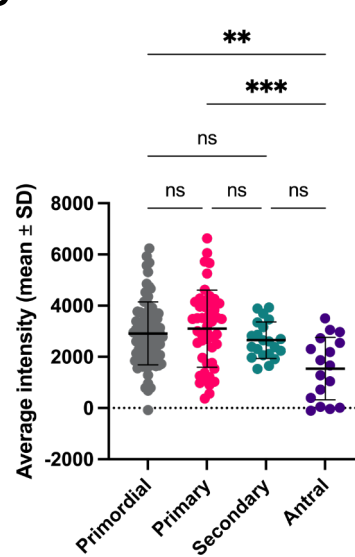
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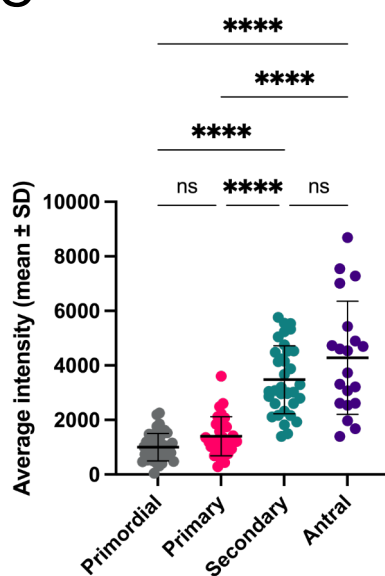
A SETD2 in WT growing oocytes



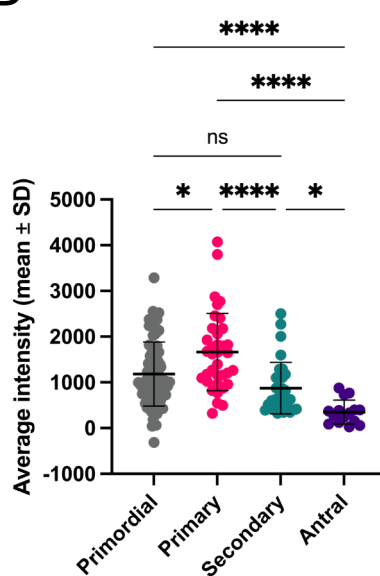
B H3K36me3 in WT growing oocytes



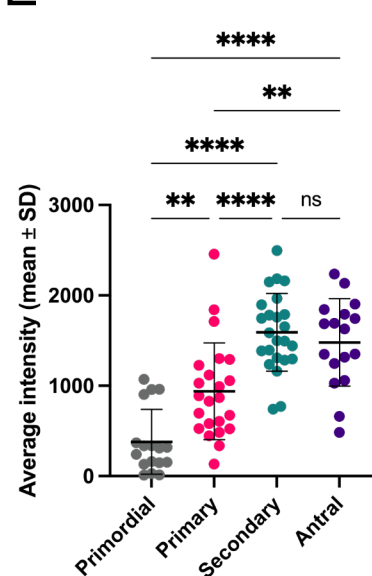
C RBBP7 in WT growing oocytes



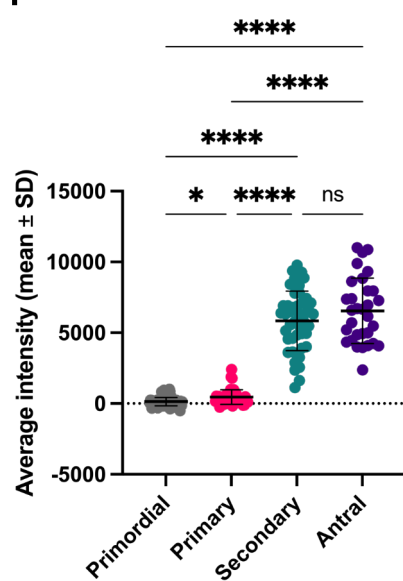
D H3K27me3 in WT growing oocytes



E KDM6A in WT growing oocytes



F DNMT3A in WT growing oocytes



G DNMT3L in WT growing oocytes

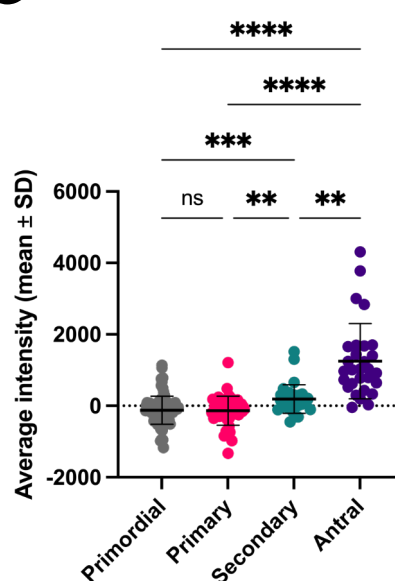


Figure S1. Assessment of the average staining intensity of epigenetic modifiers and modifications considering early and advanced antral follicles together. Quantification of SETD2 (A), H3K36me3 (B), RBBP7 (C), H3K27me3 (D), KDM6A (E), DNMT3A (F) and DNMT3L (G) within oocyte nuclei of primordial, primary, secondary, and antral follicles from wildtype adult mouse ovaries. Values represent average nuclear staining intensity with average cytoplasmic fluorescence removed to correct for non-specific background staining. Error bars represent mean \pm SD. Number of oocytes assessed at each stage in primordial, primary, secondary and antral follicle oocytes: **(A)** N=94 primordial, N=56 primary; N=32 secondary; N=30 antral; three biological replicates. **(B)** N=82 primordial; N=47 primary; N=21 secondary; N=17 antral; three biological replicates. **(C)** N=41 primordial; N=30 primary; N=35 secondary; N=20 antral; two biological replicates. **(D)** N=84 primordial; N=37 primary; N=32 secondary; N=15 antral; three biological replicates. **(E)** N=17 primordial; N=23 primary; N=24 secondary; N=17 antral; two biological replicates. **(F)** N=86 primordial; N=45 primary; N=46 secondary; N=31 antral; three biological replicates. **(G)** N=92 primordial; N=46 primary; N=31 secondary; N=29 antral; three biological replicates. **(A, E)** ns: not significant, $*P<0.05$, $**P<0.01$, $****P<0.0001$, one-way ANOVA plus Tukey's multiple comparisons test, **(B, C, D, F, G)** ns: not significant, $*P<0.05$, $**P<0.01$, $***P<0.001$, $****P<0.0001$, Kruskal-Wallis test with Dunn's multiple comparisons test.

