

RESEARCH

Open Access

Identification of a conserved linear B-cell epitope in the M protein of porcine epidemic diarrhea virus

Zhibang Zhang¹, Jianfei Chen¹, Hongyan Shi¹, Xiaojin Chen¹, Da Shi¹, Li Feng^{1*} and Bin Yang²

Abstract

Background: The major structural protein of coronaviruses, the membrane (M) protein, can elicit the formation of protective antibodies, but little information is available about the M protein of porcine epidemic diarrhea virus (PEDV). Identification of epitopes on the PEDV M protein will be helpful in the elucidation of the antigenic properties of this protein.

Results: One hybridoma cell line secreting anti-M protein monoclonal antibody (McAb) was generated and designated 4D4. To map the epitopes on the PEDV M protein, a total of 17 partially overlapping fragments covering the C-terminus of M protein were expressed as fusion proteins with a 6xHis tag or a GST tag. A linear motif, ¹⁹³TGWAFYVR²⁰⁰, was identified by enzyme-linked immunosorbent assay (ELISA) and western blot (WB) analysis using McAb 4D4. The motif ¹⁹⁵WAFYVR²⁰⁰ was the minimal requirement for reactivity, as demonstrated by removing amino acids individually from both ends of the motif ¹⁹³TGWAFYVR²⁰⁰. The result of WB analysis showed that the 4D4-defined epitope could be recognized by PEDV-positive serum, but not transmissible gastroenteritis virus (TGEV)-positive serum. Furthermore, this epitope was highly conserved among different PEDV strains, as shown by alignment and comparison of sequences.

Conclusion: A McAb, 4D4, directed against the M protein of PEDV, was obtained, and the 4D4-defined minimal epitope sequence was ¹⁹⁵WAFYVR²⁰⁰. The McAb could serve as a candidate for development of a McAb-based antigen capture ELISA for detection of PEDV. The epitope identified provides a basis for the development of epitope-based differential diagnostic techniques and may be useful in the design of epitope-based vaccines.

Background

Porcine epidemic diarrhea (PED), which is characterized by severe diarrhea, vomiting and dehydration, is a highly contagious enteric disease of swine and is caused by porcine epidemic diarrhea virus (PEDV) [1]. PED was first reported in England in 1971 [2] and the virus was identified in Belgium and United Kingdom for the first time [3]. Since then it has become prevalent in many swine-raising countries and it is one of the most important viral causes of diarrhea, resulting in heavy economic losses to the swine industry, mainly in European and

Asia [4-11]. Although the commercial vaccines are available to prevent and control of disease, damage caused by PEDV infection is serious and continuous.

PEDV, the etiologic agent of PED, belongs to the genus *Alphacoronavirus*, family *Coronaviridae*. It has a genome of single-stranded, positive-sense RNA that is approximately 28 kb in length. The spike protein (S, 180–220kDa), membrane protein (M, 27–32kDa), envelope protein (E, ~7kDa) and nucleocapsid protein (N, 55–58kDa) are the four structural proteins of PEDV [12]. The S protein and M protein are localized on the surface of the virion. The M protein of coronavirus is the most abundant component of the viral envelope. In silico analysis has suggested that the M protein consists of a triple-transmembrane segment flanked by a short amino-terminal domain on the exterior of the virion and a long carboxy-tail located inside the virion. The M

* Correspondence: fl@hvri.ac.cn

¹Division of Swine Infectious Diseases, State Key Laboratory of Veterinary Biotechnology, Harbin Veterinary Research Institute of the Chinese Academy of Agricultural Sciences, No.427 Maduan Street, Nangang District, Harbin 150001, China

Full list of author information is available at the end of the article

protein of coronaviruses is indispensable in the assembly process and budding of virions [13-15]. The immune reaction to the M protein of coronaviruses plays an important role in the induction of protection and in mediating the course of the disease [16,17]. Monoclonal antibodies against the M protein of coronaviruses have virus-neutralizing activity in the presence of complement [18]. The M protein of coronavirus can also stimulate the production of alpha-interferon (α -IFN) [19].

Until now, identification of epitopes on the M protein of PEDV has not been reported. McAb development and epitope mapping of the PEDV M protein will provide the basis for the establishment of diagnostic methods for PEDV infection and also increase understanding of the antigenic structure of the M protein.

Results

Production of the fusion protein tM

The expression strategy for the fragments of the PEDV M protein is illustrated in Figure 1. In this study, the truncated M (tM) gene, encoding the C-terminus of the M protein (amino acids [aa], 99–226), was expressed as a His₆-fusion protein and a GST-fusion protein in *E. coli* BL21(DE3), respectively. Both of the fusion proteins, tM-His₆ and GST-tM, could react with porcine anti-PEDV serum as shown by WB analysis (data not shown), which implies that they had similar antigenicity to the native M protein of PEDV.

Production and characterization of M protein-specific McAb

The tM-His₆ protein was used as an immunogen to prepare the McAb, and the GST-tM protein was used as a coating antigen to establish the indirect ELISA for screening the antibody-secreting hybridoma cell lines.

After cell fusion and screening, one hybridoma clone that secreted McAb specific for the PEDV M protein was isolated and designated 4D4. The isotype of the McAb was IgG2b with κ light chain (data not shown). The 4D4 McAb could react with authentic M protein, as shown by immunofluorescent assay (IFA) (Figure 2 A). Moreover, in the WB analysis, the 4D4 McAb recognized a 27-kDa band of the PEDV M protein, while showing no reactivity against the TGEV virion (Figure 2 B). These results demonstrated that the 4D4 McAb recognized specifically the native M protein of PEDV.

