

LETTER TO THE EDITOR

Varying effects of common tuberculosis drugs on enhancing clofazimine activity *in vitro*

Shuo Zhang^{1,2}, Wanliang Shi¹, Jie Feng¹, Wenhong Zhang³ and Ying Zhang^{1,3}*Emerging Microbes & Infections* (2017) 6, e28; doi:10.1038/emi.2017.24; published online 26 April 2017**Dear Editor,**

Clofazimine (CFZ), originally developed as an anti-tuberculosis (TB) drug in the 1950s,¹ is commonly used to treat leprosy and also nontuberculous mycobacterial infections.² Although CFZ has good activity against *Mycobacterium tuberculosis*, it was not used in the treatment of pulmonary TB mainly because it had the side effect of skin discoloration and there were other more effective drugs like isoniazid (INH), rifampin (RIF) and pyrazinamide (PZA) already available for the treatment of TB.² However, the increasing emergence of multi-drug-resistant TB (MDR-TB) has revived interest in the use of CFZ to treat MDR-TB.^{2,3}

Although resistance to CFZ has been shown to be mediated by mutations in *Rv0678*,^{4,5} *Rv1979c* or *Rv2535c* (*PepQ*),⁵ the mode of action of CFZ has remained poorly understood. CFZ appears to have multiple effects on *M. tuberculosis* including interference with redox cycling,¹ production of reactive oxygen species and membrane destabilization or dysfunction.^{6,7} CFZ is a bacteriostatic drug (MIC=0.06 µg/mL) with a slow action where it has little effect on the colony count until after 2 weeks.⁸ Heightened recent interest in this drug became apparent when CFZ added to the current MDR-TB regimen (called Bangladesh regimen), which is associated with shortening of the lengthy treatment from 18–24 to 9 months.³ Moreover, in the mouse model, CFZ was recently shown to shorten the treatment of drug susceptible TB from 6 to 3 months when added to the standard TB treatment regimen consisting of INH, RIF, PZA and ethambutol (EMB).⁹ These findings suggest that CFZ may have some unique activity on mycobacterial persisters and that certain TB drugs may synergize with CFZ or vice versa. However, so far, no information is available on the effect of other TB drugs on the activity of CFZ against *M. tuberculosis*. To address this question, in this study, we evaluated the effects of commonly used first-line and second-line TB drugs on the activity of CFZ against stationary phase *M. tuberculosis* culture enriched in persisters *in vitro* in a drug exposure assay.

A 3-week-old stationary phase *M. tuberculosis* H37Rv culture (10^{8–9} bacilli/mL) grown in 7H9 liquid medium containing 10% albumin-dextrose-catalase (ADC) was washed and diluted in 7H9 medium without ADC (5 × 10⁶ bacilli/mL), which was used for drug exposure

studies with CFZ in combination with the commonly used first-line drugs (RIF, PZA and EMB) and important second-line drugs amikacin (AMK), moxifloxacin (MXF), levofloxacin (LEV) and para-amino salicylate (PAS). Since INH is not active against stationary phase *M. tuberculosis*, INH was not included in the list of drugs evaluated. The drugs were dissolved in dimethyl sulfoxide or water as appropriate. CFZ (1 µg/mL) was incubated in combination with the following drugs at their *in vivo* relevant achievable blood concentrations: RIF (4 µg/mL), PZA (30 µg/mL at pH 6.0 or 6.8), EMB (3 µg/mL), AMK (8 µg/mL), MXF (2 µg/mL), LEV (8 µg/mL) and PAS (10 µg/mL) as CFZ containing two drug combinations, with single drug and drug-free controls, for various times (one, four, seven and fourteen days) without shaking. After drug exposure, the surviving bacteria in the above treatment groups were washed to remove drugs, diluted (undiluted, 1:10, and 1:100) and plated directly on drug-free 7H11 agar plates for colony-forming unit (CFU) counts to assess the effect of drug exposure without subculture. After incubation at 37 °C for 4 weeks, the CFU values for different treatments were determined (Table 1).

It is of interest to note that the CFZ activity was significantly enhanced at acid pH 6.0 as seen by less growth after 7-day drug exposure and no CFU remaining after 14 days (Table 1). In contrast, CFZ treatment alone at close to neutral pH 6.8 had poor activity against *M. tuberculosis* even after 14-day drug exposure (Table 1). The acid pH enhancement of CFZ activity was unexpected and not previously reported, and this is most likely caused by increased solubility of the poorly soluble CFZ (pKa=8.36) under acid pH. Future studies are needed to test this possibility in uptake experiments at acid pH with control drugs. Thus, it is possible that like PZA,¹⁰ the acid pH enhancement of CFZ activity may be relevant for *in vivo* situation during active inflammation that can produce acid pH. As a control, PZA at acid pH 6.0 was more active than at close to neutral pH (6.8), as expected (Table 1). Except RIF which had some activity against the stationary phase culture, other single drugs (AMK, MXF, LEV, PAS, CFZ and PZA at neutral pH) all had limited or poor activity against the 3-week-old stationary phase culture (Table 1).