Primordial SETD2

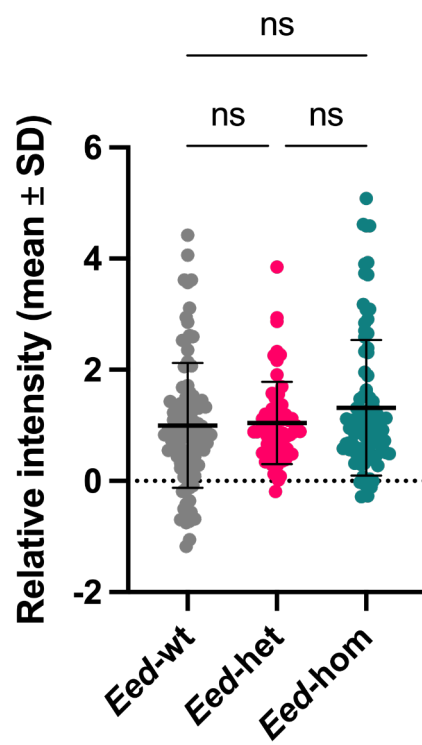


Figure S2. SETD2 levels in primordial follicle oocytes were similar between genotypes before *Zp3Cre*-mediated *Eed* deletion. Quantification of SETD2 within oocyte nuclei of primordial follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Values represent average nuclear staining intensity with average cytoplasmic fluorescence removed to correct for non-specific background staining. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. Three biological replicates with the following number of primordial follicle oocytes assessed: *Eed*-wt N=94, *Eed*-het N=63, *Eed*-hom N=91. ns: not significant, Kruskal-Wallis test with Dunn's multiple comparisons test.

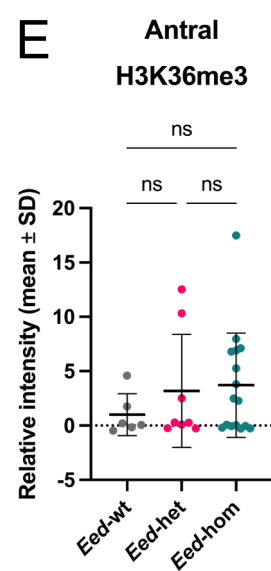
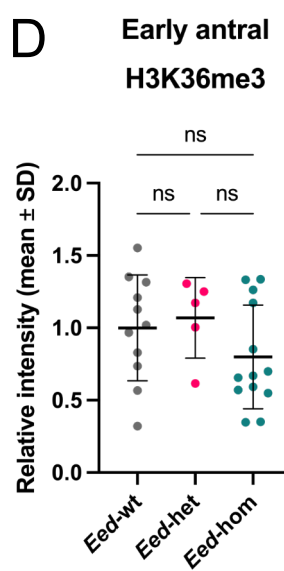
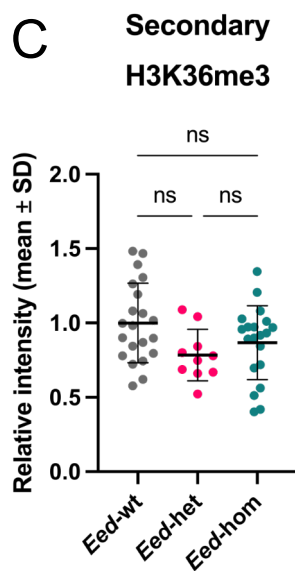
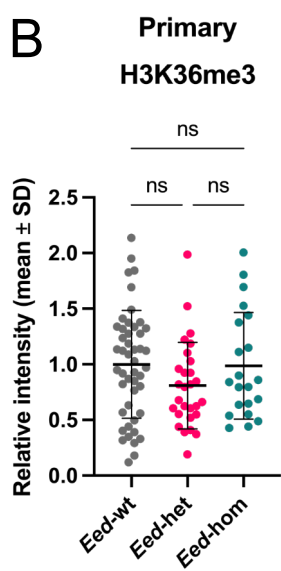
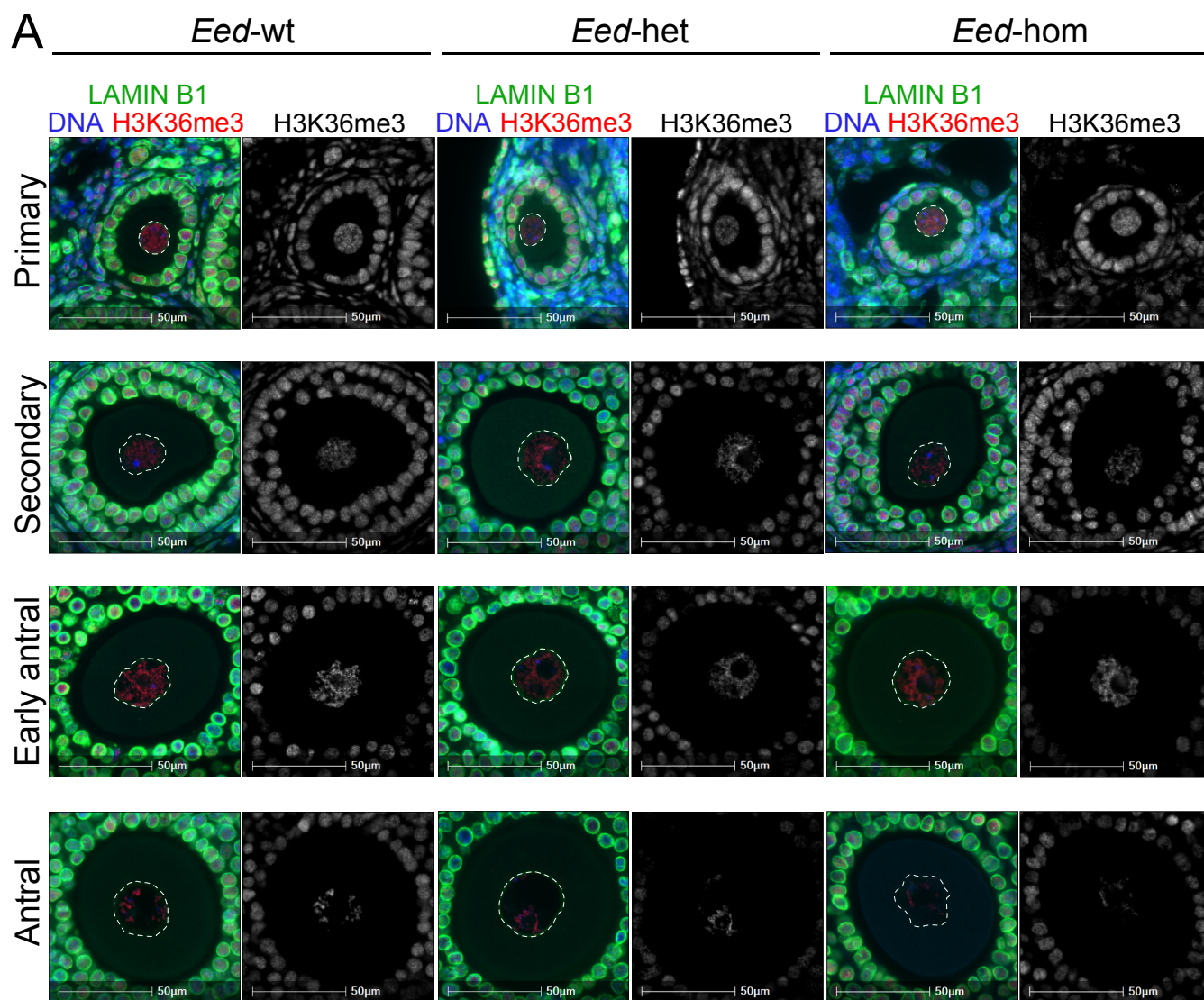