Identification of the epitope by indirect ELISA

To map the antigenic epitope of the PEDV M protein, a set of truncated peptides were expressed in prokaryotes and used to identify the epitope by indirect ELISA. All of the truncated peptides with the expected molecular weights were expressed successfully either with a 6 \times His tag or with a GST tag (data not shown).

First, three overlapping fragments (M1, M2 and M3), covering the C-terminus of the M protein, were expressed as His₆-fusion peptides, and subjected to ELISA with

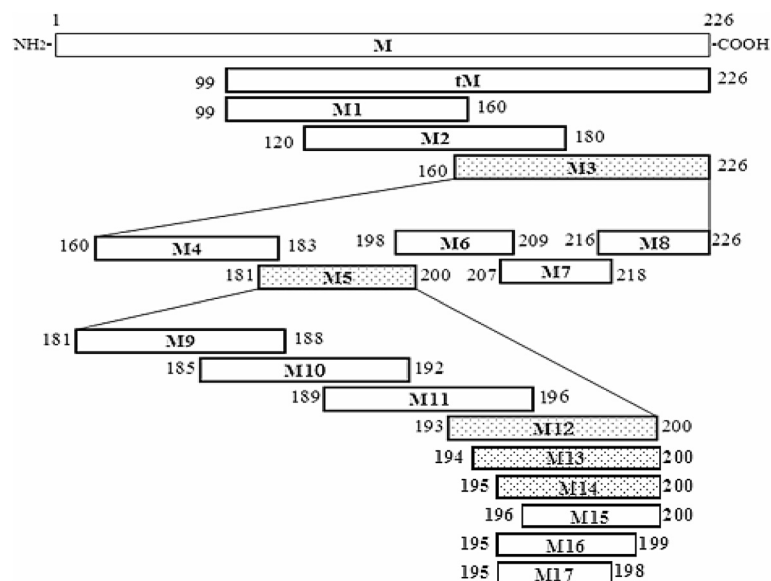
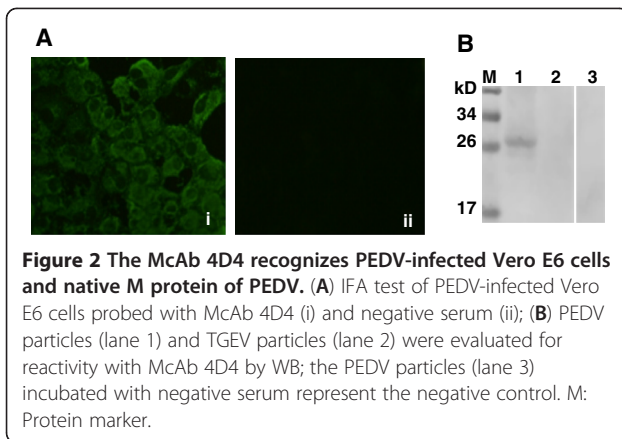


Figure 1 Schematic diagram of the relative locations of the truncated forms of the M protein of the PEDV CH/SHH/06 strain. The bars represent the truncated M proteins. The numbers represent the amino acid positions of the M protein. The bars filled with dots represent the peptides that were positive in WB analysis and ELISA with McAb 4D4 and the blank bars represent the peptides that were not recognized by McAb 4D4.



McAb 4D4 as the primary antibody. As demonstrated by ELISA, M3 showed reactivity with the McAb 4D4 (Figure 3 A). Subsequently, M3 was divided into five GST-fusion fragments (M4–M8) and the five peptides were probed by McAb 4D4. The results of the ELISA indicated that M5 harbored an antigenic epitope (Figure 3 A). To define the epitope more precisely, four 8-mer peptides (M9–M12) spanning the fragment M5 were expressed in a fusion form with GST and detected with the McAb 4D4. The results showed that M12 was recognized by the McAb 4D4, which indicates that the 4D4-specific epitope was $^{193}\text{TGWAFYVR}^{200}$ (Figure 3 A).

For fine mapping of the epitope of the M protein, we generated a panel of five shortened peptides (M13–M17) by deleting amino acids individually, at either the amino or the carboxy terminus in sequence, from the peptide M12 ($^{193}\text{TGWAFYVR}^{200}$). According to the results, the peptides M13 ($^{194}\text{GWAFYVR}^{200}$) and