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Table 1 Varying effects of commonly used TB drugs on enhancing the activity of CFZ against stationary phase *M. tuberculosis* H37Rv^a

Drug or drug combination treatment	CFU/mL ^b	CFU/mL	CFU/mL	CFU/mL
	1 day	4 days	7 days	14 days
7H9 ADC	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵
7H9 ADC pH 6	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵
CFZ	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	6.67×10 ⁴ ±1150
CFZ pH 6	>8×10 ⁵	>8×10 ⁵	3.40×10 ⁴ ±2830	0
PZA	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵
PZA pH 6	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	0
RIF	7.67×10 ⁴ ±4160	1.03×10 ⁵ ±6110	1.97×10 ⁵ ±1530	1.33×10 ² ±153
EMB	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵
AMK	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	1.70×10 ³ ±424
PAS	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵
MXF	3.03×10 ⁵ ±5860	3.83×10 ⁵ ±13 000	3.57×10 ⁵ ±7020	6.33×10 ² ±153
LEV	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	6.33×10 ² ±503
CFZ+RIF	2.47×10 ⁵ ±11 700	1.30×10 ⁵ ±13 000	1.27×10 ³ ±208	0
CFZ+EMB	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	1.03×10 ³ ±231
CFZ+AMK	>8×10 ⁵	>8×10 ⁵	6.10×10 ⁵ ±9000	0
CFZ+PAS	>8×10 ⁵	>8×10 ⁵	6.23×10 ⁵ ±10 016	0
CFZ+MXF	>8×10 ⁵	>8×10 ⁵	>8×10 ⁵	0
CFZ+LEV	>8×10 ⁵	4.73×10 ⁵ ±5030	3.00×10 ⁴ ±4360	0
CFZ+PZA	6.00×10 ⁵ ±5000	>8×10 ⁵	5.27×10 ⁵ ±5030	0
CFZ+PZA pH 6	7.27×10 ⁵ ±7020	6.77×10 ⁵ ±5030	0	0

Abbreviations: clofazimine, CFZ; rifampin, RIF; ethambutol, EMB; amikacin, AMK; para-amino salicylate, PAS; moxifloxacin, MXF; levofloxacin, LEV.

^aThe varying effects of different TB drugs alone and in combination with CFZ on the survival of a 3-week-old stationary phase culture of *M. tuberculosis* H37Rv at different time points 1, 4, 7 and 14 days were assessed by CFU counts after drug exposure.

^bThe numbers are the average CFU±s.d. per mL after drug exposure, and '0' indicates no bacterial survival detected (*n*=3).

In CFZ drug combination studies, we ranked the CFZ enhancement effects by commonly used first-line and second-line TB drugs. We found that PZA was by far the most active drug in enhancing the CFZ activity at acid pH 6.0, followed by RIF, quinolones (MXF and LEV), AMK and PAS in decreasing order of activity (Table 1). In contrast, cell wall inhibitor EMB had no apparent effect on enhancing CFZ activity (Table 1). Although we looked for other drugs that enhance CFZ activity, in fact, the combination effects can be said to be a reflection of mutual enhancements of CFZ and other TB drugs. Thus, it is noteworthy that we found in a separate study that CFZ could enhance PZA activity against *M. tuberculosis* (Niu H *et al.*, submitted).

Despite the interesting observation of varying enhancement effects of CFZ activity exhibited by different TB drugs, the mechanisms involved remain to be determined and may differ in each specific case. For example, PZA enhancement of CFZ activity may be due to their concerted effect on disrupting the mycobacterial membranes, which are a known persister target especially at acid pH.¹⁰ In addition, PZA may also enhance the CFZ activity through interfering with energy production via inhibition of PanD (aspartate decarboxylase) involved in CoA biosynthesis¹¹ such that it would deplete energy required to drive efflux of CFZ leading to increased accumulation of CFZ inside the cells to enhance its activity. We also found RIF increased the activity of CFZ (Table 1), and this could be due to the synergistic effect of RIF on causing inhibition of transcription of CFZ target leading to increased CFZ activity in the presence of RIF.