Figure S3. Average H3K36me3 levels were unchanged by loss of H3K27me3 in growing oocytes. (A) Representative IF images showing H3K36me3 (red, grey single channel) in primary, secondary, early antral and antral follicle oocytes from *Eed*-wt (left), *Eed*-het (middle), and *Eed*-hom (right) adult mouse ovaries. The oocyte nucleus is defined by Lamin B1 (green), and DNA is shown by DAPI (blue). Images are representative of multiple planes in both ovaries from three biological replicates for each genotype. Scale bars represent 50µm. (B-E) Quantification of H3K36me3 within oocyte nuclei of primary (B), secondary (C), early antral (D), and antral (E) follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Data is from three biological replicates for each genotype. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. N=47 *Eed*-wt, N=28 *Eed*-het, N=21 *Eed*-hom primary follicle oocytes. N=21 *Eed*-wt, N=10 *Eed*-het, N=20 *Eed*-hom secondary follicle oocytes. N=11 *Eed*-wt, N=5 *Eed*-het, N=13 *Eed*-hom early antral follicle oocytes. N=6 *Eed*-wt, N=8 *Eed*-het, N=16 *Eed*-hom antral follicle oocytes. ns: not significant, one-way ANOVA plus Tukey's multiple comparisons test.

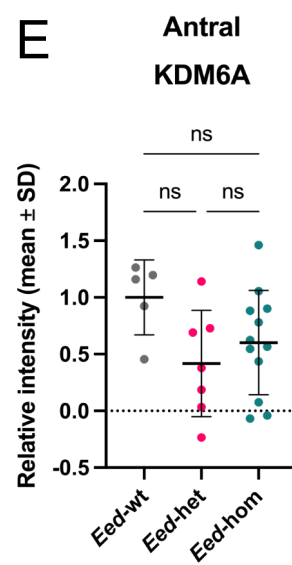
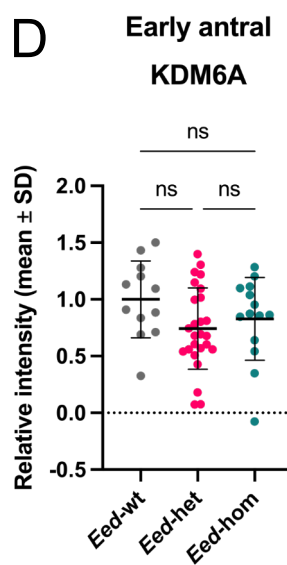