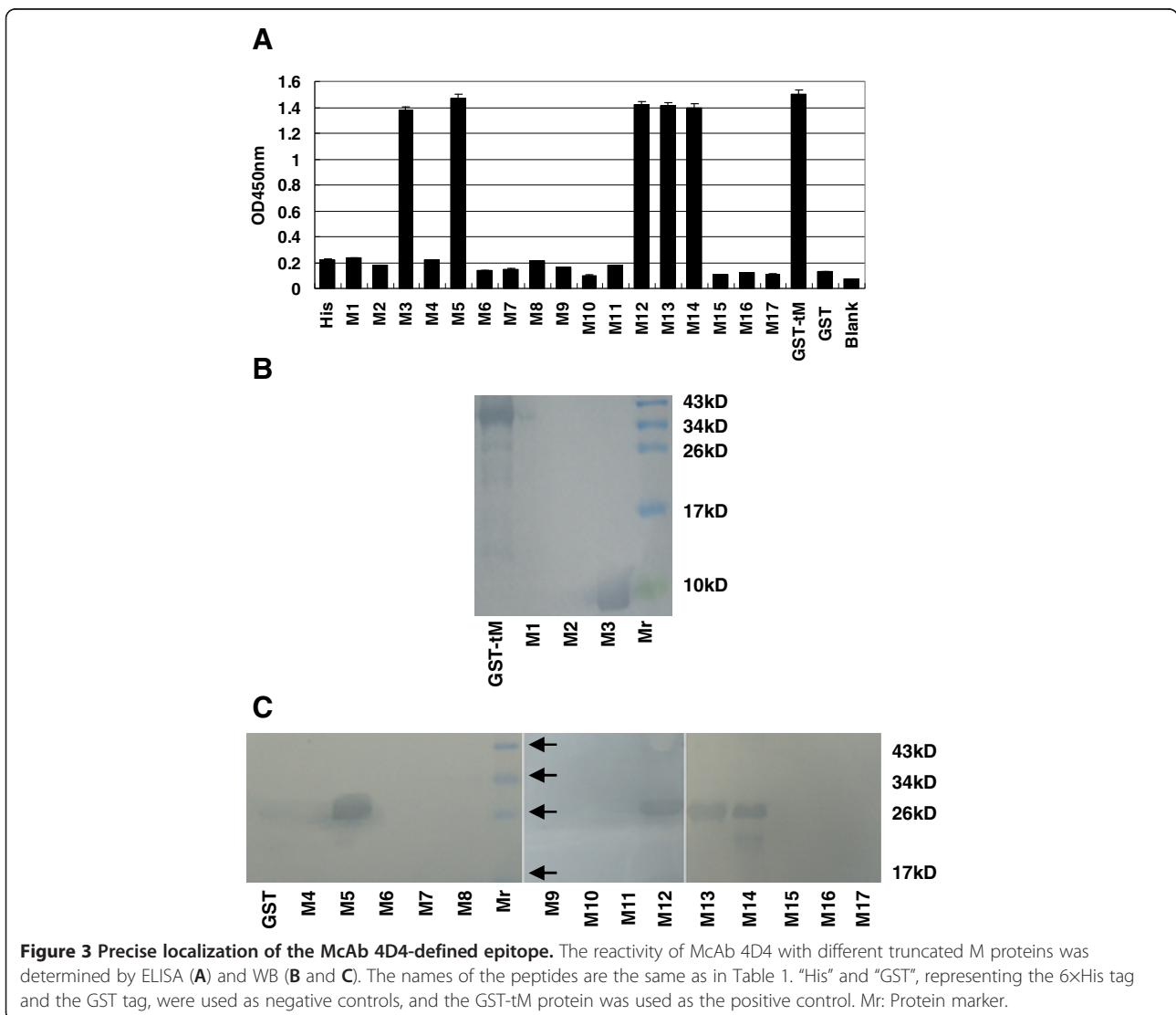


Figure 3 Precise localization of the McAb 4D4-defined epitope. The reactivity of McAb 4D4 with different truncated M proteins was determined by ELISA (A) and WB (B and C). The names of the peptides are the same as in Table 1. "His" and "GST", representing the 6xHis tag and the GST tag, were used as negative controls, and the GST-tM protein was used as the positive control. Mr: Protein marker.

M14 ($^{195}\text{WAFYVR}^{200}$) were recognized strongly by the McAb 4D4, whereas M15 ($^{196}\text{AFYVR}^{200}$), M16 ($^{195}\text{WAFYV}^{199}$) and M17 ($^{195}\text{WAFY}^{198}$) failed to show reactivity (Figure 3 A).

Mapping of the epitope by WB

The results of the ELISA were confirmed further by WB analysis and the results of WB were in accordance with the results of the ELISA (Figure 3 B and C).

Taken together, those results revealed that the minimal linear epitope required for reactivity with the McAb 4D4 was $^{195}\text{WAFYVR}^{200}$.

Reaction of the identified epitope with PEDV-positive serum

As shown by WB analysis, the epitope peptide $^{195}\text{WAFYVR}^{200}$, defined by McAb 4D4, could be recognized by PEDV-positive serum from pigs but not TGEV-positive serum, which suggests that this epitope has good reactivity and that porcine anti-PEDV serum contains antibody specific to this epitope (Figure 4 A and B).

Homology analysis

The M gene sequences for residues corresponding to this epitope from 10 different PEDV isolates were used for alignment analysis. The results showed that the epitope $^{195}\text{WAFYVR}^{200}$ is totally conserved among these PEDV strains (Figure 5 A), which indicates that the $^{195}\text{WAFYVR}^{200}$ sequence represents a conserved epitope on the M protein of PEDV. Analysis of the homologous sequences of the defined epitope from nine different coronaviruses from different groups within the family Coronaviridae demonstrated that the epitope recognized by 4D4 shares low homology among PEDV and other coronaviruses, and that only the residues A¹⁹⁶, Y¹⁹⁸ and V¹⁹⁹

are relatively conserved among all the coronaviruses selected (Figure 5 B).

Discussion

Mapping epitopes of viral proteins and defining the degree of conservation of identified epitopes may facilitate our understanding of the antigenic structure and virus-antibody interactions, and are useful for clinical application. The epitopes of the PEDV S protein have been reported, and these epitopes were shown to be able to induce the production of virus-neutralizing antibodies [20,21]. However, to date, the epitope on the M protein of PEDV has not been mapped.