Gatifloxacin or MXF and CFZ are both included in the 9-month Bangladesh regimen for treating MDR-TB.³ It is of interest to note that we found quinolone drugs MXF and LEV both enhanced the activity of CFZ against *M. tuberculosis* stationary phase cells (Table 1). In addition, we also observed AMK enhanced the activity of CFZ. Our finding that AMK enhanced the CFZ activity for *M. tuberculosis* is consistent with the previous finding that AMK was shown to enhance

CFZ activity against growing *M. abscessus in vitro*.¹² Our findings that multiple drugs including PZA, RIF (except cell wall inhibitor EMB) and second-line drugs (quinolones, AMK and PAS) enhanced the activity of CFZ or vice versa, suggest a more general or broad effect of CFZ on *M. tuberculosis*. This observation is likely due to disruption of CFZ on bacterial membranes,¹³ which is considered a good target for persister drugs.^{14,15} In addition, our findings that many frontline and second-line drugs such as PZA, new generation fluoroquinolones (gatifloxacin or MXF) and AMK all enhanced the activity of CFZ also help to explain the high efficacy of the CFZ-containing 9-month Bangladesh regimen.³

In summary, there is recent interest in understanding how CFZ might be involved in shortening the treatment of both MDR-TB and drug susceptible TB. The present study made a number of interesting observations that may help explain the unique ability of CFZ to shorten TB therapy, by demonstrating acid pH enhancement of CFZ activity, the varying degrees of enhancement of CFZ activity against stationary phase bacilli by different TB drugs, with PZA and RIF having the highest degree of enhancement, followed by fluoroquinolones (MXF and LEV), AMK and PAS. Future studies are needed to validate our *in vitro* findings reported here in animal models of TB infection.

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- Barry V, Belton JG, Conalty ML *et al.* A new series of phenazines (rimino-compounds) with high antituberculosis activity. *Nature* 1957; **179**: 1013–1015.
- Cholo MC, Steel HC, Fourie PB *et al.* Clofazimine: current status and future prospects. *J Antimicrob Chemother* 2012; **67**: 290–298.
- Van Deun A, Maug AK, Salim MA *et al.* Short, highly effective and inexpensive standardized treatment of multidrug-resistant tuberculosis. *Am J Respir Crit Care Med* 2010; **182**: 684–692.

- 4 Hartkoorn RC, Uplekar S, Cole ST. Cross-resistance between clofazimine and bedaquiline through upregulation of MmpL5 in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2014; **58**: 2979–2981.
- 5 Zhang S, Chen J, Cui P *et al*. Identification of novel mutations associated with clofazimine resistance in *Mycobacterium tuberculosis*. *J Antimicrob Chemother* 2015; **70**: 2507–2510.
- 6 Van Rensburg CE, Joone GK, O'Sullivan JF *et al*. Antimicrobial activities of clofazimine and B669 are mediated by lysophospholipids. *Antimicrob Agents Chemother* 1992; **36**: 2729–2735.
- 7 Yano T, Kassovska-Bratinova S, Teh JS *et al*. Reduction of clofazimine by mycobacterial type 2 NADH:quinone oxidoreductase: a pathway for the generation of bactericidal levels of reactive oxygen species. *J Biol Chem* 2011; **286**: 10276–10287.
- 8 Ammerman NC, Swanson RV, Tapley A *et al*. Clofazimine has delayed antimicrobial activity against *Mycobacterium tuberculosis* both in vitro and in vivo. *J Antimicrob Chemother* 2016; **72**: 455–461.
- 9 Tyagi S, Ammerman NC, Li SY *et al*. Clofazimine shortens the duration of the first-line treatment regimen for experimental chemotherapy of tuberculosis. *Proc Natl Acad Sci USA* 2015; **112**: 869–874.
- 10 Zhang Y, Shi W, Zhang W *et al*. Mechanisms of pyrazinamide action and resistance. *Microbiol Spectr* 2013; **2**: 1–12.
- 11 Zhang S, Chen J, Shi W *et al*. Mutations in panD encoding aspartate decarboxylase are associated with pyrazinamide resistance in *Mycobacterium tuberculosis*. *Emerg Microbes Infect* 2013; **2**: e34.
- 12 Ferro BE, Meletiadiis J, Wattenberg M *et al*. Clofazimine prevents the regrowth of *Mycobacterium abscessus* and *Mycobacterium avium* type strains exposed to amikacin and clarithromycin. *Antimicrob Agents Chemother* 2016; **60**: 1097–1105.
- 13 Oliva B, O'Neill AJ, Miller K *et al*. Anti-staphylococcal activity and mode of action of clofazimine. *J Antimicrob Chemother* 2004; **53**: 435–440.
- 14 Zhang Y, Wade MM, Scorpio A *et al*. Mode of action of pyrazinamide: disruption of *Mycobacterium tuberculosis* membrane transport and energetics by pyrazinoic acid. *J Antimicrob Chemother* 2003; **52**: 790–795.
- 15 Hurdle JG, O'Neill AJ, Chopra I *et al*. Targeting bacterial membrane function: an underexploited mechanism for treating persistent infections. *Nat Rev Microbiol* 2011; **9**: 62–75.



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