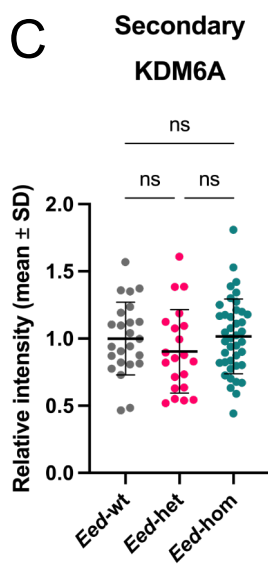
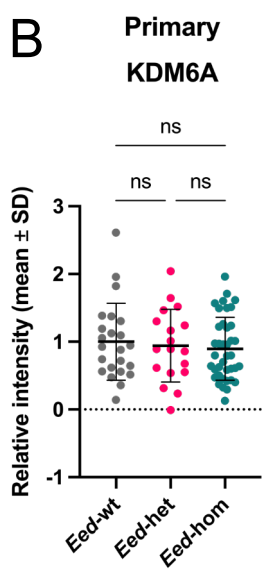
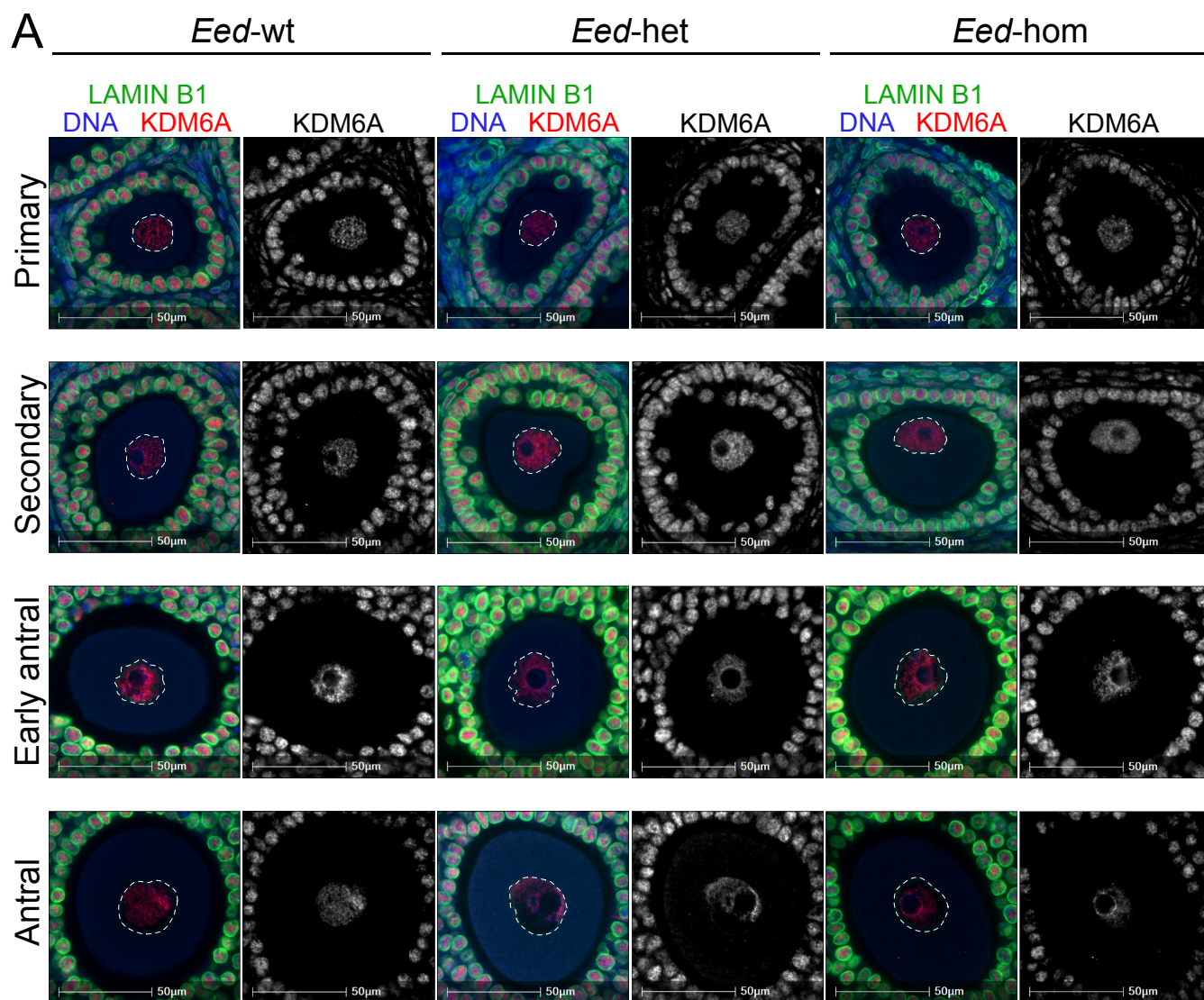


Figure S4. Loss of EED and H3K27me3 did not impact KDM6A levels throughout oocyte growth. (A) Representative IF images showing KDM6A (red, grey single channel) in primary, secondary, early antral and antral follicle oocytes from *Eed*-wt (left), *Eed*-het (middle), and *Eed*-hom (right) adult mouse ovaries. The oocyte nucleus is defined by Lamin B1 (green), and DNA is shown by DAPI (blue). Images are representative of multiple planes in both ovaries from two biological replicates for each genotype. Scale bars represent 50µm. (B-E) Quantification of KDM6A within oocyte nuclei of primary (B), secondary (C), early antral (D), and antral (E) follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Data is from two biological replicates for each genotype. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. N=23 *Eed*-wt, N=18 *Eed*-het, N=39 *Eed*-hom primary follicle oocytes. N=24 *Eed*-wt, N=21 *Eed*-het, N=41 *Eed*-hom secondary follicle oocytes. N=12 *Eed*-wt, N=26 *Eed*-het, N=14 *Eed*-hom early antral follicle oocytes. N=5 *Eed*-wt, N=7 *Eed*-het, N=12 *Eed*-hom antral follicle oocytes. ns: not significant, one-way ANOVA plus Tukey's multiple comparisons test.