Identification of B-cell epitopes on the M protein requires the preparation of McAb. To produce McAb against the M protein, we had tried to express the intact M protein, but this was unsuccessful (data not shown). As a result of unidentified but complex factors, it is very difficult to express the full-length M protein of coronaviruses [22,23]. Finally, we chose to express the truncated C-terminus of the M (tM) protein, and the tM protein was expressed in a fusion form either with a 6×His tag or a GST tag. The results of WB analysis showed that the fusion proteins tM-His₆ and GST-tM could be recognized by PEDV-positive serum, which indicates that both fusion proteins had good reactivity. In order to obtain more specific McAb, the fusion protein tM-His₆ was selected as an immunogen to elicit the formation of antibody. The fusion protein GST-tM was chosen to coat the ELISA plates to screen hybridomas, so that the coating antigen bore a different tag from the immunogen. After cell fusion and three cycles of selection, one McAb, designed 4D4, was chosen because of its specific reactivity with the fusion protein GST-tM as well as the native M protein of the PEDV CH/SHH/06 strain.

To map the epitope on the M protein, three overlapping peptides (M1, M2 and M3) covering the tM protein were expressed with a 6×His tag, respectively. Identified by ELISA and WB, the epitope was located at M3, which spans from residues 160 to 226 of the amino acid sequence of the M protein (Figure 3). Nine peptides spanning the identified epitope M3 were expressed in the form of GST-fusion peptides. Further experiments revealed that the epitope was in the M12 region ($^{193}\text{TGWAFYVR}^{200}$) of the M protein (Figure 3). To define more precisely the core epitope, amino acid residues were removed from the carboxy or amino terminals of the peptide M12 one by one to verify the minimal unit of the epitope that could be recognized by McAb 4D4. As demonstrated in Figure 3, the amino acid residues "W¹⁹⁵" and "R²⁰⁰" were essential for reactivity because deletion of either of them destroyed antibody binding. It is therefore reasonable to deduce that the minimal epitope recognized by McAb 4D4 is $^{195}\text{WAFYVR}^{200}$.

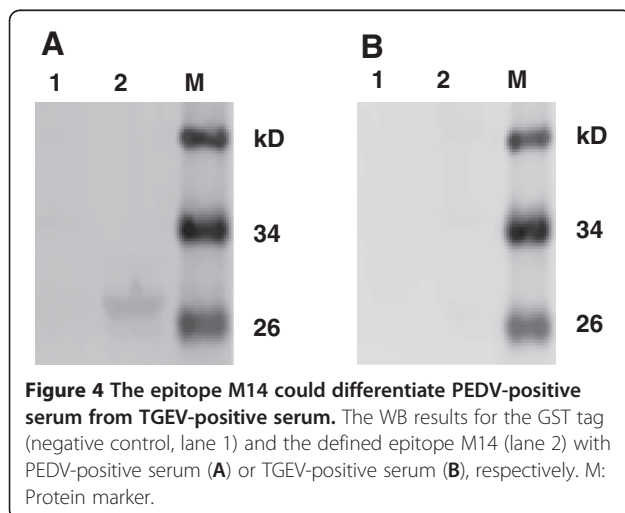


Figure 4 The epitope M14 could differentiate PEDV-positive serum from TGEV-positive serum. The WB results for the GST tag (negative control, lane 1) and the defined epitope M14 (lane 2) with PEDV-positive serum (A) or TGEV-positive serum (B), respectively. M: Protein marker.

A		VNASSGTG	WAFYVR	SKHGDYS	SAV
CH/SHH/06 (EU033966)	
JS/2004/2 (AY653205)	
LJB/03 (AY608890)	
LZC (EF185992)	.P.
QH (AY974335)	
JMe2 (D89752)	
Chinju99 (DQ845249)	
KPEDV-9 (AF015888)	
CV777 (AF353511)	
Br1/87 (Z24733)	

B		NASSGTG	WAFYVR	SKHGDYS
PEDV	CH/SHH/06 (FJ473392)
TGEV	HN2002 (AY587883)	KASSATG	WAYVVK	SKAGDYS
CCoV	68/09 (HQ450377)	KASSATG	WAYVVK	SKAGDYS
FCoV	C1Je (DQ848678)	KVTTATG	WAYVVK	SKAGDYS
HCoV	229E (M33560)	NSQNSTG	WVFVVR	VKHGDFS
HCoV	OC43 (M93390)	RISDTSG	FAVYVK	SKVGNYS
MHV	A59 (AY700211)	KVDGVS	GFAVYVK	SKVGNYS
SARS-CoV	BJ01 (AY536759)	RVGTDS	GFAAYNR	YRIGNYK
TCoV	540 (EU022525)	QSGNKKR	FATFVY	AKQSVDT
IBV	Beaudette (NC_001451)	QSGNKKR	FATFVY	AKQSVDT

Figure 5 Alignment of the amino acid sequences of the defined epitope and surrounding region with those of nine PEDV reference strains (A) and nine other coronaviruses (B). The dots represent residues that match the epitope exactly. The homologous regions of different coronaviruses that correspond to the identified epitope are in the box. Abbreviations of each virus and its strain are listed, and the corresponding GenBank accession numbers are shown in parentheses.

Sequence alignment of the amino acids of the identified epitope with those of other PEDV strains and other coronaviruses showed that this epitope was totally conserved across different strains of PEDV, but among different coronaviruses the epitope had only low homology.