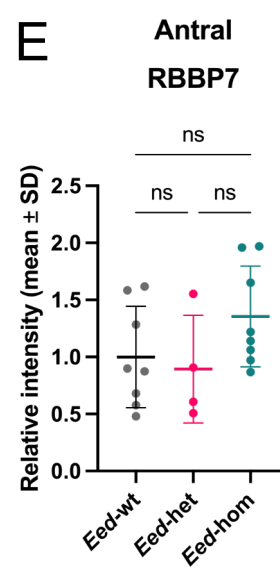
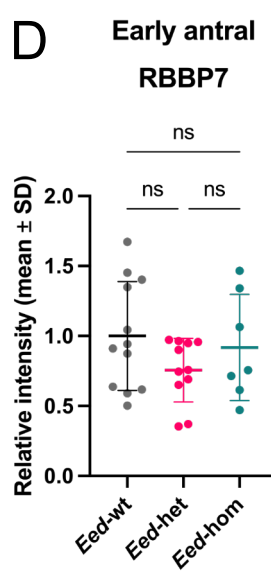
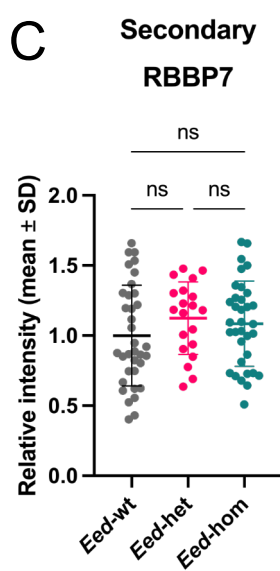
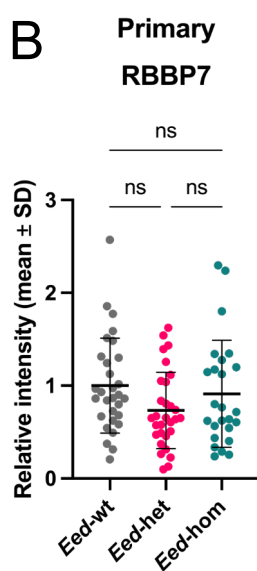
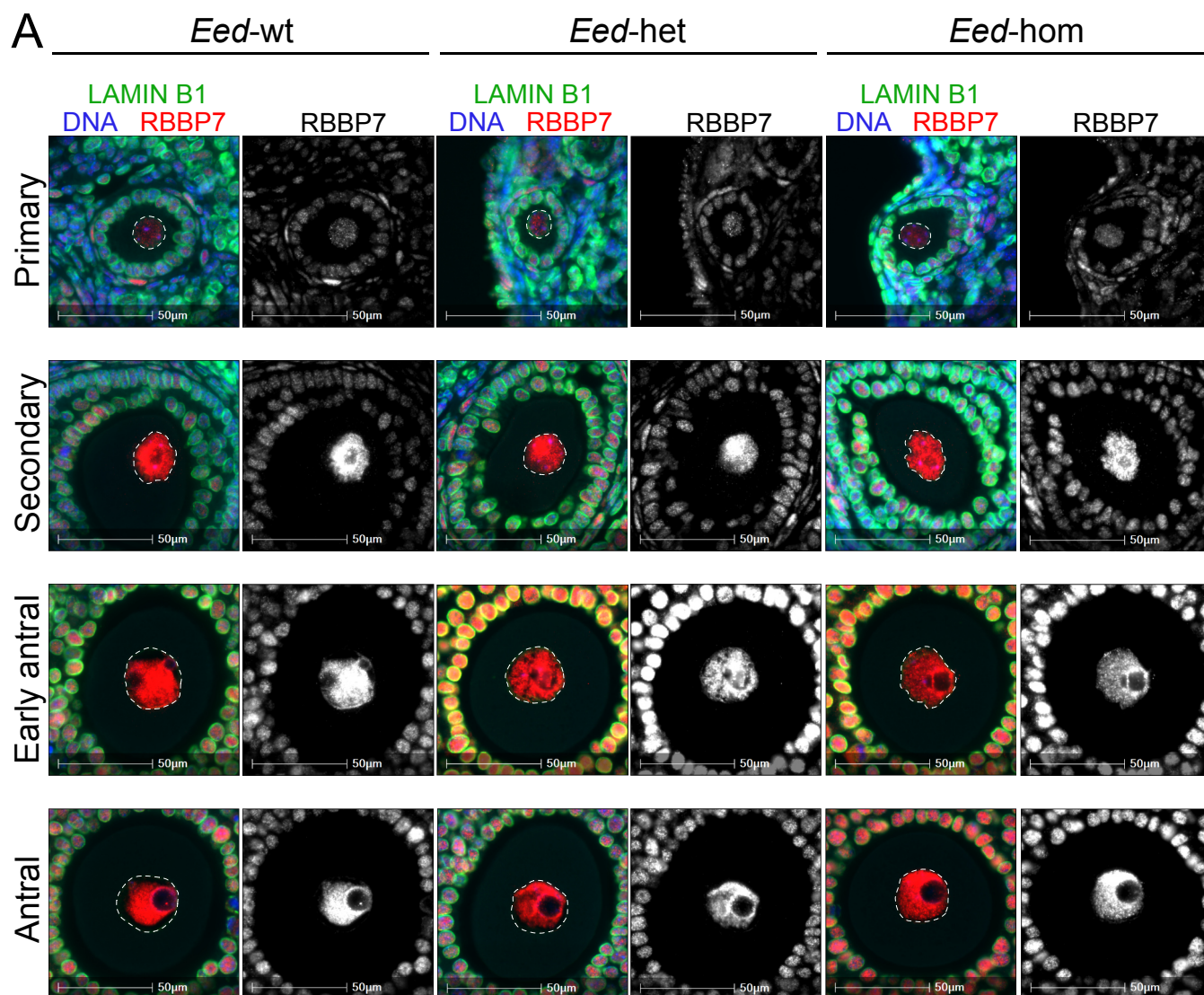