To investigate whether the identified epitope ¹⁹⁵WAFYVR²⁰⁰ is specific to PEDV, the peptide was expressed as a GST-fusion protein, and subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and WB analysis. The results demonstrated that the epitope peptide did not react with TGEV-positive serum, but it showed reactivity with PEDV-positive serum, indicating that epitope ¹⁹⁵WAFYVR²⁰⁰ is specific for PEDV. Therefore, it was possible to distinguish anti-PEDV antibody from anti-TGEV antibody by employing the epitope as antigen. This will be very helpful in distinguishing PEDV from TGEV infection, because these two diseases have similar clinical and pathological signs, which obscures the differentiation of the two diseases, and diarrhea caused by PEDV and TGEV coinfection occurs frequently in the field.

To our knowledge, the epitopes of coronaviruses that have been identified are mainly focused on the S and N proteins. In the case of the M protein of coronaviruses, Qian *et al.* identified a B-cell antigenic epitope at the N-terminus of the severe acute respiratory syndrome

coronavirus (SARS-CoV) M protein [24], and Xing *et al.* reported a linear B-cell epitope in the M protein of avian infectious bronchitis coronavirus (IBV) [25]. The epitope of the IBV M protein identified by Xing *et al.* was ¹⁹⁹FATFVYAK²⁰⁶. Comparative analysis of the epitope ¹⁹⁹FATFVYAK²⁰⁶ with the corresponding regions of other coronaviruses revealed that the homologous region of the PEDV M protein was ¹⁹⁵WAFYVRSK²⁰² [25]. The epitope of the M protein of PEDV defined in this study was ¹⁹⁵WAFYVR²⁰⁰, and by comparison the homologous region of the IBV M protein was ¹⁹⁹FATFVY²⁰⁴. So, interestingly, the results of our study are consistent with findings regarding the IBV M protein.

Conclusion

In the current study, we generated a McAb, 4D4, and mapped the epitope on the M protein of PEDV using the McAb. The linear B-cell epitope identified, ¹⁹⁵WAFYVR²⁰⁰, was highly conserved among different PEDV isolates. Furthermore, this epitope could differentiate PEDV-positive serum from TGEV-positive serum. These data could deepen understanding of the antigenic structure of the M protein. The information regarding the epitope obtained in this study could be used to

establish a differential diagnostic test and may be helpful in the design of multi-epitope vaccines.

Methods

Cell lines, viruses and serum specimens

Myeloma cell line SP2/0 and Vero E6 cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Hyclone) in humidified 5% CO₂ atmosphere at 37°C. All culture media were supplemented with 10% heat-inactivated fetal bovine serum (PAA, Somerset, UK) and antibiotics (0.1mg/ml of streptomycin and 100IU/ml of penicillin). The PEDV CH/SHH/06 strain (EU033966) and TGEV attenuated strain H (EU074218) were maintained in our laboratory. Hyperimmune serum against PEDV and TGEV were obtained from pigs that had been inoculated with purified PEDV or TGEV.

Expression of the truncated form of the PEDV M gene

A truncated form, comprising 387 bp at the C-terminus of the M gene of PEDV CH/SHH/06 strain, was cloned. The sequences of the primers used for amplification of the gene in this study are shown in Table 1.

The amplified truncated form of the M gene (tM, 387bp), encoding the C-terminus of M protein (aa, 99–226), was cloned into prokaryotic expression vectors pET-30a (Novagen, USA) and pGEX-6p-1 (Pharmacia, Belgium), respectively. The inserts in the recombinant plasmids were sequenced and the confirmed plasmids were transformed into *E. coli* BL21(DE3) and induced by Isopropyl β-D-1-thiogalactopyranoside (IPTG). The expressed fusion proteins were analyzed with SDS-PAGE and detected by staining with Coomassie blue. For preparation of purified proteins, inclusion body proteins were separated by SDS-PAGE, the proteins of interest were excised, and the gel slices were crushed and added to an appropriate volume of sterilized phosphate buffered saline (PBS). After several cycles of freeze–thawing, the fusion proteins were dissolved in PBS, and the PBS was separated from the solid gel by centrifugation. The purity of the fusion proteins generated was analyzed by SDS-PAGE, and the purified proteins were used for ELISA and WB analysis.

Preparation and characterization of McAb against the tM protein

Female six-week-old BALB/c mice were primed subcutaneously with purified tM-His₆ protein emulsified with an equal volume of Freund's complete adjuvant (Sigma, USA). Two booster immunizations were given at two-week intervals with the fusion protein tM-His₆ in Freund's incomplete adjuvant. A final immunization containing purified tM-His₆ protein without adjuvant was injected intraperitoneally. Three days after the final injection, the mice were euthanized and their

splenocytes were harvested and fused with SP2/0 myeloma cells using polyethylene glycol 1450 (PEG1450, Sigma, USA). The hybridoma cells were seeded into 96-well plates and selected in hypoxanthine–aminopterin–thymidine (HAT) medium and hypoxanthine–thymidine (HT) medium in sequence. Cell culture supernatants of surviving clones were screened for antibody reactivity and specificity by ELISA. The positive hybridoma cell clones were subcloned thrice by limiting dilution. Ascites fluid was produced in separate primed BAB/c mice. The class and subclass of the McAb was determined using a SBA Clonotyping™ System/HRP (Southern Biotechnology Associates, Inc., Birmingham, AL35260, USA).