Figure S5. Average RBBP7 levels were not significantly different between *Eed*-wt, *Eed*-het, and *Eed*-hom growing oocytes. (A) Representative IF images showing RBBP7 (red, grey single channel) in primary, secondary, early antral and antral follicle oocytes from *Eed*-wt (left), *Eed*-het (middle), and *Eed*-hom (right) adult mouse ovaries. The oocyte nucleus is defined by Lamin B1 (green), and DNA is shown by DAPI (blue). Images are representative of multiple planes in both ovaries from two biological replicates for each genotype. Scale bars represent 50µm. (B-E) Quantification of RBBP7 within oocyte nuclei of primary (B), secondary (C), early antral (D), and antral (E) follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Data is from two biological replicates for each genotype. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. N=30 *Eed*-wt, N=30 *Eed*-het, N=25 *Eed*-hom primary follicle oocytes. N=35 *Eed*-wt, N=20 *Eed*-het, N=34 *Eed*-hom secondary follicle oocytes. N=12 *Eed*-wt, N=11 *Eed*-het, N=7 *Eed*-hom early antral follicle oocytes. N=8 *Eed*-wt, N=4 *Eed*-het, N=8 *Eed*-hom antral follicle oocytes. ns: not significant, one-way ANOVA plus Tukey's multiple comparisons test.

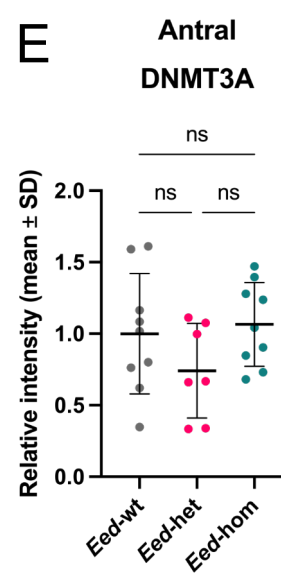
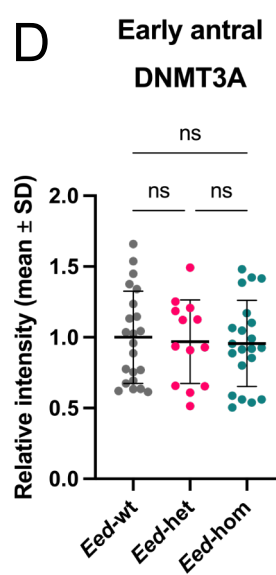
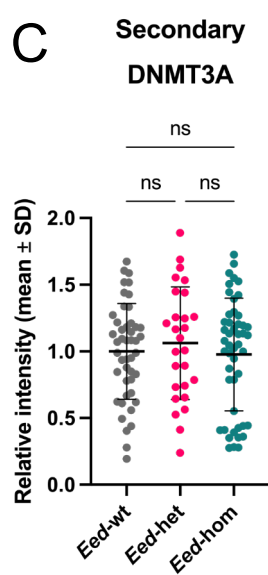
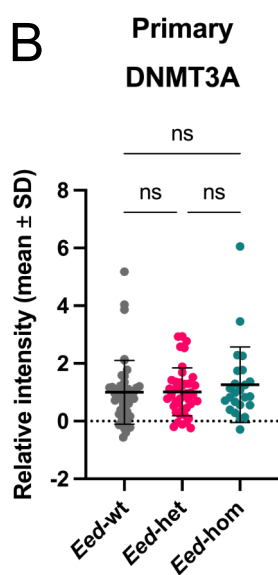
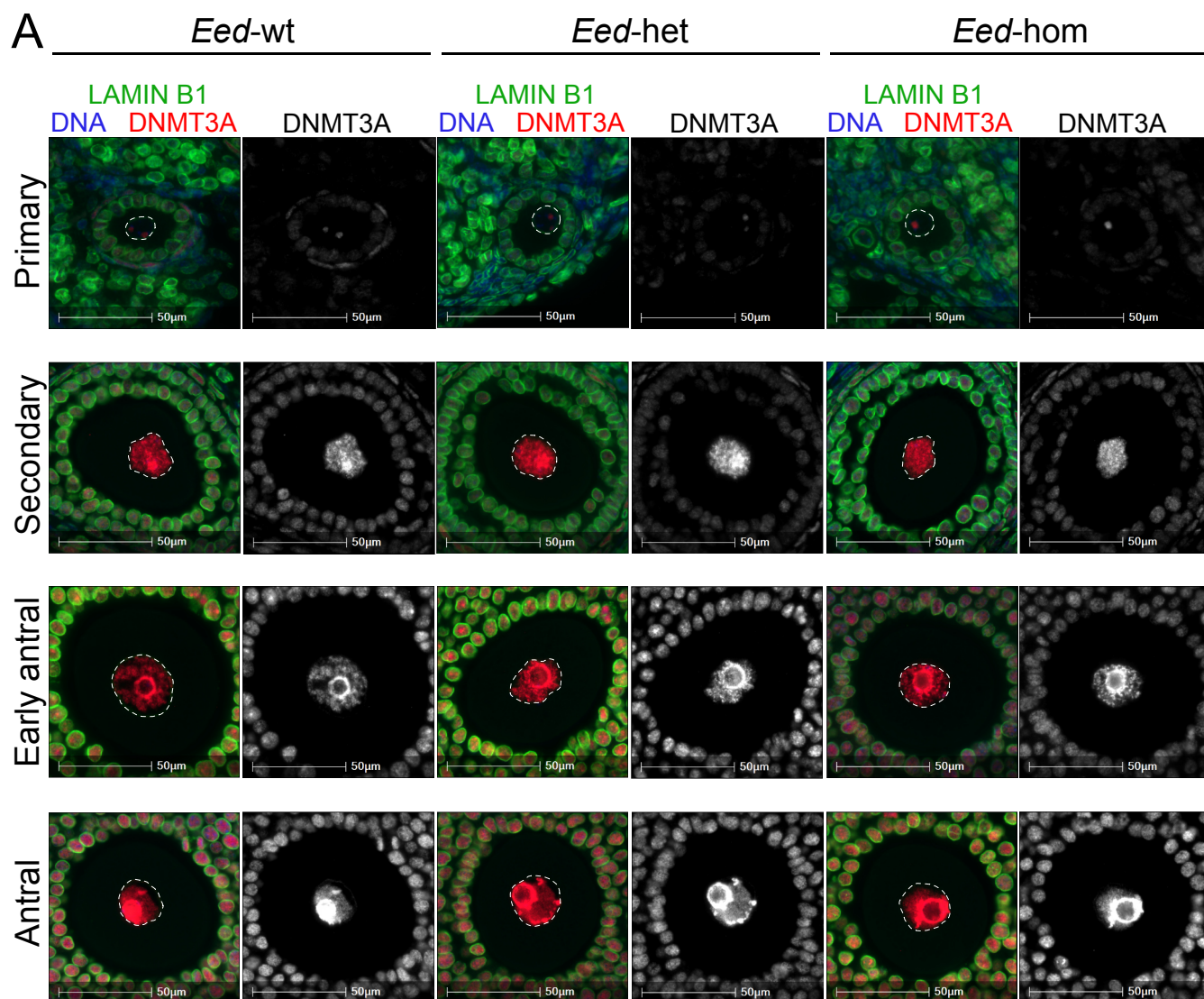


Figure S6. Loss of EED did not impact DNMT3A levels in growing oocytes. (A)

Representative IF images showing DNMT3A (red, grey single channel) in primary, secondary, early antral and antral follicle oocytes from *Eed*-wt (left), *Eed*-het (middle), and *Eed*-hom (right) adult mouse ovaries. The oocyte nucleus is defined by Lamin B1 (green), and DNA is shown by DAPI (blue). Images are representative of multiple planes in both ovaries from three biological replicates for each genotype. Scale bars represent 50µm. **(B-E)** Quantification of DNMT3A within oocyte nuclei of primary **(B)**, secondary **(C)**, early antral **(D)**, and antral **(E)** follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Data is from three biological replicates for each genotype. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. **(B)** N=45 *Eed*-wt, N=42 *Eed*-het, N=24 *Eed*-hom primary follicle oocytes. ns: not significant, Kruskal-Wallis test with Dunn's multiple comparisons test. **(C-E)** N=46 *Eed*-wt, N=27 *Eed*-het, N=48 *Eed*-hom secondary follicle oocytes. N=22 *Eed*-wt, N=13 *Eed*-het, N=21 *Eed*-hom early antral follicle oocytes. N=9 *Eed*-wt, N=7 *Eed*-het, N=9 *Eed*-hom antral follicle oocytes. ns: not significant, one-way ANOVA plus Tukey's multiple comparisons test.