Animal care and all procedures were performed in accordance with animal ethics guidelines and approved protocols. The animal experiment was approved by Harbin Veterinary Research Institute. The animal Ethics Committee approval number is Heilongjiang-SYXK-2006-032.

Immunofluorescence assay was performed to assess whether the McAb 4D4 could recognize the native M protein. Vero E6 cells were infected with the CH/SHH/06 strain of PEDV and incubated at 37°C for 24 h. The cells were fixed with cold methanol for 10 min and then probed with McAb 4D4 and negative normal mouse serum for 1 h at 37°C. A fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG was used as a secondary antibody, and bound antibodies were visualized under a fluorescence microscope.

The specificity and reactivity of the McAb was determined by WB analysis using purified virus particles of the CH/SHH/06 strain of PEDV and the attenuated H strain of TGEV.

Expression of the truncated forms of the tM gene

To map the epitope of the M protein, a panel of 17 partially overlapping peptides (M1–M17) spanning the tM protein, as shown in Figure 1, were expressed successively. For peptides M1–M3, three pairs of primers were designed to amplify the genes of the fragments, and peptides M1–M3 were expressed with a 6×His tag, respectively. For each peptide M4–M17, a pair of oligonucleotide strands was synthesized. Each pair of oligonucleotide strands was annealed, and the resultant double-stranded DNA contained a BamHI and a XhoI cohesive terminus at the 5' and 3' ends. The annealed fragments were cloned into the expression vector pGEX-6p-1 and expressed as GST fusion proteins.

The recombinant plasmids thus constructed were sequenced and the confirmed recombinant plasmids were transformed into *E. coli* BL21(DE3). A series of fusion peptides were induced by IPTG and stained with Coomassie blue after SDS-PAGE. Purification of the short fusion peptides was carried out as mentioned above.

Table 1 Sequences of the primers used in this study

Peptides	Primer sequences (5'→3')	Position	Size of amplicon
tM-His ₆	F: ATAC <u>CATATG</u> AGCATTCCGGTTGTGGCG R: CGCCTCGAGGACTAAATGAAGCACT F: TAAGGATCCAGCATTCCGGTTGTGGCGCAG	295–678	384bp
GST-tM	R: CGGCTCGAGGACTAAATGAAGCACTTTCTCAC F: TATC <u>CATATG</u> AGCATTCCGGTTGTGG	295–678	384bp
M1	R: TATCTCGAGTACGCCAGTAGCAAC F: GCGCATATGCTCACTACTTCTGTGAT	295–480	186bp
M2	R: GCGCTCGAGATAGACAATTGTTGTAG F: GCACATATGGTACAGGTAAGTCAAT	358–540	183bp
M3	R: CTACTCGAGGACTAAATGAAGCACT F: <i>gatcc</i> GTACAGGTAAGTCAATTACCTAATTCGTCACAGTCGCCA	478–678	201bp
M4	AGGCCACTACAACAATTGTCTATGGACGTGTT taac R: <i>tcgag</i> tta AACACGTCCATAGACAATTGTTGTAGTGGCCTTGCGCA CTGTGACGAAATTAGGTAATTGACTTACCTGTACg F: <i>gatcc</i> GGACGTGTTGGTCGTTCAAGTCAATGCTTCATCTGGCACTG	478–549	72bp
M5	GTTGGGCTTTCTATGTCCGG taac R: <i>tcgag</i> tta CCGGACATAGAAAGCCCAACCAGTCCAGATGAAGCA TTGACTGAACGACCAACACGTCCg	541–600	60bp
M6	F: <i>gatcc</i> TCAGCTGTGAGTAATCCGAGTCCGGTTCTCACAGAT taac R: <i>tcgag</i> tta CACAGCTGAGTAGTCGCCGTGTTTGACCGGACATAg	592–627	36bp
M7	F: <i>gatcc</i> TCAGCTGTGAGTAATCCGAGTCCGGTTCTCACAGAT taac R: <i>tcgag</i> tta ATCTGTGAGAACCAGTCCGATTACTCACAGCTGAg F: <i>gatcc</i> CTCACAGATAGTGAGAAAGTGCTTCATTTAGT taac	619–654	36bp
M8	R: <i>tcgag</i> tta GACTAAATGAAGCACTTTCTCACTATCTGTGAGg	646–678	33bp
M9	F: <i>gatcc</i> GGACGTGTTGGTCGTTCAAGTCAAT taac R: <i>tcgag</i> tta ATTGACTGAACGACCAACACGTCCg	541–564	24bp
M10	F: <i>gatcc</i> CGTTCAGTCAATGCTTCATCTGGC taac R: <i>tcgag</i> tta ATTGACTGAACGACCAACACGTCCg	553–576	24bp
M11	F: <i>gatcc</i> GCTTCATCTGGCACTGGTTGGGCT taac R: <i>tcgag</i> tta AGCCCAACCAGTCCAGATGAAGCg	565–588	24bp
M12	F: <i>gatcc</i> ACTGTTGGGCTTTCTATGTCCGG taac R: <i>tcgag</i> tta CCGGACATAGAAAGCCCAACCAGTg	577–600	24bp
M13	F: <i>gatcc</i> GGTTGGGCTTTCTATGTCCGG taac R: <i>tcgag</i> tta CCGGACATAGAAAGCCCAACCg	580–600	21bp
M14	F: <i>gatcc</i> TGGGCTTTCTATGTCCGG taac R: <i>tcgag</i> tta CCGGACATAGAAAGCCCAg F003A <i>gatcc</i> GCTTTCTATGTCCGG taac	583–600	18bp
M15	R: <i>tcgag</i> tta CCGGACATAGAAAGCg R: <i>tcgag</i> tta GACATAGAAAGCCCAg	586–600	15bp
M16	F: <i>gatcc</i> TGGGCTTTCTAT taac R: <i>tcgag</i> tta GACATAGAAAGCCCAg	583–597	15bp
M17	F: <i>gatcc</i> TGGGCTTTCTAT taac R: <i>tcgag</i> tta ATAGAAAGCCCAg	583–594	12bp

The introduced restriction enzyme sites (NdeI and XhoI or BamHI and XhoI) are underlined. At the 5' and 3' terminal of each forward strand there is a sequence "gatcc" and "taac" (in italics), respectively. At the 3' and 5' terminal of each reverse strand there is a sequence "g" and "tcgagta" (in italics), respectively. When the forward and reverse oligonucleotides annealed they form a cohesive BamHI site or NdeI at the 5' terminus and a cohesive XhoI site at the 3' terminus. The bases "taa" and "tta" (in bold) were introduced into each pair of oligonucleotides to form a stop codon.

Identification of the epitope by indirect ELISA

Ninety-six-well microtiter plates were coated with the purified proteins in 0.1 M carbonate buffer (pH 9.6) at 4°C overnight and blocked with 5% skim milk for 1 h. After washing three times with PBST, 100 µl of McAb was added to the wells and incubated at 37°C for 1 h. Subsequently, the plates were washed thrice and incubated with horseradish peroxidase (HRP)-conjugated goat anti-mouse IgG at 37°C for 1 h. The color was developed and the reaction was terminated with 2 M H₂SO₄. The absorbance was measured at 450 nm. All assays were performed in triplicate and the results depict the average of the three values.

Mapping of the epitope by WB

The reactivity of the McAb with different truncated M proteins was determined further by WB. Purified truncated proteins were subjected to electrophoresis using 12% SDS-PAGE and transferred onto a nitrocellulose (NC) membrane. Non-specific antibody binding sites were blocked with 5% skim milk in PBS overnight at 4°C. The membranes were incubated with primary antibody at 37°C for 1 h and then washed three times with PBST. Next, the stripes were probed with HRP-conjugated goat anti-mouse IgG (Sigma, USA) for 1 h at 37°C. After three final wash steps, the color was developed using 3,3'-diaminobenzidine (DAB) substrate and was stopped finally by rinsing in deionized water.

Detection of the reactivity and specificity of the epitope defined by McAb 4D4

To investigate the reactivity and specificity of the epitope peptide defined by McAb 4D4, the peptide M14 "GST-WAFYVR" was purified and subjected to WB to investigate its reactivity with PEDV-positive serum, TGEV-positive serum and pre-immune serum from pigs, respectively. The WB was performed as described above, and HRP-conjugated goat anti-swine IgG was used as the secondary antibody.

Homology analysis

To analyze the conservation of the identified epitope among PEDV reference strains, the epitope sequence and flanking sequences of the M protein were compared with those of nine other selected PEDV strains using the DNAMAN software (Lynnon BioSoft Inc., USA). Alignment analysis was also performed between the defined epitope and the corresponding regions of other associated coronavirus strains using the DNASTAR Larsergene program (Windows version; DNASTAR Inc., Madison, WI, USA).

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

LF designed the experiment. ZZ carried out most of the experiments and wrote the manuscript. XC, BY and DS prepared the McAb and analyzed the data from the ELISA. HS participated in some of the experiments. JC and LF revised the manuscript. All the authors read and approved the final manuscript.

Acknowledgements

This study was supported by grants from the National Natural Science Foundation of China (30901081, 31172350), State Key Laboratory of Veterinary Biotechnology (SKLVBP201028), Natural Science Foundation for Distinguished Young Scholars of Heilongjiang province (JC201118) and Technology Research and Development Program of Harbin (2009AA6AN029).

Author details

¹Division of Swine Infectious Diseases, State Key Laboratory of Veterinary Biotechnology, Harbin Veterinary Research Institute of the Chinese Academy of Agricultural Sciences, No.427 Maduan Street, Nangang District, Harbin 150001, China. ²College of Veterinary Medicine, Inner Mongolia Agricultural University, No.306 Zhaowuda Street, Huhhot 010018, China.