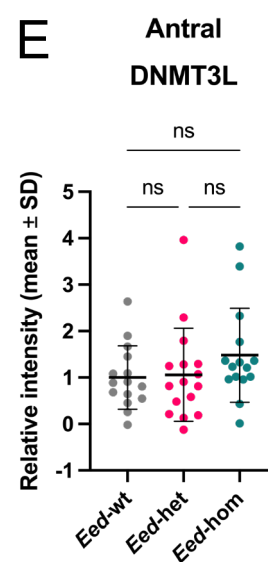
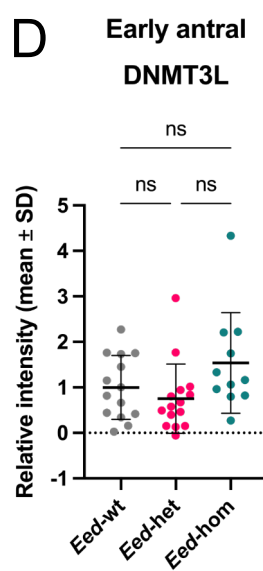
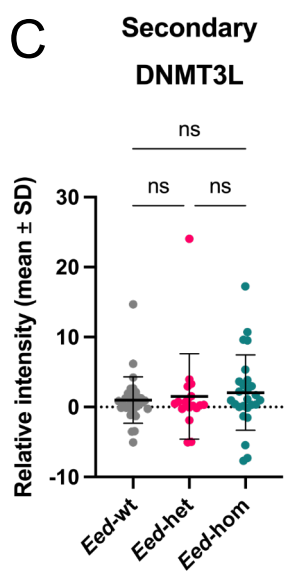
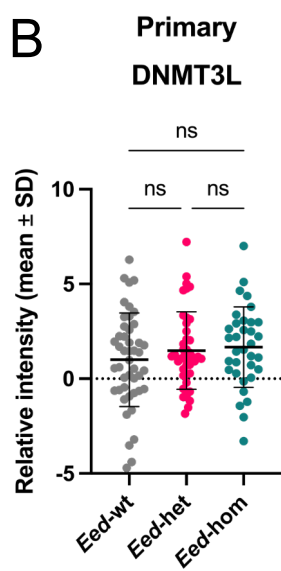
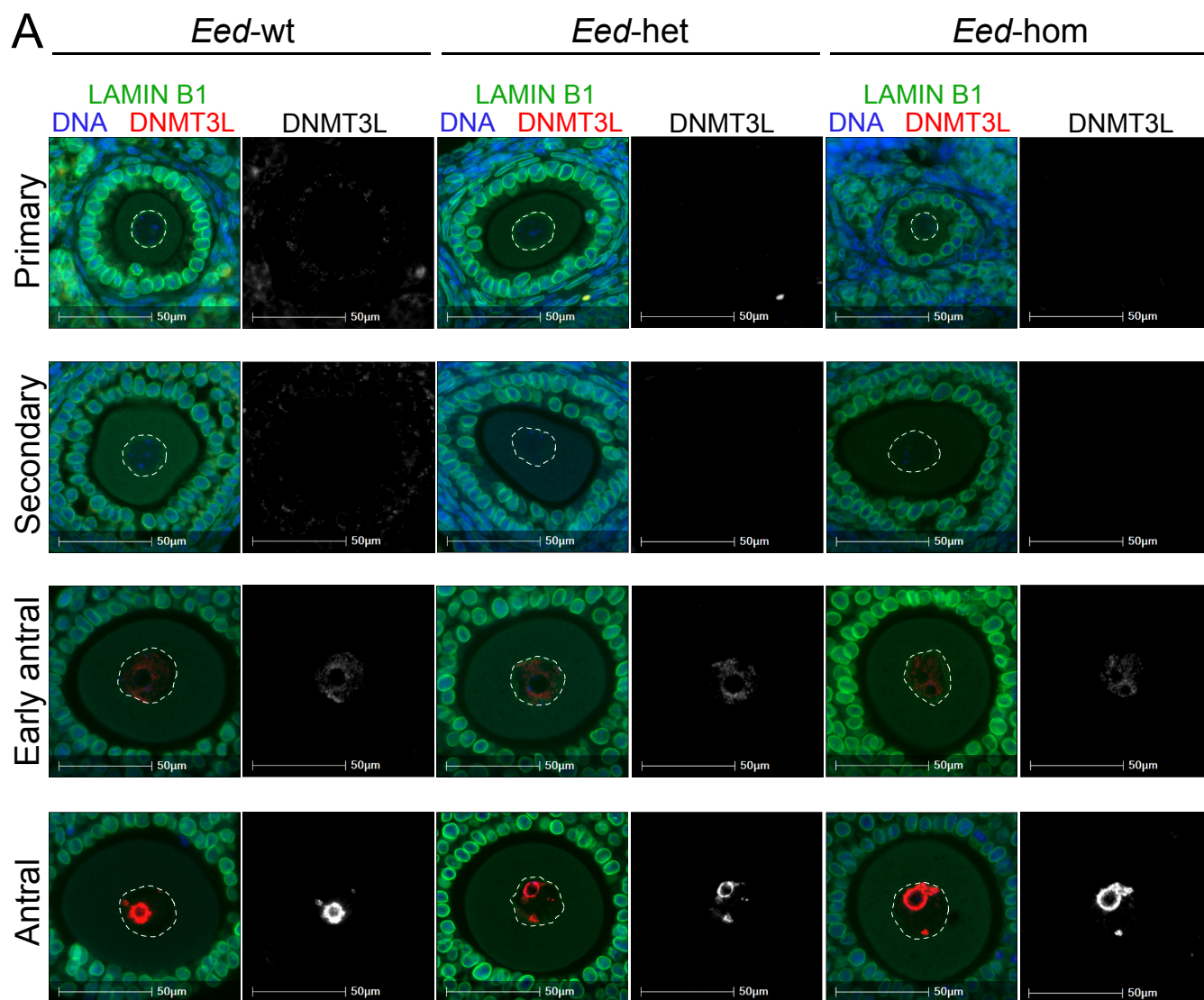


Figure S7. Loss of EED did not impact DNMT3L levels in growing oocytes. (A) Representative IF images showing DNMT3L (red, grey single channel) in primary, secondary, early antral and antral follicle oocytes from *Eed*-wt (left), *Eed*-het (middle), and *Eed*-hom (right) adult mouse ovaries. The oocyte nucleus is defined by Lamin B1 (green), and DNA is shown by DAPI (blue). Images are representative of multiple planes in both ovaries from three biological replicates for each genotype. Scale bars represent 50µm. **(B-E)** Quantification of DNMT3L within oocyte nuclei of primary **(B)**, secondary **(C)**, early antral **(D)**, and antral **(E)** follicles from *Eed*-wt, *Eed*-het, and *Eed*-hom adult mouse ovaries. Data is from three biological replicates for each genotype. Average intensity for *Eed*-het and *Eed*-hom samples are shown relative to *Eed*-wt set to 1.0. Error bars represent mean \pm SD. **(B, D, E)** N=46 *Eed*-wt, N=42 *Eed*-het, N=34 *Eed*-hom primary follicle oocytes. N=14 *Eed*-wt, N=15 *Eed*-het, N=11 *Eed*-hom early antral follicle oocytes. N=15 *Eed*-wt, N=16 *Eed*-het, N=15 *Eed*-hom antral follicle oocytes. ns: not significant, one-way ANOVA plus Tukey's multiple comparisons test. **(C)** N=31 *Eed*-wt, N=18 *Eed*-het, N=26 *Eed*-hom secondary follicle oocytes. ns: not significant, Kruskal-Wallis test with Dunn's multiple comparisons test.