Received: 22 September 2011 Accepted: 26 September 2012

Published: 1 October 2012

References

1. Pensaert MB, Debouck P: A new coronavirus-like particle associated with diarrhea in swine. *Arch Virol* 1978, **58**:243–247.
2. Wood EN: An apparently new syndrome of porcine epidemic diarrhea. *Vet Rec* 1977, **100**:243–244.
3. Chasey D, Cartwright SF: Virus-like particles associated with porcine epidemic diarrhea. *Res Vet Sci* 1978, **25**:255–256.
4. Debouck P, Callebaut P, Pensaert MB: Prevalence of the porcine epidemic diarrhea (PED) virus in the pig population of different countries. *Proc Int Pig Vet Soc Congr* 1982, **7**:53.
5. Takahashi K, Okada K, Ohshima K: An outbreak of swine diarrhea of a new-type associated with coronavirus-like particles in Japan. *Jpn J Vet Sci* 1983, **45**:829–832.
6. Jimenez G, Castro JM, Pozzo M, Correa I, Torre J, Enjuanes L: Identification of a coronavirus inducing porcine gastroenteritis in Spain. *Proc. Int. Pig Vet Soc Congr* 1986, **9**:186.
7. Hofmann M, Wyler R: Study of the occurrence of epizootic viral diarrhea in swine in Switzerland. *Switzerland Arch Tierheilkd* 1987, **129**:437–442.
8. Chae C, Kim O, Choi C, Min K, Cho WS, Kim J, Tae JH: Prevalence of porcine epidemic diarrhea virus and transmissible gastroenteritis virus infection in Korean pigs. *Vet Rec* 2000, **147**:606–608.
9. Chen JF, Sun DB, Wang CB, Shi HY, Cui XC, Liu SW, Qiu HJ, Feng L: Molecular characterization and phylogenetic analysis of membrane protein genes of porcine epidemic diarrhea virus isolates in China. *Virus Genes* 2008, **36**:355–364.
10. Puranaveja S, Poolperm P, Lertwatcharasarakul P, Kesdaengsakonwut S, Boonsoongnorn A, Urairong K, Kitikoon P, Choojai P, Kedkovid R, Teankum K, Thanawongnuwech R: Chinese-like strain of porcine epidemic diarrhea virus, Thailand. *Emerg Infect Dis* 2009, **15**:1112–1115.
11. Pyo HM, Kim IJ, Kim SH, Kim HS, Cho SD, Cho IS, Hyun BH: Escherichia coli expressing single-chain Fv on the cell surface as a potential prophylactic of porcine epidemic diarrhea virus. *Vaccine* 2009, **27**:2030–2036.
12. Park SJ, Kim HK, Song DS, Moon HJ, Park BK: Molecular characterization and phylogenetic analysis of porcine epidemic diarrhea virus (PEDV) field isolates in Korea. *Arch Virol* 2011, **156**:577–585.
13. de Haan CA, Kuo L, Masters PS, Vennema H, Rottier PJ: Coronavirus particle assembly: primary structure requirements of the membrane protein. *J Virol* 1998, **72**:6838–6850.
14. de Hann CA, Vennema H, Rottier PJ: Assembly of the Coronavirus envelope: homotypic interactions between the M proteins. *J Virol* 2000, **74**:4967–4978.
15. Arndt AL, Larson BJ, Hogue BG: A Conserved Domain in the Coronavirus Membrane Protein Tail Is Important for Virus Assembly. *J Virol* 2010, **84**:11418–11428.
16. Fleming JO, Shubin RA, Sussman MA, Casteel N, Stohlman SA: Monoclonal antibodies to the matrix (EI) glycoprotein of mouse hepatitis virus protect mice from encephalitis. *Virology* 1989, **168**:162–167.

17. Vennema H, de Groot RJ, Harbour DA, Horzinek MC, Spaan WJM: **Primary structure of the membrane and nucleocapsid protein genes of feline infectious peritonitis virus and immunogenicity of recombinant vaccinia viruses in kittens.** *Virology* 1991, **181**:327–335.
18. Woods RD, Wesley R, Kapke PA: **Neutralization of porcine transmissible gastroenteritis virus by Complement-dependent monoclonal antibodies.** *Am J Vet Res* 1988, **49**:300–304.
19. Laude H, Gelfi J, Lavenant L, Charley B: **Single amino acid changes in the viral glycoprotein M affect induction of alpha interferon by the coronavirus transmissible gastroenteritis virus.** *J Virol* 1992, **66**:743–749.
20. Sun DB, Feng L, Shi HY, Chen JF, Liu SW, Chen HY, Wang YF: **Spike protein region (aa 636–789) of porcine epidemic diarrhea virus is essential for induction of neutralizing antibodies.** *Acta Virol* 2007, **51**:149–156.
21. Sun D, Feng L, Chen J, Cui X, Chen H, Liu S, Tong Y, Wang Y, Tong G: **Identification of two novel B cell epitopes on porcine epidemic diarrhea virus spike protein.** *Vet Microbiol* 2008, **177**:231–235.
22. Gao SH, Zha EH, Li BH, Qiao XY, Tang LJ, Ge JW, Li YJ: **High-level prokaryotic expression of envelope exterior of membrane protein of porcine epidemic diarrhea virus.** *Vet Microbiol* 2007, **123**:187–193.
23. Zhang XL, Wang JR, Zhang Y, Chen ML, Zhang W, Yang S, Jiang WH: **Expression, purification and identification of recombinant SARS coronavirus membrane protein.** *Acta Biochimica et Biophysica Sinica* 2003, **35**:1140–1144.
24. Qian C, Din D, Tang Q, Zeng Y, Tang GX, Lu C: **Identification of a B-cell antigenic epitope at the N-terminus of SARS-CoV M protein and characterization of monoclonal antibody against the protein.** *Virus Genes* 2006, **33**:147–156.
25. Xing JJ, Liu SW, Han ZX, Shao YH, Li HX, Kong XG: **Identification of a novel B-cell epitope in the M protein of Avian Infectious Bronchitis Coronaviruses.** *J Microbiol* 2009, **47**:589–599.

doi:10.1186/1743-422X-9-225

Cite this article as: Zhang *et al.*: Identification of a conserved linear B-cell epitope in the M protein of porcine epidemic diarrhea virus. *Virology Journal* 2012 **9**:225.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

